

## Supplemental Information

Identification of a direct interaction between the Fab domains of IgG antibodies and human FcRn upon IgG-FcRn complex formation

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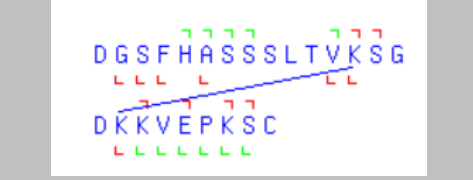
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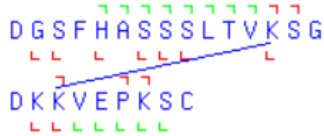


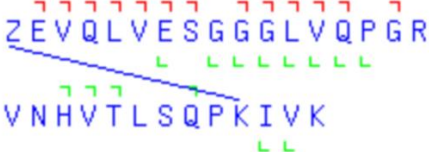
## Supplemental Methods

*Calculation and visualization of surface charge:* Surface charge analysis was done based on the crystal structures of the Fab domains of H-mAb and C-mAb (PDB ID: 4NYL and 1NGP). The surface charge maps were generated by the pdb2pqr (3.6.1) and APBS (3.4.1) servers (<https://server.poissonboltzmann.org/pdb2pqr>) with standard settings. The protonation states of side chains were calculated by the PROPKA algorithm and the PARSE forcefield was used. The resulting surface potential maps were visualised at  $\pm 3k_B T/e$  in PyMOL (2.5.2). The sequence based pH-dependent net charge of the Fab domains of H-mAb and C-mAb were calculated using the Emboss iep calculator ([http://www.bioinformatics.nl/cgi-bin/emboss/iep?\\_pref\\_hide\\_optional=0](http://www.bioinformatics.nl/cgi-bin/emboss/iep?_pref_hide_optional=0)).<sup>1</sup> All cysteines were assumed to form disulfide bridges and the Fab domains were set to contain 2 N-terminals and 1 C-termini.

*In-gel digestion of C-mAb:* The SDS-PAGE band corresponding to monomeric C-mAb was sliced of the gel, cut into smaller pieces, placed in a Eppendorf tube, solvated with 200  $\mu$ L 25 mM  $\text{NH}_4\text{HCO}_3$  in 50:50 MQ water:acetonitrile, and incubated at 37 °C under shaking for 30 min. The supernatant was discarded and the process was repeated twice to de-stain the gel pieces. The gel pieces were dehydrated with 100  $\mu$ L acetonitrile, washed with 100  $\mu$ L MQ water, dehydrated with 100  $\mu$ L acetonitrile before addition of 100  $\mu$ L 6.5 mM dithiothreitol in MQ water and incubated at 60 °C under shaking for 60 min to ensure efficient reduction of C-mAb. The solvent was discarded and the gel pieces were dehydrated in 100  $\mu$ L acetonitrile before addition of 100  $\mu$ L 54 mM iodoacetamide in MQ water and incubated at 25 °C in the dark under shaking for 30 min to alkylate C-mAb. The solvent was discarded and the gel pieces were dehydrated with 100  $\mu$ L acetonitrile and hydrated with 50 mM  $\text{NH}_4\text{HCO}_3$  in MQ water. This process was repeated 3 times, and the gel pieces were dehydrated a final time in 100  $\mu$ L acetonitrile. The supernatant was discarded and the dehydrated gel pieces were hydrated in 100  $\mu$ L of 10 ng/ $\mu$ L trypsin solution in 50 mM  $\text{NH}_4\text{HCO}_3$ . The gel pieces were incubated overnight at 37 °C under shaking. The next morning the supernatant was collected. The gel pieces were covered in 40  $\mu$ L 0.1 % formic acid in MQ water and incubated for 15 min at room temperature under shaking. The supernatant was collected. The gel pieces were covered in 40  $\mu$ L of a 50:50 mixture of 0.1 % formic acid in MQ water and acetonitrile and incubated for 15 min at room temperature under shaking. The supernatant was collected. The gel pieces were covered in 40  $\mu$ L acetonitrile and incubated for 15 min at room temperature under shaking. All collected supernatants were pooled and evaporated to dryness in a vacuum centrifuge. The samples were re-suspended in 20  $\mu$ L SEC running buffer, purified by SEC and analyzed by LC-MS as described for XL-MS samples in the methods section.

**Supporting Table S1. Table of identified cross-links including overview of identified fragment ions.** A “Z”-residue denotes the end amino group on the N-terminal of a protein chain (cross-link number 8).

	Residue 1	Residue 2	Peptide 1	Peptide 2	Id-Score	Euclidean C $\alpha$ -C $\alpha$ distance (Å)	Identified fragments ions
							Red: fragment ion containing cross-linker, Green: Fragment ions not containing cross-linker
1	$\beta$ 2m - K91	Fc HC - K292	VNHVTLSQPKIVK	FNWYVDGVEVHNAKTkPR	34.11	16.8	
2	$\beta$ 2m - K91	Fc HC - K294	VNHVTLSQPKIVK	FNWYVDGVEVHNAKTkPR	36.36	20.5	
3	$\beta$ 2m - K6	Fc HC - K292	TPKIQVYSR	FNWYVDGVEVHNAKTkPR	33.26	24.0	
4	$\alpha$ 3 - K243	Fab HC - K217	DGSFHASSSLTVKSG	DKKVEPKSC	36.16		

5	$\alpha$ 3 - K243	Fab HC - K218	DGSFHASSSLTVKSG	DKKVEPKSC	40.56	
6	$\beta$ 2m - K91	Fab HC - K59	VNHVTLSQPKIVK	IDPNSGGTKYNEK	33.41	
7	$\beta$ 2m - K94	Fab HC – K59	IVKWDRDM	IDPNSGGTKYNEK	37.86	
8	$\beta$ 2m - K91	Fab HC – E1 (N-term)	VNHVTLSQPKIVK	EVQLVESGGGLVQPGR	32.60	

**Supporting Table S2. Identification of overlenght cross-links between Fab domains and the Fc part of C-mAb.** C-mAb was cross-linked with DSS as described in the methods section. After cross-linking the cross-linking products where seperated on an SDS-PAGE as described in the methods section. The band corresponding to monomeric C-mAb was sliced out of the gel and in-gel digestion with LysC and trypsin was performed, see supporting methods. The released peptides were analysed by LC-MS as described in the methods section. 5 overlenght cross-links where identified highlighting the flexibility of the Fab domains in C-mAb. The Euclidan distances where determined in Pymol on the homology model of C-mAb. The highest Id-score observed across all data sets is shown.

Residue 1	Residue 2	Id-Score	Replicates	Euclidean C $\alpha$ -C $\alpha$ distance (Å)
<i>non overlenght XL</i>				
HC - 137	LC - 207	25.19	1/2	11.1
HC - 137	HC - 218	36.4	2/2	19.9
HC - 137	HC - 222	28.98	2/2	10.6
HC - 218	LC - 207	32.8	1/2	24.5
HC - 364	HC - 418	39.9	2/2	7.8
<i>Overlenght XL</i>				
HC - 59	HC - 344	31.75	1/2	88.3
HC - 59	HC - 418	37.59	2/2	79.5
HC - 137	HC - 330	32.54	2/2	39.8
LC - 207	HC - 344	37.94	1/2	60.0
HC - 63	HC - 418	32.22	1/2	76.0

**Supporting Table S3. Cross-links only observed in a single experimental condition or only observed in the FcRn:mutC-mAb complex.** The abbreviation in the “Residue 1 and 2” columns refers to the domains of FcRn ( $\alpha 1$ ,  $\alpha 2$ ,  $\alpha 3$  or  $\beta 2m$ ) and IgGs (LC or HC). The “Complex” column refers to in which complex the cross-link have been identified. The “Chemistry” column refers to the cross-linking chemistry applied. The “Enzymes” column refers to the used proteases. The “Id-Score / FDR” refers to the Id-score in xQuest, while the FDR was calculated in xProphet. The “n-seen” column refers to number of spectrums, where the cross-link was identified.

	Residue 1	Residue 2	Complex	Chemistry	Enzymes	Id-Score / FDR	n-seen
1	$\beta 2m$ - 6	HC - 292/294	FcRn:H-mAb	DSS	LysC + Tryp	33.26	1
2	$\alpha 2$ - 146	HC - 76	FcRn:H-mAb	DSS	LysC + Tryp	40.74	3
3	$\beta 2m$ - 91	HC - 418	FcRn:H-mAb, FcRn:C-mAb, FcRn:mutC-mAb	DSS	LysC + Tryp	36.08	1
4	$\beta 2m$ - 91	LC - 145	FcRn:H-mAb	DSS	LysC + Tryp	26.16	1
5	$\alpha 2$ - 146	HC - 418	FcRn:H-mAb	DSS	LysC + Tryp	26.83	1
6	$\alpha 2$ - 146	LC - 1	FcRn:H-mAb	DSS	LysC + Tryp	37.38	1
7	$\beta 2m$ - 91	LC - 107	FcRn:H-mAb	DSS	LysC + Tryp	37.4	1
8	$\beta 2m$ - 91	LC - 188	FcRn:H-mAb	DSS	LysC + Tryp	39.09	1
9	$\alpha 2$ - 177	LC - 1	FcRn:H-mAb	DSS	LysC + Tryp	30.76	1
10	$\alpha 1$ - 59	HC - 74	FcRn:C-mAb	DSS	LysC + Tryp	25.11	2
11	$\beta 2m$ - 91	HC - 278	FcRn:H-mAb	DSS	AspN	36.12	3
12	$\beta 2m$ - 41	HC - 343	FcRn:H-mAb	DSS	AspN	25.63	1
13	$\beta 2m$ - 94	HC - 278	FcRn:H-mAb	DSS	AspN	27.71	1
14	$\beta 2m$ - 94	HC - 46	FcRn:H-mAb	PDH-ZL	LysC + Tryp	23.24 / 0	2
15	$\alpha 2$ - 146	LC - 207	FcRn:mutC-mAb	DSS	LysC + Tryp	29.28	1
16	$\beta 2m$ - 48	LC - 72	FcRn:mutC-mAb	DSS	AspN	28.16	1
17	$\beta 2m$ - 91	HC - 217/218	FcRn:mutC-mAb	DSS	AspN	27.01	1
18	$\alpha 1$ - 67	LC - 206	FcRn:mutC-mAb	XP	LysC + Tryp	37.77	1
19	$\alpha 2$ - 168	LC - 163	FcRn:mutC-mAb	XP	LysC + Tryp	36.52	1
20	$\alpha 2$ - 178	LC71	FcRn:mutC-mAb	XP	LysC + Tryp	27.43	1
21	$\beta 2m$ - 77	HC - 46	FcRn:mutC-mAb	XP	LysC + Tryp	37.93	1
22	$\alpha 1$ - 54/62	HC - 237	FcRn:mutC-mAb	XP	AspN	32.05	2
23	$\beta 2m$ - 38	HC - 262/269	FcRn:mutC-mAb	PDH	LysC + Tryp	29.43	2
24	$\alpha 1$ - 71	LC - 71	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	40.88 / 0	2
25	$\beta 2m$ - 50/53	HC - 217/218	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	37.36 / 0	1
26	$\beta 2m$ - 50	HC - 330	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	37.29 / 0	1
27	$\beta 2m$ - 53	HC - 330	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	33.76 / 0	1
28	$\beta 2m$ - 48	LC - 206	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	33.56 / 0	1
29	$\beta 2m$ - 94	LC - 201/206	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	31.95 / 0	2
30	$\alpha 1$ - 71	LC - 80	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	31.57 / 0	1
31	$\alpha 2$ - 178	HC - 418	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	30.09 / 0.05	2
32	$\alpha 2$ - 145	HC - 67	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	26.89 / 0	1
33	$\alpha 2$ - 165	HC - 330	FcRn:mutC-mAb	XP-ZL	LysC + Tryp	31.24 / 0	3
34	$\beta 2m$ - 50	HC 324/326	FcRn:mutC-mAb	PDH-ZL	LysC + Tryp	24.37 / 0	4

