

The Adenosine Pathway and Human Immunodeficiency Virus-Associated Inflammation

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Human immunodeficiency virus (HIV) is associated with an increased risk of age-associated comorbidities and mortality compared to people without HIV. This has been attributed to HIV-associated chronic inflammation and immune activation despite viral suppression. The adenosine pathway is an established mechanism by which the body regulates persistent inflammation to limit tissue damage associated with inflammatory conditions. However, HIV infection is associated with derangements in the adenosine pathway that limits its ability to control HIV-associated inflammation. This article reviews the function of purinergic signaling and the role of the adenosine signaling pathway in HIV-associated chronic inflammation. This review also discusses the beneficial and potential detrimental effects of pharmacotherapeutic strategies targeting this pathway among people with HIV.

Keywords. adenosine; HIV; inflammation, non-AIDS comorbidities; purinergic signaling.

Despite viral suppression on antiretroviral therapy (ART), human immunodeficiency virus (HIV) has been associated with elevated levels of systemic inflammation and immune activation, which accompanies an increased risk of morbidity and mortality from non-acquired immunodeficiency syndrome (AIDS)-associated chronic diseases [1, 2]. People with HIV (PWH) experience accelerated immunologic aging and develop cardiovascular disease (CVD), liver disease, and non-AIDS-defining cancers earlier than people without HIV (PWOH) of similar age [3, 4].

In a recent study, purinergic signaling has been implicated in regulating the immunopathogenesis of HIV [5, 6]. Purinergic receptor activation impacts phagocytosis, antigen presentation, cytotoxicity, chemotaxis, chemokine and cytokine release, and T lymphocyte differentiation [7, 8]. Therefore, tailoring pharmacotherapeutic interventions to target purinergic signaling could be an important strategy in regulating chronic inflammation and persistent immune activation associated with chronic HIV. This review highlights (1) the immunologic importance of adenosine triphosphate (ATP) and its nucleoside,

adenosine, in modulating the immunologic response and inflammation in chronic HIV infection and (2) the potential role of purinergic receptor-targeted therapies in the prevention and treatment of the chronic disease events seen in PWH.

THE IMMUNOLOGIC FUNCTION OF ADENOSINE

The relationship between extracellular ATP to adenosine is a major local signal of immunoactivation versus immunosurveillance, which is governed by extracellular and intracellular purinergic metabolism (Figure 1). In response to stress, large amounts of ATP are actively and passively released into the extracellular space. Adenosine triphosphate acts as a damage-associated molecular pattern (DAMP) and activates type 2 purinergic (P2) receptors [9]. The P2 receptors consist of 2 main subtypes, P2X and P2Y receptors. The P2X receptors are ATP-gated ionotropic channels that are generally involved in proinflammatory processes. In particular, P2X₇ stimulation causes further ATP release that triggers a positive feedback loop to amplify the ATP signal while recruiting appropriate cells to the area [10]. The P2Y receptors are G-protein-coupled receptors that are implicated in a broad range of functions, including facilitating platelet aggregation, vasodilation, cell migration, and immune responses [11].

Extracellular ATP is catabolized into adenosine 5'-monophosphate (5'-AMP) by a family of enzyme ectonucleotidases, the most important being dephosphorylase-1 (CD39). Subsequently, 5'-AMP is converted to adenosine mainly by ecto-5'-nucleotidase (CD73) and other tissue nonspecific alkaline phosphatases [12]. Extracellular adenosine can be further metabolized to a proinflammatory substrate, inosine, via adenosine deaminase or be transported intracellularly

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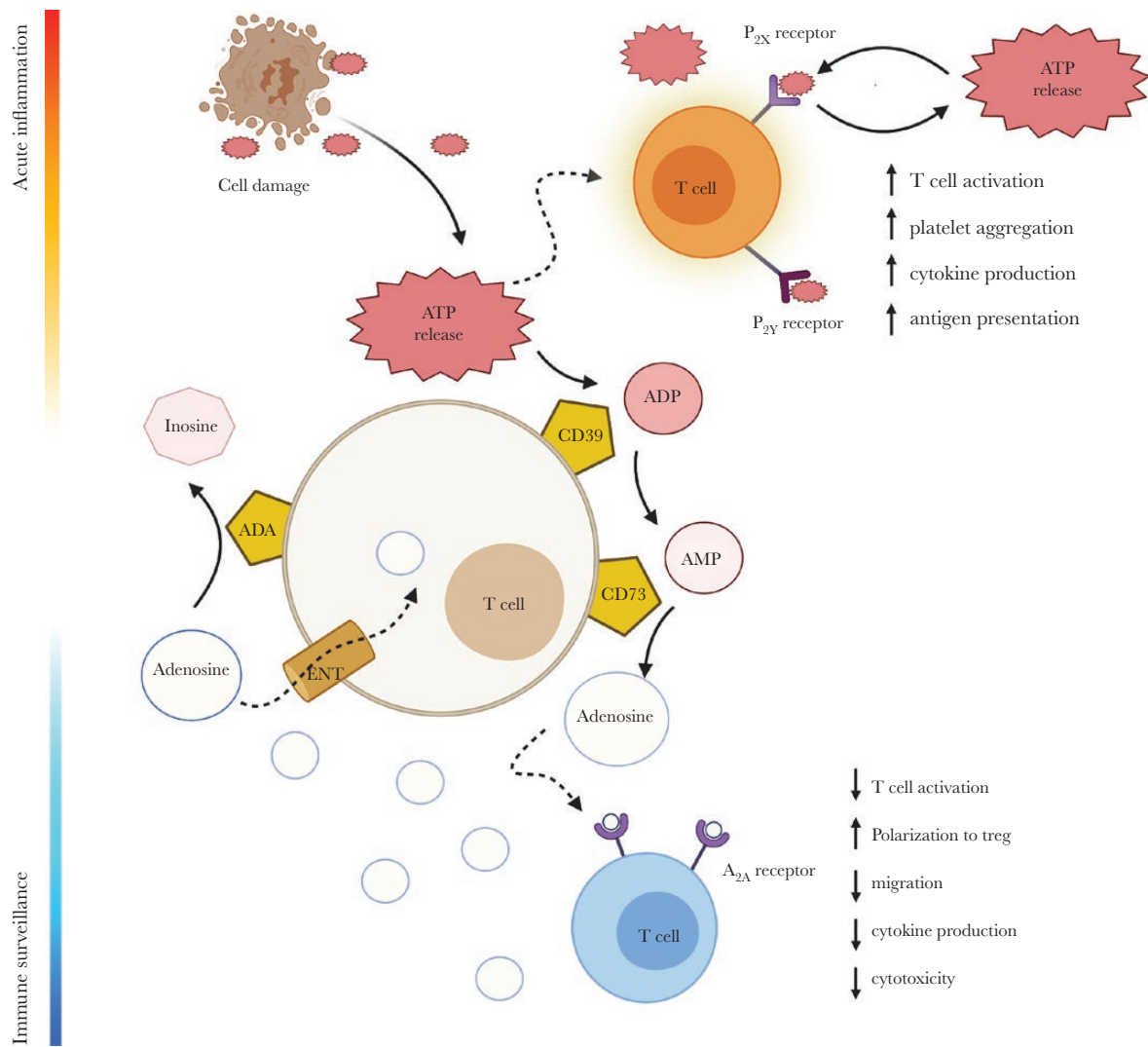


Figure 1. Schematic representation of transport pathways and extracellular enzymes involved in purinergic metabolism. The ratio of extracellular adenosine triphosphate (ATP) to adenosine signal for local immunoactivation (ATP-rich environment) versus immunosurveillance (adenosine-rich environment). The ATP is released into the extracellular space as a sign of cell damage, which promotes immune activation via purinergic 2X (P_{2X}) and purinergic 2Y (P_{2Y}) receptor activation. Extracellular ATP is broken down to adenosine diphosphate (ADP), adenosine monophosphate (AMP), and adenosine via CD39 and CD73 ectonucleotidases. Extracellular adenosine promotes local immunosuppression via activation of A_{2A} receptor. Extracellular adenosine concentration is regulated by transport into the cell via equilibrative nucleoside transporter (ENT) or conversion to inosine via adenosine deaminase (ADA).

via equilibrative nucleoside transporters (ENTs) [13, 14]. Extracellular adenosine activates type 1 purinergic (P1) receptors, which consists of subtypes A_1 , A_{2A} , A_{2B} , and A_3 , that modulate adenylyl cyclase and the 3'5'-cyclic monophosphate pathway [15]. The A_1 receptor is ubiquitous throughout the human body and typically has a proinflammatory effect [16]. However, A_2 receptors exhibit an anti-inflammatory role [17]. Specifically, activating the A_{2A} receptor suppresses neutrophil responses [18, 19], monocyte and macrophage recruitment as well as macrophage phagocytic function [20], and proinflammatory cytokine secretion [21, 22]. A_{2A} receptor agonists interfere with T-cell receptor signaling and suppress T-cell proliferation and effector function [23], ultimately producing anergic T cells [24, 25].

