

Article

Pro-Inflammatory Chemokines CCL5, CXCL12, and CX3CL1 Bind to and Activate Platelet Integrin α IIb β 3 in an Allosteric Manner

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Abstract: Activation of platelet integrin α IIb β 3, a key event for hemostasis and thrombus formation, is known to be mediated exclusively by inside-out signaling. We showed that inflammatory chemokines CX3CL1 and CXCL12 in previous studies, and CCL5 in this study, bound to the allosteric binding site (site 2) of vascular integrin α v β 3, in addition to the classical ligand binding site (site 1), and allosterically activated integrins independent of inside-out signaling. Since α IIb β 3 is exposed to inflammatory chemokines at increased concentrations during inflammation (e.g., cytokine/chemokine storm) and platelet activation, we hypothesized that these chemokines bind to and activate α IIb β 3 in an allosteric activation mechanism. We found that these chemokines bound to and activate α IIb β 3 in an allosteric activation mechanism. We found that these chemokines bound to and activate α IIb β 3 in 1 mM Ca²⁺ by binding to site 2. They activated cell-surface α IIb β 3 on CHO cells, which lack machinery for inside-out signaling or chemokine receptors, quickly (<1 min) and at low concentrations (1–10 ng/mL) compared to activation of soluble α IIb β 3, probably because chemokines bind to cell surface proteoglycans. Furthermore, activation of α IIb β 3 by the chemokines was several times more potent than 1 mM Mn²⁺. We propose that CCL5 and CXCL12 (stored in platelet granules) may allosterically activate α IIb β 3 upon platelet activation and trigger platelet aggregation. Transmembrane CX3CL1 on activated endothelial cells may mediate platelet–endothelial interaction by binding to and activating α IIb β 3. Additionally, these chemokines in circulation over-produced during inflammation may trigger α IIb β 3 activation, which is a possible missing link between inflammation and thrombosis.



Citation: Takada, Y.K.; Fujita, M.; Takada, Y. Pro-Inflammatory Chemokines CCL5, CXCL12, and CX3CL1 Bind to and Activate Platelet Integrin α IIb β 3 in an Allosteric Manner. *Cells* **2022**, *11*, 3059. <https://doi.org/10.3390/cells11193059>

Academic Editor: Sylvain G. Bourgoin

Received: 8 September 2022

Accepted: 24 September 2022

Published: 29 September 2022

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Keywords: integrin activation; chemokine; platelet

1. Introduction

Integrins are a superfamily of cell-surface heterodimers that recognize extracellular matrix (ECM) proteins, cell-surface ligands (e.g., VCAM-1 and ICAM-1), and soluble ligands (e.g., growth factors such as FGF and IGF) [1–3]. Integrins play important roles in normal biological process (e.g., wound healing and hemostasis) and in the pathogenesis of diseases [4]. By virtual screening of the protein data bank using docking simulation, we identified the chemokine domain of CX3CL1 [5] as a new ligand for integrins α v β 3, α 4 β 1, and α 5 β 1, and that CX3CL1 and integrins simultaneously bound to CX3CR1 [5]. The CX3CL1 mutant defective in integrin binding was defective in signaling and acted as an antagonist, although it still bound to CX3CR1 [5], indicating that the direct integrin binding to CX3CL1 and subsequent integrin–CX3CL1–CX3CR1 ternary complex formation is required for its signaling functions. CX3CL1 is expressed on the cell surface of IL-1 β - and TNF α -activated endothelium as a membrane-bound form [6]. Soluble CX3CL1 is released

by metalloproteinases ADAM10 and ADAM17 [7–9]. Interaction between membrane-bound CX3CL1 and CX3CR1 promotes leukocyte adhesion to the endothelium [10–12].

We unexpectedly discovered that CX3CL1 activated soluble integrin $\alpha v\beta 3$ in 1 mM Ca^{2+} in cell-free conditions by binding to the second ligand-binding site (site 2), which is distinct from the classical ligand-binding site (site 1) in the integrin headpiece [13]. We showed that peptides from site 2 of the integrin β subunit bound to CX3CL1 and blocked integrin activation by CX3CL1 [13]. Therefore, we concluded that site 2 is the allosteric binding site and involved in allosteric activation of integrins. We showed that CXCL12 activated integrins $\alpha v\beta 3$, $\alpha 4\beta 1$, and $\alpha 5\beta 1$ by binding to site 2 [14], indicating that this mechanism of integrin activation is not limited to CX3CL1. CXCL12 is a potent chemoattractant for leukocytes and is believed to regulate signaling events through CXCR4 and CXCR7 in leukocytes [15–18]. CXCL12 (Stromal cell-derived factor 1, SDF-1) is ubiquitously expressed in many tissues and cell types. Binding of CXCL12 to CXCR4 induces heterotrimeric G protein signaling leading to activation of the pro-inflammatory signaling [19,20]. CXCL12/CXCR4 are over-expressed in rheumatoid arthritis, systemic lupus erythematosus, multiple sclerosis, and inflammatory bowel disease [21–24]. CXCL12 is stored in platelet granules and rapidly transported to the surface upon platelet activation [25].

Platelet integrin $\alpha \text{IIb}\beta 3$ is a receptor for several proteins including fibronectin, fibrinogen, plasminogen, prothrombin, thrombospondin, vitronectin, and von Willebrand factor (VWF) [3]. Activation of $\alpha \text{IIb}\beta 3$ is a key event that triggers platelet aggregation by inducing $\alpha \text{IIb}\beta 3$ binding to fibrinogen leading to bridge formation between platelets [26,27]. The mechanism of integrin activation has been extensively studied in integrin $\alpha \text{IIb}\beta 3$ as a model. Integrin activation is defined by the increase in binding to monomeric ligand and we have used monomeric fragments of fibrinogen or fibronectin.

It has been proposed that activation of integrin $\alpha \text{IIb}\beta 3$ is induced exclusively by inside-out signaling by platelet agonists (e.g., thrombin, ADP, collagen), which bind to receptors on the cell surface. Signals received by other receptors induce the binding of talin and kindlin to cytoplasmic end of the integrin β subunit at sites of actin polymerization [26–28]. However, it has been proposed that canonical integrin activation pathways by platelet agonists induces integrin binding to multivalent ligands (e.g., ligand-mimetic antibody, Pac-1 IgM specific for $\alpha \text{IIb}\beta 3$, with potential 10-binding sites), but does not enhance ligand binding affinity to monovalent ligand [29,30].

We showed that the affinity to monomeric ligands is increased by chemokines CX3CL1 and CXCL12 in non- $\alpha \text{IIb}\beta 3$ integrins, but it is unclear if this occurs in platelet $\alpha \text{IIb}\beta 3$. Notably, CXCL12 is stored in platelet granules and rapidly transported to the surface upon platelet activation. We thus hypothesized that other chemokines that are stored in platelet granules (e.g., CCL5) are potentially involved in allosteric activation of platelet integrin $\alpha \text{IIb}\beta 3$ and trigger thrombus formation. So, we decided to include CCL5 in the present study. It is important to study whether the binding of chemokines CCL5, CX3CL1, and CXCL12 to site 2 activates $\alpha \text{IIb}\beta 3$, a model system for integrin activation.

CCL5 (Rantes) is a pro-inflammatory chemokine, recruiting leukocytes to the site of inflammation. CCL5 is chemotactic for T cells, eosinophils, and basophils, monocytes, natural-killer (NK) cells, dendritic cells, and mastocytes [31]. CCL5 is mainly expressed by T-cells and monocytes [32] and it is abundantly expressed by epithelial cells, fibroblasts, and thrombocytes. CCL5 is involved in numerous human diseases and disorders, including transplantation rejection [33], anti-viral immunity [31], tumor development [34], and viral hepatitis or COVID-19 [32].

In the present study, we first describe that CCL5 bound to and activated vascular integrin $\alpha v\beta 3$, as in CX3CL1 and CXCL12. We describe that CCL5, CXCL12, and CX3CL1 specifically bound to $\alpha \text{IIb}\beta 3$ to site 1, indicating that they are new ligands for $\alpha \text{IIb}\beta 3$. These chemokines activated soluble $\alpha \text{IIb}\beta 3$ by binding to site 2. In addition, cell-surface $\alpha \text{IIb}\beta 3$ on CHO cells, which lack the machinery for inside-out signaling or cognate receptors for these chemokines, was more efficiently activated by these chemokines probably because they were concentrated by binding to surface proteoglycans. These findings indicate that $\alpha \text{IIb}\beta 3$

can be activated in an allosteric manner by CXCL12 and CCL5 stored in platelet granules, and CX3CL1, which is expressed on the cell surface (e.g., activated endothelial cells). We propose a new mechanism of α IIB β 3 activation by chemokines in the platelet granules upon platelet activation. Additionally, exogenous chemokines during inflammation (e.g., cytokines storms or autoimmune diseases) might be a possible missing link between inflammation and thrombosis.