**Supporting Table S4. Identified inter protein cross-links between IdeZ-treated mAbs and FcRn.** The identified cross-links below all belong to the IdeZ treated mutC-mAb in complex with FcRn, confirming the experimental conditions. As no interprotein cross-links have been identified between the Fc-part of an mAb and FcRn in a AspN digest in the current study this is expected. The Euclidean distances were determined in Pymol on the homology model of C-mAb. The highest Id-score observed across all data sets is shown

	Residue 1	Residue 2	Id-Score	Euclidean C $\alpha$ -C $\alpha$ distance (Å)
1	$\beta$ 2m - K91	Fc HC - K292	34.37	16.8
2	$\beta$ 2m - K91	Fc HC - K294	36.59	20.5
3	$\beta$ 2m - K6	Fc HC - K294	35.79	24.0



**Supporting Table S5. Overview of recorded HDX data.** The numbers in the table refers to the number of replicate(s) of recorded HDX data for every single time point at every condition. N/A: Not applicable.

Experiment	Protein	Ligand	Molar Excess of Ligand	13 s	25 s	38 s	2.09 min	6min 17s	25min 7s	3t 25min 45s	4t 11min 11s	1d 1t 7min 8s	Maximally labelled samples
I	FcRn	Not present	N/A				3	3	3		3	3	3
	FcRn	H-mAb	7 times				3	1	1		1	1	
	FcRn	C-mAb	7 times				3	3	3		3	3	
II	FcRn	Not present	N/A	3	3	3	3						3
	FcRn	H-mAb	7 times	3	3	3	3						
	FcRn	C-mAb	7 times	3	3	3	3						
III	H-mAb	Not present	N/A				3	1	1	1			1
	H-mAb	FcRn	7 times				3	1	1	1			
IV	C-mAb	Not present	N/A				3	3	3		3	3	
	C-mAb	FcRn	7 times				3	3	3		3	3	3

**A)**

Identities:391/451(87%), Positives:415/451(92%), Gaps:1/451(0%)

H-mAb	1	EVQLVESGGGLVQPGRSRLRLSCAASGFTFDDYAMHWVRQAPGKGLEWVSAITWNSGHIDY	60
C-mAb	1	+VQL + G LV+PG S++LSC ASG+TF Y MHWV+Q PG+GLEW+ I NSG Y	60
H-mAb	61	QVQLQQPGAELVKPGASVKLSCKASGYTFTSYWMHWVKQRPGRGLEWIGRIDPNSGGTKY	60
C-mAb	61	ADSV EGRFTISRDN AKNSLYLQMN SLRAEDTAVYYCAKVS YLSTASSLDYWGQGT LTVTS	120
H-mAb	61	+ + + T++ D ++ Y+Q++SL +ED+AVYYCA+ Y +S DYWGQGT +TVS	120
C-mAb	61	NEKFKSKATLTVDKPSSTAYMQLSSLTSEDSAVYYCARYDYYG-SSYFDYWGQGTTLTVS	120
H-mAb	121	SASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQS	180
C-mAb	121	SASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQS	180
H-mAb	181	SGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLG	240
C-mAb	181	SGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLG	240
H-mAb	241	GPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQY	300
C-mAb	241	GPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQY	300
H-mAb	301	GSTYRVSVLTIVLHQLDNLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRD	360
C-mAb	301	NSTYRVSVLTIVLHQLDNLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRD	360
H-mAb	361	ELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPPVLDSDGSFFLYSKLTVDKSR	420
C-mAb	361	ELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPPVLDSDGSFFLYSKLTVDKSR	420
H-mAb	421	WQQGNVFSCSVMHEALHNHYTQKSLSLSPG	450
C-mAb	421	WQQGNVFSCSVMHEALHNHYTQKSLSLSPG	450

**B)**

Identities:206/209(99%), Positives:207/209(99%), Gaps:0/209(0%)

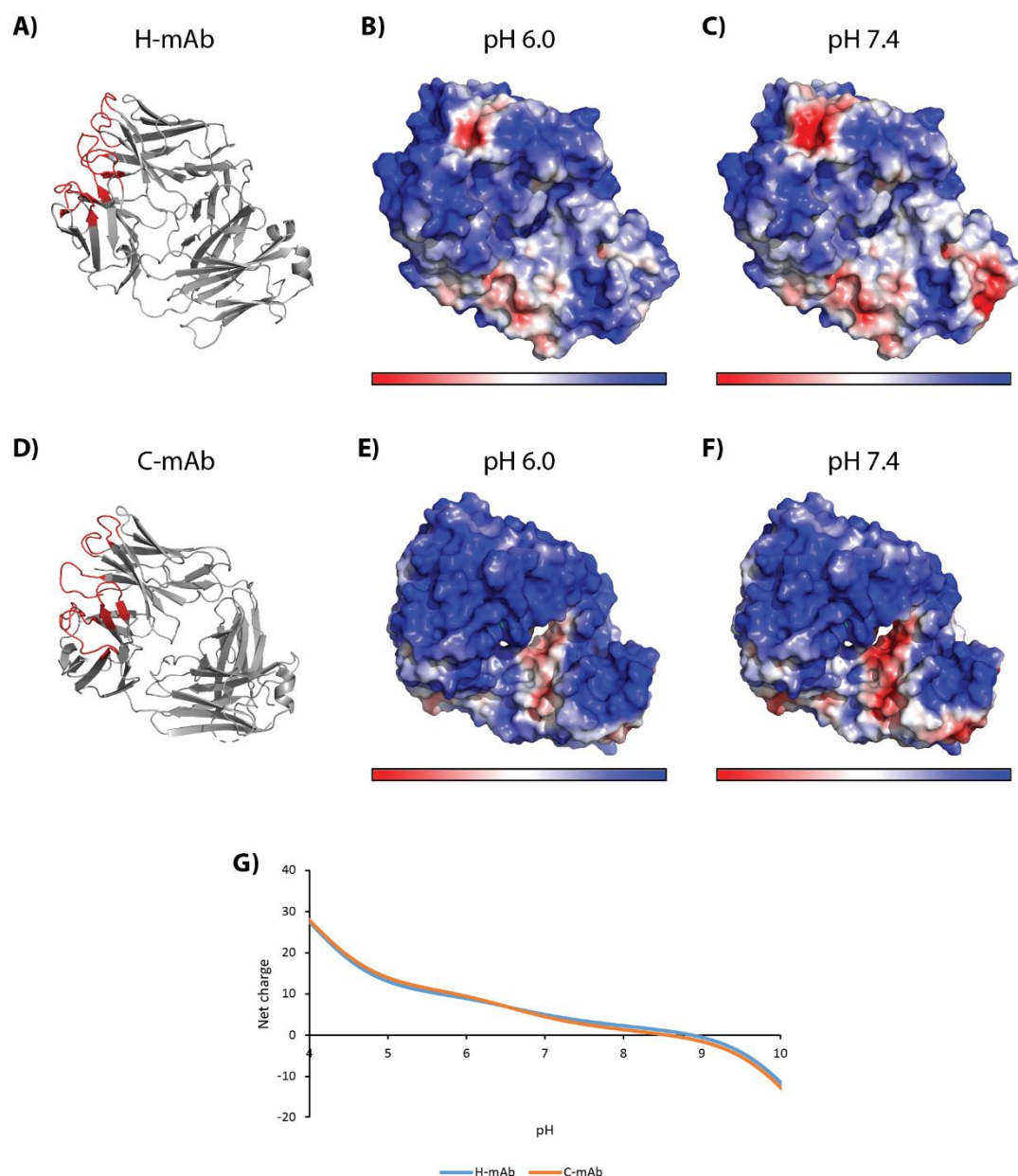
H-mAb	240	GGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQ	299
Fc-YTE	236	GGPSVFLFPPKPKDTL <b>LYIT</b> REPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQ	295
H-mAb	300	YNSTYRVSVLTIVLHQLDNLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSR	359
Fc-YTE	296	YNSTYRVSVLTIVLHQLDNLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSR	355
H-mAb	360	DELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPPVLDSDGSFFLYSKLTVDKS	419
Fc-YTE	356	DELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPPVLDSDGSFFLYSKLTVDKS	415
H-mAb	420	RWQQGNVFSCSVMHEALHNHYTQKSLSLS	447
Fc-YTE	416	RWQQGNVFSCSVMHEALHNHYTQKSLSLS	444

**Supporting Figure S1. Alignment of HCs of H-mAb with C-mAb (A) and HC of H-mAb with Fc-YTE (B).** Residues marked in bold are directly implicated in the binding of Fc-YTE to FcRn according to the crystal structure solved by Oganessian et al.<sup>2</sup> The alignment was performed by the BLAST algorithm.<sup>3,4</sup>

