



Short Communication

Lack of evidence for intermolecular epistatic interactions between adiponectin and resistin gene polymorphisms in Malaysian male subjects

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Abstract

Epistasis (gene-gene interaction) is a ubiquitous component of the genetic architecture of complex traits such as susceptibility to common human diseases. Given the strong negative correlation between circulating adiponectin and resistin levels, the potential intermolecular epistatic interactions between *ADIPOQ* (SNP+45T > G, SNP+276G > T, SNP+639T > C and SNP+1212A > G) and *RETN* (SNP-420C > G and SNP+299G > A) gene polymorphisms in the genetic risk underlying type 2 diabetes (T2DM) and metabolic syndrome (MS) were assessed. The potential mutual influence of the *ADIPOQ* and *RETN* genes on their adipokine levels was also examined. The rare homozygous genotype (risk alleles) of SNP-420C > G at the *RETN* locus tended to be co-inherited together with the common homozygous genotypes (protective alleles) of SNP+639T > C and SNP+1212A > G at the *ADIPOQ* locus. Despite the close structural relationship between the *ADIPOQ* and *RETN* genes, there was no evidence of an intermolecular epistatic interaction between these genes. There was also no reciprocal effect of the *ADIPOQ* and *RETN* genes on their adipokine levels, *i.e.*, *ADIPOQ* did not affect resistin levels nor did *RETN* affect adiponectin levels. The possible influence of the *ADIPOQ* gene on *RETN* expression warrants further investigation.

Key words: adipokine, *ADIPOQ*, epistasis, interaction, *RETN*.

Received: May 19, 2011; Accepted: August 4, 2011.

Adiponectin and resistin are important hormones in the regulation of insulin sensitivity (insulin secretion and signaling pathway), energy homeostasis (glycogen synthesis and glycogenolysis), glucose metabolism (gluconeogenesis, glucose uptake and use) and lipid metabolism (fatty acid synthesis and oxidation) (Lau and Muniandy, 2011a). We have recently shown that the mutual regulation of adiponectin and resistin is an important determinant of metabolic disturbances, cardiometabolic function and global metabolic status (Lau and Muniandy, 2011a). Moreover, the relative proportion of adiponectin-to-resistin levels reflect metabolic homeostasis and metabolic disorders much better than adiponectin and resistin levels alone, with a significant inverse correlation between the levels of these two hormones (Lau and Muniandy, 2011a). The similar domain architecture of these adipokines, despite their diametrically opposed physiological effects, suggests a common regulatory mechanism in metabolic homeostasis (Patel *et al.*, 2004). Single nucleotide polymorphisms (SNPs) have been identified in the *ADIPOQ* (SNP+45T > G, SNP+276G > T, SNP+639T > C and SNP+1212A > G) and *RETN* (SNP-420C > G and SNP+299G > A) genes. These SNPs of the *ADIPOQ* (Zhao and Zhao, 2011) and *RETN* (Wen *et al.*,

2010) genes are the most widely studied for their etiological links to the risk of type 2 diabetes mellitus (T2DM) and metabolic syndrome (MS). In general, T2DM and MS do not follow a Mendelian pattern of inheritance.

There is increasing evidence that gene-gene interactions (epistasis) play a far more important role in an individual's susceptibility to these complex diseases than single polymorphisms on their own since one gene can enhance or hinder the expression of another gene (Moore, 2003). Moreover, there is a growing awareness that the failure to replicate single-locus association studies for common diseases may reflect an underlying genetic architecture in which interactions between genes are the norm rather than the exception (Lehner, 2011). Epistasis occurs when the developmental or physiological effects of a genetic difference at one locus depend on the genotype at another locus, perhaps on a different chromosome (Desai *et al.*, 2007). This dependence is often a consequence of an underlying functional relationship between the two genes (Lehner, 2011). Since epistasis is a ubiquitous component of the genetic architecture of complex diseases, analysis of the combined effects of the *ADIPOQ* and *RETN* genes may provide more information about the risk of T2DM and MS than the analysis of a single gene.