2. Experimental Procedures

Materials. cDNA encoding (6 His tag and Fibrinogen γ -chain C-terminal residues 390-411) [HHHHHHH]NRLTIGEGQQHHLGGAKQAGDV] was conjugated with the C-terminus of GST (designated γ C390-411) in pGEXT2 vector (BamHI/EcoRI site). The protein was synthesized in *E. coli* BL21 and purified using glutathione affinity chromatography. CHO cells that express recombinant human α IIB β 3 were described [35]. The fibrinogen γ -chain C-terminal domain (γ C151-411) was generated as previously described [36]. CX3CL1 [5] and CXCL12 [14] were synthesized as described. cDNA encoding mature CCL5 (SPYSSDTPCCFAYIARPLPRAHIKEYFYTSKGKCSNPAVVVFVTRKNRQVCANPEKKWVREYINSLEMS) was synthesized and subcloned into the BamHI/EcoRI site of pET28a vector. Protein synthesis was induced by IPTG in *E. coli* BL21. Proteins were produced as insoluble inclusion bodies and purified in denaturing conditions and refolded as described [5]. The disintegrin domain of ADAM15 fused to GST (ADMA15 disintegrin) and parent GST were synthesized as previously described [37].

Cyclic β 3 site 2 peptide fused to GST-The 29-mer cyclic β 3 site 2 peptide C260-RLAGIV[QPNDGSHVGSNDNHYSASTTM]C288 was synthesized by inserting oligonucleotides encoding this sequence into the BamHI-EcoRI site of pGEX-2T vector. The positions of Cys residues for disulfide linkage were selected by using Disulfide by Design-2 (DbD2) software (<http://cptweb.cpt.wayne.edu/DbD2/>) [38]. It predicted that mutating Gly260 and Asp288 to Cys disulfide-linked cyclic site 2 peptide of β 3 does not affect the conformation of the original site 2 peptide sequence QPNDGSHVGSNDNHYSASTTM in the 3D structure. We found that the cyclic site 2 peptide bound to CX3CL1 and sPLA2-IIA to a similar extent to non-cyclized β 3 site 2 peptides in ELISA-type assays (data not shown).

2.1. Binding of Site 2 Peptide to CCL5

Wells of 96-well Immulon 2 microtiter plates (Dynatech Laboratories, Chantilly, VA, USA) were coated with 100 μ L PBS containing CCL5 for 2 h at 37 °C. Remaining protein binding sites were blocked by incubating with PBS/0.1% BSA for 30 min at room temperature. After washing with PBS, GST-tagged cyclic site 2 peptide was added to the wells and incubated in PBS for 2 h at room temperature. After unbound GST-tagged site 2 peptides were removed by rinsing the wells with PBS, bound GST-tagged site 2 peptides were measured using HRP-conjugated anti-GST antibody and peroxidase substrates.

2.2. Binding of Soluble α IIB β 3 to Chemokines

ELISA-type binding assays were performed as described previously [5]. Briefly, wells of 96-well Immulon 2 microtiter plates (Dynatech Laboratories, Chantilly, VA, USA) were coated with 100 μ L PBS containing chemokines for 2 h at 37 °C. The remaining protein binding sites were blocked by incubating with PBS/0.1% BSA for 30 min at room temperature. After washing with PBS, soluble recombinant α IIB β 3 (AgroBio, 1 μ g/mL) was added to the wells and incubated in HEPES-Tyroses buffer (10 mM HEPES, 150 mM NaCl, 12 mM NaHCO₃, 0.4 mM NaH₂PO₄, 2.5 mM KCl, 0.1% glucose, 0.1% BSA) with 1 mM MnCl₂ for 1 h at room temperature. After unbound α IIB β 3 was removed by rinsing the wells with binding buffer, bound α IIB β 3 was measured using anti-integrin β 3 mAb (AV-10) followed by HRP-conjugated goat anti mouse IgG and peroxidase substrates.

2.3. Activation of Soluble α IIb β 3 by Chemokines

ELISA-type binding assays were performed as described previously [14]. Briefly, wells of 96-well Immulon 2 microtiter plates were coated with 100 μ L PBS containing γ C390-411 for 2 h at 37 °C. Remaining protein binding sites were blocked by incubating with PBS/0.1% BSA for 30 min at room temperature. After washing with PBS, soluble recombinant α IIb β 3 (1 μ g/mL) was pre-incubated with chemokines for 10 min at room temperature and was added to the wells and incubated in HEPES-Tyroses buffer with 1 mM CaCl₂ for 1 h at room temperature. After unbound α IIb β 3 was removed by rinsing the wells with binding buffer, bound α IIb β 3 was measured using anti-integrin β 3 mAb (AV-10) followed by HRP-conjugated goat anti mouse IgG and peroxidase substrates.

2.4. Activation of Cell-Surface α IIb β 3 by Chemokines

α IIb β 3-CHO cells were cultured in DMEM/10% FCS. The cells were resuspended with HEPES-Tyroses buffer/0.02% BSA (heat-treated at 80 °C for 20 min to remove contaminating cell adhesion molecules). The α IIb β 3-CHO cells were then incubated with chemokines for 30 min on ice and then incubated with FITC-labeled γ C390-411 (50 μ g/mL) for 30 min at room temperature. The cells were washed with PBS/0.02% BSA and analyzed in BD Accuri flow cytometer (Becton Dickinson, Mountain View, CA, USA) or Attune flow cytometer (ThermoFischer Scientific, Waltham, MA, USA). For time-course studies, FITC-labeled γ C390-411 was added to cell suspension first and the mixture was kept on ice, and then incubated with chemokines for the time indicated before being analyzed in flow cytometry. The data were analyzed using FlowJo 7.6.5, BD Bioscience, Franklin Lakes, NJ, USA.

2.5. Docking Simulation

Docking simulation of interaction between CCL5 (1EQT.pdb) and integrin α v β 3 (open headpiece form 1L5G, or closed headpiece form, PDB code 1JV2) was performed using AutoDock3 as described previously [1]. We used the headpiece (residues 1–438 of α v and residues 55–432 of β 3) of α v β 3. Cations were not present in integrins during docking simulation. The ligand is presently compiled to a maximum size of 1024 atoms. Atomic solvation parameters and fractional volumes were assigned to the protein atoms by using the AddSol utility, and grid maps were calculated by using AutoGrid utility in AutoDock 3.05. A grid map with 127 \times 127 \times 127 points and a grid point spacing of 0.603 Å included the headpiece of α v β 3. Kollman ‘united-atom’ charges were used. AutoDock 3.05 uses a Lamarckian genetic algorithm (LGA) that couples a typical Darwinian genetic algorithm for global searching with the Solis and Wets algorithm for local searching. The LGA parameters were defined as follows: the initial population of random individuals had a size of 50 individuals; each docking was terminated with a maximum number of 1 \times 10⁶ energy evaluations or a maximum number of 27,000 generations, whichever came first; mutation and crossover rates were set at 0.02 and 0.80, respectively. An elitism value of 1 was applied, which ensured that the top-ranked individual in the population always survived into the next generation. A maximum of 300 iterations per local search were used. The probability of performing a local search on an individual was 0.06, whereas the maximum number of consecutive successes or failures before doubling or halving the search step size was 4.

2.6. Statistical Analysis

Treatment differences were tested using ANOVA and a Tukey multiple comparison test to control the global type I error using Prism 7 (GraphPad Software, San Diego, CA, USA).

3. Results

3.1. CCL5 Binds to and Activates Integrin $\alpha v\beta 3$

The primary goal of this study is to determine if three chemokines bind to and activate integrin $\alpha IIb\beta 3$. Active and inactive conformations, 1L5G.pdb (open headpiece $\alpha v\beta 3$) and 1JV2 (closed-headpiece $\alpha v\beta 3$), are defined in $\alpha v\beta 3$ but not in $\alpha IIb\beta 3$. We have shown that CX3CL1 binds to the classical ligand (RGD)-binding site (site 1) in open headpiece $\alpha v\beta 3$ and to the allosteric site (site 2) in a closed headpiece $\alpha v\beta 3$ in docking simulation [13]. Additionally, CXCL12 bound to site 2 and activated integrins $\alpha v\beta 3$, $\alpha 4\beta 1$, and $\alpha 5\beta 1$ [14]. It is unclear if CCL5, a major chemokine in platelet granules, binds to and activates any integrins. Thus, we first studied whether CCL5 binds to site 1 and site 2 of $\alpha v\beta 3$ before we studied CCL5 binding to $\alpha IIb\beta 3$. Our docking simulation between integrin $\alpha v\beta 3$ and CCL5 predicted that CCL5 binds to site 1 at high affinity (docking energy -24.8 kcal/mol) (Figure 1a) and to site 2 (docking energy -20.2 kcal/mol) (Figure 1b). Amino acid residues predicted to be involved in site 1 and site 2 binding are shown in Tables 1 and 2.