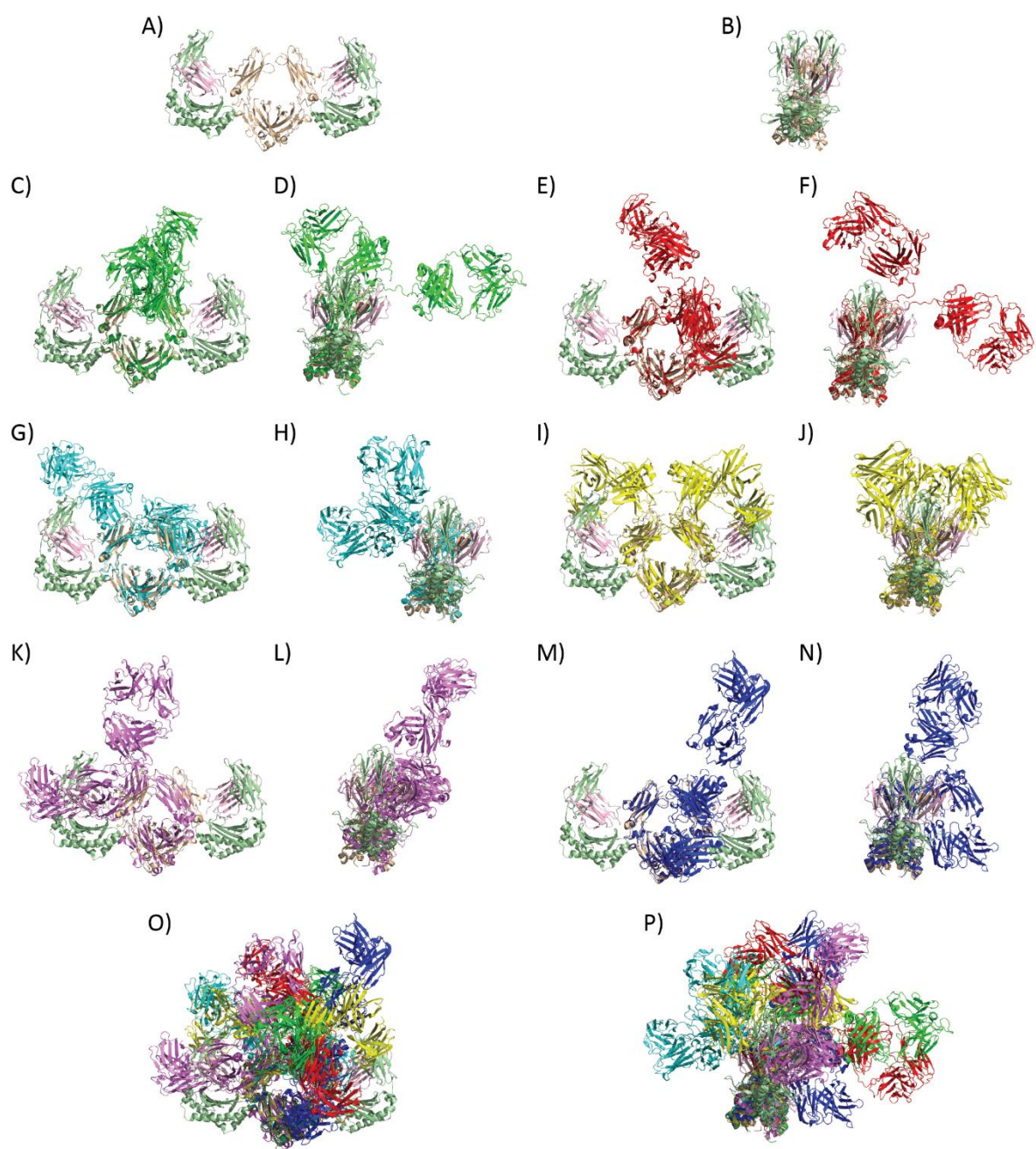
Identities:87/215(40%), Positives:123/215(57%), Gaps:11/215(5%)

H-mAb	1	DIQMTQSPSSLSASVGDRVTITCRASQG---IRNYLAWYQQKPGKAPKLLIYAAS	57
		S+L+ S G+ VT+TCR+S G       NY W Q+KP       LI + G	
C-mAb	1	-QAVVTQESALTTSPGETVTLTCRSSTGAVTTSNYANWVQEKPDLFTGLIGGTNNRAPG	59
H-mAb	58	VPSRFGSGSGTDFTLTISLQPEDVATYYCQRYNRAPYTFGQGTKVEI-KRTVAAPSVF	116
		VP+RFSGS G    LTI+ Q ED A Y+C +    + FG GTK+ + + ++PSV	
C-mAb	60	VPARFSGSLIGDKAALTITGTQTEDEAMYFCALWYSNHWVFGGGTKLTVLGQPKSSPSVT	119
H-mAb	117	IFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDST--YS	174
		+FPPS E+L++ A++VC + +FYP    V WKVD    + +Q T Q SK S Y	
C-mAb	120	LFPPSSEELETNKATLVCTITDFYPGVVTVDWKVDG---TPVTQGMETTQPSKQSNNKYM	176
H-mAb	175	LSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC	214
		SS LTL+    +E+H Y+C+VTH+G + V KS +R +C	
C-mAb	177	ASSYLTLTARAWERHSSYSQVTHEGHT--VEKSLSRADC	215

**Supporting Figure S2. Alignment of LCs of H-mAb and C-mAb.** The alignment was performed by the BLAST algorithm.<sup>3,4</sup>

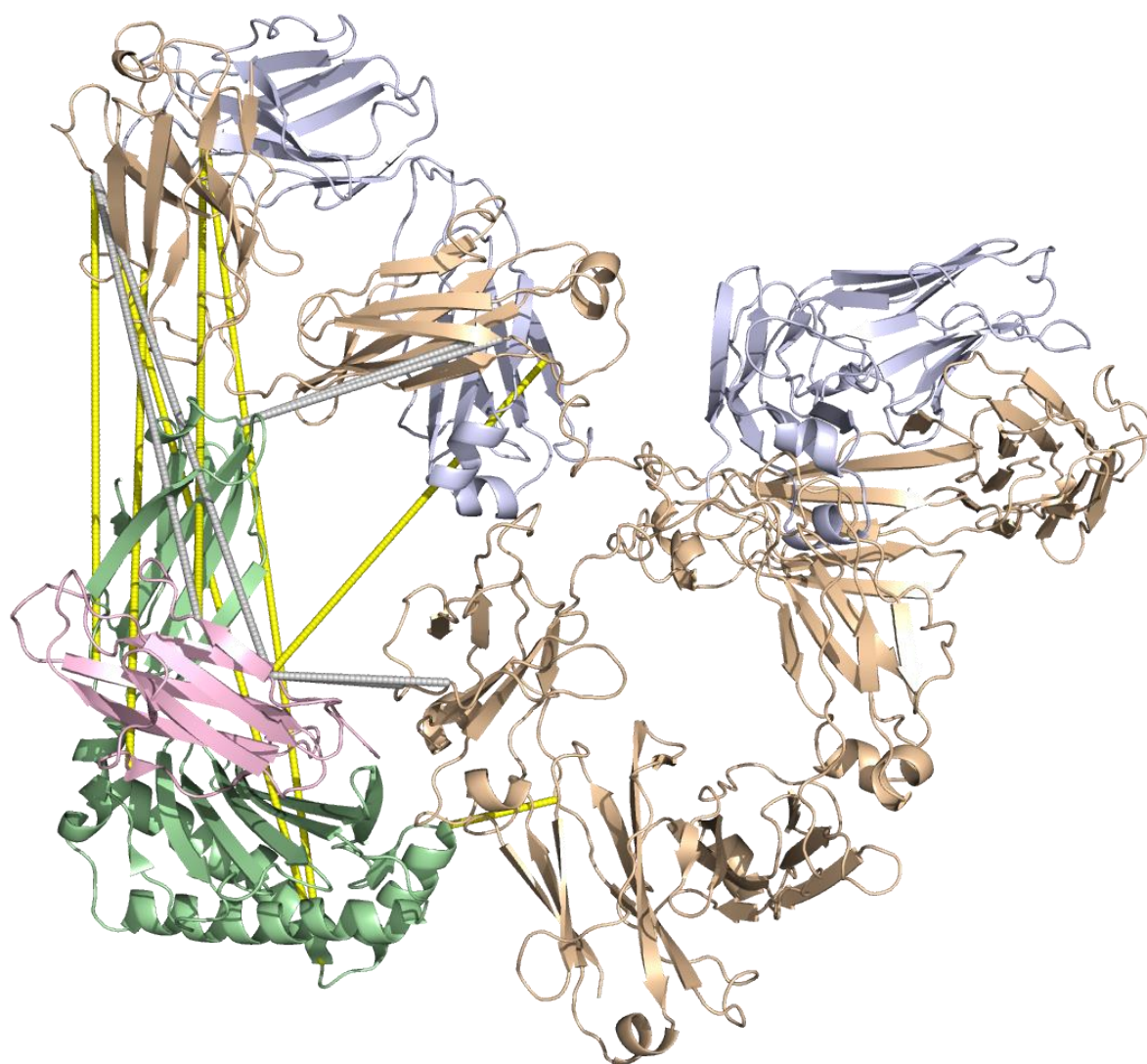


**Supporting Figure S3. Charge distribution and pH-dependent net charge of the Fab domains of H-mAb and C-mAb.** Crystal structure of the Fab domains of H-mAb (A) and C-mAb (D) (PDB ID:4NYL and 1NGP). CDRs are highlighted in red. B-C) Isoelectrical surface maps of H-mAb at pH 6.0 and pH 7.4 contoured at  $\pm 3k_B T/e$ , red; negative and blue: positive. E-F) Isoelectrical surface maps of C-mAb at pH 6.0 and pH 7.4 contoured at  $\pm 3k_B T/e$ , red; negative and blue: positive. G) pH-dependent sequence based net charge of the Fab domains of H-mAb (*blue* line) and C-mAb (*orange* line). See supplemental methods for further details.

