



In Situ CO₂ Efflux from Leaf Litter Layer Showed Large Temporal Variation Induced by Rapid Wetting and Drying Cycle

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Abstract

We performed continuous and manual in situ measurements of CO₂ efflux from the leaf litter layer (R_{LL}) and water content of the leaf litter layer (LWC) in conjunction with measurements of soil respiration (R_S) and soil water content (SWC) in a temperate forest; our objectives were to evaluate the response of R_{LL} to rainfall events and to assess temporal variation in its contribution to R_S . We measured R_{LL} in a treatment area from which all potential sources of CO₂ except for the leaf litter layer were removed. Capacitance sensors were used to measure LWC. R_{LL} increased immediately after wetting of the leaf litter layer; peak R_{LL} values were observed during or one day after rainfall events and were up to 8.6-fold larger than R_{LL} prior to rainfall. R_{LL} declined to pre-wetting levels within 2–4 day after rainfall events and corresponded to decreasing LWC, indicating that annual R_{LL} is strongly influenced by precipitation. Temporal variation in the observed contribution of R_{LL} to R_S varied from nearly zero to 51%. Continuous in situ measurements of LWC and CO₂ efflux from leaf litter only, combined with measurements of R_S , can provide robust data to clarify the response of R_{LL} to rainfall events and its contribution to total R_S .

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Introduction

Efflux of CO₂ from the soil surface (soil respiration; R_S), which is the sum of respiration by autotrophs and heterotrophs, is an important component of total CO₂ efflux from forest ecosystems [1–3]. The R_S : total ecosystem respirations varied from 58% to 76% in a mixed coniferous-deciduous forest [4], depending on interannual and seasonal changes in autotrophic and heterotrophic respiration; variability in R_S can affect the forest carbon balance on daily and seasonal time scales. To explain the cause of variability in R_S , many studies have attempted to separate differing sources of R_S and to examine factors controlling CO₂ efflux rate from each source [5–7]. Especially in forest ecosystems, heterotrophic respiration consists of CO₂ efflux from various sources (e.g., leaf and root litter, woody debris, soil organic matter) and their rates are controlled by their specific environmental condition such as water content (WC) and temperature [8], physical properties of the substrate (e.g., density and structure) [9,10], and chemical properties (e.g., labile and recalcitrant carbon) [11,12]. Moreover, CO₂ efflux from the various heterotrophic sources responds differently to these controlling factors, which illustrates the complexity of R_S . In recent decades, a variety of methods for separating components of heterotrophic respiration

and for determining their contribution to total R_S have been developed [9,13].

Among heterotrophic sources of CO₂, the leaf litter layer (L-layer) is a significant reservoir of degradable carbon and a large potential source of CO₂ efflux from forest soils [14]. In temperate forests, the contribution of CO₂ efflux from the L-layer (leaf litter respiration; R_{LL}) to R_S is reported to range from 23% to 48% [13,15,16]. The L-layer is in direct contact with rainfall, solar radiation, and wind, and environmental conditions (e.g., WC and temperature) can change more dynamically in the L-layer than in lower soil layers. Rapid and transient temporal variation in WC of the L-layer has been observed, especially in warm climates [16,17]. Heterotrophic respiration responds rapidly to changes in moisture status [17,18]; therefore, rapid and transient wetting and drying cycles would produce large temporal variations in R_{LL} . This would significantly affect variation in R_S [17,19], suggesting that R_{LL} is an important controller of temporal (daily and seasonal) patterns in the carbon balance in warm regions [19,20].

Several methods for measuring R_{LL} and for calculating its contribution to R_S have been explored. Cisneros-Dozal et al. [21] used an isotope mass balance method and reported that the contribution of R_{LL} to R_S increased from 5% to 37% in response to water addition after transient drought. Deforest et al. [15]

determined that the annual contribution of R_{LL} to R_S was 48% \pm 12% by measuring R_S with and without the L-layer, and the ratio was consistent over a range of environmental conditions. However, there is little information about temporal variation in R_{LL} in relation to rainfall events because of the difficulty of continuous and direct measurement of R_{LL} in situ.

To continuously measure CO₂ efflux from the L-layer only, in parallel with measurement of R_S , we developed an approach for measuring R_{LL} using an automated chamber method in a treatment area from which all CO₂ sources except for the L-layer were removed. In parallel with R_{LL} and R_S measurements, we continuously measured water content of the L-layer (LWC) and soil water content (SWC). LWC was measured using a method developed by Ataka et al. [22], in which intact leaf litter was attached to surrounding capacitance sensors. Sensors were also placed on top of the L-layer and at the boundary between the L- and mineral layers. From these continuous in situ measurements, we investigated the response of R_{LL} to rainfall events by comparing R_{LL} with R_S , and examined temporal variation in the contribution of R_{LL} to R_S in a warm temperate forest in Japan.

Materials and Methods

Ethics statement

The study site (Yamashiro Experimental Forest) is maintained by the Forestry and Forest Products Research Institute. All necessary permits were obtained for the field study, and the study did not involve endangered or protected species.

Study site

Our observations of R_{LL} and R_S were conducted at the Yamashiro Experimental Forest in southern Kyoto Prefecture, Japan (34°47'N, 135°50'E). The study site is a 1.7-ha watershed characterized by an annual mean air temperature of 15.5°C (maximum, 34.8°C; minimum, -3.9°C) and annual precipitation of 1449 mm [2]. The rainy season generally occurs from early June to mid-July. Daily rates of evaporation from the forest floor are 0.4–0.8 mm day⁻¹ for 1–2 days after precipitation, declining thereafter to 0.2–0.3 mm day⁻¹ [23]. The soils are Regosols with sandy loam or loamy sand texture and contain fine gravel (53% by mass) composed of residual quartz crystals from granite parent material [24]. These are immature soils in which the thickness of the A horizon is 2–3 cm. Deciduous broad-leaved, evergreen broad-leaved, and coniferous tree species account for 66%, 28%, and 6% of the living tree biomass, respectively [25]. The forest is dominated by *Quercus serrata* Thunb., which accounts for approximately 33% of the biomass. The L-layer (approximately 3–4 cm thick) consists mainly of fresh *Q. serrata* litter. There is no substantial organic horizon below the L-layer.

Automated chamber method for measuring leaf litter respiration and soil respiration

We measured R_{LL} and R_S using an automated dynamic chamber system with an infrared gas analyzer (IRGA, GMP343; Vaisala Group, Vantaa, Finland) (Fig. 1A). The system consisted of two automated circular chambers for R_{LL} and R_S measurement, four solenoid valves, a pump, mass flow meter, and IRGA. The chambers (surface area 320 cm²) were made from PVC collars with clear acrylic lids that can be opened and closed automatically using an air cylinder. Air was supplied to the cylinder from a compressor. To ensure a seal between the chamber and the closed lid, a soft rubber gasket was attached to the top edge of the chamber. Opening and closing of the chamber lid and solenoid

valves of each chamber were regulated synchronously by a control unit (ZEN, OMRON, Kyoto, Japan).

