



Article Asymmetric Synthesis of Contact Sex Pheromone of *Tetropium fuscum* and Its Enantiomer

Xueyang Wang¹, Jianan Wang¹, Fengbo Ma², Qinghua Bian¹, Min Wang¹ and Jiangchun Zhong^{1,*}

- ¹ Department of Applied Chemistry, China Agricultural University, 2 West Yuanmingyuan Road, Beijing 100193, China
- ² College of Enviromental Sciences, Sichuan Agricultural University, 211 Huimin Road, Chengdu 611130, China
- Correspondence: zhong@cau.edu.cn; Tel.: +86-010-6273-1356

Abstract: *Tetropium fuscum* is a harmful forest pest and attacks spruces. The contact sex pheromone of this pest, (*S*)-11-methyl-heptacosane, and its enantiomer were synthesized via Evans' chiral auxiliaries. The key steps of this approach included acylation of carboxylic acid, diastereoselective methylation of oxazolidinone amide, and Wittig coupling of the aldehyde with chiral phosphonium salt. The synthetic pheromones would have potential utility in the control of this pest.

Keywords: contact sex pheromone; Tetropium fuscum; chiral auxiliary; asymmetric synthesis

1. Introduction

The brown spruce longhorn beetle, *Tetropium fuscum* Fabricius (Coleoptera: Cerambycidae), is a harmful forest pest native to Europe where it attacks weakened Norway spruce, *Picea abies* L. [1,2]. It first became invasive to North America around 1990 [3,4], where it was infecting and killing healthy native spruces, such as red spruce (*Picea rubens* Sargent), white spruce (*Picea glauca* Moench), blue spruce (*Picea pungens* Engelm), and black spruce (*Picea mariana* Miller) [5,6]. Due to its subcortical feeding habits [7,8], systemic insecticides and heating are not very effective and practical [9,10].

The strategy based on pheromones for controlling agricultural pests is one of the most promising, effective, and safe solutions [11,12]. The contact sex pheromone of *Tetropium fuscum* was identified as (*S*)-11-methyl-heptacosane ((*S*)-1) (Figure 1) by Silk, meanwhile, (*S*)-1 and its enantiomer (*R*)-1 were synthesized from (*S*)- and (*R*)-citronellyl bromides [13]. To study future utilization of the contact sex pheromone [14], herein, we prepared the contact sex pheromone of *Tetropium fuscum* and its enantiomer using Evans' chiral auxiliaries. Our synthesis was easily performed and afforded the target pheromone with high enantiomeric purity.



Figure 1. Contact sex pheromone of Tetropium fuscum and its enantiomer.

2. Results and Discussion

2.1. Retrosynthetic Analysis

In view of retrosynthetic analysis of contact sex pheromone of (S)-1 (Scheme 1), the key step is to construct the chiral center. It was envisaged that Evans' chiral auxiliaries including acylation of dodecanoic acid (2) and diastereoselective methylation of oxazolidinone amide would introduce chiral methyl of amide (S,S)-5. The target pheromone (S)-1 could be synthesized via hydrogenation reduction of olefin (S)-10, which could be divided into two



Citation: Wang, X.; Wang, J.; Ma, F.; Bian, Q.; Wang, M.; Zhong, J. Asymmetric Synthesis of Contact Sex Pheromone of *Tetropium fuscum* and Its Enantiomer. *Molecules* **2022**, *27*, 6897. https://doi.org/10.3390/ molecules27206897

Academic Editor: Yu Peng

Received: 26 September 2022 Accepted: 12 October 2022 Published: 14 October 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). components, pentadecanal (9) and phosphonium salt in situ prepared by hydrocarbon bromide (*S*)-7 and triphenylphosphine. Furthermore, (*S*)-1-bromo-2-methyldodecane ((*S*)-7) could be easily prepared from chiral alcohol (*S*)-6 through Appel reaction.



Scheme 1. Retrosynthetic analysis of contact sex pheromone (S)-1.

2.2. Synthesis of Chiral Primary Bromides

Based on the retrosynthetic analysis of contact sex pheromone (*S*)-**1**, chiral primary bromides (*S*)- and (*R*)-7 were first prepared (Scheme 2). The reaction of dodecanoic acid (**2**) with oxalyl chloride afforded the corresponding crude acyl chloride [15] were then treated with oxazolidinone (*S*)-**3** and NaH to provide (*S*)-4-benzyl-3-dodecanoyloxazolidin-2-one ((*S*)-**4**) in a 96% yield [16]. In the presence of hexamethyldisilazide (NaHMDS), diastereoselective methylation of amide (*S*)-**4** with methyl iodide gave chiral methyl amide (*S*,*S*)-**5** (62% yield, dr > 99:1, determined by ¹³C NMR spectra) [17]. According to Evans' chiral auxiliary precedent [18], the absolute configuration of the new stereocenter of (*S*,*S*)-**5** was assigned as (*S*). The subsequent reduction with NaBH₄ afforded (*S*)-2-methyldodecan-1-ol ((*S*)-**6**) in an 88% yield [19,20]. Its specific rotation {[α]_D²² = -9.0 (*c* 1.37, CHCl₃)} was identical to the literature [21] value {[α]_D²³ = -8.4 (*c* 1.00, CHCl₃)}, which also supported the (*S*)-configuration of the new stereocenter. The final Appel reaction converted the chiral methyl alcohol (*S*)-**6** to (*S*)-1-bromo-2-methyldodecane ((*S*)-**7**) in a 99% yield [22,23]. Similarly, (*R*)-1-bromo-2methyldodecane ((*R*)-**7**) was prepared via acylation, diastereoselective methylation, reduction, and bromination from dodecanoic acid (**2**) and (*R*)-4-benzyloxazolidin-2-one ((*R*)-**3**).



