

# **Is There a Connection between the Metabolism of Copper, Sulfur, and Molybdenum in Alzheimer's Disease? New Insights on Disease Etiology**

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Abstract: Alzheimer's disease (AD) and other forms of dementia was ranked 3rd in both the Americas and Europe in 2019 in a World Health Organization (WHO) publication listing the leading causes of death and disability worldwide. Copper (Cu) imbalance has been reported in AD and increasing evidence suggests metal imbalance, including molybdenum (Mo), as a potential link with AD occurrence.We conducted an extensive literature review of the last 60 years of research on AD and its relationship with Cu, sulfur (S), and Mo at out of range levels.Weanalyzed the interactions among metallic elements' metabolisms;Cu and Mo are biological antagonists, Mo is a sulfite oxidase and xanthine oxidase co-factor, and their low activities impair S metabolism and reduce uric acid, respectively. We found significant evidence in the literature of a new potential mechanism linking Cu imbalance to Mo and S abnormalities in AD etiology: under certain circumstances, the accumulation of Cu not bound to ceruloplasmin might affect the transport of Mo outside the blood vessels, causing a mild Mo deficiency that might lowerthe activity of Mo and S enzymes essential for neuronal activity. The current review provides an updated discussion of the plausible mechanisms combining Cu, S, and Mo alterations in AD.

Keywords: Alzheimer's disease; metal imbalance; molybdenum; copper; sulfur

### 1. Introduction

Alzheimer's disease (AD) is the most common form of dementia globally in older adults [1]. It is highly inheritable, and diet and lifestyle are influential for increased risk [2]. Histopathologically, AD is characterized by amyloid  $\beta$  plaques (A $\beta$ ) and tau neurofibrillary tangles in the brain [3]. The world scientific community has been studying AD for decades, with some research lines investigating copper (Cu) and sulfur (S) metabolisms and, more recently, molybdenum (Mo) metabolisms.

Cu is an essential element for the functioning of enzymes linked to oxidative stress scavenging. The alteration of its homeostasis is associated with free radical production and cellular damage [4]. In the body, S is found in mineral form, sulfate  $(SO_4^{2-})$  being



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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). also present in organic molecules such as vitamins, coenzymes, and S-amino acids [5]. Mo-containing enzymes are found in human and animal tissues, and the redox potential of MoV/MoVI is suitable in redox chemistry involving electron exchange with flavin mononucleotides [6].

Many years ago, there were reports that the altered metabolism of S amino acids occurs in AD [7]. The relationship between Cu imbalance and AD is already well reported in the scientific literature [8]. Furthermore, there are many similarities between some metabolic consequences of Mo deficiency [9,10] and AD.

#### 2. Linking AD Predisposing Factors to Cu and S Metabolism

Pathologies or symptoms related to AD usually occur before and/or concomitantly with the onset of symptoms, such as memory loss. Multiple factors are associated with AD, including hypertension, diabetes, hypercholesterolemia, obesity, and depression [1], as well as sex hormones [11] and hyperhomocysteinemia [12], all of them linked in some way to Cu and S disturbances.

Consistently, people with low performance in the Digit Symbol Substitution cognitive test tended to have a higher prevalence of hypertension and diabetes, among other diseases, than people with average cognitive performance. It is of particular interest that cognitive performance in the same test decreased with an increasing total Cu intake, indicating that AD-linked risk factors could be correlated to altered Cu levels [13].

Cu can influence blood pressure regulation via tyrosinase activity, a key enzyme in synthesizing norepinephrine, a major vasoconstrictor factor [14]. Therefore, hypertension can be affected by changes in Cu concentration [14]. Additionally, a multinomial logistic model revealed that an increase of non-Ceruloplasmin Cu (nCp–Cu)in one-unit standard deviation raised the relative risk of having Type 2 Diabetes (T2D) by 9.64 [15]. Moreover, serum levels of  $Cu^{2+}$  are significantly higher in rat models of depression [16] and significant depression patients [17] compared to healthy controls [18]. Interestingly, Cu exposure exacerbates the depression-like behavior of ApoE4 mice. The mechanisms may involve the dysregulation of synaptic function and immune response and the over-activation of neuroinflammation [19].

High Cu levels have been shown influence negatively testosterone synthesis and body levels [20,21]. Testosterone levels are significantly lower in AD brains than in control ones [22]. Brain testosterone levels are also considerably reduced in men with mild neurological changes (MNC) consistent with early-stage AD. Rosario et al. [22] evaluated brain tissue at autopsy of three groups: control subjects, subjects with MNC, and subjects with AD, all with similar ages (mean (SD) ages of 70.9 (5.3), 72.9 (5.7), and 72.0 (6.5) years old, respectively). They found 1.0, 0.6 and 0.5 ng g<sup>-1</sup> of wet tissue of testosterone, respectively. In comparison with the testosterone curve, urinary Mo and serum Cu were inversely associated in a data survey, including men aged 18–55 years from NHANES 2011–2012 [23].

Changes in S compound metabolism seem implicate on the AD risk factors depression and hyperhomocysteinemia. An unhealthy dietary pattern has been associated with an increased risk of depression via decreased serum folate contents [24]. Folate is a cofactor required for S-containing amino acid metabolism, particularly important forproducing methionine (Met) from Homocysteine (Hcy), avoiding hyperhomocysteinemia. The high levels of Hcy are involved in the decreased synthesis of catecholamines and depression [24] and predisposition to AD [25]. Namekawa et al. [26] evaluated depression in the elderly and found an increased risk of AD and A $\beta$  protein metabolism.

The accumulated knowledge in the last decades has provided significant advances in understanding AD. Furthermore, the relationship of AD with Cu and S metabolism and other AD-associated diseases has had much advancement but still does not allow for a clear explanation of AD's causes. Squitti et al. [8], in a recent review, commented in-depth about the Cu imbalance in AD and its link with the A $\beta$  hypothesis. However, they did not mention S metabolism.

#### 3. Linking Cu to AD Pathology or Multiple AD-Associated Factors

Total body Cu in adult humans has been estimated to range from 70 to 150 mg, with the highest contents in the liver, brain, heart, and kidneys [27]. Cu is a nutrient for humans, which participates in the composition of essential proteins: ceruloplasmin, Cu–Zn superoxide dismutase, nitric reductase, amine oxidase, tyrosinase, bilirubin oxidase, Cytochrome c oxidase (COX), and plastocyanin, for example [28]. The main route of excretion of Cu from the body is bile [29]. However, it can lead to oxidative stress and other harmful effects when in excess.Healthy total Cu serum ranges are 700  $\mu$ g L<sup>-1</sup>–1300  $\mu$ g L<sup>-1</sup> [30].

The daily nutritional recommendations and maximum tolerable levels of Cu intake for people above 19 years of age are the same regardless of age [27,31]. More studies should be conducted to determine the Cu amount needed and the maximum limits by age groups over 19 years old. It is possible that at older ages Cu intake limits should be lower than inyounger generations because Cu can accumulate in the body with aging [32–34]. However, more than the Cu accumulation, it is the dislocation of loosely bound Cu in the brain that can cause toxicity [35,36]. In addition, some foods may increase Cu content due to the use of Cu to control plant diseases in agriculture [37].

Masaldana et al. [38] commented that cell senescence is characterized by irreversible growth arrest caused by replicative exhaustion or pro-oncogenic cell stressors (radioactivity, oxidative stress, oncogenic activation). Additionally, the same authors reported that enrichment with metals, including Cu, in the senescent cells from tissues during aging had been associated with tissue dyshomeostasis and age-related pathologies, including cancer and neurodegenerative disorders (AD, Parkinson's disease (PD)), and metabolic disorders (e.g., diabetes). Interestingly, Masaldana et al. [38] identified Cu accumulation as a universal feature of senescent cells (fibroblasts from embryonic mice, human prostate epithelial cells, and human diploid fibroblasts) in vitro. In fact, in vitro experiments showed the exposure of fibroblasts to Cu sulfate, which increases intracellular Cu, induces premature senescence through oxidative stress [39]. This fact indicates a specific association between the accumulation of Cu and the anomalies mentioned. On the other hand, a major free-radical chain-breaking antioxidant, vitamin E ( $\alpha$ -tocopherol), may interfere in the initiation and progression of Cu-induced oxidative damage [40].

Chronic exposure to Cu and its dyshomeostasis has been linked to accelerated cognitive decline and potentially increased risk of AD [41–43]. Furthermore, Cu ions, due to their redox ability, have been considered to be potential therapeutic targets in AD, and a considerable number of ligands have been developed to modulate the toxicity associated with Cu in this context via disruption of the A $\beta$ –Cu interaction [44]. Heng-Wei et al. [45] reported that recent evidence suggests that dyshomeostasis of Cu and its valency in the body are major determinants of its beneficial effects as an essential metal or its neurotoxic counterpart. Following this line, James et al. [46] appraised redox-competent Cu species (Cu<sup>2+</sup> and Cu<sup>1+</sup>) levels in AD. They quantified the burden of redox-active labile Cu in postmortem brain tissue of AD subjects and age-matched cognitively normal controls. This study found high contents of exchangeable Cu<sup>2+</sup> and increased capacity of AD cortical tissue samples to bind Cu<sup>2+</sup>.

In healthy people, 90–95% of the Cu circulation is bound to the Cu-binding protein ceruloplasmin [47]. NCp–Cu characterizes AD patients, and possible genetic defects of the ATP7B gene, which encodes for a P-type ATPase protein involved in the Cu transport, were identified in AD patients [48]. In AD, Cu decreases in the brain but increases in the serum due to the nCp–Cu component increase [36,49]. Excess Cu speeds up some early AD events, such as the induction of aggregation of A $\beta$  peptides via enhancing the amyloidogenic cleavage of amyloid precursor protein (APP) and the hyperphosphorylation of Tau by stimulating GSK3 $\beta$  kinase, disturbing brain function of wild-type mice and exacerbating neurodegenerative changes in a mouse model of AD [50]. Therefore, studies have recently tested chelators of Cu as therapeutic approaches in the AD animal models or have even used Cu to induce AD animal models [51].

