REVIEW

A role for G-CSF and GM-CSF in nonmyeloid cancers

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Granulocyte colony-stimulating factor (G-CSF) and granulocyte-macrophage

colony-stimulating factor (GM-CSF) modulate progression of certain solid

tumors. The G-CSF- or GM-CSF-secreting cancers, albeit not very common

are, however, among the most rapidly advancing ones due to a cytokine-medi-

ated immune suppression and angiogenesis. Similarly, de novo angiogenesis

and vasculogenesis may complicate adjuvant use of recombinant G-CSF or

GM-CSF thus possibly contributing to a cancer relapse. Rapid diagnostic tools

to differentiate G-CSF- or GM-CSF-secreting cancers are not well developed

therefore hindering efforts to individualize treatments for these patients. Given

an increasing utilization of adjuvant G-/GM-CSF in cancer therapy, we aimed

to summarize recent studies exploring their roles in pathophysiology of solid tumors and to provide insights into some complexities of their therapeutic

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Abstract

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Introduction

Granulocyte and granulocyte-macrophage colony-stimulating factors (G-CSF and GM-CSF, respectively) regulate maturation of progenitor cells in the bone marrow into differentiated granulocytes, macrophages, and the T cells. In clinical oncology, recombinant G- or GM-CSFs are routinely used to correct neutropenia subsequent to chemotherapy and radiation. However, adjuvant G-/GM-CSF treatments have been suggested to occasionally enable tumor growth. Such adverse treatment outcomes are thought to occur due to certain solid tumors being addicted to G-/GM-CSF-dependent signaling by expressing endogenous cytokines and/or their cognate receptors (G-/GM-CSFR). Clinical case reports reveal that newly diagnosed patients with G-/GM-CSF(R)-positive tumors

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present with an advanced often metastatic disease suggesting an accelerated progression of such cancers. The putative mechanisms of progression bear similarities to those observed with an adjuvant use of recombinant cytokines, that is, induction of immune tolerance and angiogenesis. It is therefore an imperative to explore roles of G-CSF and GM-CSF in cancer in order to improve treatment outcomes and to better define eligible patients' cohorts. In this review, we aimed to present recent advances in studies addressing putative mechanisms and therapeutic uses of G-CSF and GM-CSF in several cancers of a nonmyeloid origin.

Molecular mechanisms of G-CSF and GM-CSF signaling

G-CSF and GM-CSF receptor-ligand complexes

The G-CSF and GM-CSF are glycoproteins with molecular weights of 30 kDa and 22 kDa, respectively, secreted

by the cells of the immune system, fibroblasts, and endothelium. They function to stimulate granulopoiesis, the innate immunity, and the differentiation of neural progenitor cells [1–5]. Both cytokines require presence of their specific receptors to initiate intracellular signaling. Crystallographic studies depict activated G-CSFRs as tetramers where two receptor–ligand dimers form a complex on plasma membranes via Ig-like domains (Fig. 1A) [6]. In contrast, an activated GM-CSFR is a hexamer consisting of two ligand-selective α -subunits and the two nonselective β c subunits; each α -/ β c-subunits dimer is thought to bind one ligand molecule (Fig. 1B) [7–9]. Moreover, activation of downstream signaling requires the two hexamers to form a dodecameric complex, a feature is thought to be unique to a GM-CSFR [7].

Signal transduction pathways

In physiological conditions, for example, during maturation of granulocyte/macrophage precursors, activated

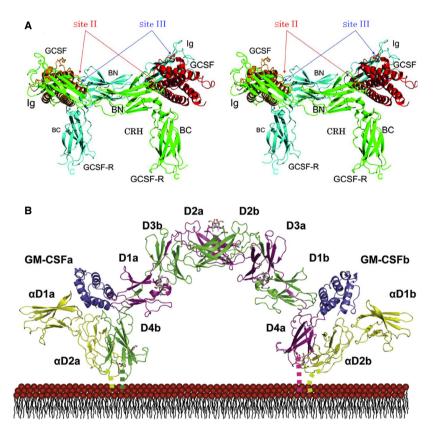


Figure 1. (A) The G-GSF receptor monomers (*green and blue*) consist of an extracellular Ig-like domain (*lg*), a cytokine receptor homologous (*CRH*) domain, and three fibronectin-type III-like domains followed by a transmembrane region and a cytoplasmic domain. Upon the G-CSF (*red*) binding, receptors polymerize via Ig-like domains to form an active signaling complex. (Modified from [6]). (B) The GM-CSF receptor is a complex of two α -subunits (*yellow*) and the two β c-subunits (*maroon and green*). The α -subunits ensure specificity of GM-CSF (*purple*) binding, whereas β c-subunits which are shared among GM-CSFR, IL-3R, and IL-5R provide high-affinity binding sites. The GM-CSFR localize extracellularly with domains of both α - and β c-subunits tethering them to the cell membranes (Modified from [7]).