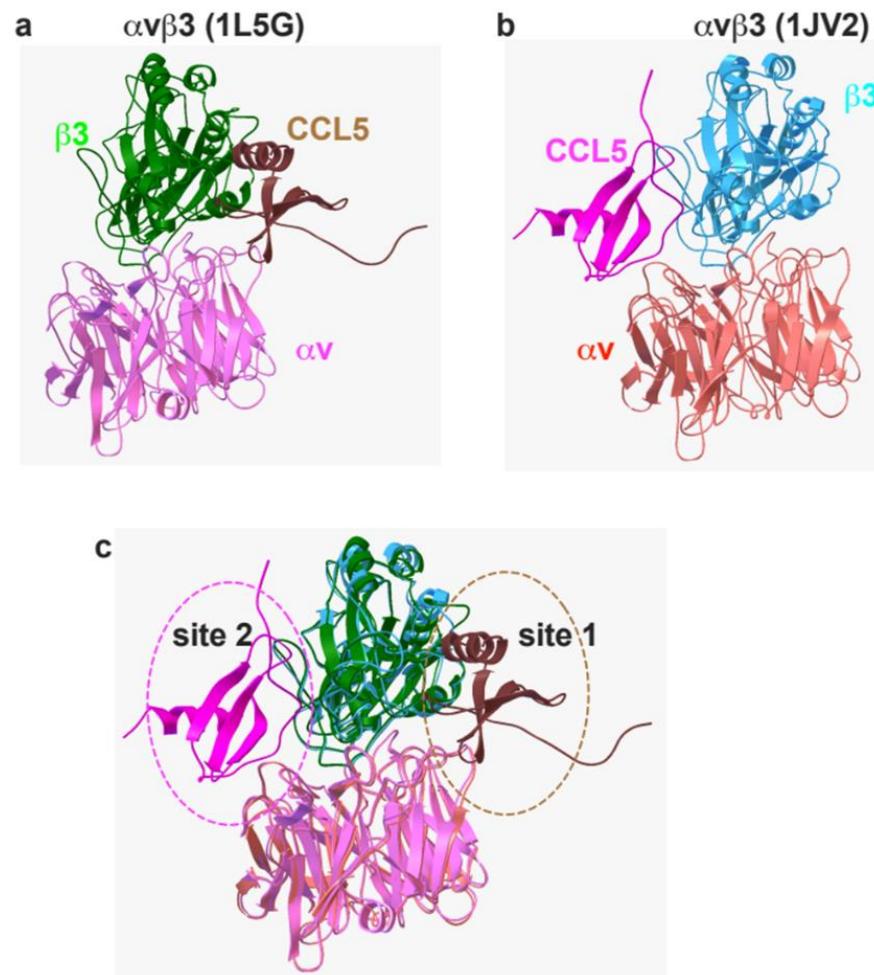


Figure 1. Model of the interaction between CCL5 and integrin $\alpha v\beta 3$. Docking simulation between CCL5 (1EQT.pdb) and integrin $\alpha v\beta 3$ (a) with open-headpiece conformation 1L5G.pdb or (b) with closed headpiece conformation, 1JV2.pdb using Autodock3, as described in the Methods section; the docking models were superimposed (c). Different colors indicate specific protein chain.

Table 1. Amino acid residues involved in CCL5 (1EQT.pdb) and $\alpha v\beta 3$ (1L5G.pdb, open headpiece) predicted by docking simulation.

CCL5	αv	$\beta 3$
Phe12, Ala13, Tyr14, Ile15, Ala16, Arg17, Pro18, Met19, Pro20, Arg21, Ala22, His23, Thr43, Lys45, Arg47, Val49, Cys50, Asn52, Glu54, Lys55, Lys56, Trp57, Arg59, Glu60 Tyr61	Met118, Lys119, Gln145, Asp146, Ile147, Asp148, Asp150, Gly151, Tyr178, Thr212, Ala215, Ile216, Phe217, Asp218, Asp219, Arg248,	Asp119, Ser121, Tyr122, Ser123, Met124, Lys125, Asp126, Asp127, Tyr166, Asp179, Met180, Thr182, Arg214, Asn215, Arg216, Asp217, Ala218, Pro219, Glu220, Asp251, Ala252, Lys253, Asn313,

Amino acid residues within 0.6 nm between CCL5 and $\alpha v\beta 3$ were selected using Swiss PDB Viewer (version 4.1) (Swiss Institute of Bioinformatics, Basel, Swiss).

Table 2. Amino acid residues involved in CCL5 and $\alpha v\beta 3$ (1JV2.pdb, closed headpiece) predicted by docking simulation.

CCL5	αv	$\beta 3$
Thr7, Thr8, Cys10, Phe12, Ala13, Tyr14, Ile15, Ala16, Arg17, Pro18, Met19, Pro20, Arg21, His23, Ser35, Asn36, Pro37, Thr43, Lys45, Arg47, Gln48, Val49, Asn52, Glu55, Lys56, Trp57, Tyr61	Glu15, Gly16, Tyr18, Lys42, Asn44, Thr45, Thr46, Gln47, Pro48, Gly49, Ile50, Val51, Glu52, Ser90, His91	Lys159, Pro160, Val161, Ser162, Met165, Glu171, Glu174, Asn175, Pro186, Met187, Phe188, Val266, Gln267, Val275, Gly276, Ser277, Asp278, His280, Tyr281, Ser282, Ala283, Thr285, Thr286

Amino acid residues within 0.6 nm between CCL5 and $\alpha v\beta 3$ were selected using Pdb Viewer (version 4.1). Amino acid residues in $\beta 3$ that are in the cyclic site 2 peptide are in bold.

We studied whether $\alpha v\beta 3$ binds to CCL5 in ELISA-type binding assays. CCL5 was immobilized to wells of 96-well microtiter plates and incubated with soluble $\alpha v\beta 3$ in 1 mM Mn^{2+} . We found that soluble $\alpha v\beta 3$ bound to immobilized CCL5 in a dose-dependent manner ($K_d = 6.7 \mu g/mL$) (Figure 2a). The $\alpha v\beta 3$ binding to CCL5 required Mn^{2+} , and EDTA, Mg^{2+} , and Ca^{2+} (1 mM) did not support the binding (Figure 2b). Inhibitory anti- $\alpha v\beta 3$ mAb LM609 weakly but significantly suppressed the binding of soluble $\alpha v\beta 3$ to CCL5 in 1mM Mn^{2+} (Figure 2c), indicating that the binding of CCL5 to $\alpha v\beta 3$ is specific. Previous studies showed that the disintegrin domain of ADAM15 (ADAM15 disintegrin) specifically bound to integrin $\alpha v\beta 3$ [37] and later $\alpha IIb\beta 3$ [39]. We found that ADAM15 disintegrin suppressed the binding of soluble $\alpha v\beta 3$ to the immobilized CCL5, but GST did not (Figure 2d), indicating that CCL5 specifically bound to $\alpha v\beta 3$. These findings indicate that CCL5 is a new ligand for $\alpha v\beta 3$. These findings suggest that CCL5 binds to site 1 of $\alpha v\beta 3$ in Mn^{2+} -dependent manner.

Since docking simulation predicts that CCL5 binds to site 2 of $\alpha v\beta 3$ (docking energy -20.2) (Figure 1b), we studied whether CCL5 activates soluble $\alpha v\beta 3$ by binding to site 2 in ELISA-type activation assays. To study whether CCL5 activates $\alpha v\beta 3$, soluble $\alpha v\beta 3$ was pre-incubated with CCL5 for 10 min at room temperature and then incubated with immobilized $\gamma C399tr$, a specific ligand for $\alpha v\beta 3$, in the presence of 1 mM Ca^{2+} . CCL5 enhanced the binding of soluble $\alpha v\beta 3$ to $\gamma C399tr$ in a dose-dependent manner (Figure 2e), but required high concentrations ($>1 \mu g/mL$) of CCL5 for detection. We found that CCL5 activated soluble $\alpha v\beta 3$ in 1 mM Ca^{2+} in a time-dependent manner (Figure 2f).