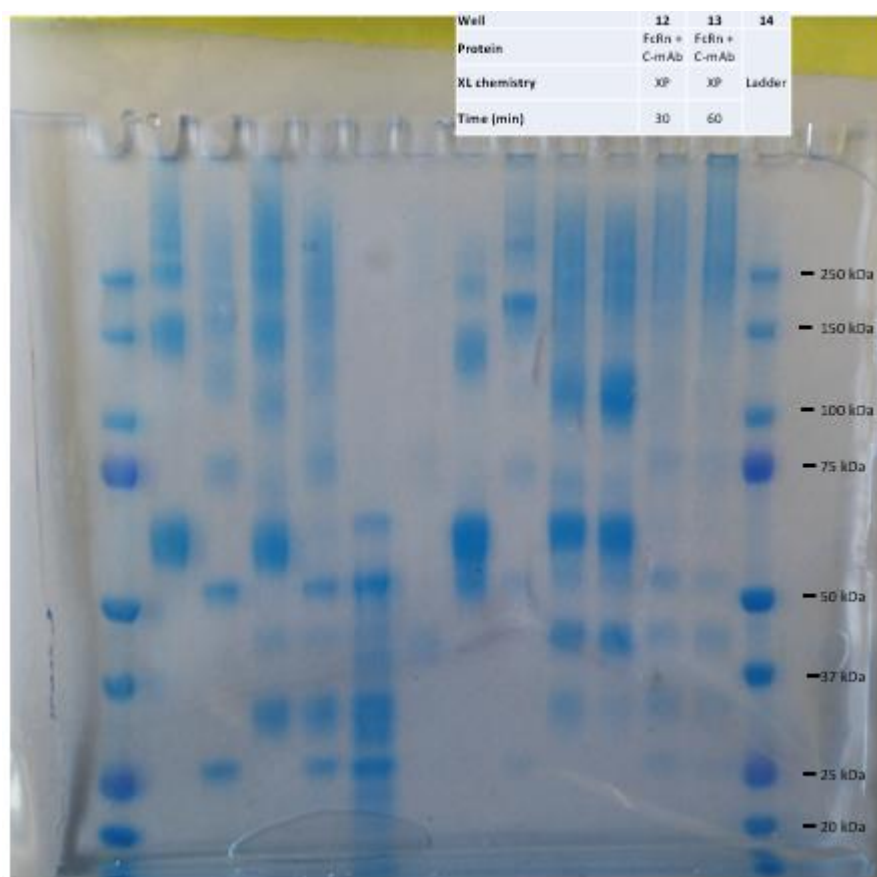


**Supporting Figure S4: Conformational flexibility of full-length IgGs.** Six crystal structures of full-length IgGs (PDB ID: 1IGT, 1HZH, 6GFE, 1MCO, 5FK3, and 1IGY) were aligned with the Fc part of the crystal structure of Fc-YTE in complex with FcRn (PDB ID: 4N0U, human serum albumin was omitted to simplify the figure). A-B) FcRn in complex with Fc-YTE (PDB ID: 4N0U) C-N) Individual IgGs shown with FcRn in complex with Fc-YTE. O-P) All IgG structures are shown at the same time. *Red*: 1IGT, *green*: 1HZH, *blue*: 6GFE, *yellow*: 1MCO, *violet*: 5DK3, *cyan*: 1IGY, *pale green*: FcRn, *light pink*:  $\beta$ -2-microglobulin, *wheat*: Fc-YTE. All figures are presented in two different views: a front view (A, C, E, G, I, K, M and O) and a view where the front view is rotated 90° counter clockwise along the horizontal axis (B, D, F, H, J, L, N and P).

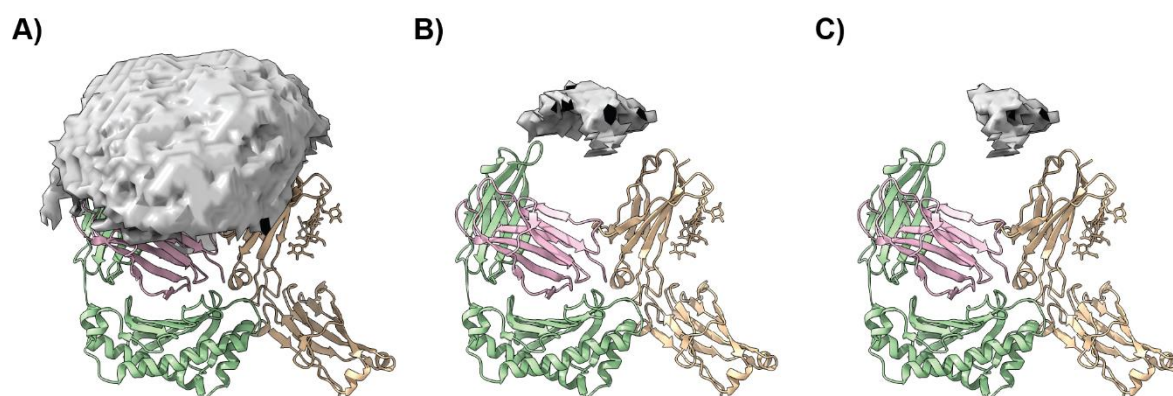




**Supporting Figure S5: Visualization of cross-links identified in the complex between mutC-mAb and FcRn.** Cross-links also found in the complex of C-mAb and FcRn are colored gray, while the cross-links only identified in the mutC-mAb:FcRn complex are colored yellow. The cross-links between mutC-mAb and FcRn illustrates several different binding modes not in line with the published crystal structure of FcRn:Fc-YTE complex (PDB ID: 4N0U). *Pale green*:  $\alpha$  domains of FcRn, *light pink*:  $\beta$ 2m, *wheat*: HC of mutC-mAb, *light blue*: LC of mutC-mAb.



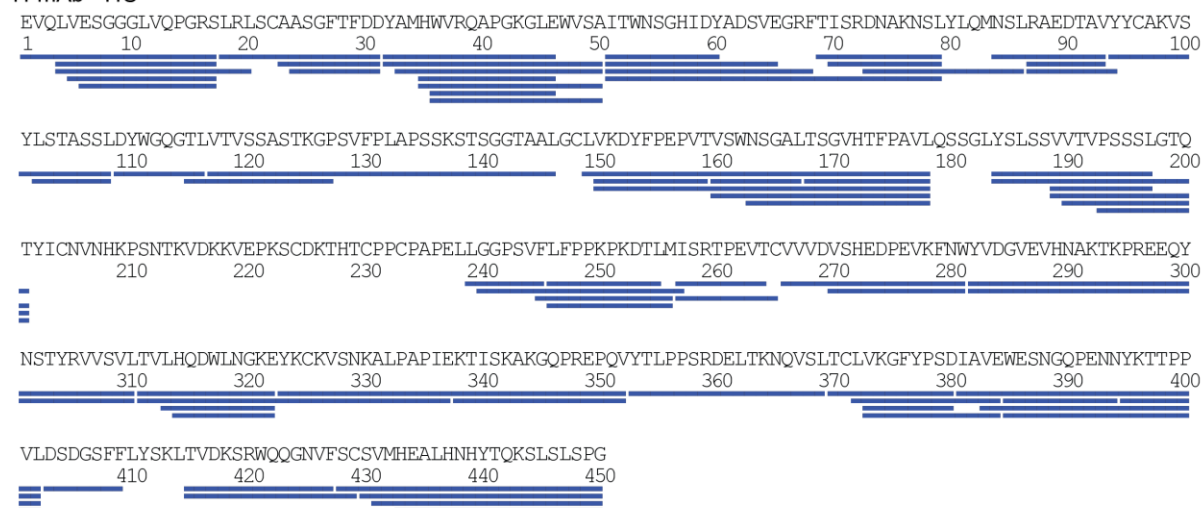
**Supporting Figure S6: Observation of higher order oligomers after cross-linking a solution of FcRn and C-mAb with XPlex cross-linking (XP).**



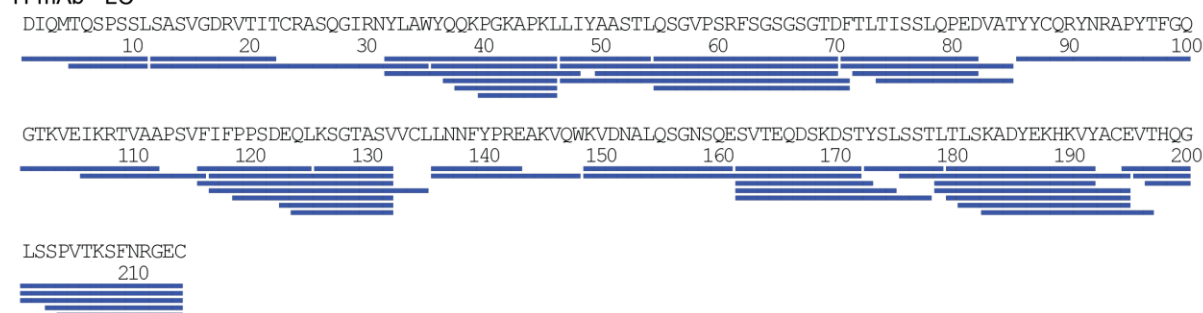
**Supporting Figure S7: DisVis analysis of mAbs Fab domain interaction with FcRn.** A) Interaction space of the Fab domain of H-mAb defined by the identified cross-links between H-mAb Fab domains and FcRn. B) Interaction space of the C-mAb Fab domain defined by the identified cross-links between C-mAb Fab domains and FcRn. C) Interaction space of a generic Fab domain defined by combining all cross-links between the Fab domains of both C-mAb and H-mAb and FcRn. *Pale green*:  $\alpha$  domains of FcRn, *light pink*:  $\beta$ 2m, *wheat*: Fc-YTE.



### H-mAb - HC

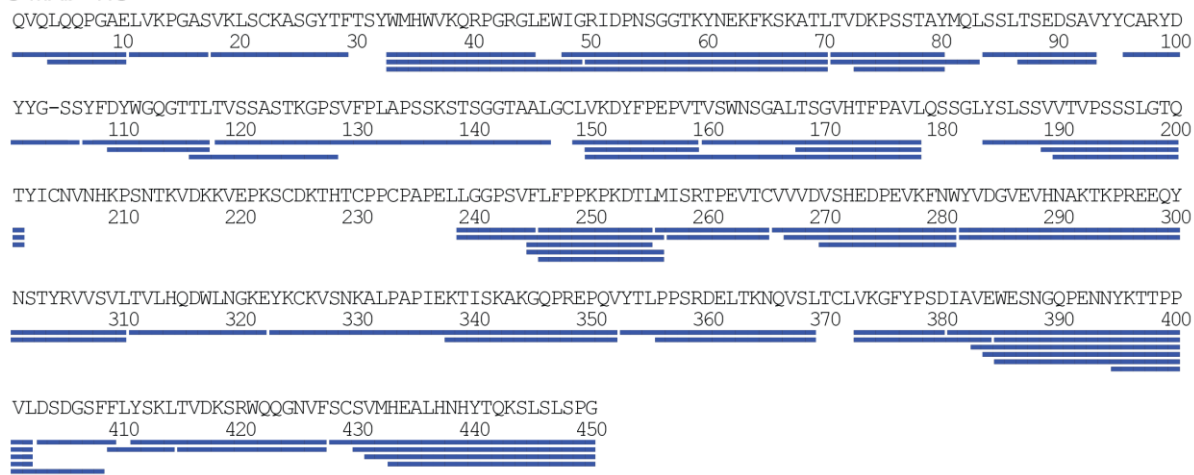


### H-mAb - LC

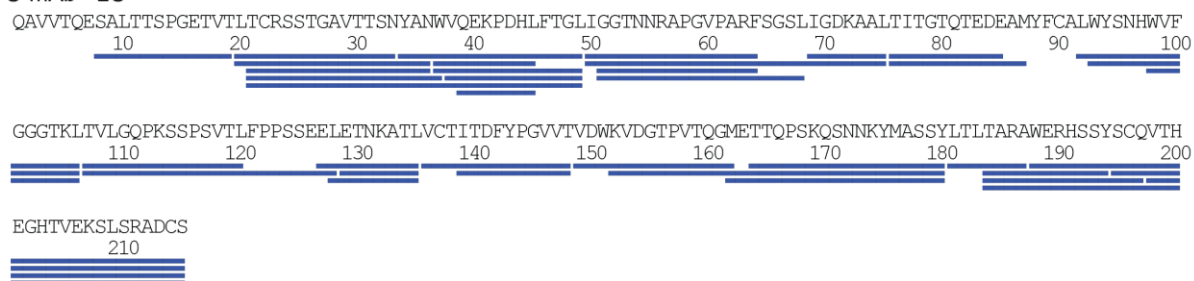


**Supporting Figure S8: Effective HDX-MS coverage map of H-mAb.** HDX data were obtained for 77 peptides covering 89.1% of the HC and for 51 peptides covering 100% of the LC.