G-/GM-CSFR elicit phosphorylation of JAK kinases and subsequent recruitment of STAT5 transcription factor to effect cellular differentiation [7, 10, 11]. G-CSF has also been shown to convey neuroprotection to central neurons upon increases in phosphorylation of PI3K/Akt pathway [7, 10, 11]. In cancers of nonmyeloid origin, however, the G-/GM-CSF signaling cascades are less known. Highly annotated automatic pathways analysis tools, such as MetaCoreTM (Thompson Reuters, New York) therefore may become indispensable in predicting such novel cascades. For example, Figure 2 outlines a map of putative signal transduction pathways whereby G-CSF or GM-CSF regulate epithelial to mesenchymal transition (EMT), a crucial event in malignant transformation. The map suggests that a recruitment of *c-jun* proto-oncogene may occur downstream from Lyn/JAK/STAT3 or, alternatively, MAPK/ERK1/2 pathways upon their activation by cytokines. Recruitment of *c-jun* would promote reorganization of actin cytoskeleton, secretion of matrix metalloproteases, and a loss of cell to cell contact to increase cell motility and hence to facilitate dissemination. Indeed, G-/GM-CSF-dependent phosphorylation of JAK2 and recruitment of STAT-3 have previously been reported as required steps in tumor angiogenesis and vascularization [12, 13]. Moreover, signaling cascades regulating EMT have been shown to convey stem cell phenotype to neoplastic cells which would account for their ability to metastasize and for their multidrug resistance [14]. To date, the contributions of G-/GM-CSF to cancer stem cell phenotypes are not clearly defined [15]. It is noteworthy

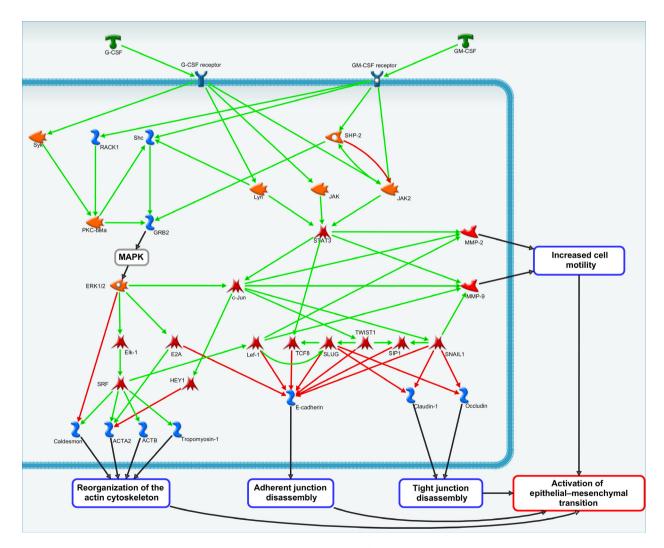


Figure 2. MetaCore[™] pathways analysis has been utilized to generate a map of putative signal transduction pathways for G-CSF or GM-CSF to regulate epithelial to mesenchymal transition in cancer. The green arrows indicate activated signaling pathways, whereas red arrows depict inhibited pathways.

therefore that the MetaCoreTM analysis proposes a role for G-/GM-CSF in maintaining a pool of stem cells in solid cancers via *c-jun*-dependent activation of SLUG, SNAIL1, or TWIST-1 transcription factors [16, 17]. Given a propensity of G-/GM-CSF(R)-expressing cancers toward accelerated growth and an early dissemination, it is feasible to predict that the experimental evidence will emerge as to their roles in sustaining a cancer stem cell phenotype.

latrogenic induction of G-/GM-CSFdependent tumor growth

Clinical reports and animal cancer models attest to the chemo- and/or antiangiogenic therapies or a radiation treatment inadvertently promoting cancer progression in part via G-CSF. Shaked et al. demonstrate a G-CSFdependent mobilization of endothelial progenitor cells and a tumor regrowth in murine models of melanoma and a lung cancer following treatments with vascular disrupting agents (VDA) [18]. Similar increases in plasma G-CSF levels have been detected in patients with solid tumors receiving VDA [19]. Remarkably, G-CSF fails to mobilize endothelial progenitor cells in mice not bearing tumors suggesting modulatory effects of tumor microenvironment on bone marrow responses to cytokines [20]. Gr1+CD11b+ myeloid-derived suppressor cells (MDSC) recruited to the tumor site may in part mediate these effects [21]. It has also been proposed that G-CSF, but not GM-CSF, expression with simultaneous infiltration of Gr1+CD11b+ MDSC render tumors refractory to the subsequent antiangiogenic treatments [21]. In murine models of pancreatic adenocarcinoma expressing RAS oncogene, the G-CSF-mediated resistance to antivascular endothelial growth factor (VEGF) therapies occurs through activation of RAS/MEK/ERK pathways and an Ets-induced overexpression of G-CSF [22]. Furthermore, studies suggest that by recruiting MDSC, G-CSF induces a VEGF-independent angiogenesis in addition to increasing resistance to anti-VEGF drugs [22]. It becomes apparent therefore that correction of cancer therapy-related neutropenia using recombinant G-/GM-CSF carries risks of disease recurrence thus necessitating stricter eligibility criteria for patients requiring these treatments.

Tumors secreting G-/GM-CSF

Lung cancer

Primary and metastatic lung cancers are by far the most frequently occurring type of malignancy driven by the ectopically secreted G-/GM-CSF [23–26]. Different histopathological types of a non-small-cell lung cancer (NSCLC) comprise a majority of cytokine-producing lung tumors although a case of G-CSF-secreting lung sarcoma has also been described [27, 28]. At a time of diagnosis, patients present with an advanced disease and a profound paraneoplastic leukocytosis. Elevated G-CSF levels have been proposed as a marker of shorter survival in NSCLC patients even if a subsequent resection of a cytokinesecreting tumor has been successful [24, 29].

