



# Latent Toxoplasmosis Effects on Rodents and Humans: How Much is Real and How Much is Media Hype?

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ABSTRACT Toxoplasma gondii is a ubiquitous, intracellular protozoan parasite with a broad range of intermediate hosts, including humans and rodents. In many hosts, T. gondii establishes a latent long-term infection by converting from its rapidly dividing or lytic form to its slowly replicating and encysting form. In humans and rodents, the major organ for encystment is the central nervous system (CNS), which has led many to investigate how this persistent CNS infection might influence rodent and human behavior and, more recently, neurodegenerative diseases. Given the interest in this topic, here we seek to take a global approach to the data for and against the effects of latent T. gondii on behavior and neurodegeneration and the proposed mechanisms that might underlie behavior modifications.

KEYWORDS Toxoplasma gondii, behavior, central nervous system infections, human, murine, neurodegeneration

oxoplasma gondii is a common, intracellular protozoan parasite in the Apicomplexa family and is found worldwide. While T. gondii only undergoes sexual reproduction in felids, most warm-blood animals, including humans and rodents, are natural intermediate hosts for T. gondii (1). Most intermediate hosts ingest T. gondii as either a sporozoite, which is contained in the oocysts shed by newly infected cats, or a bradyzoite, which is the tissue-persistent form of the parasite. Once ingested, parasites convert to the fast-replicating asexual form, the tachyzoite, which disseminates throughout the host and is controlled by the immune response. In certain organs (e.g., skeletal muscle or central nervous system [CNS]), T. gondii switches to its bradyzoite form, which is characterized by slow replication and encystment. Intracellular bradyzoite-filled cysts evade the immune response and have the potential to persist for the entire life of the host (2-5). Like other intermediate hosts, humans and rodents primarily become infected with T. gondii through ingestion of contaminated food and water, although humans can also pass T. gondii vertically or through organ transplantation (6). In humans and rodents, the major organ for encystment is the CNS (7, 8).

While T. gondii is established to cause symptomatic disease in immunocompromised hosts (e.g., toxoplasmic encephalitis in AIDS patients [9-12]) and occasionally in immunocompetent hosts (e.g., mono-like syndrome, chorioretinitis), the parasite's ability to cause a latent long-term CNS infection has generated continued interest in understanding how this persistent CNS infection might influence host behavior and outcomes. The potential for parasite-induced behaviors is especially appealing given the recognition that other parasites cause host behavioral changes (e.g., parasite-driven fish behavior or "the zombie ant fungus," Ophiocordyceps unilateralis) (13, 14). While a number of studies have investigated this possibility, with reports in both humans and rodents linking T. gondii infection to a wide range of behavioral modifications and,

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TABLE 1 List of tested behaviors and methods<sup>a</sup>

Behavior	Test method
Motor activity	Deep maze
	Social interaction test
	Open field
	Elevated plus maze
Memory	Deep-maze retention
	Spontaneous alteration in Y maze
	Barnes maze short-term memory
	STFP <sup>b</sup> , olfactory memory
	Morris water maze
	Novel object recognition
	Object placement
Fear	Cat urine, cat fur
	Bobcat urine versus mink or rabbit urine
	In square pen or rectangular arena

<sup>a</sup>For a comprehensive overview, see Worth et al. (17).

<sup>b</sup>STFP, Social transmission of food preference.

more recently, neurodegenerative disease, conflicting data also exist. These contradictory findings make it unclear as to what extent *T. gondii* affects host behavior or secondary disease. Below, we will review a sample of key studies that exemplify the data for and against latent *T. gondii* to modify host behavior and outcomes.

## T. GONDII EFFECTS ON RODENT MODELS

A variety of behavioral abnormalities has been reported in *T. gondii*-infected rodents. Memory deficits were the first behavioral abnormality linked to *T. gondii*, beginning with an observational study in wild rats, which was later substantiated by experimental studies in both mice and rats (15–17). Motor abnormalities have also been reported, which include both increased activity (18–25) and decreased activity (16, 23, 26–29). Finally, infected rodents have also shown diminished fear responses as assessed by several assays, including (i) loss of aversion to cat urine (22, 30–34), (ii) decreased fear of open spaces (18, 35), and (iii) increased likelihood of eating novel food (20). Tests used for memory, fear, and motor abnormalities are listed in Table 1. Decreased fear, especially to cat urine, has been postulated to increase the chance of predation of rodents by the definitive host, thus offering a potentially plausible reason for *T. gondii* infection to alter this behavior.

While these studies suggesting a strong effect of *T. gondii* infection on behavior have garnered attention in both the scientific community and in the general press, it is important to note that negative studies also exist. A subset of studies reported no changes in memory (22, 29, 32, 36), no motor abnormalities (20–22, 26–28, 32, 36), and no fear differences associated with *T. gondii* infection (22, 23, 31, 32). Even within those studies reporting positive results, the results are more heterogenous than often recognized, with most studies reporting a mix of both positive and negative results. For instance, one study found that infected rats that showed decreased fear in one context (feline urine) had normal fear responses in another scenario (feline fur odor) (32). In addition, a more recent study reported a lack of fear response to predator and nonpredator urine, suggesting that this behavioral modification might be more generalized and not based on predation risks (37).

A range of factors—including how a given behavior was assayed or when behavior testing was performed—may account for these contradictory findings. For example, a number of different tasks have been used to assess memory performance with variable results. These tasks include performance on the deep maze (16), Barnes maze (36), Y maze (22), and the Morris water maze (32). In addition, the genotype of the infecting *T. gondii* strain may also affect behavioral outcomes. For example, a study that utilized two different type II/haplotype 2 strains (Me49 and Prugniaud [Pru]) that differ genetically only by <0.5% (38) identified memory impairments in rodents infected with Me49

but not with Pru (22). Similarly, a variety of rodent strains have been used, which might also impact behavioral phenotypes (see neurotransmitter studies below). Studies that reported a lack of aversion to cat urine were conducted between 4 and 16 weeks postinfection (23, 32, 39), while negative studies that reported normal aversion ranged from 4 weeks to as long as 28 weeks postinfection (22, 23, 31, 32). Finally, the method through which rodents were infected (e.g., intraperitoneally, congenitally, or orally) also fluctuates between studies, which in turn can affect host immune response and thus, potentially, outcomes of infection (40). Overall, such study heterogeneity likely contributes to the conflicting behavioral outcomes.