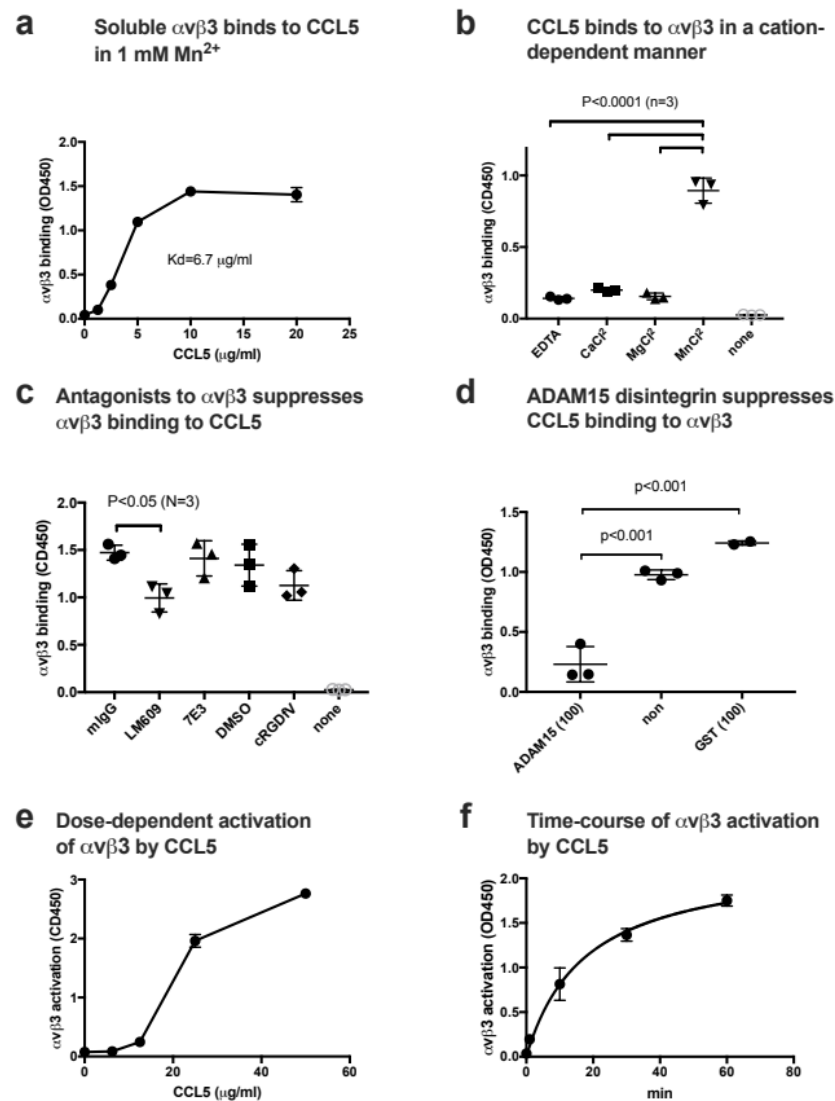


Figure 2. CCL5 binds to and activates integrin $\alpha v\beta 3$. (a) Binding of soluble integrin $\alpha v\beta 3$ to CCL5 in 1 mM Mn^{2+} in cell-free conditions. Wells of 96-well microtiter plates were coated with CCL5 and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble $\alpha v\beta 3$ (1 $\mu g/mL$) in Tyrode-HEPES buffer with 1 mM Mn^{2+} (to activate $\alpha v\beta 3$) for 1 h at room temperature. After washing the wells to remove unbound integrin, bound $\alpha v\beta 3$ was quantified using anti- $\beta 3$ mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (b) Cation dependency of $\alpha v\beta 3$ binding to CCL5. Binding of soluble integrin $\alpha v\beta 3$ to CCL5 was measured in the presence of different cations (1 mM) in ELISA-type binding assays. Data are shown as means \pm SD in triplicate experiments. (c) Effect of antagonists to $\alpha v\beta 3$ on CCL5 binding to $\alpha v\beta 3$. The concentrations of antagonists used are 10 $\mu g/mL$ for LM609 and 7E3, and 10 μM for cRGDFV. Data are shown as means \pm SD in triplicate experiments. (d) ADAM15 disintegrin, a specific ligand for $\alpha v\beta 3$, suppresses CCL5 binding to $\alpha v\beta 3$. ADAM15 disintegrin fused to GST or control GST (100 $\mu g/mL$ each) were included in the binding assay as described in (a). Data are shown as means \pm SD in triplicate experiments. (e) Activation of soluble $\alpha v\beta 3$ by CCL5 in 1 mM Ca^{2+} . Wells of 96-well microtiter plates were coated with $\gamma C399tr$, a specific $\alpha v\beta 3$ -ligand (20 $\mu g/mL$) and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble $\alpha v\beta 3$ (1 $\mu g/mL$) in Tyrode-HEPES buffer with 1 mM Ca^{2+} for 1 h at room temperature in the presence of CCL5. After removing the unbound integrin, bound $\alpha v\beta 3$ was quantified using anti- $\beta 3$

mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (f) Time-course of activation of soluble α v β 3 by CCL5. Wells of 96-well microtiter plates were coated with γ C399tr (a specific ligand for α v β 3) (20 μ g/mL) and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α v β 3 (1 μ g/mL) and CCL5 (20 μ g/mL) at room temperature in Tyrode-HEPES buffer with 1 mM Ca^{2+} for the time indicated. After washing the wells to remove unbound integrin, bound α v β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments.

3.2. CCL5 Binds to and Activates Soluble α IIb β 3

Since docking simulation predicted that CCL5 binds to site 1 and site 2 of integrin α v β 3, and bound to and activated integrin α v β 3, we studied whether CCL5 binds to and activates integrin α IIb β 3. We found that CCL5 bound to soluble α IIb β 3 in a dose-dependent manner (Figure 3a). However, heat treatment (80 °C 10 min) or specific inhibitors for α IIb β 3, anti- β 3 mAb 7E3, or eptifibatid (0.6 μ g/mL), did not affect the binding of soluble α IIb β 3 to CCL5 (Supplementary Figure S1). However, ADAM15 disintegrin fused to GST, a specific ligand to α IIb β 3, suppressed the binding of soluble α IIb β 3 to the immobilized CCL5, but control GST did not (Figure 3b), indicating that CCL5 specifically bound to α IIb β 3. These findings indicate that CCL is a new ligand for α IIb β 3.