### 4. Linking S Metabolism to AD Pathology or Multiple AD-Associated Factors

In humans, S inorganic species, such as  $SO_4^{2-}$ , and S-biomolecules, including Scontaining amino acids, their derivatives, S-vitamins, biotin or S-coenzymes, and co-factors, as coenzyme A and  $\alpha$ -lipoid acid, are obtained from a regular and equilibrated diet [5]. Met is an essential S amino acid that functions as a precursor for the synthesis of Hcy and cysteine (Cys).

Varadarajan et al. [52] commented that  $A\beta$  is a source of oxidative stress from free radicals that can lead to neurodegeneration in AD. It may be partly due to the oxidative stress associated with free radicals from  $A\beta$ , which involves Met. Met-SO (methioninesulfoxide), an oxidized form of Met, is an indicator of systemic oxidative stress. Weng et al. [53] found serum levels of Met-SO, significantly higher in early-stage AD patients than in mild cognitive impairment (MCI) patients.

In AD, there is in plasma an increase in the Cys content [7] and a decrease in  $SO_4^{2-}$  [7], folate [54], and choline contents [55]. Moss and Waring [56] evaluated that plasma Cys/ $SO_4^{2-}$  ratio is a possible clinical biomarker for investigating chronic health disorders including AD, PD, multiple sclerosis, myalgic encephalomyelitis, irritable bowel syndrome, autism, depression, motor neuron disease, and hemolytic anemia. It is interesting to note that although Cys contents are high in the plasma in AD [7], there is no consensus yet about its level inside the brain cells. While Luo et al. [57] found decreased Cys by ca. 3-fold in an animal model of AD, Mandal et al. [58] found higher ones using another detection method.

Individuals with low folate contents and high Hcy contents are at increased risk of AD [54,59–62]. Increased Hcy accompanies folate deprivation because folate is a necessary co-factor for the enzyme 5,10-methylenetetrahydrofolate reductase that mediates the conversion of Hcy to Met [63].

Irizarry et al. [64] investigated a link between high Hcy and AD. They found that elevated plasma total Hcy was positively correlated with plasma A $\beta$  40 and A $\beta$  42 levels. Additionally, prolonged hyperhomocysteinemia can lead to the onset of cardiovascular disease, atherosclerosis, stroke, inflammatory syndromes like osteoporosis and rheumatism, and neuronal pathologies, including AD and PD [65].

The relationship between folate and choline deficiencies with AD has been extensively studied [66–69]. Choline deficiency, for example, besides causing lower acetylcholine synthesis, contributes to mitochondrial dysfunction and, therefore, to the rate of fatty acid oxidation deficiencies and has also been linked to increased fatty acid uptake [69].

### 5. Hypothetical Relationship between Mo and AD Pathology or Multiple AD-Associated Factors

Mo is considered essential for animals and plants. It is a co-factor of enzymes (molybdoenzymes) xanthine oxidase-dehydrogenase (XO), aldehyde oxidase (AO), sulfite oxidase (SUOX), and mitochondrial amidoxime reducing component (mARC) [70], also being a reversible activator of adenylyl cyclase [71–73] in animals. Additionally, Mo is a cofactor of nitrate reductase in all plants and nitrogenase in Fabaceae plants associated with bacteria-fixing atmospheric nitrogen [74,75]. These enzymes possess a unique co-factor (Mo co-factor—Moco) that contains a novel ligand in bioinorganic chemistry, the pyranopterin ene-1,2-dithiolate [76]. Moco comprises Mo covalently bound to two S atoms of a unique tricyclic pterin moiety called molybdopterin [77] (Figure 1). It is still noteworthy that S required for Moco biosynthesis is obtained from Cys.

For humans, Mo is required in small amounts; however, it is worth considering that studies that assess daily nutrient requirements are performed onhealthy individuals and do not take aging into account. The tolerable upper intake level for the United States and Canada was set at 2 mg day<sup>-1</sup> [31]. The European Commission suggested an upper limit of 0.6 mg day<sup>-1</sup> [78]. Healthy Mo serum ranges are 0.28  $\mu$ g L<sup>-1</sup>–1.17  $\mu$ g L<sup>-1</sup> [79]. Giussani et al. [80] verified that Mo is rapidly and efficiently absorbed into the systemic circulation. When it is administered dissolved in black tea and composite meals, the absorption rate is lower and delayed than dissolved in water.



Figure 1. Mo co-factor—Moco.

The Mo deficiency results in high levels of sulfite and xanthine and low levels of  $SO_4^{2-}$  and uric acid in the blood and urine due to impaired SUOX and XO activity, respectively [81]. XO is an enzyme that generates reactive oxygen species, such as superoxide and peroxide, by reacting with  $O_2$  [82]. This enzyme catalyzes the oxidation of hypoxanthine to xanthine and can further catalyze the oxidation of xanthine to uric acid. Moreover, XO plays an essential role in purine catabolism in some species, including humans, impacting adenosine and guanosine production [83,84]. Hereditary xanthinuria (type I) is caused by an inherited XO deficiency characterized by low blood and urine uric acid and high urinary xanthine contents [85].

Alonso-Andrés et al. [86] found significantly decreased adenosine, guanosine, hypoxanthine, and xanthine contents in the frontal cortex from the early stages of AD pathology. Additionally, Du et al. [87] showed using meta-analysis that those patients with AD had lower uric acid contents than healthy controls and verified that high serum uric acid contents were significantly associated with decreased risk of AD. The authors concluded an inverse association between serum uric acid contents and AD risk. This allows us to hypothesize that a lower XO activity, maybe linked to Mo deficiency, correlates to higher AD risk. Interestingly, Tana et al. [88] reviewed the related literature and concluded that serum uric acid might modulate cognitive function. Then, current studies indeed demonstrate that uric acid may exert neuroprotective actions in AD. In addition, Latourte et al. [89] commented that patients with gout (a form of inflammatory arthritis characterized by persistently elevated contents of uric acid in the blood) may have decreased risk for AD.

The reaction catalyzed by SUOX is the terminal step in the excretory metabolism of S-containing amino acids such as Met and Cys, by which sulfite produced from Met and Cys metabolism is detoxified [90]. Additionally, via the H<sub>2</sub>S pathways, obtained during metabolism of Hcy in Cys, significant quantities of sulfite are formed that require oxidation by SUOX [91]. In the nervous system, H<sub>2</sub>S acts as a neuromodulator influencing synaptic activity under regulation of steroid hormones and neurotransmitters; it may be involved in associative learning [92]. An interesting occurrence is that H<sub>2</sub>S in the brain is severely reduced in AD [93]. These authors commented that the reduction of H<sub>2</sub>S may be involved in some aspects of the cognitive decline associated with AD.

SUOX is the key enzyme in SO<sub>4</sub><sup>2-</sup> production. Due to low SUOX activity, biochemical abnormalities include high urinary sulfite, thiosulfate, and S-sulfocysteine excretion, which are abnormal metabolic products of degradation of S amino acids and low SO<sub>4</sub><sup>2-</sup> content [9,10]. SUOX is located in the mitochondrial intermembrane space [74]. Cytochrome c is the electron acceptor for the SUOX [94], with oxygen and potassium ferrocyanide as alternative oxidizing substrates. Bosetti et al. [95] studied COX of isolated mitochondria from platelets and postmortem motor cortex and hippocampus from AD patients and age-matched control subjects. Compared with controls, COX activity from AD was decreased significantly in platelets (-30%, p < 0.01, n = 20) and the hippocampus (-35 to -40%, p < 0.05, n = 6). Similarly, Maurer et al. [96] also worked with postmortem brain tissues and found a selective defect of COX in the AD brain tissue versus the normal human brain. In general, the low SUOX activity can be due to problems in the enzyme synthesis or problems in the Moco synthesis [97]. The lowest synthesis of the enzymes or Moco occurs in newborns or infants, and these are acute health problems that can lead to death [98]. However, it alsoappears as "late-onset" or "mild" [99]. A third explanation for low SUOX activity may be a Mo deficiency.

Every unbalanced chemistry candidate contributor to AD, such as lower Mo contents, is caused by other metabolic, physiological, or dietary imbalances. Given that Mo deficiencies are very rare, there is a need for further studies prior to concluding that they are the cause of AD. The only link between Mo levels and AD that observed is that SUOX, a Mo-containing protein, could be dysregulated in AD. High levels of sulfites due to low levels of SUOX could dysregulate glutaminergic neurotransmission, contributing with one of the main aspects of AD [100].

We suggested relationships between the actions of Mo as a co-factor or enzyme stimulator and metabolites to which it is related and AD. There are many similarities between Mo deficiency and AD metabolic symptoms (Table 1).

In addition, Xia and Storm [101] reported that adenylyl cyclases, type 1 and type 8, are essential during memory formation. It is important to emphasize that AD patients/models demonstrate reductions in adenylyl cyclase activity [102,103]. Nonaka et al. [104] recently reviewed the interactions between adenylyl cyclase and dementia.

**Table 1.** Similarities among lower activity or synthesis of XO, AO, SUOX, and Moco or Mo deficiency and symptoms of Alzheimer's disease.

	Lower Activity or Synthesis of XO, AO, SUOX or Moco, or Mo Deficiency		Alzheimer's Disease
	Similarities		
Sulfate	Low urinary $SO_4^{2-}$ content [9,10].	1. 2.	High Cys/SO <sub>4</sub> <sup>2-</sup> in blood plasma [7]; AD's plasma had only 22% SO <sub>4</sub> <sup>2-</sup> than standard control [7].
Methionine	High plasma Met [105].	1. 2.	Alterations in pathways related to Met metabolism [106]; Serum levels of Met-SO are higher in early-stage AD as compared with MCI patients [53].
Cysteine	Sulfite loading stops the conversion of Met to Cys, and this causes Cys levels to fall [105].	1. 2.	Cys content decreased inside brain cells [60]; Cys is high in brain AD [58].
Uric acid	Serum and urinary uric acid contents are reduced [107].	1. 2.	Lower contents of uric acid in AD patients than healthy controls [87]; Patients with gout with elevated uric acid contents in the blood may have decreased risk for AD [89].

