Research Article

# Ethnobotanical Study of Medicinal Plants Used by Traditional Healers to Treat Cancer-Like Symptoms in Eleven Districts, Ethiopia

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There is no ethnobotanical study conducted specifically on medicinal plants traditionally used to treat cancer in Ethiopia. Yet, traditional herbalists in different parts of the country claim that they have been treating cancer-like symptoms using herbal remedies. The objective of this study was to document medicinal plants traditionally used to treat cancer-like symptoms in eleven districts, Ethiopia. Traditional herbalists were interviewed using semistructured questionnaires, and field visits were also carried out to collect claimed plants for identification purpose. Seventy-four traditional herbalists, who claimed that they knew about and/ or had used medicinal plants to treat cancer-like symptoms, were selected using the snowball method and interviewed. Herbalists used their intuition and relied on the chronicity, growth of external mass, and spreading of the disease to other parts of the body, as a means to characterize cancer symptoms. Furthermore, in some of the study districts, herbalists reported that they treat patients who had already been diagnosed in modern healthcare institutions prior to seeking help from them. The inventory of medicinal plants is summarized in a synoptic table, which contains the scientific and vernacular names of the plants, their geographical location, the parts of the plants, and the methods used to prepare the remedies. A total of 53 traditionally used anticancer plants, belonging to 30 families, were identified during the survey. The most frequently reported anticancer plants were Acmella caulirhiza Del (Asteraceae), Clematis simensis Fresen. (Ranunculaceae), Croton macrostachyus Del. (Euphorbiaceae), and Dorstenia barnimiana Schweinf. (Moraceae). Organizing traditional healers, documenting their indigenous knowledge, and scientifically validating it for the development of better cancer therapeutic agents constitute an urgent and important task for policymakers and scientists.

# 1. Introduction

Cancer is a complex disease that is very heterogenic and variable at cellular level and also differs from one patient to the other in its behaviour, development, and outcome [1]. Physical, metabolic, and behavioural variations of cancer cells from normal ones arise through the accumulation of genetic modifications and help them to proliferate rapidly, escape from host immune surveillance, and ultimately invade distant tissues [2]. Histopathological, genetic, and epigenetic and clinical outcome variations between and within different types of cancers have been the greatest challenge to understand the disease and develop novel therapies [3].

Surgery and radiation therapy were the most preferred means of treatment to control cancer before 1950 and after 1960, respectively [4]. Chemotherapy can be done before surgery to shrink the tumor or after surgery to kill the remaining cancer cells [5]. However, most of the chemotherapeutic drugs lack specificity and tend to rapidly damage normal dividing tissues, causing side effects such as immunosuppression, neurotoxicity, and hair loss [6]. Moreover, resistance has also reduced therapeutic efficacy of some anticancer chemotherapeutic drugs [7]. In order to address these limitations, tapping nature as a major source of chemically diverse novel anticancer compounds is a consistently proven track [8]. Screening natural products yield more hit with more "drug-like" characteristics (absorption and metabolism) as compared to screening of rationally designed compounds [9]. Furthermore, screening medicinal plants based on traditional use provides a higher chance of finding active plants relative to the random approach [10].

Ethiopia has a rich and diverse heritage of traditional medical practices, known for using plants to prepare more than 90% of the remedies [11]. In addition, the country has more than 6,500 higher plant species of which, around 12% are endemic [12]. Reports indicate that up to 80% of the population relies on traditional remedies as a primary source of health care [13]. Only few ethnobotanical reports from different agroecological zones of Ethiopia are available in the literature regarding medicinal plants used for cancer treatment. These include Bersama abyssinica, Buddleja polystachya, Clerodendrum myricoides, Dovyalis abyssinica, Ekebergia capensis, Myrsine melanophloeos, Olea capensis, Pentas lanceolata, Sideroxylon oxyacanthum, and Zingiber officinale [14]; Bidens macroptera, Clematis simensis, Ferula communis, and Punica granatum [15]; Rumex abyssinicus [16]; Zanthoxylum chalybeum [17]; Phytolacca dodecandra and Vinca rosea [18]; Kalanchoe lanceolata, Stephania abyssinica, and Vernonia hymenolepis [19]; Plumbago zeylanica [20–22]; Acalypha acrogyna, Carissa spinarum, Maytenus ovatus, and Salvia nilotica [23]; Croton macrostachyus [24]; and Dorstenia barnimiana [25, 26].

In view of this fact and considering the weak traditional recording and knowledge transfer system and an alarming rate of environmental degradation, finding anticancer plants and documenting their ethnobotanical information constitute an urgent and indispensable task. Therefore, the main aim of this study was to establish an inventory of medicinal plants traditionally used to treat cancer in eleven districts of Ethiopia.

### 2. Materials and Methods

2.1. Description of the Study Areas. This ethnobotanical study was conducted in four national regional states of Ethiopia: Oromia, Amhara, Afar, and Southern Nations, Nationalities, and People. The survey included different districts from each region, namely, Bale Robe and Goba from Oromia, Bahir Dar Zuria and Filiklik from Amhara, Gewane from Afar, and Wondo Genet, Sodo Zuria, Doyo Gena, North Bench, Mizan Aman, and Shako from Southern Nations, Nationalities, and People Regional State (Figure 1). These geographically, culturally, and agroecologically different study areas (Table 1) were selected mainly based on the availability of traditional healers and recommendations from health workers.

2.2. Data Collection. A team comprising a botanist and researchers from Addis Ababa University was set up, and health authorities were contacted for permission and

identification of traditional herbalists living in each study area. Altogether, 117 traditional healers were approached using the snowball technique and 74 traditional healers who used herbs to manage cancer-like symptoms were selected. Ethnobotanical data were collected between January and August 2016, mainly through individual interviews with the selected traditional herbalists using a semistructured interview questionnaire. The questionnaire was prepared in Amharic language and translated to different local languages for traditional healers who do not speak Amharic. This questionnaire was designed to obtain information in the following areas: (i) general data on the informant, (ii) school attendance, (iii) use of plants for cancer treatment, (iv) source of the plant material, (v) part of the plant used, (vi) method of medicinal preparation, (vii) route of administration, and (viii) side effects.

A traditional healer for the purpose of this study is "a person who is recognized by the community in which s/he lives as competent to provide healthcare by using plants and plant products." Each traditional healer was approached, briefed about the purpose of the research, and asked for his/ her verbal consent in talking about cancer and its treatment. They were assured of the confidentiality of the information they provided. If plants were mentioned for their anticancer purposes, a botanical sample was collected. These specimens were pressed and preserved for later identification at the National Herbarium, Addis Ababa University, Addis Ababa, and a voucher specimen of each plant was deposited in the institute. All botanical names have been transcribed according to the nomenclature system used by the Plant List (http://www.theplantlist.org).

2.3. Data Analysis. The relative importance of medicinal plants used in the management of cancer-like symptoms in study areas was assessed using the relative frequency of citation (RFC), use value (UV), informants consensus factor (ICF), and cultural importance index (CI).

2.3.1. Relative Frequency of Citation (RFC). The RFC was calculated by dividing the number of informants that cite a particular plant species (FC) by the total number of informants in the survey (N) [29]:

$$RFC = \frac{FC}{N}.$$
 (1)

2.3.2. Use Value (UV). The use value demonstrates the relative importance of plant species to treat particular ailment, and it is determined by the following formula [30]:

$$UV = \sum \frac{U_i}{N_i},$$
 (2)

where "UV" stands for the use value of a species, " $U_i$ " stands for the number of use reports cited by informants for that plant species, and " $N_i$ " is the total number of informers who reported the particular plant species *i*.

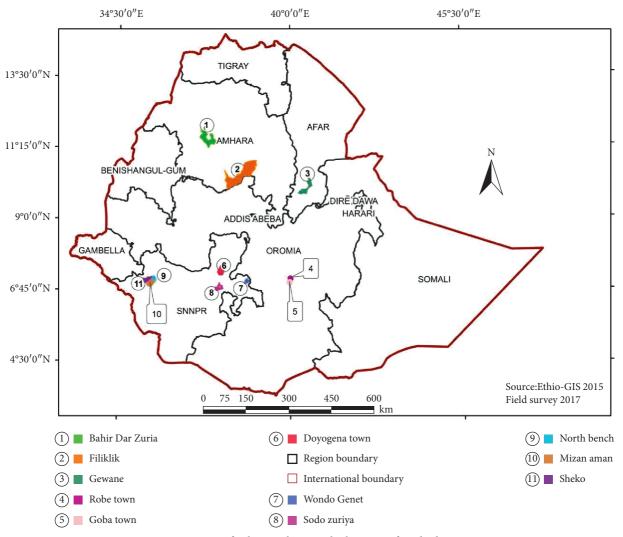


FIGURE 1: Map of Ethiopia showing the location of study districts.

2.3.3. Informant Consensus Factor (ICF). Informant consensus factor (ICF) was calculated to determine the homogeneity of the information collected about particular plant species to treat specific ailment. It was estimated using the following formula [31]:

$$ICF = \frac{Nur - Nt}{Nur - 1},$$
(3)

where Nur is the number of use reports of informants for particular ailment category and Nt refers to the number of species used for the ailment category by all informants.

2.3.4. Cultural Importance Index (CI). Cultural importance index (CI) is calculated by the sum of the use reports (UR) of informants mentioning each species use (from  $i_1$  to  $i_N$ ) in each use category and adding all the UR of each category (from  $u_1$  to  $u_{\rm NC}$ ) divided by the total number of informants N. This index is determined by the following formula [29]:

$$\operatorname{CI}_{i}\sum_{u=u_{1}}^{u_{\mathrm{NC}}}\sum_{i=i_{1}}^{i_{N}}\frac{\mathrm{UR}u_{i}}{N},$$
(4)

where CI is an ethnobotanical index that indicates the spread of the use along with the diversity of uses of each species.

# 3. Results

The informants consisted of 66 male and 8 female traditional healers and they were divided into three age groups: 20-40, 41-60, and  $\geq$  61 years. Out of 74 interviewed traditional healers, most of them (N%) were adults aged between 41 and 60 years. Majority of the respondents (70.2%) gained their knowledge from family members and 82% of all interviewed respondents practiced ethnomedicine for more than 25 years. More than 70% of the respondents were either only at their primary level of education or did not have a formal education at all (Figure 2). Traditional healers usually used their intuition and relied on the chronicity and growth of external mass, as a means to diagnose cancer. Lumpy growth was the most commonly cited criteria used to diagnose cancer, followed by ulcerative wounds and bleeding (Table 2). However, there were instances where some of the healers claimed to have treated patients already diagnosed with cancer at modern health institutions. Traditional 4

TABLE 1: Vegetation type, climatic condition, and demographic data of the study areas [27, 28] (source: National Meteorological Service Agency of Ethiopia).

	Distance					Average			c condition 2014)
District	from capital city (km)	Approximate population (2015)	Number of interviewed healers	Area size (km <sup>2</sup> )	Geographical location	elevation above sea level (m.a.s.l)	Vegetation type	Annual average rainfall (mm)	Annual average temperature range (°C)
Bale Robe	432	65,284	2	8.87	7°07′11.65″ N 40°00′24.82″ E	2480	DAF	745.6	9.2-23.2
Goba	444	47,135	7	20.15	7°00′41.66″ N 39°58′33.96″ E	2614	DAF	736.3	9.5-23.8
Bahir Dar Zuria	578	206,708	16	1443.37	11°34′27.15″ N 37°21′40.87″ E	1800	CTW, DAF, and FLV/ MFS	1547.1	12.7–27.6
Filiklik	188	142,722	7	806.98	10°02′12.74″ N 38°14′27.65″ E 10°29′59.99″	1853	CTW and DAF	880.2	12.9–22.0
Gewane	344	39,186	6	967.85	N 40°44′59.99″ E	568	ACB	586.7	19.5–36.7
Wondo Genet	270	196,277	12	226.45	7°05′3.01″ N 38°37′8.02″ E	1742	DAF	928.7	15-29.6
Sodo Zuria	383	145,092	2	25.62	6°51′10.11″ N 37°45′39.49″ E	1854	CTW and DAF	1569.2	14.8-25.2
Doyo Gena	258	95,393	14	130.57	7°21′20.22″ N 37°47′07.15″ E	2300	DAF	1334.5	11-22.8
North Bench	587	126,308	4	392.65	6°37′53.43″ N 35°33′56.83″ E	2367	CTW	1671.8	16-33.3
Mizan Aman	565	64,996	3	24.45	6°59′37.13″ N 35°34′55.92″ E	1441	CTW and MAF	1963.7	14.8-28.8
Shako	617	51,195	1	48,089.63 ha	7°33′42.37″ N 35°39′11.83″ E	1800	CTW and MAF	1906.9	11.4-22.4

*Note.* Vegetation type: DAF: dry evergreen Afromontane forest and grassland complex; CTW: Combretum-Terminalia woodland and wooded grassland; FLV/MFS: freshwater marshes and swamps, floodplains, and lake shore vegetation; ACB: Acacia-Commiphora woodland and bushland proper; MAF: moist evergreen Afromontane forest. m.a.s.l: meter above sea level; mm: millimeter; °C: degree Celsius; km<sup>2</sup>: kilometer square.

healers identified cancer as "*Nekersa*" in Bahir Dar Zuria and Filiklik, "*Naqarsa*" in Bale Robe and Goba, "*Sissac*" in Gewane, "*Xoka* or *Toka*" in Doyo Gena, "*Balamo*" in Wondo Genet, "*Kums* or *niamt*" in North Bench, and "*Kanser*" in Sheko and Sodo Zuria district. Out of the 6 specific cancer types (skin, breast, lung, cervical, throat, and intestinal) claimed to be treated by the respondents, skin cancer was a dominant one followed by breast cancer.