The duration of measurement of CO₂ concentration inside each chamber was 6 min and was performed twice per hour. The CO₂ concentration in each chamber was recorded at 1-s intervals using a data logger (GL220, Graphtec, Kanagawa, Japan). We calculated R_{LL} and R_S from the increase in CO₂ concentration (ΔC_{CO_2}) using linear regression. Data from the first 2 min were discarded to avoid effects of closing the chamber. R_{LL} and R_S were calculated using the following equation:

$$R = \frac{\Delta C_{CO_2}}{10^6} \times \frac{V}{V_{air}} \frac{273.2}{273.2 + T} \times M_{CO_2} \times \frac{1}{A}, \quad (1)$$

where R is respiration (mg CO₂ m⁻² s⁻¹), ΔC_{CO_2} is the change in CO₂ concentration per unit time (CO₂ ppm s⁻¹), V is the volume of the system (L), V_{air} is the standard gas volume (22.41 L mol⁻¹), T is temperature inside the chamber (°C), M_{CO_2} is the molecular weight of CO₂ (44.01 g mol⁻¹), and A is the soil surface area covered by the chamber (m²).

To continuously measure CO₂ efflux from the L-layer only, we developed an approach for measuring R_{LL} by using an automated chamber method in a treatment area in which all potential CO₂ sources (e.g., organic soil and fine roots) except for the L-layer were replaced with combusted granite soil (Fig. 1B). To prepare the treatment area (1 m²), we removed surface soil (approximately 5 cm). An acrylic board was placed on the bottom and sides of the treatment area to prevent penetration of roots; a drain tube was located at the bottom of the board to prevent the treatment area from flooding with rainwater. The treatment area was then filled with granite soil combusted in a muffle furnace (500°C for 1 day). For R_{LL} measurement, we placed a PVC collar (320-cm² surface area) and acrylic board below the collar. The board was set at a slight incline to drain rainwater from the collar. We added 15 g of newly fallen leaf litter, which represents the average litterfall mass per unit ground surface area at this site, to the collar. We added the leaf litter to each chamber on January 2012. To acquire data on the temporal variation in R_{LL} of fresh leaf litter, we replaced the litter with newly fallen leaf litter in January 2013. The collar for measurement of R_S was placed near the treatment area for R_{LL} measurement and the L-layer inside the collar was removed and leaf litter was supplied similarly as for measurement of R_{LL} . To prevent incorporation of newly fallen litter, we placed a mesh sheet (1 × 1 mm mesh) on the L-layer inside the chamber, and fallen litter was removed weekly. CO₂ efflux from combusted granite soil was measured 6 months from the start of the R_{LL} measurements. The mean CO₂ flux rate (\pm standard deviation) was 0.00063 \pm 0.00068 mg CO₂ m⁻² s⁻¹ ($n = 16$) when SWC ranged from 0.05 to 0.3 m³ m⁻³ at temperatures of 24°C. Thus, we assumed that CO₂ efflux from the combusted granite soil was negligible throughout the measurement period.

For continuous in situ measurement of LWC, we used capacitance sensors as described by Ataka et al. [22]. The measurements were performed on the top surface of the L-layer and at the boundary between the L-layer and mineral soil (Fig. 1B), to capture the large vertical distribution of WC within the L-layer. We estimated average LWC from the output voltage (V) of the two sensors using the conversion equation LWC = 12.73 V - 3.42 presented by Ataka et al. [22]. LWC at the forest floor shows spatial variability associated with tree canopy conditions. Thus, to reflect the LWC of the L-layer by direct measurement, two capacitance sensors were placed on the L-layer inside the chamber. To check the validity of continuous LWC monitoring, we compared the sensor values with LWC measured