In addition to T. gondii's effect on rodent behavior, a small number of studies examined T. gondii's effect on secondary CNS insults. Mice infected with type II (Me49) T. gondii had decreased stroke sizes compared to those in uninfected mice. This protection against stroke was thought to be secondary to a diminished stroke-related immune response and increased oxidative capacity in chronically infected mice (41). More recently, three studies in three different Alzheimer's disease mouse models that overexpress mutant human amyloid precursor protein determined that infection with a type II strain (Me49 in two studies, Pru in one study) was protective against amyloid beta (A $\beta$ ) deposition (42–44). One of the studies also documented prevention of memory deficits (42) and one study showed that protection appeared to be strain specific as infection with a type III strain did not decrease A $\beta$  deposition (44). The postulated mechanisms for this protection include the induction of anti-inflammatory cytokines (interleukin 10 [IL-10] and transforming growth factor  $\beta$  [TGF- $\beta$ ]) (42) and an increase in infiltrating phagocytic immune cells (43) in infected mice. In summary, in a very limited number of studies in mice, type II infection appears to be protective against secondary CNS insults, suggesting that we may be able to use T. gondii infection to find new pathways for treating other neurologic diseases.

#### **MECHANISMS BY WHICH T. GONDII MIGHT AFFECT RODENT BEHAVIOR**

Though the extent to which *T. gondii* infection universally influences rodent behavior is unclear, several mechanisms have been proposed to account for the observed behavioral changes. Possible mechanisms include variations in secreted neurotransmitters, the neuroanatomic location of *T. gondii* cysts, or modulation of the CNS by the neuroinflammatory response (34, 45).

The modulation of neurotransmitters represents an obvious mechanism for altering behavior, as these molecules are responsible for the cell-to-cell communication that drives CNS circuits and behavior. An early study focused on examining changes in dopamine (DA), norepinephrine (NE), and serotonin (5-HT) in acute and chronically infected mice because these neurotransmitters are involved in learning, memory, fear modulation, and mood (46-48). The study found that NE decreased in an acute infection and DA increased in chronically infected mice (49). These findings were replicated by a more recent study that reported similar changes in neurotransmitter levels in whole brain homogenates as well as positive behavioral changes in mice, though it is unclear if the neurotransmitter levels were measured in the same mice that underwent behavioral testing (50). One proposed mechanism for these changes is a downregulation of dopamine  $\beta$ -hydroxylase (DBH), an enzyme that converts DA to NE. Consistent with this hypothesis, in a recent study, NE protein and DBH mRNA were decreased in male rats, but not female rats, though neither group showed behavior changes compared to uninfected rats. The study then moved to mice, where infected male mice displayed behavioral abnormalities and decreased DBH mRNA, while infected female mice had no changes in DBH mRNA; it is unclear if behavior changes were observed in the infected female mice. In addition, the relevant neurotransmitter levels (i.e., DA and NE) were not assayed in the male or female mice (51). Another study, investigating DA, DA metabolites, NE, and 5-HT found increased levels of DA metabolites (but not DA itself) and NE in the cortices of infected mice as well as decreased 5-HT in the amygdala (52). In this study, DA metabolite levels were correlated with freezing behaviors. It is important to note, however, that a separate study that used the

same strain of *T. gondii* but a different mouse strain found no behavior abnormality and no difference in DA levels (53). Given that immune cells make and release DA (54), the opposing outcomes between these studies may arise from the positive study using a mouse strain that develops high levels of neuroinflammation (C57BL/6 mice) while the negative study used a mouse strain that generates very little neuroinflammation (CD-1 mice). Finally, a single study has shown that glutamate levels are increased in infected mice secondary to a decrease in expression of astrocytic GLT-1, the transporter that removes glutamate released by neurons. In this study, ceftriaxone treatment restored levels of astrocytic GLT-1 and normalized brain glutamate levels but had no effects on *T. gondii* cyst burden, the neuroinflammatory response, or the identified behavioral abnormalities (35). These findings suggest that glutamate, the major excitatory neurotransmitter in the cortex, does not drive the *T. gondii*-associated behavioral abnormalities identified in this study.

The second commonly postulated mechanism for *T. gondii*-associated rodent behavioral changes is that the prevalence and neuroanatomical location of cysts dictate both the type and magnitude of behavioral changes that accompany infection. Again, this hypothesis makes intuitive sense, as different brain regions mediate specific functions (e.g., hippocampus and memory; occipital cortex and vision). Despite its appeal, there is very little evidence to support this hypothesis: cyst location remains highly variable from mouse to mouse, with no preponderance of cysts in brain regions implicated in behavioral changes (e.g., motor cortex or limbic system) (55). In addition, a lack of fear aversion to feline urine was reported in mice infected with a *T. gondii* strain that does not produce cysts (39), making the cyst hypothesis very unlikely as a cause of the altered behaviors in those mice. Finally, a very recent study showed that the administration of the drug guanabenz normalized infection-associated hyperactivity in infected C57BL/6 mice without affecting cyst number (25).

A related hypothesis is that cysts cause a local increase in DA, which in turn drives behavioral changes. This idea arose because *T. gondii* contains two amino hydroxylases, AAH1 and AAH2, which could convert tyrosine to L-dopa, the rate limiting step in DA synthesis (56, 57). AAH1 is an essential gene that is expressed in both tachyzoites and bradyzoites, while AAH2 is expressed only in bradyzoites and amenable to gene deletion. Three studies using two independently generated AAH2 knockout (KO) strains have laid this theory to rest, as all three studies found that mice infected with either parental or AAH2 KO *T. gondii* parasites showed the same behavioral changes or neurotransmitter changes regardless of the infecting *T. gondii* strain (24, 53, 58).

A final common hypothesis for *T. gondii*-associated behavioral change is that the neuroinflammatory response drives these changes. This possibility is consistent with the previously mentioned study that reported behavioral changes in association with a *T. gondii* strain that does not form cysts but does provokes a neuroinflammatory response (39). This hypothesis is also consistent with the guanabenz study. In this study, which used both BALB/c and C57BL/6 mice, drug treatment rescued *T. gondii*-associated behavioral deficits and decreased the neuroinflammatory response in both mouse strains but only decreased the cyst burden in BALB/c mice while increasing the cyst burden in C57BL/6 mice (25).