A total of 53 plant species belonging to 30 families were reported for their anticancer use (Table 3). The result of this study showed that shrubs (49.1%), herbs (33.9%), trees (13.2%), and climbers (3.8%) were the main sources of potential anticancer medicinal plants. This study also indicated that leaves (56.7%) were the most commonly used plant parts followed by roots (21.7%), bark (6.7%), stem (1.7%), seeds (1.7%), whole plant (1.7%), leaves and roots (5%), leaves or stem (1.7%), and leaves or seeds (1.7%) (Figure 3). Most of the reported plants occurred naturally in wild (96.2%); however, cultivation was also a source (3.8%). Reported medicinal plants have been traditionally claimed to be used to treat different types of ailments including cancer. However, only few have been scientifically investigated for their antiproliferative or cytotoxic activity (Table 4). While comparing the amount and distribution of anticancer plants in the past ten years,

regardless of the study areas, all respondents believed that the amount and distribution of these plants are reduced.

In the current study, the highest UVs were recorded for Aloe spp. (6), Albizia schimperiana (4), Sida schimperiana (4), Achyranthes aspera (4), Brucea antidysenterica (4), Cleome brachycarpa (3), Leonotis ocymifolia (3), and Prunus africana (3). The lowest UVs were obtained for Acokanthera schimperi, Acmella caulirhiza, Cineraria abyssinica, and Gnidia involucrata (Table 3). A total of 228 use reports have been documented and categorized into seven categories (Table 5). Among these, other ailments (46.3%) and skin cancer (26.5%) had the highest use reports. Furthermore, ICF values were also calculated and ranged from 0 to 0.42. The highest ICF values were recorded in other ailments (0.42) and breast cancer (0.32) followed by skin cancer (0.23) category (Table 5). The other ailments category comprises of diseases such as stomach ache, malaria, wart, swelling, wounds, evil eye, toothache, bleeding, gastrointestinal disorder, headache, bone fracture, cough, snake bite, herpes simplex, tonsillitis, hypertension, dandruff, fever, and hemorrhoid. The ICF value of the remaining four categories (lung cancer, colon cancer, cervical cancer, and throat cancer) was zero. Quantitative ethnobotanical indexes such as RFC and CI were calculated in this study to

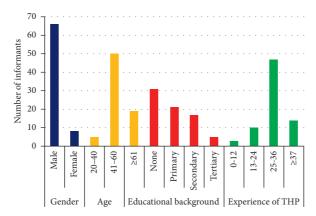


FIGURE 2: Demographic details of the interviewed informants.

TABLE 2: Symptoms that	t are used by traditional	healers to diagnose cancer.

Cancer types	Reported symptoms	Number of traditional healers
	Lumpy growth	32
Skin	Spreading pea-sized growth	1
	Ulcerative growth and oozing blood	1
	Lumpy growth	17
	Lumpy growth on one breast and progressive weight loss	1
Breast	Ulcerative wounds on breast	5
	Ulcerative wounds on breast and swelling on armpit and neck	1
	The patient was receiving anticancer treatment for breast cancer in hospital	12
Cervical	Foul-smelling bloody vaginal discharge, pain during sexual intercourse, and weight loss	1
Colon	Chronic rectal bleeding and weight loss	1
Lung	Coughing up blood	1
Throat	Coughing and swelling on the neck	1

analyze the ethnobotanical information. According to RFC values, *Croton macrostachyus* (0.1), *Vernonia auriculifera* (0.04), *Clematis simensis* (0.04), and *Acmella caulirhiza* (0.04) are the most frequently cited among all reported plants. *Croton macrostachyus* (0.16), *Dorstenia barnimiana* (0.12), and *Aloe* spp. (0.08) rank 1<sup>st</sup>, 2<sup>nd</sup>, and 3<sup>rd</sup> in position, respectively, according to the CI reference. Our result also shows that the Pearson correlation coefficient of RFC was positively and negatively correlated to CI and UV, respectively (Table 6).

Most of the reported remedies, prepared from these plants, were either applied topically (50%) or taken orally (41.7%). The remaining remedies were prepared to be administered either topically or orally (3.3%), both topically and orally (1.7%), and intranasally (1.7%). Usually, fresh plants were finely chopped, dried, and pounded to powder form. Then, the powder of either one or the combination of more than one plant was either mixed with drinking water or pasted and applied topically. In other cases, fresh plant parts were decocted and taken orally or crushed and applied topically. Water was the main medium in preparation of most remedies and additives like honey, milk, and butter were also used. To determine the amount of plant parts used to prepare remedies, traditional healers used spoon, fingertip, and number (in case of fresh leaves). Adverse effects reported by respondents include vomiting, diarrhea, and skin ulcers.

## 4. Discussion

Despite the rich biodiversity of the study areas, broad acceptability, and centuries-old tradition of using traditional medicines, the number of anticancer plants reported in this study is far less than expected. As it was reported by different ethnobotanical studies conducted in different parts of Ethiopia, this could be attributed to the attitude of many traditional healers to guard their indigenous medical knowledge as a family secret and hence hesitant to share with the researchers [13, 32, 73]. Justifying the lower number of female traditional healers (8, 11%) participated in this study, these studies also inferred that traditional healers usually pass their knowledge to the first son of the family.

In this study, in agreement with the studies conducted in Fiche district [35], Ghimbi district [20], and Hawassa city [17] of Ethiopia, the predominant botanical families recorded, listing over 5 plant species each, were Asteraceae, Fabaceae, and Lamiaceae. This could be due to the fact that these families are the largest in the flora of Ethiopia and Eritrea [15, 21, 143]. Moreover, cytotoxicity studies conducted on different Mexican plants reported that the highest number of plant species with both *in vitro* and *in vivo* antineoplasic activities was from these families [20].

The highest UVs recorded in this study include *Aloe* spp. (6), *Achyranthes aspera* L. (4), *Albizia schimperiana* (4), *Sida schimperiana* (4), and *Brucea antidysenterica* (4). The

		TABLE 3: 1	List of candi	idate medic	unal plants	traditional	TABLE 3: List of candidate medicinal plants traditionally used for cancer treatment in the study areas.	areas.				
Voucher number	Botanical name (family)	Vernacular name	Districts	Growth form	Habitat	Parts used	Preparation	Type of cancer treated	Application	UV	RFC (	CI
	Acanthaceae											
Bele-047	Justicia schimperiana (Hochst. ex Nees) T. Anderson	Kitkit	North Bench	Shrub	Wild	Roots	Fresh roots are crashed and boiled, and the cool decoction is drunk before meal	Lung	Oral	1	0.027 0.	0.067
Bele-057	Justicia schimperiana (Hochst. ex Nees) T. Anderson	Gulbana	Doyo Gena	Shrub	Wild	Leaves	Fresh leaves are pounded, and the juice is applied on the affected area	Skin	Topical			
	Aloaceae											
Bele-060	Aloe sp.	Gurta waqota	Doyo Gena	Shrub	Wild	Leaves	Fresh roots are crashed, and the sap is applied on the affected area	Skin	Topical	6	0.014 0.	0.081
	Amaranthaceae											
Bele-044	Achyranthes aspera L.	Koch ashite	Mizan Aman	Herb	Wild	Leaves	Leaves are roasted on metal plate, pounded into powder, mixed with animal butter, and smeared on the affected part	Skin	Topical	4	0.014 0.	0.054
	Apiaceae											
Bel-046	<i>Centella asiatica</i> (L.) Urb.	Gorongoch	Sheko	Herb	Wild	Leaves	Young leaves are crashed, and the sap sniffed	Throat	Intranasal	2	0.014 0.	0.027
Bel-002	Hydrocotyle mannii Hook.f	Ye'ti medhanit	North Bench	Herb	Wild	Leaves	Young leaves are crashed and applied on the affected area	Skin	Topical	1	0.014 0.	0.014
	Apocynaceae											
Bel-003	Acokanthera schimperi (A.DC.) Schweinf.	Merenz	Bahir Dar Zuria	Shrub	Wild	Leaves	Young leaves are crashed and applied	Skin	Topical	0.5	0.027 0.	0.027
Bel-009	Carissa spinarum L.	Agam	Bahir Dar Zuria	Shrub	Wild	Leaves	Leaves are crashed and infused in cold water overnight and drunk before meal and applied on the affected area	Skin	Oral	1	0.027 0.	0.027
	Asclepiadaceae											
Bel-040	Calotropis procera (Aiton) Dryand.	Qumbo	Gewane	Shrub	Wild	Roots	Fresh roots are crashed, and the sap is applied on the affected area	Breast	Topical	б	0.014 0.	0.027
Bel-036	Pentarrhinum insipidum E. Mey.	Barohula	Gewane	Shrub	Wild	Roots	Fresh roots are crashed, and the sap is applied on the affected area	Breast and skin	Topical	1	0.014 0.	0.014
Bel-037	dammanniana Sprenger	Mureli	Gewane	Herb	Wild	Stem	Stems are cut, and the sap is applied	Skin	Topical	5	0.014 0.	0.027
	Asphodelaceae											
Bel-020	<i>Kniphofia foliosa</i> Hochst. Asteraceae	Shushube	Bale Goba	Shrub	Wild	Roots	Dry roots are pounded, and the powder is mixed with honey	Cervical and breast	Oral	1	0.027 0.	0.027

	CI	0.054	0.027	0.014	0.014	0.054	0.014	0.027	0.081			0.014		0.027		0.041		0.16		
	RFC	0.04	0.014	0.014	0.014	0.027	0.014	0.014	0.041			0.014		0.014		0.027		0.1		
	UV	0.67	7	1	1	1.5	1	2	1.33			3		2		1.5		0.75		
	Application	Topical	Oral Oral	Oral	Oral	Topical	Topical	Topical	Topical	Topical		Topical		Topical		Topical		Topical	Topical	Oral
	Type of cancer treated	Breast	Breast Breast	Breast	Breast	Skin	Skin	Skin	Skin	Skin		Breast and skin		Skin		Breast and skin		Breast and skin	Skin	Breast and skin
itinued.	Preparation	Young leaves are chewed by the healer and spit on Fresh leaves are crashed and infused in	cold water Dried leaves are ground and morensted in coffee or tea	Dried leaves are ground and macerated in coffee or tea	Dried leaves will be ground and decocted in hot water	Fresh leaves are pounded, and the sap is applied on the affected area	Fresh leaves are pounded, and the sap is applied on the affected area	Fresh leaves are pounded and the sap is applied on the affected area	Fresh leaves are pounded, and the sap is applied on the affected area	Fresh leaves are chewed by the healer and spit on		Fresh leaves are pounded, and the sap is applied on the affected area		Fresh roots are pounded, and the sap is applied on the affected area		Fresh leaves are roasted for 2 minutes and applied on the affected area		Fresh leaves or succulent stems are crashed, and the sap is applied on the affected area	Dry bark is pounded, and the powder is applied on the affected area	Fresh leaves are crashed, macerated in cold water, and drunk
TABLE 3: Continued	Parts used	Leaves	Leaves Leaves	Leaves	Leaves	Leaves	Leaves	Leaves	Leaves	Leaves		Leaves		Roots		Leaves		Leaves or stem	Bark	Leaves
TABI	Habitat	Wild	Wild Wild	Wild	Cultivated	Wild	Wild	Wild	Wild	Wild		Wild		Cultivated		Wild		Wild	Wild	Wild
	Growth form	Shrub	Shrub Herb	Herb	Tree	Herb	Shrub	Shrub	Shrub	Shrub		Herb		Herb		Shrub		Tree	Tree	Tree
	Districts	Mizan Aman Wondo	Genet Sodo Zuria	Doyo Gena	Sodo Zuria	Bale Robe	Doyo Gena	Doyo Gena	Doyo Gena	Wondo Genet		Gewane		Doyo Gena		Bale Goba		Filiklik	Doyo Gena	Wondo Genet
	Vernacular name	Kust asht	Bitisa Natrara	Agufa	Artemisia	Unknown	Sheshota	Arbaba	Barawa	Reji		Berbere		Laluncha		Anchura		Bisana	Besena	Masichoo
	Botanical name (family)	Acmella caulirhiza Delile Acmella caulirhiza	Delile Artemisia absinthium I	Artemisia afra Jacq. ex Willd.	Artemisia annua L.	Cineraria abyssinica Sch.Bip. ex A.Rich.	Guizotia scabra (Vis.) Chiov.	Solanecio gigas (Vatke) C. Jeffrey	<i>Vernonia auriculifera</i> Hiern	Vernonia auriculifera Hiern	Capparidaceae	<i>Cleome brachycarpa</i> (Forssk.) Vahl ex DC.	Commelinaceae	Commelina benghalensis L.	Crassulaceae	Kalanchoe petitiana A. Rich.	Euphorbiaceae	<i>Croton macrostachyus</i> Hochst. ex Delile	Croton macrostachyus Hochst. ex Delile	Croton macrostachyus Hochst. ex Delile
	Voucher number	Bel-045	Bel-049 Bel-030	Bel-029	Bel-031	Bel-021	Bel-058	Bel-034	Bel-025	Bel-056		Bel-039		Bel-026		Bel-019		Bel-012	Bel-035	Bel-048