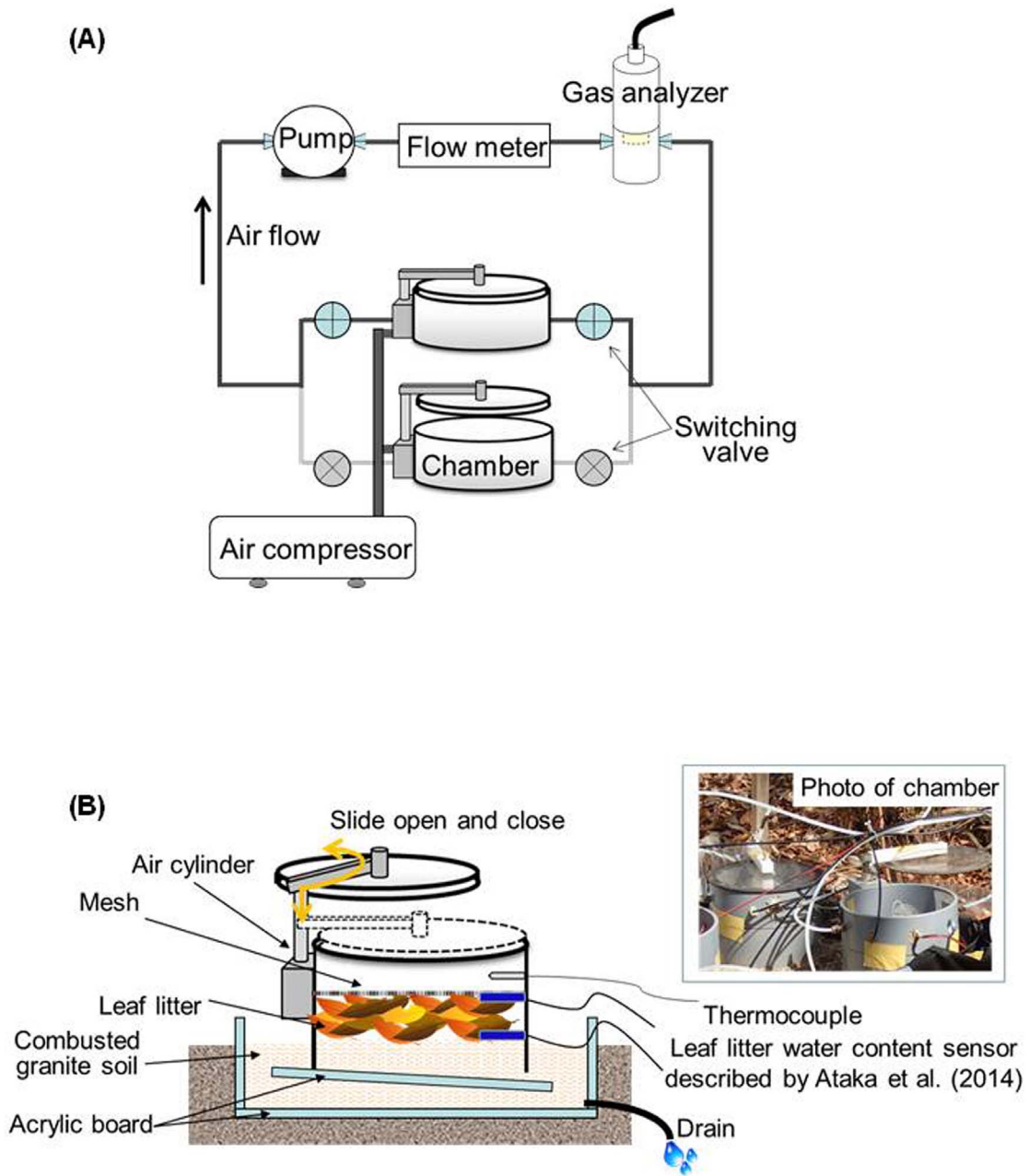


Figure 1. Schematic of the automated chamber system and the experimental design for measurement of CO₂ efflux from the leaf litter layer. **A.** Schematic of the automated dynamic-closed chamber system for measuring leaf litter respiration and soil respiration. **B.** The experimental design for continuous measurement of CO₂ efflux from the leaf litter layer only using automated chamber system. doi:10.1371/journal.pone.0108404.g001

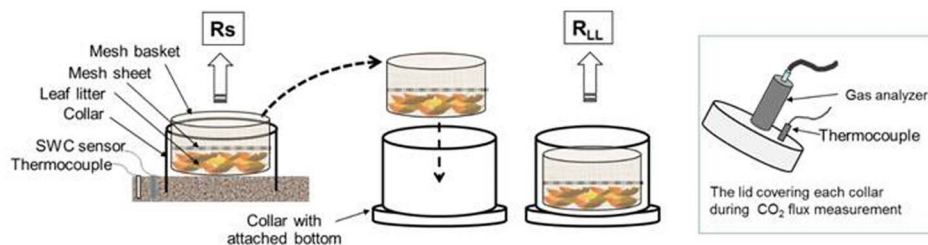


Figure 2. Schematic of the manual chamber system and the experimental design for measurement of CO₂ efflux from the leaf litter layer (R_{LL}) and soil (R_s). doi:10.1371/journal.pone.0108404.g002

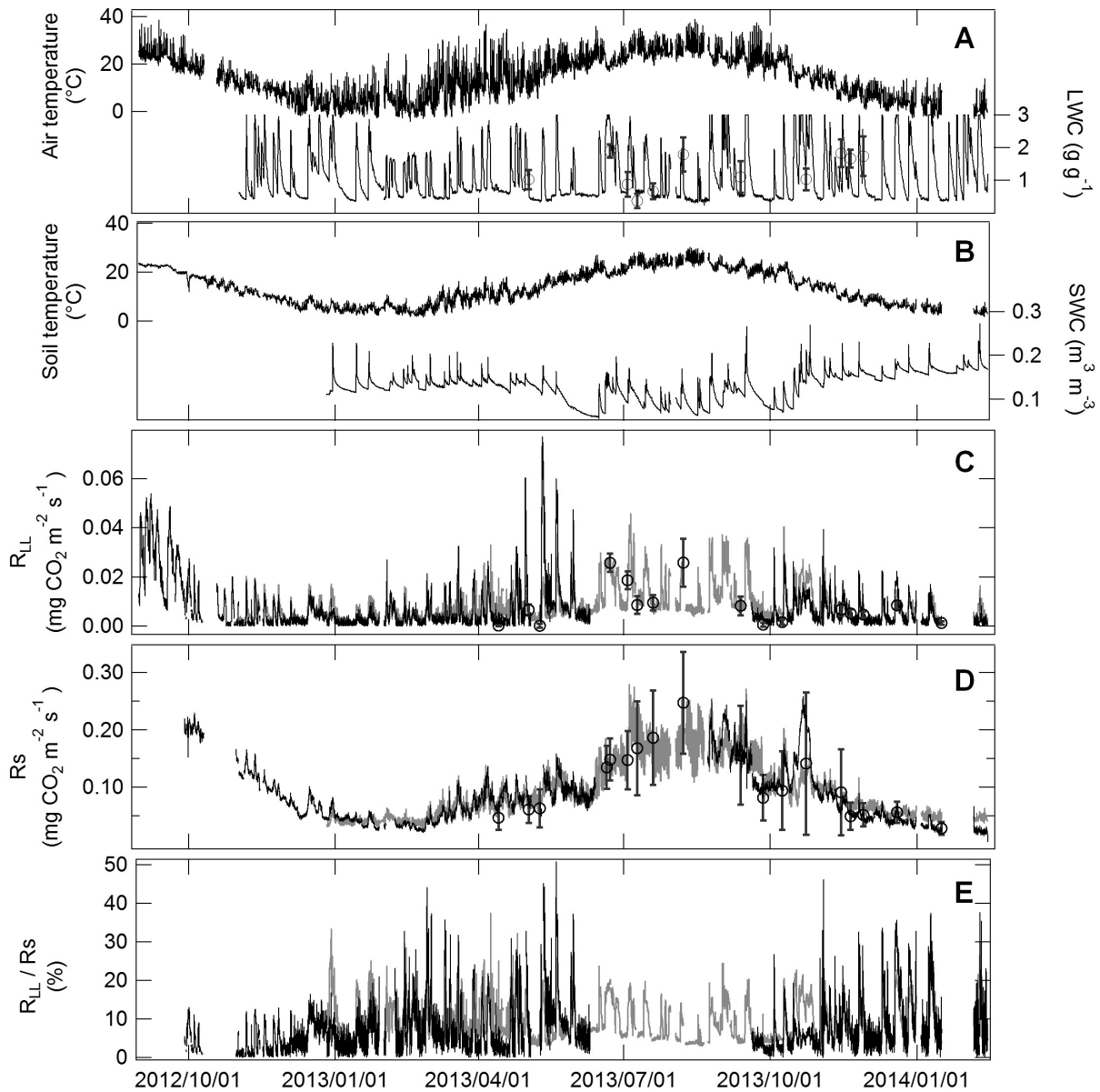


Figure 3. Seasonal variation in environmental factors, CO₂ efflux from the leaf litter layer (R_{LL}), and soil respiration (R_S). Data were measured every 30 min between September 2012 and January 2014. **A.** Bold and fine lines show air temperature and water content of the leaf litter layer (LWC), respectively. **B.** Bold and fine lines show soil temperature and soil water content (SWC), respectively. **C.** Black and grey lines show observed and estimated R_{LL} , respectively. **D.** Black and grey lines show observed and estimated R_S , respectively. Circles and bars show mean values and standard deviation of manual measurements. Estimated R_{LL} and R_S were calculated from regression equations using temperature (T) and water content (WC): $R_{LL} = 0.29e^{0.059T}[WC/(95.04+WC)]$ and $R_S = 0.031e^{0.10T}[WC/(0.032+WC)]$. doi:10.1371/journal.pone.0108404.g003

Table 1. Q_{10} of leaf litter respiration (R_{LL}) and soil respiration (R_S) for different water contents of the leaf litter layer (LWC) and soil (SWC).