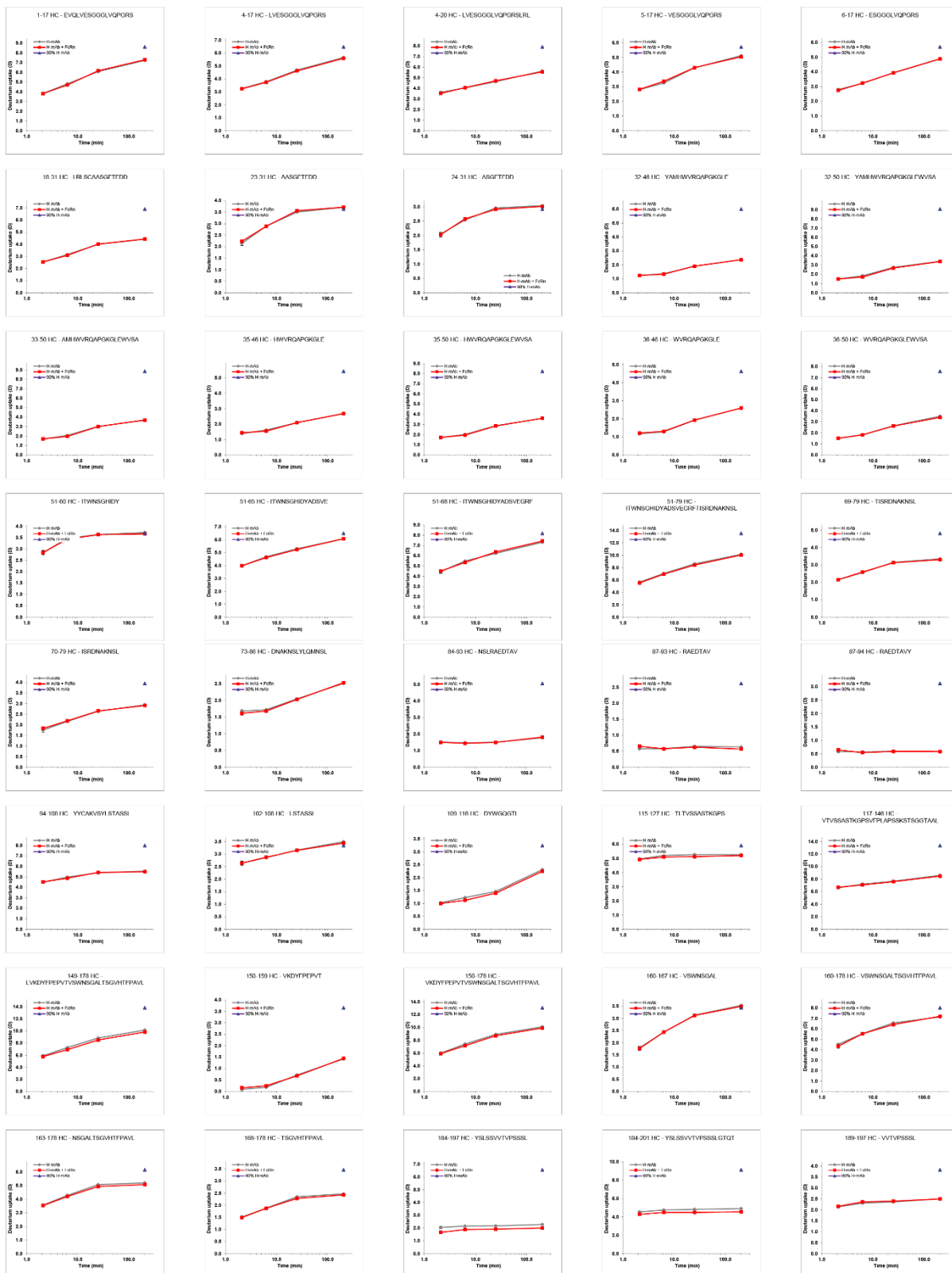
### C-mAb - HC

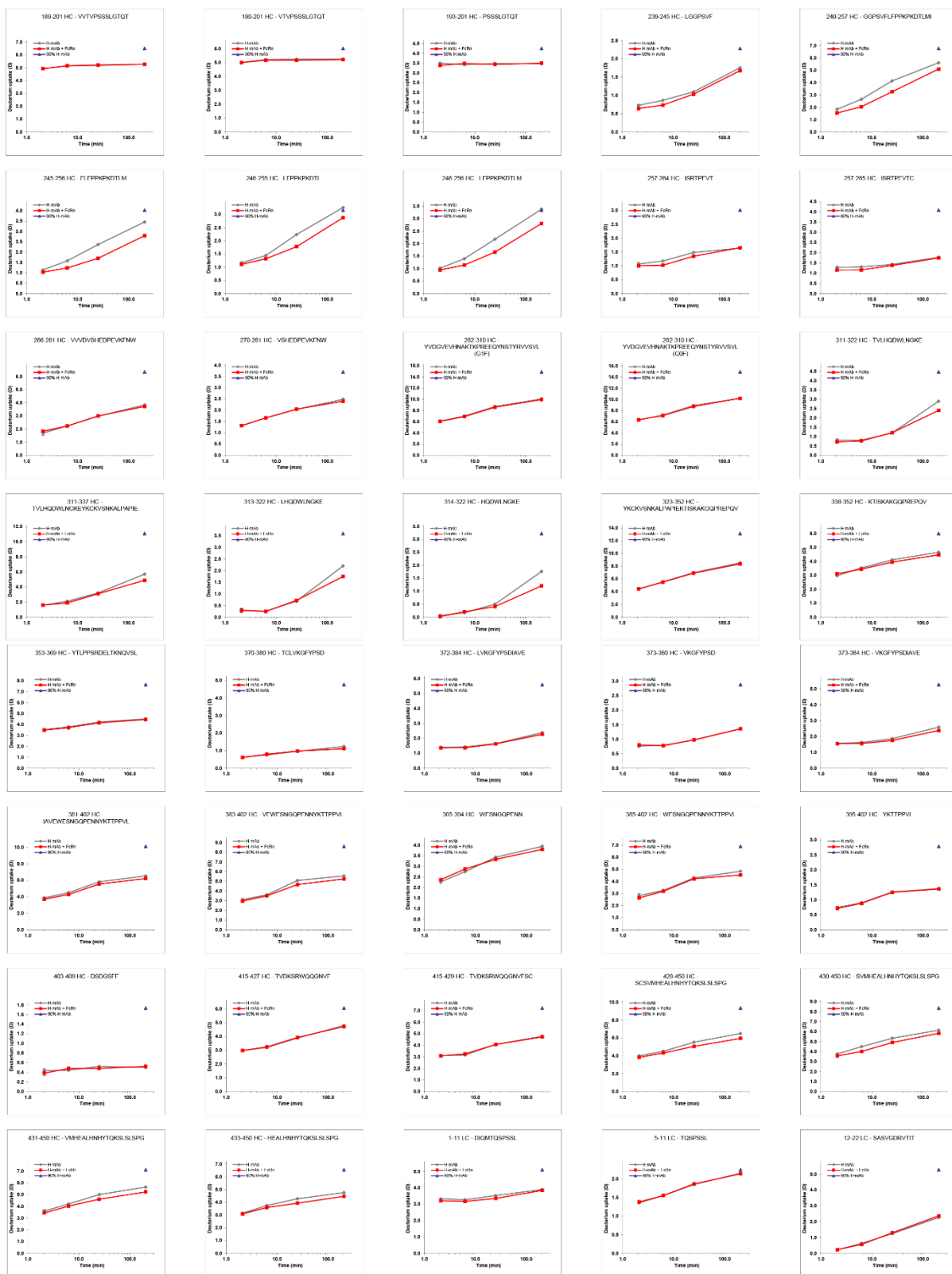


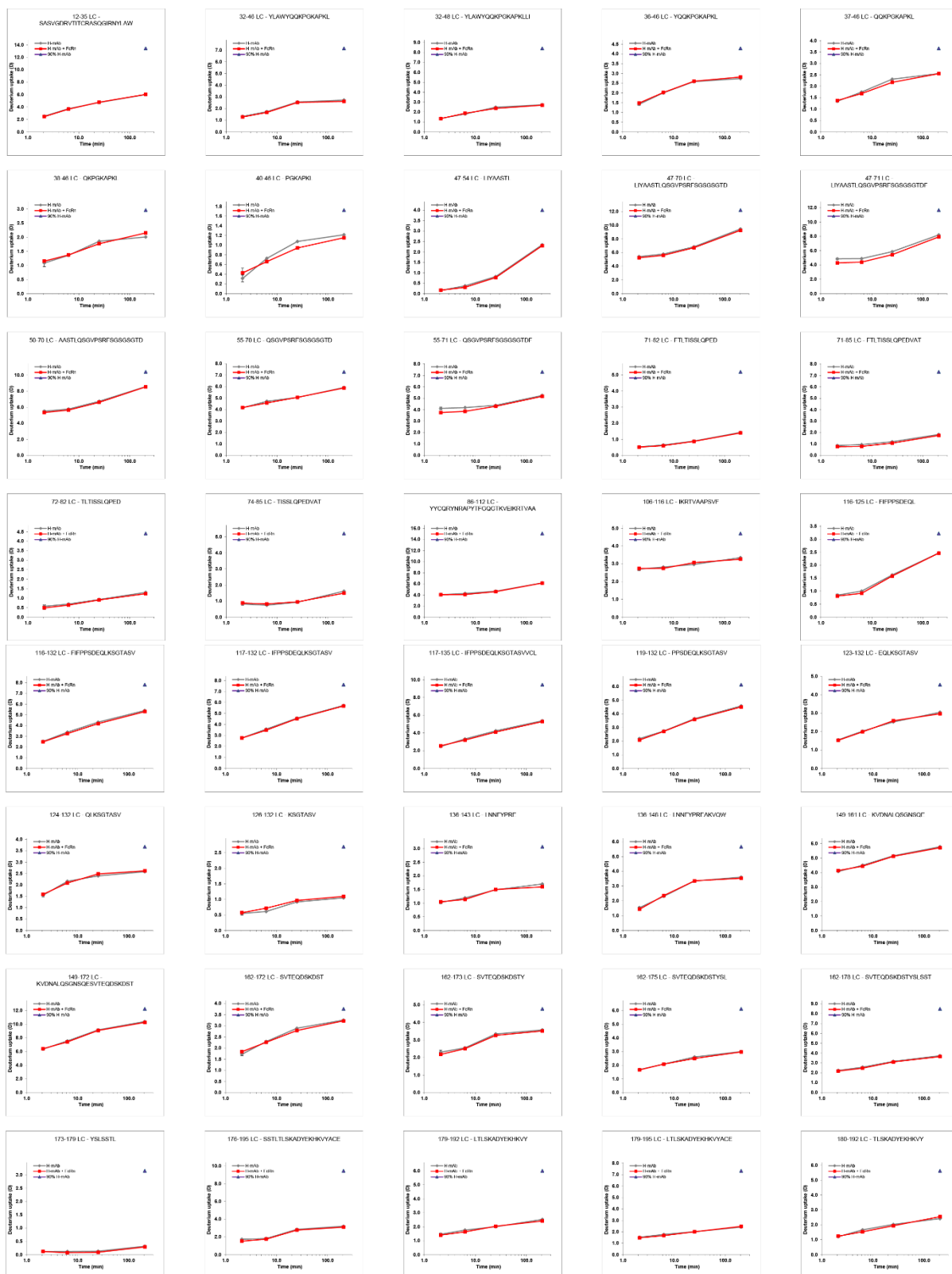
### C-mAb - LC

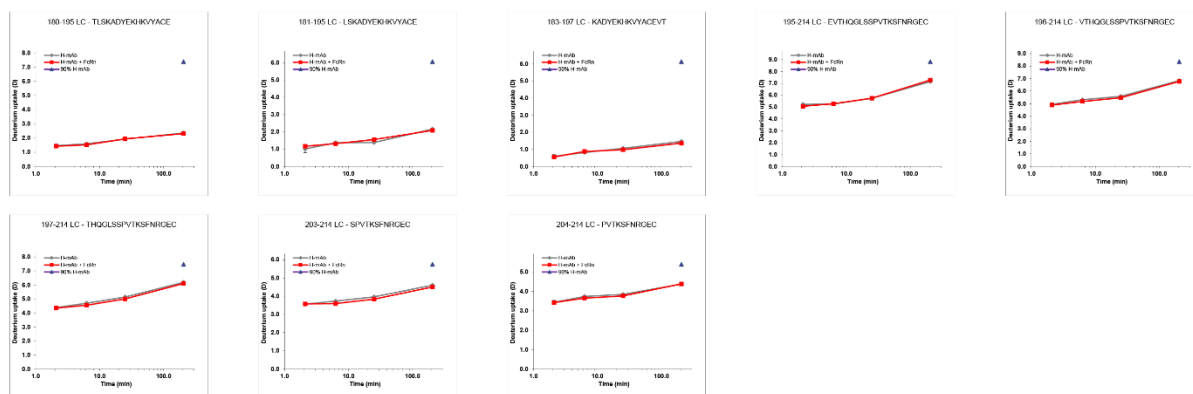


**Supporting Figure S9: Effective HDX-MS coverage map of C-mAb.** HDX data were obtained for 61 peptides covering 88.4% of the HC and for 39 peptides covering 94.9% of the LC.