Scheme 2. Synthesis of chiral primary bromides (S)- and (R)-7.

2.3. Research on the Enantiomeric Purity of Chiral Alcohols

To explore the optical purity of key intermediate chiral methyl alcohols (*S*)-**6** and (*R*)-**6**, we synthesized their MBT derivatives (Scheme 3). According to the similar sequence of (*S*)-**6**, the racemic alcohol **6** was prepared from dodecanoic acid and oxazolidin-2-one. Then, the Mitsunobu reaction of methyl alcohols **6** with benzo[*d*]thiazole-2(*3H*)-thione (MBT) afforded their MBT derivatives **8** [24,25]. Ee of (*S*)-**8** and (*R*)-**8** was 98–99% determined by HPLC with a Daciel Chiralcel OD-H column, which indicated that the enantiomeric purity of (*S*)-**6** and (*R*)-**6** were also 98–99%.



Scheme 3. Synthesis of MBT derivatives of alcohols 6.

2.4. Synthesis of the Target Compounds

With chiral primary bromides in hand, we focused on the synthesis of the target pheromone (*S*)-1 and its enantiomer (*R*)-1 (Scheme 4). The *Z*/*E* mixtures of olefin (*S*)-10 were achieved through Wittig coupling of *n*-pentadecanal (9) and phosphonium salt (53% yield, *Z*:*E* = 5.6:1, determined by ¹³C NMR spectra) [26,27], which was prepared in situ from hydrocarbon bromide (*S*)-7 and triphenylphosphine[28]. The final palladium-catalyzed hydrogenation of (*S*)-10 afforded the target compound (*S*)-11-methylheptacosane ((*S*)-1) [29,30]. The NMR spectrum and specific rotation of (*S*)-1 were matched with the literature [13]. Using the similar approach of pheromone (*S*)-1, we synthesized its enantiomer,

(*R*)-11-methylheptacosane ((*R*)-1). Moreover, the structure of (*R*)-1 was characterized with ¹H NMR, ¹³C NMR, and EIMS spectra, which were consistent with reference 13.



Scheme 4. Synthesis of contact sex pheromone (*S*)-1 and its enantiomer (*R*)-1.

3. Materials and Methods

3.1. General Information

All reactions were performed in a Schlenk system under an argon atmosphere unless otherwise indicated. All commercial reagents were used directly, whereas solvents were purified following the standard strategies before use. Polarimetric measurements were taken on a Perkin–Elmer PE-341 polarimeter. Enantiomeric excesses were determined by an Agilent 1200 HPLC system with a Daicel Chiralcel OD-H column with the eluents of *n*-hexane and isopropanol. ¹H and ¹³C NMR spectra were recorded on a Bruker DP-X500 MHz spectrometer. Chemical shifts were reported in ppm relative to tetramethylsilane for ¹H NMR and CDCl₃ (77.16 ppm) for ¹³C NMR. High resolution mass spectra were collected on Waters LCT Premier[™] with an ESI mass spectrometer. Low-resolution mass spectra were obtained from an Exactive GC-MS (EI).

3.2. Synthesis of (S)-4-Benzyl-3-dodecanoyloxazolidin-2-one ((S)-4) (CAS 198649-20-6)

The catalytic amount of DMF was added to a stirred solution of dodecanoic acid (2) (5.00 g, 24.97 mmol) in DCM (40 mL) at 0 °C. Oxalyl chloride (4.75 g, 37.43 mmol) was then added dropwise, and the reaction mixture was stirred for 1 h at 0 °C. After being warmed to room temperature and maintained for another 1 h, the solvent was removed under reduced pressure. The crude dodecanoyl chloride (3.49 g, 64% yield) was obtained as a slight yellow solid.

NaH (0.99 g, 60% in mineral oil, 24.75 mmol) was added in portions to a stirred solution of (S)-4-benzyloxazolidin-2-one ((S)-3) (2.95 g, 16.64 mmol, >99% ee) in THF (20 mL) at 0 °C. The resulting mixture was warmed to room temperature and stirred for 2 h, followed by the addition of the crude dodecanoyl chloride. The reaction mixture was maintained for another 3 h at the same temperature, then quenched with saturated aqueous NH₄Cl (10 mL). After the layers were separated, the aqueous phase was extracted with EtOAc (3 \times 50 mL). The EtOAc extracts were combined with organic layer, washed with saturated aqueous NaCl (50 mL), dried over anhydrous Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the residue was purified by silica gel column chromatography (EtOAc/petroleum ether 2:8) to afford (S)-4-benzyl-3-dodecanoyloxazolidin-2-one ((S)-4) as a colorless oil (5.73 g, 96% yield). $[\alpha]_D^{22} = +32.8$ (c = 2.67, CHCl₃). ¹H NMR (500 MHz, CDCl₃) § 7.33–7.20 (m, 5H), 4.67 (ddt, J = 10.7, 7.3, 3.2 Hz, 1H), 4.20–4.14 (m, 2H), 3.29 (dd, *J* = 13.4, 3.2 Hz, 1H), 3.00–2.86 (m, 2H), 2.77 (dd, *J* = 13.4, 9.6 Hz, 1H), 1.72–1.66 (m, 2H), 1.38–1.23 (m, 16H), 0.88 (t, J = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 173.56, 153.55, 135.44, 129.51, 129.01, 127.39, 66.23, 55.22, 38.00, 35.62, 31.99, 29.70, 29.57, 29.48, 29.42, 29.22, 24.37, 22.77, 14.19; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) *m/z* calcd. for C₂₂H₃₃NO₃Na (M + Na)⁺: 382.2353, found 382.2354.