Various cell types have been proposed as putative sources of G-/GM-CSF in lung cancers. Specifically, tumor-associated endothelial cells may secrete cytokines and thus promote angiogenesis and metastasis via increases in expression of cell adhesion molecules [30]. In addition, studies in animal models propose such function for Gr1+CD11b+ MDSC [31]. Surprisingly, microarray data reveal augmentations of a GM-CSF gene in small-cell lung cancers but not in NSCLC possibly suggesting posttranscriptional mechanisms for an increased secretion of this cytokine [32]. Given that G-/GM-CSF may accelerate progression and distant metastases in lung cancers, caution is warranted when using recombinant cytokines as an adjuvant treatment in these patients [33].

Glioma

Glioma is the most common type of a primary malignant brain tumor in adults with universally poor prognosis. Gliomas more often than other G-/GM-CSF-secreting cancers also express intratumoral cognate receptors; augmented G-/GM-CSF(R) levels have been found to correlate with higher tumor grade [34-36]. In these cancers, the G-/GM-CSF(R) promote progression possibly using auto-/paracrine activation of antiapoptotic and pro-angiogenic pathways via activation of STAT-3 transcription factor or an increased expression of VEGF/VEGFR [37-41]. The origins of cytokine- secreting cells in glioma are not completely known. For example, tumor cells and tumorassociated microglia, but not mesenchymal stromal cells, have been proposed for this role [42-44]. In experimental models of glioblastoma, decreasing G-/GM-CSF levels attenuate invasiveness and cell proliferation thus suggesting modulatory effects of these cytokines on tumor microenvironment [42]. Consistent with these findings, accumulation of MDSC concomitant with increases in G-CSF levels has been shown in patients with glioma [45]. Remarkably, a combination treatment consisting of a recombinant G-CSF and an interferon-gamma promotes maturation of tumor-associated dendritic cells [46, 47]. The induction of immune response by this treatment shows relative efficacy in prolonging survival in experimental gliomas [46, 47]. However, G-CSF monotherapy does not provide similar benefits which may be consistent with the aforementioned oncogenic roles of G-CSF

[45–48]. The lack of understanding of G-/GM-CSF roles in gliomas necessitates further research in pursuit of safer G-/GM-CSF-based therapies for patients with these cancers.

Bladder cancer

A G-CSF was initially purified from the human bladder carcinoma cell line 5637 [49] thus implying a role for this cytokine in progression of bladder malignancies. Tachibana et al. reported an autocrine growth induction after heterologous G-CSFR has been engineered into the G-CSF-secreting tumor cells from a resected bladder carcinoma [50]. Growth potentiation may have possibly occurred via beta1-integrin-dependent augmentation of cell adhesion and invasion [50-52]. In English scientific literature, clinical cases of bladder cancers secreting G-/GM-CSF or expressing their cognate receptors are not common [53-57]. Patients present with an advanced disease and a marked leukocytosis in the absence of secondary infection or a myeloproliferative disorder [53-57]. Peripheral blood smears show an abundance of differentiated neutrophils without left shift consistent with an ectopic induction of normal granulopoiesis [54]. Responses to treatments are noticeably variable. Normalization of the white blood cell counts and cytokine levels upon successful resection of a tumor has been reported [53-57]. Conversely, rapid metastatic spread and patient's demise despite therapeutic interventions has also been documented [53-57]. Refractory cases are speculated to reflect possible involvement of endogenous tumor G-/GM-CSFRs [54]. The microarray data report small but statistically significant increases in gene expression for both GM-CSF and a-subunits of GM-CSFR in bladder cancers compared to normal tissues [58, 59]. To the contrary, changes in G-CSF or its receptor gene expression have not been found [58, 59]. Given the rarity of G-/GM-CSF-secreting bladder tumors and an increasing use of recombinant G-/GM-CSF, appropriate diagnostic approaches are needed to identify patients to whom such therapies may be detrimental [60-62].

Colorectal cancers

The elevated GM-CSF plasma levels have been found in certain patients with colorectal cancers thus implying this cytokine contributions to a disease progression [63]. Gene expression arrays identified more than one third of human and murine colorectal cancers as secreting GM-CSF [64]. Unlike in many other cancers, however, ectopic secretion of GM-CSF driven by demethylation of its gene promoter conveys antitumor effects that are

both immune mediated and immune independent [64]. The immune-independent effects require GM-CSFR which upon ligand binding significantly attenuates tumor formation [64]. Moreover, patients with colorectal cancers whose tumors test positive for GM-CSF/GM-CSFR show improved overall 5-year survival [64]. In contrast, clinical case reports of G-CSF-secreting cancers of colon and rectum describe patients presenting with large tumors and distal metastases [65, 66]. Despite surgical resection resulting in reduced G-CSF plasma levels, the overall survival of these patients is poor implying oncogenic functions for G-CSF [65, 66]. The aforementioned studies thus propose differential roles for GM-CSF and G-CSF in cancers of colon and rectum. This knowledge would undoubtedly translate into the individualized use of (neo)adjuvant cytokines depending on a molecular profile of a particular tumor.