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	Botanical name (family)	Vernacular name	Districts	Growth form	Habitat	Parts used	Preparation	Type of cancer treated	Application	UV	RFC	CI
Š	Euphorbia schimperiana Scheele	Gendalelata	Doyo Gena	Shrub	Wild	Roots	Fresh roots are pounded, and the sap is applied on the affected area	Skin	Topical	1	0.014 (	0.014
	Fabaceae											
1	Albizia schimperiana Oliv.	Sessa	Filiklik	Tree	Wild	Leaves	The mixture of fresh leaves of <i>Albizia</i> schimperiana and <i>Carissa spinarum</i> is macerated in cold water for 2 days, and the macerated liquid is drunk	Breast, intestinal, and skin	Oral	4	0.014 (	0.014
	Calpurnia aurea (Aiton) Benth.	Digita	Bahir Dar Zuria	Shrub	Wild	Leaves or seeds	Dry leaves or seeds are ground, macerated in cold water, and drunk	Breast	Oral	5	0.014 0.027	0.027
č	Crotalaria agatiflora Schweinf.	Unknown	Bale Goba	Shrub	Wild	Seeds	Dry seeds are ground, mixed with honey, and applied	Skin	Topical	1	0.014 (	0.014
-	Crotalaria incana L.	Chelke	Doyo Gena	Shrub	Wild	Leaves	Fresh leaves are crashed, and the sap is applied on the affected area	Skin	Topical	1	0.014 (	0.014
	<i>Senna singueana</i> (Delile) Lock	Gefa	Bahir Dar Zuria	Shrub	Wild	Leaves	Fresh leaves are crashed, macerated, and drunk	Skin	Oral	7	0.014 (	0.027
	Lamiaceae											
	Ajuga leucantha Lukhoba	Tiks asht	North Bench	Herb	Wild	Leaves	Fresh leaves are crushed, and the sap is applied on the affected area	Breast	Topical	Ч	0.014 (	0.014
	<i>Leonotis ocymifolia</i> (Burm.f.) Iwarsson	Armagusa	Bale Goba	Herb	Wild	Leaves	Fresh leaves are crashed, macerated overnight, and drunk	Breast and skin	Oral	$\tilde{\mathbf{\omega}}$	0.014 (	0.014
$\cup$	Ocimum gratissimum L.	Mekedesisa	Wondo Genet	Herb	Wild	Roots	Fresh roots are crushed, boiled, and drunk	Skin	Oral	7	0.014 (	0.027
	Pycnostachys abyssinica Fresen.	Tontona	Doyo Gena	Herb	Wild	Leaves	Fresh leaves are crushed, and the sap is applied on the affected area	Skin	Topical	2	0.014 (	0.027
S	Salvia nilotica Juss. ex Jacq.	Barnbanch	North Bench	Shrub	Wild	Whole plant	Dry plant parts are ground, mixed with honey, and applied	Breast	Topical	2	0.014 (	0.027
	Thymus schimperi Ronniger	Tosigne	Bale Goba	Herb	Wild	Leaves	Dry leaves are decocted and drunk	Breast	Oral	5	0.014 (	0.027
	Malvaceae											
	Sida schimperiana Hochst. ex A. Rich.	Kotijebessa	Wondo Genet	Shrub	Wild	Roots and leaves	Fresh leaves and roots are crashed, macerated, and drunk	Breast and skin	Oral	4	0.014 (	0.027
	Melianthaceae											
	Bersama abyssinica Fresen.	Azamir	Bahir Dar Zuria	Shrub	Wild	Bark	Dry bark is ground, macerated, and drunk before meal	Breast	Oral	-	0.014 (	0.014

	CI		0.12		0.027		0.027		0.041	0.041			0.054		0.014	0.027		0.027
	RFC		0.068		0.014 0.027		0.014 0.027		0.014	0.027			0.041		0.014	0.014		0.027
	UV		0.6		7		2		3	1.5			0.67		3	2		2
	Application		Oral or topical		Oral		Topical		Oral	Oral	Topical		Topical		Oral	Oral		Oral
	Type of cancer treated		Breast		Skin		Breast		Skin	Colon	Skin		Breast		Breast and skin	Breast		Breast
ntinued.	Preparation		Dry roots are ground, mixed with water and honey, and drunk, or dry roots are ground, mixed with honey, and applied on the affected area		Dry leaves and roots of <i>Syzygium</i> <i>guineense</i> and dry leaves of <i>Osyris</i> <i>quadripartita</i> are ground, mixed, decocted, and drunk		Fresh leaves and roots are crashed and applied with a bandage		Dry roots are ground, macerated, and drunk	Dry roots are ground and taken with food	Fresh bark is crashed and squeezed, and the sap is applied		Fresh roots of <i>Dorstenia barnimiana</i> mixed with fresh leaves of <i>Clematis</i> <i>simensis</i> , pounded, and applied		Dry bark is ground, decocted, and drunk	Dry leaves are ground, mixed with honey, and eaten		Dry leaves are ground, decocted, and drunk
TABLE 3: Continued.	Parts used		Roots		Leaves and roots		Leaves and roots		Roots	Roots	Bark		Leaves		Bark	Leaves		Leaves
TAI	Habitat		Wild		Wild		Wild		Wild	Wild	Wild		Wild		Wild	Wild		Wild
	Growth form		Herb		Tree		Herb		Shrub	Herb	Herb		Climber		Tree	Shrub		Shrub
	Districts		Bahir Dar Zuria		Bahir Dar Zuria		Wondo Genet		Filiklik	Doyo Gena	Wondo Genet		Bahir Dar Zuria		Bahir Dar Zuria	Filiklik		Filiklik
	Vernacular name		Work Bemeda		Dokima		Qinta		Emboacho	Goecho	Sharibicho		Yeazo Hareg		Tikur enchet	Limich		Keret
	Botanical name (family)	Moraceae	Dorstenia barnimiana Schweinf.	Myrtaceae	Syzygium guineense (Willd.) DC.	Oxalidaceae	Oxalis corniculata L.	Polygonaceae	Rumex nervosus Vahl	Rumex nepalensis Spreng.	Rumex nepalensis Spreng.	Ranunculaceae	<i>Clematis simensis</i> Fresen.	Rosaceae	Prunus africana (Hook.f.) Kalkman	Kutaceae Clausena anisata (Willd.) Hook.f. ex Benth.	Santalaceae	Osyris quadripartita Salzm. ex Decne.
	Voucher number		Bel-008		Bel-006		Bel-052		Bel-018	Bel-033	Bel-053		Bel-010		Bel-011	Bel-016		Bel-013

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Voucher Bo number s											
J	Botanical name (family)	Vernacular name	Districts	Growth form	Habitat	Parts used	Preparation	Type of cancer treated	Application UV RFC	UV	RFC CI
0	Sapindaceae										
Bel-005 Subs (L.	<i>Dodonaea viscosa</i> subsp. <i>angustifolia</i> (L.f.) J.G.West	Kitkita	Bahir Dar Zuria	Tree	Wild	Roots	Dry roots are ground, mixed with honey, and applied or dry roots are ground, decocted, and drunk	Breast, skin and cervical	Topical or oral	1	0.014 0.041
Sir	Simaroubaceae										
Bel-017 Brucea	Brucea antidysenterica J.F.Mill.	Abalo	Filiklik	Tree	Wild	Leaves	Dry leaves are ground, pasted with cold water, and applied	Skin	Topical	4	0.014 0.054
	Solanaceae										
Bel-027 D	Discopodium penninervium Hochst.	Chechanga	Doyo Gena	Shrub	Wild	Leaves	Fresh leaves are crashed and applied	Skin	Topical	1	0.014 0.014
Th	Thymelaeaceae										
Bel-055 Grid Steu	<i>Gnidia involucrata</i> Steud. ex A.Rich.	Bito	Bahir Dar Zuria	Herb	Wild	Roots	Dry roots are ground, mixed with honey, and eaten	Breast	Oral	0.5	0.027 0.027
Λ	Verbenaceae										
Bel-050 Lant	Lantana trifolia L.	Hanshebello	Wondo Genet	Shrub	Wild	Leaves	Fresh leaves are ground, macerated in cold spring water, and drunk	Breast and skin	Oral	2	0.014 0.014
Bel-015 Lip	<i>Lippia adoensis</i> Hochst.	Kessie	Filiklik	Shrub	Wild	Leaves	Dry leaves are ground, macerated in cold water, and drunk	Skin	Oral	2	0.014 0.027
	Vitaceae										
C <i>ypho</i> Bel-038 (Hocl	<i>Cyphostemma serpens</i> (Hochst. ex A.Rich.) Desc.	Eiriti	Gewane	Climber	Wild	Roots	Dry roots are ground, pasted with honey and eaten, and applied	Skin	Oral and topical	1	0.014 0.014
UV = use value; RFC = relative frequency of citation; CI = cultural importance index.	= relative frequency	of citation; CI =	cultural imp	ortance inde	x.						

TABLE 3: Continued.

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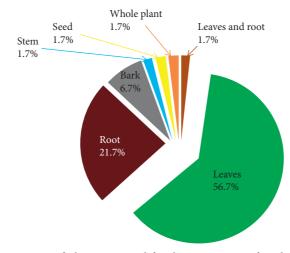


FIGURE 3: Frequency of plant parts used for the preparation of medicinal remedy.

T 10 6 6 1 1 1	1.1 . 1	11 ( 1 ( 1 ) 1	
TABLE 4: Cross-reference of cancer treatment	candidate plant species	collected from the stud	v areas with the published literature
indee in Groot reference of current freuthient	culture plant opecies	concerca nonn the staa	y areas with the published interature.

Botanical name (family)	Biological activity/chemical constituents	Illnesses/symptoms claimed to be treated traditionally
<i>Justicia schimperiana</i> (Hochst. ex Nees) T. Anderson (Acanthaceae)	Saponins, alkaloids, terpenoids and flavonoids [32] <i>In vitro</i> cytotoxicity [33]; <i>in vitro</i> antioxidant activity on DPPH assay [34]; <i>in vivo</i> suppression of parasitaemia on <i>Plasmodium berghei</i> -infected mice in the 4-day suppressive test [32]; and <i>in vivo</i> hepatoprotective activity in mice intoxicated with CCL <sub>4</sub> [35]	<ul> <li>Wound [15, 21]; rabies [15, 18–20, 36–39]; jaundice</li> <li>[15, 16, 21, 23, 38, 40]; gonorrhea [17, 36, 39]; liver cirrhosis [18, 26]; seizure [19, 41]; stomach ache</li> <li>[15, 25, 38]; helminths [15, 42, 43]; skin burn/lesion</li> <li>[23, 44]; arthritis [21, 23]; hepatitis [45, 46]; evil eye</li> <li>[15, 46]; dysentery [15, 21]; malaria [36, 39]; common cold, asthma, and headache [36, 39, 47]; otitis [48]; toothache [49]; and rheumatism [50]</li> </ul>
<i>Aloe</i> sp. (Aloaceae)	Anthrones and chromones [51], pyrones, coumarins, alkaloids, glycoproteins, naphthalenes, and flavonoids [52] 7-O-methylaloeresin showed <i>in vitro</i> antioxidant activity in DPPH assay [51], and methanol and ethanol extract showed <i>in vivo</i> parasitaemia suppression on <i>Plasmodium berghei</i> -infected mice in the 4-day suppressive test [53, 54]	<ul> <li>Wound [21, 55]; eye disease [21, 46, 48, 56]; snake bite</li> <li>[21, 48, 56]; malaria [20, 21, 44, 48, 54]; easing labour</li> <li>[44]; tropical ulcer, colon cleaner, and gallstone [48]; amoeba, abdominal pain, impotence, and urine</li> <li>retention [21]; dandruff [46, 56], hemorrhoids and</li> <li>hepatitis B [46]; ascariasis [21]; diabetes [54]; asthma</li> <li>[55]; foot strain [57, 58]; wart and anthrax [20];</li> <li>external injury [59]; and liver swelling, splenomegaly, and skin inflammation [56]</li> </ul>
<i>Achyranthes aspera</i> L. (Amaranthaceae)	Phytosteroids, polyphenols, and saponins [60] Methanol extracts have showed <i>in vivo</i> wound healing activity [61]	Bleeding [21, 24, 26, 62–64]; retained placenta [21, 62]; stomach ache and external swelling [17]; rhesus factor incompatibility in pregnancy [40, 55]; epistaxis [19]; hepatitis and evil eye [24]; tonsillitis [21, 57]; snake bite and paralysis [21]; dysentery [59]; herpes zoster [26]; anthrax [21, 49]; nasal infection and ophthalmic infection [64]; excessive menstruation and tape worm infection [15]; and gonorrhea [65]
<i>Centella asiatica</i> (L.) Urb. (Apiaceae)	Terpenoids (triterpenes, asiaticoside, centelloside, madecassoside, brahmoside, brahminoside (saponin glycosides), asiaticentoic acid, centellic acid, centoic acid, madecassic acid, terminolic acid, betulic acid, $\beta$ -caryophyllene, trans- $\beta$ -farnesene and germacrene D (sesquiterpenes), $\alpha$ -pinene, and $\beta$ -pinene [66, 67] Methanol extract inhibited the proliferation of human gastric adenocarcinoma (MK-1), human uterine carcinoma (HeLa), and murine melanoma (B16F10) cells <i>in vitro</i> [68]; aqueous extracts induced apoptosis in colonic crypts and exerted chemopreventive effect on colon tumorigenesis in male F344 rats [69]	Genital infection and lymphadenitis [63]; topical swelling [26, 70]; gastritis, headache, and evil eye [70]; bleeding [40]; wound [24]; abdominal ache [71]; meningitis [72]; and tinea corporis [47]
<i>Hydrocotyle mannii</i> Hook.f (Apiaceae)	No previous reports	Eye infection [63] and cataract [72]

