



# Whole-Genome Sequence of the Soil Bacterium *Micrococcus* sp. KBS0714

V. Kuo,<sup>a</sup> W. R. Shoemaker,<sup>a</sup> M. E. Muscarella,<sup>b</sup> J. T. Lennon<sup>a</sup>

Department of Biology, Indiana University, Bloomington, Indiana, USA<sup>a</sup>; Department of Plant Biology, University of Illinois, Urbana-Champaign, Illinois, USA<sup>b</sup>

**ABSTRACT** We present here a draft genome assembly of *Micrococcus* sp. KBS0714, which was isolated from agricultural soil. The genome provides insight into the strategies that *Micrococcus* spp. use to contend with environmental stressors such as desiccation and starvation in environmental and host-associated ecosystems.

Microorganisms have evolved traits that allow them to cope with a wide range of environmental conditions. For example, chemoorganotrophic representatives of the genus *Micrococcus* (class *Actinobacteria*, family *Micrococcaceae*) are commonly found in temperate soil (1), water (2), mammalian skin (3), Antarctic ice (4), and desert soil (5). Survival and reproduction in some of these habitats has been linked to the ability of *Micrococcus* spp. to form biofilms or enter dormant stages in response to conditions such as desiccation and starvation (6–8). Genome sequencing may illuminate additional traits that allow *Micrococcus* spp. to persist when challenged with environmental stressors. However, outside of isolates from contaminated ecosystems (1, 9–12), very few *Micrococcus* genomes have been sequenced from soil. Here, we present the draft genome of *Micrococcus* sp. KBS0714, isolated from never-tilled agricultural soil at the Kellogg Biological Station Long-Term Ecological Research site (6, 13).

*Micrococcus* sp. KBS0714 genomic DNA was prepared with the Illumina TruSeq DNA sample prep kit using an insert size of 250 bp for sequencing on an Illumina HiSeq 2500 with 100-bp paired-end reads (Illumina, San Diego, CA, USA). Raw sequences were processed by removing the TruSeq adaptors and the first 10 bp using Cutadapt version 1.9 (14), interleaving the paired reads using khmer version 2.0 (15), and quality-filtering with an average Phred score of 30 using the FASTX-Toolkit version 0.0.13 (Hannon Lab, [http://hannonlab.cshl.edu/fastx\\_toolkit](http://hannonlab.cshl.edu/fastx_toolkit)). The coverage was normalized to 25 based on a *k*-mer size of 25 bp using khmer, resulting in a total of 1,614,974 unmapped paired-end reads. The genome was assembled using Velvet version 1.2.10 (16) with the following parameters: a *k*-mer size of 55, expected coverage of 18×, and a coverage cutoff of 2.29. Contigs longer than 200 bp were annotated using Prokka version 1.12 (<https://github.com/tseemann/prokka>) (17). Finally, we used MAPLE version 2.3.0 with bidirectional best-hit matches (<http://www.genome.jp/tools/maple>) (18) to predict metabolic and physiological functions.

The draft assembly of *Micrococcus* sp. KBS0714 comprises 2,489,009 bp. It consists of 63 contigs with an  $N_{50}$  of 122,407 and a G+C content of 69%. Based on our gene annotation, the genome contains 2,210 protein-coding sequences, 3 rRNAs, 52 tRNAs, 2,267 genes, and 2 transfer-messenger RNAs (tmRNAs).

The KBS0714 genome highlights traits that may be used by *Micrococcus* spp. in soil. For example, we found genes involved with general stress response (*relA*) (19, 20) and biofilm formation (*brpA*) (21) in both the genome of KBS0714 and the closely related *M. luteus* NCTC 2662 (99% 16S rRNA sequence similarity, NCBI CP001628). Additionally, we detected pathways for desiccation and starvation resistance, such as genes for osmotic stress response (*mtrB-mtrA*, *rpoE*) (22, 23),

Received 7 June 2017 Accepted 8 June 2017 Published 10 August 2017

**Citation** Kuo V, Shoemaker WR, Muscarella ME, Lennon JT. 2017. Whole-genome sequence of the soil bacterium *Micrococcus* sp. KBS0714. *Genome Announc* 5:e00697-17. <https://doi.org/10.1128/genomeA.00697-17>.

**Copyright** © 2017 Kuo et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to J. T. Lennon, [lennonj@indiana.edu](mailto:lennonj@indiana.edu).

phosphate starvation response (*senX3-regX3*), and membrane lipid fluidity regulation (*desK-desR*) (24, 25). In summary, the KBS0714 genome has features that may provide a selective advantage under stressful conditions in soils and other environments.

**Accession number(s).** This draft genome assembly has been deposited in DDBJ/EMBL/GenBank under the accession number [MVDF00000000](https://doi.org/10.1128/genomeA.00627-15). The version described here is the second version, MVDF02000000. The code used for assembly and annotation is available online at <https://github.com/LennonLab/Micrococcus>.

## ACKNOWLEDGMENTS

We thank B. K. Lehmkuhl for technical assistance.

Our research was supported by National Research Initiative grants (2011-67019-30225) from the USDA National Institute of Food and Agriculture, by the National Science Foundation (1442246 to J.T.L.), and by the U.S. Army Research Office (W911NF-14-1-0411).