Given the strong negative correlation between circulating adiponectin and resistin levels, in this work we examined the potential intermolecular epistatic interactions

between *ADIPOQ* and *RETN* gene polymorphisms in the genetic risk underlying T2DM and MS. The potential mutual influence of the *ADIPOQ* and *RETN* genes on their adipokine levels was also examined. The results of this study provide a better understanding of the intermolecular interactions between *ADIPOQ* and *RETN* gene polymorphisms that underlie the pathogenesis of T2DM and MS. Such an analysis of genetic interactions can reveal implicit structural and functional relationships between the *ADIPOQ* and *RETN* genes and may provide useful insights for developing therapeutic approaches to treat T2DM and MS.

All subjects were native Malaysian males 40-70 years old from three major ethnic groups (Malay, Chinese and Indian). This study was approved by the University of Malaya Medical Centre (UMMC) Ethics Committee (Ethical Approval Letter no. 612.17). Informed consent was obtained from each participant after thorough explanation of the study. Each subject received a detailed questionnaire about his personal and family history of disease and demographic data. Eight hundred and nine subjects were enrolled in a case-control study in which there were 208 controls, 174 in-

dividuals with MS but no T2DM, 171 individuals with T2DM but no MS and 256 individuals with T2DM and MS. MS was defined according to the International Diabetes Federation 2005 diagnostic criteria (Alberti *et al.*, 2005). The sampling method used and the characteristics of this study have been described elsewhere (Lau and Muniandy, 2011a,b).

The subjects were screened for metabolic parameters that included serum fasting levels of adiponectin, resistin, insulin, total cholesterol, LDL cholesterol, HDL cholesterol, triglycerides, plasma glucose and whole blood HbA1C. The anthropometric parameters that were measured or calculated included blood pressure, body mass index (BMI), waist circumference and waist-to-hip ratio (WHR). β -Cell function (HOMA% β index), insulin sensitivity (HOMA%S index) and insulin resistance (HOMA-IR index) were calculated with HOMA calculator software version 2.2.2 (Diabetes Trials Unit, Oxford University). The clinical characteristics and demographic parameters of the subjects have been reported elsewhere (Lau and Muniandy, 2011a).