	TABLE 4: Continued.				
Botanical name (family)	Biological activity/chemical constituents	Illnesses/symptoms claimed to be treated traditionally			
Acokanthera schimperi (A.DC.) Schweinf. (Apocynaceae)	In vitro cytotoxicity [73]; in vitro antiviral activity against coxsackie B3, influenza A, and herpes simplex type1 virus [74]; in vitro antimicrobial activity against Staphylococcus aureus, Pseudomonas aeruginosa, Trichophyton mentagrophytes [75]; and in vivo parasitaemia suppression in Plasmodium berghei-infected mice [76]	Wound [16, 44, 77, 78]; hepatitis [15, 16, 22, 44]; gonorrhea [19, 25]; evil eye [62]; bone fracture [24]; hemorrhoids [44]; scabies [21]; malaria and tonsillitis [48, 56]; psychiatric disease [55]; and skin diseases [65]			
Carissa spinarum L. (Apocynaceae)	<i>In vitro</i> antioxidant activity on DPPH assay and <i>in</i> antiproliferative activity [79]	Throat cancer [23, 80]; evil eye [16, 21, 24, 49, 62, 70, 72, 81]; snake bite [23, 80]; gonorrhea [20, 65]; stomach ache [20, 70]; impotence and headache [20]; tonsillitis [17, 56, 70]; wound and febrile illness [16]; bleeding after delivery [44]; muscle cramps [49]; toothache [47]; and premature ejaculation [56]			
<i>Calotropis procera</i> (Aiton) Dryand. (Asclepiadaceae)	Latex contains phytochemicals such as alkaloids, sterols, fatty acids, starches, sugars, oils, tannins, resins, and gums, and enzymatic proteins such as proteases, chitenases, lipases, peptidases, esterase, peroxidases, papain, hevein, and lectins [82] <i>In vivo</i> hepatoprotective [83]; hypoglycemic effect [84]; strong anti-implantation (antifertility) [85]; crude latex showed antioxidant and antiapoptotic activities against the toxicity of 4-nonylphenol [86]	Wound [16, 21, 81]; hemorrhoids [16, 19, 44]; wart [16, 57]; snake bite [23, 87]; kidney stone, tuberculosis, and scabies [16]; swelling [58]; skin rash [21, 49]; tinea capitis [21]			
<i>Pentarrhinum insipidum</i> E. Mey. (Asclepiadaceae)	No prev	vious reports			
<i>Echidnopsis dammanniana</i> Sprenger (Asclepiadaceae)	No previous reports	Snake bite [56]			
<i>Kniphofia foliosa</i> Hochst. (Asphodelaceae)	2-Acetyl-1-hydroxy-8-methoxy-3- methylnaphthalene, 10-(chrysophanol-7'-yl)-10- (ξ)-hydroxychrysophanol-9-anthrone, chryslandicin, knipholone, and chrysophanol [88] 10-(Chrysophanol-7'-yl)-10-(ξ)- hydroxychrysophanol-9-anthrone showed <i>in vitro</i> antiplasmodial activity against chloroquine- sensitive 3D7 strain of <i>Plasmodium falciparum</i> and knipholone selectively inhibited leukotriene metabolism in <i>in vitro</i> a human blood assay [88]; knipholone anthrone showed <i>in vitro</i> cytotoxicity [89] and antioxidant activity on DPPH assay [90]	No previous reports			
<i>Acmella caulirhiza</i> Delile (Asteraceae)	Unsaturated alkylamides like spilanthol and N- isobutylnona-2E,4E-dien-8ynamide [91] <i>In vitro</i> antiplasmodial activity [92]	Swelling [15]; tonsillitis [20, 63]; and toothache [40, 87]			
Artemisia absinthium L. (Asteraceae)	Camphor, davanone, ethyl (E)-cinnamate, (E)- nerolidol, and chamazulene [93] Essential oils showed <i>in vitro</i> antiparasitic effects	Hypertension, stomach ache, severe abdominal cramp [18] and sour throat [40]			
<i>Artemisia afra</i> Jacq. ex Willd. (Asteraceae)	Epoxylinalol and dihydrocostunolide [94]; camphor, davanone, bornyl acetate, 4-terpineol, and chamazulene [95] <i>In vitro</i> cytotoxicity on human leukaemia cell lines [73]; and <i>in vitro</i> antioxidant effect on DPPH assay [95]	Stomach ache [18, 42]; evil eye [16, 17, 62]; headache [42, 77]; eye disease, tinea capitis infection, hematuria, and stabbing pain [77]; antifertility agent [33]; malaria [42, 62]; ascariasis [18]; epilepsy and febrile illness [46, 65]			

TABLE 4: Continued.

	TABLE 4: Continued.	
Botanical name (family)	Biological activity/chemical constituents	Illnesses/symptoms claimed to be treated traditionally
<i>Artemisia annua</i> L. (Asteraceae)	In vitro inhibition of immune mediators of angiogenesis [96]; the sesquiterpene (Z)-7- acetoxy-methyl-11-methyl-3-methylene-dodeca- 1,6,10-triene showed moderate cytotoxic activities against the human tumor cell lines of HO8910 (ovary), 95-D (lung), QGY (liver), and HeLa (cervix) by MTT assay and induced apoptosis on 95-D tumor cells [97]; artemisinin and quercetagetin 6,7,3',4'-tetramethyl ether showed significant cytotoxicity against P-388, A-549, HT- 29, MCF-7, and KB tumor cells [98]	No previous reports
<i>Cineraria abyssinica</i> Sch.Bip. ex A.Rich. (Asteraceae)	<i>In vitro</i> radical scavenging activity on DPPH assay [99]; flavonoidal glycoside (rutin) showed <i>in vitro</i> antibacterial activity [100]	No previous reports
<i>Guizotia scabra</i> (Vis.) Chiov. (Asteraceae)	<i>In vitro</i> cytotoxicity on human leukaemia cell lines [73], and <i>in vitro</i> antiviral activity [101]	Wound [20]; epilepsy [40]; and ectoparasite infestation [47]
<i>Solanecio gigas</i> (Vatke) C. Jeffrey (Asteraceae)	<i>In vitro</i> antiviral activity against human immunodeficiency virus type 1 and type 2 cytotoxicity on human T-lymphocytic MT-4 cell lines [102]	Skin diseases [62]; retained placenta [40]; hepatitis [64]; evil eye [15]
<i>Vernonia auriculifera</i> Hiern (Asteraceae)	Tannins, flavonoids, terpenoids, and saponins [103]	Toothache [72]; snake bite [42]; skin cut [47]
<i>Cleome brachycarpa</i> (Forssk.) Vahl ex DC. (Capparidaceae)	No prev	vious reports
<i>Commelina benghalensis</i> L. (Commelinaceae)	Phlobatannins, carbohydrates, tannins, glycosides, volatile oils, resins, balsams, flavonoids, and saponins [104] Ethanol extract showed <i>in vivo</i> sedative and anxiolytic activity [105]	Helminths [65]; skin infection [72]
<i>Kalanchoe petitiana</i> A. Rich. (Crassulaceae)	Polyphenols, alkaloids, flavonoids, tannins, saponins, and steroids [106] In vitro antimicrobial activity against Escherichia coli, Pseudomonas aeruginosa, and Staphylococcus aureus [75]; and in vivo wound healing activity [106]	Breast and skin cancer [107]; swelling [40, 77]; tapeworm infection, trachoma, and syphilis [77]; lymphadenopathy and evil eye [22]; sore muscles [108]; itching skin [63]; and bone fracture [23]
<i>Croton macrostachyus</i> Hochst. ex Delile (Euphorbiaceae)	Ethanol extract showed <i>in vitro</i> antioxidant activity on DPPH assay [79]	Tumor, rabies, and wart [24]; skin cancer and wound [17]; gonorrhea [20, 23, 62]; headache [18, 109]; snake bite [18, 72]; malaria [16, 18–20, 110]; helminths [18, 111]; tinea nigra [40]; ringworm [17, 62]; tinea versicolor [16, 25]; heart failure [62]; bleeding [18, 24]; hepatitis [16, 18, 24]; stomach ache [16, 18, 23]; diarrhea [16, 18]; lymph adenitis and rheumatism [18]; bloat, scabies, and urine retention [16]; retained placenta and leprosy [19]
Euphorbia schimperiana Scheele (Euphorbiaceae)	<i>In vitro</i> cytotoxic effect against breast cancer (MCF7), hepatocellular carcinoma (HEPG2), and cervix cancer (HELA) cells [112]	Syphilis [108]
Albizia schimperiana Oliv. (Fabaceae)	<i>In vitro</i> cytotoxicity on human leukaemia cells [73]	Evil eye [20]; kidney infection and liver cirrhosis [18]

TABLE	4.	Continued.

TABLE 4: Continued.					
Botanical name (family)	Biological activity/chemical constituents	Illnesses/symptoms claimed to be treated traditionally			
<i>Calpurnia aurea</i> (Aiton) Benth. (Fabaceae)	$3\beta,4\alpha,13\alpha$ -Trihydroxylupanine, calpaurine, lupinine, and epilupinine calpurmenine and calpurmenine pyrrolecarboxylic acid ester, 13- hydroxylupanine, its tiglate and pyrrolecarboxylic acid esters (calpumine), virgiline and virgiline pyrrolecarboxylic acid ester [113]; $4\beta$ -hydroxy- $13\alpha$ -O-(2'-pyrrolylcarbonyl)-lupanine (digittine) and $4\beta,13\alpha$ -dihydroxylupanine [114]; alkaloids, tannins, flavonoids, and saponins [35] Methanol extract showed <i>in vitro</i> antimicrobial activity against <i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , and <i>Pseudomonas aeruginosa</i> [75] and type 1 and type 2 human immunodeficiency virus and showed cytotoxicity on human T-lymphocytic MT-4 cell lines [102]; methanol and dichloromethane crude extracts showed <i>in vitro</i> cytotoxicity on human leukaemia cells [73]; and ethanol extracts showed <i>in vitro</i> antioxidant activity on DPPH assay [79]	Tumor [22, 26, 80]; stomach ache [21, 62, 70, 81]; wound and skin infection [62]; Gonorrhoea and syphilis [16], amoebiasis [16, 80]; ascariasis and gastric ulcer [23]; diarrhea [21, 38, 70]; scabies and pubic hair louse [40]; diabetes mellitus and hypertension [19]; herpes zoster, hemorrhoids and tinea capitis [21]; and swelling and tuberculosis [58]			
Crotalaria agatiflora Schweinf. (Fabaceae) Crotalaria incana L. (Fabaceae)	Methanol and dichloromethane crude extracts showed <i>in vitro</i> cytotoxicity on human leukaemia cells [73] Dihydrosenecionine isomer, nemorensine isomer, integerrimine and anacrotine [115] Methanol and dichloromethane crude extracts showed <i>in vitro</i> cytotoxicity on human leukaemia cell lines [73]	No previous reports			
<i>Senna singueana</i> (Delile) Lock (Fabaceae)	Methanol extracts showed <i>in vitro</i> antioxidant activity on DPPH assay [116]	Stomach ache [58, 62, 70]; wound and swellings [62]; teeth infection and sprain [58]			
<i>Ajuga leucantha</i> Lukhoba (Lamiaceae)	No previous reports	Diarrhea [70]			
<i>Leonotis ocymifolia</i> (Burm.f.) Iwarsson (Lamiaceae)	Methanol and dichloromethane crude extracts showed <i>in vitro</i> cytotoxicity on human leukaemia cells [73]	Ascariasis [62], febrile illness [16, 62]; eye disease [16]; headache and neck ulcer [55]; and snake bite [15]			
Ocimum gratissimum L. (Lamiaceae)	Essential oil contains constitutes $\gamma$ -terpinene, $\beta$ -phellandrene, limonene, and thymol and showed <i>in vivo</i> antiplasmodial activity against <i>Plasmodium</i> <i>berghei</i> infection [117]	Allergy reaction [18, 20]; rheumatism, headache and eye disease [18]; febrile illness and general malaise [40]; sun stroke [24]; malaria [44]			
<i>Pycnostachys abyssinica</i> Fresen. (Lamiaceae)	No previous reports	Eye disease [18, 47]; ascariasis and wound [18]; diarrhea, stomach ache, amoebiasis, stomach bloating, and food poisoning [70]; headache [63]			
<i>Salvia nilotica</i> Juss. ex Jacq. (Lamiaceae)	Essential oil contains germacrene D, guaiol, and <i>trans</i> -caryophyllene as major constituents and showed activity against both Gram-positive and Gram-negative pathogenic bacteria; the oil also showed <i>in vitro</i> antioxidant activity on DPPH assay [118]	Tonsillitis and constipation [62]; herpes simplex [18, 38]; wound [40]; lymphadenitis [63]; and hemorrhoids and diarrhea [65]			
<i>Thymus schimperi</i> Ronniger (Lamiaceae)	Phenol and flavonoid compounds, and aqueous methanol extract showed <i>in vitro</i> radical scavenging ability, iron reducing power, and total antioxidant capacity [119]	Diabetes [62]; hypertension [18, 40]; tonsillitis [18]; toothache [18, 21]; abdominal pain [21]; and cough [38, 55]			
Sida schimperiana Hochst. ex A. Rich. (Malvaceae)	No previous reports	"Shotelaye" (hydrops fetalis) [21, 22]; cough and fever [62]; diarrhea [18]; wound [25, 62]; bleeding and evil eye [24]; glandular disease and rabies [40]; amoebic dysentery, and liver disease [65]; paralysis [21]; epilepsy [43]			

TABLE 4: Continued.