Constitutively low levels of extracellular ATP and adenosine are maintained by nucleoside and purine transporters under normal physiological conditions [26]. On most cells, including lymphocytes and endothelial cells, the surface expression of CD39 and CD73 is regulated by external stimuli that ultimately influence the concentration of adenosine in the local environment to mediate paracrine signaling [27]. The concentrations of extracellular ATP and adenosine are intrinsically regulated during inflammation and immune responses, which modulates the functions of myeloid and lymphoid cells [28]. During acute inflammatory responses, high levels of extracellular ATP act as a DAMP and trigger proinflammatory effector functions in a setting of low extracellular adenosine levels, including T lymphocyte migration and proliferation [29]. In the ATP-rich,

adenosine-poor environment, A_1 receptors work synergistically with P2 receptors to promote cell migration, cytotoxicity, apoptosis, and proinflammatory cytokine secretion in neutrophils [30], monocytes [31, 32], and macrophages [33, 34].

As inflammation persists, the concentration of extracellular adenosine increases as a result of the breakdown of ATP [35, 36]. Immune cells in the most injured areas produce an adenosine-rich environment to inhibit themselves, and other local immune cells, while allowing neighboring cells to continue eliminating the pathogen [37]. The protective increase in extracellular adenosine inhibits effector functions of neutrophils, macrophages, dendritic cells, and T lymphocytes [28]. Thus, a rise in extracellular adenosine and A_{2A} receptor expression provides a negative feedback mechanism to prevent further tissue damage [38]. These characteristics, combined with the short half-life of adenosine *in vivo*, allow for efficient paracrine and autocrine adenosine signaling among immune cells [39].

THE ADENOSINE PATHWAY IN HUMAN IMMUNODEFICIENCY VIRUS INFECTION

Given the established ability of the adenosine pathway in modulating immune function, multiple studies have examined the relationship between adenosine and HIV (Table 1). Specifically, antagonism of the P2X receptor inhibits HIV infection of CD4⁺ T lymphocytes [52]. For example, CD34⁺ hematopoietic progenitor cells from PWH who are immunologic nonresponders (INRs) overexpress P2X₇R, and inhibition of these receptors promotes maturation of CD4⁺ T cells [51]. Moreover, macrophages, which can act as HIV reservoirs, release HIV-1 virions from stored vacuoles when stimulated with P2X₇R agonists [53]. Human immunodeficiency virus infection directly impairs purinergic metabolism on inflammatory immune cells, which skews towards an adenosine-poor local environment, thereby promoting chronic immune activation [6]. In our study comparing CD39 and CD73 expression on Tregs, we found higher frequencies of ectonucleotidase coexpression as well as higher levels of adenosine in gut mucosal tissue in the nonprogressive model of simian immunodeficiency virus (SIV) infection in African green monkeys (AGM) compared with the progressive model in pigtailed macaques (PTM) [5]. This finding suggests a potential role for adenosine in AGM through the control of immune activation and inflammation, despite SIV infection, that prevents them from progressing to AIDS. When examining the functionality of adenosine *ex vivo*, we found that adenosine significantly suppressed cytokine production of CD4⁺ and CD8⁺ T cells in both AGM and PTM [5].

In addition to chronic immune activation, changes to the purinergic pathway during HIV infection are associated with T-cell exhaustion, immunosenescence, and immunosuppression, which mitigates the ability of the immune system to effectively manage chronic viral infection or cancer [54–56]. Human immunodeficiency virus infection is associated with

the downregulation of CD73⁺ and upregulation of CD39⁺ on CD8⁺ T cells, the latter of which has been identified as a marker of terminal exhaustion [50, 57]. Among viremic PWH, there is greater CD39, but not CD73, expression in natural killer cells, which correlates with viral load and markers of systemic inflammation [58].

Although the adenosine pathway could be protective in HIV-associated chronic inflammation and immune activation, its immunosuppressive function could have important implications in HIV persistence. Regulatory T cells (Tregs) represent important viral reservoirs during chronic HIV infection, and the frequency of CD39⁺ Tregs is correlated with Treg HIV deoxyribonucleic acid levels [59]. The Treg cells coexpress CD39 and CD73, which make them highly efficient in generating adenosine [60]. Because CD39 expression is upregulated in Tregs for PWH, this creates an adenosine-Treg positive feedback loop to promote a local adenosine-rich, immunosuppressant environment [48]. In addition, A_{2A} receptor activation increases Treg suppressive activity [25, 61, 62]. There is evidence *in vitro* that CD4⁺CD25⁺ Tregs in ART-treated PWH diminishes CD4⁺ and CD8⁺ T-cell function and proinflammatory cytokine production [63, 64]. Specifically, the adenosine/Treg-mediated suppression of CD4⁺ T cells inhibits interleukin (IL)-2 and interferon- γ release as well as gag-stimulated CD8⁺ T-cell cytotoxic activity [65–67]. Antibodies that block CD39 activity inhibit Treg-mediated suppression of CD8⁺ T-cell cytokine production, suggesting that adenosine metabolism is integral in the suppressive effects of Tregs in HIV infection [48]. Therefore, although HIV infection is associated with a persistent immunoactivated state, local enhanced production of adenosine by Treg cells may mediate inappropriate immune tolerance.

IMPACT OF ADENOSINE SIGNALING ON INFLAMMATION AND COMORBIDITIES IN TREATED HUMAN IMMUNODEFICIENCY VIRUS

Despite the extended survival of PWH on ART, virally suppressed individuals experience a greater rate of age-associated non-AIDS events compared with PWOH, and this is believed to be due to higher levels of inflammation [3]. Multiple factors contribute to this persistent inflammation. We have previously shown that PWH, regardless of viremia level or CD4⁺ T-cell reconstitution, have lower frequencies of CD4⁺ T cells expressing the rate-limiting enzyme CD73 and that CD4⁺CD73⁺ T-cell frequencies are associated with lower T-cell activation and C-reactive protein levels [49]. Likewise, Tóth et al [50] showed decreased frequencies of CD8⁺ T cells expressing CD73, and this correlated with immune activation and T-cell exhaustion. These findings suggest that alterations in the adenosine pathway are playing an important role in chronic HIV-associated inflammation.