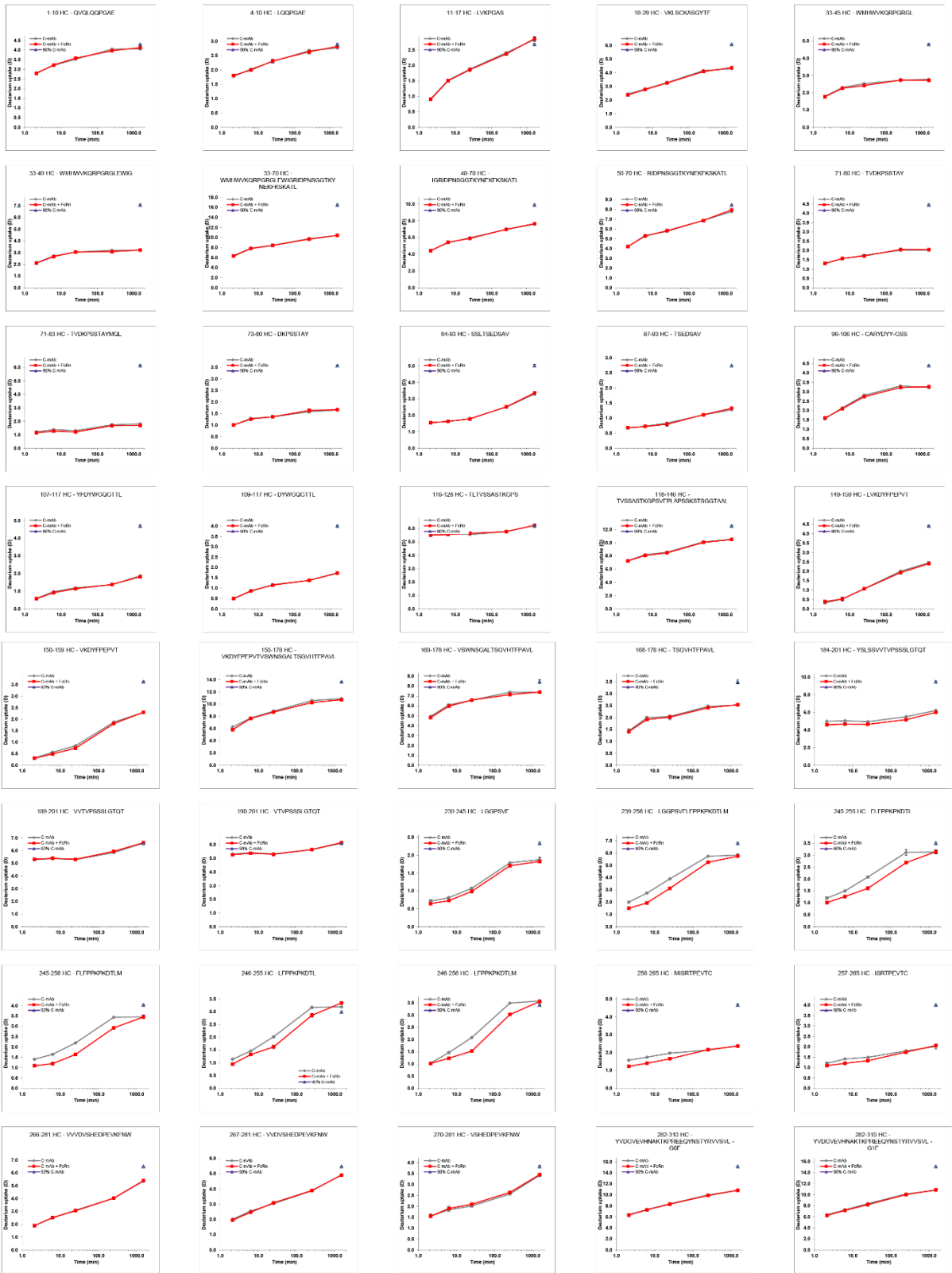


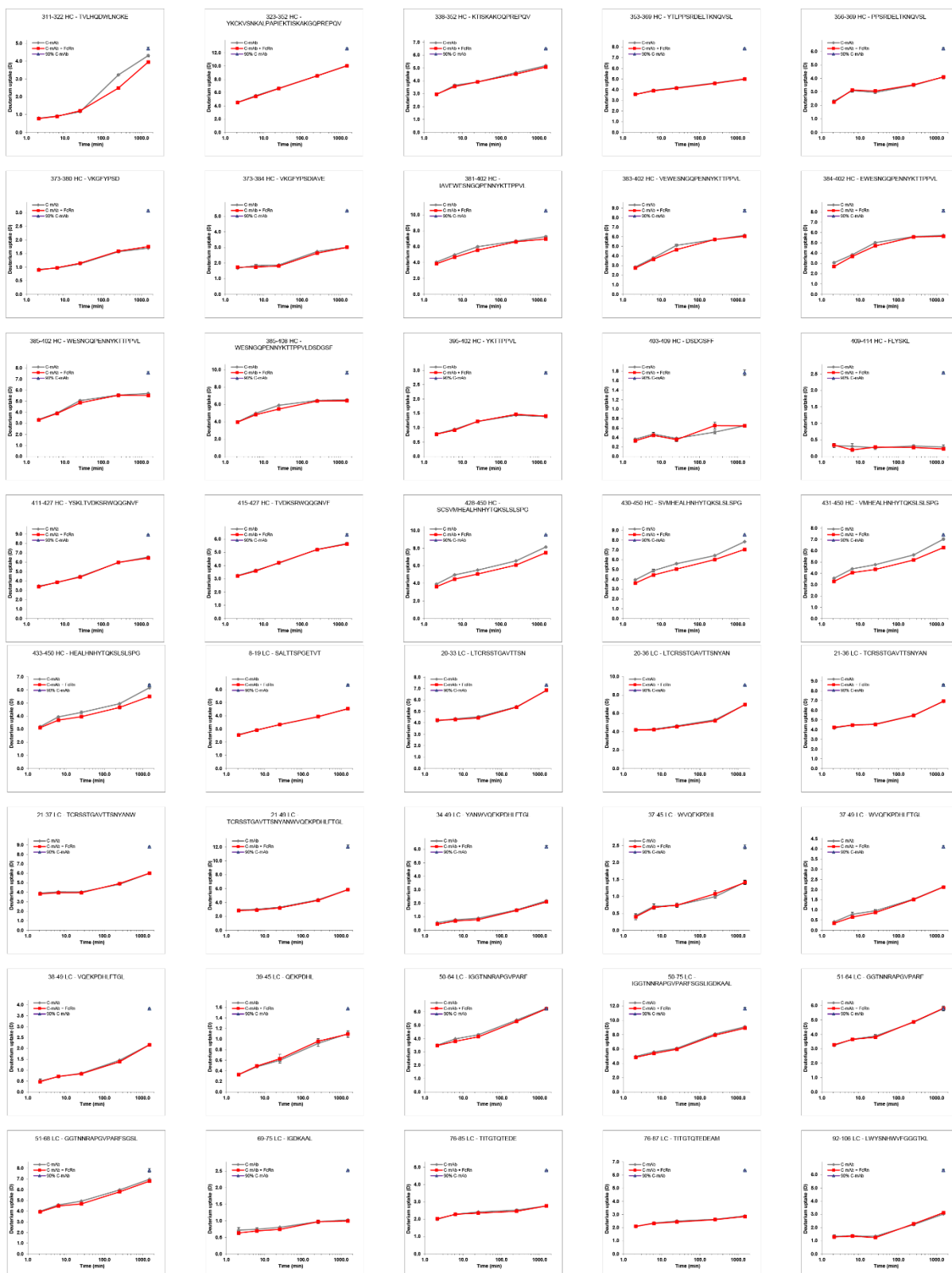




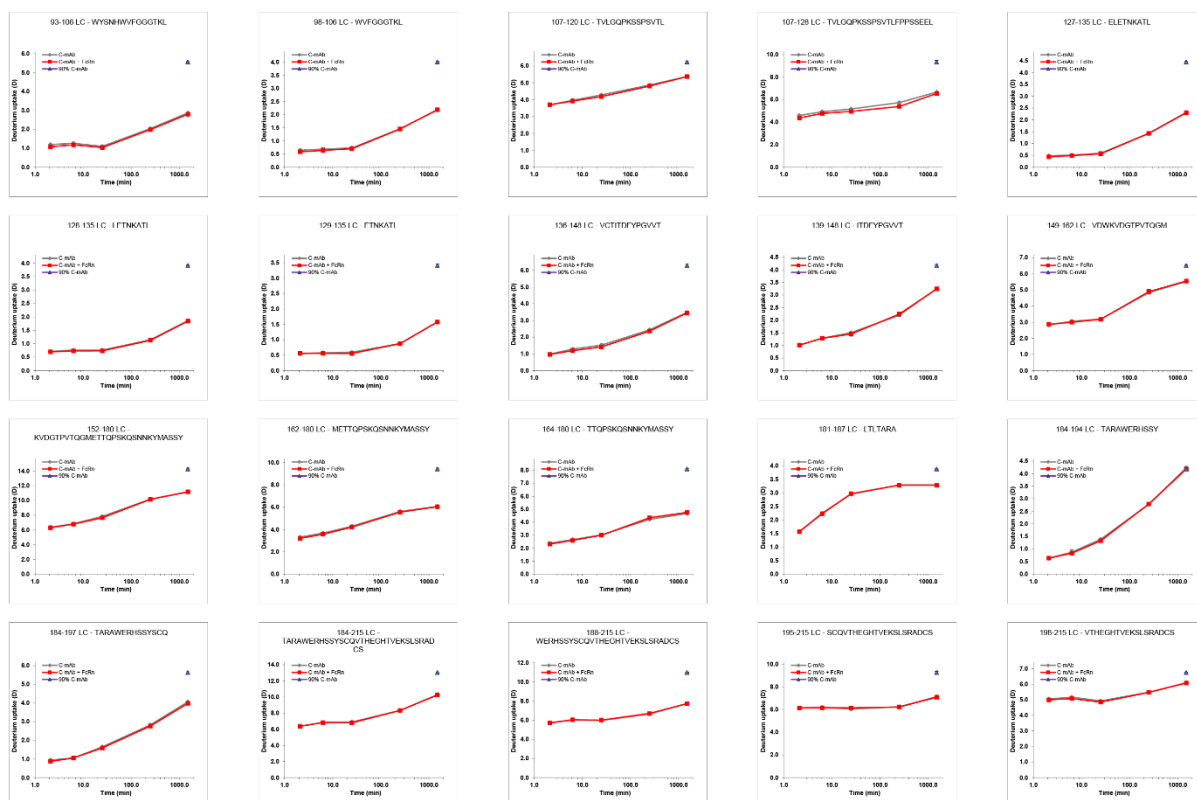


**Supporting Figure S10: HDX plots of H-mAb in the absence and presence of FcRn.** Absolute deuterium incorporation is plotted as a function of time for H-mAb (*grey* curves) and H-mAb in the presence of FcRn (*red* curves). Maximally labelled (90 %) control samples are plotted as *purple* triangles at the longest time points. SD is plotted as error bars (only slightly visible). ( $n = 1$  for all data points, except the first time point, where  $n=3$ ).