	TABLE 4: Continued.	
Botanical name (family)	Biological activity/chemical constituents	Illnesses/symptoms claimed to be treated traditionally
<i>Bersama abyssinica</i> Fresen. (Melianthaceae)	Flavonol glycosides isoquercetrin, hyperoside, quercetin-3- <i>O</i> -arabinopyranoside, kaempferol-3- O-arabinopyranoside, xanthone glycoside, mangiferin [115] Ethanol water extracts showed <i>in vitro</i> antioxidant activity on DPPH assay and antiproliferative activity on human liver carcinoma cell line and normal human fetal lung cells [79]; methanol extract showed <i>in vitro</i> antioxidant activity on DPPH assay [115], and antiviral activity against type 1 human immunodeficiency virus [102]	Tumor, dysentery and roundworms [107, 109]; ascariasis [15, 38, 81, 109]; wound [20]; stomach ache [17]; snake bite and liver diseases [70]; tonsillitis [72]; bronchitis and febrile illness [42, 43]
<i>Dorstenia barnimiana</i> Schweinf. (Moraceae)	Phytochemical screening showed the presence of coumarins [34]	Cancer [26]; hepatitis, syphilis and rabies [25, 26]; skin cancer, dysentery, wart and fever [25]; pulmonary tuberculosis, leprosy, and stomach illness [22]
<i>Syzygium guineense</i> (Willd.) DC. (Myrtaceae)	Methanol and dichloromethane crude extracts showed <i>in vitro</i> cytotoxicity on human leukaemia cells [73] and antimicrobial activity [120]	Stomach ache [17–19, 23]; diarrhea [15, 18, 19, 24], kidney infection, liver cirrhosis, and tonsillitis [18]; syphilis [23, 80]; malaria, hemorrhoid, internal worms, snake bite, and gonorrhea [65]
<i>Oxalis corniculata</i> L. (Oxalidaceae)	In vivo antitumor activity against Ehrlich ascites carcinoma on mice [121]	Wound [17]; arthritis [63]; tape worm infection [21]
<i>Rumex nervosus</i> Vahl (Polygonaceae)	Alkaloids, flavonoids, terpenoids, tannins, glycosides, and volatile oils [122]	Breast cancer, gastritis, and snake bite [16]; wart [15, 22]; hepatitis [49, 55]; skin rash [16, 21]; bleeding [15, 40, 81, 109]; wound [40, 49, 55, 62, 109, 110]; scabies and acne vulgaris [62]; ascariasis and herpes simplex [21]; stomach ache and dysentery [22]; diarrhea [49]; eye problems and round worm [55]
<i>Rumex nepalensis</i> Spreng. (Polygonaceae)	Anthraquinones, naphthalenes, tannins, stilbenoids [123] Ethanol water extracts showed <i>in vitro</i> antiproliferative activity on human liver carcinoma cell line and on normal human fetal lung cells and antioxidant activity on DPPH assay [79], and methanol and dichloromethane crude extracts showed <i>in vitro</i> cytotoxicity on human leukaemia cells [73]	Wound, ascariasis, abdominal bleeding, gastric ulcer, and hemorrhage [23, 80]; gastritis [18]; stomach problems [108]; leishmaniasis [25]; abdominal cramp and ear infection [63]; tonsillitis [18, 25]
<i>Clematis simensis</i> Fresen. (Ranunculaceae)	Triterpenoids, saponins, alkaloids, polyphenols, and unsaturated sterols [120] <i>In vivo</i> anti-inflammatory and antinociceptive activities [124]	Cancer and hemorrhoid [15]; wart and evil eye [24, 40]; wound [15, 24, 40, 63, 81]; tonsillitis [62]; eye infection [63]; leg swelling, malaria, and mental illness [49]; stomach ache [47]
<i>Prunus africana</i> (Hook.f.) Kalkman (Rosaceae)	No previous reports	Benign prostatic hyperplasia and prostate gland hypertrophy [20]; cancer, respiratory disorders, bad breathe, diarrhea, gonorrhea, tuberculosis, and ear problems [22]; swelling [40]; wounds [19, 22]; tonsillitis [23, 80]
<i>Clausena anisata</i> (Willd.) Hook.f. ex Benth. (Rutaceae)	Carbazole alkaloids, peptide derivatives, sitosterol, and stigmasterol [125] Methanol and dichloromethane crude extracts showed <i>in vitro</i> cytotoxicity on human leukaemia cells [73]	Skin irritation [20]; toothache [40]; ascariasis [19]; evil eye [24, 25, 63]
<i>Osyris quadripartita</i> Salzm. ex Decne. (Santalaceae)	Alkaloids, phenols, terpenoids, tannins, saponins, and flavonoids [126] Methanol extracts showed <i>in vitro</i> antimicrobial activity against <i>Escherichia coli, Pseudomonas</i> <i>aeruginosa, Staphylococcus aureus, Candida</i> <i>albicans</i> , and <i>Trichophyton mentagrophytes</i> [11]; <i>in</i> <i>vitro</i> inhibition of NO production and cytotoxicity against MCF-7 and NCI-H460 cell lines [127]	Cancer [62]; anaphylactic shock, evil eye, and epilepsy [18]; eczema [40]; toothache [46]

TABLE 4: Continued.

	TABLE 4: Continued.			
Botanical name (family)	Biological activity/chemical constituents	Illnesses/symptoms claimed to be treated traditionally		
<i>Dodonaea viscosa</i> subsp. <i>angustifolia</i> (L.f.) J.G.West (Sapindaceae)	Alkaloids, terpenoids, saponins, tannins, sugars, phenolics, and flavonoids [128] Methanol extracts showed <i>in vivo</i> nonsensitizer effect in mice using the mouse ear swelling test method [129], <i>in vitro</i> antiviral effect against type 1 human immunodeficiency virus [102], and <i>in vitro</i> free radical scavenging activity on DPPH assay [128]	Malaria [57]		
Brucea antidysenterica J.F.Mill. (Simaroubaceae)Flavonoids, amino acids, and vitamin C [130] In vitro antiplasmodial activity against Plasmodium berghei infection [131]		Cancer/tumor [107]; wart [24]; rabies [18, 62]; leprosy [62]		
<i>Discopodium penninervium</i> Hochst. (Solanaceae)	5α,17β-Dihydroxy-6α,7α-epoxy-1-oxowitha-2,24- dienolide, withanone, and withanolide A [132], 5,6-epoxy-16-oxygenated withanolides, jaborosalactone-L, and 17-epiacnistin-A [133, 134]; 6α,7α-epoxy-1-oxo-5α,12α,17α- trihydroxywitha-2,24-dienolide and a coloratane sesquiterpene, 7α,11α-dihydroxy-4(13),8- coloratadien-12,11-olide, withanone, 5α,17β- dihydroxy-6α,7α-epoxy-1-oxowitha-2,24- dienolide, 7α,11α-dihydroxy-8-drimen-12,11- olide, withasomnine, and (E,Z)-9- hydroxyoctadeca-10,12-dienoic acid [135] Jaborosalactone-L showed cytotoxicity only to the murine macrophage cell line, RAW 264.7, but the 16α-oxygenated withanolides exhibited cytotoxicity to both human (COR-L23 and ECV 304) and murine (L929 and RAW 264.7) carcinoma cell lines with IC <sub>50</sub> values ranging from 1.2 to 150 μM [136]. 6α,7α-Epoxy-1-oxo- 5α,12α,17α-trihydroxy-witha-2,24-dienolide inhibited COX-2 and LTB4 formation; 7α,11α- dihydroxy-4(13),8-coloratadien-12,11-olide and withasomnine inhibited LTB <sub>4</sub> biosynthesis but showed minor inhibition of COX-1 and COX-2 [135]	Skin detoxification [62]; and liver disease [70]		
<i>Gnidia involucrata</i> Steud. ex A.Rich. (Thymelaeaceae)	Flavonoids and glycosides [137]	Ascariasis, evil eye, anthrax, intestinal helminths, and gland swelling [18]		
<i>Lantana trifolia</i> L. (Verbenaceae)	Flavone glycosides (scutellarein-7- $O$ - $\beta$ -D- apiofuranoside and apigenin-7- $O$ - $\beta$ -D- apiofuranosyl-(1 $\longrightarrow$ 2)- $\beta$ -D-apiofuranoside) and the flavone celtidifoline (5,6,40,50-tetrahydroxy- 7,30-dimethoxyflavone) [138, 139]	Headache [70]; malaria [71]		
<i>Lippia adoensis</i> Hochst. (Verbenaceae)	Limonene, perillaldehyde, piperitenone, and 2- methyl-6-methylene-2,7-octadien-4-one [140], sesquiterpene hydrocarbon (germacrene D) [141] Methanol extract showed <i>in vitro</i> cytotoxicity on human leukaemia cell lines [73], and antimicrobial activity against <i>Staphylococcus aureus</i> , <i>Escherichia</i> <i>coli</i> , and <i>Pseudomonas aeruginosa</i> [75]; water extracts showed <i>in vivo</i> protection/relieve against acetic acid induced writhing in mice model [142]	Eczema, fungal infections, common cold, and cough [62]; intestine swelling [18]; gastrointestinal disorder [40]; abdominal irritation and acute stomach illness [46]		
Cyphostemma serpens (Hochst. ex A.Rich.) Desc. (Vitaceae)	No prev	No previous reports		

TABLE 4: Continued.

highest ICF value (0.42) recorded for "other ailments" category, in this study, suggests that informants are in agreement with the use of particular plant species to treat

ailments in this category. The lowest ICF value (0) obtained was for lung, colon, cervical, and throat cancer categories. This might be due to the cultural and ecological differences

No.	Category	No. of species	% of all species	No. of use reports	% of all use reports	ICF
1	Skin	25	30.5	32	26.5	0.23
2	Breast	20	24.4	29	23.9	0.32
3	Cervical	1	1.22	1	0.83	0
4	Colon	1	1.22	1	0.83	0
5	Lung	1	1.22	1	0.83	0
6	Throat	1	1.22	1	0.83	0
7	Other disease	33	40.2	56	46.3	0.42
	Total	82*		121		

TABLE 5: Informants consensus factor for different ailment categories.

\*Each plant species may be listed in several categories.

TABLE 6: Summary of stats for relative frequency of citation (RFC) and cultural importance index (CI).

	Mean	Standard deviation	Minimum	Maximum
UV	1.8	1.1	0.5	6
RFC	0.02	0.015	0.014	0.1
CI	0.034	0.027	0.014	0.16
Association between RFC and CI by using Pearson correlation method				

	UV	RFC	CI	
UV	1			
RFC	-0.36*	1		
CI	0.003	0.858**	1	
* 0	1		1	

\*Correlation is significant at 0.05 level. \*\*Correlation is significant at 0.001 level.

of the study sites and the difficulty to pinpoint the physical symptoms of lung, colon, cervical, and throat cancer as compared to the breast and skin cancer.

The present study also revealed that RFC and CI values of some reported species are similar. However, there is a distinct difference in species ranking using each index. C. macrostachyus is placed in the first position according to both RFC and CI index. This could be due to the fact that this species is mentioned by many informants and is the most recognized plant in most study areas. Furthermore, CI value of C. macrostachyus is also high, suggesting the diversified use of the plant. V. auriculifera and C. simensis ranked next to C. macrostachyus, according to RFC index. On the other hand, *D. barnimiana* and *Aloe* spp. ranked 2<sup>nd</sup> and 3<sup>rd</sup> by CI index. It has been suggested that UV value is a good measure of use diversity, than the number of citations [144]. In agreement with this, UV value in our study is driven by species with greatest number of use rather than those cited by more informants. The Pearson correlation coefficient of -0.36, between RFC and UV, shows significant negative association between the local importance of each medicinal plant and relative importance of use of plants. This result is in contrast to previous studies that reported a significant positive correlation between RFC and UV [145, 146]. On the other hand, there is a significant positive correlation between RFC and CI ( $r^2 = 0.74$ , p < 0.001) implying that their pattern matches across species. The species with larger RFC value usually have higher CI, such as Croton macrostachyus and Vernonia auriculifera.

Leaves and roots are the most commonly used plant parts in the preparation of remedies in the study districts. Similarly, other ethnobotanical studies conducted in different parts of Ethiopia also reported that leaves are the dominant plant part followed by root [16–18, 20]. The preference towards leaves may be because leaves are the main photosynthetic organs in plants and the primary reservoirs for secondary metabolites with medicinal values [36]. In contrary to other ethnobotanical studies [17, 18], where the common use of concoctions and oral route were reported, in the current study majority of the reported remedies are prepared from a single plant species and applied topically.

Comparative analysis of this study with other ethnobotanical surveys of plants used traditionally in treating and managing cancer in Ethiopia [18], Kenya [147], Cameroon [37], Nigeria [19, 38], South Africa [39], and Bangladesh [148] revealed some similarities in the plants cited in these surveys. Of the 30 plant species cited to be used in Ethiopia [18], 7 species are identified in our study: *Bersama abyssinica* Fresen., *Brucea antidysenterica* JF. Mill., *Calpurnia aurea* (Ait.) Benth. *Dodonaea angustifolia* L.f., Dorstenia barnimiana Schweinf, *Kalanchoe petitiana* A. Rich., and *Prunus africana* (Hook, f) Kalkm.

Although herbal remedies are believed by the general public to be safe [46], some research findings suggested otherwise. For instance, traditionally used Thai anticancer plants Ganoderma lucidum (Fr.) Karst., Houttuynia cordata Thunb., and Saussurea involucrata Matsum. & Koidz. were reported to cause side effects such as headache, insomnia, constipation, and diarrhea [62]. Similarly, side effects such as vomiting, diarrhea, and skin necrosis, associated with the use of traditional herbal remedies, were reported in this and other ethnobotanical studies conducted in Ethiopia [149, 150]. Few side effects reported in this study, as compared to other ethnobotanical studies conducted in Ethiopia, could be attributed to the frequent use of the topical route of administration. Nevertheless, considering the probability of underreporting adverse effects, extensive toxicological investigations should be conducted to protect the public.

In vitro cytotoxicity and antioxidant properties of some of the plants reported in our study have also been studied. Among these plants, potent cytotoxic activity was reported for knipholone anthrone isolated from *Kniphofia foliosa*, with  $IC_{50}$  value that ranges between  $0.9 \pm 0.1$  and  $3.3 \pm 0.4 \,\mu$ g/mL [89]. Similarly, Nibret and Wink reported the cytotoxic activity of the crude extract of *Acokanthera* 

schimperi with IC<sub>50</sub> value of 7.1  $\mu$ g/mL [73]. Studies conducted on the leaves of *Cineraria abyssinica* [100], bark of *Senna singueana* [116], and bark and leaves of *Rumex nepalensis* [79] also revealed potent radical scavenging activity of these plants.