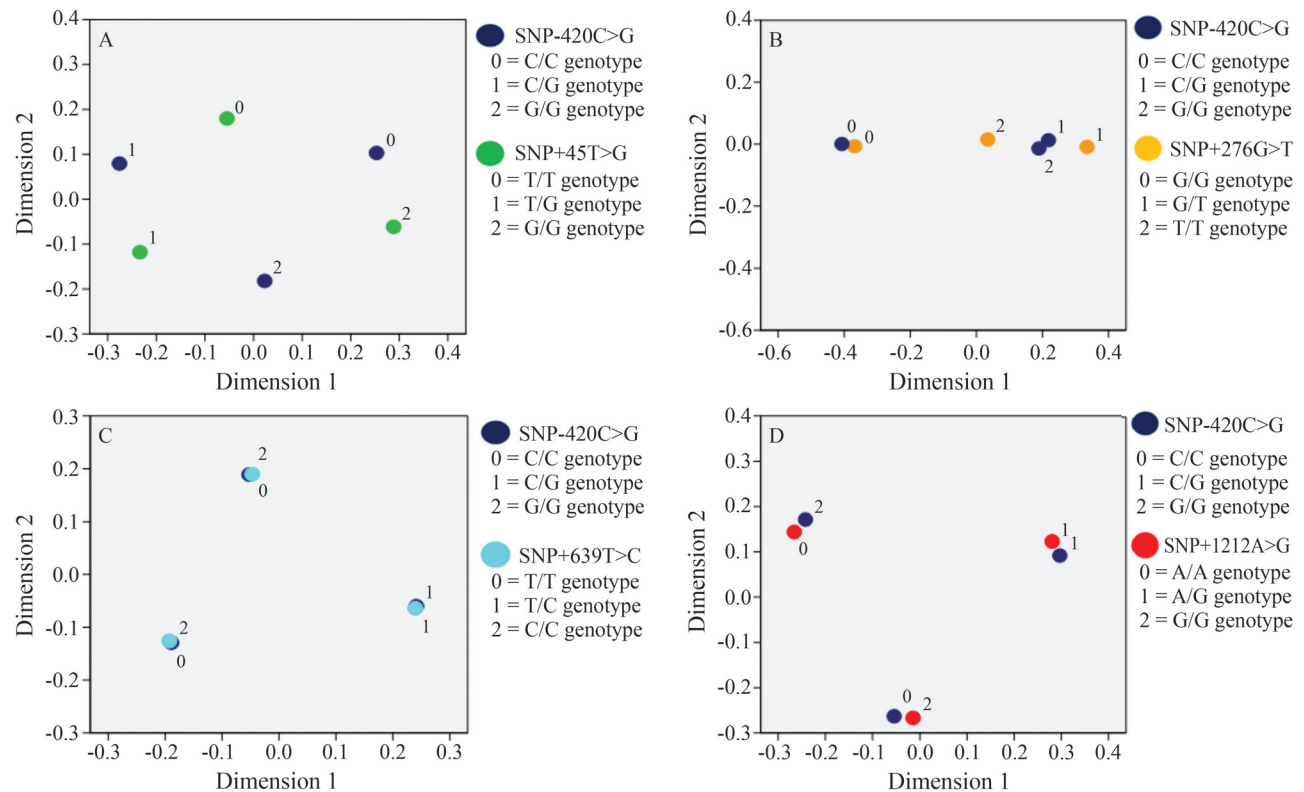


Figure 1 - Dimension reduction correspondence analysis of SNP-420C > G showing biplot results for (A) SNP-420C > G and SNP+45T > G, (B) SNP-420C > G and SNP+276G > T, (C) SNP-420C > G and SNP+639T > C and (D) SNP-420C > G and SNP+1212A > G. The total sample size was 809. The model settings included two dimensions in solution, Euclidean distance measurements, default standardization and symmetrical normalization methods. A biplot illustrates the underlying structural relationships between two SNPs. Projecting the genotypes for one SNP on the vector from the origin to a genotype for the other SNP describes the structural relationship between the SNPs. The distance between genotypes in a plot reflects the strength of the relationship between the genotypes, with associated genotypes being plotted close to each other. Genotypes that are closer to each other are more alike (co-inherited) than those that are farther apart. Note: *RETN* (SNP-420C > G); *ADIPOQ* (SNP+45T > G, SNP+276G > T, SNP+639T > C, SNP+1212A > G).

Genomic DNA was extracted from whole blood using a Wizard Genomic DNA purification kit (Promega Corporation, Madison, WI, USA). SNPs were genotyped using Custom TaqMan[®] SNP Genotyping assay kits (Applied Biosystems, Foster City, CA, USA) according to the manufacturer's standard protocol. Allelic discrimination analysis was done with a StepOnePlus real-time PCR system version 2.0 (Applied Biosystems). All of the SNPs passed the pre-defined threshold for genotyping call rate. Only subjects with a high genotyping call rate (quality value $\geq 98\%$) were included in the study. Primers and PCR conditions were designed to amplify the DNA region of each SNP marker. Thirty samples were randomly selected for forward and reverse DNA sequencing (AITBiotech Pte Ltd., Singapore) to verify the genotyping results for each SNP marker. Details of the genotyping and DNA sequencing procedures are described elsewhere (Lau and Munianidy, 2011b).

Dimension reduction correspondence analysis to examine the structural relationships between two SNPs in low-dimensional space was done using the PASW Statistics 18 Program (SPSS Inc, Chicago, IL, USA). Quantitative traits analysis for alleles was done using the HAPSTAT program, version 3.0 (Chapel Hill, NC, USA).

In the additive model, an empirical p value was generated after Bonferroni correction with adjusted known confounding factors. The potential nonlinear interactions between *ADIPOQ* and *RETN* gene polymorphisms were analyzed with the Generalized Multifactor Dimensionality Reduction (GMDR) Beta program, version 0.7 (University of Virginia, Charlottesville, VA, USA). GMDR analysis included a combined cross-validation/permutation testing procedure that prevented model over-fitting and minimized false-positive results generated by multiple comparisons. Ethnicity was stratified before analysis. All p values were two-tailed, and p values < 0.05 were considered statistically significant.