The hypothesis regarding a link between Mo and A $\beta$  is indirect, because Mo is not directly involved in the formation of A $\beta$  in the same way as Cu. However, Kong et al. [108] found that, similar to MoS<sub>2</sub> nanoparticles, MoS<sub>2</sub> nanosheets possessed therapeutic effects on AD by inhibiting A $\beta$  aggregations and degrading the previously formed A $\beta$  fibrils. Similarly, Manna et al. [109] found the disappearance of fibrillar structures and recovery from neurodegenerative disorders in molybdate-treated A $\beta$  42-mutant Drosophila flies as compared to the untreated ones, corroborating the therapeutic ability of (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub> for treating AD. Thus, it is possible that, in normal conditions, Mo may have a destabilizing effect on A $\beta$  fibril formation and that its deficiency may impair this natural process, facilitating A $\beta$  formation.

### 6. Linking Metabolic Interaction of Cu, Mo, and S to AD Pathology or Multiple AD-Associated Factors

Many researchers have studied the interaction of Cu–Mo–S in non-ruminant nutrition. The majority of the studies are about dietary Mo excess causing depletion in Cu contents in rats, swine, etc. [110,111]. Conversely, studies demonstrated that excess Cu causes Mo deficiency. In rats with excess Cu supply, a non-absorbable complex of Cu and Mo in the gastrointestinal tract might reduce Mo absorption [112]. Suttle [113] commented that it is possible that at low Mo concentrations, the primary site of Cu–Mo interaction is in the gut, resulting in a decreased uptake of Cu. In contrast, at high Mo contents, high levels of Mo in the bloodstream and tissues favor the formation of inorganic and organic Cu–Mo complexes, which accumulate in the tissues.

Experimental studies indicate that Cu from the Cu–Mo complex is unavailable for ceruloplasmin synthesis. Dowdy and Matrone [114] showed a lower level of ceruloplasmin activity in the serum of pigs receiving the Cu–Mo complex (in a value very close to those not receiving Cu) than with Cu sulfate-fed controls. Dowdy et al. [115] have suggested that this Cu–Mo complex might be the mineral lindgrenite (2CuMo0<sub>4</sub>. Cu(OH)<sub>2</sub>). Arthur [116] reported lower Mo content in the liver of Guinea Pigs supplemented with Cu. They found  $3.4 \pm 0.4$  and  $6.2 \pm 1.0$  mg kg<sup>-1</sup> of Mo in the liver of animals fed with 10 mg kg<sup>-1</sup> or no Cu supplemented diet, respectively.

In vitro studies indicated a competition between high levels of Cu and Mo during Moco formation [97,117]. Inhibition of Moco synthesis by Cu could be explained by inhibiting the Mg-dependent Mo insertion reaction [97]. Hadizadeh et al. [117] found stimulation and inhibition of XO activity by Cu<sup>2+</sup>. Less than 5  $\mu$ M Cu<sup>2+</sup> resulted in stimulating the XO enzymatic activity, while in a concentration-dependent manner, 5–700  $\mu$ M, 700 to 2000  $\mu$ M, and 2000  $\mu$ M inhibited moderate, drastic, and ultimately, respectively. Mendel and Kruse [118] commented that Moco deficiency could not be seen as an isolated defect but in a context with cellular Cu metabolism since increased cellular Cu contents might be causal for a decreased rate of Moco synthesis [97].

Suzuki et al. [119] investigated Cu–S metabolism interaction. They concluded that nCp–Cu ions in the bloodstream bind primarily to the significant form of albumin to form the mercaptalbumin-Cu complex. The latter sequesters Cys to form the albumin–Cu–Cys complex. Only Cu in the albumin–Cu–Cys complex must be transferred to the liver with albumin remaining in the blood serum as an albumin–Cys complex.

A high Cu/Mo ratio is present in children with Wilson disease. Mahoney et al. [120] demonstrated decreased uric acid production, suggesting reduced XO activity and possible Mo deficiency. The deleterious effect of high ratio Cu/Mo with impact on low XO activity has been commented on in human nutrition since many years ago [121]. Seelig [121] highlighted excess Cu levels in blood and liver associated with decreased Mo levels and/or low XO activity.

Bar-Or et al. [122] demonstrated the relationship between Cu and S metabolism has a direct effect in AD. They show that high nCp–Cu causes Hcy accumulation due to a decrease in Cystathionine b-synthase (CBS) activity: 10  $\mu$ M and 25  $\mu$ M Cu decreased CBS activity by 50% and 70%, respectively. Additionally, low CBS activity results in low Cys synthesis and consequently low glutathione (GSH) synthesis, since Cys is the amino acid rate-limiting in its production. GSH concentration is reduced in MCI and AD in brain regions (hippocampi and frontal cortices) affected by AD pathology [58]. Another effect is a low synthesis of H<sub>2</sub>S endogenously produced in the brain from Cys by CBS. H<sub>2</sub>S in the brain is severely reduced in AD, and, as mentioned, its reduction may be involved in cognitive decline because it functions as a neuromodulator [93].

Table 2 summarizes possible links between AD and Mo deficiency or/and Cu–Mo–S altered metabolism.

Event	Research Results
NCp–Cu accumulation in serum or plasma.	<ol> <li>Chronic exposure to Cu and its dyshomeostasis has been linked to accelerated cognitive decline and potentially increased the risk of AD [41–43];</li> <li>High nCp–Cu in the serum characterizes AD patients [36];</li> <li>Serum Cu was inversely associated with testosterone [23] and Rosario et al. [22] found that brain levels of testosterone are significantly lower in AD.</li> </ol>
Cu and Mo are antagonists, so excessive Cu cause Mo deficiency and vice versa.	<ol> <li>The formation of a Cu–Mo complex can be the place of nutritional interaction between the two metal ions [116];</li> <li>Cu–Mo complex was unavailable for ceruloplasmin synthesis [117];</li> <li>Excess in Cu supply and a non-absorbable complex of Cu and Mo in the gastrointestinal tract might reduce Mo absorption [114];</li> <li>Reducing uptake or retention of Mo in Cu-deficient induces Cu-excess in rats [114];</li> <li>Increased cellular Cu contents might be causal for a decreased rate of Moco synthesis [97];</li> <li>Competition between Cu, at a high level, and Mo occurs during Moco formation [119];</li> <li>Cu/Mo ratio imbalance affects S metabolism [123].</li> </ol>
Products of enzymes' activity that have Mo as a co-factor are in lower content in AD than in healthy people	<ol> <li>A high Cu/Mo ratio affects xanthine oxidase activity [121];</li> <li>Decreased contents of adenosine, guanosine, hypoxanthine, and xanthine in early AD frontal cortex [86];</li> <li>High serum uric acid contents were significantly associated with decreased risk of AD, so patients with gout may have reduced risk for AD [86,89];</li> <li>AD patients/models demonstrate reductions in adenylyl cyclase activity [102,103].</li> </ol>
Altered S amino acids metabolism occurs in AD	<ol> <li>Mo is the sulfite oxidase (SUOX) co-factor: its low activity damages S metabolism [9,10];</li> <li>AD has a high Cys/SO<sub>4</sub><sup>2-</sup> ratio in plasma [7].;</li> <li>In AD, there is a plasma decrease of SO<sub>4</sub><sup>2-</sup> level [7] (low SO<sub>4</sub><sup>2-</sup> content indicates low SUOX activity due to possible Mo or Moco deficiency);</li> <li>Cys levels are high in AD plasma [7];</li> <li>Cys decreased by ca. 3-fold in the brain of a rat model of AD [57];</li> <li>Cytochrome c is the electron acceptor for the SUOX. COX activity from AD was decreased significantly in platelets and the hippocampus [95];</li> <li>High nCp-Cu causes an accumulation of Hcy due to a decrease in Cystathionine b-synthase activity [122].</li> </ol>
Mo contents	<ol> <li>AD: serum Mo level increased progressively, passing from healthy subjects (HS) through subjective memory complaint, mild cognitive impairment up to AD [124];</li> <li>Diabetic (T2D) with severe complications: serum Mo contents increased [30];</li> <li>T2D with severe complications: lower urine Mo contents [30].</li> </ol>

**Table 2.** Possible links between Alzheimer's Disease (AD) and Mo deficiency or/and Cu–Mo–S altered metabolism.

## 7. Cu and Mo Increase in the Blood Vessels and Evidence Sustaining a Decreased Cu and Mo Transport in the Brain with Aging

Paglia et al. [124] evaluated four groups of 118 people aged between 54 and 87 years old (HS, healthy subjects—n = 34; SMC, subjective memory complaint—n = 20; MCI, mild cognitive impairment—n = 24; AD, Alzheimer disease—n = 40). Mo content increased in serum progressively, passing from HS through SMC, MCI, up to AD, and the difference between HS and AD was statistically significant (0.83 ± 0.26; 0.99 ± 0.24; 1.09 ± 0.36; and 1.20 ± 0.52 µg L<sup>-1</sup> of Mo in HS, SMC, MCI, and AD, respectively). Additionally, serum Cu concentration was higher in SMC than in HS (p > 0.05) 858.96 ± 224.19 and 703.88 ± 244.03 µg L<sup>-1</sup>, respectively.

The results of Mo content [124] indicate that there is a progressive Mo accumulation in serum throughout the worsening phases of AD symptoms. On the other hand, Cu increase in serum is likely associated with the increase of the nCp–Cu component [43], and nCp–Cu increases the susceptibility to AD [125,126]. Additionally, Cu decreases in the brain in AD [36] and, possibly, Mo content decreases in tissues in AD, as the decreased levels of molybdoenzymes products suggest [7,87].

There are some similarities between Cu and Mo metabolism in AD, so their serum and tissues contents have a similar pattern, as shown in Table 3.

	Serum Levels	Brain Tissue Levels	
	Similarities		
Cu	High content of nCp–Cu in serum [36].	In brain tissues, the Cu content is usually low [36,68].	
Мо	Serum Mo content increased progressively, passing from healthy subjects (HS) through subjective memory complaint, mild cognitive impairment up to AD, and the difference between HS and AD was statistically significant [124].	<ol> <li>Evidence suggesting a low level of Mo in AD brain tissues:</li> <li>High serum uric acid contents were significantly associated with decreased risk of AD, so patients with gout may have decreased risk for AD [87,89];</li> <li>Brain tissues from AD patients/models demonstrate reductions in adenylyl cyclase activity [102,103].</li> </ol>	

Table 3. Similarities between Cu and Mo contents in Alzheimer's disease (AD).

