## **Supplementary information**

The cyanobacterial oxadiazine nocuolin A shows broad-spectrum toxicity against protozoans and the nematode *C. elegans* 

Ana R. Vieira<sup>1</sup>, Francisco Camacho<sup>1,2</sup>, Maria L. Sousa<sup>1</sup>, Sara Luelmo<sup>3</sup>, Nuno Santarém<sup>3,4</sup>, Anabela Cordeiro-da-Silva<sup>3,4</sup>, Pedro N. Leão<sup>1,\*</sup>

<sup>1</sup>Interdisciplinary Centre of Marine and Environmental Research (CIIMAR/CIMAR), University of Porto, Matosinhos, Portugal

<sup>2</sup>Department of Biology and Chemistry, Faculty of Sciences, University of Porto, Porto, Portugal

<sup>3</sup>Institute for Research and Innovation in Health (i3S), University of Porto, Porto, Portugal

<sup>4</sup>Laboratory of Microbiology, Department of Biological Sciences, Faculty of Pharmacy, University of Porto, Porto, Portugal

**Supplementary Figure S1.** Schematic highlighting *noc* locus and its proposed relationship with nocuolin A, chlorosphaerolactylate A and the nocuolactylate A in the cyanobacterial strain *Nodularia* sp. LEGE 06071.

**Supplementary Figure S2.** The cyanobacterial compound nocuolin A affects *Acanthamoeba castellanii* viability.

**Supplementary Figure S3.** 1H NMR (400 MHz) spectra of nocuolin A isolation in chloroform-d.

**Supplementary Figure S4.** Dose response curves used to generate the IC50 of chlorhexidine digluconate and nocuolin A in *Acanthamoeba castellanii* and *Dictyostelium discoideum*.

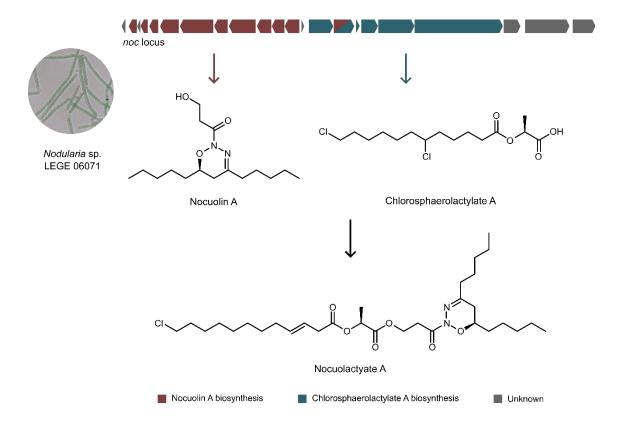
**Supplementary Figure S5.** Microscope pictures of *A. castellanii* cells after treatment with chlorhexidine digluconate and nocuolin A before and after its removal overtime.

**Supplementary Figure S6:** *Nodularia* sp. LEGE 06071 is resistant to *D. discoideum* grazing in a solid medium drop grazing assay.

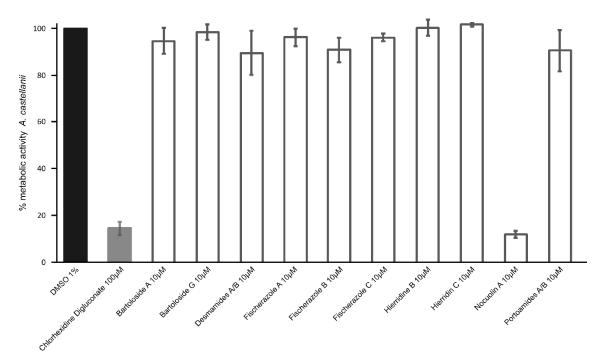
**Supplementary Figure S7:** Detection of nocuolactylates and chlorosphaerolactylates in the lawns of the monocultures and co-cultures of *Nodularia* sp. LEGE 06071 and *Sphaerospermopsis* sp. LEGE 00249 with amoebae.

**Supplementary Figure S8:** Dose response curves used to generate the IC50 of nocuolin A and the control compounds miltefosine and pentamidine in *Trypanosoma brucei*, *Leishmania infantum* and THP-1 cells.