SNP-420C $>$ G of the *RETN* gene showed a strong structural relationship with SNP+639T $>$ C and SNP+1212A $>$ G of the *ADIPOQ* gene (Figure 1). The heterozygous genotype of SNP-420C $>$ G tended to be co-inherited together with the heterozygous genotypes of SNP+639T $>$ C (Figure 1C) and SNP+1212A $>$ G (Figure 1D). SNP+299G $>$ A of the *RETN* gene shared a strong structural relationship with SNP+639T $>$ C of the *ADIPOQ* gene (Figure 2). The common homozygous genotypes of SNP+299G $>$ A and SNP+639T $>$ C tended to be co-inherited (Figure 2C). Interestingly, the rare homozygous

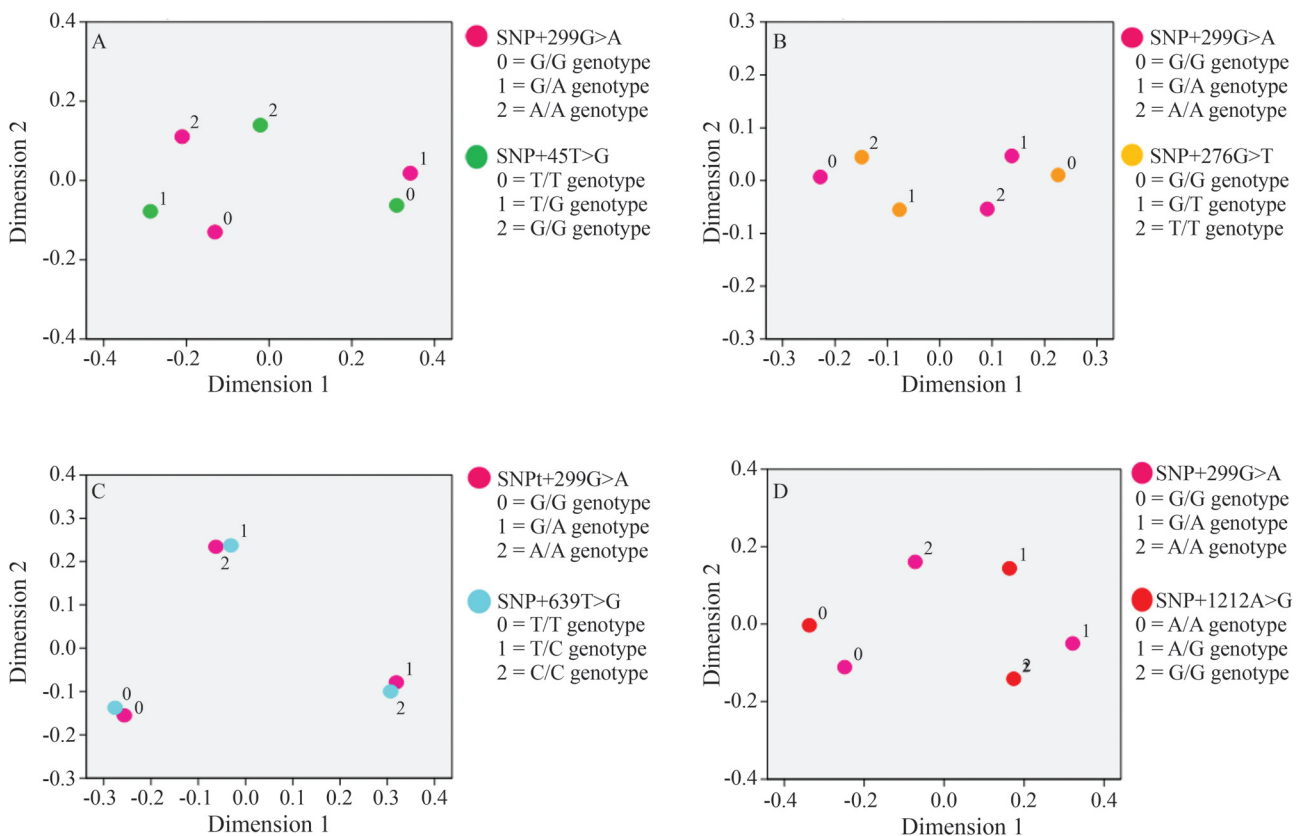


Figure 2 - Dimension reduction correspondence analysis of SNP+299G $>$ A showing biplot results for (A) SNP+299G $>$ A and SNP+45T $>$ G, (B) SNP+299G $>$ A and SNP+276G $>$ T, (C) SNP+299G $>$ A and SNP+639T $>$ C and (D) SNP+299G $>$ A and SNP+1212A $>$ G. The total sample size was 809. See the legend for Figure 1 for details of the model settings and the interpretation of biplots. Note: *RETN* (SNP+299G $>$ A); *ADIPOQ* (SNP+45T $>$ G, SNP+276G $>$ T, SNP+639T $>$ C, SNP+1212A $>$ G).

genotype of SNP-420C > G was generally co-inherited with the common homozygous genotypes of SNP+639T < C (Figure 1C) and SNP+1212A < G (Figure 1D). Furthermore, the rare homozygous genotypes of these SNPs at the *ADIPOQ* and *RETN* loci were unlikely to be co-inherited (Figures 1 and 2), which suggested a possible antagonistic relationship between the *ADIPOQ* and *RETN* genes.