Melanoma and skin carcinoma

Melanoma is a rapidly progressing incurable skin cancer with a propensity to metastasize early due to immunosuppression and growth induction in part occurring via G-/GM-CSF. Clinical case studies report existence of are albeit severe G-CSF-secreting melanomas [67, 68]. That necessitates their consideration in a differential diagnosis given the use of adjuvant GM-CSF in these patients [67, 68]. Melanoma cells in vitro have been found to express G-CSFR transcript; increases in cell proliferation, however, do not occur upon G-CSF stimulation possibly reflecting an absence of a G-CSFR protein [69]. In contrast, a role of GM-CSF in dissemination of melanoma is controversial. Studies using a murine model indicate that under hypoxic conditions tumor-associated macrophages upon stimulation with GM-CSF secrete a soluble VEGF receptor [70]. The receptor inactivates VEGF and thus exerts antiangiogenic effects [70]. A simultaneous stabilization of a hypoxia-induced transcription factor HIF-2a augments transcription of a VEGF receptor gene and enhances the antiangiogenic actions of GM-CSF in this model [70]. In agreement with these observations, clinical trials of adjuvant GM-CSF monotherapy in patients with locally advanced melanoma demonstrate a decrease in the melanoma-specific deaths without improvements in a disease-free survival [71]. However, other studies also utilizing murine models of melanoma have found positive correlations between increased GM-CSF levels and growth of lesions [72]. Furthermore, a cytokine-dependent infiltration of tumors with Gr1+CD11+ MDSC consistent with tumorigenic effects of GM-CSF has also been documented [72]. Controversies concerning the roles of G-/GM-CSF in melanoma may arise in part due to their complex synergistic inputs into a disease progression

similar to those found in skin carcinoma [73]. Authors of this study demonstrate that synergy between G-CSF and GM-CSF augments cell proliferation and invasiveness in addition to an early recruitment of tumor-associated macrophages to a tumor site [73]. Thus, G-CSF in melanoma and skin carcinomas exacerbates disease progression due to pro-angiogenic and immunosuppressive actions. To the contrary, GM-CSF may demonstrate antitumor activity via modulating recruitment of tumor-associated macrophages and their VEGFR secretion.

Bone metastases in cancers of prostate and breast

A role for G- or GM-CSF in cancers of prostate is less defined. Reports of G-/GM-CSF-secreting prostate tumors in English scientific literature are uncommon thus implying lesser significance of these cytokines for a prostate cancer progression. However, in vitro and animal models of this disease implicate G-/GM-CSF in promoting dissemination and bone metastasis. Namely, costimulation with G-CSF and a stem cell factor enhances cancer stem cell phenotype via upregulation of Oct3/4 transcription factor, NANOGP8 pseudogene and ABCG2 transporter [74]. In murine xenograft models, GM-CSF has been found to facilitate metastatic seeding of prostate cancer cells in the bone by enhancing osteoclastic activity [75]. Interestingly, this phenomenon has been observed while animals were undergoing a treatment with GM-CSF for a

 $\label{eq:constraint} \textbf{Table 1.} \ \text{Summary of pro- and antitumorigenic roles of G-CSF and GM-CSF.}$

Tumor type	G-CSF	GM-CSF
NSCLC	Angiogenic, immunosuppressive via MDSC	
Glioma	Auto-/paracrine growth stimulation	
Bladder carcinoma	Auto-/paracrine growth stimulation	?
Colorectal cancer	Tumorigenic	Immune-mediated and immune-independent tumor suppression
Melanoma	Tumorigenic	Antiangiogenic via soluble VEGFR Tumorigenic via MDSC
Skin cancer	Synergistically tumorigenic and angiogenic	
Bone metastases	Sustain cancer stem cell phenotype	

G-CSF and GM-CSF differentially regulate tumor growth and metastasis in solid cancers. NSCLC, non-small-cell lung cancer; VEGFR, vascular endothelial growth factor receptor; MDSC, myeloid-derived suppressor cells; ?, no discernible evidence of an oncogenic or a tumor suppressor role has been found. chemotherapy-induced leukopenia hence suggesting growth-promoting effects of a therapeutic GM-CSF [75]. Similar exacerbation of an osteoclastic bone resorption has been reported in patients with breast cancer that metastasized to the bone [76]. Specifically, in metastatic tissues the NF-kappaB transcription factor has been found to target GM-CSF gene to activate osteoclastogenesis and thus to facilitate homing of malignant cells in bone tissues. Given that cancer dissemination reflects a presence of cancer stem cells, their activation via administration of G-CSF has been proposed as a therapeutic strategy to augment their chemosensitivity [77]. In this study, authors speculate that an increased relapse-free survival of breast cancer patients who received an adjuvant G-CSF is due to a diminished drug resistance of neoplastic cells populating bone micrometastases.

Conclusions

Solid tumors of every origin present with many dissimilarities in their cellular composition and the molecular mechanisms of progression. Tumors addicted to G-/GM-CSF(R) signaling represents a very distinct molecular subset among other malignancies. For example, they utilize signaling pathways of normal hematopoiesis and the cytokine-mediated auto-/paracrine growth augmentation in addition to immune suppression. The aforementioned signaling modalities albeit not common in solid neoplasms contribute to some of the most advanced cases. Table 1 briefly summarizes the pro- and antitumorigenic roles of G-/GM-CSF in cancers of different origins. Prompt diagnosis based on a tumor cytokine/receptor profile would aid in individualizing the anticancer treatment choices for these patients. In particular, stricter eligibility criteria for adjuvant use of G-/GM-CSF would prevent certain adverse effects, for example, exacerbation of tumor growth in cancers addicted to G-/GM-CSF.