# 5. Conclusion

The present study showed that traditional healers in eleven districts of Ethiopia use different medicinal plants to manage cancer-like symptoms. Frequency of citation value ranked Croton macrostachyus Del., Clematis simensis Fresen., Dorstenia barnimiana Schweinf, Vernonia auriculifera Hiern, and Acmella caulirhiza Del. as most cited plant species in study areas. Hence, based on these findings, we are currently evaluating the *in vitro* antiproliferative activities of reported medicinal plant species with a higher frequency of citation against human breast adenocarcinoma (MCF-7), human uterine cervical adenocarcinoma (SiSo), human lung carcinoma (A-427), and human bladder cancer (RT-4) cell lines using crystal violate assay. However, considering the rapid disappearance of the traditional knowledge of medicinal plants and an urgent need for new anticancer agents, additional studies have to be conducted to document and scientifically validate traditionally used Ethiopian anticancer plants.

# **Data Availability**

The authors declare that all data supporting the finding of this study are included in this article and its supplementary information files.

# **Ethical Approval**

Ethical approval was obtained from Addis Ababa University, College of Health Sciences Ethics Review Board (Ref no. ERB/SOP/126/12/2015).

#### Consent

Each participant consented before the interview.

# **Conflicts of Interest**

The authors declare that they have no conflicts of interest.

# **Authors' Contributions**

A.B., E.E., and K.A. jointly conceived the study. S.T. conducted the ethnobotanical study and taxonomical identification. S.T., A.B., E.E., T.G., and K.A. enriched the draft manuscript for its intellectual content. All authors read and approved the final manuscript.

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# References

- J. Zhang, S. S. Späth, S. L. Marjani, W. Zhang, and X. Pan, "Characterization of cancer genomic heterogeneity by next- generation sequencing advances precision medicine in cancer treatment," *Precision Clinical Medicine*, vol. 1, no. 1, pp. 29–48, 2018.
- [2] H. H. Heng, J. B. Stevens, S. W. Bremer, K. J. Ye, G. Liu, and C. J. Ye, "The evolutionary mechanism of cancer," *Journal of Cellular Biochemistry*, vol. 109, no. 6, pp. 1072–1084, 2014.
- [3] G. Finak, N. Bertos, F. Pepin et al., "Stromal gene expression predicts clinical outcome in breast cancer," *Nature Medicine*, vol. 14, no. 5, pp. 518–527, 2008.
- [4] H.-C. Wu, D.-K. Chang, and C.-T. Huang, "Targeted therapy for cancer," *Journal of Molecular Cancer*, vol. 2, no. 2, pp. 57–66, 2010.
- [5] J. A. Roth, F. Fossella, R. Komaki et al., "A randomized trial comparing perioperative chemotherapy and surgery with surgery alone in resectable stage IIIA non-small-cell lung cancer," *JNCI Journal of the National Cancer Institute*, vol. 86, no. 9, pp. 673–680, 1994.
- [6] K. Gregory and A. Tutt, "Managing side-effects of cancer therapy," in *Cancer in Primary Care*, M. E. Gore and D. Russell, Eds., pp. 47–88, Taylor and Francis Publishers, London, UK, 2003.
- [7] M. M. Gottesman, "Mechanisms of cancer drug resistance," Annual Review of Medicine, vol. 53, no. 1, pp. 615–627, 2002.
- [8] G. M. Cragg, D. J. Newman, and K. M. Snader, "Natural products in drug discovery and development," *Journal of Natural Products*, vol. 60, no. 1, pp. 52–60, 1997.
- [9] A. Harvey, "Medicines from nature: are natural products still relevant to drug discovery?" *Trends in Pharmacological Sciences*, vol. 20, no. 5, pp. 196–198, 1999.
- [10] K. Hostettmann, A. Marston, K. Ndjoko, and J.-L. Wolfender, "The potential of African plants as a source of drugs," *Current Organic Chemistry*, vol. 4, no. 10, pp. 973–1010, 2000.
- [11] S. Taddese, K. Asres, and T. Gebre-Mariam, "In vitro antimicrobial activities of some selected topically applied medicinal plants of Ethiopia," *Ethiopian Pharmaceutical Journal*, vol. 21, pp. 39–46, 2003.
- [12] S. D. E. Kelbessa, Z. Woldu, and S. Edwards, "Some threatened endemic plants of Ethiopia," in *The Status of Some Plants in Parts of Tropical Africa*, Z. A. S. Edwards, Ed., pp. 35–55, Addis Ababa University, Addis Ababa, Ethiopia, 1992.
- [13] K. D. Kassaye, A. Amberbir, B. Getachew, and Y. Mussema, "A historical overview of traditional medicine practices and policy in Ethiopia," *Ethiopian Journal of Health Development*, vol. 20, no. 2, pp. 127–134, 2007.
- [14] N. Tuasha, B. Petros, and Z. Asfaw, "Medicinal plants used by traditional healers to treat malignancies and other human ailments in Dalle District, Sidama Zone, Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 14, no. 1, p. 15, 2018.
- [15] G. Chekole, Z. Asfaw, and E. Kelbessa, "Ethnobotanical study of medicinal plants in the environs of Tara-gedam and Amba remnant forests of Libo Kemkem district, northwest Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 11, no. 1, p. 4, 2015.
- [16] S. Araya, B. Abera, and M. Giday, "Study of plants traditionally used in public and animal health management in

Seharti Samre district, Southern Tigray, Ethiopia," Journal of Ethnobiology and Ethnomedicine, vol. 11, no. 1, p. 22, 2015.

- [17] R. Regassa, "Assessment of indigenous knowledge of medicinal plant practice and mode of service delivery in Hawassa city, southern Ethiopia," *Journal of Medicinal Plants Research*, vol. 7, no. 9, pp. 517–535, 2013.
- [18] M. Agize, S. Demissew, and Z. Asfaw, "Ethnobotany of medicinal plants in Loma and Gena bosa districts (woredas) of dawro zone, southern Ethiopia," *Topclass Journal of Herbal Medicine*, vol. 2, no. 9, pp. 194–212, 2012.
- [19] S. Suleman and T. Alemu, "A survey on utilization of ethnomedicinal plants in Nekemte town, East Wellega (Oromia), Ethiopia," *Journal of Herbs, Spices & Medicinal Plants*, vol. 18, no. 1, pp. 34–57, 2012.
- [20] B. Abera, "Medicinal plants used in traditional medicine by Oromo people, Ghimbi district, Southwest Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 10, no. 1, p. 40, 2014.
- [21] A. Teklay, B. Abera, and M. Giday, "An ethnobotanical study of medicinal plants used in Kilte Awulaelo district, Tigray region of Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 9, no. 1, p. 65, 2013.
- [22] T. Teklehaymanot, M. Giday, G. Medhin, and Y. Mekonnen, "Knowledge and use of medicinal plants by people around Debre Libanos monastery in Ethiopia," *Journal of Ethnopharmacology*, vol. 111, no. 2, pp. 271–283, 2007.
- [23] M. Ragunathan and S. M. Abay, "Ethnomedicinal survey of folk drugs used in Bahirdar Zuria district, Northwestern Ethiopia," *Indian Journal of Traditional Knowledge*, vol. 8, pp. 281–284, 2009.
- [24] T. Mekuanent, A. Zebene, and Z. Solomon, "Ethnobotanical study of medicinal plants in Chilga district, Northwestern Ethiopia," *Journal of Natural Remedies*, vol. 15, no. 2, pp. 88–112, 2015.
- [25] T. Teklehaymanot, "Ethnobotanical study of knowledge and medicinal plants use by the people in Dek Island in Ethiopia," *Journal of Ethnopharmacology*, vol. 124, no. 1, pp. 69–78, 2009.
- [26] T. Teklehaymanot and M. Giday, "Ethnobotanical study of medicinal plants used by people in Zegie Peninsula, northwestern Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 3, no. 1, p. 12, 2007.
- [27] CSA, "The 2007 population and housing census of Ethiopia: administrative report," Central Statistical Agency, Addis Ababa, Ethiopia, 2012.
- [28] I. Friis, S. Demissew, and P. Van Breugel, "Atlas of the potential vegetation of Ethiopia," Det Kongelige Danske Videnskabernes Selskab, Copenhagen, Denmark, 2010.
- [29] J. Tardío and M. Pardo-de-Santayana, "Cultural importance indices: a comparative analysis based on the useful wild plants of southern cantabria (northern Spain)1," *Economic Botany*, vol. 62, no. 1, pp. 24–39, 2008.
- [30] O. Phillips and A. H. Gentry, "The useful plants of Tambopata, Peru: I. Statistical hypotheses tests with a new quantitative technique," *Economic Botany*, vol. 47, no. 1, pp. 15–32, 1993.
- [31] M. Heinrich, A. Ankli, B. Frei, C. Weimann, and O. Sticher, "Medicinal plants in Mexico: healers' consensus and cultural importance," *Social Science & Medicine*, vol. 47, no. 11, pp. 1859–1871, 1998.
- [32] J. Abdela, E. Engidawork, and W. Shibeshi, "In vivo antimalarial activity of solvent fractions of the leaves of justicia schimperiana hochst. Ex Nees against Plasmodium berghei

in Mice," *Ethiopian Pharmaceutical Journal*, vol. 30, no. 2, pp. 95–108, 2014.

- [33] B. Desta, "Ethiopian traditional herbal drugs—part III: antifertility activity of 70 medicinal plants," *Journal of Ethnopharmacology*, vol. 44, no. 3, pp. 199–209, 1994.
- [34] R. A. Mothana, R. Gruenert, P. Bednarski, and U. Lindequist, "Evaluation of the in vitro anticancer, antimicrobial and antioxidant activities of some Yemeni plants used in folk medicine," *Die Pharmazie—An International Journal of Pharmaceutical Sciences*, vol. 64, no. 4, pp. 260–268, 2009.
- [35] S. Umer, K. Asres, and C. Veeresham, "Hepatoprotective activities of two Ethiopian medicinal plants," *Pharmaceutical Biology*, vol. 48, no. 4, pp. 461–468, 2010.
- [36] H. Agisho, M. Osie, and T. Lambore, "Traditional medicinal plants utilization, management and threats in Hadiya Zone, Ethiopia," *Journal of Medicinal Plants Research*, vol. 2, no. 2, pp. 94–108, 2014.
- [37] A. Enyew, Z. Asfaw, E. Kelbessa, and R. Nagappan, "Status of medico-cultural commercial plants at Fiche town market, Ethiopia," *International Journal of Pharmaceuticals and Health Care Research*, vol. 1, no. 4, pp. 227–236, 2013.
- [38] Z. Kassa, Z. Asfaw, and S. Demissew, "Ethnobotanical study of medicinal plants used by the local people in tulu korma and its surrounding areas of ejere district, western shewa zone of oromia regional state, Ethiopia," *Journal of Medicinal Plants Studies*, vol. 4, no. 2, pp. 24–47, 2016.
- [39] S. Zerabruk and G. Yirga, "Traditional knowledge of medicinal plants in Gindeberet district, Western Ethiopia," *South African Journal of Botany*, vol. 78, pp. 165–169, 2012.
- [40] A. Kefalew, Z. Asfaw, and E. Kelbessa, "Ethnobotany of medicinal plants in ada'a district, east shewa zone of oromia regional state, Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 11, no. 1, p. 25, 2015.
- [41] T. T. Ayele, M. B. Regasa, and D. A. Delesa, "Antibacterial and antagonistic activity of selected traditional medicinal plants and herbs from East Wollega zone against clinical isolated human pathogens," *Science, Technology and Arts Research Journal*, vol. 4, no. 3, pp. 175–179, 2016.
- [42] G. Bekele and P. R. Reddy, "Ethnobotanical study of medicinal plants used to treat human ailments by Guji Oromo tribes in Abaya District, Borana, Oromia, Ethiopia," Universal Journal of Plant Science, vol. 3, no. 1, pp. 1–8, 2015.
- [43] F. Mesfin, S. Demissew, and T. Teklehaymanot, "An ethnobotanical study of medicinal plants in Wonago Woreda, SNNPR, Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 5, no. 1, p. 28, 2009.
- [44] M. Giday, T. Teklehaymanot, A. Animut, and Y. Mekonnen, "Medicinal plants of the shinasha, agew-awi and Amhara peoples in northwest Ethiopia," *Journal of Ethnopharmacology*, vol. 110, no. 3, pp. 516–525, 2007.
- [45] M. A. Eshete, E. Kelbessa, and G. Dalle, "Ethnobotanical study of medicinal plants in guji agro-pastoralists, blue Hora district of Borana zone, Oromia region, Ethiopia," *Journal of Medicinal Plants Studies*, vol. 4, no. 2, pp. 170–184, 2016.
- [46] H. Yineger, E. Kelbessa, T. Bekele, and E. Lulekal, "Plants used in traditional management of human ailments at Bale mountains national park, southeastern Ethiopia," *Journal of Medicinal Plants Research*, vol. 2, no. 6, pp. 132–153, 2008.
- [47] M. Megersa, Z. Asfaw, E. Kelbessa, A. Beyene, and B. Woldeab, "An ethnobotanical study of medicinal plants in Wayu Tuka district, east Welega zone of oromia regional state, West Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 9, no. 1, p. 68, 2013.