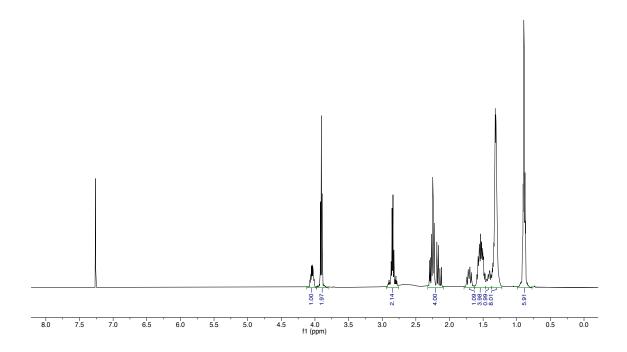
**Supplementary Figure S9:** Nocuolin A toxicity evaluation using *C. elegans*.



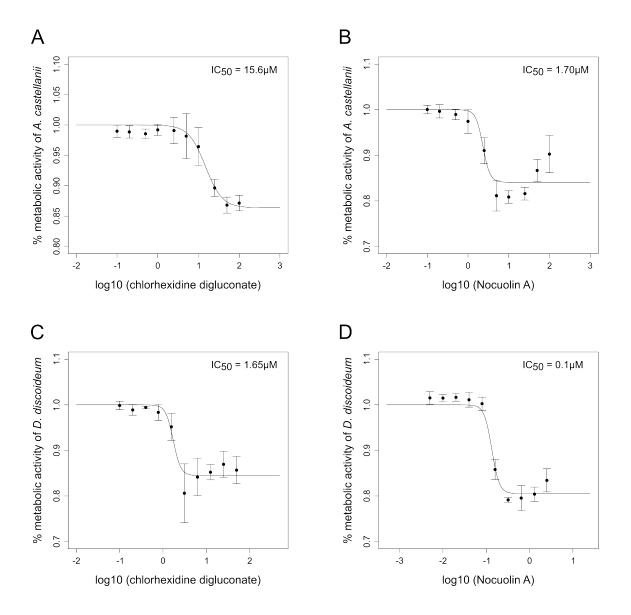
Supplementary Fig. S1. Schematic highlighting *noc* locus and its proposed relationship with nocuolin A, chlorosphaerolactylate A and the nocuolactylate A in the cyanobacterial strain *Nodularia* sp. LEGE 06071. A portion of the  $\approx$ 40 kb BGC putatively assigned to nocuolin A (in red), and another portion putatively assigned to the chlorosphaerolactylates (in green). The nocuolactylates could be products of both red and green represented BGC portion, which colocalize in cyanobacterial genomes, however, further evidence is required to unequivocally connect *noc* genes to these metabolites (Figueiredo *et al.*, 2021; Martins *et al.*,2022). The cyanobacterial strain *Sphaerospermopsis* sp. LEGE 00249 also contain the *noc* locus but only the chlorosphaerolactylates were reported for this strain.



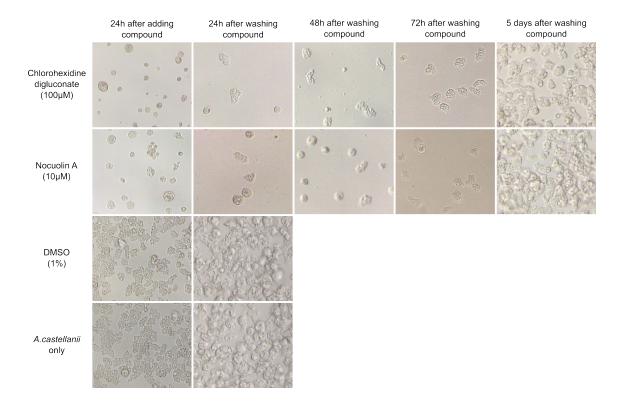
Supplementary Fig. S2 The cyanobacterial compound nocuolin A affects *Acanthamoeba castellanii* viability. The toxicity of diverse cyanobacterial pure compounds ( $10 \mu M$ ) was tested against *A. castellanii*. The detergent chlorhexidine digluconate was used as the positive control. Average of three assays. Black bar represents *A. castellanii* cells treated with DMSO 1 % (negative control); grey bar represents *A. castellanii* cells treated with chlorhexidine digluconate (positive control) and white bars represent *A. castellanii* cells treated with the diverse cyanobacterial compounds.