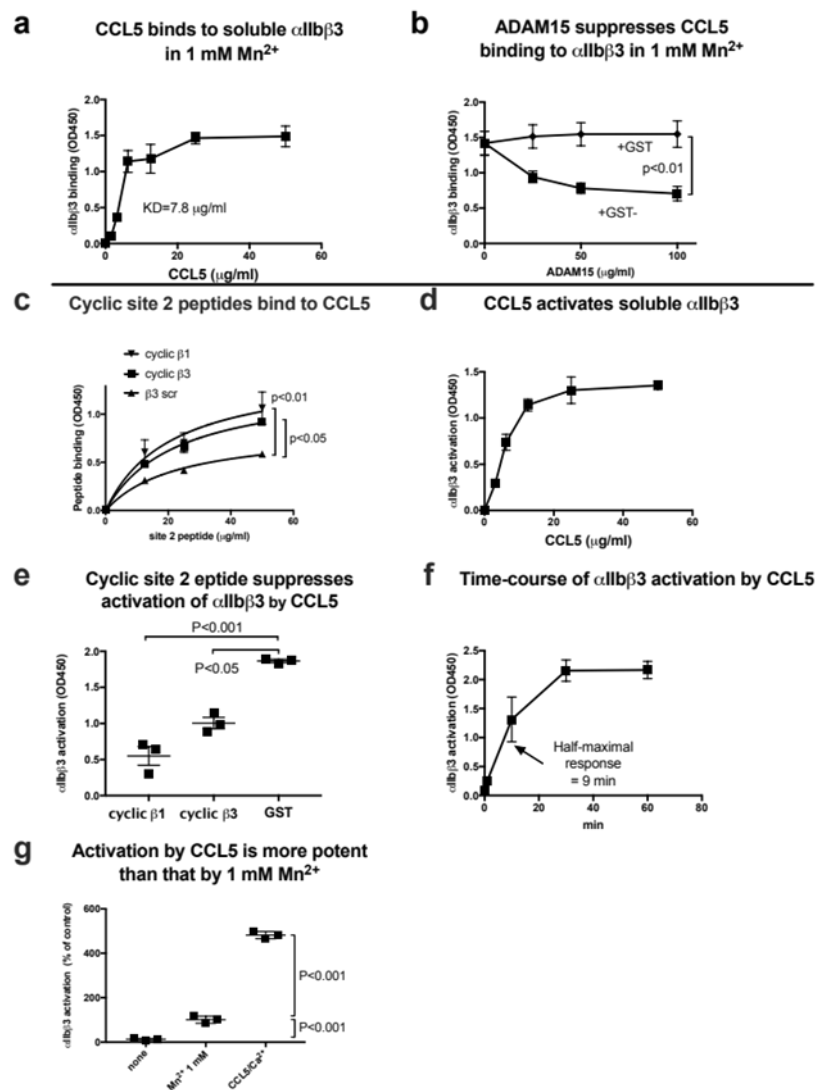


Figure 3. CCL5 binds to and activates soluble integrin α IIb β 3 in cell-free conditions. (a) Binding of soluble integrin α IIb β 3 to CCL5 in 1 mM Mn^{2+} in cell-free conditions. Wells of 96-well microtiter plates were coated with CCL5 and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) in Tyrode-HEPES buffer with 1 mM Mn^{2+} (to activate α IIb β 3) for 1 h at room temperature. After washing the wells to remove unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (b) ADAM15 disintegrin, another ligand for α IIb β 3, suppressed CCL5 binding to α IIb β 3. ADAM15 disintegrin fused to GST or control GST were included in the binding assay as described in (a). (c) Binding of cyclic site 2 peptides to CCL5. Wells of 96-well microtiter plates were coated with CCL5 (20 μ g/mL) and remaining protein binding sites were coated with BSA. Wells were then incubated with cyclic β 3 or β 1 site 2 peptide fused to GST or control β 3 scrambled peptide for 1 h at room temperature and bound site 2 peptide was quantified using HRP-conjugated anti-GST antibodies. Data are shown as means \pm SD in triplicate experiments. (d) Activation of soluble α IIb β 3 by CCL5 in 1 mM Ca^{2+} . Wells of 96-well microtiter plates were coated with γ C390-411 (the α IIb β 3-ligand peptide conjugated to GST) (20 μ g/mL) and the remaining protein-binding site were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) in Tyrode-HEPES buffer with 1 mM Ca^{2+} for 1 h at room temperature in the presence of CCL5. After removing the unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (e) Effect of site 2 peptide on α IIb β 3 activation by CCL5. Activation of α IIb β 3 by CCL5 was assayed as described in (d) except that cyclic site 2 peptides or control GST (100 μ g/mL each) were included as a competitor. Data are shown as means \pm SD in triplicate experiments. (f) Time-course of activation of soluble α IIb β 3 by CCL5. Wells of 96-well microtiter plates were coated with γ C390-411 (20 μ g/mL) and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) and CCL5 (20 μ g/mL) at room temperature in Tyrode-HEPES buffer with 1 mM Ca^{2+} for the time indicated. After washing the wells to remove unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (g) Comparison of α IIb β 3 activation by CCL5 and that by 1 mM Mn^{2+} . Soluble α IIb β 3 was activated with only 1 mM Mn^{2+} or CCL5 (50 μ g/mL) in 1 mM Ca^{2+} . Data are shown as means \pm SD in triplicate experiments.

Docking simulation predicts that CCL5 binds to the allosteric site (site 2) of α v β 3 at high affinity. We previously showed that peptides from site 2 bind to CX3CL1, CXCL12, and several integrin activators (e.g., sPLA2-IIA and CD40L), and blocked allosteric activation of integrins α v β 3, α 4 β 1 or α 5 β 1 [13,14,40], indicating that they actually bind to site 2 and binding to site 2 is required for allosteric activation. We found that cyclic site 2 peptides from β 3 and β 1 bound to CCL5, indicating that CCL5 actually binds to site 2 (Figure 3c). We thus studied if CCL5 activates soluble integrin α IIb β 3.

We previously developed ELISA-type activation assays, in which soluble integrins are incubated with ligands that are immobilized to plastic in the presence of chemokines or other activators in 1 mM Ca^{2+} (to keep integrins inactive) in cell-free conditions [13]. Integrin activation is defined as the increase in binding of soluble integrins to immobilized ligand. To measure the increase in ligand binding affinity, we need to use monovalent ligand. Pac-1 IgM specific for α IIb β 3, multivalent ligand-mimetic antibody with potential 10-binding sites, has been widely used for detecting α IIb β 3 ligand binding. Since monovalent Fab of Pac-1 IgM (kindly provided by Sandy Shattil, UC San Diego, La Jolla, CA, USA) showed very weak affinity to α IIb β 3, we generated the C-terminal residues 390-411 of fibrinogen γ -chain fused to GST (designated γ C390-411) as a ligand for α IIb β 3. α IIb β 3 binds to the C-terminal 400HHLGGAKQAGDV411 sequence of fibrinogen γ -chain C-terminal domain (γ C) [41]. We showed that soluble recombinant α IIb β 3 (activated by 1 mM Mn^{2+}) bound to γ C390-411 in ELISA-type assays in a dose dependent manner (Supplementary Figure S2). Integrin α v β 3 does not recognize this sequence for binding to γ C [36,42]. Using the whole fibrinogen γ -chain C-terminal domain (γ C151-411) [36] as

a ligand for soluble α IIb β 3 in the ELISA-type activation assay, we obtained results very similar to those with γ C390-411 (Supplementary Figure S3). Since γ C151-411 also binds to other integrins such as α v β 3 and α M β 2 [36,43], we used γ C390-411 as a ligand for α IIb β 3 throughout the entire project.

To study whether CCL5 activate α IIb β 3, soluble α IIb β 3 was pre-incubated with CCL5 for 10 min at room temperature and then incubated with immobilized γ C390-411 in the presence of 1 mM Ca^{2+} . CCL5 enhanced the binding of soluble α IIb β 3 to γ C390-411 in a dose-dependent manner (Figure 3d). We found that cyclic site 2 peptides from β 3 and β 1 effectively suppressed CCL5-induced α IIb β 3 activation (Figure 3e), indicating that CCL5 is required to bind to site 2 of α IIb β 3 to activate α IIb β 3. We found that it took >30 min to fully activate soluble α IIb β 3 by CCL5 (Figure 3f). This is consistent with the previous findings that the activation of soluble integrin α v β 3 by CXCL12 was slow and required high CXCL12 concentrations [14]. These findings suggest that activation of soluble α IIb β 3 by CCL5 does not require inside-out signaling, but requires high concentrations of CCL5 and takes >60 min to complete.

It has been assumed that 1 mM Mn^{2+} fully activates integrins [44–47]. We compared the levels of activation of soluble α IIb β 3 by CCL5 with that of 1 mM Mn^{2+} as a standard integrin activator. Unexpectedly, we found that CCL5 was much more potent (4.5-fold) than 1 mM Mn^{2+} (Figure 3g).

3.3. CXCL12 Binds to and Activates Soluble α IIb β 3

CXCL12 is another chemokines that is stored in platelet granules and rapidly transported to the surface upon platelet activation [48]. We studied if CXCL12 binds to and activates integrin α IIb β 3. We obtained results similar to that of CCL5. Briefly, CXCL12 bound to α IIb β 3 in 1 mM Mn^{2+} (Figure 4a) and this interaction was suppressed by another α IIb β 3 ligand, ADAM15 (Figure 4b). CXCL12 activated α IIb β 3 in a dose-dependent manner in cell-free conditions. A high concentration of CXCL12 (>50 $\mu\text{g}/\text{mL}$) was required to fully activate soluble α IIb β 3 (Figure 4c). Previous studies showed that activation of α v β 3, α 4 β 1 or α 5 β 1 by CXCL12 was blocked by site 2-derived peptides [14]. Activation of α IIb β 3 by CXCL12 was also suppressed by cyclic site 2 peptides (Figure 4d), indicating that CXCL12 binding to site 2 is required for α IIb β 3 activation. It took more than 60 min to fully activate soluble α IIb β 3 by CXCL12 (Figure 4e), indicating that this activation process is relatively slow. CXCL12 was more potent (3.5-fold) in activating α IIb β 3 than that of Mn^{2+} (Figure 4f). These findings suggest that CXCL12 also play a role in α IIb β 3 activation.