- [48] A. Belayneh, Z. Asfaw, S. Demissew, and N. F. Bussa, "Medicinal plants potential and use by pastoral and agropastoral communities in Erer Valley of Babile Wereda, Eastern Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 8, no. 1, p. 42, 2012.
- [49] T. Wondimu, Z. Asfaw, and E. Kelbessa, "Ethnobotanical study of medicinal plants around 'Dheeraa' town, Arsi Zone, Ethiopia," *Journal of Ethnopharmacology*, vol. 112, no. 1, pp. 152–161, 2007.
- [50] H. Yineger, D. Yewhalaw, and D. Teketay, "Ethnomedicinal plant knowledge and practice of the Oromo ethnic group in southwestern Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 4, no. 1, p. 11, 2008.
- [51] G. Asamenew, D. Bisrat, A. Mazumder, and K. Asres, "In vitro antimicrobial and antioxidant activities of anthrone and chromone from the latex of Aloe harlana Reynolds," *Phytotherapy Research*, vol. 25, no. 12, pp. 1756–1760, 2011.
- [52] E. Dagne, D. Bisrat, A. Viljoen, and B.-E. Van Wyk, "Chemistry of Aloe species," *Current Organic Chemistry*, vol. 4, no. 10, pp. 1055–1078, 2000.
- [53] T. Deressa, Y. Mekonnen, and A. Animut, "In vivo antimalarial activities of Clerodendrum myricoides, Dodonea angustifolia and Aloe debrana against Plasmodium berghei," *Ethiopian Journal of Health Development*, vol. 241 pages, 2010.
- [54] A. Mesfin, M. Giday, A. Animut, and T. Teklehaymanot, "Ethnobotanical study of antimalarial plants in Shinile district, Somali region, Ethiopia, and in vivo evaluation of selected ones against Plasmodium berghei," *Journal of Ethnopharmacology*, vol. 139, no. 1, pp. 221–227, 2012.
- [55] E. d'Avigdor, H. Wohlmuth, Z. Asfaw, and T. Awas, "The current status of knowledge of herbal medicine and medicinal plants in Fiche, Ethiopia," *Journal of Ethnobiology* and Ethnomedicine, vol. 10, no. 1, p. 38, 2014.
- [56] A. Belayneh and N. F. Bussa, "Ethnomedicinal plants used to treat human ailments in the prehistoric place of Harla and Dengego valleys, eastern Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 10, no. 1, p. 18, 2014.
- [57] G. Yirga, "Assessment of indigenous knowledge of medicinal plants in central zone of Tigray, northern Ethiopia," *African Journal of Plant Science*, vol. 4, no. 1, pp. 6–11, 2010.
- [58] G. Zenebe, M. Zerihun, and Z. Solomon, "An ethnobotanical study of medicinal plants in Asgede Tsimbila district, northwestern Tigray, northern Ethiopia," *Ethnobotany Research and Applications*, vol. 10, pp. 305–320, 2012.
- [59] T. Teklehaymanot and M. Giday, "Ethnobotanical study of wild edible plants of kara and kwego semi-pastoralist people in lower omo river valley, debub omo zone, SNNPR, Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 6, no. 1, p. 23, 2010.
- [60] W. Shibeshi, E. Makonnen, L. Zerihun, and A. Debella, "Effect of Achyranthes aspera L. on fetal abortion, uterine and pituitary weights, serum lipids and hormones," African Health Sciences, vol. 6, no. 2, pp. 108–112, 2006.
- [61] A. Fikru, E. Makonnen, T. Eguale, A. Debella, and G. Abie Mekonnen, "Evaluation of in vivo wound healing activity of methanol extract of *Achyranthes aspera* L," *Journal of Ethnopharmacology*, vol. 143, no. 2, pp. 469–474, 2012.
- [62] A. Enyew, Z. Asfaw, E. Kelbessa, and R. Nagappan, "Ethnobotanical study of traditional medicinal plants in and around Fiche district, Central Ethiopia," *Current Research Journal of Biological Sciences*, vol. 6, no. 4, pp. 154–167, 2014.

- [63] M. Giday, Z. Asfaw, and Z. Woldu, "Ethnomedicinal study of plants used by Sheko ethnic group of Ethiopia," *Journal of Ethnopharmacology*, vol. 132, no. 1, pp. 75–85, 2010.
- [64] E. Lulekal, Z. Asfaw, E. Kelbessa, and P. Van Damme, "Ethnoveterinary plants of Ankober district, north Shewa zone, Amhara region, Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 10, no. 1, p. 21, 2014.
- [65] F. Mesfin, T. Seta, and A. Assefa, "An ethnobotanical study of medicinal plants in Amaro Woreda, Ethiopia," *Ethnobotany Research and Applications*, vol. 12, pp. 341–354, 2014.
- [66] J. Barnes, L. A. Anderson, J. D. Phillipson, and C. A. Newall, *Herbal Medicines*, Pharmaceutical Press London, London, UK, 2007.
- [67] S. S. Jamil, Q. Nizami, and M. Salam, "Centella asiatica (Linn.) Urban: a review," Natural Product Radiance, vol. 6, no. 2, pp. 158–170, 2007.
- [68] M. Yoshida, M. Fuchigami, T. Nagao et al., "Antiproliferative constituents from Umbelliferae plants VII: active triterpenes and rosmarinic acid from *Centella asiatica*," *Biological & Pharmaceutical Bulletin*, vol. 28, no. 1, pp. 173–175, 2005.
- [69] P. Bunpo, K. Kataoka, H. Arimochi et al., "Inhibitory effects of *Centella asiatica* on azoxymethane-induced aberrant crypt focus formation and carcinogenesis in the intestines of F344 rats," *Food and Chemical Toxicology*, vol. 42, no. 12, pp. 1987–1997, 2004.
- [70] B. Kidane, T. van Andel, L. van der Maesen, and Z. Asfaw, "Use and management of traditional medicinal plants by Maale and Ari ethnic communities in southern Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 10, no. 1, p. 46, 2014.
- [71] K. Tolossa, E. Debela, S. Athanasiadou, A. Tolera, G. Ganga, and J. G. Houdijk, "Ethno-medicinal study of plants used for treatment of human and livestock ailments by traditional healers in south Omo, southern Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 9, no. 1, p. 32, 2013.
- [72] M. Giday, Z. Asfaw, and Z. Woldu, "Medicinal plants of the Meinit ethnic group of Ethiopia: an ethnobotanical study," *Journal of Ethnopharmacology*, vol. 124, no. 3, pp. 513–521, 2009.
- [73] E. Nibret and M. Wink, "Trypanocidal and cytotoxic effects of 30 Ethiopian medicinal plants," *Zeitschrift für Naturforschung C*, vol. 66, no. 11-12, pp. 0541–0546, 2011.
- [74] T. Gebre-Mariam, R. Neubert, P. Schmidt, P. Wutzler, and M. Schmidtke, "Antiviral activities of some Ethiopian medicinal plants used for the treatment of dermatological disorders," *Journal of Ethnopharmacology*, vol. 104, no. 1-2, pp. 182–187, 2006.
- [75] H. Tadeg, E. Mohammed, K. Asres, and T. Gebre-Mariam, "Antimicrobial activities of some selected traditional Ethiopian medicinal plants used in the treatment of skin disorders," *Journal of Ethnopharmacology*, vol. 100, no. 1-2, pp. 168–175, 2005.
- [76] T. Mohammed, B. Erko, and M. Giday, "Evaluation of antimalarial activity of leaves of Acokanthera schimperi and Croton macrostachyus against Plasmodium berghei in Swiss albino mice," *BMC Complementary and Alternative Medicine*, vol. 14, no. 1, p. 314, 2014.
- [77] D. Abebe and A. Ayehu, Medicinal Plants and Enigmatic Health Practices of Northern Ethiopia, Berhanina Selam Printing Press, Addis Ababa, Ethiopia, 1993.
- [78] G. Meaza, B. Tadesse, A. S. Maria, B. Piero, and Y. Gidey, "Traditional medicinal plants used by Kunama ethnic group in northern Ethiopia," *Journal of Medicinal Plants Research*, vol. 9, no. 15, pp. 494–509, 2015.

- [79] J. Tauchen, I. Doskocil, C. Caffi et al., "In vitro antioxidant and anti-proliferative activity of Ethiopian medicinal plant extracts," *Industrial Crops and Products*, vol. 74, pp. 671–679, 2015.
- [80] Z. Birhanu, "Ethno-botanical survey on medicinal plants used by ethnic groups of Denbia district, north-western Ethiopia," *Journal of Natural Remedies*, vol. 11, no. 2, pp. 119–123, 2011.
- [81] G. Seyoum and G. Zerihun, "An ethnobotanical study of medicinal plants in Debre Libanos Wereda, Central Ethiopia," *African Journal of Plant Science*, vol. 8, no. 7, pp. 366–379, 2014.
- [82] H. Heli, M. Amani, A. A. Moosavi-Movahedi, A. Jabbari, G. Floris, and A. Mura, "Electroactive centers in Euphorbia latex and lentil seedling amine oxidases," *Bioscience, Biotechnology, and Biochemistry*, vol. 72, no. 1, pp. 29–36, 2008.
- [83] S. R. Setty, A. A. Quereshi, A. V. Swamy et al., "Hepatoprotective activity of Calotropis procera flowers against paracetamol-induced hepatic injury in rats," *Fitoterapia*, vol. 78, no. 7-8, pp. 451–454, 2007.
- [84] S. Roy, R. Sehgal, B. M. Padhy, and V. L. Kumar, "Antioxidant and protective effect of latex of Calotropis procera against alloxan-induced diabetes in rats," *Journal of Ethnopharmacology*, vol. 102, no. 3, pp. 470–473, 2005.
- [85] J. V. Kamath and A. C. Rana, "Preliminary study on antifertility activity of Calotropis procera roots in female rats," *Fitoterapia*, vol. 73, no. 2, pp. 111–115, 2002.
- [86] A. E.-D. H. Sayed, N. H. Mohamed, M. A. Ismail, W. M. Abdel-Mageed, and A. A. M. Shoreit, "Antioxidant and antiapoptotic activities of Calotropis procera latex on Catfish (*Clarias gariepinus*) exposed to toxic 4-nonylphenol," *Ecotoxicology and Environmental Safety*, vol. 128, pp. 189– 194, 2016.
- [87] T. Flatie, T. Gedif, K. Asres, and T. Gebre-Mariam, "Ethnomedical survey of Berta ethnic group Assosa zone, Benishangul-Gumuz regional state, mid-west Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 5, no. 1, p. 14, 2009.
- [88] A. A. Wube, F. Bucar, K. Asres, S. Gibbons, L. Rattray, and S. L. Croft, "Antimalarial compounds from *Kniphofia foliosa* roots," *Phytotherapy Research*, vol. 19, no. 6, pp. 472–476, 2005.
- [89] S. Habtemariam, "Knipholone anthrone from Kniphofia foliosa induces a rapid onset of necrotic cell death in cancer cells," Fitoterapia, vol. 81, no. 8, pp. 1013–1019, 2010.
- [90] S. Habtemariam, "Antioxidant activity of Knipholone anthrone," *Food Chemistry*, vol. 102, no. 4, pp. 1042–1047, 2007.
- [91] N. R. Crouch, A. Langlois, D. A. Mulholland, J. J. Nair, and P. Houghton, "A novel alkylamide from the leaves of *Acmella caulirhiza* (Asteraceae), a traditional surface analgesic," *South African Journal of Botany*, vol. 71, no. 2, pp. 228–230, 2005.
- [92] B. O. Owuor, J. O. Ochanda, J. O. Kokwaro et al., "In vitro antiplasmodial activity of selected Luo and Kuria medicinal plants," *Journal of Ethnopharmacology*, vol. 144, no. 3, pp. 779–781, 2012.
- [93] Y. Tariku, A. Hymete, A. Hailu, and J. Rohloff, "In vitro evaluation of antileishmanial activity and toxicity of essential oils of Artemisia absinthium and Echinops kebericho," *Chemistry & Biodiversity*, vol. 8, no. 4, pp. 614–623, 2011.
- [94] E. Nibret and M. Wink, "Volatile components of four Ethiopian Artemisia species extracts and their in vitro

antitrypanosomal and cytotoxic activities," *Phytomedicine*, vol. 17, no. 5, pp. 369–374, 2010.