Based on the evidence they reported, they suggested a higher Mo level in serum [124] and a lower one in urine [30] and brain. Our hypothesis posits that there is little Cu-Mo complex formation inside blood vessels when there is an adequate nCp–Cu /Mo ratio in youth. Consequently, Cu and Mo are efficiently transported out of the vessels, reaching the tissue cells as depicted by Smith and Wright [127] for sheep with Cu–Mo balanced diet. With aging, people with a genetic predisposition for AD may have progressive nCp–Cu accumulation in the body that can likely increase the formation of Cu-Mo complexes inside the blood vessels. Under these conditions, the transport of Mo out of the vessels might be reduced, causing a mild Mo deficiency within the tissues. We posit that at older ages, in AD, the accumulation of nCp–Cu is increased and, consequently, the formation of the Cu–Mo complex is significantly increased inside the vessels, leading to an increase in the serum Mo content and a great decrease in the Mo transport out of the vessels, a mechanism observed by Smith and Wright [127]. This leads to an increase in the Mo deficiency symptoms, with a significant drop in the activity of enzymes that contain Mo as a co-factor, as reported by Alonso-Andrés et al. [86], Du et al. [87], Latourte et al. [89], Yamamoto et al. [102], and Kelly [103].

We propose that AD-susceptible people may have a higher nCp–Cu/Mo ratio in serum resulting in a lesser Cu/Mo ratio in food in adult age. In this case, Mo should possibly be supplemented in the daily diet. We propose future research reformulation in the daily human requirements for Cu and Mo, including the Cu/Mo ratio for different ages. For example, for ruminants, the ideal Cu/Mo ratio is >6:1 and <10:1 in the finished feed [128]. Thus, in this group of animals, if the Cu/Mo ratio is <6:1 or >10:1 in feed, it causes metabolic problems. Moreover, our discussion focuses on nCp–Cu/Mo ratio in AD blood (serum or plasma): in AD, serum nCp–Cu is higher than in healthy people [15]. The nCp–Cu/Mo ratio may be higher in serum than in healthy subjects in AD. Thus, Mo might be progressively captured by nCp–Cu and accumulated in serum, decreasing the Mo bioavailability for the Mo enzymes/proteins.

### 8. Mo Deficiency and Low SUOX and Cytochrome c Activities Are Possibly Linked to AD

As previously mentioned, in individuals with a predisposition towards AD the content of nCp–Cu in serum increases due to aging [36], which possibly causes greater Mo demands or its deficiency. One strong indication for this is the low content of molybdoenzyme

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products in AD [7,56,86–89]. In addition, there is an interaction of deficient Mo with disturbed S metabolism in AD [57,105].

Under Mo deficiency, the SUOX activity decreases, which leads to an increase in sulfite contents (which is toxic) [74] and endogenous S metabolism disturbance [7,54,106] with decreased contents of intracellular Cys [57] and decreases contents of  $SO_4^{-2}$ , folate, choline, and  $H_2S$ .

The terminal step in the excretory metabolism of Met and Cys needs SUOX. If SOUX does not work well, toxic sulfite increases, and the pathway for Cys synthesis decreases. [105]. Accordingly, lower intracellular Cys confirm this idea in AD brain [57]. As Cys is required in CoA [129] and choline synthesis, the acetylcholine production is also lower, which is consistent with disturbed cholinergic neurotransmission seen in AD. Another disturbance is the excess of Met, which can increase A $\beta$  neurotoxicity [52]. High Hcy contents in AD result from the low activity of the trans-sulphuration pathway [130]. When in low SUOX activity, sulfite loading blocks the standard conversion of Met to Cys, and this causes Cys values to fall, possibly due to accumulated sulfite [105]. We commented that Cys contents are high in the plasma in AD [7], but it may be decreased inside the brain cells [57].

In our view, the possible primary link between S and Mo metabolic interaction in AD is the low SUOX activity. It is well documented that low SUOX activity causes neurological damage [74]. Anything that negatively affects the activity of SUOX will also affect the activity of Cytochrome c. That is because sulfite induces Cytochrome c release from brain mitochondria in the presence of  $Ca^{2+}$  [131]. In addition, given that Cytochrome c and SUOX are linked, it makes sense to think that what negatively affects Cytochrome c activity will also negatively affect SUOX activity. Thus, the SUOX–Cytochrome c complex has to be working well for neurological functions to occur satisfactorily. In addition, in AD, there is a decrease in SO<sub>4</sub><sup>2–</sup> content [7], indicating low SUOX activity due to possible Mo deficiency.

We must consider that newborns' acute low SUOX activity results in low plasma Hcy contents [132]. On the other hand, increased plasma Hcy contents are a risk factor for AD, and a typical AD symptom is high plasma Hcy contents [54,65,68]. The high Hcy contents in AD may be due to the direct high nCp–Cu effect demonstrated by Bar-Or et al. [124] when they showed that high nCp–Cu causes Hcy accumulation due to a decrease in CBS activity.

In conclusion, we hypothesize that the excessive nCp–Cu in the body can result in a higher requirement for Mo, which becomes deficient. It results in low SUOX activity, disrupted Met metabolism and higher A $\beta$  content. Consequently, the excess of Met can increase A $\beta$  neurotoxicity [52]. So, although Mo is not directly involved in the formation of A $\beta$ , such as Cu, the link with the A $\beta$  hypothesis is indirect (Figure 2). Its harmful effects on metabolism trigger the other symptoms of AD-related diseases and, later, AD symptoms. That hypothesis is in line with the findings of the current review.



**Figure 2.** Summary of proposed cellular/molecular mechanisms for the Cu–Mo–S circuitry in AD pathology or multiple AD-associated factors.

### 9. Clinical Perspective for AD and the "Cu-Mo-S Circuitry"

Despite considerable social and economic interest in AD, which make this disease one of the most studied in neuroscience, developing an efficient therapeutic protocol is very difficult. Basic research has improved the knowledge about physiopathology and metabolic pathways involved in AD. However, this knowledge was still not able to reach an effective therapy. It has been proposed as a possible strategy for a therapeutic approach to developing disease-modifying therapies (DMT), which aim to promote a significant change in the course of AD, preventing, delaying its onset, or slowing its progression [133]. However, Liu et al. [134] confirmed the effectiveness of eye-tracking in diagnosing a cognitive disorder and concluded that it is a viable marker for recognizing that.

Up to 2021, there were very few drugs approved for AD, and all of them act only in ameliorating the symptoms. None was developed to treat the causes of AD. This is mainly because the real cause for this disease is far from being ultimately discovered. The high failure rate in clinical trials clearly demonstrates this. From 2002 to 2012, the failure rate was 99.6% [133,135].

However, the FDA approved a new treatment in 2021. Aducanumab is an amyloidtargeting monoclonal antibody applied by intravenous infusions and acts on the extracellular accumulation of A $\beta$  protein [136]. Although it might be considered the first approved disease-modifying drug for AD, its approval is highly controversial. Several researchers and scientific societies in the fieldhave stated that there is little evidence of its absolute safety and efficacy and that it has a high financial cost [137–139]. Therefore, despite the effort to find an effective treatment for AD, this has not been accomplished to date. Regarding the main point of this review, few clinical trials involving the "Cu–Mo–S circuitry" were performed. Cu chelating agents were tested, but no satisfactory results were obtained [140].

Given that the search for a pharmacologic treatment has been ineffective, new proposals have been made. Dietary supplementation has been recently proposed to treat AD, specifically those related to the "Cu–Mo–S circuitry", exploring the known metabolic pathways involved in AD [100]. This proposal agrees with our hypotheses of the connection between Mo and AD. It might be expected that a multifactorial disease should need a multifactorial treatment, including drugs and dietary supplementation.

Another critical point to be considered is the fact that there is deep overlap in the molecular mechanisms involved in all neurodegenerative diseases. For example, the same "Cu–Mo–S circuitry" described here for AD is also engaged in PD [141–143]. The therapy with Cu chelating agents was also performed for PD, with no positive result [144]. Therefore, although neurodegenerative diseases have different causes and physiopathological features, their courses involve several common metabolic pathways, which might mean that the therapeutic intervention focused on the "Cu–Mo–S circuitry" would not act in the onset of the development of all neurodegenerative diseases. By the same reasoning, this circuitry might not be the origin but a converging circuitry during neurodegeneration. Future research in this field is needed to shed light on these questions.

Another point concernscholine, folate, and Mo supplementation effects in AD. Choline supplementation significantly reduced A $\beta$  load and improved spatial memory in APP/PS1 mice [67]. Regardingfolate, the conclusions are inconsistent while supplementing the elderly with different folic acid states [145]. To date, no research related to Mo supplementation has been conducted in humans and this would bean interesting subject for future research.

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#### Abbreviations

AD, Alzheimer's disease; AO, aldehyde oxidase; APP, Amyloid Precursor Protein; Aβ, amyloid β plaques; CBS, cystathionine b-synthase; COX, cytochrome c oxidase; Cu, copper; Cys, cysteine; DMT, disease-modifying therapies; GSH, glutathione; Hcy, homocysteine; HS, healthy subjects; mARC, mitochondrial amidoxime reducing component; MCI, mild cognitive impairment; Met, methionine; Met-SO, methionine-sulfoxide; MNC, mild neurological changes; Mo, molybdenum; Moco, Mo co-factor; nCp–Cu, non-ceruloplasmin Cu; PD, Parkinson's disease; S, sulfur; SMC, subjective memory complaint; SUOX, sulfite oxidase; T2D, type 2 diabetes; XO, xanthine oxidase-dehydrogenase.