### T. GONDII AND HUMANS

The first study to link human behavior to *T. gondii* seropositivity found that men positive for IgG antibodies fell within distinct personality categories compared to those for uninfected controls (59). Since the original study, well over 200 papers have been published on *T. gondii* infection and its influence on human behavior and secondary disease. Before we review the data, we will first outline several caveats that are important to consider when evaluating these studies. We will then briefly review the data correlating *T. gondii* infection to personality and risky behavior, mental health disorders (e.g., schizophrenia), neurodegeneration (e.g., Parkinson's disease), and cognition.

While rodent studies allow one to experimentally manipulate which animals are

infected, human studies rely on using serologic testing (e.g., antibodies to *T. gondii*) to identify latently infected persons and then compare a given behavior between the seropositive and seronegative groups. Consequently, such approaches only allow for investigations of the correlation between *T. gondii* infection (or really, seropositivity) and behavior but cannot directly assess the causal role that infection may play in a given behavior. This difference between correlation and causality is important, because it raises a key question: does a given behavior or disease increase the likelihood of *T. gondii* infection rather than the other way around? At present, the published studies cannot address this critical question. In addition, serology (IgG positivity usually) does not distinguish prior exposure from ongoing chronic infection, a point highlighted by an autopsy study which found no histopathologic evidence of *T. gondii* in seropositive patients with behavioral abnormalities (60).

Some of the human studies are also hampered by other limitations inherent to their design. For instance, many utilize survey responses and voluntary correspondence, which decrease the sample size and may lead to sampling bias. Additionally, in places with low *T. gondii* seropositivity, large statistical effects can be driven by very few subjects. Reporting bias may also be a problem, as small studies with positive findings are more numerous, while negative findings are largely only published from large census data. Furthermore, many of these studies do not perform multivariate analyses or ensure that the baseline groups are matched. Finally, akin to the limitations described for the rodent studies, there is marked heterogeneity in the behavioral assays employed in investigating *T. gondii*-associated behavioral or cognitive change in humans.

**Personality and risky behavior.** Differences in personality types in *T. gondii*seropositive individuals were the first reported change in otherwise healthy individuals, leading to a multitude of follow-up studies (59, 61–67). Since then, various other behaviors have been associated with *T. gondii* infection, including increased aggression (65, 68), increased risk for motor vehicle accidents (65, 69–72), decreased reaction time (73, 74), and excessive ethanol consumption (75). While risky behavior is a popular subject (76, 77), publication bias is likely a contributing factor, and most studies lack multifactorial analyses. Notably, in a population-representative birth cohort that examined poor impulse control, personality, and cognition, only suicide maintained a statistically significant correlation with *T. gondii* seropositivity (78).

Mental health. While T. gondii seropositivity has been studied in association with a number of mental health disorders, the association between T. gondii seropositivity or serointensity (the amount of IgG antibody) (79) and schizophrenia has been the most studied and publicized. While most studies focus on seroprevalence, it is often the case that if no correlation is found between seroprevalence and schizophrenia, serointensity is then considered and reported (80). A recent meta-analysis by Sutterland et al. (77) examined the association between T. gondii and mental health disorders. In this analysis, studies were only included if the original publication was in English, French, or Dutch; healthy subjects were used as controls, patients were diagnosed according to the diagnostic and statistical manual of mental disorders, third edition (DSM-III) or a higher standard, and seropositivity was determined by well-established diagnostic tests. Of the 50 studies that met these criteria, 42 were for schizophrenia. Overall, they found a significant association between T. gondii seroprevalence and schizophrenia (odds ratio [OR], 1.81) but noted that serointensity was heterogenous and results were region specific (77). Considerations for age (seroprevalence generally increases with age) and publication bias lowered the OR to 1.43, meaning that those who were seropositive for T. gondii IgG were 1.43 times more likely to also be diagnosed with schizophrenia than seronegative controls. For context, a maternal family history of schizophrenia has an OR of 9.31, paternal age >54 years has an OR as high as 5.92, and cannabis use has a reported OR of 2.93 (81). As noted above, these studies cannot determine if T. gondii infection caused or increased the likelihood of developing

schizophrenia or if high-risk behaviors associated with schizophrenia increase the likelihood of becoming infected with *T. gondii*.

In addition to schizophrenia, bipolar disorder has also been commonly studied in association with *T. gondii* seropositivity, though there are far fewer published studies. In a recent meta-analysis of 118 studies on *T. gondii* and bipolar disorder, only 8 met their criteria: original articles, published from January 1980 to June 2015, in English, Portuguese, or Spanish, and measured *T. gondii* serology. This study concluded that patients with bipolar disorder were more likely to be infected by *T. gondii* but with a low OR of 1.26, which was likely driven by the substantial overlap in *T. gondii* seropositivity prevalence in the included studies: 15.6% to 95.3% in bipolar patients and 16.3% to 87.3% in controls (82).

**Neurodegeneration and cognition.** The association between *T. gondii* and dementia has been investigated in Parkinson's disease (PD), Alzheimer's disease (AD), and overall cognition. Several publications reported a positive correlation between PD and *T. gondii* seropositivity (83, 84), although they were quickly followed by negative studies (85, 86). In a recent meta-analysis on PD and *T. gondii* seropositivity, Zhou et al. included seven studies with 1,086 subjects total and found no correlation between PD and *T. gondii* seropositivity (87). This meta-analysis highlighted that the initial positive studies for PD had small sample sizes and likely some degree of sampling bias related to the geographical region in which the studies were conducted.

The investigation between Alzheimer's disease (AD) and *T. gondii* has been driven, in part, by the idea that CNS pathogens may be drivers or potentiators of AD pathology. This concept arose for several reasons: (i) the neuropathological findings of AD (neurofibrillary tangles and amyloid beta plaques) can be seen in CNS infections; (ii) amyloid-beta has been shown to have antimicrobial properties; and (iii) DNA from a variety of pathogens has been found in the brains of AD patients (88, 89). Collectively, these findings suggested that amyloid beta accumulation might occur in response to pathogens and then cause deleterious cognitive effects.

A recent meta-analysis reviewed 8 AD studies for a total of 3,239 subjects across seven countries (90). They excluded all study types except for cross-sectional (n = 2) and case-control (n = 6) studies. Only two of the eight studies reported a significance in *T. gondii* seroprevalence and AD. One of the positive studies had a very high OR (49.9), but this high OR was driven by the fact that the study included only 2 patients with AD, both of whom were *T. gondii* seropositive, while the control population (n = 180) had a seroprevalence rate of 9%. The remaining studies had ORs ranging from 0.94 to 5.75 with *P* values that ranged from 0.18 to 0.99. However, the combined analysis of all 8 studies resulted in a common OR of 1.53, though the authors note that this OR may be influenced by a publication bias for positive results.