3.4. CX3CL1 Binds to and Activates Soluble α IIb β 3

Transmembrane CX3CL1 is not stored in platelet granules but is expressed on the surface of activated endothelial cells [6]. Previous studies showed that platelet–endothelial interaction plays a role in vascular inflammation (e.g., atherosclerosis) [25]. We hypothesized that CX3CL1 on endothelial cells mediates platelet–endothelial interaction by binding to and activating α IIb β 3. We obtained results with the chemokine domain of CX3CL1 similar to that of CCL5. Briefly, we found that CX3CL1 bound to α IIb β 3 in 1 mM Mn^{2+} (Figure 5a) and this interaction was suppressed by another α IIb β 3 ligand, ADAM15 (Figure 5b). CX3CL1 activated soluble α IIb β 3 in ELISA-type activation assays in 1 mM Ca^{2+} and high concentrations of CXCL1 (>50 $\mu\text{g}/\text{mL}$) was required to fully activate α IIb β 3 (Figure 5c). This activation was suppressed by cyclic site 2 peptides (Figure 5d), indicating that CX3CL1 binding to site 2 is required for α IIb β 3 activation. It took more than 60 min to fully activate soluble α IIb β 3 by CX3CL1 (Figure 5e), indicating that this activation process is relatively slow. CX3CL1 was several times more potent in activating α IIb β 3 than that of Mn^{2+} (Figure 5f). These findings suggest that CX3CL1 on activated endothelial cells mediates platelet–endothelial interaction by activating and binding to platelet α IIb β 3. Moreover, α IIb β 3 can be activated by soluble CX3CL1 in circulation.

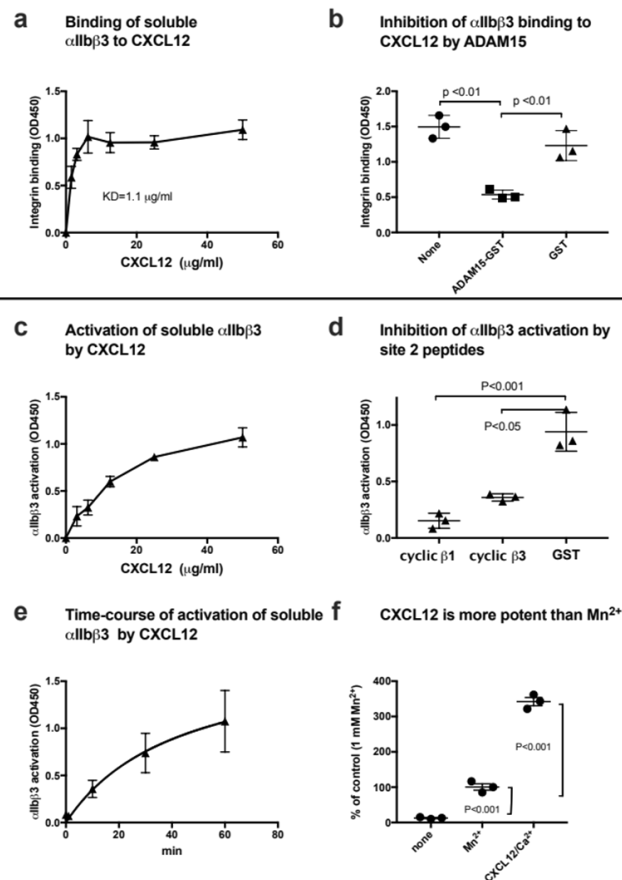


Figure 4. CXCL12 binds to and activates soluble integrin α IIb β 3 in cell-free conditions. (a) Binding of soluble integrin α IIb β 3 to CXCL12 in 1 mM Mn^{2+} in cell-free conditions. Wells of 96-well microtiter plates were coated with CXCL12 and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) in Tyrode-HEPES buffer with 1 mM Mn^{2+} (to activate α IIb β 3) for 1 h at room temperature. After washing the wells to remove unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (b) ADAM15 disintegrin, another ligand for α IIb β 3, suppressed CXCL12 binding to α IIb β 3. ADAM15 disintegrin fused to GST or control GST (100 μ g/mL each) were included in the binding assay as described in (a). Data are shown as means \pm SD in triplicate experiments. (c) Activation of soluble α IIb β 3 by CXCL12 in 1 mM Ca^{2+} . Wells of 96-well microtiter plates were coated with γ C390-411 (20 μ g/mL) and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) in Tyrode-HEPES buffer with 1 mM Ca^{2+} for 1 h at room temperature in the presence of CXCL12. After removing the unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (d) Effect of site 2 peptide on α IIb β 3 activation by CXCL12. Activation of α IIb β 3 by CXCL12 was assayed as described in (c) except that cyclic site 2 peptides fused to GST or control GST (100 μ g/mL) were included as a competitor. Data are shown as means \pm SD in triplicate experiments. (e) Time-course of activation of soluble α IIb β 3 by CXCL12. Wells of 96-well microtiter plates were coated with γ C390-411 (20 μ g/mL) and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) and CXCL12 (20 μ g/mL) at room temperature in Tyrode-HEPES buffer with 1 mM Ca^{2+} for the time indicated. After washing the wells to remove unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (f) Comparison of α IIb β 3 activation by CXCL12 and that by 1 mM Mn^{2+} . Soluble α IIb β 3 was activated with only 1 mM Mn^{2+} or CXCL12 (50 μ g/mL) in 1 mM Ca^{2+} . Data are shown as means \pm SD in triplicate experiments.

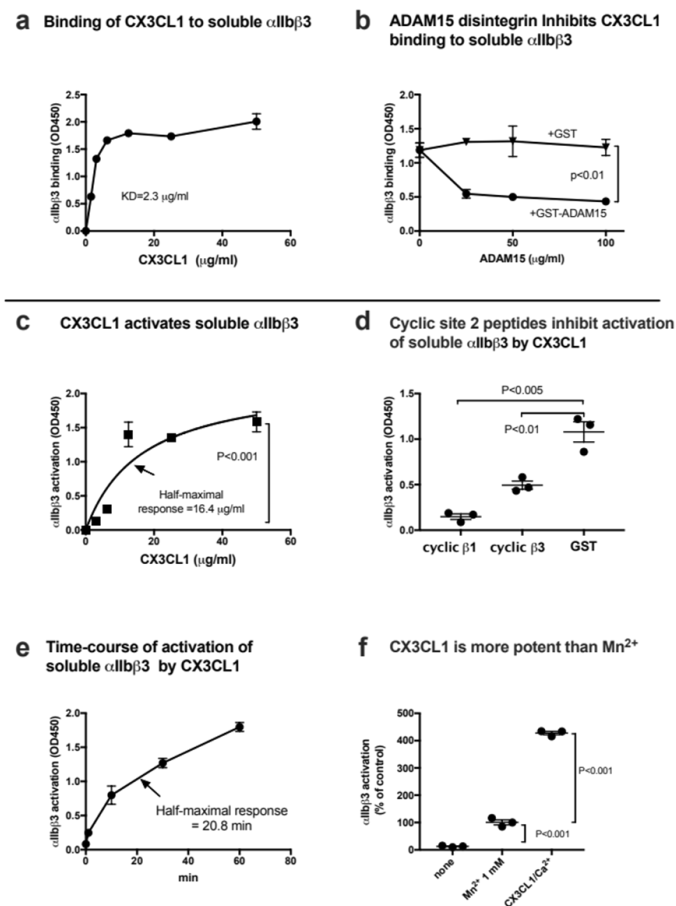


Figure 5. CX3CL1 binds to and activates soluble integrin α IIb β 3 in cell-free conditions. (a) Binding of soluble integrin α IIb β 3 to CX3CL1 in 1 mM Mn^{2+} in cell-free conditions. Wells of 96-well microtiter plates were coated with CX3CL1 and remaining protein-binding sites were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) in Tyrode-HEPES buffer with 1 mM Mn^{2+} (to activate α IIb β 3) for 1 h at room temperature. After washing the wells to remove unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (b) ADAM15 disintegrin suppressed CX3CL1 binding to α IIb β 3. ADAM15 disintegrin fused to GST or control GST were included in the binding assay as described in (a). (c) Activation of soluble α IIb β 3 by CX3CL1 in 1 mM Ca^{2+} . Wells of 96-well microtiter plates were coated with γ C390-411 (20 μ g/mL) and the remaining protein-binding site were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) in Tyrode-HEPES buffer with 1 mM Ca^{2+} for 1 h at room temperature in the presence of CX3CL1. After removing the unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (d) Effect of site 2 peptide on α IIb β 3 activation by CX3CL1. Activation of α IIb β 3 by CX3CL12 was assayed as described in (c) except that cyclic site 2 peptides (100 μ g/mL) were included as a competitor. Data are shown as means \pm SD in triplicate experiments. (e) Time-course of activation of soluble α IIb β 3 by CX3CL1. Wells of 96-well microtiter plates were coated with γ C390-411 (20 μ g/mL) and the remaining protein-binding site were blocked with BSA. Wells were incubated with soluble α IIb β 3 (1 μ g/mL) and CX3CL1 (20 μ g/mL) at room temperature in Tyrode-HEPES buffer with 1 mM Ca^{2+} for the time indicated. After washing the wells to remove unbound integrin, bound α IIb β 3 was quantified using anti- β 3 mAb (AV10) and HRP-conjugated anti mouse IgG. Data are shown as means \pm SD in triplicate experiments. (f) Comparison of α IIb β 3 activation by CX3CL1 and that by 1 mM Mn^{2+} . Soluble α IIb β 3 was activated with only 1 mM Mn^{2+} or CX3CL1 (50 μ g/mL) in 1 mM Ca^{2+} . Data are shown as means \pm SD in triplicate experiments.

