



Fundamental and Applied Advances in Stem Cell Therapeutic Research

Makram Merimi ^{1,2,†}, Saida Rahmani ^{1,2,†}, Ahmed Afailal Tribak ^{1,†}, Fatima Bouhtit ^{1,2,‡}, Hassan Fahmi ^{3,‡} and Mehdi Najar ^{3,4,*,‡}

- Experimental Hematology, Jules Bordet Institute, Unive and nd nd ité Libre de Bruxelles, 1070 Bruxelles, Belgium; makram.merimi.cri@gmail.com (M.M.); saida.ramani@gmail.com (S.R.); tribakafailal.ahmed@gmail.com (A.A.T.); bouhtitfatima@gmail.com (F.B.)
- ² Genetics and Immune Cell Therapy Unit, Faculty of Sciences, University Mohammed Premier, Oujda 60000, Morocco
- ³ Osteoarthritis Research Unit, University of Montreal Hospital Research Center (CRCHUM), Department of Medicine, University of Montreal, Montreal, QC H2X 0A9, Canada; h.fahmi@umontreal.ca
- ⁴ Laboratory of Clinical Cell Therapy, Jules Bordet Institute, Université Libre de Bruxelles, 1070 Brussels, Belgium
- * Correspondence: mnajar@ulb.ac.bCe
- + These authors contributed equally to this work.
- ‡ Equal senior authors.

check for updates

Citation: Merimi, M.; Rahmani, S.; Afailal Tribak, A.; Bouhtit, F.; Fahmi, H.; Najar, M. Fundamental and Applied Advances in Stem Cell Therapeutic Research. *Cells* **2022**, *11*, 1976. https://doi.org/10.3390/ cells11121976

Received: 13 May 2022 Accepted: 6 June 2022 Published: 20 June 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

We are pleased to present this Special Issue of Cells, entitled 'Feature Papers in Stem Cells'. We hope that this collection of papers may contribute greatly to this field by discussing and presenting new outcomes of basic and translational stem cell-based regenerative medicine research. The rapid progress in the field of stem cell research has laid strong foundations for their use in regenerative medicine applications involving injured or diseased tissues. Cellular therapy aims to replace damaged resident cells by restoring cellular and molecular environments suitable for tissue repair and regeneration. Growing evidence indicates that some of the observed therapeutic outcomes of stem cell-based therapy are due to paracrine effects (including extracellular vesicles), rather than long-term engraftment or the survival of transplanted cells [1]. Embryonic and induced pluripotent stem cells (ESCs and iPSCs), as well as adult stem cells, hold great promise for future cell replacement therapies. Among other candidates, mesenchymal stem/stromal cells (MSCs) represent a critical component of stromal niches known to be involved in tissue homeostasis [2]. Additional evidence suggests that MSCs originate from perivascular cells-principally pericytes that are vascular mural cells—within multiple human organs including lung, adipose tissue and placenta [3]. Accordingly, MSCs play a crucial role during lung development by interacting with the airway epithelium, and also during lung regeneration and remodeling after injury, particularly in chronic obstructive pulmonary disease [4]. During tissue healing, MSCs may exhibit several therapeutic functions to support the repair and regeneration of injured tissue. The process underlying these effects likely involves the migration and homing of MSCs, as well as their immune-tropic functions [5]. Interestingly, tissue-nonspecific alkaline phosphatase (ALP) (TNSALP), a ubiquitous membrane-bound glycoprotein capable of providing inorganic phosphate by catalyzing the hydrolysis of organic phosphate esters, or removing inorganic pyrophosphate that inhibits calcification, is highly expressed in juvenile cells, such as pluripotent stem cells (i.e., ESCs (iPSCs) and somatic stem cells (i.e., MSCs), and is involved in their maintenance and differentiation [6]. Understanding and controlling these cellular products requires in-depth knowledge of their maintenance mechanisms and their exit from undifferentiated states in specific biomaterials mimicking native niches. An interesting approach has been established for differentiating porcine epiblast stem cells (pEpiSCs) into proliferating and functional endothelial cells (ECs). Functional tests revealed that the generated ECs could be used in in vitro assays to examine angiogenesis or cellular