Finally, we will conclude with a review of the National Health and Nutrition Examination Survey (NHANES) III from the Centers for Disease Control because of the size and long-term follow-up of the studied cohort. This cohort has been used in multiple studies correlating *T. gondii* serology to cognition. A variety of cognitive tests were recorded from 4,178 participants, ages 20 to 59 years, from 1966 to 1994, in which 19.1% were seropositive for *T. gondii*. The breadth of this cohort allowed for many correlations between host behaviors and other factors with *T. gondii* seropositivity. These studies nicely illustrate the impact of multivariate analyses. While at first glance, seropositivity was correlated with decreased cognitive performance in the total population, when a multivariate analysis was performed, the positive correlation became smaller and was more associated with socioeconomic and immigration status (91, 92). Multivariate analysis allows us to have additional insights into what other factors may be driving the correlation with *T. gondii* seropositivity and the NHANES studies demonstrate the utility of rigorously designed and analyzed studies.

In summary, the validity of seropositivity, population incidence rates, study design limitations, and publication bias all must be considered when evaluating the link between *T. gondii* infection and human behavior and disease.

### DISCUSSION

Toxoplasma gondii is a unique parasite in that it causes a long-term latent infection in the CNS of humans and rodents. As such, much work has been devoted to determining if infection with *T. gondii* can lead to changes in rodent and human behavior and cognition and, more recently, neurodegeneration. The rodent work shows heterogeneity, with studies suggesting a positive linkage and studies showing no linkage, a finding that is also true for studies investigating the mechanisms postulated to underlie these behavioral changes—alterations in neurotransmitter release, cyst location, and neuroinflammation. As discussed in detail above, this heterogeneity may arise from the many important differences between these studies, which include differences in the behavioral assays, the timing of the behavioral assays, and the *T. gondii* and rodent strains used. Collectively, these data suggest that *T. gondii* infection does not universally cause changes in rodent behaviors and that the underlying mechanism(s) for the observed behavioral changes remains unclear.

Akin to the rodent work, the human studies also show a great deal of heterogeneity. Unlike the rodent work, in which causality can be established, human studies rely on correlating T. gondii seropositivity and behavioral or cognitive outcomes. In addition to the lack of causality, many of the human studies are confounded by issues such as small sample sizes and a lack of multivariate analyses. Overall, these conflicting data and study limitations make drawing definitive conclusions difficult, though the relatively low ORs identified in meta-analyses suggest that T. gondii's effect on human behavior or disease may be quite mild. Consistent with T. gondii playing a limited role in modifying human behavior and disease, prevalence rates of potentially T. gondiiassociated diseases (e.g., schizophrenia and Alzheimer's disease) do not vary across countries with high and low T. gondii seropositivity (93-95). In fact, among developed countries, the United States has one of the highest prevalence and incidence rates for AD at age 65, with 4.9% and 10.5%, respectively, but one of the lower rates of T. gondii seropositivity (~10%) (94, 95). In summary, the current data suggest that if T. gondii influences human behavior or disease, the effect is likely subtle and/or maybe highly dependent on the genetic background of the individual or the context of the infection (e.g., T. gondii strain type, route of infection, and how long the individual has been infected).

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#### REFERENCES

- 1. Dubey JP. 2010. Toxoplasmosis of animals and humans. CRC Press, Boca Raton, FL.
- Kim S-K, Boothroyd JC. 2005. Stage-specific expression of surface antigens by *Toxoplasma gondii* as a mechanism to facilitate parasite persistence. J Immunol 174:8038–8048. https://doi.org/10.4049/jimmunol.174 .12.8038.
- Pittman KJ, Knoll LJ. 2015. Long-term relationships: the complicated interplay between the host and the developmental stages of *Toxoplasma gondii* during acute and chronic infections. Microbiol Mol Biol Rev 79:387–401. https://doi.org/10.1128/MMBR.00027-15.
- Jeffers V, Tampaki Z, Kim K, Sullivan WJ. 2018. A latent ability to persist: differentiation in *Toxoplasma gondii*. Cell Mol Life Sci 75:2355–2373. https://doi.org/10.1007/s00018-018-2808-x.
- Salvioni A, Belloy M, Lebourg A, Bassot E, Cantaloube-Ferrieu V, Vasseur V, Blanié S, Liblau RS, Suberbielle E, Robey EA, Blanchard N. 2019. Robust control of a brain-persisting parasite through MHC | presentation by infected neurons. Cell Rep 27:3254–3268. https://doi.org/10.1016/j.celrep .2019.05.051.
- Dabritz HA, Conrad PA. 2010. Cats and *Toxoplasma*: implications for public health. Zoonoses Public Health 57:34–52. https://doi.org/10.1111/j .1863-2378.2009.01273.x.

- Remington JS, Cavanaugh EN. 1965. Isolation of the encysted form of *Toxoplasma gondii* from human skeletal muscle and brain. N Engl J Med 273:1308–1310. https://doi.org/10.1056/NEJM196512092732404.
- Alvarado-Esquivel C, Sánchez-Anguiano LF, Mendoza-Larios A, Hernández-Tinoco J, Pérez-Ochoa JF, Antuna-Salcido El, Rábago-Sánchez E, Liesenfeld O. 2015. Prevalence of *Toxoplasma gondii* infection in brain and heart by immunohistochemistry in a hospital-based autopsy series in Durango, Mexico. Eur J Microbiol Immunol (Bp) 5:143–149. https://doi.org/10.1556/1886 .2015.00014.
- Roth A, Roth B, Höffken G, Steuber S, Khalifa KI, Janitschke K. 1992. Application of the polymerase chain reaction in the diagnosis of pulmonary toxoplasmosis in immunocompromised patients. Eur J Clin Microbiol Infect Dis 11:1177–1181. https://doi.org/10.1007/bf01961141.
- Porter SB, Sande MA. 1992. Toxoplasmosis of the central nervous system in the acquired immunodeficiency syndrome. N Engl J Med 327: 1643–1648. https://doi.org/10.1056/NEJM199212033272306.
- 11. Luft BJ, Remington JS. 1992. Toxoplasmic encephalitis in AIDS. Clin Infect Dis 15:211–222. https://doi.org/10.1093/clinids/15.2.211.
- Vijaykumar BR, Lekshmi SU, Sai Kant R, Vaigundan D, Mahadevan A, Rajendran C, Shankar SK, Jayshree RS. 2016. Genetic characterization of *Toxoplasma gondii* from autopsy proven cases of AIDS associated cere-

bral toxoplasmosis in South India. Infect Genet Evol 39:106–112. https:// doi.org/10.1016/j.meegid.2016.01.014.