An important factor contributing to the persistent inflammation in PWH is microbial translocation resulting from the

Table 1. Summary of Findings on the Adenosine Pathway and HIV Infection

Reference	Study Population	Endpoint	Major Findings
Blanco et al [40]	In vitro; murine clones expressing human CD26, CD4, and CXCR4	Investigation of gp120-induced inhibition of ADA binding to human CD26	<ul style="list-style-type: none"> Soluble gp120 and HIV particles able to inhibit ADA-CD26 binding CXCR4 cells enhanced gp120 inhibitory effects CXCR4 cells dependent on CD4 expression for inhibition
Fotheringham et al [41]	In vitro; primary human monocytes	Adenosine receptor influence on HIV Tat-induced intracellular calcium and TNF- α production	<ul style="list-style-type: none"> A_{2A}R activation inhibited Tat-induced calcium release, and reduction of intracellular calcium inhibited TNF-α production in monocytes Inhibitory actions of adenosine receptors relied on protein phosphatase activity
Pingle et al [42]	In vitro; PC12 cells and rat cerebellar granule neuron cultures	Neuroprotective potential of A ₁ receptor activation against HIV Tat-induced toxicity	<ul style="list-style-type: none"> A₁R activation suppressed the increase in calcium and nitric oxide mediated by HIV Tat protein A₁R inhibition of inducible nitric-oxide synthase expression dependent on NF-κB Activation of A₁R displayed protection against Tat-induced apoptosis in PC12 cells and cerebellar granule cells
By et al [43]	In vitro; CEM cells, a CD4 ⁺ human T-lymphoma cell line expressing A _{2A} R, CXCR4, and CCR5	Analysis of the influence of an agonist-like monoclonal antibody to A _{2A} R, Adonis, on CD4 ⁺ CEM T cells	<ul style="list-style-type: none"> Adonis assisted activation of AZAR and inhibited CEM cell growth A_{2A}R upregulation reduced CXCR4 and CCR5 expression without altering CD4 expression on CEM cells
Moreno-Fernandez et al [44]	Ex vivo; PBMCs from HIV ⁻ participants	Functional inspection of the suppressive abilities of Tregs including extracellular adenosine formation	<ul style="list-style-type: none"> Treg suppression occurred through gap junctions via a cAMP-dependent mechanism that activated protein kinase A in conventional T cells CD39 expression on Treg also played an important role during suppression
Martinez-Navio et al [45]	Ex vivo; 36 HIV ⁺ and 10 HIV ⁻ participants	Exploration of the influence of HIV on ADA costimulation in T cells	<ul style="list-style-type: none"> ADA increased T-cell proliferation and positively correlated with CD4⁺ percentage and count while negatively correlated with viral load HIV reduced ADA-induced cytokine production (IFN-γ, IL6, and IL-10) in T cells gp120 impaired ADA-CD26 interaction in HIV
Climent et al [46]	Ex vivo; 8 HIV ⁺ ; HIV ⁻ participants	Measurement of the immunologic usefulness of ADA as an adjuvant in HIV dendritic cell-based therapeutic vaccines	<ul style="list-style-type: none"> ADA-induced enhancement of CD4⁺ and CD8⁺ T-cell proliferation ADA increased cytokine production (IFN-γ, TNF-α, and IL6) to promote a Th1 response to improve T helper and CTL responses
Parish et al [47]	Ex vivo; PBMCs from HIV ⁻ participants	Investigation of the role of ADA on replicative senescence in human CD8 ⁺ T cells	<ul style="list-style-type: none"> ADA expression on CD8⁺ T lymphocytes was lost after successive cultures CD8⁺CD28⁺ T lymphocytes that are ADA⁺ had greater telomerase activity than ADA⁻ Lack of ADA expression subjected CD8⁺ T lymphocytes to prolonged adenosine exposure, which accelerated senescence and loss of CD28 expression
Nikolova et al [48]	Ex vivo; 39 HIV ⁺ , c-ART naive; 39 HIV ⁺ , c-ART stable; 25 HIV ⁻ participants	Examination of the CD39/adenosine axis involvement in the pathogenesis and progression of HIV	<ul style="list-style-type: none"> Downregulation of CD39 expression on Treg caused a higher CD8⁺ T-cell proliferation and cytokine production in HIV⁺ vs HIV⁻ individuals Higher level of A_{2A}R expression on untreated HIV⁺ individuals, which caused a higher susceptibility to CD39/adenosine-mediated inhibition In HIV⁺ participants, CD39⁺ Treg positively correlated with immune activation and inversely with CD4⁺ T-cell absolute counts A CD39 gene polymorphism, causing lower levels of expression, associated with LTNPs and may indicate slower progression of HIV-1 disease
Schuler et al [49]	Ex vivo; 36 HIV ⁺ and 10 HIV ⁻ participants	Investigation of adenosine-induced immunomodulation and CD4 ⁺ CD73 ⁺ T-cell involvement in HIV-associated immune activation	<ul style="list-style-type: none"> Absolute numbers of CD4⁺CD73⁺ T cells are lower in HIV⁺ individuals compared to HIV⁻ regardless of viral suppression Absolute numbers of CD4⁺CD73⁺ T cells inversely correlate with activated CD4⁺ T cells, activated CD8⁺ T cells, and plasma CRP in HIV⁺ individuals Circulating CD4⁺CD39⁺ T cells frequency did not correlate with the frequency of activated CD4⁺ or CD8⁺ T cells or with the plasma CRP levels CD4⁺ T cells require the presence of both CD4⁺CD39⁺ and CD4⁺CD73⁺ T cells to hydrolyze exogenous ATP to adenosine Exogenous adenosine decreases in the percentage of cytokine-expressing CD4⁺ T cells
Tóth et al [50]	Ex vivo; 95 HIV ⁺ and 27 HIV ⁻ participants	Immunophenotypic analysis of T-cell populations were compared across 5 groups: health controls, ECs, LTNPs, ART patients, and viremic patients	<ul style="list-style-type: none"> In HIV⁺ individuals, %CD73⁺ cells was significantly lower in CD8⁺ T cells and CD4⁺ non-Tregs compared with HIV⁻ individuals Among the HIV⁺ participants, ECs and ART-treated participants showed the highest percentages of CD73⁺ cells Viremic participants displayed the lowest expression levels of CD73⁺, followed by LTNPs

Table 1. Continued

Reference	Study Population	Endpoint	Major Findings
Menkova-Garnier et al [51]	Ex vivo; 16 HIV ⁺ INRs; 16 HIV ⁻ INRs; 18 HIV ⁻ participants	Limiting dilution assays of circulating CD34 ⁺ hematopoietic progenitor cells; frequency of recent thymic emigrants (RTEs), defined as CD31 ^{hi} CD27 ⁺ CCR7 ⁺ CD45RA ⁺ CD4 ⁺ cells; RT-qPCR analysis of FAS, P2X ₇ , and CD73; mRNA levels in purified CD34 cells; transcriptomic analysis of CD3 ⁺ cells in HIV INRs versus INRs.	<ul style="list-style-type: none"> • There was no difference in the frequency of CD34⁺ cells between the 3 groups • The T-cell potential of CD34⁺ cells was significantly lower in INRs compared to HIV- participants and INRs • P2X₇ was more strongly expressed in INRs than in INRs and HIV⁻ participants • CD73 expression was undetectable in all the INRs studied • P2X₇ inhibition with PPAD significantly improved the potential of CD34⁺ cells from INRs to differentiate into T cells
He et al [5]	In vivo; 14 PTM (pathogenic SIV host) and 15 AGM (nonpathogenic SIV host)	Changes of markers related to ADO production (CD39 and CD73) and breakdown (CD26 and ADO deaminase) on T cells from blood, lymph nodes, and intestine after SIV acute infection	<ul style="list-style-type: none"> • Coexpression of CD38 and CD73 was low in circulating CD4⁺ Tregs and CD8⁺ Tregs in both AGMs and PTMs before infection • Coexpression of CD39 and CD73 was highest in intestinal Treg cells and significantly higher in AGMs compared with PTMs • Intestinal levels of adenosine increased during acute SIV infection in AGMs but not in PTMs

Abbreviations: ADA, adenosine deaminase; ADO, adenosine; AGM, African green monkey; ART, antiretroviral therapy; ATP, adenosine triphosphate; cAMP, 3',5'-cyclic monophosphate; c-ART, combination ART; CRP, C-reactive protein; CTL, cytotoxic T lymphocyte; EC, elite controllers; HIV, human immunodeficiency virus; IFN, interferon; IL, interleukin; INR, immunological nonresponder; LTNP, long-term nonprogressor; mRNA, messenger ribonucleic acid; PBMC, peripheral blood mononuclear cell; PTM, pigtail macaques; SIV, simian immunodeficiency virus; RT-qPCR, quantitative reverse-transcription polymerase chain reaction; TNF, tumor necrosis factor; Treg, regulatory T cells.

disruption of the intestinal epithelial integrity and mucosal immune dysfunction [68–71]. Multiple markers of microbial translocation and innate immune activation have been associated with HIV comorbidities, including CVD [72–74]. Extracellular adenosine signaling has been implicated in promoting restoration of the epithelial barrier in inflammatory bowel disease (IBD) [75, 76], and enhanced migration of antigen-presenting cells (APCs) to the gut mucosa can be induced via preferential activation of A₁ receptors [77]. Given these findings, decreased adenosine levels in the gut epithelium could be an important contributing factor to the disrupted epithelial barrier caused by HIV.