### References

- 1. WHO. *Risk Reduction of Cognitive Decline and Dementia: WHO Guidelines;* Licence: CC BY-NC-SA 3.0 IGO; World Health Organization: Geneva, Switzerland, 2019; ISBN 978-92-4-155054-3.
- Socha, K.; Klimiuk, K.; Naliwajko, S.K.; Soroczyńska, J.; Puścion-Jakubik, A.; Markiewicz-Żukowska, R.; Kochanowicz, J. Dietary Habits, Selenium, Copper, Zinc and Total Antioxidant Status in Serum in Relation to Cognitive Functions of Patients with Alzheimer's Disease. *Nutrients* 2021, 13, 287. [CrossRef]
- Stocker, H.; Nabers, A.; Perna, L.; Möllers, T.; Rujescu, D.; Hartmann, A.; Holleczek, B.; Schöttker, B.; Stockmann, J.; Gerwert, K.; et al. Genetic predisposition, Aβ misfolding in blood plasma, and Alzheimer's disease. *Transl. Psychiatry* 2021, 11, 261. [CrossRef]
- Lalioti, V.; Muruais, G.; Tsuchiya, Y.; Pulido, D.; Sandoval, I.V. Molecular mechanisms of copper homeostasis. *Front. Biosci.* 2009, 14, 4878–4903. [CrossRef]
- 5. Palego, L.; Betti, L.; Giannaccini, G. Sulfur Metabolism and Sulfur-Containing Amino Acids: I-Molecular Effectors. *Biochem. Pharmacol.* **2015**, *4*, 1. [CrossRef]
- 6. World Health Organization (WHO). *Trace Elements in Human Nutrition and Health;* Prepared in Collaboration with the FAO of the UN and the IAEA; World Health Organization: Geneva, Switzerland, 1996.
- 7. Heafield, M.T.; Fearn, S.; Steventon, G.B.; Waring, R.H.; Williams, A.C. Plasma cysteine and sulphate levels in patients with motor neurone, Parkinson's and Alzheimer's disease. *Neurosci. Lett.* **1990**, *110*, 216–220. [CrossRef]
- Squitti, R.; Faller, P.; Hureau, C.; Granzotto, A.; White, A.R.; Kepp, K.P. Copper Imbalance in Alzheimer's Disease and Its Link with the Amyloid Hypothesis: Towards a Combined Clinical, Chemical, and Genetic Etiology. J. Alzheimers Dis. 2021, 83, 23–41. [CrossRef]
- Irreverre, F.; Mudd, H.; Heizer, W.D.; Laster, L. Sulfite oxidase deficiency: Studies of a patient with mental retardation, dislocated ocular lenses, and abnormal urinary excretion of S-sulfo-l-cysteine, sulfite, and thiosulfate. *Biochem. Med.* 1967, 1, 187–217. [CrossRef]
- 10. Brown, O.K.; Scholem, R.D.; Croll, H.B.; Wraith, J.E.; McGill, J.J. Sulfite oxidase deficiency. Clinical, neuroradiologic, and biochemical features in two new patients. *Neurology* **1989**, *39*, 252. [CrossRef]
- 11. Bianchi, V.E. Impact of Testosterone on Alzheimer's Disease. World J. Mens Health 2022, 40, 243–256. [CrossRef]
- 12. Ingenbleek, Y. Implications of protein malnutrition and inflammatory disorders in the pathophysiology of Alzheimer's disease. *Asia Pac. J. Clin. Nutr.* **2020**, *29*, 450–461. [CrossRef]
- Li, S.; Sun, W.; Zhang, D. Association of Zinc, Iron, Copper, and Selenium Intakes with Low Cognitive Performance in Older Adults: A Cross-Sectional Study from National Health and Nutrition Examination Survey (NHANES). J. Alzheimers Dis. 2019, 72, 1145–1157. [CrossRef]
- 14. Gouaref, I.; Bellahsene, Z.; Zekri, S.; Alamir, B.; Koceir, E.A. The link between trace elements and metabolic syndrome/oxidative stress in essential hypertension with or without type 2 diabetes. *Ann. Biol. Clin.* **2016**, *74*, 233–243. [CrossRef]
- 15. Squitti, R.; Mendez, A.J.; Simonelli, I.; Ricordi, C. Diabetes and Alzheimer's Disease: Can Elevated Free Copper Predict the Risk of the Disease? *J. Alzheimers Dis.* **2017**, *56*, 1055–1064. [CrossRef]
- 16. Zhou, H.H.; Lu, F.; Chen, S.D.; Zhou, Z.H.; Han, Y.Z.; Hu, J.Y. Effect of electroacupuncture on serum copper, zinc, calcium and magnesium levels in the depression rats. *J. Tradit. Chin. Med.* **2011**, *31*, 112–114. [CrossRef]
- 17. Salustri, C.; Squitti, R.; Zappasodi, F.; Ventriglia, M.; Bevacqua, M.G.; Fontana, M.; Tecchio, F. Oxidative stress and brain glutamate-mediated excitability in depressed patients. J. Affect. Disord. 2010, 127, 321–325. [CrossRef]
- Ni, M.; You, Y.; Chen, J.; Zhang, L. Copper in depressive disorder: A systematic review and meta-analysis of observational studies. *Psychiatry Res.* 2018, 267, 506–515. [CrossRef]
- 19. Xu, J.; He, K.; Zhang, K.; Yang, C.; Nie, L.; Dan, D.; Liu, J.; Zhang, C.; Yang, X. Low-Dose Copper Exposure Exacerbates Depression-Like Behavior in ApoE4 Transgenic Mice. *Oxidative Med. Cell. Longev.* **2021**, *25*, 6634181. [CrossRef]
- 20. Chatopadhyay, A.; Sarkar, M.; Biswas, N.M. Dose-dependent effect of copper chloride on malereproductive function in immature rats. *Kathmandu Univ. Med. J.* 2005, *3*, 392–400.
- Chang, C.S.; Choi, J.B.; Kim, H.J.; Park, S.B. Correlation between serum testosterone level and concentrations of copper and zinc in hair tissue. *Biol. Trace Elem. Res.* 2011, 144, 264–271. [CrossRef]
- 22. Rosario, E.R.; Stanczyk, F.Z.; Pike, C.J. Age-Related Testosterone Depletion and the Development of Alzheimer Disease. *JAMA J. Am. Med. Assoc.* 2004, 292, 1431–1432. [CrossRef]
- 23. Lewis, R.C.; Meeker, J.D. Biomarkers of Exposure to Molybdenum and Other Metals in Relation to Testosterone among Men from NHANES 2011–2012. *Fertil. Steril.* 2015, 103, 172–178. [CrossRef]
- 24. Khosravi, M.; Sotoudeh, G.; Amini, M.; Raisi, F.; Mansoori, A.; Hosseinzadeh, M. The relationship between dietary patterns and depression mediated by serum levels of Folate and vitamin B12. *Psychiatry* **2020**, *20*, *63*. [CrossRef]
- 25. Bhatia, P.; Singh, N. Homocysteine excess: Delineating the possible mechanism of neurotoxicity and depression. *Fundam. Clin. Pharmacol.* **2015**, *29*, 522–528. [CrossRef]
- Namekawa, Y.; Baba, H.; Maeshima, H.; Nakano, Y.; Satomur, E.; Takebayashi, N.; Nomoto, H.; Suzuki, T.; Arai, H. Heterogeneity of elderly depression: Increased risk of Alzheimer's disease and Aβ protein metabolism. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 2013, 43, 203–208. [CrossRef]