- [95] M. Burits, K. Asres, and F. Bucar, "The antioxidant activity of the essential oils of *Artemisia afra*, *Artemisia abyssinica* and *Juniperus procera*," *Phytotherapy Research*, vol. 15, no. 2, pp. 103–108, 2001.
- [96] X. X. Zhu, L. Yang, Y. J. Li et al., "Effects of sesquiterpene, flavonoid and coumarin types of compounds from Artemisia annua L. on production of mediators of angiogenesis," *Pharmacological Reports*, vol. 65, no. 2, pp. 410–420, 2013.
- [97] D.-D. Zhai, K. Supaibulwatana, and J.-J. Zhong, "Inhibition of tumor cell proliferation and induction of apoptosis in human lung carcinoma 95-D cells by a new sesquiterpene from hairy root cultures of *Artemisia annua*," *Phytomedicine*, vol. 17, no. 11, pp. 856–861, 2010.
- [98] G.-Q. Zheng, "Cytotoxic terpenoids and flavonoids from Artemisia annua," Planta Medica, vol. 60, no. 1, pp. 54–57, 1994.
- [99] T. Teshome, B. Sintayehu, H. Yohannes et al., "Radical scavenging activity and preliminary phytochemical screening on aerial part extracts of Cineraria abyssinica sch. bip," *Journal of Pharmacognosy and Phytochemistry*, vol. 3, no. 6, pp. 239–243, 2015.
- [100] B. Sintayehu, K. Asres, and Y. Raghavendra, "Radical scavenging activities of the leaf extracts and a flavonoid glycoside isolated from Cineraria abyssinica Sch. Bip. Exa. Rich," *Journal of Applied Pharmaceutical Science*, vol. 2, no. 4, pp. 44–49, 2012.
- [101] P. Cos, N. Hermans, T. De Bruyne et al., "Further evaluation of Rwandan medicinal plant extracts for their antimicrobial and antiviral activities," *Journal of Ethnopharmacology*, vol. 79, no. 2, pp. 155–163, 2002.
- [102] K. Asres, F. Bucar, T. Kartnig, M. Witvrouw, C. Pannecouque, and E. De Clercq, "Antiviral activity against human immunodeficiency virus type 1 (HIV-1) and type 2 (HIV-2) of ethnobotanically selected Ethiopian medicinal plants," *Phytotherapy Research*, vol. 15, no. 1, pp. 62–69, 2001.
- [103] B. Albejo, M. Endale, B. Kibret, and M. Anza, "Phytochemical investigation and antimicrobial activity of leaves extract of Vernonia auriculifera Hiern," *Journal of Pharmacy* & *Pharmacognosy Research*, vol. 3, no. 6, pp. 141–147, 2015.
- [104] J. Ibrahim, V. C. Ajaegbu, and H. O. Egharevba, "Pharmacognostic and phytochemical analysis of *Commelina benghalensis* L," *Ethnobotanical Leaflets*, vol. 2010, no. 5, p. 7, 2010.
- [105] S. Hasan, M. Hossain, R. Akter et al., "Sedative and anxiolytic effects of different fractions of the Commelina benghalensis Linn," *Drug Discoveries & Therapeutics*, vol. 3, no. 5, 2009.
- [106] A. Mekonnen, T. Sidamo, K. Asres, and E. Engidawork, "In vivo wound healing activity and phytochemical screening of the crude extract and various fractions of Kalanchoe petitiana A. Rich (Crassulaceae) leaves in mice," *Journal of Ethnopharmacology*, vol. 145, no. 2, pp. 638–646, 2013.
- [107] D. Abebe, A. Debella, and K. Urga, *Medicinal Plants and other Useful Plants of Ethiopia*, Camerapix Publishers International, Addis Ababa, Ethiopia, 2013.
- [108] R. W. Bussmann, P. Swartzinsky, A. Worede, and P. Evangelista, "Plant use in odo-bulu and demaro, Bale region, Ethiopia," *Journal of Ethnobiology and Ethnomedicine*, vol. 7, no. 1, p. 28, 2011.
- [109] Z. Birhanu, "Traditional use of medicinal plants by the ethnic groups of Gondar Zuria district, north-western Ethiopia," *Journal of Natural Remedies*, vol. 13, no. 1, pp. 46–53, 2011.

- [110] Y. Limenih, S. Umer, and M. Wolde-Mariam, "Ethnobotanical study on traditional medicinal plants in Dega damot woreda, Amhara region, north Ethiopia," *International Journal of Research in Pharmacy and Chemistry*, vol. 5, pp. 258–273, 2015.
- [111] G. Belay, Y. Tariku, T. Kebede, A. Hymete, and Y. Mekonnen, "Ethnopharmacological investigations of essential oils isolated from five Ethiopian medicinal plants against eleven pathogenic bacterial strains," *Phytopharmacology*, vol. 1, no. 5, pp. 133–143, 2011.
- [112] H. Almehdar, H. M. Abdallah, A.-M. M. Osman, and E. A. Abdel-Sattar, "In vitro cytotoxic screening of selected Saudi medicinal plants," *Journal of Natural Medicines*, vol. 66, no. 2, pp. 406–412, 2012.
- [113] K. Asres, W. A. Gibbons, J. D. Phillipson, and P. Mascagni, "Alkaloids of Ethiopian Calpurnia aurea subsp. aurea," *Phytochemistry*, vol. 25, no. 6, pp. 1443–1447, 1986.
- [114] K. Asres, J. Phillipson, and P. Mascagni, "Two novel minor alkaloids from Ethiopian Calpurnia aurea sp.aurea," *Planta Medica*, vol. 52, no. 4, pp. 302–304, 1986.
- [115] K. Asres, F. Sporer, and M. Wink, "Patterns of pyrrolizidine alkaloids in 12 Ethiopian Crotalaria species," *Biochemical Systematics and Ecology*, vol. 32, no. 10, pp. 915–930, 2004.
- [116] M. Gebrelibanos, K. Asres, and C. Veeresham, "In vitro radical scavenging activity of the leaf and bark extracts of Senna singueana (Del). Lock," *Ethiopian Pharmaceutical Journal*, vol. 25, pp. 77–84, 2008.
- [117] F. Tchoumbougnang, P. H. Zollo, E. Dagne, and Y. Mekonnen, "In Vivo antimalarial activity of essential oils from *Cymbopogon citratus* and *Ocimum gratissimumon* mice infected with *Plasmodium berghei*," *Planta Medica*, vol. 71, no. 1, pp. 20–23, 2005.
- [118] K. Asres, H. Asfaha, A. Mazumder, and F. Bucar, "Leaf essential oils of Salvia nilotica and Salvia schimperi: their antimicrobial and antioxidant activities," *Ethiopian Pharmaceutical Journal*, vol. 26, no. 1, pp. 49–58, 2008.
- [119] D. Engeda, B. Geremew, D. H. Gulelat, and P. V. R. H, "Antioxidant and -amylase inhibition activities in vitro of various solvent extracts of Thymus schimperi Ronniger," *Journal of Medicinal Plants Research*, vol. 9, no. 15, pp. 515–524, 2015.
- [120] A. Geyid, D. Abebe, A. Debella et al., "Screening of some medicinal plants of Ethiopia for their anti-microbial properties and chemical profiles," *Journal of Ethnopharmacology*, vol. 97, no. 3, pp. 421–427, 2005.
- [121] A. Kathiriya, K. Das, E. Kumar, and K. Mathai, "Evaluation of antitumor and antioxidant activity of oxalis corniculata linn: against ehrlich ascites carcinoma on mice," *Iranian Journal of Cancer Prevention*, vol. 34 pages, 2010.
- [122] B. B. Kasimala, V. R. Anna, and U. R. Mallu, "Reverse phase liquid chromatographic method for the simultaneous estimation of antibiotic drugs: metronidazole, nalidixic acid, tinidazole and norfloxacin," *Der Pharmacia Lettre*, vol. 6, pp. 411–419, 2014.
- [123] A. Vasas, O. Orbán-Gyapai, and J. Hohmann, "The genus rumex: review of traditional uses, phytochemistry and pharmacology," *Journal of Ethnopharmacology*, vol. 175, pp. 198–228, 2015.
- [124] A. Tadele, K. Asres, D. Melaku, and W. Mekonnen, "In vivo anti-inflammatory and antinociceptive activities of the leaf extracts of Clematis simensis Fresen," *Ethiopian Pharmaceutical Journal*, vol. 27, pp. 33–41, 2010.
- [125] J. Songue, E. Kouam, E. Dongo, T. Mpondo, and R. White, "Chemical constituents from stem bark and roots of

Clausena anisata," *Molecules*, vol. 17, no. 11, pp. 13673–13686, 2012.

- [126] M. Abebaw, B. Mishra, and D. A. Gelayee, "Evaluation of anti-ulcer activity of the leaf extract of," *Journal of Experimental Pharmacology*, vol. 9, pp. 1–11, 2017.
- [127] W. Rached, R. C. Calhelha, Â. Fernandes et al., "Phytochemical characterization and bioactive properties of Osyris quadripartita Salzm. ex Decne. leaves from Algeria," *RSC Advances*, vol. 6, no. 76, pp. 72768–72776, 2016.
- [128] T. Riaz, M. A. Abbasi, T. Shahzadi, M. Ajaib, and K. M. Khan, "Phytochemical screening, free radical scavenging, antioxidant activity and phenolic content of Dodonaea viscosa," *Journal of the Serbian Chemical Society*, vol. 771 pages, 2012.
- [129] K. Teshome, T. Gebre-Mariam, K. Asres, and E. Engidawork, "Toxicity studies on dermal application of plant extract of Dodonaea viscosa used in Ethiopian traditional medicine," *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, vol. 24, no. 1, pp. 60–69, 2010.
- [130] A. Amuamuta, Z. Mekonnen, and E. Gebeyehu, "Traditional therapeutic uses and phytochemical screening of some selected indigenous medicinal plants from Northwest Ethiopia," *African Journal of Pharmacology and Therapeutics*, vol. 4, no. 3, 2015.
- [131] A. Kefe, M. Giday, H. Mamo, and B. Erko, "Antimalarial properties of crude extracts of seeds of Brucea antidysenterica and leaves of Ocimum lamiifolium," *BMC Complementary and Alternative Medicine*, vol. 16, no. 1, p. 118, 2016.
- [132] S. Habtemariam and A. I. Gray, "Withanolides from the roots of *Discopodium penninervium*," *Planta Medica*, vol. 64, no. 3, pp. 275-276, 1998.
- [133] S. Habtemariam, A. I. Gray, and P. G. Waterman, "16-oxygenated withanolides from the leaves of Discopodium penninervium," *Phytochemistry*, vol. 34, no. 3, pp. 807–811, 1993.
- [134] S. Habtemariam, B. W. Skelton, P. G. Waterman, and A. H. White, "17-Epiacnistin-A, a further withanolide from the leaves of discopodium p enninervium," *Journal of Natural Products*, vol. 63, no. 4, pp. 512-513, 2000.
- [135] A. A. Wube, E.-M. Wenzig, S. Gibbons, K. Asres, R. Bauer, and F. Bucar, "Constituents of the stem bark of *Discopodium penninervium* and their LTB4 and COX-1 and-2 inhibitory activities," *Phytochemistry*, vol. 69, no. 4, pp. 982–987, 2008.
- [136] S. Habtemariam, "Cytotoxicity and immunosuppressive activity of withanolides from Discopodium penninervium," *Planta Medica*, vol. 63, no. 1, pp. 15–17, 1997.
- [137] J. Ferrari, C. Terreaux, S. Sahpaz, J. D. Msonthi, J.-L. Wolfender, and K. Hostettmann, "Benzophenone glycosides from Gnidia involucrata," *Phytochemistry*, vol. 54, no. 8, pp. 883–889, 2000.
- [138] P. S. Imbenzi, Y.-Z. He, Z.-X. Yan, E. K. Osoro, and P. K. Cheplogoi, "Chemical constituents in extracts from leaves of *Lantana trifolia* and their in vitro anti-oxidative activity," *Chinese Herbal Medicines*, vol. 6, no. 3, pp. 242– 246, 2014.
- [139] A. Valerino, T. González, and I. Spengler, "Bio-directed phytochemical study of the allelopathic activity of Lantana trifolia L. leaves. Part I," in *Proceedings of the XVII Congreso de la Asociación Latinoamericana de Malezas (ALAM) I Congreso Iberoamericano de Ciencia de las Malezas, IV Congreso Nacional de Ciencia de Malezas*, Asociación Latinoamericana de Malezas (ALAM), Matanzas, Cuba, pp. 652–660, November 2005.

- [140] B. Abegaz, N. Asfaw, and W. Lwande, "Constituents of the essential oils from wild and cultivated Lippia adoensis Hochst. ex Walp," *Journal of Essential Oil Research*, vol. 5, no. 5, pp. 487–491, 1993.
- [141] K. Asres and F. Bucar, "Lippia adoensis var. adoensis: studies on the essential oil composition and antioxidant activity," *Ethiopian Pharmaceutical Journal*, vol. 20, pp. 32–38, 2002.
- [142] A. Debella, E. Makonnen, D. Abebe, F. Teka, and A. Kidanemariam, "Pain management in mice using the aqueous and ethanol extracts of four medicinal plants," *East African Medical Journal*, vol. 80, no. 8, pp. 435–439, 2004.
- [143] A. Tesfaye, E. Makonnen, and S. Gedamu, "Hypoglycemic and antihyperglycemic activity of aqueous extract of Justicia Schimperiana leaves in normal and streptozotocin-induced diabetic mice," *International Journal of Pharma Sciences and Research*, vol. 7, no. 2, pp. 110–113, 2016.
- [144] V. A. Da Silva, L. D. H. C. Andrade, and U. P. De Albuquerque, "Revising the cultural significance index: the case of the Fulni-ô in northeastern Brazil," *Field Methods*, vol. 18, no. 1, pp. 98–108, 2006.
- [145] K. S. Ahmad, A. Hamid, F. Nawaz et al., "Ethnopharmacological studies of indigenous plants in Kel village, Neelum Valley, Azad Kashmir, Pakistan," *Journal of Ethnobiology and Ethnomedicine*, vol. 13, no. 1, p. 68, 2017.
- [146] A. Bano, M. Ahmad, T. B. Hadda et al., "Quantitative ethnomedicinal study of plants used in the skardu valley at high altitude of Karakoram-Himalayan range, Pakistan," *Journal* of *Ethnobiology and Ethnomedicine*, vol. 10, no. 1, p. 43, 2014.
- [147] Z. Birhanu, A. Endale, and Z. Shewamene, "An ethnomedicinal investigation of plants used by traditional healers of Gondar town, north-western Ethiopia," *Journal of Medicinal Plants Studies*, vol. 3, no. 2, pp. 36–43, 2015.
- [148] B. Kebu, K. Ensermu, and A. Zemede, "Indigenous medicinal utilization, management and threats in Fentale area, Eastern Shewa, Ethiopia," *Ethiopian Journal of Biological Sciences*, vol. 3, no. 1, pp. 37–58, 2015.
- [149] O. Ryding, "Lamiaceae," in *Flora of Ethiopia and Eritrea*, E. K. I. Hedberg, S. Edwards, S. Demissew, and E. Persson, Eds., pp. 516–604, The National Herbarium, Addis Ababa University, Addis Ababa, Ethiopia, 2006.
- [150] M. Tadesse, "Asteraceae (compositae)," in *Flora of Ethiopia* and Eritrea, I. F. I. Hedberg and S. Edwards, Eds., pp. 222-223, Addis Ababa and Uppsala: The National Herbarium, Addis Ababa University and Uppsala University, Addis Ababa, Ethiopia, 2004.