Despite the above relationships, there were no interactions between *ADIPOQ* (SNP+45T > G, SNP+276G > T, SNP+639T > C and SNP+1212A > G) and *RETN* (SNP-420C > G and SNP+299G > A) gene polymorphisms and their adipokine levels (Table 1). GMDR analysis showed that none of these SNP combinations was associated with the risk of T2DM and MS among Malaysian Malay, Chinese and Indian men (Table 2). Despite the close structural relationship between the *ADIPOQ* and *RETN* genes (Figures 1 and 2), the functional relationship between these genes was unclear (Tables 1 and 2). The extent to which the structural relationship observed here was related to gene transcriptional activity (adipokine levels) and/or metabolic disturbances (risk of T2DM and MS) remains to be determined.

Genome-based linkage and association studies have shown marked variations in the circulating levels of adipone-

nectin (Henneman *et al.*, 2010) and resistin (Menzaghi *et al.*, 2006). We have previously reported a strong association between *RETN* gene polymorphisms and resistin levels, but only a marginal association between *ADIPOQ* gene polymorphisms and adiponectin levels (Lau and Muniandy, 2011b). Recent comprehensive linkage disequilibrium mapping studies have shown that *ADIPOQ* (Heid *et al.*, 2010) and *RETN* (Asano *et al.*, 2010) gene polymorphisms, including the SNPs examined here, account for the high variability in the levels of their respective adipokines. Nevertheless, the genetic epistasis of adiponectin and resistin remain largely unexplored.

The results described here show that the *ADIPOQ* and *RETN* genes did not have a reciprocal effect on their adipokine levels, *i.e.*, *ADIPOQ* did not affect resistin levels nor did *RETN* affect adiponectin levels (Table 1). To our knowledge, an association between *RETN* gene polymorphisms and adiponectin levels has been reported in only one study in which the G-allele of SNP-420C > G at the *RETN* locus was associated with lower circulating adiponectin levels in Japanese patients (Miyamoto *et al.*, 2009). The present study is the first to examine the association between *ADIPOQ* gene polymorphisms and resistin levels. Although not statistically significant, the C-allele of

Table 1 - Estimative effect of alleles on adipokine levels.

<i>ADIPOQ</i> gene polymorphisms	Resistin (ng/mL) (n = 809)	Nominal p value	Empirical p value
SNP+45T > G (rs2241766)			
T-allele (78.18%)	+0.3415 (0.7756)	0.6597	1.0000
G-allele (21.82%)	-0.3415 (0.7756)		
SNP+276G > T (rs1501299)			
G-allele (70.52%)	+1.0562 (0.7161)	0.1402	0.4157
T-allele (29.48%)	-1.0562 (0.7161)		
SNP+639T > C (rs3821799)			
T-allele (56.37%)	-1.0448 (0.6485)	0.1072	0.3078
C-allele (43.63%)	+1.0448 (0.6485)		
SNP+1212A > G (rs6773957)			
A-allele (51.17%)	-1.1663 (0.6435)	0.0699	0.1910
G-allele (48.83%)	+1.1663 (0.6435)		
<i>RETN</i> gene polymorphisms	Adiponectin (µg/mL) (n = 809)	Nominal p value	Empirical p value
SNP-420C > G (rs1862513)			
C-allele (58.65%)	+0.0654 (0.2270)	0.7733	1.0000
G-allele (41.35%)	-0.0654 (0.2270)		
SNP+299G > A (rs3745367)			
G-allele (63.84%)	+0.2470 (0.2256)	0.2735	0.7912
A-allele (36.16%)	-0.2470 (0.2256)		

The results are expressed as the estimative effect (standard error in parentheses). The nominal p value was generated after adjusting for covariate age, ethnicity, body mass index (BMI), type 2 diabetes mellitus (T2DM) and metabolic syndrome (MS). The empirical p value was generated after performing the Bonferroni correction and adjusting for covariate age, ethnicity, BMI, T2DM and MS. The Bonferroni correction was used to counteract the problem of multiple comparisons by controlling for false positive results (type I errors). The estimative effect reflects the change in the mean serum resistin or adiponectin levels associated with a single allele.