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Conflict of Interest

None declared.

References

- 1. Fleetwood, A. J., A. D. Cook, and J. A. Hamilton. 2005. Functions of granulocyte-macrophage colony-stimulating factor. Crit. Rev. Immunol. 25:405–428.
- Shibasaki, T., N. Katayama, K. Ohishi, A. Fujieda, F. Monma, K. Nishi, et al. 2007. IL-3 cannot replace GM-CSF in inducing human monocytes to differentiate into langerhans cells. Int. J. Oncol. 30:549–555.
- 3. Metcalf, D. 2010. The colony-stimulating factors and cancer. Nat. Rev. Cancer 10:425–434.
- Codarri, L., G. Gyülvészi, V. Tosevski, L. Hesske, A. Fontana, L. Magnenat, et al. 2011. RORγt drives production of the cytokine GM-CSF in helper T cells, which is essential for the effector phase of autoimmune neuroinflammation. Nat. Immunol. 12:560–567.
- El-Behi, M., B. Ciric, H. Dai, Y. Yan, M. Cullimore, F. Safavi, et al. 2011. The encephalitogenicity of T(H)17 cells is dependent on IL-1- and IL-23-induced production of the cytokine GM-CSF. Nat. Immunol. 12:568–575.
- Tamada, T., E. Honjo, Y. Maeda, T. Okamoto, M. Ishibashi, M. Tokunaga, et al. 2006. Homodimeric cross-over structure of the human granulocyte colony-stimulating factor (GCSF) receptor signaling complex. Proc. Natl Acad. Sci. USA 103:3135–3140.
- Hansen, G., T. R. Hercus, B. J. McClure, F. C. Stomski, M. Dottore, J. Powell, et al. 2008. The structure of the GM-CSF receptor complex reveals a distinct mode of cytokine receptor activation. Cell 134:496–507.
- Hercus, T. R., D. Thomas, M. A. Guthridge, P. G. Ekert, J. King-Scott, M. W. Parker, et al. 2009. The granulocyte-macrophage colony-stimulating factor receptor: linking its structure to cell signaling and its role in disease. Blood 114:1289–1298.
- Hercus, T. R., S. E. Broughton, P. G. Ekert, H. S. Ramshaw, M. Perugini, M. Grimbaldeston, et al. 2012. The GM-CSF receptor family: mechanism of activation and implications for disease. Growth Factors 30:63–75.
- Hamilton, J. A. 2008. Colony-stimulating factors in inflammation and autoimmunity. Nat. Rev. Immunol. 8:533–544.
- 11. Kaushansky, K. 2006. Lineage-specific hematopoietic growth factors. N. Engl. J. Med. 354:2034–2045.
- Valdembri, D., G. Serini, A. Vacca, D. Ribatti, and F. Bussolino. 2002. In vivo activation of JAK2/STAT-3 pathway during angiogenesis induced by GM-CSF. FASEB J. 16:225–227.
- Zgheib, A., S. Lamy, and B. Annabi. 2013. Epigallocatechin-gallate targeting of membrane type-1 matrix metalloproteinase-mediated Src and JAK/STAT3 signalling inhibits transcription of colony stimulating factors-2 and -3 in mesenchymal stromal cells. J. Biol. Chem. 288:13378–13386.

- West, N. R., J. I. Murray, and P. H. Watson. 2013. Oncostatin-M promotes phenotypic changes associated with mesenchymal and stem cell-like differentiation in breast cancer. Oncogene 33:1485–1494.
- Levina, V., A. M. Marrangoni, R. DeMarco, E. Gorelik, and A. E. Lokshin. 2008. Drug-selected human lung cancer stem cells: cytokine network, tumorigenic and metastatic properties. PLoS ONE 3:e3077.
- Banerjee, A., P. Qian, Z. S. Wu, X. Ren, M. Steiner, N. M. Bougen, et al. 2012. Artemin stimulates radio- and chemo-resistance by promoting TWIST1-BCL-2-dependent cancer stem cell-like behavior in mammary carcinoma cells. J. Biol. Chem. 287:42502–42515.
- Izumiya, M., A. Kabashima, H. Higuchi, T. Igarashi, G. Sakai, H. Iizuka, et al. 2012. Chemoresistance is associated with cancer stem cell-like properties and epithelial-to-mesenchymal transition in pancreatic cancer cells. Anticancer Res. 32:3847–3853.
- Shaked, Y., T. Tang, J. Woloszynek, L. G. Daenen, S. Man, P. Xu, et al. 2009. Contribution of granulocyte colony-stimulating factor to the acute mobilization of endothelial precursor cells by vascular disrupting agents. Cancer Res. 69:7524–7528.
- Langenberg, M. H., M. W. Nijkamp, J. M. Roodhart, N. Snoeren, T. Tang, Y. Shaked, et al. 2010. Liver surgery induces an immediate mobilization of progenitor cells in liver cancer patients: a potential role for G-CSF. Cancer Biol. Ther. 9:743–748.
- 20. Okazaki, T., S. Ebihara, M. Asada, A. Kanda, H. Sasaki, and M. Yamaya. 2006. Granulocyte colony-stimulating factor promotes tumor angiogenesis via increasing circulating endothelial progenitor cells and Gr1+CD11b+ cells in cancer animal models. Int. Immunol. 18:1–9.
- 21. Shojaei, F., X. Wu, X. Qu, M. Kowanetz, L. Yu, M. Tan, et al. 2009. G-CSF-initiated myeloid cell mobilization and angiogenesis mediate tumor refractoriness to anti-VEGF therapy in mouse models. Proc. Natl Acad. Sci. USA 106:6742–6747.
- 22. Phan, V. T., X. Wu, J. H. Cheng, R. X. Sheng, A. S. Chung, G. Zhuang, et al. 2013. Oncogenic RAS pathway activation promotes resistance to anti-VEGF therapy through G-CSF-induced neutrophil recruitment. Proc. Natl Acad. Sci. USA 110:6079–6084.
- Lammel, V., C. Stoeckle, B. Padberg, R. Zweifel, D. L. Kienle, W. H. Reinhart, et al. 2012. Hypereosinophilia driven by GM-CSF in large-cell carcinoma of the lung. Lung Cancer 76:493–495.
- Fukutomi, T., M. Kohno, Y. Izumi, M. Watanabe, Y. Hayashi, and H. Nomori. 2012. Pulmonary pleomorphic carcinoma producing granulocyte-macrophage colony-stimulating factor: report of a case. Surg. Today 42:288–291.
- Bahar, B., B. Acedil Ayc Iota, U. Çoşkun, S. Büyükberber, M. Benekli, and R. Yildiz. 2010. Granulocyte colony