- Barber I, Hoare D, Krause J. 2000. Effects of parasites on fish behaviour: a review and evolutionary perspective. Rev Fish Biol Fish 10:131–165. https://doi.org/10.1023/A:1016658224470.
- Hughes DP, Araújo JPM, Loreto RG, Quevillon L, de Bekker C, Evans HC.
  2016. Chapter eleven from so simple a beginning: the evolution of behavioral manipulation by fungi, p 437–469. *In* Lovett B, St. Leger RJ (ed), Advances in genetics, vol 94. Academic Press, Waltham, MA.
- Piekarski G, Zippelius HM, Witting PA. 1978. Auswirkungen einer latenten*Toxoplasma*-Infektion auf das Lernvermögen von weißen Laboratoriumsratten und -mäusen. Z Parasitenkd 57:1–15. https://doi.org/10 .1007/BF00927625.
- 16. Witting PA. 1979. Learning capacity and memory of normal and *Toxoplasma*-infected laboratory rats and mice. Z Parasitenkd 61:29–51. https://doi.org/10.1007/bf00927085.
- Worth AR, Andrew Thompson RC, Lymbery AJ. 2014. Chapter three reevaluating the evidence for *toxoplasma gondii*-induced behavioural changes in rodents, p 109–142. *In* Rollinson D, Stothard JR (ed), Advances in parasitology. Academic Press, Waltham, MA.
- Hay J, Hutchison WM, Aitken PP, Graham DI. 1983. The effect of congenital and adult-acquired *Toxoplasma* infections on activity and responsiveness to novel stimulation in mice. Ann Trop Med Parasitol 77:483–495. https://doi.org/10.1080/00034983.1983.11811741.
- Hay J, Aitken PP, Hair DM, Hutchison WM, Graham DI. 1984. The effect of congenital *Toxoplasma* infection on mouse activity and relative preference for exposed areas over a series of trials. Ann Trop Med Parasitol 78:611–618. https://doi.org/10.1080/00034983.1984.11811872.
- Webster JP. 1994. The effect of *Toxoplasma gondii* and other parasites on activity levels in wild and hybrid *Rattus norvegicus*. Parasitology 109: 583–589. https://doi.org/10.1017/S0031182000076460.
- Hodkova H, Kodym P, Flegr J. 2007. Poorer results of mice with latent toxoplasmosis in learning tests: impaired learning processes or the novelty discrimination mechanism? Parasitology 134:1329–1337. https:// doi.org/10.1017/S0031182007002673.
- Kannan G, Moldovan K, Xiao J-C, Yolken RH, Jones-Brando L, Pletnikov MV. 2010. *Toxoplasma gondii* strain-dependent effects on mouse behaviour. Folia Parasitol (Praha) 57:151–155. https://doi.org/10.14411/fp.2010 .019.
- Xiao J, Kannan G, Jones-Brando L, Brannock C, Krasnova IN, Cadet JL, Pletnikov M, Yolken RH. 2012. Sex-specific changes in gene expression and behavior induced by chronic *Toxoplasma* infection in mice. Neuroscience 206:39–48. https://doi.org/10.1016/j.neuroscience.2011.12.051.
- Afonso C, Paixão VB, Klaus A, Lunghi M, Piro F, Emiliani C, Di Cristina M, Costa RM. 2017. *Toxoplasma*-induced changes in host risk behaviour are independent of parasite-derived AaaH2 tyrosine hydroxylase. Sci Rep 7:1–14. https://doi.org/10.1038/s41598-017-13229-y.
- Martynowicz J, Augusto L, Wek RC, Boehm SL, Sullivan WJ. 2019. Guanabenz reverses a key behavioral change caused by latent toxoplasmosis in mice by reducing neuroinflammation. mBio 10:e00381-19. https:// doi.org/10.1128/mBio.00381-19.
- Hrdá Š, Votýpka J, Kodym P, Flegr J. 2000. Transient nature of *Toxoplasma gondii*-induced behavioral changes in mice. J Parasitol 86: 657–663. https://doi.org/10.2307/3284944.
- Skallová A, Kodym P, Frynta D, Flegr J. 2006. The role of dopamine in *Toxoplasma*-induced behavioural alterations in mice: an ethological and ethopharmacological study. Parasitology 133:525–535. https://doi.org/ 10.1017/S0031182006000886.
- Gonzalez LE, Rojnik B, Urrea F, Urdaneta H, Petrosino P, Colasante C, Pino S, Hernandez L. 2007. *Toxoplasma gondii* infection lower anxiety as measured in the plus-maze and social interaction tests in rats: a behavioral analysis. Behav Brain Res 177:70–79. https://doi.org/10.1016/j.bbr .2006.11.012.
- Gulinello M, Acquarone M, Kim JH, Spray DC, Barbosa HS, Sellers R, Tanowitz HB, Weiss LM. 2010. Acquired infection with *Toxoplasma gondii* in adult mice results in sensorimotor deficits but normal cognitive behavior despite widespread brain pathology. Microbes Infect 12:528–537. https://doi .org/10.1016/j.micinf.2010.03.009.
- Berdoy M, Webster JP, Macdonald DW. 2000. Fatal attraction in rats infected with *Toxoplasma gondii*. Proc Biol Sci 267:1591–1594. https:// doi.org/10.1098/rspb.2000.1182.
- Vyas A, Kim S-K, Giacomini N, Boothroyd JC, Sapolsky RM. 2007. Behavioral changes induced by *Toxoplasma* infection of rodents are highly

specific to aversion of cat odors. Proc Natl Acad Sci U S A 104: 6442–6447. https://doi.org/10.1073/pnas.0608310104.