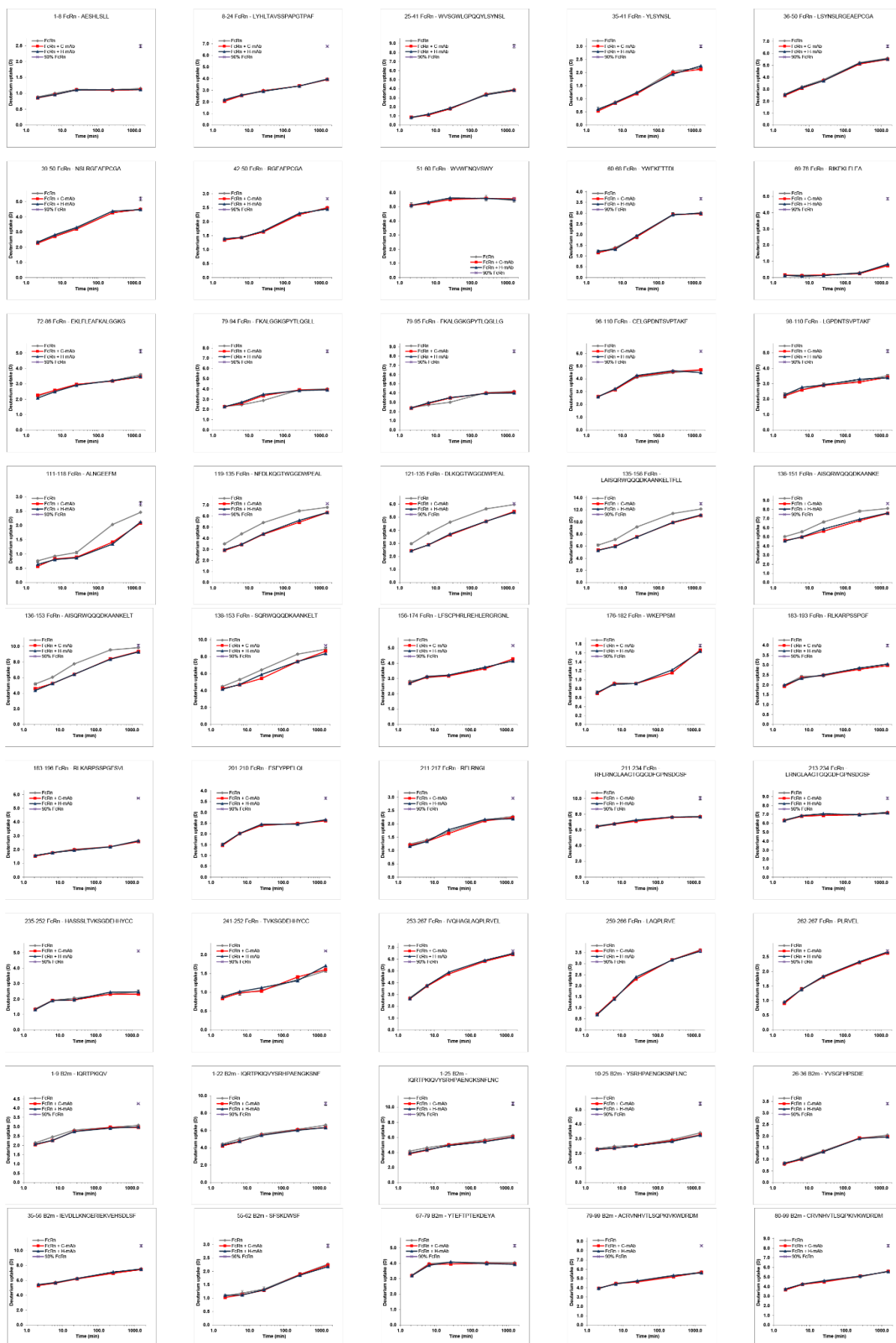


### Supporting Figure S11 HDX plots of C-mAb in the absence and presence of FcRn.

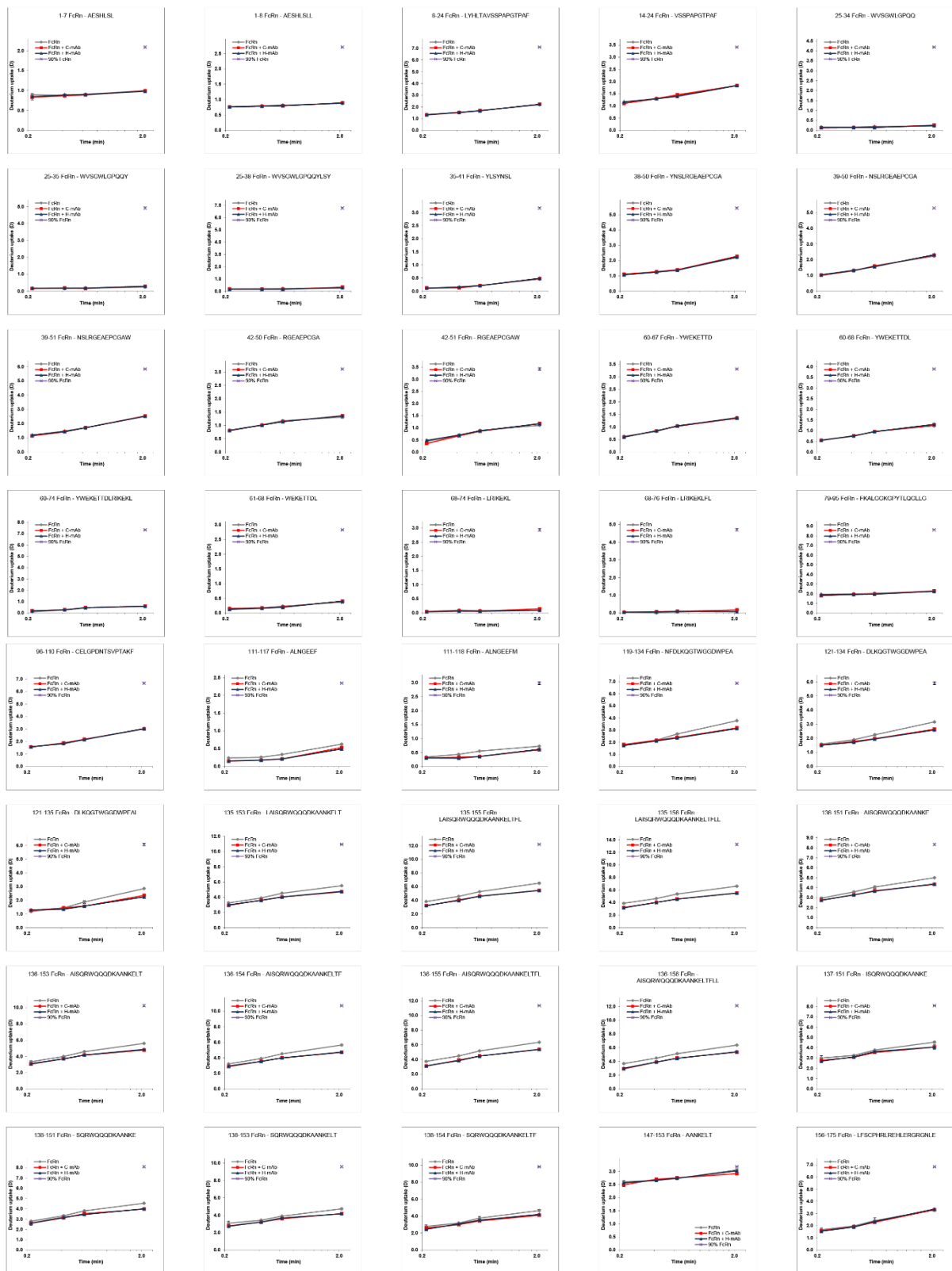
Absolute deuterium incorporation is plotted as a function of time for C-mAb (*grey curves*) and C-mAb in the presence of FcRn (*red curves*). Maximally labelled (90 %) control samples are plotted as *purple triangles* at the longest time points. SD is plotted as error bars (only slightly visible). (n=3).



