

# Response and survival predictors in a cohort of 319 patients with Waldenström macroglobulinemia treated with ibrutinib monotherapy

Jorge J. Castillo,<sup>1,2</sup> Shayna R. Sarosiek,<sup>1,2</sup> Joshua N. Gustine,<sup>1,3</sup> Catherine A. Flynn,<sup>1</sup> Carly R. Leventoff,<sup>1</sup> Timothy P. White,<sup>1</sup> Kirsten Meid,<sup>1</sup> Maria L. Guerrero,<sup>1</sup> Amanda Kofides,<sup>1</sup> Xia Liu,<sup>1</sup> Manit Munshi,<sup>1</sup> Nicholas Tsakmaklis,<sup>1</sup> Zachary R. Hunter,<sup>1,2</sup> Christopher J. Patterson,<sup>1</sup> Andrew R. Branagan,<sup>2,4</sup> and Steven P. Treon<sup>1,2</sup>

<sup>1</sup>Bing Center for Waldenström Macroglobulinemia, Dana-Farber Cancer Institute, Boston, MA; <sup>2</sup>Department of Medicine, Harvard Medical School, Boston, MA; <sup>3</sup>Department of Medicine, Boston University School of Medicine, Boston, MA; and <sup>4</sup>Division of Hematology and Oncology, Massachusetts General Hospital, Boston, MA

## Key Points

- *CXCR4* mutations are associated with lower rates of major (67% vs 86%) and deep responses (16% vs 35%) in ibrutinib-treated WM patients.
- *CXCR4* mutations and platelet count 100 K/uL or less were associated with worse PFS, and a scoring system using these 2 factors is proposed.

Bruton tyrosine kinase (BTK) inhibitors are the only FDA-approved treatments for Waldenström macroglobulinemia (WM). Factors prognostic of survival and predictive of response to BTK inhibitors remained to be clarified. We evaluated 319 patients with WM to identify predictive and prognostic factors on ibrutinib monotherapy. Logistic and Cox proportional-hazard regression models were fitted for response and survival. Multiple imputation analyses were used to address bias associated with missing data. Major (partial response or better) and deep responses (very good partial response or better) were attained in 78% and 28% of patients. *CXCR4* mutations were associated with lower odds of major (odds ratio [OR], 0.2; 95% confidence interval [CI], 0.1-0.5;  $P < .001$ ) and deep response (OR, 0.3; 95% CI, 0.2-0.6;  $P = .001$ ). *CXCR4* mutations (hazard ratio [HR], 2.0; 95% CI, 1.2-3.4;  $P = .01$ ) and platelet count 100 K/uL or less (HR, 2.5; 95% CI, 1.3-4.9;  $P = .007$ ) were associated with worse progression-free survival (PFS). We proposed a scoring system using these 2 factors. The median PFS for patients with 0, 1, and 2 risk factors were not reached, 5 years and 3 years ( $P < .001$ ). Patients with 2 risk factors had HR 2.2 (95% CI, 1.3-3.8;  $P = .004$ ) compared with 1 factor, and patients with 1 factor had HR 2.3 (95% CI, 1.1-5.1;  $P = .03$ ) compared with 0 factors. Age  $\geq 65$  years was the only factor associated with overall survival (HR, 3.2; 95% CI, 1.4-7.0;  $P = .005$ ). Multiple imputation analyses did not alter our results. Our study confirms the predictive and prognostic value of *CXCR4* mutations in patients with WM treated with ibrutinib monotherapy.

## Introduction

Waldenström macroglobulinemia (WM) is an indolent lymphoma composed of malignant IgM-secreting lymphoplasmacytic cells that accumulate in the bone marrow (BM) and other organs.<sup>1</sup> The identification of recurrent somatic mutations in *MYD88* and *CXCR4*, detected in 90% and 40% of WM patients, respectively, have helped our understanding of the biology of the disease.<sup>2,3</sup> Bruton tyrosine kinase (BTK) activation is apparent in *MYD88* mutated WM cells,<sup>4</sup> supporting the development of BTK inhibitors in WM patients, while *CXCR4* signaling suggested resistance mechanisms in WM cells despite

Submitted 7 September 2021; accepted 6 December 2021; prepublished online on *Blood Advances* First Edition 29 December 2021; final version published online 4 February 2022. DOI 10.1182/bloodadvances.2021006106.

Requests for data sharing may be submitted to Jorge J. Castillo (jorgej\_castillo@dfci.harvard.edu).

The full-text version of this article contains a data supplement.