- Vyas A, Kim S-K, Sapolsky RM. 2007. The effects of toxoplasma infection on rodent behavior are dependent on dose of the stimulus. Neuroscience 148:342–348. https://doi.org/10.1016/j.neuroscience.2007.06.021.
- Lamberton PHL, Donnelly CA, Webster JP. 2008. Specificity of the *Tox-oplasma gondii*-altered behaviour to definitive versus non-definitive host predation risk. Parasitology 135:1143–1150. https://doi.org/10.1017/S0031182008004666.
- 34. Kaushik M, Lamberton PHL, Webster JP. 2012. The role of parasites and pathogens in influencing generalised anxiety and predation-related fear in the mammalian central nervous system. Horm Behav 62:191–201. https://doi.org/10.1016/j.yhbeh.2012.04.002.
- David CN, Frias ES, Szu JI, Vieira PA, Hubbard JA, Lovelace J, Michael M, Worth D, McGovern KE, Ethell IM, Stanley BG, Korzus E, Fiacco TA, Binder DK, Wilson EH. 2016. GLT-1-dependent disruption of CNS glutamate homeostasis and neuronal function by the protozoan parasite *Toxoplasma gondii*. PLoS Pathog 12:e1005643. https://doi.org/ 10.1371/journal.ppat.1005643.
- Goodwin D, Hrubec TC, Klein BG, Strobl JS, Werre SR, Han Q, Zajac AM, Lindsay DS. 2012. Congenital infection of Mice with *Toxoplasma gondii* induces minimal change in behavior and no change in neurotransmitter concentrations. J Parasitol 98:706–712. https://doi.org/10.1645/ GE-3068.1.
- Boillat M, Hammoudi P-M, Dogga SK, Pagès S, Goubran M, Rodriguez I, Soldati-Favre D. 2020. Neuroinflammation-associated aspecific manipulation of mouse predator fear by *Toxoplasma gondii*. Cell Rep 30: 320–334. https://doi.org/10.1016/j.celrep.2019.12.019.
- Hassan MA, Melo MB, Haas B, Jensen KDC, Saeij J. 2012. *De novo* reconstruction of the *Toxoplasma gondii* transcriptome improves on the current genome annotation and reveals alternatively spliced transcripts and putative long non-coding RNAs. BMC Genomics 13:696. https://doi .org/10.1186/1471-2164-13-696.
- Ingram WM, Goodrich LM, Robey EA, Eisen MB. 2013. Mice infected with low-virulence strains of *Toxoplasma gondii* lose their innate aversion to cat urine, even after extensive parasite clearance. PLoS One 8:e75246. https://doi.org/10.1371/journal.pone.0075246.
- Szabo EK, Finney CAM. 2017. *Toxoplasma gondii*: one organism, multiple models. Trends Parasitol 33:113–127. https://doi.org/10.1016/j.pt.2016 .11.007.
- Arsenijevic D, de Bilbao F, Vallet P, Hemphill A, Gottstein B, Richard D, Giannakopoulos P, Langhans W. 2007. Decreased infarct size after focal cerebral ischemia in mice chronically infected with *Toxoplasma gondii*. Neuroscience 150:537–546. https://doi.org/10.1016/j.neuroscience.2007 .09.080.
- 42. Jung B-K, Pyo K-H, Shin KY, Hwang YS, Lim H, Lee SJ, Moon J-H, Lee SH, Suh Y-H, Chai J-Y, Shin E-H. 2012. *Toxoplasma gondii* infection in the brain inhibits neuronal degeneration and learning and memory impairments in a murine model of Alzheimer's disease. PLoS One 7:e33312. https://doi.org/10.1371/journal.pone.0033312.
- Möhle L, Israel N, Paarmann K, Krohn M, Pietkiewicz S, Müller A, Lavrik IN, Buguliskis JS, Schott BH, Schlüter D, Gundelfinger ED, Montag D, Seifert U, Pahnke J, Dunay IR. 2016. Chronic *Toxoplasma gondii* infection enhances β-amyloid phagocytosis and clearance by recruited monocytes. Acta Neuropathol Commun 4:25. https://doi.org/10.1186/s40478-016 -0293-8.
- 44. Cabral CM, McGovern KE, MacDonald WR, Franco J, Koshy AA. 2017. Dissecting amyloid beta deposition using distinct strains of the neurotropic parasite *Toxoplasma gondii* as a novel tool. ASN Neuro 9:1759091417724915. https://doi.org/10.1177/1759091417724915.
- Tedford E, McConkey G. 2017. Neurophysiological changes induced by chronic *Toxoplasma gondii* infection. Pathogens 6:E19. https://doi.org/ 10.3390/pathogens6020019.
- Beninger RJ. 1983. The role of dopamine in locomotor activity and learning. Brain Res Rev 6:173–196. https://doi.org/10.1016/0165-0173(83) 90038-3.
- Strasser B, Gostner JM, Fuchs D. 2016. Mood, food, and cognition: role of tryptophan and serotonin. Curr Opin Clin Nutr Metab Care 19:55–61. https://doi.org/10.1097/MCO.0000000000237.
- Oueghlani Z, Juvin L, Lambert FM, Cardoit L, Courtand G, Masmejean F, Cazalets J-R, Barrière G. 18 October 2019. Serotonergic modulation of sacral dorsal root stimulation-induced locomotor output in newborn rat. Neuropharmacology https://doi.org/10.1016/j.neuropharm.2019 .107815.