	R_{LL}			R_S		
	LWC ≤ 1	1 < LWC ≤ 2	2 < LWC	SWC ≤ 0.1	0.1 < SWC ≤ 0.15	0.15 < SWC
Q₁₀	1.54	1.88	2.07	1.97	2.12	2.73
a	0.0019	0.0044	0.0064	0.027	0.032	0.025
b	0.043	0.063	0.073	0.068	0.075	0.10

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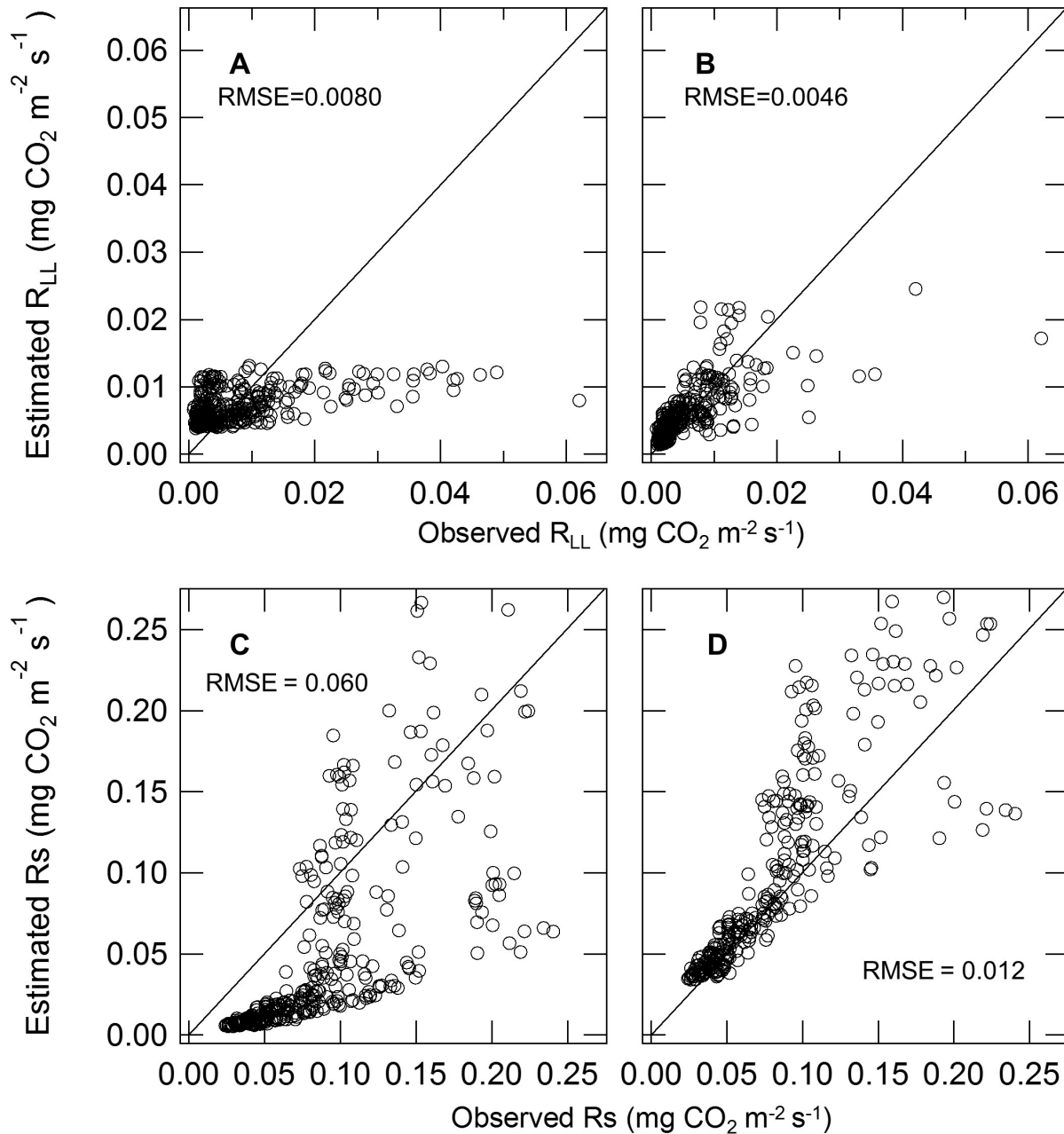


Figure 4. Relationship between observed and estimated CO₂ efflux rate from leaf litter respiration (R_{LL}) and soil respiration (R_S). R_{LL} (A, B) and R_S (C, D) show daily mean values. Estimated respiration rates were calculated using a function of temperature (A, C) from Eq. (5,6) and a function of temperature and water content (B, D) from Eq. (7,8) in the Results. Lines represent the 1:1 ratio. RMSE: root mean square error. doi:10.1371/journal.pone.0108404.g004

manually as described in the following section. In parallel with LWC measurement, soil temperature (copper-constantan thermocouple) and soil volumetric water content (ECH₂O EC-5 sensors; Decagon Devices, Pullman, WA, USA) were measured at 5-cm depth near each chamber. The output voltage of all environmental data was recorded every 1 min with a data logger (Datamark LS-3000 PtV; Hakusan, Japan) and average values were computed every 30 min. The environmental data, R_{LL} , and R_S were measured continuously between September 2012 and January 2014. Malfunction of IRGA resulted in a lack of data for R_{LL} and R_S for 31% of the measurements.

Manual chamber method for measuring leaf litter respiration and soil respiration

To determine the validity of R_{LL} and R_S measured using the automated chamber method, respiration was measured using the manual chamber method. We assumed that manual chamber method allow to measure under conditions that were closer to natural than the automated chamber method. We measured R_{LL} and R_S manually using a static chamber system at midday on 18 days between April 2013 and January 2014. Twelve PVC collars (320 cm² surface area) were placed in a 2×4 m area in January 2013. The edges of the collars were inserted approximately 1.5 cm into the soil. To measure R_{LL} , mesh baskets (1×1 mm mesh, the