responses to various vascular diseases [7]. In another setting, a male mouse model for high running performance recruited myogenic precursor cells/SATCs with lower activation thresholds that responded more rapidly to external stimuli and were more primed for differentiation at the expense of more primitive cells. Satellite cells (SATCs), known as the most abundant skeletal muscle stem cells, play a main role in muscle plasticity, including in the adaptive response following physical activity [8]. In parallel, using pluripotent stem cells (PSCs) to generate hepatocytes is preferable because of their availability and scalability. However, the efficient maturation of PSC-derived hepatocytes towards functional units in liver organoids (LOs) remains a challenging subject. The incorporation of cell-sized microparticles (MPs) derived from the liver extracellular matrix (ECM) provides an enriched tissue-specific microenvironment for the further maturation of hepatocytes inside LOs [9]. This approach has led to the improvement of hepatocyte-like cells in terms of gene expression and function, CYP activities, albumin secretion, and the metabolism of xenobiotics. An experimental basis for the application of stem cells in the treatment of keloids, a pathological scar observed during wound healing, has been developed. Moreover, a co-culture method has been set up to investigate the influence and mechanism of human dental pulp stem cells (HDPSCs) on keloid fibroblast properties [10]. HDPSCs inhibited the migration, the synthesis of the extracellular matrix, and the expression of pro-fibrotic genes within human keloid fibroblasts (HKFs), while promoting the expression of anti-fibrotic genes. Therefore, it can be concluded that HDPSCs can themselves be used as a tool for restraining/hindering the initiation or progression of fibrotic tissue. Mechanistically, new findings have established ten eleven translocation 1 (Tet1) as a regulator of embryonic stem cell (ESC) proliferation by suppressing p21 to ensure a rapid G1-to-S progression [11]. Moreover, Zscan4, which is highly upregulated in telomerase-deficient late-generation mouse ESCs and human alternative lengthening of telomeres (ALT) cancer cells, has been shown to contribute to the telomere maintenance of those cells without telomerase activities [12]. Several features are still to be identified and resolved for improving the safety and efficiency of stem cell-based therapy, in particular for the use of biological delivery systems. Thus, a systematic literature review investigates the potential of therapy with MSCs associated with fibrin glue on the regeneration of the central or peripheral nervous system [13]. Recently, various strategies using a hydrogel-based system, both as encapsulated stem cells and as biocompatible patches loaded with stem cells and applied at the tissue damage site, were developed for regenerating the infarcted myocardium [14]. Joint engineering, representing a potential tool for cartilage regeneration, is an interdisciplinary field that aims to recreate a neo-tissue whose physical and biochemical properties are close to those of the native tissue. It combines cells, biomaterials, and environmental factors. A particular focus on the extrusion bioprinting of cellularized hydrogels for articular cartilage tissue engineering has been discussed [15]. Furthermore, approaches for optimizing standard MSC culture protocols during this essential primary step of in vitro expansion are required. Several studies suggest some improvements in culture media components (amino acids, ascorbic acid, glucose level, growth factors, lipids, platelet lysate, trace elements, serum, and xenogeneic components) as well as culture conditions and processes (hypoxia, cell seeding, and dissociation during passaging) in order to preserve MSC phenotypes and functionality during the primary phase of in vitro culture [16]. Collectively, this Special Issue, managed and supervised by Dr. Mehdi Najar, successfully gathers a great collection of research articles and reviews highlighting recent fundamental and applied advances in different types of stem cells.

Author Contributions: M.M. and M.N. conceived and designed the editorial. M.M., M.N., S.R., A.A.T., F.B. and H.F. have made a substantial, direct and intellectual contribution to the work. All authors listed contributed to manuscript writing, revision, reading, and approved the submitted version. All authors have read and agreed to the published version of the manuscript.

Funding: This work was funded by Generation Life Foundation, 'Fonds Lambeau-Marteaux', 'Fonds National de la Recherche Scientifique (FNRS)', 'Télévie', 'Les Amis de l'Institut Jules Bordet', 'La Chaire en Arthrose de l'Université de Montréal, The Arthritis Society (SOG-20-0000000046) and The Canadian Institutes of Health Research (PJT 175-1110).

Acknowledgments: We would like to thank the Cell Therapy Unit Team for their inspiring dialogs.