NLRP3 inflammasome activation and IL-1β production are factors that contribute to HIV-associated inflammation, and these are strongly implicated in the development of atherosclerosis and hypertension [78, 79]. Activation of the NLRP3 inflammasome and release of IL-1β are strongly tied to purinergic signaling, particularly P2X₇R stimulation [78, 80]. In addition, the inflammasome system may contribute to the loss of CD4⁺ T-cell populations and overall lack of immune recovery seen in PWH [81]. For example, INRs upregulate NLRP3 inflammasome and caspase-1 expression compared with those who experience immune recovery on ART [82]. Antagonism of P2X₇R may therefore help to suppress inflammasome activity and IL-1β release [78]. The exact role of how other components of the adenosine pathway can influence inflammasome activity is still unclear. In non-HIV studies, adenosine has been shown to be necessary for sustained inflammasome activation via the A_{2A} receptor [83]. In animal models of hypertension, proinflammatory cytokines, including IL-1β, downregulate CD39 expression [84]. However, CD39 has recently been implicated to serve a protective role by limiting NLRP3 activation and IL-1β release [85]. Upregulation of CD39⁺/CD73⁺ Tregs are thought to be protective against CVD and unstable plaque rupture [86–89]. Antiretroviral therapy-treated PWH with coronary artery disease have depressed levels of CD39⁺/CD73⁺ Tregs, which may predispose them to atherosclerotic development [90]. How the adenosine pathway affects inflammasome activation in treated HIV will require additional studies to fully understand its role in HIV pathogenesis.

It has recently been shown that purinergic signaling is linked to metabolic disease and dyslipidemia, which contributes to CVD and liver disease among PWH [91, 92]. P2X₇R expression is upregulated in the peripheral blood mononuclear cells from patients with type 2 diabetes and in adipocytes of patients with metabolic syndrome [93, 94]. In addition, P2X₇R expression correlates with low-density lipoprotein cholesterol, which is a major component of metabolic syndrome and elevated cardiovascular risk [93]. Furthermore, extracellular nucleotides have been associated with insulin resistance and dysregulated lipoprotein synthesis [95]. There is also emerging evidence that P2Y₆R activation may contribute to obesity [96].

From our current knowledge of the adenosine pathway, focusing on the role of adenosine as an anti-inflammatory agent could prove beneficial in developing safe and effective interventions in clinical settings. However, although chronic immunoactivation has an integral role in HIV-associated chronic conditions, local immunosuppression plays a key role in cancer risk and viral persistence. Although ART has reduced the mortality of AIDS-related cancers [97], PWH still experience accelerated aging and other risk factors for cancer diagnoses [98]. Long-term immunosuppression is likely the main contributor to non-AIDS-defining cancers [99, 100]. Enhanced adenosine activation in tumor microenvironments generates an immunosuppressant environment that supports tumor growth and evasion from T-cell immune defense [101]. Therefore, modulating adenosine activation in PWH, who are at risk for both solid and liquid malignancies, should be closely monitored.

Viruses that are commonly comorbid with HIV can also upregulate the expression and activity of CD39 and CD73 to facilitate infection [102, 103]. Endothelial cells infected with cytomegalovirus demonstrate an increase in local adenosine production due to the upregulation of both ectonucleotidases [102]. This is thought to facilitate viral entry into target cells by creating a locally immunosuppressive environment. In addition, compared with people with resolved hepatitis B virus (HBV) infection, HBV carriers have higher proportions of Tregs, and increase proportions of circulating CD39⁺ Tregs correlated with serum viral load, thus, suggesting that CD39⁺ Tregs contribute to chronic viral persistence [103].

Finally, modifying the adenosine pathway can have important implications on quality of life among PWH. Adenosine is a well established sleep regulatory substance, and enhanced extracellular concentrations in the brain are associated with sleep deprivation and promotion of sleep, particularly via stimulation of A₁ and A_{2A} receptors [104]. Indeed, caffeine, an adenosine antagonist, is commonly used to thwart sleepiness in the general population [105]. Sleep disturbances are highly prevalent among PWH and, therefore, they may be especially vulnerable to worsened fatigue [106]. Enhancing adenosine activation may facilitate insomnia and nocturnal sleep quality among PWH. However, adenosine activation may promote daytime sleepiness and fatigue as well. Alternatively, changes in sleep patterns have been demonstrated to augment P2X₇ and A_{2A} receptor expression on circulating leukocytes [107]; this suggests that modifying sleep behavior may be a novel nonpharmacologic mechanism to alter purinergic signaling in PWH.

POTENTIAL OF PHARMACOTHERAPIES

There is an understandably heightened interest in developing effective pharmacotherapies to reduce the incidence of non-AIDS-related comorbidities that lead to early mortality in

PWH. Targeting the purinergic signaling pathway to shift the balance away from proinflammatory P2 activation towards anti-inflammatory activation of adenosine receptors is an attractive model to test pharmacotherapeutics. For example, A_{2A} receptors are a critical part in the negative feedback loop of limiting and inhibiting inflammatory responses, providing a rationale to develop A_{2A} receptor-targeted therapeutics to either inhibit or enhance immune responses [38, 108, 109]. There has been emerging literature on pharmacological approaches that target purinergic signaling in various ways to reconstitute the subsequent immune damage of HIV-1 infection [110, 111]. This review focuses on potential therapies to reduce inflammation and promote viral clearance (Figure 2).

Investigation into several potential therapies to curb chronic inflammation in ART-treated PWH and show promising preliminary results [112]. Given that T-cell expression of CD73 is reduced among PWH, attempts have been made to assess whether modulating the adenosine signaling pathway may decrease the persistent chronic inflammatory profile experienced in PWH. In a double-blind, placebo-controlled study, we randomized 40 ART-controlled PWH to 12 weeks of dipyridamole versus placebo, followed by 12 weeks of open-label dipyridamole [113]. Dipyridamole is a nucleoside transport inhibitor and phosphodiesterase 3 inhibitor used clinically in patients with a history of peripheral vascular disease and stroke patients to prevent future thrombotic events. It increases extracellular adenosine by blocking ENTs and preventing transport of adenosine intracellularly down its concentration gradient [114, 115]. Initial data showed that dipyridamole decreased CD8⁺ T-cell activation in the treatment arm versus placebo arm. In pooled analyses, after 12 weeks of dipyridamole, there was a significant decrease in CD4⁺ T-cell activation and a trend toward decreased CD8⁺ T-cell activation in blood [113]. In a substudy, we collected rectosigmoid biopsies from 18 participants to further assess the effect of dipyridamole on mucosal immune cells. Those receiving dipyridamole had (1) a median 70.2% decrease from baseline in the Treg population and (2) an 11.3% increase in CD8⁺ T cells. There were also trends towards decreased CD4⁺ T-cell activation and CD8⁺ T-cell activation [116]. Because the population of Tregs increased in response to heightened inflammation, these data suggest that there is a decrease in gut inflammation that obviates a compensatory Treg response.