3.3. Synthesis of (S)-4-Benzyl-3-((S)-2-methyldodecanoyl)oxazolidin-2-one ((S,S)-5) (CAS 2771300-96-8)

NaHMDS (13 mL, 2.0 M in THF, 26.00 mmol) was added to a stirred solution of oxazolidinone amide 4 (6.25 g, 17.38 mmol) in dry THF (60 mL) at -78 °C over 15 min via syringe pump. The resulting mixture was stirred for 1 h at -78 °C, followed by slow addition of MeI (12.34 g, 86.93 mmol). The reaction mixture was maintained for 2 h at the same temperature, then quenched with saturated aqueous NH_4Cl (20 mL). After the layers were separated, the aqueous phase was extracted with EtOAc (3 \times 75 mL). The EtOAc extracts were combined with organic layer, washed with saturated aqueous NaCl (150 mL), dried over anhydrous Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the residue was purified by silica gel column chromatography (EtOAc/petroleum ether 1:10) to afford (S)-4-benzyl-3-((S)-2-methyldodecanoyl) oxazolidin-2-one ((*S*,*S*)-5) (4.03 g, 62 % yield, dr > 99:1, based on 13 C MNR spectra) as a white solid. $[\alpha]_D^{22} = +56.5$ (c = 2.04, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 7.34–7.21 (m, 5H), 4.68 (dd, J = 9.7, 7.2 Hz, 1H), 4.21–4.15 (m, 2H), 3.72–3.68 (m, 1H), 3.27 (dd, J = 13.4, 3.2 Hz, 1H), 2.77 (dd, J = 13.3, 9.6 Hz, 1H), 1.75–1.72 (m, 1H), 1.42–1.41 (m, 1H), 1.29–1.25 (m, 16H), 1.22 (d, *J* = 6.8 Hz, 3H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 177.49, 153.19, 135.49, 129.57, 129.05, 127.45, 66.12, 55.49, 38.04, 37.84, 33.57, 32.03, 29.79, 29.72, 29.64, 29.45, 27.40, 22.80, 17.48, 14.24; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) m/z calcd. for C₂₃H₃₅NO₃Na (M + Na)⁺: 396.2509, found 396.2518.

3.4. Synthesis of (S)-2-Methyldodecan-1-ol ((S)-6) (CAS 57289-26-6)

NaBH₄ (2.02 g, 53.68 mmol) in water (8 mL) was added dropwise to a stirred solution of oxazolidinone amide **5** (4.01 g, 10.74 mmol) in THF (64 mL) at 0 °C over 30 min. After the reaction mixture was stirred for 3 h at room temperature, it was neutralized with aqueous HCl (1 M) until the pH was 6. The resulting mixture was then extracted with Et₂O (3 × 30 mL). The combined organic layers were washed sequentially with saturated aqueous NaHCO₃ (20 mL) and saturated aqueous NaCl (2 × 20 mL), dried over anhydrous Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the residue was purified by silica gel column chromatography (EtOAc/petroleum ether 1:10) to afford (*S*)-2-methyldodecan-1-ol ((*S*)-**6**) (1.90 g, 88% yield, 98% ee, determined by chiral HPLC of its MBT derivative (*S*)-**8**). $[\alpha]_D^{22} = -9.0$ (c = 1.37, CHCl₃), literature [21] $[\alpha]_D^{23} = -8.4$ (c = 1.00, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 3.51 (dd, *J* = 10.4, 5.8 Hz, 1H), 3.41 (dd, *J* = 10.4, 6.6 Hz, 1H), 1.64–1.59 (m, 2H), 1.26–1.16 (m, 17H), 0.91 (d, *J* = 6.7 Hz, 3H), 0.88 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 68.56, 35.91, 33.30, 32.06, 30.09, 29.81, 29.79, 29.78, 29.49, 27.13, 22.83, 16.72, 14.24. HRMS (ESI) *m*/z calcd. for C₁₃H₂₈OK (M + K)⁺: 239.1772, found 239.1786.

3.5. Synthesis of (S)-1-Bromo-2-methyldodecane ((S)-7) (CAS 1333499-05-0)

PPh₃ (1.56 g, 5.97 mmol) and CBr₄ (1.98 g, 5.97 mmol) were added to a stirred solution of alcohol **6** (1.14 g, 5.69 mmol) in DCM (30 mL) at 0 °C. The reaction mixture was warmed to room temperature and stirred for 6 h. The solvent was removed under reduced pressure and the residue was purified by silica gel column chromatography (*n*-hexane) to afford (*S*)-1-bromo-2-methyldodecane ((*S*)-7) (1.48 g, 99% yield) as a colorless oil. $[\alpha]_D^{22} = -0.90$ (c = 5.33, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 3.39 (dd, *J* = 9.8, 4.9 Hz, 1H), 3.32 (dd, *J* = 9.8, 6.3 Hz, 1H), 1.80–1.76 (m, 1H), 1.31–1.18 (m, 18H), 1.01 (d, *J* = 6.6 Hz, 3H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 41.75, 35.37, 35.04, 32.07, 29.86, 29.78, 29.74, 29.50, 27.04, 22.84, 18.95, 14.26; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) *m*/z calcd. for C₁₃H₂₇Br (M)⁺: 262.1291, found 262.1279.