stimulating factor (G-CSF) and macrophage colony stimulating factor (M-CSF) as potential tumor markers in non small cell lung cancer diagnosis. Asian Pac. J. Cancer Prev. 11:709–712.

- Shalom, G., N. Sion-Vardy, J. Dudnik, and S. Ariad. 2010. Leukemoid reaction in lung cancer patients. Isr. Med. Assoc. J. 12:255–256.
- Granger, J. M., and D. P. Kontoyiannis. 2009. Etiology and outcome of extreme leukocytosis in 758 nonhematologic cancer patients: a retrospective, single-institution study. Cancer 115:3817–4041.
- Jardin, F., M. Vasse, M. Debled, S. Dominique, P. Courville, F. Callonnec, et al. 2005. Intense paraneoplastic neutrophilic leukemoid reaction related to a G-CSF-secreting lung sarcoma. Am. J. Hematol. 80:243– 245.
- Stathopoulos, G. P., A. Armakolas, T. Tranga, H. Marinou, J. Stathopoulos, and H. Chandrinou. 2011. Granulocyte colony-stimulating factor expression as a prognostic biomarker in non-small cell lung cancer. Oncol. Rep. 25:1541–1544.
- 30. Chen, C., C. A. Duckworth, Q. Zhao, D. M. Pritchard, J. M. Rhodes, and L. G. Yu. 2013. Increased circulation of galectin-3 in cancer induces secretion of metastasis-promoting cytokines from blood vascular endothelium. Clin. Cancer Res. 19:1693–1704.
- Younos, I., M. Donkor, T. Hoke, A. Dafferner, H. Samson, S. Westphal, et al. 2011. Tumor- and organ-dependent infiltration by myeloid-derived suppressor cells. Int. Immunopharmacol. 11:816–826.
- Bhattacharjee, A., W. G. Richards, J. Staunton, C. Li, S. Monti, P. Vasa, et al. 2001. Classification of human lung carcinomas by mRNA expression profiling reveals distinct adenocarcinoma subclasses. Proc. Natl Acad. Sci. USA 98:13790–13795.
- 33. Kowanetz, M., X. Wu, J. Lee, M. Tan, T. Hagenbeek, X. Qu, et al. 2010. Granulocyte-colony stimulating factor promotes lung metastasis through mobilization of Ly6G+Ly6C+ granulocytes. Proc. Natl Acad. Sci. USA 107:21248–21255.
- 34. Wang, J., L. Yao, S. Zhao, X. Zhang, J. Yin, Y. Zhang, et al. 2012. Granulocyte-colony stimulating factor promotes proliferation, migration and invasion in glioma cells. Cancer Biol. Ther. 13:389–400.
- Revoltella, R. P., M. Menicagli, and D. Campani. 2012. Granulocyte-macrophage colony-stimulating factor as an autocrine survival-growth factor in human gliomas. Cytokine 57:347–359.
- Karcher, S., H. H. Steiner, R. Ahmadi, S. Zoubaa, G. Vasvari, H. Bauer, et al. 2006. Different angiogenic phenotypes in primary and secondary glioblastomas. Int. J. Cancer 118:2182–2189.
- Brantley, E. C., L. B. Nabors, G. Y. Gillespie, Y. H. Choi, C. A. Palmer, K. Harrison, et al. 2008. Loss of protein

inhibitors of activated STAT-3 expression in glioblastoma multiforme tumors: implications for STAT-3 activation and gene expression. Clin. Cancer Res. 14:4694–4704.