Conflicts of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

- Jarrige, M.; Frank, E.; Herardot, E.; Martineau, S.; Darle, A.; Benabides, M.; Domingues, S.; Chose, O.; Habeler, W.; Lorant, J.; et al. The Future of Regenerative Medicine: Cell Therapy Using Pluripotent Stem Cells and Acellular Therapies Based on Extracellular Vesicles. *Cells* 2021, *10*, 240. [CrossRef] [PubMed]
- Najar, M.; Melki, R.; Khalife, F.; Lagneaux, L.; Bouhtit, F.; Moussa Agha, D.; Fahmi, H.; Lewalle, P.; Fayyad-Kazan, M.; Merimi, M. Therapeutic Mesenchymal Stem/Stromal Cells: Value, Challenges and Optimization. *Front. Cell Dev. Biol.* 2021, 9, 716853. [CrossRef] [PubMed]
- Li, C.; Zhao, H.; Wang, B. Mesenchymal stem/stromal cells: Developmental origin, tumorigenesis and translational cancer therapeutics. *Transl. Oncol.* 2021, 14, 100948. [CrossRef] [PubMed]
- 4. Nasri, A.; Foisset, F.; Ahmed, E.; Lahmar, Z.; Vachier, I.; Jorgensen, C.; Assou, S.; Bourdin, A.; De Vos, J. Roles of Mesenchymal Cells in the Lung: From Lung Development to Chronic Obstructive Pulmonary Disease. *Cells* **2021**, *10*, 3467. [CrossRef] [PubMed]
- Merimi, M.; El-Majzoub, R.; Lagneaux, L.; Moussa Agha, D.; Bouhtit, F.; Meuleman, N.; Fahmi, H.; Lewalle, P.; Fayyad-Kazan, M.; Najar, M. The Therapeutic Potential of Mesenchymal Stromal Cells for Regenerative Medicine: Current Knowledge and Future Understandings. *Front. Cell Dev. Biol.* 2021, 9, 661532. [CrossRef] [PubMed]
- Sato, M.; Saitoh, I.; Kiyokawa, Y.; Iwase, Y.; Kubota, N.; Ibano, N.; Noguchi, H.; Yamasaki, Y.; Inada, E. Tissue-Nonspecific Alkaline Phosphatase, a Possible Mediator of Cell Maturation: Towards a New Paradigm. *Cells* 2021, 10, 3338. [CrossRef] [PubMed]
- Shin, J.-H.; Seo, B.-G.; Lee, I.-W.; Kim, H.-J.; Seo, E.-C.; Lee, K.-M.; Jeon, S.-B.; Baek, S.-K.; Kim, T.-S.; Lee, J.-H.; et al. Functional Characterization of Endothelial Cells Differentiated from Porcine Epiblast Stem Cells. *Cells* 2022, 11, 1524. [CrossRef] [PubMed]
- Petkov, S.; Brenmoehl, J.; Langhammer, M.; Hoeflich, A.; Röntgen, M. Myogenic Precursor Cells Show Faster Activation and Enhanced Differentiation in a Male Mouse Model Selected for Advanced Endurance Exercise Performance. *Cells* 2022, *11*, 1001. [CrossRef] [PubMed]
- Zahmatkesh, E.; Ghanian, M.H.; Zarkesh, I.; Farzaneh, Z.; Halvaei, M.; Heydari, Z.; Moeinvaziri, F.; Othman, A.; Ruoß, M.; Piryaei, A.; et al. Tissue-Specific Microparticles Improve Organoid Microenvironment for Efficient Maturation of Pluripotent Stem-Cell-Derived Hepatocytes. *Cells* 2021, *10*, 1274. [CrossRef] [PubMed]
- 10. Yan, M.; Fu, L.-L.; Nada, O.A.; Chen, L.-M.; Gosau, M.; Smeets, R.; Feng, H.-C.; Friedrich, R.E. Evaluation of the Effects of Human Dental Pulp Stem Cells on the Biological Phenotype of Hypertrophic Keloid Fibroblasts. *Cells* **2021**, *10*, 1803. [CrossRef] [PubMed]
- 11. Chrysanthou, S.; Flores, J.C.; Dawlaty, M.M. Tet1 Suppresses p21 to Ensure Proper Cell Cycle Progression in Embryonic Stem Cells. *Cells* **2022**, *11*, 1366. [CrossRef] [PubMed]
- 12. Dan, J.; Zhou, Z.; Wang, F.; Wang, H.; Guo, R.; Keefe, D.L.; Liu, L. Zscan4 Contributes to Telomere Maintenance in Telomerase-Deficient Late Generation Mouse ESCs and Human ALT Cancer Cells. *Cells* **2022**, *11*, 456. [CrossRef] [PubMed]
- Ortiz, A.d.C.; Fideles, S.O.M.; Pomini, K.T.; Bellini, M.Z.; Pereira, E.d.S.B.M.; Reis, C.H.B.; Pilon, J.P.G.; de Marchi, M.Â.; Trazzi, B.F.d.M.; da Silva, W.S.; et al. Potential of Fibrin Glue and Mesenchymal Stem Cells (MSCs) to Regenerate Nerve Injuries: A Systematic Review. *Cells* 2022, *11*, 221. [CrossRef] [PubMed]
- Sharma, V.; Dash, S.K.; Govarthanan, K.; Gahtori, R.; Negi, N.; Barani, M.; Tomar, R.; Chakraborty, S.; Mathapati, S.; Bishi, D.K.; et al. Recent Advances in Cardiac Tissue Engineering for the Management of Myocardium Infarction. *Cells* 2021, 10, 2538. [CrossRef]
- 15. Messaoudi, O.; Henrionnet, C.; Bourge, K.; Loeuille, D.; Gillet, P.; Pinzano, A. Stem Cells and Extrusion 3D Printing for Hyaline Cartilage Engineering. *Cells* **2021**, *10*, 2. [CrossRef]
- Nikolits, I.; Nebel, S.; Egger, D.; Kreß, S.; Kasper, C. Towards Physiologic Culture Approaches to Improve Standard Cultivation of Mesenchymal Stem Cells. Cells 2021, 10, 886. [CrossRef] [PubMed]