Modulating ectonucleotidase activity, particularly CD39 and CD73 activity, is an attractive therapeutic target to reduce proinflammatory extracellular ATP concentrations in favor of anti-inflammatory adenosine. Methotrexate and sulfasalazine, immunosuppressants commonly used in IBD, may be partially effective in treating IBD by enhanced CD73 production of adenosine [117]. Among PWH, although low doses of methotrexate had no effect on systemic inflammatory endothelial markers, there were improvements in brachial artery ultrasound measurements, which may indicate favorable vasculature changes

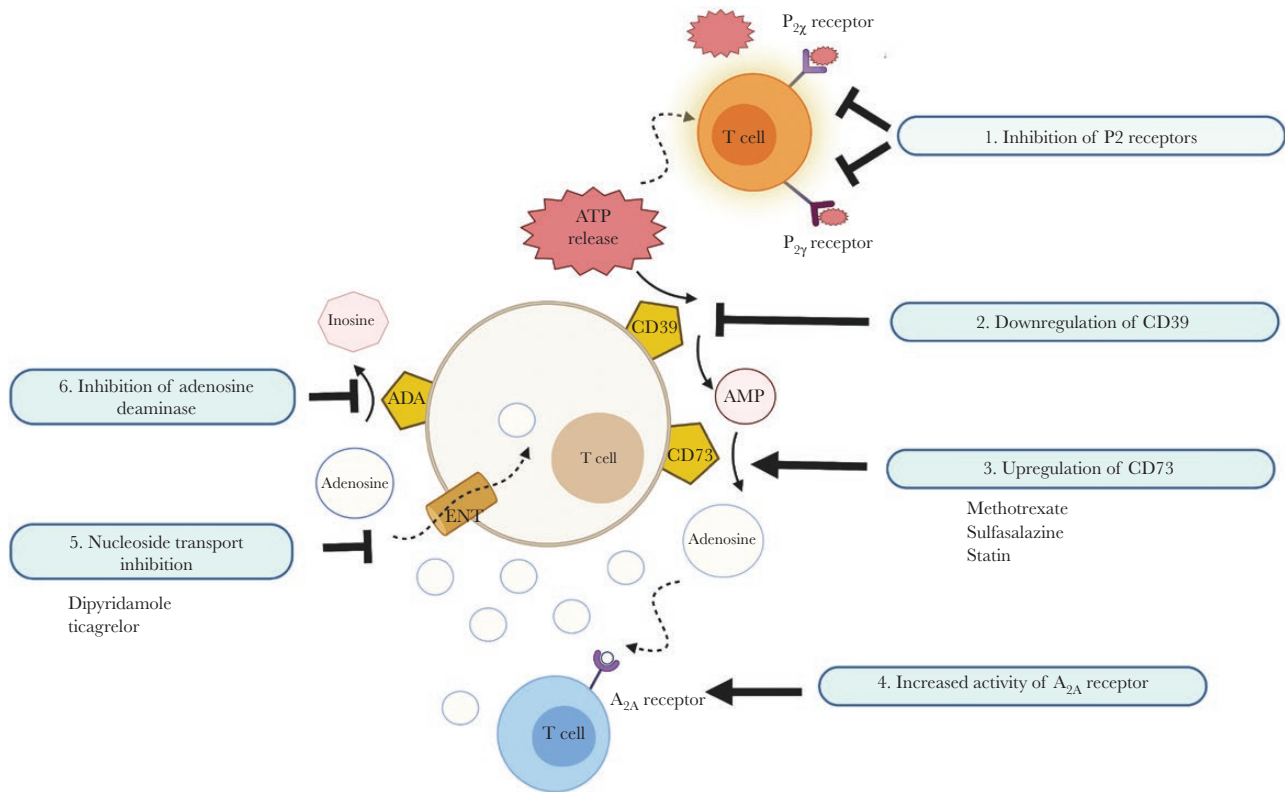


Figure 2. Pharmacologic targets to target adenosine metabolism in human immunodeficiency virus infection. Sites and available therapeutics that decrease purinergic 2 (P2) receptor activity and promote extracellular adenosine and adenosine 2A (A_{2A}) receptor activation are shown. A2A receptor, adenosine 2A receptor; ADA, adenosine deaminase; ADP, adenosine diphosphate; AMP, adenosine monophosphate; ATP, adenosine triphosphate; ENT, equilibrative nucleoside transporter; P2Y receptor, purinergic 2Y receptor.

[118, 119]. Rosuvastatin, typically used to treat high cholesterol and triglyceride levels, can also increase extracellular adenosine formation via upregulation of CD73, and it has shown in vivo protection against inflammation [120–122]. The Randomized Trial to Prevent Vascular Events in HIV (REPRIEVE) is an ongoing, prospective, randomized placebo-controlled clinical trial of a pitavastatin strategy for the primary CVD prevention among PWH [123]. In addition to informing the field on the efficacy of statin strategy among PWH, it includes multiple measurements of plaque stability, immune activation, and inflammation [124]. This trial may provide valuable insights on the role of statins in modifying purinergic metabolism.

Selectively inhibiting $P2Y_2$ receptor expression with small interfering ribonucleic acids reduces the HIV-induced inflammatory response and cell death [125, 126]. Inhibiting $P2X_7$ receptors to restore T-cell differentiation from $CD34^+$ hematopoietic progenitor cells could be a potential strategy in PWH, who experience reduced immune recovery while on ART, to regenerate new T-cell populations [51]. Due to the ubiquitous nature of purinergic receptors, there should be a narrowed focus on refining the characterization of cellular patterns and molecular control of expression of crucial enzymes in the purinergic signaling pathway to minimize unwanted side

effects. It is important to understand receptor regulation to design and improve purinergic receptor strategies to effectively prevent accelerated aging and control systemic inflammation in PWH.

CONCLUSIONS

Although ART is effective in viral suppression and prolonging the development of AIDS, the concern lies in patient susceptibilities to morbidity and early mortality of age-associated diseases due to immunological dysfunction caused by HIV. Adenosine agonists provide immune advantage by inhibiting T-cell effector function, in conjunction with Tregs, to reduce the chronic immune activation and dysfunction seen in PWH. However, although enhancing CD39 and CD73 activity may improve inflammation and immunoactivation-related comorbidities in HIV, modulating ectonucleotidase activity is a double-edged sword. The adenosine/Treg axis can be detrimental by suppressing HIV-specific immune responses. Additional studies are necessary to determine the proper balance between controlling inflammation but still allowing the generation of an effective immune response against the virus. Novel and innovative strategies targeting these 2 contrasting functions of the adenosine pathway can lead to a decreased risk

for non-AIDS-associated chronic disease and, at the same time, target viral persistence.