3.6. Synthesis of (R)-4-Benzyl-3-dodecanoyloxazolidin-2-one ((R)-4) (CAS 185803-85-4)

According to the same manner of (*S*)-4, dodecanoic acid (**2**) (5.00 g, 24.97 mmol), oxalyl chloride (4.75 g, 37.43 mmol), and (*R*)-4-benzyloxazolidin-2-one ((*R*)-3) (2.95 g, 16.64 mmol, >99% ee) afforded (*R*)-4-benzyl-3-dodecanoyloxazolidin-2-one ((*R*)-4) as a

colorless oil (5.67 g, 95% yield). $[\alpha]_D^{22} = -31.0$ (c = 2.18, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 7.35–7.17 (m, 5H), 4.70–4.67 (m, 1H), 4.21–4.15 (m, 2H), 3.30 (dd, *J* = 13.4, 3.3 Hz, 1H), 3.00–2.86 (m, 2H), 2.76 (dd, *J* = 13.4, 9.6 Hz, 1H), 1.73–1.66 (m, 2H), 1.30–1.26 (m, 16H), 0.87 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 173.62, 153.61, 135.48, 129.57, 129.09, 127.47, 66.28, 55.29, 38.06, 35.68, 32.04, 29.76, 29.72, 29.62, 29.54, 29.47, 29.27, 24.40, 22.82, 14.25. HRMS (ESI) *m/z* calcd. for C₂₂H₃₃NO₃Na (M + Na)⁺: 382.2353, found 382.2364.

3.7. Synthesis of (R)-4-Benzyl-3-((R)-2-methyldodecanoyl)oxazolidin-2-one ((R,R)-5) (CAS 185803-86-5)

According to the same manner of (*S*,*S*)-**5**, oxazolidinone amide (*R*)-**4** (6.25 g, 17.4 mmol) and MeI (12.34 g, 86.93 mmol) afforded (*R*)-4-benzyl-3-((*R*)-2-methyldodecanoyl)oxazolidin-2-one ((*R*,*R*)-**5**) (4.85 g, 75% yield, dr > 99:1, based on ¹³C MNR spectra) as a white solid. [α]22D = -71.3 (c = 2.31, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 7.34–7.21 (m, 5H), 4.67 (ddt, *J* = 10.3, 6.8, 3.1 Hz, 1H), 4.21–4.15 (m, 2H), 3.72–3.67 (m, 1H), 3.26 (dd, *J* = 13.3, 3.3 Hz, 1H), 2.77 (dd, *J* = 13.3, 9.5 Hz, 1H), 1.76–1.70 (m, 1H), 1.44–1.38 (m, 1H), 1.28–1.23 (m, 16H), 1.22 (d, *J* = 6.9 Hz, 3H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 177.49, 153.18, 135.49, 129.57, 129.03, 127.43, 66.10, 55.46, 38.01, 37.82, 33.55, 32.01, 29.77, 29.71, 29.62, 29.43, 27.37, 22.79, 17.46, 14.21; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) *m*/*z* calcd. for C₂₃H₃₅NO₃Na (M + Na)⁺: 396.2509, found 396.2516.

3.8. Synthesis of (R)-2-Methyldodecan-1-ol ((R)-6) (CAS 109034-03-9)

According to the same manner of (*S*)-6, oxazolidinone amide (*R*,*R*)-5 (3.50 g, 9.37 mmol) and NaBH₄ (1.77 g, 46.85 mmol) afforded (*R*)-2-methyldodecan-1-ol ((*R*)-6) (1.60 g, 85% yield, 99% ee, determined by chiral HPLC of its MBT derivative (*R*)-8) as a colorless oil. $[\alpha]_D^{22} = +10.8$ (c = 1.52, CHCl₃), literature [31] $[\alpha]_D^{22} = +9.1$ (c = 0.80, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 3.50 (dd, *J* = 10.5, 5.8 Hz, 1H), 3.42 (dd, *J* = 10.5, 6.5 Hz, 1H), 1.63–1.35 (m, 2H), 1.27–1.13 (m, 17H), 0.91 (d, *J* = 6.8 Hz, 3H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 68.55, 35.90, 33.30, 32.06, 30.09, 29.81, 29.78, 29.48, 27.12, 22.82, 16.71, 14.24; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) *m*/z calcd. for C₁₃H₂₈OK (M + K)⁺: 239.1772, found 239.1785.

3.9. Synthesis of (R)-1-Bromo-2-methyldodecane ((R)-7) (CAS 1643618-98-7)

According to the same manner of (*S*)-7, alcohol (*R*)-6 (1.50 g, 7.49 mmol), PPh₃ (2.06 g, 7.86 mmol), and CBr₄ (2.61 g, 7.86 mmol) afforded (*R*)-1-bromo-2-methyldodecane ((*R*)-7) (1.87 g, 95% yield) as a colorless oil. $[\alpha]_D^{22} = +0.29$ (c = 3.99, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 3.40 (dd, *J* = 9.8, 4.9 Hz, 1H), 3.32 (dd, *J* = 9.8, 6.3 Hz, 1H), 1.81–1.75 (m, 1H), 1.45–1.42 (m, 1H), 1.33–1.19 (m, 18H), 1.01 (d, *J* = 6.6 Hz, 3H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 41.77, 35.36, 35.03, 32.06, 29.86, 29.77, 29.73, 29.48, 27.03, 22.83, 18.95, 14.25; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) *m*/*z* calcd. for C₁₃H₂₈Br (M + H)⁺: 263.1369, found 263.1369.