## REFERENCES

- Sims GK, Sommers LE, Konopka A. 1986. Degradation of pyridine by *Micrococcus luteus* isolated from soil. *Appl Environ Microbiol* 51:963–968.
- Dib JR, Angelov A, Liebl W, Döbber J, Voget S, Schuldes J, Gorriti M, Farias ME, Meinhardt F, Daniel R. 2015. Complete genome sequence of the linear plasmid pJD12 hosted by *Micrococcus* sp. D12, isolated from a high-altitude volcanic lake in Argentina. *Genome Announc* 3(3):e00627-15. <https://doi.org/10.1128/genomeA.00627-15>.
- Kloos WE, Tornabene TG, Schleifer KH. 1974. Isolation and characterization of micrococci from human skin, including two new species: *Micrococcus lylae* and *Micrococcus kristinae*1. *Int J Syst Evol Microbiol* 24:79–101. <https://doi.org/10.1099/00207713-24-1-79>.
- Liu H, Xu Y, Ma Y, Zhou P. 2000. Characterization of *Micrococcus antarcticus* sp. nov., a psychrophilic bacterium from Antarctica. *Int J Syst Evol Microbiol* 50:715–719. <https://doi.org/10.1099/00207713-50-2-715>.
- Azua-Bustos A, González-Silva C. 2014. Biotechnological applications derived from microorganisms of the Atacama Desert. *Biomed Res Int* 2014:909312. <https://doi.org/10.1155/2014/909312>.
- Lennon JT, Aanderud ZT, Lehmkuhl BK, Schoolmaster DR. 2012. Mapping the niche space of soil microorganisms using taxonomy and traits. *Ecology* 93:1867–1879. <https://doi.org/10.1890/11-1745.1>.
- Rickard AH, Leach SA, Hall LS, Buswell CM, High NJ, Handley PS. 2002. Phylogenetic relationships and coaggregation ability of freshwater biofilm bacteria. *Appl Environ Microbiol* 68:3644–3650. <https://doi.org/10.1128/AEM.68.7.3644-3650.2002>.
- Lennon JT, Jones SE. 2011. Microbial seed banks: the ecological and evolutionary implications of dormancy. *Nature Reviews Microbiology* 9:119–130. <https://doi.org/10.1038/nrmicro2504>.
- Ghazali FM, Rahman RNZA, Salleh AB, Basri M. 2004. Biodegradation of hydrocarbons in soil by microbial consortium. *Int Biodeterior Biodegrad* 54:61–67. <https://doi.org/10.1016/j.ibiod.2004.02.002>.
- Tallur PN, Megadi VB, Ninnekar HZ. 2008. Biodegradation of cypermethrin by *Micrococcus* sp. strain CPN 1. *Biodegradation* 19:77–82. <https://doi.org/10.1007/s10532-007-9116-8>.
- Tuleva B, Christova N, Cohen R, Antonova D, Todorov T, Stoineva I. 2009. Isolation and characterization of trehalose tetraester biosurfactants from a soil strain *Micrococcus luteus* BN56. *Proc Biochem* 44:135–141. <https://doi.org/10.1016/j.procbio.2008.09.016>.
- Zhang JY, Liu XY, Liu SJ. 2010. *Agrococcus terreus* sp. nov. and *Micrococcus terreus* sp. nov., isolated from forest soil. *Int J Syst Evol Microbiol* 60:1897–1903. <https://doi.org/10.1099/ijs.0.013235-0>.
- Grandy AS, Robertson GP. 2006. Aggregation and organic matter protection following tillage of a previously uncultivated soil. *Soil Sci Soc Am J* 70:1398–1406. <https://doi.org/10.2136/sssaj2005.0313>.
- Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnetJ* 17:10–12. <https://doi.org/10.14806/ej.17.1.200>.
- McDonald E, Brown CT. 2013. khmer: working with big data in bioinformatics, preprint, arXiv:1303.2223:1-8.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <https://doi.org/10.1101/gr.074492.107>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
- Takami H, Taniguchi T, Arai W, Takemoto K, Moriya Y, Goto S. 2016. An automated system for evaluation of the potential functionome: MAPLE version 2.1.0. *DNA Res* 23:467–475. <https://doi.org/10.1093/dnares/dsw030>.
- Primm TP, Andersen SJ, Mizrahi V, Avarbock D, Rubin H, Barry CE. 2000. The stringent response of *Mycobacterium tuberculosis* is required for long-term survival. *J Bacteriol* 182:4889–4898. <https://doi.org/10.1128/JB.182.17.4889-4898.2000>.
- Aertsen A, Michiels CW. 2004. Stress and how bacteria cope with death and survival. *Crit Rev Microbiol* 30:263–273. <https://doi.org/10.1080/10408410490884757>.
- Wen ZT, Baker HV, Burne RA. 2006. Influence of BrpA on critical virulence attributes of *Streptococcus mutans*. *J Bacteriol* 188:2983–2992. <https://doi.org/10.1128/JB.188.8.2983-2992.2006>.
- Hiratsu K, Amemura M, Nashimoto H, Shinagawa H, Makino K. 1995. The *rpoE* gene of *Escherichia coli*, which encodes sigma E, is essential for bacterial growth at high temperature. *J Bacteriol* 177:2918–2922. <https://doi.org/10.1128/jb.177.10.2918-2922.1995>.
- Möker N, Krämer J, Unden G, Krämer R, Morbach S. 2007. In vitro analysis of the two-component system MtrB-MtrA from *Corynebacterium glutamicum*. *J Bacteriol* 189:3645–3649. <https://doi.org/10.1128/JB.01920-06>.
- Aguilar PS, Hernandez-Arriaga AM, Cybulski LE, Erazo AC, de Mendoza D. 2001. Molecular basis of thermosensing: a two-component signal transduction thermometer in *Bacillus subtilis*. *EMBO J* 20:1681–1691. <https://doi.org/10.1093/emboj/20.7.1681>.
- Rietkötter E, Hoyer D, Mascher T. 2008. Bacitracin sensing in *Bacillus subtilis*. *Mol Microbiol* 68:768–785. <https://doi.org/10.1111/j.1365-2958.2008.06194.x>.