- WHO. Evaluations of the Joint FAO/WHO Expert Committee on Food Additives (JECFA). Copper. CAS Number: 7440-50-8. Tox Monograph: FAS 17-JECFA 26/265. 1982. Available online: http://www.inchem.org/documents/jecfa/jecmono/v17je31.htm (accessed on 12 January 2020).
- 28. Pohanka, M. Copper and copper nanoparticles toxicity and their impact on basic functions in the body. *Bratisl. Med. J.* 2019, 120, 397–409. [CrossRef]
- Di Donato, M.; Sarkar, B. Copper transport and its alterations in Menkes and Wilson diseases. *Biochim. Biophys. Acta* 1997, 1360, 3–16. [CrossRef]
- Flores, C.R.; Puga, M.P.; Wrobel, K.; Sevilla, M.E.G.; Wrobel, K. Trace elements status in diabetes mellitus type 2: Possible role of the interaction between molybdenum and copper in the progress of typical complications. *Diabetes Res. Clin. Pract.* 2011, *91*, 333–341. [CrossRef]
- Institute of Medicine—US. Panel on Micronutrients. Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc; National Academy Press: Washington, DC, USA, 2002; 773p.
- Massie, H.R.; Aiello, V.R.; Iodice, A.A. Changes with age in copper and superoxide dismutase levels in brains of C57BL/6J mice. Mech. Ageing Dev. 1979, 10, 93–99. [CrossRef]
- 33. Zatta, P.; Drago, D.; Zambenedetti, P.; Bolognin, S.; Nogara, E.; Peruffo, A.; Cozzi, B. Accumulation of copper and other metal ions, and metallothionein I/II expression in the bovine brain as a function of aging. *J. Chem. Neuroanat.* **2008**, *36*, 1–5. [CrossRef]
- Vasudevaraju, P.; Bharathi; Jyothsna, T.; Shamasundar, N.M.; Rao, K.S.; Balaraj, B.M.; Rao, K.S.J.; Sathyanarayana, R.T.S. New evidence on iron, copper accumulation and zinc depletion and its correlation with DNA integrity in aging human brain regions. *Indian J. Psychiatry* 2010, 52, 140–144. [CrossRef]
- 35. Kepp, K.P.; Squitti, R. Copper imbalance in Alzheimer's disease: Convergence of the chemistry and the clinic. *Coord. Chem. Rev.* **2019**, *397*, 168–187. [CrossRef]
- Squitti, R.; Ventriglia, M.; Simonelli, I.; Bonvicini, C.; Costa, A.; Perini, G.; Binetti, G.; Benussi, L.; Ghidoni, R.; Koch, G.; et al. Copper Imbalance in Alzheimer's Disease: Meta-Analysis of Serum, Plasma, and Brain Specimens, and Replication Study Evaluating ATP7B Gene Variants. *Biomolecules* 2021, 11, 960. [CrossRef]
- Coelho, F.C.; Squitti, R.; Ventriglia, M.; Cerchiaro, G.; Daher, J.P.; Rocha, J.G.; Rongioletti, M.C.A.; Moonen, A.C. Agricultural Use of Copper and Its Link to Alzheimer's Disease. *Biomolecules* 2020, 10, 897. [CrossRef]
- Masaldana, S.; Sharnel, A.S.; Clatworthya, C.G.; Smithd, Z.M.; Francis, P.S.; Denoyera, D.; Meggyesya, P.M.; Fontainea, S.L.; Cater, M.A. Copper accumulation in senescent cells: Interplay between copper transporters and impaired autophagy. *Redox Biol.* 2018, 16, 322–333. [CrossRef]
- Boilan, E.; Winant, V.; Dumortier, E.; Piret, J.P.; Bonfitto, F.; Osiewacz, H.D.; Debacq-Chainiaux, F.; Toussaint, O. Role of p38MAPK and oxidative stress in copper-induced senescence. Age 2013, 35, 2255–2271. [CrossRef]
- 40. Gaetke, L.M.; Chow, C.K. Copper toxicity, oxidative, and antioxidant nutrients. Toxicology 2003, 189, 147–163. [CrossRef]
- 41. Brewer, G.J. Copper-2 Hypothesis for Causation of the Current Alzheimer's Disease Epidemic Together with Dietary Changes That Enhance the Epidemic. *Chem. Res. Toxicol.* **2017**, *30*, 763–768. [CrossRef]
- 42. Squitti, R.; Ghidoni, R.; Simonelli, I.; Ivanova, I.D.; Colabufo, N.A.; Zuin, M.; Benussi, L.; Binetti, G.; Cassetta, E.; Rongioletti, M.; et al. Copper dyshomeostasis in Wilson disease and Alzheimer's disease as shown by serum and urine copper indicators. *J. Trace Elem. Med. Biol.* **2018**, *45*, 181–188. [CrossRef]
- Hsu, H.; Rodriguez-Ortiz, C.J.; Lam Lim, S.; Zumkehr, J.; Kilian, J.G.; Vidal, J.; Kitazawa, M. Copper-Induced Upregulation of MicroRNAs Directs the Suppression of Endothelial LRP1 in Alzheimer's Disease Model. *Toxicol. Sci.* 2019, 170, 144–156. [CrossRef]
- 44. Esmieu, C.; Guettas, D.; Conte-Daban, A.; Sabater, L.; Faller, P.; Hureau, C. Copper-Targeting Approaches in Alzheimer's Disease: How to Improve the Fallouts Obtained from In Vitro Studies. *Inorg. Chem.* **2019**, *58*, 13509–13527. [CrossRef]
- 45. Hsu, H.-W.; Stephen, C.B.; Masashi, K. Environmental and Dietary Exposure to Copper and Its Cellular Mechanisms Linking to Alzheimer's Disease. *Toxicol. Sci.* 2018, 163, 338–345. [CrossRef]
- 46. James, S.A.; Volitakis, I.; Adlard, P.A.; Duce, J.A.; Masters, C.L.; Cherny, R.A.; Bush, A.I. Elevated labile Cu is associated with oxidative pathology in Alzheimer's Disease. *Free Radic. Biol. Med.* **2012**, *52*, 298–302. [CrossRef]
- Pérez-Aguilar, F.; Burguera, J.A.; Benlloch, S.; Berenguer, M.; Rayón, J.M. Aceruloplasminemia in an asymptomatic patient with a new mutation. *Diagn. Fam. Genet. Anal.* 2005, 42, 947–949. [CrossRef]
- 48. Squitti, R.; Siotto, M.; Arciello, M.; Rossi, L. Non-ceruloplasmin bound copper and ATP7B gene variants in Alzheimer's disease. *Metallomics* **2016**, *8*, 863–873. [CrossRef]
- 49. Bagheri, S.; Squitti, R.; Haertlé, T.; Siotto, M.C.; Saboury, A.A. Role of Copper in the Onset of Alzheimer's Disease Compared to Other Metals. *Front. Aging Neurosci.* 2018, 9, 446. [CrossRef]
- Chen, C.; Jiang, X.; Li, Y.; Yu, H.; Li, S.; Zhang, Z.; Xu, H.; Yang, Y.; Liu, G.; Zhu, F.; et al. Low-dose oral copper treatment changes the hippocampal phosphoproteomic profile and perturbs mitochondrial function in a mouse model of Alzheimer's disease. *Free Radic. Biol. Med.* 2019, 135, 144–156. [CrossRef]
- 51. Zhao, J.; Shi, Q.; Tian, H.; Li, Y.; Liu, Y.; Xu, Z.; Robert, A.; Liu, Q.; Meunier, B. TDMQ20, a Specific Copper Chelator, Reduces Memory Impairments in Alzheimer's Disease Mouse Models. *ACS Chem. Neurosci.* 2021, 12, 140–149. [CrossRef]

- 52. Varadarajan, S.; Yatin, S.; Kanski, J.; Jahanshahi, F.; Butterfield, D.A. Methionine residue 35 is important in amyloid b peptideassociated free radical oxidative stress. *Brain Res. Bull.* **1999**, *50*, 133–141. [CrossRef]
- 53. Weng, W.; Huang, W.; Tang, H.; Cheng, M.; Chen, K. The Differences of Serum Metabolites between Patients with Early-Stage Alzheimer's Disease and Mild Cognitive Impairment. *Front. Neurol.* **2019**, *10*, 1223. [CrossRef]
- 54. Kruman, I.I.; Kumaravel, T.S.; Lohani, A.; Pedersen, W.A.; Cutler, R.G.; Kruman, Y.; Haughey, N.; Lee, J.; Evans, M.; Mattson, M.P. Folic Acid Deficiency and Homocysteine Impair DNA Repair in Hippocampal Neurons and Sensitize Them to Amyloid Toxicity in Experimental Models of Alzheimer's Disease. J. Neurosci. 2002, 22, 1752–1762. [CrossRef]
- Leermakers, E.T.M.; Moreira, E.M.; Kiefte-de Jong, J.C.; Darweesh, S.K.L.; Visser, T.; Voortman, T.; Bautista, P.K.; Chowdhury, R.; Gorman, D.; Bramer, W.M.; et al. Effects of choline on health across the life course: Asystematic review. *Nutr. Rev.* 2015, 73, 500–522. [CrossRef] [PubMed]
- 56. Moss, M.; Waring, R.H. The Plasma Cysteine/Sulphate Ratio: A Possible Clinical Biomarker. J. Nutr. Environ. Med. 2003, 13, 215–229. [CrossRef]
- 57. Luo, Y.; Zhang, L.; Liu, W.; Yu, Y.; Tian, Y. A Single Biosensor for Evaluating the Levels of Copper Ion and L-Cysteine in a Live Rat Brain with Alzheimer's Disease. *Angew. Chem. Int. Ed.* **2015**, *54*, 14053–14056. [CrossRef] [PubMed]
- Mandal, P.K.; Saharan, S.; Tripathi, M.; Murari, G. Brain Glutathione Levels—A Novel Biomarker for Mild Cognitive Impairment and Alzheimer's Disease. *Biol. Psychiatry* 2015, 79, 71. [CrossRef]
- 59. Quadri, P.; Fragiacomo, C.; Pezzati, R.; Zanda, E.; Forloni, G.; Tettamanti, M.; Lucca, U. Homocysteine, folate, and vitamin B-12 in mild cognitive impairment, Alzheimer disease, and vascular dementia. *Am. J. Clin. Nutr.* **2004**, *80*, 114–122. [CrossRef]
- Babiloni, C.; Bosco, P.; Ghidoni, R.; Del Percio, C.; Squitti, R.; Binetti, G.; Benussi, L.; Ferri, R.; Frisoni, G.; Lanuzza, B.; et al. Homocysteine and electroencephalographic rhythms in Alzheimer disease: A multicentric study. *Neuroscience* 2007, 145, 942–954. [CrossRef]
- 61. Gorgone, G.; Ursini, F.; Altamura, C.; Bressi, F.; Tombini, M.; Curcio, G.; Chiovenda, P.; Squitti, R.; Silvestrini, M.; Ientile, R.; et al. Hyperhomocysteinemia, intima-media thickness and C677T MTHFR gene polymorphism: A correlation study in patients with cognitive impairment. *Atherosclerosis* **2009**, *206*, 309–313. [CrossRef]
- Ma, F.; Wu, T.; Zhao, J.; Ji, L.; Song, A.; Zhang, M.; Huang, G. Plasma Homocysteine and Serum Folate and Vitamin B12 Levels in Mild Cognitive Impairment and Alzheimer's Disease: A Case-Control Study. *Nutrients* 2017, 9, 725. [CrossRef]
- 63. Pietrzik, K.; Brönstrup, A. Vitamins B12, B6 and folate as determinants of homocysteine concentration in the healthy population. *Eur. J. Pediatr.* **1998**, 157 (Suppl. 2), S135–S138. [CrossRef]
- Irizarry, M.C.; Gurol, M.E.; Raju, S.; Diaz-Arrastia, R.; Locascio, J.J.; Tennis, M.; Hyman, B.T.; Growdon, J.H.; Greenberg, S.M.; Bottiglieri, T. Association of homocysteine with plasma amyloid protein in aging and neurodegenerative disease. *Neurology* 2005, 65, 1402–1408. [CrossRef]
- 65. Tinelli, C.; Di Pino, A.; Ficulle, E.; Marcelli, S.; Feligioni, M. Hyperhomocysteinemia as a Risk Factor and Potential Nutraceutical Target for Certain Pathologies. *Front. Nutr.* **2019**, *6*, 49. [CrossRef] [PubMed]
- 66. Robinson, N.; Grabowski, P.; Rehman, I. Alzheimer's disease pathogenesis: Is there a role for folate? *Mech. Ageing Dev.* **2018**, 174, 86–94. [CrossRef] [PubMed]
- Mellott, T.J.; Huleatt, O.M.; Shade, B.N.; Pender, S.M.; Liu, Y.B.; Slack, B.E.; Blusztajn, J.K. Perinatal Choline Supplementation Reduces Amyloidosis and Increases Choline Acetyltransferase Expression in the Hippocampus of the APPswePS1dE9 Alzheimer's Disease Model Mice. *PLoS ONE* 2017, 12, e0170450. [CrossRef]
- 68. Wilde, M.C.; Vellas, B.; Giraulta, E.; Yavuza, A.C.; Sijbena, J.W. Lower brain and blood nutrient status in Alzheimer's disease: Results from meta-analyses. *Alzheimers Dement.* **2017**, *3*, 416–431. [CrossRef]
- Caballero, F.; Fernandez, A.; Matias, N.; Martínez, L.; Fucho, R.; Elena, M.; Caballeria, J.; Morales, A.; Fernández-Checa, J.C.; García-Ruiz, C. Specific contribution of methionine and choline in nutritional nonalcoholic steatohepatitis: Impact on mitochondrial S-adenosyl-L-methionine and glutathione. J. Biol. Chem. 2010, 285, 18528–18536. [CrossRef]
- 70. Novotny, J.A. Molybdenum Nutriture in Humans. J. Evid.-Based Complement. Altern. Med. 2011, 16, 164–168. [CrossRef]
- Richards, J.M.; Swislocki, N.I. Activation of Adenylate Cyclase by Molybdate. J. Biol. Chem. 1979, 254, 6857–6860. Available online: https://www.jbc.org/article/S0021-925850251-9/pdf (accessed on 20 November 2020). [CrossRef]
- 72. Richards, J.M.; Swislocki, N.I. Mechanism of molybdate activation of adenylate cyclase. *Biochim. Biophys. Acta* **1981**, 678, 180–186. [CrossRef]
- Sim, A.T.R.; White, M.D.; Denborough, M.A. Effects of adenylate cyclase activators on porcine skeletal muscle in malignant hyperpyrexia. Br. J. Anaesth. 1987, 59, 1557–1562. [CrossRef]
- 74. Rajagopalan, K.V. Molybdenum: An essential trace element in human nutrition. Ann. Rev. Nutr. 1988, 8, 401–427. [CrossRef]
- 75. Coelho, F.C.; Vieira, C.; Mosquim, P.R.; Cassini, S.T.A. Nitrogênio e molibdênionasculturas do milho e do feijãoemmonocultivos e emconsórcios: Efeitossobre o feijão. *Rev. Ceres* **1998**, *45*, 393–407.
- 76. Khadanand, K.; Kirk, M.L. Mo and W Cofactors and the Reactions they Catalyze. Met. Ions Life Sci. 2020, 23, 20. [CrossRef]
- 77. Schwarz, G.; Mendel, R.R. Molybdenum co-factor biosynthesis and molybdenum enzymes. *Annu. Rev. Plant Biol.* 2006, 57, 623–647. [CrossRef]
- Scientific Committee on Food, European Commission. Opinion of the Scientific Committee on Food on the Tolerable Upper Intake Level of Molybdenum; SCF/SC/NUT/UPPERLEV/22; Scientific Committee on Food, European Commission: Brussels, Belgium, 2000; pp. 1–15.