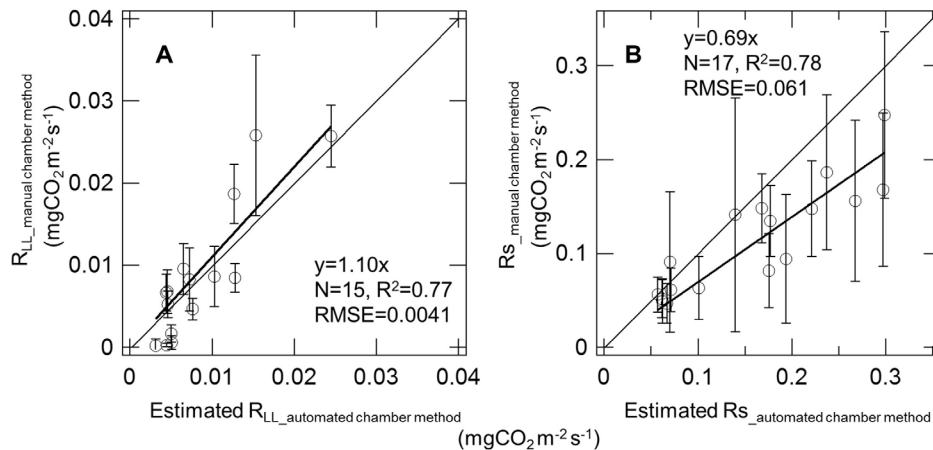


Figure 5. Relationship between respirations measured using a manual chamber method and estimated from automated chamber data. Respiration rate measured with the manual chamber method ($R_{LL_manual\ chamber\ method}$) show mean value obtained from measurement of 12 collars. Bars show standard deviation. Respiration estimated from automated chamber data (estimated $R_{LL_automated\ chamber\ method}$) shows daily mean respiration. The estimated R was calculated using a function based on temperature and water content (Eq. 8, 9). doi:10.1371/journal.pone.0108404.g005

same diameter as the PVC collars; 20 cm) were set into each collar and 15 g (dry weight) of newly fallen leaf litter was placed on the L-layer inside each basket (Fig. 2). To prevent supply of newly fallen litter, we placed a mesh sheet (1 × 1 mm mesh) on the L-layer inside the chamber, and fallen litter was removed weekly.

For measurement of R_S , the collars were completely covered with lids to which an IRGA and copper-constantan thermocouple were attached. Soil temperature and SWC (5 cm depth) were measured close to the collars when R_S was measured. After completing the measurements of R_S , the mesh baskets were carefully removed from the collars and placed in PVC chambers (20 cm diameter, 7 cm high; Fig. 2). We measured R_{LL} using the same methods as used for R_S measurement. The temperature and CO₂ concentrations in the chamber were recorded at 1-s intervals using a data logger (GL220). Linearity of the CO₂ flux was checked on the data logger monitor at each measurement. The measurement period for each chamber was 10 min and CO₂ data for the middle 5-min intervals were used to determine R_{LL} according to Eq. (1), excluding data from the first 3 min.

For measurement of LWC in the mesh baskets, four or five leaves were removed from each basket and immediately placed in sealed plastic bags. Fresh weight of the leaf litter was measured in the laboratory within 24 h of sampling. Leaf litter samples were oven dried at 65°C for 48 h, and water content (WC; g g⁻¹) was calculated using Eq. 2 as follows:

$$WC = \frac{(FW - DW)}{DW}, \quad (2)$$

where FW is the fresh mass of the sample (g), and DW is the dry mass of the sample (g). Samples were returned to each mesh basket within 1 week after sampling.

Leaf litter respiration and soil respiration rates as a function of environmental factors

Respiration models are fundamentally described by nonlinear functions. We used the following function to investigate the response of respiration to temperature:

$$R = a \exp(bT), \quad (3)$$

where T is temperature (leaf litter temperature for R_{LL} measurement or soil temperature for R_S measurement) and a and b are constants. Leaf litter temperature was assumed to be same as air temperature. b is related to the Q_{10} parameter ($Q_{10} = e^{10b}$). To determine the effects of temperature and water content on R_{LL} and R_S , we used a function that was previously applied to estimate soil respiration by Subke and Schlesinger [26]:

$$R = a \exp(bT) \left(\frac{WC}{c + WC} \right), \quad (4)$$

where a , b , and c are constants. LWC or SWC was used as WC in this equation. These nonlinear regressions were performed using a modified Levenberg–Marquardt method with Igor Pro 6.0 software (WaveMetrics, Lake Oswego, OR, USA). The estimated respiration values presented in this manuscript were calculated using Eq. 4.

Short-term changes in R_{LL} and LWC on wetting and drying cycle

To evaluate short-term changes in R_{LL} and LWC after rainfall events, we chose eight typical periods that included one wetting and drying cycle and had consecutive no rainfall days for at least 3 days. We used daily mean R_{LL} and LWC before the day on which precipitation occurred as the pre-wetting condition, and these values after precipitation as the post-wetting condition. Daily mean R_{LL} was calculated from R_{LL} values observed using the automated chamber method.

Effect of wetting and drying cycle of the L-layer on R_{LL} and R_S on the annual time scale

To investigate the effects of wetting and drying of the L-layer on R_{LL} on the annual time scale, we separated the estimated daily mean R_{LL} in 2013 into ‘Dry’ and ‘Wet’ periods based on daily mean LWC as a threshold value. The threshold LWC value that separated ‘Dry’ and ‘Wet’ periods for R_{LL} was estimated by the abovementioned short-term analyses. Daily mean R_{LL} was calculated from the estimated R_{LL} values because there were gaps in the continuous R_{LL} data observed using the automated