3.10. Synthesis of 2-((2-Methyldodecyl)thio)benzo[d]thiazole (rac-8) (New Compound)

Ph₃P (0.38 g, 1.44 mmol) was added to a stirred solution of alcohol *rac*-**6** (0.24 g, 1.20 mmol) in THF (6 mL) at 0 °C. Benzo[*d*]thiazole-2(*3H*)-thione (MBT) (0.24 g, 1.44 mmol) and DIAD (0.29 g, 1.44 mmol) were added sequentially. The reaction mixture was warmed to room temperature and stirred for 4 h, followed by removal of solvent under reduced pressure. The residue was purified by silica gel column chromatography (hexanes: ethyl acetate = 50:1) to afford 2-((2-methyldodecyl)thio)benzo[*d*]thiazole (*rac*-**8**) (0.31 g, 75% yield) as a yellow oil. ¹H NMR (500 MHz, CDCl₃) δ 7.77 (dd, *J* = 8.1, 1.0 Hz, 1H), 7.66 (dd, *J* = 8.0, 1.2 Hz, 1H), 7.32 (td, *J* = 8.2, 7.7, 1.2 Hz, 1H), 7.20 (td, *J* = 7.6, 1.1 Hz, 1H), 3.33 (dd, *J* = 12.7, 5.6 Hz, 1H), 3.11 (dd, *J* = 12.7, 7.5 Hz, 1H), 1.87–1.83 (m, 1H), 1.45–1.41 (m, 1H), 1.21–1.18 (m, 17H), 1.06 (d, *J* = 6.7 Hz, 3H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 167.89, 153.51, 135.31, 126.10, 124.20, 121.57, 121.02, 36.24, 33.37, 32.05, 29.89, 29.79,29.78, 29.74,

29.48, 27.03, 22.82, 19.51, 14.26; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) m/z calcd for C₂₀H₃₂NS₂ (M + H)⁺: 350.1971, found 350.1960.

3.11. Synthesis of (S)-2-((2-Methyldodecyl)thio)benzo[d]thiazole ((S)-8) (New Compound)

According to the same manner of *rac*-**8**, alcohol (*S*)-**6** (0.080 g, 0.40 mmol) and benzo[*d*] thiazole-2(*3H*)-thione (0.080 g, 0.48 mmol) afforded (*S*)-2-((2-methyldodecyl)thio) benzo[*d*] thiazole ((*S*)-**8** (0.11 g, 79% yield, 98% ee) as a yellow oil. The ee was determined by chiral HPLC (Daicel Chiralcel OD-H column, 1% isopropanol in *n*-hexane, 0.7 mL/min, 254 nm, major t_r = 8.53 min (*S*), minor t_r = 9.40 min (*R*)). $[\alpha]_D^{22} = +7.7$ (c = 1.46, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 7.78–7.76 (m, 1H), 7.65 (dd, *J* = 8.0, 1.2 Hz, 1H), 7.33–7.29 (m, 1H), 7.21–7.17 (m, 1H), 3.33 (dd, *J* = 12.7, 5.7 Hz, 1H), 3.11 (dd, *J* = 12.7, 7.5 Hz, 1H), 1.87–1.83 (m, 1H), 1.45–1.40 (m, 1H), 1.21–1.18 (m, 17H), 0.98 (d, *J* = 6.7 Hz, 3H), 0.80 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 167.89, 153.49, 135.29, 126.09, 124.19, 121.55, 121.01, 40.84, 36.23, 33.35, 32.05, 29.89, 29.78, 29.75, 29.49, 27.03, 22.83, 19.51, 14.26; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) *m*/*z* calcd for C₂₀H₃₂NS₂ (M + H)⁺: 350.1971, found 350.1973.

3.12. Synthesis of (R)-2-((2-Methyldodecyl)thio)benzo[d]thiazole ((R)-8) (New Compound)

According to the same manner of *rac*-8, alcohol (*R*)-6 (0.080 g, 0.40 mmol) and benzo[*d*]thiazole-2(*3H*)-thione (0.080 g, 0.48 mmol) afforded (*R*)-2-((2-methyldodecyl)thio) benzo[*d*]thiazole ((*R*)-8) (0.11 g, 79% yield, 99% ee) as a yellow oil. The ee was determined by chiral HPLC (Daicel Chiralcel OD-H column, 1% isopropanol in *n*-hexane, 0.7 mL/min, 254 nm, minor $t_r = 8.78 \text{ min}$ (*S*), major $t_r = 9.44 \text{ min}$ (*R*)). [α]_D²² = -8.0 (c = 1.84, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 7.79–7.77 (m, 1H), 7.65 (dd, *J* = 8.0, 1.2 Hz, 1H), 7.34–7.31 (m, 1H), 7.21–7.19 (m, 1H), 3.33 (dd, *J* = 12.7, 5.7 Hz, 1H), 3.11 (dd, *J* = 12.7, 7.5 Hz, 1H), 1.86–1.84 (m, 1H), 1.46–1.40 (m, 1H), 1.18–1.15 (m, 17H), 0.98 (d, *J* = 6.7 Hz, 3H), 0.81 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 167.92, 153.51, 135.30, 126.11, 124.20, 121.56, 121.02, 40.86, 36.24, 33.37, 32.06, 29.89, 29.78, 29.75, 29.49, 27.03, 22.83, 19.52, 14.26; one resonance was not observed due to coincidence of the chemical shifts. HRMS (ESI) *m*/*z* calcd for C₂₀H₃₂NS₂ (M + H)⁺: 350.1971, found 350.1972.