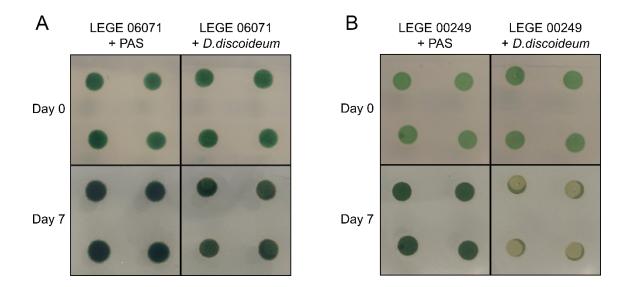
**Supplementary Fig. S3.** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) spectrum of nocuolin A (> 99% purity) used in this study. The non-integrated broad signal at  $\sim$ 2.75 ppm corresponds to the exchangeable alcohol proton.



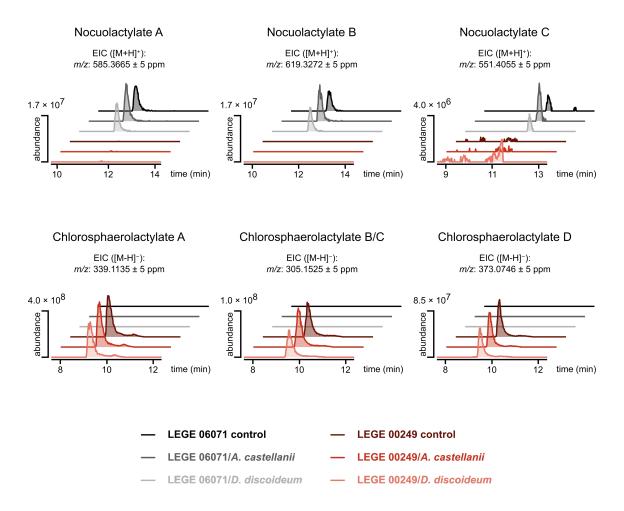
**Supplementary Fig. S4**: Dose response curves used to generate the IC50 of A) chlorhexidine digluconate and B) nocuolin A in *Acanthamoeba castellanii* and, C) chlorhexidine digluconate and D) nocuolin A in *Dictyostelium discoideum*. Average of three assays. The IC50 was 15.6  $\mu$ M and 1.70  $\mu$ M for chlorhexidine diglocunate and nocuolin A in *A. castellanii* and 1.65  $\mu$ M and 0.1  $\mu$ M, respectively for *D. discoideum*.



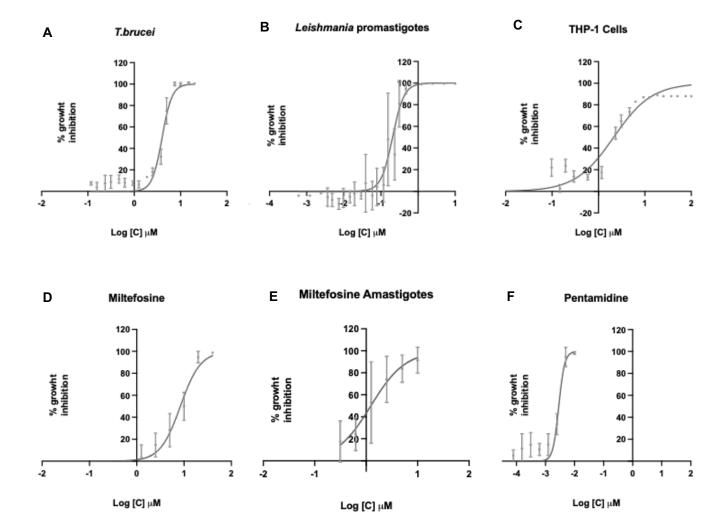
**Supplementary Fig. S5**: Microscope pictures of *A. castellanii* cells after treatment with chlorhexidine digluconate and nocuolin A before and after its removal overtime. Amoeba cells encysted after adding the positive control chlorhexidine diglocunate and nocuolin A, but could recover after a few days of washing out these compounds. *A. castellanii* cells without treatment and treated with DMSO 1 % were used as negative controls and are observed as active trophozoites.