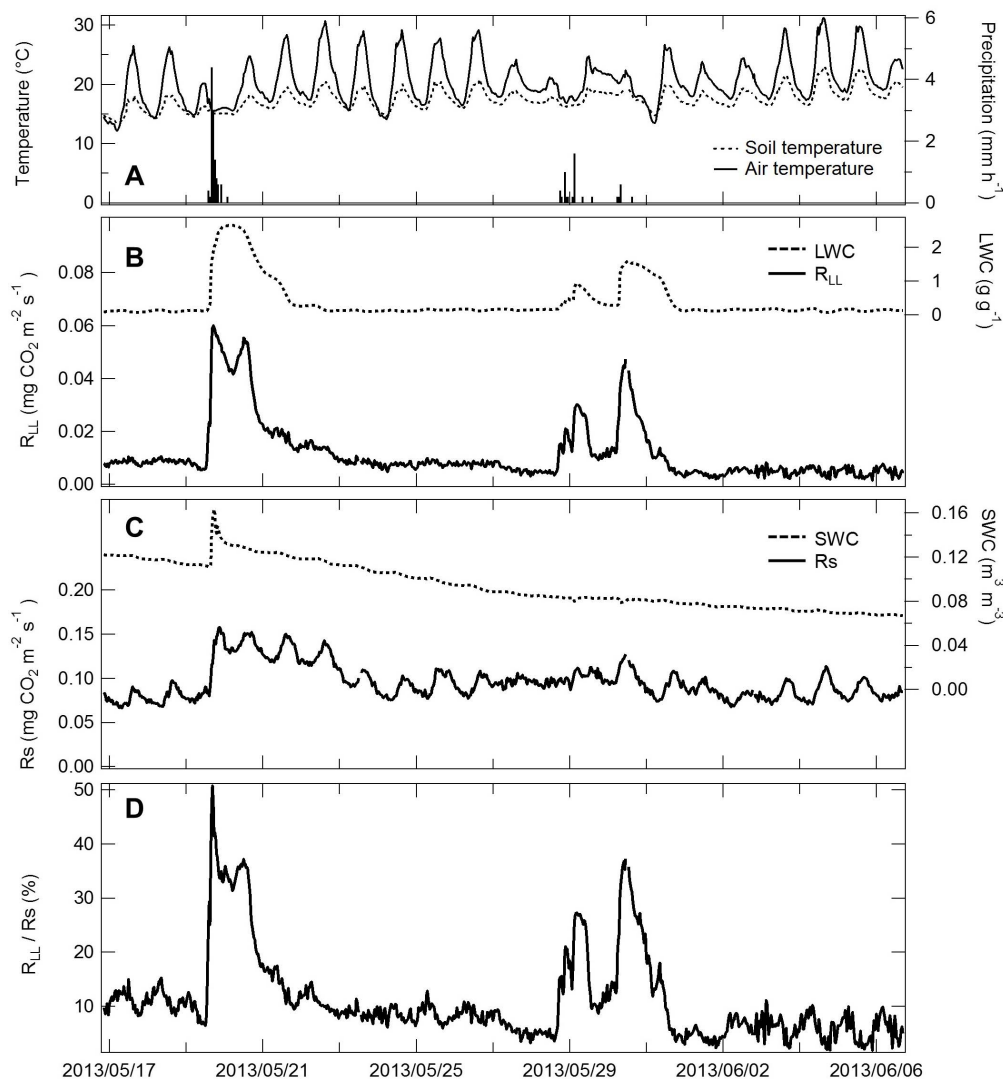


Figure 6. Temporal variation in environmental factors, CO₂ efflux from the leaf litter layer (R_{LL}), soil respiration (R_S), and the ratio of R_{LL} to R_S . Data was measured at one collar every 30 min between May 17 and June 6, 2013. **A.** Soil and air temperature. Spikes on the x-axis indicate precipitation events (mm h^{-1}). **B.** R_{LL} and water content of the leaf litter layer (LWC). **C.** R_S and soil water content (SWC). **D.** The ratio of R_{LL} to R_S (%). doi:10.1371/journal.pone.0108404.g006

chambers. We estimated the contribution of R_{LL} accumulated during the wet and dry period to total R_S .

Results

Seasonal variation in R_{LL} and R_S

The magnitude of the peak in the observed R_{LL} pulse was higher in summer than in winter (Fig. 3C). R_{LL} values were low when LWC was low (Fig. 3A, C). R_S changed substantially according to temperature (Fig. 3B, D), with higher values in summer than in winter. The relationships between respiration and temperature were described by the following functions:

$$R_{LL}(\text{mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}) = 0.0038 \exp(0.065 \times T_{LL}), \quad (5)$$

$$R_S(\text{mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}) = 0.0031 \exp(0.19 \times T_S), \quad (6)$$

where T_{LL} is leaf litter temperature and T_S is soil temperature ($^{\circ}\text{C}$).

To evaluate effect of WC on the temperature sensitivity of respiration, the measured respiration data was separated into three groups based on WC (Table 1). More than 14% of total respiration data was included in each WC group. R_{LL} showed low values when WC values were low in spite of high temperature. Consequently, calculated Q_{10} values for not only R_{LL} but also R_S decreased with decreasing WC. The relationships between respiration and temperature and WC were described by the following functions:

$$R_{LL}(\text{mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}) = 0.29 \exp(0.059 \times T_{LL}) \left(\frac{LWC}{95.04 + LWC} \right), \quad (7)$$

$$R_S(\text{mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}) = 0.031 \exp(0.10 \times T_S) \left(\frac{SWC}{0.032 + SWC} \right), \quad (8)$$

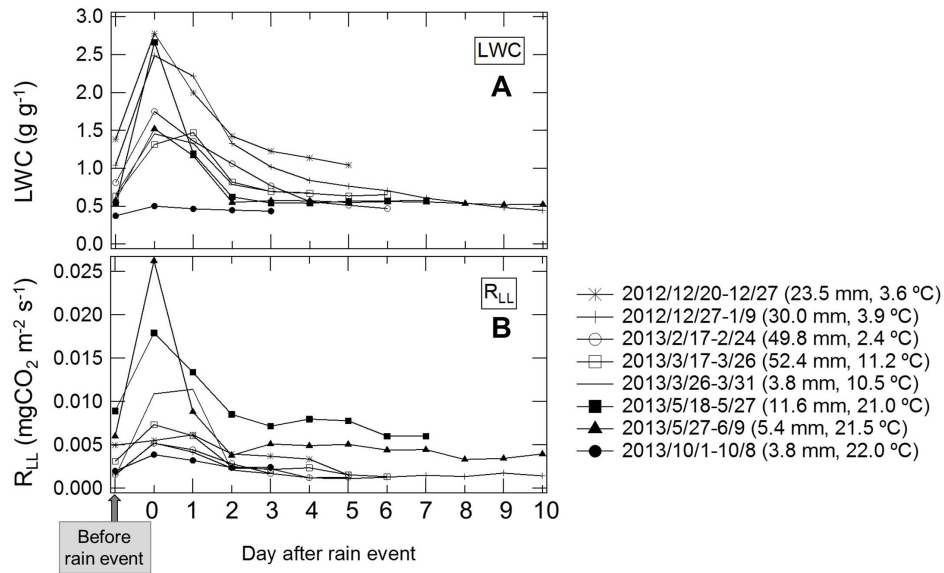


Figure 7. Temporal variation in water content of the leaf litter layer (LWC) and CO₂ efflux from the leaf litter layer (R_{LL}) after rainfall events. LWC (A) and R_{LL} (B) show the daily mean values. The rainfall intensity of each precipitation event was 23.5 mm in 2 days (2012/12/20–12/27, mean air temperature; 3.6 °C); 30.0 mm in 3 days (2012/12/27–1/9, 3.9 °C); 49.8 mm in 2 days (2013/2/17–2/24, 2.4 °C); 52.4 mm in 3 days (2013/3/17–3/26, 11.2 °C); 3.8 mm in 2 days (2013/3/26–3/31, 10.5 °C); 11.6 mm in 2 days (2013/5/18–5/27, 21.0 °C); 5.4 mm in 3 days (2013/5/27–6/9, 21.5 °C); and 3.8 mm in 4 days (2013/10/1–10/8, 22.0 °C). doi:10.1371/journal.pone.0108404.g007

where LWC (g g^{-1}) and SWC ($\text{m}^3 \text{m}^{-3}$) are water content of leaf litter and soil, respectively. The RMSE between observed and estimated daily mean respiration based on temperature (R_{LL} , $0.0080 \text{ mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}$; R_S , $0.060 \text{ mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}$) was larger than that based on temperature and WC (R_{LL} , $0.0046 \text{ mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}$; R_S , $0.012 \text{ mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}$) (Fig. 4). Estimated respiration was calculated using the equation based on temperature and WC because of the lower RMSE. Throughout the measurement period, the contribution of observed R_{LL} to variation in R_S changed from nearly zero to 51% following a rainfall event (Fig. 3E).