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References

- Cai CW, Sereti I. Residual immune dysfunction under antiretroviral therapy. *Semin Immunol* **2021**;101471.
- Elvstam O, Marrone G, Medstrand P, et al. All-cause mortality and serious non-AIDS events in adults with low-level human immunodeficiency virus viremia during combination antiretroviral therapy: results from a Swedish Nationwide Observational Study. *Clin Infect Dis* **2021**; 72:2079–86.
- Babu H, Ambikan AT, Gabriel EE, et al. Systemic inflammation and the increased risk of inflamm-aging and age-associated diseases in people living with HIV on long term suppressive antiretroviral therapy. *Front Immunol* **2019**; 10:1965.
- Effros RB, Fletcher CV, Gebo K, et al. Aging and infectious diseases: workshop on HIV infection and aging: what is known and future research directions. *Clin Infect Dis* **2008**; 47:542–53.
- He T, Brocca-Cofano E, Gillespie DG, et al. Critical role for the adenosine pathway in controlling simian immunodeficiency virus-related immune activation and inflammation in gut mucosal tissues. *J Virol* **2015**; 89:9616–30.
- Passos DF, Bernardes VM, da Silva JLG, et al. Adenosine signaling and adenosine deaminase regulation of immune responses: impact on the immunopathogenesis of HIV infection. *Purinergic Signal* **2018**; 14:309–20.
- Milne GR, Palmer TM. Anti-inflammatory and immunosuppressive effects of the A2A adenosine receptor. *ScientificWorldJournal* **2011**; 11:320–39.
- Kumar V, Sharma A. Adenosine: an endogenous modulator of innate immune system with therapeutic potential. *Eur J Pharmacol* **2009**; 616:7–15.
- Rayah A, Kanellopoulos JM, Di Virgilio F. P2 receptors and immunity. *Microbes Infect* **2012**; 14:1254–62.
- Jarvis MF, Khakh BS. ATP-gated P2X cation-channels. *Neuropharmacology* **2009**; 56:208–15.
- Jacobson KA, Delicado EG, Gachet C, et al. Update of P2Y receptor pharmacology: IUPHAR Review 27. *Br J Pharmacol* **2020**; 177:2413–33.
- Zimmermann H. Extracellular metabolism of ATP and other nucleotides. *Naunyn Schmiedebergs Arch Pharmacol* **2000**; 362:299–309.
- Alam MS, Costales MG, Cavanaugh C, Williams K. Extracellular adenosine generation in the regulation of pro-inflammatory responses and pathogen colonization. *Biomolecules* **2015**; 5:775–92.
- Coleman MS, Hutton JJ. Micromethod for quantitation of adenosine deaminase activity in cells from human peripheral blood. *Biochem Med* **1975**; 13:46–55.
- Jacobson KA, Gao ZG. Adenosine receptors as therapeutic targets. *Nat Rev Drug Discov* **2006**; 5:247–64.
- Dhalla AK, Shryock JC, Shreeniwars R, Belardinelli L. Pharmacology and therapeutic applications of A1 adenosine receptor ligands. *Curr Top Med Chem* **2003**; 3:369–85.
- Panther E, Corinti S, Idzko M, et al. Adenosine affects expression of membrane molecules, cytokine and chemokine release, and the T-cell stimulatory capacity of human dendritic cells. *Blood* **2003**; 101:3985–90.
- Fortin A, Harbour D, Fernandes M, et al. Differential expression of adenosine receptors in human neutrophils: up-regulation by specific Th1 cytokines and lipopolysaccharide. *J Leukoc Biol* **2006**; 79:574–85.
- Zhang Y, Palmblad J, Fredholm BB. Biphasic effect of ATP on neutrophil functions mediated by P2U and adenosine A2A receptors. *Biochem Pharmacol* **1996**; 51:957–65.
- Haskó G, Pacher P, Deitch EA, Vizi ES. Shaping of monocyte and macrophage function by adenosine receptors. *Pharmacol Ther* **2007**; 113:264–75.
- Haskó G, Kuhel DG, Chen JF, et al. Adenosine inhibits IL-12 and TNF-[alpha] production via adenosine A2a receptor-dependent and independent mechanisms. *FASEB J* **2000**; 14:2065–74.
- Sipka S, Kovács I, Szántó S, et al. Adenosine inhibits the release of interleukin-1beta in activated human peripheral mononuclear cells. *Cytokine* **2005**; 31:258–63.
- Huang S, Apasov S, Koshiba M, Sitkovsky M. Role of A2a extracellular adenosine receptor-mediated signaling in adenosine-mediated inhibition of T-cell activation and expansion. *Blood* **1997**; 90:1600–10.
- Ohta A, Ohta A, Madasu M, et al. A2A adenosine receptor may allow expansion of T cells lacking effector functions in extracellular adenosine-rich microenvironments. *J Immunol* **2009**; 183:5487–93.
- Zarek PE, Huang CT, Lutz ER, et al. A2A receptor signaling promotes peripheral tolerance by inducing T-cell anergy and the generation of adaptive regulatory T cells. *Blood* **2008**; 111:251–9.
- Lazarowski ER, Boucher RC, Harden TK. Constitutive release of ATP and evidence for major contribution of ecto-nucleotide pyrophosphatase and nucleoside diphosphokinase to extracellular nucleotide concentrations. *J Biol Chem* **2000**; 275:31061–8.
- Antonoli L, Pacher P, Vizi ES, Haskó G. CD39 and CD73 in immunity and inflammation. *Trends Mol Med* **2013**; 19:355–67.
- Bours MJ, Swennen EL, Di Virgilio F, et al. Adenosine 5'-triphosphate and adenosine as endogenous signaling molecules in immunity and inflammation. *Pharmacol Ther* **2006**; 112:358–404.
- Baricordi OR, Melchiorri L, Adinolfi E, et al. Increased proliferation rate of lymphoid cells transfected with the P2X(7) ATP receptor. *J Biol Chem* **1999**; 274:33206–8.
- Kaneider NC, Mosheimer B, Reinisch N, et al. Inhibition of thrombin-induced signaling by resveratrol and quercetin: effects on adenosine nucleotide metabolism in endothelial cells and platelet-neutrophil interactions. *Thromb Res* **2004**; 114:185–94.
- Aga M, Johnson CJ, Hart AP, et al. Modulation of monocyte signaling and pore formation in response to agonists of the nucleotide receptor P2X(7). *J Leukoc Biol* **2002**; 72:222–32.
- Kaufmann A, Musset B, Limberg SH, et al. "Host tissue damage" signal ATP promotes non-directional migration and negatively regulates toll-like receptor signaling in human monocytes. *J Biol Chem* **2005**; 280:32459–67.
- Aga M, Watters JJ, Pfeiffer ZA, et al. Evidence for nucleotide receptor modulation of cross talk between MAP kinase and NF-kappa B signaling pathways in murine RAW 264.7 macrophages. *Am J Physiol Cell Physiol* **2004**; 286:C923–30.
- Ichinose M. Modulation of phagocytosis by P2-purinergic receptors in mouse peritoneal macrophages. *Jpn J Physiol* **1995**; 45:707–21.
- Deussen A, Stappert M, Schäfer S, Kelm M. Quantification of extracellular and intracellular adenosine production: understanding the transmembranous concentration gradient. *Circulation* **1999**; 99:2041–7.
- Haskó G, Cronstein B. Regulation of inflammation by adenosine. *Front Immunol* **2013**; 4:85.
- Drygiannakis I, Ernst PB, Lowe D, Glomski IJ. Immunological alterations mediated by adenosine during host-microbial interactions. *Immunol Res* **2011**; 50:69–77.
- Ohta A, Sitkovsky M. Role of G-protein-coupled adenosine receptors in downregulation of inflammation and protection from tissue damage. *Nature* **2001**; 414:916–20.
- Zsuga J, Erdei T, Szabó K, et al. Methodical challenges and a possible resolution in the assessment of receptor reserve for adenosine, an agonist with short half-life. *Molecules* **2017**; 22:839.
- Blanco J, Valenzuela A, Herrera C, et al. The HIV-1 gp120 inhibits the binding of adenosine deaminase to CD26 by a mechanism modulated by CD4 and CXCR4 expression. *FEBS Lett* **2000**; 477:123–8.
- Fotheringham J, Mayne M, Holden C, et al. Adenosine receptors control HIV-1 Tat-induced inflammatory responses through protein phosphatase. *Virology* **2004**; 327:186–95.
- Pingle SC, Jajoo S, Mukherjee D, et al. Activation of the adenosine A1 receptor inhibits HIV-1 tat-induced apoptosis by reducing nuclear factor-kappaB activation and inducible nitric-oxide synthase. *Mol Pharmacol* **2007**; 72:856–67.
- By Y, Durand-Gorde JM, Condo J, et al. Monoclonal antibody-assisted stimulation of adenosine A2A receptors induces simultaneous downregulation of CXCR4 and CCR5 on CD4+ T-cells. *Hum Immunol* **2010**; 71:1073–6.
- Moreno-Fernandez ME, Rueda CM, Rusie LK, Chougnnet CA. Regulatory T cells control HIV replication in activated T cells through a cAMP-dependent mechanism. *Blood* **2011**; 117:5372–80.
- Martinez-Navio JM, Climent N, Pacheco R, et al. Immunological dysfunction in HIV-1-infected individuals caused by impairment of