3.5. CCL5, CXCL12, and CX3CL1 more Efficiently Activate Cell-Surface α IIb β 3 Than Soluble α IIb β 3

α IIb β 3 on CHO cells is not activated by platelet agonists, since intracellular machineries for inside-out activation are missing [49]. Previous studies have shown that integrin activation by CX3CL1 or CXCL12 in CHO cells is independent of their receptors (CX3CR1 and CXCR4, respectively) [13,14]. CHO cells do not express CCR3 [50]. Additionally, response to CCL5 in CHO cells required transfection of CCR1 or CCR5 [51,52]. Therefore, it is highly likely that activation of integrin α IIb β 3 by CCL5, CXCL12, and CX3CL1 in CHO cells occurs independent of their cognate receptors. We studied whether CCL5, CXCL12, and CX3CL1 activate cell-surface α IIb β 3 on CHO cells using FITC-labeled γ C390-411 in 1 mM Ca^{2+} in flow cytometry. We found that CCL5, CXCL12, or CX3CL1 markedly enhanced the binding of γ C390-411 to cell-surface α IIb β 3 (Figure 6a). Notably, the levels of activation by CCL5, CXCL12, and CX3CL1 was much higher than 1 mM Mn^{2+} (Figure 6b), which is consistent with the results in the ELISA-type activation assays. We studied the time-course of activation of cell-surface α IIb β 3 by CCL5, CXCL12, and CX3CL1. CCL5, CXCL12, and CX3CL1 activated cell-surface α IIb β 3 on CHO cells much more rapidly (half-maximal response < 1 min) (Figure 6c) than in soluble α IIb β 3. Activation of soluble α IIb β 3 by CCL5, CXCL12, or CX3CL1 was slow and required more than 60 min to reach maximum).

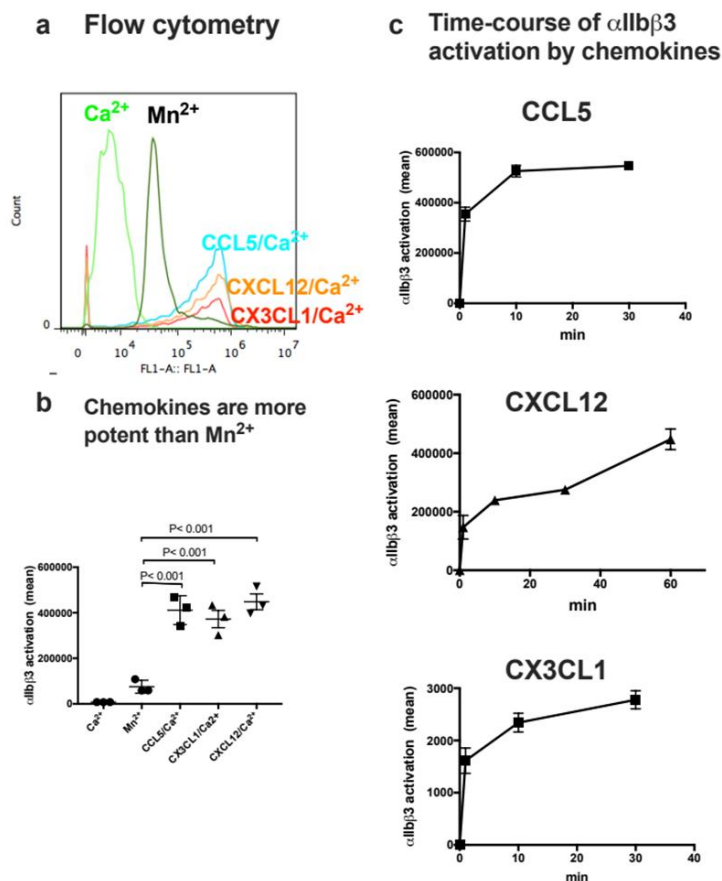


Figure 6. Chemokines activate cell-surface α IIb β 3 in 1 mM Ca^{2+} . (a) Activation of cell-surface α IIb β 3 on CHO cells. α IIb β 3-CHO cells were incubated with chemokines (50 $\mu\text{g}/\text{mL}$) for 30 min on ice and then incubated with FITC-labeled γ C390-411 for 30 min at room temperature. The cells were washed with PBS/0.02% BSA and analyzed by flow cytometer. Data are shown as mean fluorescence intensity \pm SD in triplicate experiments. (b) Comparison of activation of cell-surface α IIb β 3 by 1 mM Mn^{2+} and chemokines in 1 mM Ca^{2+} . α IIb β 3-CHO cells were incubated with chemokines as described in (a).

Data are shown as mean fluorescence intensity \pm SD in triplicate experiments. Ca^{2+} only and Mn^{2+} only are significantly different in unpaired t test ($n = 3$), $p < 0.001$. (c) Time-course of activation of cell-surface $\alpha\text{IIb}\beta 3$ on $\alpha\text{IIb}\beta 3$ -CHO cells. $\alpha\text{IIb}\beta 3$ -CHO cells were incubated with chemokines (50 $\mu\text{g}/\text{mL}$) and FITC-labeled $\gamma\text{C390-411}$ and incubated for the time indicated at room temperature. The cells were washed with PBS/0.02% BSA and analyzed by flow cytometer. Data are shown as mean fluorescence intensity \pm SD in triplicate experiments.

We studied the activation of cell-surface $\alpha\text{IIb}\beta 3$ as a function of CCL5, CXCL12, and CX3CL1 concentrations. We found that activation of cell-surface $\alpha\text{IIb}\beta 3$ was detectable at 1–10 ng/mL concentrations (Figure 7a–c). We suspect that this is probably because soluble chemokines are rapidly concentrated on the cell surface by binding to cell-surface proteoglycans [14]. These findings suggest that activation of platelet surface $\alpha\text{IIb}\beta 3$ can be efficiently activated by these chemokines in an allosteric manner in the absence of canonical inside-out signaling by these chemokines, and this process is biologically potentially important.

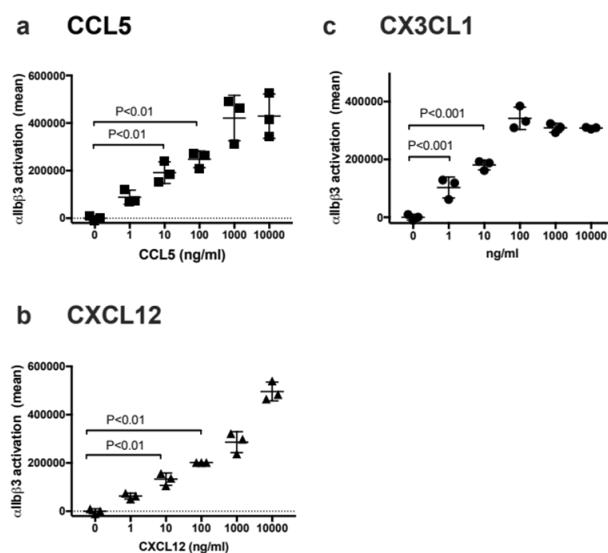


Figure 7. Dose-response of chemokine-induced activation of cell-surface $\alpha\text{IIb}\beta 3$. $\alpha\text{IIb}\beta 3$ -CHO cells were incubated with chemokines (a) CCL5, (b) CXCL12, or (c) CX3CL1 at indicated concentrations for 30 min on ice and then incubated with FITC-labeled $\gamma\text{C390-411}$ for 30 min at room temperature. The cells were washed with PBS/0.02% BSA and analyzed by flow cytometer. Data are shown as mean fluorescence intensity \pm SD in triplicate experiments.