3.13. Synthesis of (S)-11-Methylheptacos-9-ene ((S)-10) (New Compound)

Ph₃P (3.31 g, 12.62 mmol) was added to a stirred solution of hydrocarbon bromide (*S*)-7 (2.20 g, 8.35 mmol) in CH₃CN (60 mL) at room temperature. After the reaction solution was heated to 85 °C and stirred for 48 h, it was cooled to room temperature and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (DCM/MeOH 10:1) to afford the corresponding phosphonium salt (2.23 g, 51% yield) as a colorless oil.

n-BuLi (0.25 mL, 2.4 M in *n*-hexane, 0.60 mmol) was added dropwise to a stirred solution of phosphonium salt (0.24 g, 0.45 mmol) in dry THF (5 mL) at room temperature via syringe. After the reaction mixture was maintained for 2 h at the same temperature, it was cooled to -35 °C. *n*-Pentadecanal (9) (0.068 g, 0.30 mmol) in dry THF (3 mL) was then added. The reaction mixture was stirred for 5 h at -35 °C and quenched with saturated aqueous NH₄Cl (5 mL). After the layers were separated, the aqueous phase was extracted with Et₂O (3 × 5 mL). The ether extracts were combined with the organic layer, washed with saturated aqueous NaCl (20 mL), dried over anhydrous Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the residue was purified by silica gel column chromatography (petroleum ether) to give the Z/E mixtures of (*S*)-11-methylheptacos-9-ene ((*S*)-10) (0.062 g, 53% yield, *Z*:*E* = 5.1:1, determined by ¹³C NMR spectra) as a colorless liquid. [α]_D²² = -1.09 (c = 1.47, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 5.31–5.26 (m, 1H), 5.12–5.08 (m, 1H), 2.41–2.40 (m, 1H), 2.03–1.95 (m, 2H), 1.29–1.24 (m, 42H), 0.91 (d, *J* = 6.7 Hz, 3H), 0.88 (t, *J* = 6.9 Hz, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 135.63, 135.58, 127.59, 127.50, 76.42, 76.16, 75.91, 36.77, 31.10, 30.78, 29.12, 29.01, 28.87, 28.84, 28.75, 28.54, 26.70,

26.65, 21.86, 20.60, 13.28. EIMS (*m*/*z*(%)): 392.5(8, M⁺), 266.3(9), 167.1(19), 139.1(12), 83.1(78), 71.1(52), 57.1(100), 43.1(59).

3.14. Synthesis of (S)-11-Methylheptacosane ((S)-1) (CAS 1370709-05-9)

Pd (0.010 g, 10% on carbon) was placed in a 25-mL Schlenk tube, and hydrogen was charged at room temperature. Olefine (*S*)-**10** (0.033 g, 0.084 mmol) in EtOH (5 mL) was then added dropwise. The reaction mixture was maintained for 12 h, during which time hydrogen was bubbled into the Schlenk tube from a hydrogen balloon. After the reaction mixture was filtered, the filter was washed with *n*-hexane (10 mL). The filtrate and washing were combined and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (*n*-hexane) to obtain (*S*)-11-methylheptacosane ((*S*)-1) (0.018 g, 54% yield) as a colorless oil. $[\alpha]_D^{22} = -3.33$ (c = 0.60, CHCl₃), literature [13] $[\alpha]_D^{20} = -0.06$ (c = 3.33, hexanes). ¹H NMR (500 MHz, CDCl₃) δ 1.38–1.35 (m, 1H), 1.31–1.26 (m, 48H), 0.88 (t, *J* = 6.8 Hz, 6H), 0.83 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 37.25, 32.90, 32.08, 30.18, 29.89, 29.85, 29.81, 29.51, 27.24, 22.84, 19.88, 14.27; sixteen resonances were not observed due to coincidence of the chemical shifts. EIMS (*m*/*z*(%)): 379.5(3, (M–Me)⁺), 252.3(14), 168.2(34), 99.1(26), 85.1(65), 71.1(81), 57.1(100), 43.1(54).

3.15. Synthesis of (R)-11-Methylheptacos-9-ene ((R)-10) (New Compound)

According to the same manner of (*S*)-**10**, hydrocarbon bromide (*R*)-7 (2.86 g, 10.86 mmol) and Ph₃P (4.30 g, 16.39 mmol) provided the corresponding phosphonium salt (2.90 g, 51% yield) as a colorless oil, followed by the reaction with *n*-pentadecanal (**9**) (0.15 g, 0.66 mmol) affording the *Z*/*E* mixtures of (*R*)-11-methylheptacos-9-ene ((*R*)-**10**) (0.020 g, 77% yield, *Z*:*E* = 2.2:1, determined by ¹³C NMR spectra) as a colorless liquid. $[\alpha]_D^{22} = +7.09$ (c = 0.73, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 5.34–5.27 (m, 1H), 5.26–5.07 (m, 1H), 2.04–1.94 (m, 2H), 1.33–1.25 (m, 43H), 0.94–0.90 (m, 3H), 0.88 (t, *J* = 6.8 Hz, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 136.64, 136.59, 128.60, 128.51, 37.77, 37.41, 36.87, 32.74, 32.10, 31.77, 30.11, 30.00, 29.95, 29.87, 29.83, 29.74, 29.69, 29.53, 29.29, 27.69, 27.65, 27.51, 22.86, 21.59, 21.12, 14.27. EIMS (*m*/*z*(%)): 392.4(7, M⁺), 266.3(10), 167.2(19), 139.1(12), 83.1(80), 71.1(51), 57.1(100), 43.1(58).

3.16. Synthesis of (R)-11-Methylheptacosane ((R)-1) (CAS 1370709-06-0)

According to the same manner of (*S*)-1, Pd (0.023g, 10% on carbon) and olefine (*R*)-10 (0.077 g, 0.20 mmol) afforded (*R*)-11-methylheptacosane ((*R*)-1) (0.052 g, 66% yield) as a colorless oil. $[\alpha]_D^{22} = +3.31$ (c = 1.43, CHCl₃), literature [13] $[\alpha]_D^{20} = +0.09$ (c = 4.68, hexanes). ¹H NMR (500 MHz, CDCl₃) δ 1.37–1.35 (m, 1H), 1.31–1.26 (m, 48H), 0.88 (t, *J* = 6.8 Hz, 6H), 0.83 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 37.25, 32.90, 32.08, 30.19, 29.89, 29.85, 29.82, 29.52, 27.24, 22.85, 19.88, 14.27. Sixteen resonances were not observed due to coincidence of the chemical shifts. EIMS (*m*/*z*(%)): 379.5(2, (M–Me)⁺), 252.3(14), 168.2(32), 99.1(25), 85.1(65), 71.1(79), 57.1(100), 43.1(51).