- adenosine deaminase-induced costimulation of T-cell activation. *Immunology* **2009**; 128:393–404.
46. Climent N, Martinez-Navio JM, Gil C, et al. Adenosine deaminase enhances T-cell response elicited by dendritic cells loaded with inactivated HIV. *Immunol Cell Biol* **2009**; 87:634–9.
 47. Parish ST, Kim S, Sekhon RK, et al. Adenosine deaminase modulation of telomerase activity and replicative senescence in human CD8 T lymphocytes. *J Immunol* **2010**; 184:2847–54.
 48. Nikolova M, Carriere M, Jenabian MA, et al. CD39/adenosine pathway is involved in AIDS progression. *PLoS Pathog* **2011**; 7:e1002110.
 49. Schuler PJ, Macatangay BJ, Saze Z, et al. CD4+CD73+ T cells are associated with lower T-cell activation and C reactive protein levels and are depleted in HIV-1 infection regardless of viral suppression. *AIDS* **2013**; 27:1545–55.
 50. Tóth I, Le AQ, Hartjen P, et al. Decreased frequency of CD73+CD8+ T cells of HIV-infected patients correlates with immune activation and T cell exhaustion. *J Leukoc Biol* **2013**; 94:551–61.
 51. Menkova-Garnier I, Hocini H, Foucat E, et al. P2X7 Receptor inhibition improves CD34 T-cell differentiation in HIV-infected immunological nonresponders on c-ART. *PLoS Pathog* **2016**; 12:e1005571.
 52. Swartz TH, Esposito AM, Durham ND, et al. P2X-selective purinergic antagonists are strong inhibitors of HIV-1 fusion during both cell-to-cell and cell-free infection. *J Virol* **2014**; 88:11504–15.
 53. Graziano F, Desdouts M, Garzetti L, et al. Extracellular ATP induces the rapid release of HIV-1 from virus containing compartments of human macrophages. *Proc Natl Acad Sci U S A* **2015**; 112:E3265–73.
 54. Deeks SG. HIV infection, inflammation, immunosenescence, and aging. *Annu Rev Med* **2011**; 62:141–55.
 55. Deeks SG, Verdin E, McCune JM. Immunosenescence and HIV. *Curr Opin Immunol* **2012**; 24:501–6.
 56. Serrano-Villar S, Sainz T, Lee SA, et al. HIV-infected individuals with low CD4/CD8 ratio despite effective antiretroviral therapy exhibit altered T cell subsets, heightened CD8+ T cell activation, and increased risk of non-AIDS morbidity and mortality. *PLoS Pathog* **2014**; 10:e1004078.
 57. Gupta PK, Godec J, Wolski D, et al. CD39 expression identifies terminally exhausted CD8+ T cells. *PLoS Pathog* **2015**; 11:e1005177.
 58. Dierks P, Wroblewski R, Eberhard JM, et al. Brief report: increased frequency of CD39+ CD56bright natural killer cells in HIV-1 infection correlates with immune activation and disease progression. *J Acquir Immune Defic Syndr* **2017**; 74:467–72.
 59. Song JW, Huang HH, Zhang C, et al. Expression of CD39 is correlated with HIV DNA levels in naïve tregs in chronically infected ART naïve patients. *Front Immunol* **2019**; 10:2465.
 60. Rueda CM, Jackson CM, Choungnet CA. Regulatory T-cell-mediated suppression of conventional T-cells and dendritic cells by different cAMP intracellular pathways. *Front Immunol* **2016**; 7:216.
 61. Deaglio S, Dwyer KM, Gao W, et al. Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. *J Exp Med* **2007**; 204:1257–65.
 62. Ohta A, Kini R, Ohta A, et al. The development and immunosuppressive functions of CD4(+) CD25(+) FoxP3(+) regulatory T cells are under influence of the adenosine-A2A adenosine receptor pathway. *Front Immunol* **2012**; 3:190.
 63. Kinter A, McNally J, Riggan L, et al. Suppression of HIV-specific T cell activity by lymph node CD25+ regulatory T cells from HIV-infected individuals. *Proc Natl Acad Sci U S A* **2007**; 104:3390–5.
 64. Weiss L, Donkova-Petrini V, Caccavelli L, et al. Human immunodeficiency virus-driven expansion of CD4+CD25+ regulatory T cells, which suppress HIV-specific CD4 T-cell responses in HIV-infected patients. *Blood* **2004**; 104:3249–56.
 65. Jenabian MA, Seddiki N, Yatim A, et al. Regulatory T cells negatively affect IL-2 production of effector T cells through CD39/adenosine pathway in HIV infection. *PLoS Pathog* **2013**; 9:e1003319.
 66. Kinter AL, Horak R, Sion M, et al. CD25+ regulatory T cells isolated from HIV-infected individuals suppress the cytolytic and nonlytic antiviral activity of HIV-specific CD8+ T cells in vitro. *AIDS Res Hum Retroviruses* **2007**; 23:438–50.
 67. Macatangay BJ, Szajnik ME, Whiteside TL, et al. Regulatory T cell suppression of Gag-specific CD8 T cell polyfunctional response after therapeutic vaccination of HIV-1-infected patients on ART. *PLoS One* **2010**; 5:e9852.
 68. Baroncelli S, Galluzzo CM, Pirillo MF, et al. Microbial translocation is associated with residual viral replication in HAART-treated HIV+ subjects with <50copies/ml HIV-1 RNA. *J Clin Virol* **2009**; 46:367–70.
 69. Brenchley JM, Douek DC. Microbial translocation across the GI tract. *Annu Rev Immunol* **2012**; 30:149–73.
 70. Brenchley JM, Price DA, Schacker TW, et al. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Nat Med* **2006**; 12:1365–71.
 71. Klatt NR, Funderburg NT, Brenchley JM. Microbial translocation, immune activation, and HIV disease. *Trends Microbiol* **2013**; 21:6–13.
 72. Alcaide ML, Parmigiani A, Pallikkuth S, et al. Immune activation in HIV-infected aging women on antiretrovirals—implications for age-associated comorbidities: a cross-sectional pilot study. *PLoS One* **2013**; 8:e63804.
 73. Burdo TH, Lo J, Abbasa S, et al. Soluble CD163, a novel marker of activated macrophages, is elevated and associated with noncalcified coronary plaque in HIV-infected patients. *J Infect Dis* **2011**; 204:1227–36.
 74. Sandler NG, Wand H, Roque A, et al; INSIGHT SMART Study Group. Plasma levels of soluble CD14 independently predict mortality in HIV infection. *J Infect Dis* **2011**; 203:780–90.
 75. Aherne CM, Collins CB, Rapp CR, et al. Coordination of ENT2-dependent adenosine transport and signaling dampens mucosal inflammation. *JCI Insight* **2018**; 3:e121521.
 76. Aherne CM, Saeedi B, Collins CB, et al. Epithelial-specific A2B adenosine receptor signaling protects the colonic epithelial barrier during acute colitis. *Mucosal Immunol* **2015**; 8:1324–38.
 77. Antonioli L, Fornai M, Colucci R, et al. Regulation of enteric functions by adenosine: pathophysiological and pharmacological implications. *Pharmacol Ther* **2008**; 120:233–53.
 78. Ferrari D, la Sala A, Milani D, et al. Purinergic signaling in controlling macrophage and T cell functions during atherosclerosis development. *Front Immunol* **2020**; 11:617804.
 79. Hsue PY, Li D, Ma Y, et al. IL-1 β inhibition reduces atherosclerotic inflammation in HIV infection. *J Am Coll Cardiol* **2018**; 72:2809–11.
 80. Ferrari D, Pizzirani C, Adinolfi E, et al. The P2X7 receptor: a key player in IL-1 processing and release. *J Immunol* **2006**; 176:3877–83.
 81. Zhang C, Song JW, Huang HH, et al. NLRP3 inflammasome induces CD4+ T cell loss in chronically HIV-1-infected patients. *J Clin Invest* **2021**; 131.
 82. Bandera A, Masetti M, Fabbiani M, et al. The NLRP3 inflammasome is upregulated in HIV-infected antiretroviral therapy-treated individuals with defective immune recovery. *Front Immunol* **2018**; 9:214.
 83. Ouyang X, Ghani A, Malik A, et al. Adenosine is required for sustained inflammasome activation via the A_{2A} receptor and the HIF-1 α pathway. *Nat Commun* **2013**; 4:2909.
 84. Roy C, Tabiasco J, Caillon A, et al. Loss of vascular expression of nucleoside triphosphate diphosphohydrolase-1/CD39 in hypertension. *Purinergic Signal* **2018**; 14:73–82.
 85. Yadav V, Chi L, Zhao R, et al. Ectonucleotidase tri(di)phosphohydrolase-1 (ENTPD-1) disrupts inflammasome/interleukin 1 β -driven venous thrombosis. *J Clin Invest* **2019**; 129:2872–7.
 86. Covarrubias R, Chepurko E, Reynolds A, et al. Role of the CD39/CD73 purinergic pathway in modulating arterial thrombosis in mice. *Arterioscler Thromb Vasc Biol* **2016**; 36:1809–20.
 87. Koszalka P, Ozüyan B, Huo Y, et al. Targeted disruption of cd73/ecto-5'-nucleotidase alters thromboregulation and augments vascular inflammatory response. *Circ Res* **2004**; 95:814–21.
 88. Rohm I, Atiskova Y, Drobniak S, et al. Decreased regulatory T cells in vulnerable atherosclerotic lesions: imbalance between pro- and anti-inflammatory cells in atherosclerosis. *Mediators Inflamm* **2015**; 2015:364710.
 89. Zerneck A, Bidzhekov K, Ozüyan B, et al. CD73/ecto-5'-nucleotidase protects against vascular inflammation and neointima formation. *Circulation* **2006**; 113:2120–7.
 90. Rothan C, Yero A, Shi T, et al; Canadian HIV and Aging Cohort Study. Antiretroviral therapy-treated HIV-infected adults with coronary artery disease are characterized by a distinctive regulatory T-cell signature. *AIDS* **2021**; 35:1003–14.
 91. Sparks DL, Chatterjee C. Purinergic signaling, dyslipidemia and inflammatory disease. *Cell Physiol Biochem* **2012**; 30:1333–9.
 92. Teixeira GP, Faria RX. Influence of purinergic signaling on glucose transporters: A possible mechanism against insulin resistance? *Eur J Pharmacol* **2021**; 892:173743.
 93. García-Hernández MH, Portales-Cervantes L, Cortez-Espinosa N, et al. Expression and function of P2X(7) receptor and CD39/Entpd1 in patients with type 2 diabetes and their association with biochemical parameters. *Cell Immunol* **2011**; 269:135–43.
 94. Madec S, Rossi C, Chiarugi M, et al. Adipocyte P2X7 receptors expression: a role in modulating inflammatory response in subjects with metabolic syndrome? *Atherosclerosis* **2011**; 219:552–8.
 95. Chatterjee C, Sparks DL. Extracellular nucleotides inhibit insulin receptor signaling, stimulate autophagy and control lipoprotein secretion. *PLoS One* **2012**; 7:e36916.
 96. Zhang Y, Ecelbarger CM, Lesniewski LA, et al. P2Y2 receptor promotes high-fat diet-induced obesity. *Front Endocrinol (Lausanne)* **2020**; 11:341.
 97. Park LS, Tate JP, Sigel K, et al. Time trends in cancer incidence in persons living with HIV/AIDS in the antiretroviral therapy era: 1997–2012. *AIDS* **2016**; 30:1795–806.