- Baj, J.; Forma, A.; Sitarz, E.; Karakuła, K.; Flieger, W.; Sitarz, M.; Grochowski, C.; Maciejewski, R.; Karakula-Juchnowicz, H. Beyond the Mind-Serum Trace Element Levels in Schizophrenic Patients: A Systematic Review. *Int. J. Mol. Sci.* 2020, 21, 9566. [CrossRef]
- 80. Giussani, A.; Arogunjo, A.M.; Claire Cantone, M.; Tavola, F.; Veronese, I. Rates of intestinal absorption of molybdenum in humans. *Appl. Radiat. Isot.* **2006**, *64*, 639–644. [CrossRef]
- Johnson, L.E. Molybdenum Deficiency. Merck Manual Professional Version. Available online: https://www.merckmanuals.com/ professional/nutritional-disorders/mineral-deficiency-and-toxicity/molybdenum-deficiency# (accessed on 20 November 2021).
- 82. Ardan, T.; Kovaceva, J.; Cejková, J. Comparative histochemical and immunohistochemical study on xanthine oxidoreductase/xanthine oxidase in mammalian corneal epithelium. *Acta Histochem.* **2004**, *106*, 69–75. [CrossRef]
- 83. Hille, R. Molybdenum-containing hydroxylases. Arch. Biochem. Biophys. 2005, 433, 107–116. [CrossRef]
- 84. Harrison, R. Structure and function of xanthine oxidoreductase: Where are we now? *Free Radic. Biol. Med.* **2002**, *33*, 774–797. [CrossRef]
- 85. Sebesta, I.; Stiburkova, B.; Krijt, J. Hereditary xanthinuria is not so rare disorder of purine metabolism. *Nucleosides Nucleotides Nucleic Acids* **2018**, *37*, 324–328. [CrossRef]
- Alonso-Andrés, P.; Albasanz, J.L.; Ferrer, I.; Martín, M. Purine-related metabolites and their converting enzymes are altered in frontal, parietal and temporal cortex at early stages of Alzheimer's disease pathology. *Brain Pathol.* 2018, 28, 933–946. [CrossRef]
- 87. Du, N.; Xu, D.; Hou, X.; Song, X.; Liu, C.; Chen, Y.; Wang, Y.; Li, X. Inverse Association between Serum Uric Acid Levels and Alzheimer's Disease Risk. *Mol. Neurobiol.* **2016**, *53*, 2594–2599. [CrossRef]
- Tana, C.; Ticinesi, A.; Prati, B.; Nouvenne, A.; Meschi, T. Uric Acid and Cognitive Function in Older Individuals. *Nutrients* 2018, 10, 975. [CrossRef]
- 89. Latourte, A.; Bardin, T.; Richette, P. Uric acid and cognitive decline: A double-edge sword? *Curr. Opin. Rheumatol.* **2018**, 30, 183–187. [CrossRef]
- 90. Griffith, O.W. Mammalian sulfur amino acid metabolism: An overview. Methods Enzymol. 1987, 143, 366–376. [CrossRef]
- Mellis, A.; Misko, A.L.; Arjune, S.; Liang, Y.; Erdélyi, K.; Ditrói, T.; Kaczmarek, A.T.; Nagy, P.; Schwarz, G. The role of glutamate oxaloacetate transaminases in sulfite biosynthesis and H<sub>2</sub>S metabolism. *Redox Biol.* 2021, *38*, 101800. [CrossRef]
- 92. Abe, K.; Kimura, H. The Possible Role of Hydrogen Sulfide as an Endogenous Neuromodulator. J. Neurosci. 1996, 16, 1066–1071. [CrossRef]
- 93. Eto, K.; Asada, T.; Arima, K.; Makifuchi, T.; Kimura, H. Brain hydrogen sulfide is severely decreased in Alzheimer's disease. *Biochem. Biophys. Res. Commun.* 2002, 293, 1485–1488. [CrossRef]
- 94. Velayutham, M.; Hemann, C.F.; Cardounel, A.J.; Zweiera, J.L. Sulfite oxidase activity of cytochrome c: Role of hydrogen peroxide. *Biochem. Biophys. Rep.* **2016**, *5*, 96–104. [CrossRef]
- Bosetti, F.; Brizzi, F.; Barogi, S.; Mancuso, M.; Siciliano, G.; Tendi, E.A.; Murri, L.; Rapoport, S.L.; Solaini, G. Cytochrome c oxidase and mitochondrial F1F0-ATPase (ATP synthase) activities in platelets and brain from patients with Alzheimer's disease. *Neurobiol. Aging* 2002, 23, 371–376. [CrossRef]
- Maurer, I.; Zierz, S.; Möller, H. A selective defect of cytochrome c oxidase is present in brain of Alzheimer disease patients. *Neurobiol. Aging* 2000, 21, 455–462. [CrossRef]
- Kuper, J.; Llamas, A.; Hecht, H.; Mendel, R.R.; Schwarz, G. Structure of the molybdopterin-bound Cnx1G domain links molybdenum and copper metabolism. *Nature* 2004, 430, 803–806. [CrossRef] [PubMed]
- Alonzo-Martínez, M.C.; Cazorla, E.; Cánovas, E.; Anniuk, K.; Cores, A.E.; Serrano, A.M. Molybdenum Cofactor Deficiency: Mega Cisterna Magna in Two Consecutive Pregnancies and Review of the Literature. *Appl. Clin. Genet.* 2020, 13, 49–55. [CrossRef] [PubMed]
- 99. Misko, A.L.; Liang, Y.; Kohl, J.B.; Eichler, F. Delineating the phenotypic spectrum of sulfite oxidase and molybdenum co-factor deficiency. *Neurol. Genet.* 2020, *6*, e486. [CrossRef] [PubMed]
- Berry, T.; Abohamza, E.; Moustafa, A.A. A disease-modifying treatment for Alzheimer's disease: Focus on the trans-sulfuration pathway. *Rev. Neurosci.* 2020, *31*, 319–334. [CrossRef]
- 101. Xia, Z.; Storm, D. Role of Circadian Rhythm and REM sleep for memory consolidation. Neurosci. Res. 2017, 118, 13–20. [CrossRef]
- Yamamoto, M.; Gotz, M.E.; Ozawa, H.; Luckhaus, C.; Saito, T.; Rosler, M.; Riederer, P. Hippocampal level of neural speci¢c adenylyl cyclase type I is decreased in Alzheimer's disease. *Biochim. Biophys. Acta* 2000, 1535, 60–68. [CrossRef]
- Kelly, M.P. Cyclic nucleotide signaling changes associated with normal aging and age-related diseases of the brain. *Cell. Signal.* 2018, 42, 281–291. [CrossRef]
- Nonaka, N.; Banks, A.W.; Shioda, S. Pituitary adenylate cyclase-activating polypeptide: Protective effects in stroke and dementia. *Peptides* 2020, 130, 17033. [CrossRef]
- 105. Abumrad, N.N.; Schneider, A.J.; Steel, D.; Rogers, L.S. Amino acid intolerance during prolonged total parenteral nutrition reversed by molybdate therapy. *Am. J. Clin. Nutr.* **1981**, *34*, 2551–2559. [CrossRef]
- 106. Hajjar, I.; Liu, C.; Jones, D.P.; Uppal, K. Untargeted metabolomics reveal dysregulations in sugar, methionine, and tyrosine pathways in the prodromal state of AD. *Alzheimers Dement.* **2020**, *12*, e12064. [CrossRef]
- 107. Atwal, P.S.; Scaglia, F. Molybdenum co-factor deficiency. Mol. Genet. Metab. 2016, 117, 1–4. [CrossRef] [PubMed]
- Kong, L.; Zhou, X.; Shi, G.; Yu, Y. Molybdenum disulfide nanosheets based fluorescent "off-to-on" probe for targeted monitoring and inhibition of Î<sup>2</sup>-amyloid oligomers. *Analyst* 2020, 145, 6369–6377. [CrossRef] [PubMed]