To consider the validity of R_{LL} and R_S estimated from continuous measurement, we compared these values with respiration rates measured using the manual chamber method (Fig. 5). Estimated respiration was very similar to that observed using manual measurements. The RMSE between estimated and observed respiration were 0.0041 and $0.061 \text{ mg CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ for R_{LL} and R_S , respectively.

Temporal changes in R_{LL} and R_S on the short-term scale

To show clear temporal variation in R_{LL} and R_S , the period between May 17 and June 6, 2013 (Fig. 6) was chosen because this

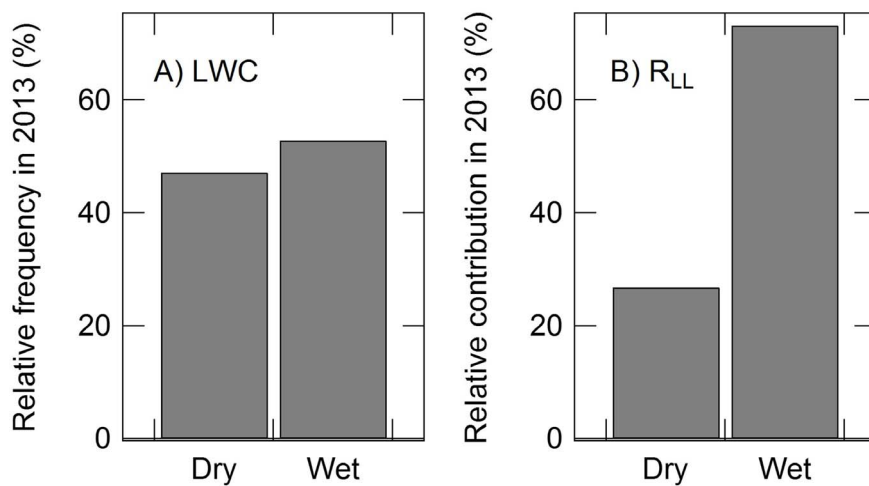


Figure 8. Histograms of the relative frequency of “Dry” and “Wet” periods in relation to water content of the leaf litter layer (LWC), and the relative contribution of estimated leaf litter respiration (R_{LL}) in 2013. The daily mean LWC (A) and R_{LL} (B) were used to present histograms. Estimated respiration rates were calculated using a function based on temperature (T) and water content (WC). $R_{LL} = 0.29e^{0.059T}[\text{WC}/(95.04+\text{WC})]$. The daily mean LWC and R_{LL} were defined as Dry or Wet based on LWC. Days in which daily mean LWC $< 0.75 \text{ g g}^{-1}$ were defined as Dry periods, while days in which daily mean LWC $\geq 0.75 \text{ g g}^{-1}$ were defined as Wet periods. doi:10.1371/journal.pone.0108404.g008

period included two characteristic rainfall events. The rainfall intensity was 11.6 mm over 13 h during the first event and 5.4 mm over 46 h during the second event. LWC and SWC increased from 0.11 to 2.64 g g⁻¹ and from 0.11 to 0.16 m³ m⁻³, respectively, following the first rainfall event (Fig. 6B, C). LWC increased from 0.16 to 1.58 g g⁻¹ but SWC did not increase after the second rainfall event.

Temporal variation in R_{LL} measured using the automated chamber system changed according to wetting and drying of the L-layer (Fig. 6B), reaching a maximum of 0.060 and 0.047 mg CO₂ m⁻² s⁻¹ during first and second rainfall events, respectively. R_S increased following the increase in SWC and subsequently decreased gradually with diurnal variation according to temperature (Fig. 6C). Between May 17 and June 6, 2013, the contribution of R_{LL} to R_S increased from 6.5% to 51%, with a peak value of 51% during the first rainfall event and 37% during the second rainfall event (Fig. 6D).

Both R_{LL} and LWC reached a peak during or one day after rainfall events (Fig. 7). The peak of R_{LL} and LWC varied from 0.0020 to 0.026 mg CO₂ m⁻² s⁻¹ and from 0.50 to 2.66 g g⁻¹, respectively. Peak value of each rainfall event highly depended on air temperature. High peaks of R_{LL} were observed in the warm season (0.017 mg CO₂ m⁻² s⁻¹; 2013/5/18–5/27, 0.026 mg CO₂ m⁻² s⁻¹; 2013/5/27–6/9 in Fig. 7). Also, the peak value was related to LWC: low peak of R_{LL} was observed when LWC was low (0.004 mg CO₂ m⁻² s⁻¹; 2013/10/1–10/8 in Fig. 7). The relationship between LWC and amount of precipitation was not clear. In the cold season, peak values of R_{LL} were relatively low (e.g., 0.005 mg CO₂ m⁻² s⁻¹; 2013/2/17–2/24, 0.006 mg CO₂ m⁻² s⁻¹; 2012/12/20–12/27 in Fig. 7) even when the L-layer was wet enough (LWC more than 1.5 g g⁻¹). The peak values of R_{LL} were 1.2- to 8.6-fold higher than the R_{LL} values before rainfall events, and R_{LL} fell to pre-wetting levels within 2–4 days after rainfall events and peak LWC values were 1.3- to five-fold higher than LWC before rainfall, and LWC also dropped to pre-wetting levels within 2–4 days after rainfall events. We defined R_{LL} from the period just after rainfall events through 2–4 days later as the “ R_{LL} pulse”.

Effects of wetting and drying of the L-layer on R_{LL} and R_S on the annual time scale

Estimated daily mean R_{LL} in 2013 was separated into ‘Dry’ and ‘Wet’ periods based on daily mean LWC. Days for which mean LWC was <0.75 g g⁻¹ were categorized as Dry, while days for which mean LWC ≥0.75 g g⁻¹ were categorized as Wet. The threshold value (0.75 g g⁻¹) was obtained from mean LWC 3 days after a rainfall event (Fig. 7A). The relative frequency of Dry and Wet periods in 2013 were 47.2% and 52.8%, respectively, while the relative contributions of daily mean R_{LL} during the Dry and Wet periods in 2013 were 26.9% and 73.2%, respectively (Fig. 8). Annual R_{LL} and R_S in 2013 were estimated to be 0.69 and 7.94 t C ha⁻¹ y⁻¹, respectively. The RMSE between continuous respiration measured and estimated based on temperature and WC was 0.011 and 0.029 t C ha⁻¹ y⁻¹, respectively.

The contribution of annual R_{LL} to R_S was 8.6%. The relative frequency of LWC was similar during Dry and Wet periods, while the contribution of R_{LL} during the Wet period was approximately three-fold higher than that during the Dry period (Fig. 8).