- Weissenberger, J., S. Loeffler, A. Kappeler, M. Kopf, A. Lukes, T. A. Afanasieva, et al. 2004. IL-6 is required for glioma development in a mouse model. Oncogene 23:3308–3316.
- 39. Niola, F., C. Evangelisti, L. Campagnolo, S. Massalini, M. C. Buè, A. Mangiola, et al. 2006. A plasmid-encoded VEGF siRNA reduces glioblastoma angiogenesis and its combination with interleukin-4 blocks tumor growth in a xenograft mouse model. Cancer Biol. Ther. 5:174–179.
- 40. Jung, K. H., K. Chu, S. T. Lee, S. J. Kim, D. I. Sinn, S. U. Kim, et al. 2006. Granulocyte colony-stimulating factor stimulates neurogenesis via vascular endothelial growth factor with STAT activation. Brain Res. 1073–1074:190–201.
- Ohki, Y., B. Heissig, Y. Sato, H. Akiyama, Z. Zhu, D. J. Hicklin, et al. 2005. Granulocyte colony-stimulating factor promotes neovascularization by releasing vascular endothelial growth factor from neutrophils. FASEB J. 19:2005–2007.
- 42. Curran, C. S., M. D. Evans, and P. J. Bertics. 2011. GM-CSF production by glioblastoma cells has a functional role in eosinophil survival, activation, and growth factor production for enhanced tumor cell proliferation. J. Immunol. 187:1254–1263.
- 43. Gabrusiewicz, K., A. Ellert-Miklaszewska, M. Lipko, M. Sielska, M. Frankowska, and B. Kaminska. 2011. Characteristics of the alternative phenotype of microglia/ macrophages and its modulation in experimental gliomas. PLoS ONE 6:e23902.
- 44. Kucerova, L., M. Matuskova, K. Hlubinova, V. Altanerova, and C. Altaner. 2010. Tumor cell behaviour modulation by mesenchymal stromal cells. Mol. Cancer 9:129.
- Raychaudhuri, B., P. Rayman, J. Ireland, J. Ko, B. Rini, E. C. Borden, et al. 2011. Myeloid-derived suppressor cell accumulation and function in patients with newly diagnosed glioblastoma. Neuro. Oncol. 13:591–599.
- 46. Smith, K. E., S. Fritzell, W. Badn, S. Eberstál, S. Janelidze, E. Visse, et al. 2009. Cure of established GL261 mouse gliomas after combined immunotherapy with GM-CSF and IFNgamma is mediated by both CD8+ and CD4+ T-cells. Int. J. Cancer 124:630–637.
- 47. Smith, K. E., S. Janelidze, E. Visse, W. Badn, L. Salford, P. Siesjö, et al. 2007. Synergism between GM-CSF and IFNgamma: enhanced immunotherapy in mice with glioma. Int. J. Cancer 120:75–80.
- Mariani, C. L., D. Rajon, F. J. Bova, and W. J. Streit. 2007. Nonspecific immunotherapy with intratumoral lipopolysaccharide and zymosan A but not GM-CSF leads to an effective anti-tumor response in subcutaneous RG-2 gliomas. J. Neurooncol. 85:231–240.
- 49. Welte, K., E. Platzer, L. Lu, J. L. Gabrilove, E. Levi, R. Mertelsmann, et al. 1985. Purification and biochemical

characterization of human pluripotent hematopoietic colony-stimulating factor. Proc. Natl Acad. Sci. USA 82:1526–1530.

- Tachibana, M., A. Miyakawa, A. Uchida, M. Murai, K. Eguchi, K. Nakamura, et al. 1997. Granulocyte colony-stimulating factor receptor expression on human transitional cell carcinoma of the bladder. Br. J. Cancer 75:1489–1496.
- Chakraborty, A., S. M. White, and S. Guha. 2006. Granulocyte colony-stimulating receptor promotes beta1-integrin-mediated adhesion and invasion of bladder cancer cells. Urology 68:208–213.
- Chakraborty, A., and S. Guha. 2007. Granulocyte colony-stimulating factor/granulocyte colony-stimulating factor receptor biological axis promotes survival and growth of bladder cancer cells. Urology 69:1210–1215.
- 53. Wetzler, M., Z. Estrov, M. Talpaz, A. Markowitz, J. U. Gutterman, and R. Kurzrock. 1993. Granulocyte-macrophage colony-stimulating factor as a cause of paraneoplastic leukaemoid reaction in advanced transitional cell carcinoma. J. Intern. Med. 234:417–420.
- Perez, F. A., C. L. Fligner, and E. Y. Yu. 2009. Rapid clinical deterioration and leukemoid reaction after treatment of urothelial carcinoma of the bladder: possible effect of granulocyte colony-stimulating factor. J. Clin. Oncol. 27:e215–e217.
- Turalic, H., F. D. Deamant, and J. H. Reese. 2006. Paraneoplastic production of granulocyte colony-stimulating factor in a bladder carcinoma. Scand. J. Urol. Nephrol. 40:429–432.
- Kitayama, S., Y. Fujii, and K. Kihara. 2004. Urothelial cancer producing granulocyte colony-stimulating factor: possible induction of splenomegaly. Urology 63:377–378.
- Ueno, M., S. Ban, T. Ohigashi, T. Nakanoma, S. Nonaka, R. Hirata, et al. 2000. Simultaneous production of granulocyte colony-stimulating factor and parathyroid hormone-related protein in bladder cancer. Int. J. Urol. 7:72–75.
- Sanchez-Carbayo, M., N. D. Socci, J. Lozano, F. Saint, and C. Cordon-Cardo. 2006. Defining molecular profiles of poor outcome in patients with invasive bladder cancer using oligonucleotide microarrays. J. Clin. Oncol. 24:778–789.
- Lee, J. S., S. H. Leem, S. Y. Lee, S. C. Kim, E. S. Park, S. B. Kim, et al. 2010. Expression signature of E2F1 and its associated genes predict superficial to invasive progression of bladder tumors. J. Clin. Oncol. 28:2660–2667.
- 60. Theano, T., S. Pelagia, N. Konstantinos, K. Petros, B. Alfred, D. Konstantinos, et al. 2002. Lymphocyte activation by granulocyte macrophage-colony stimulating factor in human bladder cancer. J. Exp. Ther. Oncol. 2:153–157.
- 61. Wu, Q., R. Mahendran, and K. Esuvaranathan. 2003. Nonviral cytokine gene therapy on an orthotopic bladder cancer model. Clin. Cancer Res. 9:4522–4528.