- Stibbs HH. 1985. Changes in brain concentrations of catecholamines and indoleamines in *Toxoplasma gondii* infected mice. Ann Trop Med Parasitol 79:153–157. https://doi.org/10.1080/00034983.1985.11811902.
- Wang T, Sun X, Qin W, Zhang X, Wu L, Li Y, Zhou C, Zhou H, He S, Cong H. 2019. From inflammatory reactions to neurotransmitter changes: implications for understanding the neurobehavioral changes in mice chronically infected with *Toxoplasma gondii*. Behav Brain Res 359: 737–748. https://doi.org/10.1016/j.bbr.2018.09.011.
- Alsaady I, Tedford E, Alsaad M, Bristow G, Kohli S, Murray M, Reeves M, Vijayabaskar MS, Clapcote SJ, Wastling J, McConkey GA. 2018. Downregulation of the central noradrenergic system by *Toxoplasma gondii* infection. Infect Immun 87:e00789-18. https://doi.org/10.1128/IAI .00789-18.
- Ihara F, Nishimura M, Muroi Y, Mahmoud ME, Yokoyama N, Nagamune K, Nishikawa Y. 2016. *Toxoplasma gondii* infection in mice impairs long-term fear memory consolidation through dysfunction of the cortex and amygdala. Infect Immun 84:2861–2870. https://doi.org/10.1128/IAI .00217-16.
- Wang ZT, Harmon S, O'Malley KL, Sibley LD. 2015. Reassessment of the role of aromatic amino acid hydroxylases and the effect of infection by *Toxoplasma gondii* on host dopamine. Infect Immun 83:1039–1047. https://doi.org/10.1128/IAI.02465-14.
- 54. Levite M. 2016. Dopamine and T cells: dopamine receptors and potent effects on T cells, dopamine production in T cells, and abnormalities in the dopaminergic system in T cells in autoimmune, neurological and psychiatric diseases. Acta Physiol (Oxf) 216:42–89. https://doi.org/10 .1111/apha.12476.
- McConkey GA, Martin HL, Bristow GC, Webster JP. 2013. *Toxoplasma gondii* infection and behaviour location, location, location? J Exp Biol 216:113–119. https://doi.org/10.1242/jeb.074153.
- Gaskell EA, Smith JE, Pinney JW, Westhead DR, McConkey GA. 2009. A unique dual activity amino acid hydroxylase in *Toxoplasma gondii*. PLoS One 4:e4801. https://doi.org/10.1371/journal.pone.0004801.
- Webster JP, McConkey GA. 2010. *Toxoplasma gondii*-altered host behaviour: clues as to mechanism of action. Folia Parasitol (Praha) 57:95–104. https://doi.org/10.14411/fp.2010.012.
- McFarland R, Wang ZT, Jouroukhin Y, Li Y, Mychko O, Coppens I, Xiao J, Jones-Brando L, Yolken RH, Sibley LD, Pletnikov MV. 2018. AAH2 gene is not required for dopamine-dependent neurochemical and behavioral abnormalities produced by *Toxoplasma* infection in mouse. Behav Brain Res 347:193–200. https://doi.org/10.1016/j.bbr.2018.03.023.
- Flegr J, Hrdý I. 1994. Influence of chronic toxoplasmosis on some human personality factors. Folia Parasitol (Praha) 41:122–126.
- 60. Conejero-Goldberg C, Torrey EF, Yolken RH. 2003. Herpesviruses and *Toxoplasma gondii* in orbital frontal cortex of psychiatric patients. Schizophr Res 60:65–69. https://doi.org/10.1016/S0920-9964(02)00160-3.
- 61. Flegr J, Havlicek J. 1999. Changes in the personality profile of young women with latent toxoplasmosis. Folia Parasitol (Praha) 46:22–28.
- Flegr J, Preiss M, Klose J, Havlícek J, Vitáková M, Kodym P. 2003. Decreased level of psychobiological factor novelty seeking and lower intelligence in men latently infected with the protozoan parasite *Toxoplasma gondii* Dopamine, a missing link between schizophrenia and toxoplasmosis? Biol Psychol 63:253–268. https://doi.org/10.1016/s0301 -0511(03)00075-9.
- Novotná M, Hanusova J, Klose J, Preiss M, Havlicek J, Roubalová K, Flegr J. 2005. Probable neuroimmunological link between Toxoplasma and cytomegalovirus infections and personality changes in the human host. BMC Infect Dis 5:54. https://doi.org/10.1186/1471-2334-5-54.
- Skallová A, Novotná M, Kolbeková P, Gasová Z, Veselý V, Sechovská M, Flegr J. 2005. Decreased level of novelty seeking in blood donors infected with *Toxoplasma*. Neuro Endocrinol Lett 26:480–486.
- Flegr J. 2007. Effects of *Toxoplasma* on human behavior. Schizophr Bull 33:757–760. https://doi.org/10.1093/schbul/sbl074.
- Flegr J, Novotná M, Fialová A, Kolbeková P, Gasová Z. 2010. The influence of RhD phenotype on toxoplasmosis- and age-associated changes in personality profile of blood donors. Folia Parasitol (Praha) 57:143–150. https://doi.org/10.14411/fp.2010.018.
- Lindová J, Příplatová L, Flegr J. 2012. Higher extraversion and lower conscientiousness in humans infected with *Toxoplasma*. Eur J Pers 26: 285–291. https://doi.org/10.1002/per.838.
- Cook TB, Brenner LA, Cloninger CR, Langenberg P, Igbide A, Giegling I, Hartmann AM, Konte B, Friedl M, Brundin L, Groer MW, Can A, Rujescu D, Postolache TT. 2015. "Latent" infection with *Toxoplasma gondii*: asso-

ciation with trait aggression and impulsivity in healthy adults. J Psychiatr Res 60:87–94. https://doi.org/10.1016/j.jpsychires.2014.09.019.