Discussion

As seen in Fig. 6, R_{LL} immediately increased with wetting of the L-layer and decreased to pre-wetting levels within 2–4 days after rainfall events, which was consistent with observations made in

previous studies [17,19]. R_{LL} showed no diurnal variation despite a diurnal temperature range >10°C. Consequently, the Q_{10} of R_{LL} increased with increasing LWC (Table. 1). The variation in Q_{10} would be directly related to water stress experienced by microorganism. This indicated that LWC can reach to adequate low value, suspected as water stress for microorganism, within several days after rainfall. On the one hand, R_S increased during rainfall and subsequently decreased, showing diurnal variation. The Q_{10} of R_S also increased with increasing SWC. Dannoura et al. [27] reported that root respiration showed little change with variation in SWC compared with changes in R_S . Therefore, the increased Q_{10} of R_S with increasing SWC might be highly affected by not only R_{LL} but also by respiration from other heterotrophic sources.

Although the relative frequency of LWC was similar during Dry and Wet periods, the contribution of annual R_{LL} during the Wet period was approximately three-fold higher than that during the Dry period (Fig. 8), indicating strong effect of rainfall on R_{LL} . Although the R_{LL} pulse can last for only 3–4 days after a rainfall event, this pulse would determine a large part of annual R_{LL} . This suggests that the magnitude of total R_{LL} may be influenced by the frequency of rainfall events, especially in summertime, rather than the intensity of rainfall. Still, the cumulative R_{LL} in the Dry period contributed 26.9% of annual R_{LL} in 2013, even though instantaneous R_{LL} was very low. There may be large vertical variability in WC and R_{LL} within the L-layer, indicating that higher WC and R_{LL} occur in lower parts of the L-layer during the drying process because the upper L-layer dries more rapidly [28]. In that case, although the mean WC of the L-layer was very low, local wetting in lower sections would produce small CO₂ fluxes. Despite low instantaneous R_{LL} , the accumulation of R_{LL} over a long time period (approximately 6 mo) resulted in a substantial contribution (27%) of Dry-period respiration to annual R_S .

Raindrops first reach the L-layer and then percolate to the soil layers below. Small amounts of precipitation caused no change in SWC or R_S , but R_{LL} increased rapidly with increasing LWC (Fig. 6). In semi-arid and arid ecosystems, wetting of the L-layer and surface soil by small fog-drop pulses during the dry season can contribute up to 35% of R_S [29]. Although such small water inputs (e.g., brief rain showers and fog), which mainly affect the surface of the forest floor, can be significant drivers of temporal variation in R_S , the soil water content sensors (generally inserted at depths > 5 cm) could not capture these inputs. Continuous measurement of LWC allowed for realistic modeling of the effects of rapid changes in LWC on R_{LL} .

Although the annual contribution of R_{LL} to R_S was relatively small (8.6%), this contribution showed large temporal variation according to rainfall, ranging from nearly zero to 51%. Several other studies have described similar results [17,21]. For example, Borken et al. [17] reported that peaks in R_{LL} during addition of water ranged from 0.031 to 0.071 mg CO₂ m⁻² s⁻¹ in vitro, which represented 11–26% of maximum in situ R_S in the Harvard forest, although R_{LL} before addition of water was nearly zero. These findings indicate that R_{LL} is a significant component of rapid and transient temporal variation in R_S in relation to rainfall events. Although numerous studies have examined CO₂ efflux from mineral soils in relation to the intensity, duration, and frequency of rainfall [30,31], few studies have focused on R_{LL} because of the difficulty in measuring this dynamic. Here, R_{LL} pulses were observed only during and several days after rainfall events. Thus, periodic sampling (e.g., twice per week) might be insufficient to capture the contribution of the R_{LL} pulse to R_S . Moreover, manual flux measurements are usually not performed during precipitation events because of difficulties that can occur

with electronic instruments and sampling methods. In our view, conducting in situ measurements of CO₂ efflux from the L-layer only over short time intervals (e.g., up to 1 h) produces robust data for understanding the response of R_{LL} to rainfall events and its contribution to R_S .

The contribution of R_{LL} to annual R_S was 8.6% in our site. In an oak forest, the contribution of R_{LL} to R_S was 23%, according to model simulation based on temperature and LWC by Hanson et al. [13]. Ngao et al. [32] reported a lower contribution (8%) in a beech forest, estimated using an isotope mass balance approach, which was close to the value observed at our site (8.6%). However, simple quantitative comparisons between studies are difficult because of the use of different methods. In addition, some technical problems remain at our site. First, we performed R_{LL} measurements in the treatment area in which the mineral soil below the L-layer was replaced with combusted granite soil. This treatment may have affected the microbial community and environmental conditions in the L-layer. Secondly, each continuous measurement of R_{LL} and R_S was performed with single chambers, so spatial heterogeneity in R_{LL} and R_S were not considered. Automated chamber methods allowed high-interval measurements of temporal variation in respiration but had poorer spatial distribution compared with the manual chamber method. The balance of trade-offs between automated and manual chamber method is subject to the relative importance of characterizing temporal and spatial variability of individual CO₂ sources. The number of chambers used can enhance the accuracy of measured mean values. Loescher et al. [33] reported that the number of chambers needs to be >100 to adequately represent spatial variability. However, this is not a feasible experimental design because of practical limitations to sampling efforts. To improve estimation of R_{LL} and R_S at the forest stand level, and to better understand the soil carbon budget, a comprehensive comparison of the diverse C pools and fluxes in forest soils is required.

Conclusions

In our study, the rapid and transient variation in R_{LL} induced by rainfall; the peak R_{LL} was observed during or one day after rainfall, and R_{LL} subsequently decreased to pre-wetting levels

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within 2–4 days after rainfall events, following the decrease in LWC. On the one hand, CO₂ efflux from coarse woody debris found in our site decreased during rainfall events, and subsequently, a gradual increase in CO₂ efflux continued for at least 14 days until next rainfall [34]. Therefore, coarse woody debris was a CO₂ efflux source over longer time scales, while R_{LL} approached nearly zero within a few days after rainfall events, even at high temperatures. Such specific temporal CO₂ efflux patterns for each heterotrophic source when subjected to wetting and drying cycles would be a result of substrate properties (e.g., specific surface area). In our view, continuous and direct measurements of CO₂ efflux and environmental conditions characterized by substrate properties of individual CO₂ sources could improve understanding of the processes that regulate variation in heterotrophic respiration and R_S and enable progress beyond empirical models that are primarily based on simple temperature and SWC relationships.

Moreover, the magnitude of heterotrophic respiration under wetting and drying cycles is strongly related to microbial physiology and community composition. For example, Schnurer et al. [35] showed that longer-duration wetting could promote microbial biomass, causing an increase in basal respiration. Fierer et al. [36] showed the influence of drying and rewetting frequency on microbial (fungi and bacteria) community composition. To improve understanding of heterotrophic respiration associated with response and adaptation of microorganisms under climatic changes, collected continuous in situ data for CO₂ efflux and environmental conditions (e.g., temperature and WC) of individual CO₂ sources should be combined with analyses of microbial physiology and community composition.

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Author Contributions

Conceived and designed the experiments: MA YK TM. Performed the experiments: MA KY MT. Analyzed the data: MA YK MT. Contributed reagents/materials/analysis tools: MA MJ. Contributed to the writing of the manuscript: MA.

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