4. Discussion

The present study establishes that CCL5 bound to site 1 of $\alpha\text{v}\beta 3$, and activated $\alpha\text{v}\beta 3$ by binding to site 2. This indicates that CCL5 is similar to CX3CL1 and CXCL12 in binding to and activating integrin $\alpha\text{v}\beta 3$ [13,14]. Next, we showed that chemokines CCL5, CXCL12, and CX3CL1 bound to platelet integrin $\alpha\text{IIb}\beta 3$, indicating that they are new ligands for $\alpha\text{IIb}\beta 3$ (Figure 8a). Furthermore, activation of soluble integrin $\alpha\text{IIb}\beta 3$ was suppressed by cyclic site 2 peptides, suggesting that the binding of these chemokines to site 2 is required. Since this activation is observed in cell-free conditions, inside-out signaling or their cognate receptors is not involved in this activation. High concentrations ($>10 \mu\text{g}/\text{mL}$) of the chemokines were required to detect activation of soluble $\alpha\text{IIb}\beta 3$ and this activation was a slow process. These chemokines activated cell-surface $\alpha\text{IIb}\beta 3$ on CHO cells, which lack machinery for inside-out signaling and cognate receptors for chemokines. In contrast to activation of soluble $\alpha\text{IIb}\beta 3$, activation of cell-surface $\alpha\text{IIb}\beta 3$ can be detected at very low concentrations of chemokines (1–10 ng/mL) and much more quickly (half maximal response <1 min) than that of soluble $\alpha\text{IIb}\beta 3$, probably because soluble chemokines can be quickly concentrated on the cell surface

by binding to cell-surface proteoglycans. These findings are the first evidence that platelet integrin $\alpha\text{IIb}\beta_3$ can be efficiently activated by chemokines independent of canonical inside-out signaling. We propose that this is one of the mechanisms for enhancing ligand binding affinity of $\alpha\text{IIb}\beta_3$, which is distinct from the canonical inside-out signaling (Figure 8b). CCL5 and CXCL12, which are stored in platelet granules, and CX3CL1 on the surface of activated endothelial cells (Figure 8c), and those in circulation (e.g., during inflammation and cytokine storm) (Figure 8b) may possibly contribute to $\alpha\text{IIb}\beta_3$ activation. We propose that chemokine binding to site 2 of $\alpha\text{IIb}\beta_3$ is a potential target for drug discovery and site 2 peptides have potential as therapeutics for thrombosis.

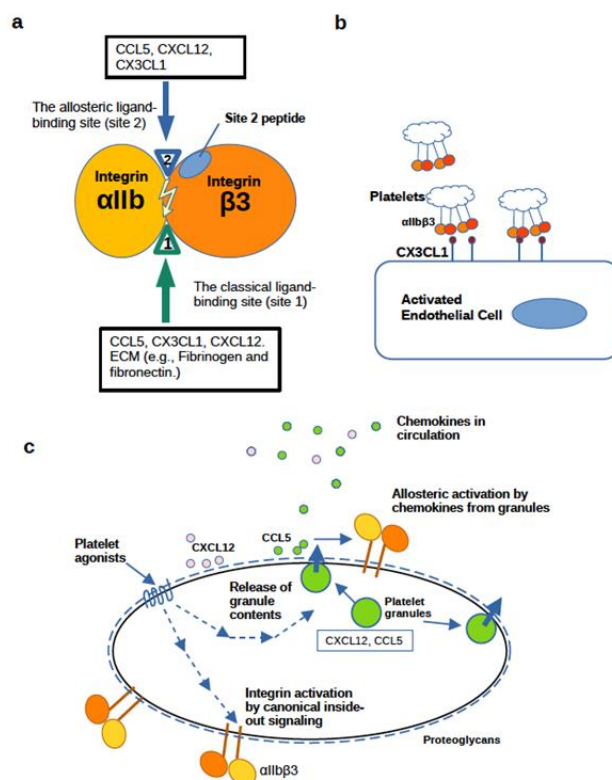


Figure 8. Possible biological role of allosteric activation of $\alpha\text{IIb}\beta_3$ by CCL5, CXCL12, and CX3CL1. (a) A model of the binding of CCL5, CXCL12, and CX3CL1 to the classical ligand binding site (site 1) and to the allosteric site (site 2) in integrin $\alpha\text{IIb}\beta_3$. CCL5, CXCL12, and CX3CL1 bind to site 1 and activate integrins by binding to site 2. The site 2 peptide inhibits integrin activation by chemokines. (b) Activation of $\alpha\text{IIb}\beta_3$ is induced by CXCL12 or CCL5 stored in platelet granules upon platelet activation or circulating CCL5, CXCL12, or CX3CL1 overproduced during inflammation. Several chemokines are stored in platelet granules and are transported to the platelet surface upon platelet activation by platelet agonists (e.g., thrombin, collagen, and ADP). Platelet integrin $\alpha\text{IIb}\beta_3$ is inactive in resting platelets but quickly activated upon platelet activation. This activation is mediated by canonical inside-out signaling. In the present study we showed that the ligand-binding affinity of $\alpha\text{IIb}\beta_3$ is enhanced by the binding of chemokines to site 2. Note that transfer of granular contents to the surface is mediated by inside-out signaling by platelet agonists. Levels of pro-inflammatory chemokines in circulation increase during inflammation and they may also allosterically activate $\alpha\text{IIb}\beta_3$. This may be a missing link between inflammation and thrombosis. (c) Potential role of CX3CL1 in platelet–endothelial cell interaction. Platelet–endothelial cell interaction is involved in vascular inflammation (e.g., atherosclerosis). Transmembrane CX3CL1 is expressed on endothelial cells activated by pro-inflammatory cytokines (e.g., IL-1 β and TNF α). CX3CL1 on the cell surface can activate $\alpha\text{IIb}\beta_3$ by binding to site 2 and then support platelet–endothelial interaction by binding to site 1.

Since activation of integrins by binding of chemokines to site 2 (allosteric activation) enhanced the binding of monovalent ligand to integrins, we propose that site 2-mediated allosteric integrin activation enhanced ligand affinity of α IIb β 3. It is possible that inside-out signaling induces integrin binding to multivalent ligands (e.g., integrin clustering) and site 2-mediated allosteric binding enhances ligand affinity. Since transport of platelet granules and their contents to the surface is known to require inside-out signaling, allosteric activation by chemokines also requires inside-out signaling. It is imperative to determine the role of each pathway in the future studies.

Binding of integrins to their ligands requires the presence of divalent cations, and different cations can markedly alter integrin affinities to fibronectin [44], and Mn^{2+} produced the most striking increase in integrin ligand affinity compared with other divalent cations (Mg^{2+} or Ca^{2+}) in a wide variety of integrins. Subsequently, Mn^{2+} has been widely used as a positive control for integrin activation. Mn^{2+} -induced integrin activation was thought to mimic physiologic integrin activation because Mn^{2+} activates integrins in the absence of a bound ligand and induces similar epitope exposure [53]. It is surprising that CX3CL1, CXCL12, and CCL5 are more effective in activating α IIb β 3 than 1 mM Mn^{2+} . It is likely that α IIb β 3 is less sensitive to Mn^{2+} than other integrins. It is possible to use chemokines as the standard of α IIb β 3 activation in future studies.

It is unclear if α IIb β 3 activation mediated by the binding of chemokines site 2 requires global conformational changes in α IIb β 3. Previous studies have shown that sPLA2-IIA allosterically activated integrins α v β 3, α 4 β 1, and α 5 β 1 [37]. sPLA2-IIA did not change reactivity to activation-dependent anti- β 1 antibodies HUTS4 and HUTS21 [54,55]. In our preliminary experiments, chemokines did not change reactivity of conformation-dependent anti- β 3 integrin antibody LIBS2 [56] in 1 mM Ca^{2+} (Supplementary Figure S4). It is thus possible that allosteric activation of α IIb β 3 by chemokines may not induce global conformational changes as in allosteric activation of β 1 integrins [40]. It has recently been reported that the binding of allosteric activating anti β 1 mAb TS2/16 to integrin α 6 β 1 did not change the 3D structure of the integrin at all [57]. It would be important to study whether the binding of chemokines to site 2 induces changes in 3D structure of integrins in future experiments.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/cells11193059/s1>, Figure S1: Effect of antagonists to α IIb β 3 on chemokine binding to α IIb β 3; Figure S2: Chemokines enhanced binding of soluble α IIb β 3 to immobilized fibrinogen fragment γ C390-411 fused to GST; Figure S3: Chemokines enhanced binding of soluble α IIb β 3 to immobilized γ C151-411 peptide fused to GST; Figure S4: Activation of α IIb β 3 on CHO cells (α IIb β 3-CHO cells) by chemokines does not induce LIBS2 epitopes.

Author Contributions: Conceptualization, Y.T.; data curation, Y.K.T.; writing—original draft, Y.T.; writing—review and editing, M.F. All authors have read and agreed to the published version of the manuscript.

Funding: This project was supported by Pilot funding from Comprehensive Cancer Center at UC Davis School of Medicine. This work is partly supported by the UC Davis Comprehensive Cancer Center Support Grant (CCSG) awarded by the National Cancer Institute (NCI P30CA093373).

Acknowledgments: We thank Barry Collar (Rockefeller University) for insightful discussion, and Sandy Shattil (University of California San Diego) for reagent. We also thank Samuel Hwang (UC Davis) for the access to flow cytometry.

Conflicts of Interest: The authors declare that they have no conflicts of interest with the contents of this article.

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