- 109. Manna, S.; Karmakar, P.; Kisan, B.; Mishra, M.; Barooah, N.; Bhasikuttan, A.C.; Mohant, J. Fibril-induced neurodegenerative disorders in an Ab-mutant Drosophila model: Therapeutic targeting using ammonium molybdate. *Chem. Commun.* 2021, 57, 8488. [CrossRef]
- Kulwich, R.; Hansard, S.L.; Comar, C.L.; Davis, G.K. Copper, Molybendurn and Zinc Interrelationships in Rats and Swine. *Exp. Biol. Med.* 1953, 84, 2. [CrossRef]
- 111. Halverson, A.W.; Phifer, J.H.; Monty, K.J. A mechanism for the copper-molybdenum interrelationship. *J. Nutr.* **1960**, *71*, 95–100. [CrossRef] [PubMed]
- 112. Gray, L.F.; Daniel, L.J. Effect of the Copper Status of the Rat on the Copper-Molybdenum-Sulfate Interaction. *J. Nutr.* **1964**, *84*, 31–37. [CrossRef]
- 113. Suttle, N.F. Copper Imbalances in Ruminants and Humans: Unexpected Common Ground. Adv. Nutr. 2012, 3, 666–674. [CrossRef]
- 114. Dowdy, R.P.; Matrone, G. A Copper-Molybdenum Complex: Its effects and movement in the piglet and sheep. *J. Nutr.* **1968**, *95*, 197–201. [CrossRef]
- Dowdy, R.P.; Kunz, G.A.; Sauberlich, H.E. Effect of a Copper-Molybdenum Compound upon Copper Metabolism in the Rat. J. Nutr. 1969, 99, 491–496. [CrossRef]
- 116. Arthur, D. Interrelationships of Molybdenum and Copper in the Diet of the Guinea Pig. J. Nutr. 1965, 87, 69–76. [CrossRef]
- 117. Hadizadeh, M.; Keyhani, E.; Keyhani, J.; Khodadadi, C. Functional and structural alterations induced by copper in xanthine oxidase. *Acta Biochim. Biophys. Sin.* 2009, 41, 603–617. [CrossRef] [PubMed]
- 118. Mendel, R.R.; Kruse, T. Cell biology of molybdenum in plants and humans. *Biochim. Biophys. Acta (BBA)-Mol. Cell Res.* 2012, 1823, 1568–1579. [CrossRef] [PubMed]
- 119. Suzuki, K.T.; Karasawa, A.; Yamanaka, K. Binding of copper to albumin and participation of cysteine in vivo and in vitro. *Arch. Biochem. Biophys.* **1989**, 273, 572–577. [CrossRef]
- Mahoney, J.P.; Sandberg, A.A.; Gubler, C.J.; Cartwright, G.E.; Wintrobe, M.M. Uric Acid Metabolism in Hepatolenticular Degeneration. *Exp. Biol. Med.* 1955, 88, 427–430. [CrossRef]
- Seelig, M.S. Proposed role of copper—Molybdenum interaction in iron-deficiency and iron-storage diseases. *Am. J. Clin. Nutr.* 1973, 26, 657–672. [CrossRef] [PubMed]
- 122. Bar-Or, D.; Rael, L.; Thomas, G.; Kraus, J. Inhibitory Effect of Copper on Cystathionine β-Synthase Activity: Protective Effect of an Analog of the Human Albumin N-Terminus. *Protein Pept. Lett.* **2005**, *12*, 271–273. [CrossRef]
- 123. Vasquez, E.F.A.; Herrera, A.P.N.; Santiago, G.S. Interaçãocobre, molibdênio e enxofreemruminantes. *Cienc. Rural* 2001, *31*, 1101–1106. [CrossRef]
- 124. Paglia, G.; Miedico, O.; Cristofano, A.; Vitale, M.; Angiolillo, A.; Chiaravalle, A.E.; Corso, G.; Costanzo, A.D. Distinctive Pattern of Serum Elements during the Progression of Alzheimer's. *Dis. Sci. Rep.* **2016**, *9*, 22769. [CrossRef]
- 125. Squitti, R.; Pasqualetti, P.; Polimanti, R.; Salustri, C.; Moffa, F.; Cassetta, E.; Lupoi, D.; Ventriglia, M.; Cortesi, M.; Siotto, M.; et al. Metal-score as a potential non-invasive diagnostic test for Alzheimer's disease. *Curr. Alzheimer Res.* 2013, *10*, 191–198. [CrossRef]
- 126. Squitti, R.; Ghidoni, R.; Siotto, M.; Ventriglia, M.; Benussi, L.; Paterlini, A.; Magri, M.; Binetti, G.; Cassetta, E.; Caprara, D.; et al. Value of serum nonceruloplasmin copper for prediction of mild cognitive impairment conversion to Alzheimer disease. *Ann. Neurol.* 2014, 75, 574–580. [CrossRef]
- 127. Smith, B.S.W.; Wright, H. Copper: Molybdenum interaction: Effect of dietary molybdenum on the binding of copper to plasma proteins in sheep. *J. Comp. Pathol.* **1975**, *85*, 299–305. [CrossRef]
- 128. Post, L.O.; Bailey, E.M., Jr.; Williams, S.; Applegate, C.; Jones, B.; Herrman, T.J. Risk Assessment of Copper and Molybdenum and Other Minerals in Feed Ingredients and Finished Feeds. *J. Regul. Sci.* **2019**, *7*, 1–9. [CrossRef]
- 129. Badgley, M.A.; Kremer, D.M.; Maurer, H.C.; DelGiorno, K.E.; Lee, H.; Purohit, V.; Sagalovskiy, I.R.; Ma, A.; Kapilian, J.; Firl, C.E.M.; et al. Cysteine depletion induces pancreatic tumor ferroptosis in mice. *Science* **2020**, *368*, 85–89. [CrossRef]
- 130. Paul, B.D. Neuroprotective Roles of the Reverse Transsulfuration Pathway in Alzheimer's Disease. *Front. Aging Neurosci.* **2021**, 13, 659402. [CrossRef]
- 131. Grings, M.; Moura, A.P.; Amaral, A.U.; Parmeggiani, B.; Gasparotto, J.; Moreira, J.C.F.; Gelain, D.P.; Wyse, A.T.S.; Wajner, M.; Leipnitz, G. Sulfite disrupts brain mitochondrial energy homeostasis and induces mitochondrial permeability transition pore opening via thiol group modification. *Biochim. Biophys. Acta* 2014, 1842, 1413–1422. [CrossRef] [PubMed]
- 132. Tan, W.; Eichler, F.S.; Hoda, S.; Lee, M.S.; Baris, H.; Hanley, C.A.P.; Grant, E.; Krishnamoorthy, K.S.; Shih, V.E. Isolated Sulfite Oxidase Deficiency: A Case Report with a Novel Mutation and Review of the Literature. *Pediatrics* **2005**, *116*, 757. [CrossRef]
- 133. Cummings, J.; Ritter, A.; Zhong, K. Clinical Trials for Disease-Modifying Therapies in Alzheimer's Disease: A Primer, Lessons Learned, and a Blueprint for the Future. *J. Alzheimers Dis.* **2018**, *64*, S3–S22. [CrossRef]
- Liu, Z.; Yang, Z.; Gu, Y.; Liu, H.; Wang, P. The effectiveness of eye tracking in the diagnosis of cognitive disorders: A systematic review and meta-analysis. *PLoS ONE* 2021, *16*, e0254059. [CrossRef]
- 135. Cummings, J.L.; Morstorf, T.; Zhong, K. Alzheimer's disease drug-development pipeline: Few candidates, frequent failures. *Alzheimers Res. Ther.* **2014**, *6*, 37. [CrossRef]
- 136. Yang, P.; Sun, F. Aducanumab: The first targeted Alzheimer's therapy. Drug Discov. Ther. 2021, 15, 166–168. [CrossRef]
- 137. Cummings, J.; Aisen, P.; Apostolova, L.G.; Atri, A.; Salloway, S.; Weiner, M.J. Aducanumab: Appropriate Use Recommendations. *Prev. Alzheimers Dis.* **2021**, *8*, 398–410. [CrossRef]
- 138. Fillit, H.; Green, A. Aducanumab and the FDA—Where are we now? Nat. Rev. Neurol. 2021, 17, 129–130. [CrossRef] [PubMed]

- 139. Rabinovici, G.D. Controversy and Progress in Alzheimer's Disease—FDA Approval of Aducanumab. *N. Engl. J. Med.* **2021**, *385*, 771–774. [CrossRef] [PubMed]
- 140. Baldari, S.; Di Rocco, G.; Toietta, G. Current Biomedical Use of Copper Chelation Therapy. Int. J. Mol. Sci. 2020, 21, 1069. [CrossRef]
- 141. Montes, S.; Rivera-Mancia, S.; Diaz-Ruiz, A.; Tristan-Lopez, L.; Rios, C. Copper and Copper Proteins in Parkinson's Disease. Oxidative Med. Cell. Longev. 2014, 2014, 147251. [CrossRef]
- 142. Pal, A.; Kumar, A.; Prasad, R. Predictive association of copper metabolism proteins with Alzheimer's disease and Parkinson's disease: A preliminary perspective. *BioMetals* 2014, 27, 25–31. [CrossRef] [PubMed]
- 143. Mezzaroba, L.; Alfieri, D.F.; Simão, A.N.C.; Reiche, E.M.V. The role of zinc, copper, manganese and iron in neurodegenerative diseases. *Neuro Toxicol.* 2019, 74, 230–241. [CrossRef]
- 144. Tosato, M.; Marco, V.D. Metal Chelation Therapy and Parkinson's Disease: A Critical Review on the Thermodynamics of Complex Formation between Relevant Metal Ions and Promising or Established Drugs. *Biomolecules* **2019**, *9*, 269. [CrossRef]
- 145. Huang, L.; Zhao, J.; Chen, Y.; Ma, F.; Huang, G.; Li, W. Baseline folic acid status affects the effectiveness of folic acid supplements in cognitively relevant outcomes in older adults: A systematic review. *Aging Ment. Health* **2021**, *26*, 457–463. [CrossRef]