- Flegr J, Havlícek J, Kodym P, Malý M, Smahel Z. 2002. Increased risk of traffic accidents in subjects with latent toxoplasmosis: a retrospective case-control study. BMC Infect Dis 2:11. https://doi.org/10.1186/1471 -2334-2-11.
- Yereli K, Balcioğlu IC, Ozbilgin A. 2006. Is *Toxoplasma gondii* a potential risk for traffic accidents in Turkey? Forensic Sci Int 163:34–37. https:// doi.org/10.1016/j.forsciint.2005.11.002.
- 71. Kocazeybek B, Oner YA, Turksoy R, Babur C, Cakan H, Sahip N, Unal A, Ozaslan A, Kilic S, Saribas S, Aslan M, Taylan A, Koc S, Dirican A, Uner HB, Oz V, Ertekin C, Kucukbasmaci O, Torun MM. 2009. Higher prevalence of toxoplasmosis in victims of traffic accidents suggest increased risk of traffic accident in *Toxoplasma*-infected inhabitants of Istanbul and its suburbs. Forensic Sci Int 187:103–108. https://doi.org/10.1016/j.forsciint .2009.03.007.
- Alvarado-Esquivel C, Torres-Castorena A, Liesenfeld O, Estrada-Martínez S, Urbina-Álvarez JD. 2012. High seroprevalence of *Toxoplasma gondii* infection in a subset of Mexican patients with work accidents and low socioeconomic status. Parasit Vectors 5:13. https://doi.org/10.1186/1756 -3305-5-13.
- Havlíček J, GAšOVÁ Z, Smith AP, Zvára K, Flegr J. 2001. Decrease of psychomotor performance in subjects with latent 'asymptomatic' toxoplasmosis. Parasitology 122:515–520. https://doi.org/10.1017/s0031182001007624.
- 74. Novotná M, Havlícek J, Smith AP, Kolbeková P, Skallová A, Klose J, Gasová Z, Písacka M, Sechovská M, Flegr J. 2008. *Toxoplasma* and reaction time: role of toxoplasmosis in the origin, preservation and geographical distribution of Rh blood group polymorphism. Parasitology 135:1253–1261. https://doi.org/10.1017/S003118200800485X.
- Samojłowicz D, Twarowska-Małczyńska J, Borowska-Solonynko A, Poniatowski ŁA, Sharma N, Olczak M. 2019. Presence of *Toxoplasma gondii* infection in brain as a potential cause of risky behavior: a report of 102 autopsy cases. Eur J Clin Microbiol Infect Dis 38:305–317. https://doi.org/ 10.1007/s10096-018-3427-z.
- Flegr J. 2013. Influence of latent *Toxoplasma* infection on human personality, physiology and morphology: pros and cons of the *Toxoplasma*-human model in studying the manipulation hypothesis. J Exp Biol 216:127–133. https://doi.org/10.1242/jeb.073635.
- 77. Sutterland AL, Fond G, Kuin A, Koeter MWJ, Lutter R, van Gool T, Yolken R, Szoke A, Leboyer M, de Haan L. 2015. Beyond the association. *Toxoplasma gondii* in schizophrenia, bipolar disorder, and addiction: systematic review and meta-analysis. Acta Psychiatr Scand 132:161–179. https://doi.org/10.1111/acps.12423.
- Sugden K, Moffitt TE, Pinto L, Poulton R, Williams BS, Caspi A. 2016. Is *Toxoplasma gondii* infection related to brain and behavior impairments in humans? Evidence from a population-representative birth cohort. PLoS One 11:e0148435. https://doi.org/10.1371/journal.pone.0148435.
- Okusaga O, Langenberg P, Sleemi A, Vaswani D, Giegling I, Hartmann AM, Konte B, Friedl M, Groer MW, Yolken RH, Rujescu D, Postolache TT. 2011. *Toxoplasma gondii* antibody titers and history of suicide attempts in patients with schizophrenia. Schizophr Res 133:150–155. https://doi .org/10.1016/j.schres.2011.08.006.
- Hinze-Selch D, Däubener W, Eggert L, Erdag S, Stoltenberg R, Wilms S. 2007. A controlled prospective study of *Toxoplasma gondii* infection in individuals with schizophrenia: beyond seroprevalence. Schizophr Bull 33:782–788. https://doi.org/10.1093/schbul/sbm010.
- Torrey EF, Bartko JJ, Yolken RH. 2012. *Toxoplasma gondii* and other risk factors for schizophrenia: an update. Schizophr Bull 38:642–647. https:// doi.org/10.1093/schbul/sbs043.
- de Barros JLVM, Barbosa IG, Salem H, Rocha NP, Kummer A, Okusaga OO, Soares JC, Teixeira AL. 2017. Is there any association between *Toxoplasma gondii* infection and bipolar disorder? A systematic review and meta-analysis. J Affect Disord 209:59–65. https://doi.org/10.1016/j.jad .2016.11.016.
- Miman O, Kusbeci OY, Aktepe OC, Cetinkaya Z. 2010. The probable relation between *Toxoplasma gondii* and Parkinson's disease. Neurosci Lett 475:129–131. https://doi.org/10.1016/j.neulet.2010.03.057.
- Ramezani M, Shojaii M, Asadollahi M, Karimialavijeh E, Gharagozli K. 2016. Seroprevalence of *Toxoplasma gondii* in Iranian patients with idiopathic Parkinson's disease. Clin Exp Neuroimmunol 7:361–365. https://doi.org/10.1111/cen3.12329.
- Mahami Oskouei M, Hamidi F, Talebi M, Farhoudi M, Taheraghdam AA, Kazemi T, Sadeghi-Bazargani H, Fallah E. 2016. The correlation between *Toxoplasma gondii* infection and Parkinson's disease: a case-control

study. J Parasit Dis 40:872–876. https://doi.org/10.1007/s12639-014 -0595-3.

- Alvarado-Esquivel C, Méndez-Hernández EM, Salas-Pacheco JM, Ruano-Calderón LÁ, Hernández-Tinoco J, Arias-Carrión O, Sánchez-Anguiano LF, Castellanos-Juárez FX, Sandoval-Carrillo AA, Liesenfeld O, Ramos-Nevárez A. 2017. *Toxoplasma gondii* exposure and Parkinson's disease: a case-control study. BMJ Open 7:e013019. https://doi.org/10.1136/ bmjopen-2016-013019.
- Zhou Z, Zhou R, Li K, Wei W, Zhang Z, Zhu Y. 2019. The association between *Toxoplasma gondii* infection and risk of Parkinson's disease: a systematic review and meta-analysis. Biomed Res Int 2019:8086017. https://doi.org/10.1155/2019/8186017.
- Miklossy J. 2011. Emerging roles of pathogens in Alzheimer disease. Expert Rev Mol Med 13:e30. https://doi.org/10.1017/S1462399411002006.
- Mawanda F, Wallace R. 2013. Can infections cause Alzheimer's disease? Epidemiol Rev 35:161–180. https://doi.org/10.1093/epirev/mxs007.
- Nayeri Chegeni T, Sarvi S, Moosazadeh M, Sharif M, Aghayan SA, Amouei A, Hosseininejad Z, Daryani A. 2019. Is *Toxoplasma gondii* a potential risk factor for Alzheimer's disease? A systematic review and meta-

analysis. Microb Pathog 137:103751. https://doi.org/10.1016/j.micpath .2019.103751.

- Pearce BD, Kruszon-Moran D, Jones JL. 2014. The association of *Toxoplasma gondii* infection with neurocognitive deficits in a population-based analysis. Soc Psychiatry Psychiatr Epidemiol 49:1001–1010. https://doi.org/10.1007/s00127-014-0820-5.
- Gale SD, Brown BL, Erickson LD, Berrett A, Hedges DW. 2015. Association between latent toxoplasmosis and cognition in adults: a cross-sectional study. Parasitology 142:557–565. https://doi.org/10.1017/S0031182014001577.
- 93. van Os J, Kapur S. 2009. Schizophrenia. Lancet 374:635–645. https://doi .org/10.1016/S0140-6736(09)60995-8.
- 94. Mayeux R, Stern Y. 2012. Epidemiology of Alzheimer disease. Cold Spring Harb Perspect Med 2:a006239. https://doi.org/10.1101/cshperspect .a006239.
- Pappas G, Roussos N, Falagas ME. 2009. Toxoplasmosis snapshots: global status of *Toxoplasma gondii* seroprevalence and implications for pregnancy and congenital toxoplasmosis. Int J Parasitol 39:1385–1394. https://doi.org/10.1016/j.ijpara.2009.04.003.