4. Conclusions

In summary, we have developed a novel and efficient synthesis of (*R*)-11-methylheptacosane, the contact sex pheromone of *Tetropium fuscum*, and its enantiomer. The central element to our strategy involved Evans' chiral auxiliary to construct the stereocenter, and Wittig coupling to connect aldehyde with chiral phosphonium salt. The synthetic pheromones would be helpful for the development of the pest control.

Supplementary Materials: ¹H NMR and ¹³C NMR spectra for all the synthetic compounds and chiral HPLC chromatography of the MBT derivatives 8 can be downloaded at: https://www.mdpi.com/article/10.3390/molecules27206897/s1.

Author Contributions: Conceptualization, M.W. and J.Z.; performing the experiments and writing—original draft preparation, X.W.; participating in the chemical synthesis, J.W. and F.M.;

writing—review and editing, Q.B.; funding acquisition, J.Z. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Key Technology Research and Development Program of China (No. 2017YFD0201404).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this article are available in the Supplementary Materials.

Acknowledgments: We thank the National Key Technology Research and Development Program of China (No. 2017YFD0201404).

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Anderson, J.L.; Heard, S.B.; Sweeney, J.; Pureswaran, D.S. Mate choice errors may contribute to slow spread of an invasive Eurasian longhorn beetle in North America. *NeoBiota* **2022**, *71*, 71–89. [CrossRef]
- Lemay, M.A.; Silk, P.J.; Sweeney, J. Calling behavior of *Tetropium fuscum* (Coleoptera: Cerambycidae: Spondylidinae). *Can. Entomol.* 2010, 142, 256–260. [CrossRef]
- 3. Schroeder, M.; Cocos, D.; Johansson, H.; Sweeney, J. Attraction of the cerambycid beetles *Tetropium gabrieli*, *T. castaneum* and *T. fuscum* to pheromones and host tree volatiles. *Agr. Forest Entomol.* **2021**, *23*, 203–211. [CrossRef]
- 4. Smith, G.; Hurley, J.E. First North American record of the Palearctic species *Tetropium fuscum* (Fabricius) (Coleoptera: Cerambycidae). *Coleopts. Bull.* 2000, *54*, 540. [CrossRef]
- Sweeney, J.; De Groot, P.; MacDonald, L.; Smith, S.; Cocquempot, C.; Kenis, M.; Gutowski, J.M. Host volatile attractants and traps for detection of *Tetropium fuscum* (F.), *Tetropium castaneum* L., and other longhorned beetles (Coleoptera: Cerambycidae). *Environ. Entomol.* 2004, 33, 844–854. [CrossRef]
- 6. Dearborn Kenneth, W.; Heard Stephen, B.; Sweeney, J.; Pureswaran Deepa, S. Displacement of *Tetropium cinnamopterum* (Coleoptera: Cerambycidae) by its invasive congener *Tetropium fuscum*. *Environ*. *Entomol*. **2016**, 45, 848–854. [CrossRef]
- Heustis, A.; Heard, S.B.; Moise, E.R.D.; Johns, R.; Pureswaran, D.S. Impact of an invasive longhorned beetle, *Tetropium fuscum* (Coleoptera: Cerambycidae), on community structure of subcortical and wood-associated insects in Eastern Canada. *Environ. Entomol.* 2018, 47, 39–47. [CrossRef]
- 8. MacKay, C.A.; Sweeney, J.D.; Hillier, N.K. Olfactory receptor neuron responses of a longhorned beetle, *Tetropium fuscum* (Fabr.) (Coleoptera: Cerambycidae), to pheromone, host, and non-host volatiles. *J. Insect Physiol.* **2015**, *83*, 65–73. [CrossRef] [PubMed]
- Sweeney, J.; Silk, P.J.; Rhainds, M.; Mackay, W.; Hughes, C.; Van Rooyen, K.; Mackinnon, W.; Leclair, G.; Holmes, S.; Kettela, E.G. First report of mating disruption with an aggregation pheromone: A case study with *Tetropium fuscum* (Coleoptera: Cerambycidae). J. Econ. Entomol. 2017, 110, 1078–1086. [CrossRef] [PubMed]
- 10. Mushrow, L.; Morrison, A.; Sweeney, J.; Quiring, D. Heat as a phytosanitary treatment for the brown spruce longhorn beetle. *Forest. Chron.* **2004**, *80*, 224–228. [CrossRef]
- 11. Petkevicius, K.; Lofstedt, C.; Borodina, I. Insect sex pheromone production in yeasts and plants. *Curr. Opin. Biotechnol.* 2020, 65, 259–267. [CrossRef]
- 12. Yew, J.Y.; Chung, H. Insect pheromones: An overview of function, form, and discovery. *Prog. Lipid. Res.* 2015, 59, 88–105. [CrossRef] [PubMed]
- Silk, P.J.; Sweeney, J.; Wu, J.; Sopow, S.; Mayo, P.D.; Magee, D. Contact sex pheromones identified for two species of longhorned beetles (Coleoptera: Cerambycidae) *Tetropium fuscum* and *T. cinnamopterum* in the subfamily Spondylidinae. *Environ. Entomol.* 2011, 40, 714–726. [CrossRef]
- 14. Fujiwara-Tsujii, N.; Yasui, H. Improving contagion and horizontal transmission of entomopathogenic fungi by the white-spotted longicorn beetle, *Anoplophora malasiaca*, with help of contact sex pheromone. *Insects* **2021**, *12*, 383. [CrossRef] [PubMed]
- Ohkubo, Y.; Akasaka, K.; Masuda, Y.; Konishi, S.; Yang, C.Y.; Takikawa, H.; Mori, K. Pheromone synthesis. Part 265: Synthesis and stereochemical composition of two pheromonal compounds of the female Korean apricot wasp, *Eurytoma maslovskii*. *Tetrahedron* 2020, 76, 131410. [CrossRef]
- Yang, Z.; Jin, X.; Guaciaro, M.; Molino, B.F. Asymmetric synthesis and absolute configuration of streptophenazine G. J. Org. Chem. 2012, 77, 3191–3196. [CrossRef]
- Tsakos, M.; Clement, L.L.; Schaffert, E.S.; Olsen, F.N.; Rupiani, S.; Djurhuus, R.; Yu, W.; Jacobsen, K.M.; Villadsen, N.L.; Poulsen, T.B. Total synthesis and biological evaluation of rakicidin A and discovery of a simplified bioactive analog. *Angew. Chem. Int. Ed.* 2016, 55, 1030–1035. [CrossRef]
- 18. Evans, D.A.; Ennis, M.D.; Mathre, D.J. Asymmetric alkylation reactions of chiral imide enolates. A practical approach to the enantioselective synthesis of α-substituted carboxylic acid derivatives. *J. Am. Chem. Soc.* **1982**, *104*, 1737–1739. [CrossRef]
- 19. Wu, Y.; Shen, X.; Tang, C.-J.; Chen, Z.-L. An expeditious access to enantiomerically pure *cis*-dialkyl-substituted γ and δ -lactones. *Helv. Chim. Acta.* **2001**, *84*, 3428–3432. [CrossRef]