SNP+639T > C and the G-allele of SNP+1212A > G at the *ADIPOQ* locus possibly provided a small contribution to the elevated serum resistin levels in individuals with these polymorphisms (Table 1). Recent large scale genome-wide association studies have suggested that these rare alleles are also associated with an increased risk of hypoadiponecตินemia (Ling *et al.*, 2009; Heid *et al.*, 2010). Since the *ADIPOQ* and *RETN* genes encode functionally-related proteins, a deleterious mutation in the *ADIPOQ* gene could indirectly interfere with *RETN* expression (probably through adipokine interaction or metabolic pathway rather than via genetic regulation). In agreement with this conclusion, genes with similar molecular functions normally have similar epistatic interactions. Thus, if in a particular condition

two metabolic pathways can produce the same metabolite, then inactivation of either pathway alone will have little effect on fitness (Lehner, 2011).

Paradoxically, the rare homozygous genotype (risk alleles) of SNP-420C > G at the *RETN* locus tended to be co-inherited with the common homozygous genotypes (protective alleles) of SNP+639T > C (Figure 1C) and SNP+1212A > G (Figure 1D) at the *ADIPOQ* locus. The T-allele of SNP+639T > C and the A-allele of SNP+1212A > G have a protective effect against T2DM and MS (Heid *et al.*, 2010; Siitonen *et al.*, 2011), whereas the G-allele of SNP-420C > G triggers these conditions (Ochi *et al.*, 2007; Miyamoto *et al.*, 2009). Recombination between SNPs in these genes may prevent disease-causing alleles of these

Table 2 - Generalized multifactor dimensionality reduction analysis.

Group comparisons	The best combination of SNPs (Optimum model)	Training balance accuracy	Testing balance accuracy	Significance test n (p ^E)	Cross-validation consistency
Malay men (n = 281)					
Control (n = 75) vs. MS without T2DM (n = 62)	SNP+299G > A, SNP+45T > G, SNP+639T > C	68.85%	62.02%	7 (0.1719)	10/10
Control (n = 75) vs. T2DM without MS (n = 43)	SNP+299G > A, SNP+45T > G, SNP+276G > T	69.26%	53.82%	6 (0.3770)	9/10
Control (n = 75) vs. T2DM with MS (n = 101)	SNP+45T > G, SNP+639T > C	63.37%	59.27%	7 (0.1719)	7/10
Chinese men (n = 264)					
Control (n = 73) vs. MS without T2DM (n = 54)	SNP-420C > G, SNP+299G > A, SNP+45T > G SNP+639T > C	72.61%	48.62%	4 (0.8281)	6/10
Control (n = 73) vs. T2DM without MS (n = 62)	SNP-420C > G, SNP+299G > A, SNP+639T > C	72.66%	63.76%	8 (0.0547)	10/10
Control (n = 73) vs. T2DM with MS (n = 75)	SNP-420C > G, SNP+299G > A, SNP+276G > T	68.70%	51.70%	5 (0.6230)	8/10
Indian men (n = 264)					
Control (n = 60) vs. MS without T2DM (n = 58)	SNP+299G > A, SNP+276G > T	66.23%	55.33%	6 (0.3770)	10/10
Control (n = 60) vs. T2DM without MS (n = 66)	SNP-420C > G, SNP+299G > A, SNP+1212A > G	66.89%	59.60%	7 (0.1719)	9/10
Control (n = 60) vs. T2DM with MS (n = 80)	SNP-420C > G, SNP+299G > A, SNP+276G > T, SNP+639T > C	68.08%	51.02%	5 (0.6230)	10/10

Significance test: n indicates the number of tests with accuracies greater than 50% and, in parentheses (p^E), the empirical p value (after correction for multiple comparisons) computed from 1,000,000 permutations. The exhaustive search method configuration was used. With a 10-fold cross-validation, the data are divided into 10 equal parts and the model is developed on 9/10 of the data (training set) and then tested on 1/10 of the remaining data (testing set). This is repeated for each possible combination of 9/10 (training balance accuracy) and 1/10 (testing balance accuracy) of the data, with the resulting 10 testing accuracies then being averaged. The cross-validation consistency is a measure of how many times out of 10 divisions of the data that GMDR found the same best model. The model with the combination of SNPs that maximized the average cross-validation consistency and minimized the average prediction error was selected. Note: *RETN* (SNP-420C > G and SNP+299G > A); *ADIPOQ* (SNP+45T > G, SNP+276G > T, SNP+639T > C and SNP+1212A > G); T2DM = type 2 diabetes mellitus; MS = metabolic syndrome.