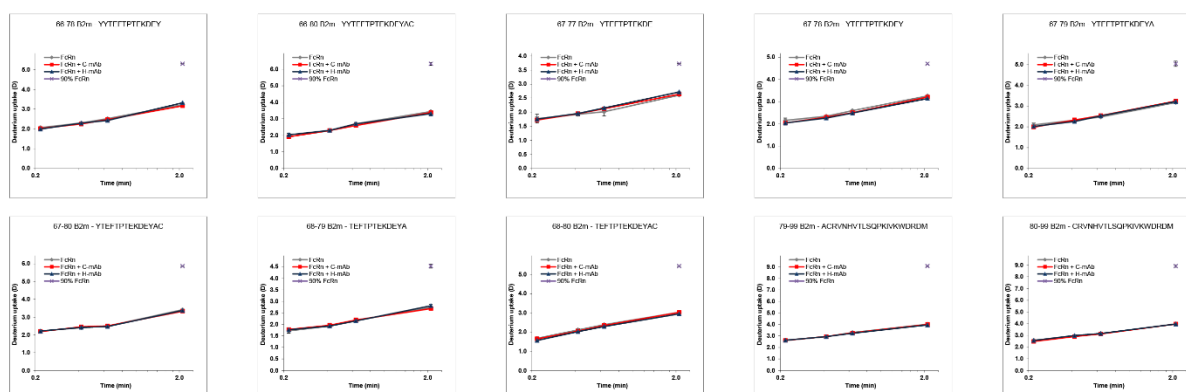
**Supporting Figure S12: Effective HDX-MS coverage map of FcRn.** Two separate data sets investigating two different exchange windows of FcRn in presence and absence of C-mAb and H-mAb were obtained. The first data set covered exchange times from 2.09 min to 1507.13 min (A), while the second data set covered exchange time from 13 s to 2.09 min (B). A) HDX data were obtained for 35 peptides covering 96.0% of the FcRn HC and for 10 peptides covering 96.0% of the β2m sequence. B) HDX data were obtained for 62 peptides covering 92.7% of the FcRn HC and for 28 peptides covering 97.0% of the β2m sequence.



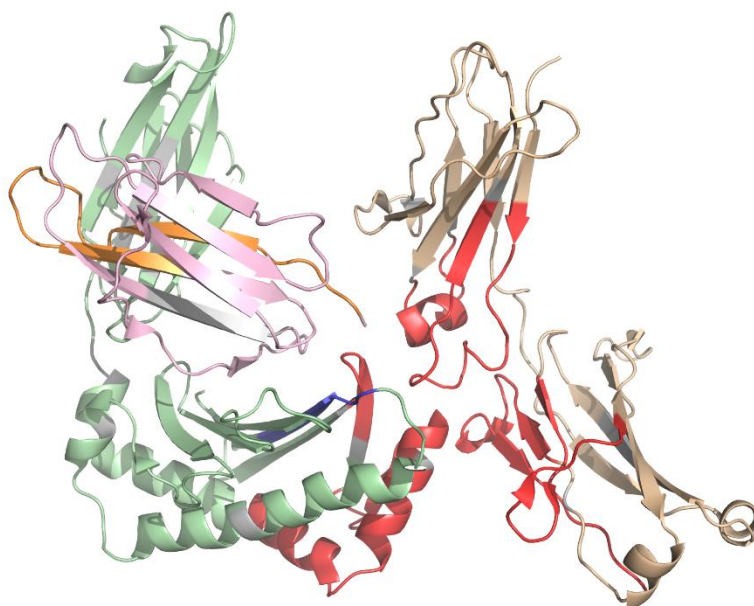
**Supporting Figure S13: HDX plots of FcRn in the absence and presence of either H-mAb or C-mAb (exchange time: 2.09 min to 1507.13 min).** Absolute deuterium incorporation is plotted as a function of time for FcRn (*grey* curves), FcRn in the presence of C-mAb (*red lines*) and FcRn in the presence of H-mAb (*blue* curves). Maximally labelled (90 %) control samples are plotted as *purple* crosses at the longest time points. SD is plotted as error bars (only slightly visible). (n = 3 for all data points, except the last four time points of FcRn in presence of H-mAb, where n=1).





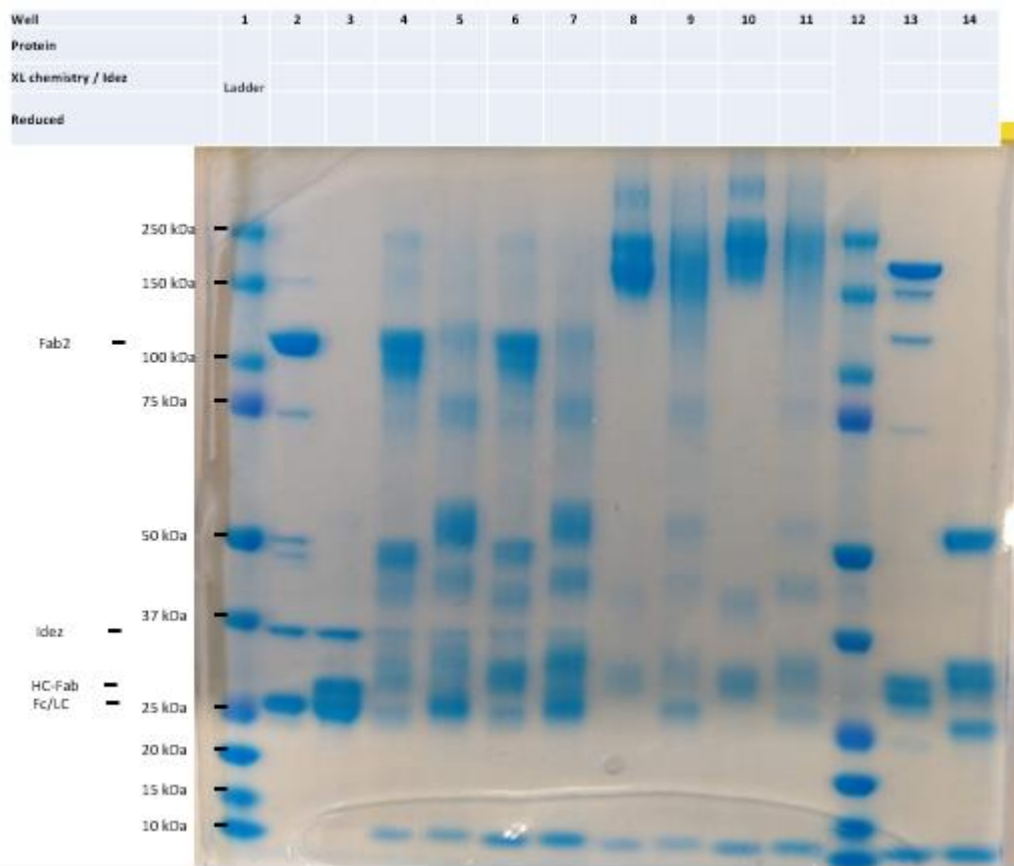


**Supporting Figure S14: HDX plots of FcRn in the absence and presence of either H-mAb or C-mAb (exchange time: 13 s to 2.09 min).** Absolute deuterium incorporation is plotted as a function of time for FcRn (*grey lines*), FcRn in the presence of C-mAb (*red lines*) and FcRn in the presence of H-mAb (*blue lines*). Maximally labelled (90 %) control samples are plotted as *purple crosses* at the longest time points. SD is plotted as error bars (only slightly visible). (n = 3 for all data points).



**Supporting Figure S15: Conformational response of complex formation between FcRn and C-mAb.** Differences in HDX mapped onto the crystal structure of FcRn in complex with Fc-YTE ( $\alpha$ -domain: *pale green*,  $\beta$ 2m: *light pink*, Fc-YTE: *wheat*, PDB ID: 4N0U). Regions in the  $\alpha$ -domain and  $\beta$ 2m of FcRn displaying a significant protection from exchange in the presence of H-mAb or C-mAb are colored *orange* and *red*, respectively. Regions displaying a significant deprotection from exchange in the presence of H-mAb or C-mAb are colored *blue*. Regions in the Fc-part of C-mAb displaying a significant protection from exchange in the presence of FcRn are colored *red*, while regions for which no HDX information could be obtained are colored *grey*.





**Supporting Figure S16: SDS-PAGE of IdeZ treated C-mAb.** Well 2 (non-reduced conditions) and 3 (reduced conditions) illustrate the successful cleavage of C-mAb by IdeZ.

## Supplemental References

1. Rice, P., Longden, I. & Bleasby, A. EMBOSS: the European Molecular Biology Open Software Suite. *Trends Genet* **16**, 276–7 (2000).
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3. Altschul, S. F. *et al.* Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res* **25**, 3389–402 (1997).
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