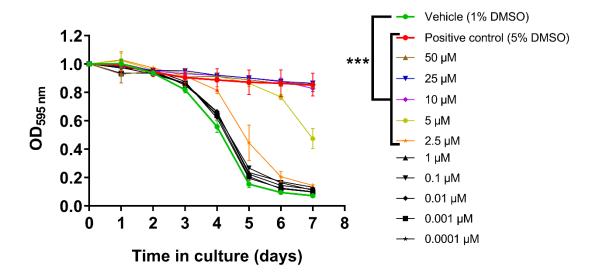
**Supplementary Fig. S6**: *Nodularia* sp. LEGE 06171 is resistant to *Dictyostelium discoideum* grazing. Cyanobacterial drop grazing assay of A) *Nodularia* sp. LEGE 06071 and B) *Sphaerospermopsis* sp. LEGE 00249 against *D. discoideum* grazing over a period of one week. Yellow areas on top of the co-culture drop indicates grazing of the cyanobacteria. The control drops are cyanobacteria with only PAS amoeba medium on top. This assay was performed in triplicate. *Nodularia* sp. LEGE 06071, the producer strain of nocuolin A, shows resistance to *D. discoideum* grazing, as opposite to *Sphaerospermopsis* sp. LEGE 00249, that is sensitive to this amoebae strain.



**Supplementary Fig. S7**: Detection of nocuolactylates and chlorosphaerolactylates in the lawns of the monocultures and co-cultures of *Nodularia* sp. LEGE 06071 and *Sphaerospermopsis* sp. LEGE 00249 with amoebae. LC-HRMS-derived extracted ion chromatograms (EICs) of nocuolactylates A-C detected in the monocultures of LEGE 06071 and its co-cultivation with both *A. castellanii* and *D. discoideum*, and the chlorosphaerolactylates A-D detected in the monocultures of LEGE 00249 and its co-cultivation with *A. castellanii* and *D. discoideum*.



Supplementary Fig. S8: Dose response curves used to generate the IC50 of nocuolin A and the control compounds miltefosine and pentamidine in *Trypanosoma brucei*, *Leishmania infantum* and THP-1 cells. Dose-response curve of nocuolin A in A) *Trypanosoma brucei*, B) *Leishmania infantum* promastigotes, and C) THP-1 cells. Dose-response curve of miltefosine in D) *Leishmania infantum* promastigotes and E) amastigotes and F) pentamidine in *Trypanosoma brucei*. Miltefosine and pentamidine were tested as reference drugs in each correspondent assay. The IC50 for nocuolin A was 3,990  $\mu$ M, 0,2062  $\mu$ M and 2,127  $\mu$ M for *Trypanosoma brucei*, *Leishmania infantum* and THP-1 cells, respectively. The IC50 for miltefosine was 8,369  $\mu$ M and 2,12  $\mu$ M for *Leishmania infantum* promastigotes and intracellular amastigotes, respectively and, the IC50 for pentamidine was 0,002864  $\mu$ M for *Trypanosoma brucei*. Average of three assays.



**Supplementary Fig. S9:** Nocuolin A toxicity evaluation using *C. elegans*. The OD<sub>595nm</sub> of the *E. coli* OP50 suspension was measured daily of WT animals treated with nocuolin A at the indicated concentrations. For each condition tested, the mean OD<sub>595nm</sub> was calculated for each day from five replicates and plotted over time. Toxic concentrations of 10  $\mu$ M and 5  $\mu$ M was defined as food clearance differs from vehicle (DMSO 1 %). Control DMSO (1 % DMSO) corresponds to drug vehicle and DMSO at 5 % (5 % DMSO) was used as positive (toxic compound) control. WT, wild-type. Average of tree assays.