SNPs from being co-inherited. Indeed, some studies have reported a prevalence of antagonistic epistasis, *i.e.*, the tendency of multiple mutations to have a mitigating/alleviating effect rather than a reinforcing/aggravating effect (Desai *et al.*, 2007). Recombination and mutations provide the basis for less synergistic or more antagonistic epistasis (Desai *et al.*, 2007). The results described here provide no conclusive evidence for a link between the structural similarity and/or functional similarity (synergistic or antagonistic epistasis) between these two genes. Indeed, the extent to which the *ADIPOQ* gene interferes with *RETN* expression remains to be determined.

Although *ADIPOQ* and *RETN* are located on different chromosomes, the intermolecular epistatic interaction between these genes is possible (Charalampos and Richard, 2006). Epistatic interactions between mutations in different genes may be caused by different underlying molecular mechanisms (Lehner, 2011). If two genes share a common function, then the loss of one gene could be compensated for by continued activity of the second gene (Lehner, 2011). Since adiponectin and resistin have opposite physiological effects (Lau and Muniandy, 2011a), polymorphisms in *ADIPOQ* and *RETN* are likely to affect disease outcome by acting antagonistically during natural selection. Despite the structural similarity between *ADIPOQ* and *RETN* (Figures 1 and 2), there was no evidence of an intermolecular epistatic interaction between these genes (Table 2). The *ADIPOQ* and *RETN* genes may not, by themselves, exhibit a significant association with the risk of T2DM and MS because their combined effect may be relatively small and dependent on genotypes at other loci; the latter may compensate for variation in the loci under study and may differ across populations. Indeed, recent large-scale systematic reviews and meta-analyses have shown marked heterogeneity among SNPs associated with the *ADIPOQ* (Zhao and Zhao, 2011) and *RETN* (Wen *et al.*, 2010) loci.

The findings of this study relate to Malaysian men and may not be applicable to other groups because of genetic differences. Fine-mapping studies are needed since our investigation did not cover upstream and downstream variations in the *ADIPOQ* and *RETN* genes. Comprehensive linkage disequilibrium mapping and haplotype-based data mining would allow a detailed analysis of haplotype-haplotype interactions and more insight into the genetic risk factors associated with T2DM and MS. In addition to the *ADIPOQ* and *RETN* genes, other genes involved in the susceptibility to T2DM and MS should be included when modelling high-order gene-gene interactions. The availability of more powerful computer-based approaches will allow large scale genome-proteome interaction-based analyses to map epistatic interactions on a wider scale. In this context, understanding the role of “gene-environment” interactions is crucial for the characterization of low-penetrance genes. Further studies with larger sample sizes

than that used here are required in order to reach more reliable conclusions.

Acknowledgments

This work was supported by University of Malaya postgraduate research grants (grant nos. PS102-2009A and PS201-2010A), a short-term research university grant (grant no. FS232-2008C) and an e-science fund grant (grant no. 12-02-03-2044). We thank all those who participated in this project, including the nurses of the University of Malaya Medical Centre (UMMC), particularly Madam Farahwahidah who helped with blood sample collection. We also thank members of our laboratory who provided support and helpful discussions throughout this work.

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Internet Resources

- HOMA calculator software ver. 2.2.2, Diabetes Trials Unit, Oxford University, <http://www.dtu.ox.ac.uk/homa> (April 25, 2011).
- HAPSTAT program, ver. 3.0, <http://www.bios.unc.edu/~lin/hapstat> (April 3, 2011).
- Generalized Multifactor Dimensionality Reduction (GMDR) Beta program, ver. 0.7, <http://sourceforge.net/projects/gmdr> (May 18, 2011).

Associate Editor: Mara Hutz

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