- Gofrit, O. N., K. C. Zorn, S. Shikanov, and G. D. Steinberg. 2010. Marker lesion experiments in bladder cancer–what have we learned? J. Urol. 183:1678–1684.
- Demirci, U., U. Coskun, B. Sancak, B. Ozturk, B. Bahar, M. Benekli, et al. 2009. Serum granulocyte macrophage-colony stimulating factor: a tumor marker in colorectal carcinoma? Asian Pac. J. Cancer Prev. 10:1021– 1024.
- 64. Urdinguio, R. G., A. F. Fernandez, A. Moncada-Pazos, C. Huidobro, R. M. Rodriguez, C. Ferrero, et al. 2013. Immune-dependent and independent antitumor activity of GM-CSF aberrantly expressed by mouse and human colorectal tumors. Cancer Res. 73:395–405.
- 65. Matsuda, A., K. Sasajima, T. Matsutani, H. Maruyama, M. Miyamoto, T. Yokoyama, et al. 2009. Aggressive undifferentiated colon carcinoma producing granulocyte-colony stimulating factor: report of a case. Surg. Today 39:990–993.
- 66. Fujiwara, Y., O. Yamazaki, S. Takatsuka, R. Kaizaki, and T. Inoue. 2011. Granulocyte colony-stimulating factor-producing ascending colon cancer as indicated by histopathological findings: report of a case. Osaka City Med. J. 57:79–84.
- Davis, J. L., R. T. Ripley, T. L. Frankel, I. Maric, J. N. Lozier, and S. A. Rosenberg. 2010. Paraneoplastic granulocytosis in metastatic melanoma. Melanoma Res. 20:326–329.
- 68. Schniewind, B., M. Christgen, A. Hauschild, R. Kurdow, H. Kalthoff, and H. J. Klomp. 2005. Paraneoplastic leukemoid reaction and rapid progression in a patient with malignant melanoma: establishment of KT293, a novel G-CSF-secreting melanoma cell line. Cancer Biol. Ther. 4:23–27.
- Moon, H. W., T. Y. Kim, B. R. Oh, S. M. Hwang, J. Kwon, J. L. Ku, et al. 2012. Effects of granulocyte-colony stimulating factor and the expression of its receptor on various malignant cells. Korean J. Hematol. 47:219–224.
- 70. Roda, J. M., Y. Wang, L. A. Sumner, G. S. Phillips, C. B. Marsh, and T. D. Eubank. 2012. Stabilization of HIF- 2α induces sVEGFR-1 production from tumor-associated macrophages and decreases tumor growth in a murine melanoma model. J. Immunol. 189:3168–3177.
- Grotz, T. E., L. Kottschade, E. S. Pavey, S. N. Markovic, and J. W. Jakub. 2013. Adjuvant GM-CSF Improves Survival in High-risk Stage IIIC Melanoma: a Single-center Study. Am. J. Clin. Oncol. Epub ahead of print.
- 72. Meyer, C., A. Sevko, M. Ramacher, A. V. Bazhin, C. S. Falk, W. Osen, et al. 2011. Chronic inflammation promotes myeloid-derived suppressor cell activation blocking antitumor immunity in transgenic mouse melanoma model. Proc. Natl Acad. Sci. USA 108: 17111–17116.
- 73. Obermueller, E., S. Vosseler, N. E. Fusenig, and M. M. Mueller. 2004. Cooperative autocrine and paracrine

7801-7812.

- 74. Ma, Y., D. Liang, J. Liu, K. Axcrona, G. Kvalheim, K. E. Giercksky, et al. 2012. Synergistic effect of SCF and G-CSF on stem-like properties in prostate cancer cell lines. Tumour Biol. 33:967–978.
- 75. Dai, J., Y. Lu, C. Yu, J. M. Keller, A. Mizokami, J. Zhang, et al. 2010. Reversal of chemotherapy-induced leukopenia using granulocyte macrophage colony-stimulating factor promotes bone metastasis that

can be blocked with osteoclast inhibitors. Cancer Res. 70:5014-5023.

- 76. Park, B. K., H. Zhang, Q. Zeng, J. Dai, E. T. Keller, T. Giordano, et al. 2007. NF-kappaB in breast cancer cells promotes osteolytic bone metastasis by inducing osteoclastogenesis via GM-CSF. Nat. Med. 13:62–69.
- 77. Altundag, K., O. Altundag, E. T. Elkiran, M. Cengiz, and Y. Ozisik. 2004. Addition of granulocyte-colony stimulating factor (G-CSF) to adjuvant treatment may increase survival in patients with operable breast cancer: interaction of G-CSF with dormant micrometastatic breast cancer cells. Med. Hypotheses 63:56–58.