98. Shiels MS, Engels EA. Evolving epidemiology of HIV-associated malignancies. *Curr Opin HIV AIDS* **2017**; 12:6–11.
99. Hernández-Ramírez RU, Shiels MS, Dubrow R, Engels EA. Cancer risk in HIV-infected people in the USA from 1996 to 2012: a population-based, registry-linkage study. *Lancet HIV* **2017**; 4:e495–504.
100. Wang CC, Silverberg MJ, Abrams DI. Non-AIDS-defining malignancies in the HIV-infected population. *Curr Infect Dis Rep* **2014**; 16:406.
101. Soleimani A, Bahreyni A, Roshan MK, et al. Therapeutic potency of pharmacological adenosine receptors agonist/antagonist on cancer cell apoptosis in tumor microenvironment, current status, and perspectives. *J Cell Physiol* **2019**; 234:2329–36.
102. Kas-Deelen AM, Bakker WW, Olinga P, et al. Cytomegalovirus infection increases the expression and activity of ecto-ATPase (CD39) and ecto-5'nucleotidase (CD73) on endothelial cells. *FEBS Lett* **2001**; 491:21–5.
103. Tang Y, Jiang L, Zheng Y, et al. Expression of CD39 on FoxP3+ T regulatory cells correlates with progression of HBV infection. *BMC Immunol* **2012**; 13:17.
104. Huang ZL, Zhang Z, Qu WM. Roles of adenosine and its receptors in sleep-wake regulation. *Int Rev Neurobiol* **2014**; 119:349–71.
105. Faudone G, Arifi S, Merk D. The medicinal chemistry of caffeine. *J Med Chem* **2021**; 64:7156–78.
106. Wu J, Wu H, Lu C, et al. Self-reported sleep disturbances in HIV-infected people: a meta-analysis of prevalence and moderators. *Sleep Med* **2015**; 16:901–7.
107. Chennaoui M, Arnal PJ, Drogou C, et al. Leukocyte expression of type 1 and type 2 purinergic receptors and pro-inflammatory cytokines during total sleep deprivation and/or sleep extension in healthy subjects. *Front Neurosci* **2017**; 11:240.
108. de Lera Ruiz M, Lim YH, Zheng J. Adenosine A2A receptor as a drug discovery target. *J Med Chem* **2014**; 57:3623–50.
109. Sitkovsky MV, Lukashev D, Apasov S, et al. Physiological control of immune response and inflammatory tissue damage by hypoxia-inducible factors and adenosine A2A receptors. *Annu Rev Immunol* **2004**; 22:657–82.
110. Freeman TL, Swartz TH. Purinergic receptors: elucidating the role of these immune mediators in HIV-1 fusion. *Viruses* **2020**; 12:290.
111. Swartz TH, Dubyak GR, Chen BK. Purinergic receptors: key mediators of HIV-1 infection and inflammation. *Front Immunol* **2015**; 6:585.
112. Taiwo B, Barcena L, Tressler R. Understanding and controlling chronic immune activation in the HIV-infected patients suppressed on combination antiretroviral therapy. *Curr HIV/AIDS Rep* **2013**; 10:21–32.
113. Macatangay BJC, Jackson EK, Abebe KZ, et al. A randomized, placebo-controlled, pilot clinical trial of dipyridamole to decrease HIV-associated chronic inflammation. *J Infect Dis* **2019**; 221:1598–1606.
114. Galabov AS, Mastikova M. Dipyridamole is an interferon inducer. *Acta Virol* **1982**; 26:137–47.
115. Newby AC. How does dipyridamole elevate extracellular adenosine concentration? Predictions from a three-compartment model of adenosine formation and inactivation. *Biochem J* **1986**; 237:845–51.
116. Mallarino-Haeger C, Abebe KZ, Jackson EK, et al. Dipyridamole decreases gut mucosal regulatory T cell frequencies among people with HIV on antiretroviral therapy. *JAIDS* **2020**; 85:665–9.
117. Eltzschig HK, Sitkovsky MV, Robson SC. Purinergic signaling during inflammation. *N Engl J Med* **2012**; 367:2322–33.
118. Hsue PY, Ribaldo HJ, Deeks SG, et al. Safety and impact of low-dose methotrexate on endothelial function and inflammation in individuals with treated human immunodeficiency virus: AIDS clinical trials group study A5314. *Clin Infect Dis* **2019**; 68:1877–86.
119. Stein JH, Yeh E, Weber JM, et al. Brachial artery echogenicity and grayscale texture changes in HIV-infected individuals receiving low-dose methotrexate. *Arterioscler Thromb Vasc Biol* **2018**; 38:2870–8.
120. Calza L, Vanino E, Salvadori C, et al. Tenofovir/emtricitabine/efavirenz plus rosuvastatin decrease serum levels of inflammatory markers more than antiretroviral drugs alone in antiretroviral therapy-naive HIV-infected patients. *HIV Clin Trials* **2014**; 15:1–13.
121. Eckard AR, Jiang Y, Debanne SM, et al. Effect of 24 weeks of statin therapy on systemic and vascular inflammation in HIV-infected subjects receiving antiretroviral therapy. *J Infect Dis* **2014**; 209:1156–64.
122. Funderburg NT, Jiang Y, Debanne SM, et al. Rosuvastatin treatment reduces markers of monocyte activation in HIV-infected subjects on antiretroviral therapy. *Clin Infect Dis* **2014**; 58:588–95.
123. Grinspoon SK, Fitch KV, Overton ET, et al; REPRIEVE Investigators. Rationale and design of the randomized trial to prevent vascular events in HIV (REPRIEVE). *Am Heart J* **2019**; 212:23–35.
124. Hoffmann U, Lu MT, Olalere D, et al; REPRIEVE Investigators. Rationale and design of the mechanistic substudy of the randomized trial to prevent vascular events in HIV (REPRIEVE): effects of pitavastatin on coronary artery disease and inflammatory biomarkers. *Am Heart J* **2019**; 212:1–12.
125. Hübner W, Chen P, Del Portillo A, et al. Sequence of human immunodeficiency virus type 1 (HIV-1) Gag localization and oligomerization monitored with live confocal imaging of a replication-competent, fluorescently tagged HIV-1. *J Virol* **2007**; 81:12596–607.
126. Jolly C, Kashefi K, Hollinshead M, Sattentau QJ. HIV-1 cell to cell transfer across an Env-induced, actin-dependent synapse. *J Exp Med* **2004**; 199:283–93.