- Wu, Y.; Shen, X.; Tang, C.-J.; Chen, Z.-L.; Hu, Q.; Shi, W. Synthesis of natural fragrant molecules *cis*-3-methyl-4-decanolide and aerangis lactone. General enantioselective routes to β, γ-cis-disubstituted γ-lactones and γ,δ-cis-disubstituted δ-lactones. *J. Org. Chem.* 2002, 67, 3802–3810. [CrossRef] [PubMed]
- Schlamp, K.K.; Gries, R.; Khaskin, G.; Brown, K.; Khaskin, E.; Judd, G.J.; Gries, G. Pheromone components from body scales of female *Anarsia lineatella* induce contacts by conspecific males. *J. Chem. Ecol.* 2005, 31, 2897–2911. [CrossRef]
- Appel, R. Tertiary phosphane/tetrachloromethane, a versatile reagent for chlorination, dehydration, and phosphorus-nitrogen linking. Angew. Chem. Int. Ed. 1975, 14, 801–811. [CrossRef]
- 23. Favaro, C.F.; Millar, J.G.; Zarbin, P.H.G. Identification and synthesis of the male-produced sex pheromone of the stink bug, *Pellaea* stictica. J. Chem. Ecol. 2015, 41, 859–868. [CrossRef] [PubMed]
- 24. Zhang, H.L.; Sun, Z.F.; Zhou, L.N.; Liu, L.; Zhang, T.; Du, Z.T. Synthesis of the sex pheromone of the tea tussock moth based on a resource chemistry strategy. *Molecules* **2018**, *23*, 1347. [CrossRef] [PubMed]
- Mitsunobu, O.; Wada, M.; Sano, T. Stereospecific and stereoselective reactions. I. Preparation of amines from alcohols. J. Am. Chem. Soc. 1972, 94, 679–680. [CrossRef]
- Woo, S.H.; Frechette, S.; Khalil, E.A.; Bouchain, G.; Vaisburg, A.; Bernstein, N.; Moradei, O.; Leit, S.; Allan, M.; Fournel, M.; et al. Structurally simple trichostatin A-like straight chain hydroxamates as potent histone deacetylase inhibitors. *J. Med. Chem.* 2002, 45, 2877–2885. [CrossRef] [PubMed]
- 27. Wittig, G.; Schollkopf, U. Triphenylphosphinemethylene as an olefin-forming reagent. I. Chem. Ber. 1954, 97, 1318–1330. [CrossRef]
- Bruns, H.; Herrmann, J.; Mueller, R.; Wang, H.; Wagner Doebler, I.; Schulz, S. Oxygenated *N*-acyl alanine methyl esters (NAMEs) from the marine bacterium *Roseovarius tolerans* EL-164. *J. Nat. Prod.* 2018, *81*, 131–139. [CrossRef] [PubMed]
- 29. Patil, P.S.; Cheng, T.-J.R.; Zulueta, M.M.L.; Yang, S.-T.; Lico, L.S.; Hung, S.-C. Total synthesis of tetraacylated phosphatidylinositol hexamannoside and evaluation of its immunomodulatory activity. *Nat. Commun.* **2015**, *6*, 7239. [CrossRef] [PubMed]
- Ramachandran, P.V.; Srivastava, A.; Hazra, D. Total synthesis of potential antitumor agent, (–)-dictyostatin. Org. Lett. 2007, 9, 157–160. [CrossRef] [PubMed]
- Cao, J.; Perlmutter, P. "One-pot" reductive lactone alkylation provides a concise asymmetric synthesis of chiral isoprenoid targets. Org. Lett. 2013, 15, 4327–4329. [CrossRef] [PubMed]