© 2022 by The American Society of Hematology. Licensed under Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0), permitting only noncommercial, nonderivative use with attribution. All other rights reserved.

effective *MYD88* inhibition.<sup>5</sup> Mounting evidence supports an impact of genomic profile in the clinical features of WM patients as well as response to BTK inhibitors.<sup>6-9</sup>

Ibrutinib, a first-in-class oral BTK inhibitor, was approved by the US Food and Drug Administration and the European Medicines Agency in 2015 for the treatment of symptomatic WM patients, based on a response rate of 90% observed in a prospective investigator-initiated phase 2 study evaluating 63 previously treated WM patients.<sup>10</sup> Similar response rates to ibrutinib monotherapy were reported in rituximab-refractory and treatment-naïve WM patients.<sup>11,12</sup> A recent study from our group reported similar outcomes to ibrutinib in WM patients on and off clinical trials,<sup>13</sup> but long-term data on the safety and efficacy of ibrutinib monotherapy are limited.<sup>14</sup> The prognostic value of the International Prognostic Scoring System for WM (IPSSWM) has not yet been validated in patients treated with ibrutinib.

We designed a retrospective study to evaluate the long-term safety and efficacy of ibrutinib monotherapy in WM patients. The main objective of our study was to evaluate factors predictive of response and prognostic of progression-free survival (PFS) and overall survival (OS).

## Patients and methods

### Patient selection

The analytical cohort was composed of patients treated with ibrutinib monotherapy who participated in 2 prospective clinical trials (NCT01614821 and NCT02604511) and consecutive patients from a prospectively maintained database at our institution. The study inclusion period ran from January 2012 through December 2018, with follow-up through December 2020. Data on the individual cohorts of patients in clinical trials have been previously published.<sup>10,12</sup> All patients met clinicopathological criteria for the diagnosis of WM and for treatment initiation based on the Second International Workshop on WM guidelines.<sup>15,16</sup> All patients provided consent to having blood and marrow samples and data collected for research. Patients with central nervous system involvement by WM (Bing-Neel syndrome), non-IgM lymphoplasmacytic lymphoma, or who received ibrutinib in combination with other agents were excluded from the study.

### Data collection

Pertinent data were collected at the time of ibrutinib monotherapy initiation. Data were categorized as follows: age (>65, ≤65 years), sex (male, female), hemoglobin level (<11.5, ≥11.5 g/dl), platelet count (<100, ≥100 K/uL), serum IgM level (>4000, ≤4000 mg/dl, and >7000, ≤7000 mg/dl), serum β2-microglobulin level (>3, ≤3 mg/l), serum albumin level (≤3.5, >3.5 g/dl), BM involvement (≥60%, <60%), IPSSWM (low, intermediate, high), *CXCR4* mutational status (mutated, wildtype), *CXCR4* mutational status subtype (nonsense, frameshift), and previously treated (yes, no). Responses were assessed using modified criteria from the Sixth International Workshop on WM.<sup>17</sup> A decrease of 25% to 49%, 50% to 89%, and ≥90% in serum IgM levels denoted minor (MR), partial (PR), and very good partial (VGPR) responses. Normalization of serum IgM level; no monoclonal IgM spike, BM disease involvement, or pathological adenopathy or splenomegaly was required for complete response (CR). Overall response rate included MR or better, major response included PR or better, and deep response included VGPR or better. Disease progression was defined as an increase in serum

IgM level of ≥25% with an increase of ≥500 mg/dl from the lowest serum IgM level attained on therapy. Transient increases in serum IgM level while on a temporary hold of ibrutinib therapy were not considered disease progression. At our institution, *MYD88* and *CXCR4* mutations were assessed using the Rapid Heme Panel, a custom next-generation sequencing (NGS) panel for hematologic malignancies.<sup>18</sup> Additionally, *MYD88 L265P* and nonsense *CXCR4* mutations were assessed using allele-specific polymerase chain reaction (PCR) assays and frameshift *CXCR4* mutations by Sanger sequencing in CD19-selected BM samples, as previously reported.<sup>2,3,19,20</sup> PFS was defined as the time between ibrutinib initiation and disease progression, last follow-up or death from any cause, and OS after ibrutinib as the time between ibrutinib initiation and last follow-up or death from any cause. The IPSSWM was estimated as previously reported.<sup>21</sup> Missing data had a random distribution and rates were as follows: hemoglobin level (n = 4; 1%), platelet count (n = 6; 2%), serum β2-microglobulin level (n = 68; 21%), serum albumin level (n = 16; 5%), BM involvement (n = 41; 13%), IPSSWM (n = 51; 16%), *MYD88* mutational status (n = 46; 14%), *CXCR4* mutational status (n = 75; 24%).

**Statistical analysis.** Patients' characteristics and response rates are presented using descriptive statistics. Differences between categorical variables were assessed using Fisher exact test or χ-square test, depending on the number of events. Survival curves were generated using the Kaplan-Meier method and compared using the log-rank test. Univariate and multivariate logistic regression models were fitted to identify predictive factors of major and deep response to ibrutinib. Logistic regression outcomes are reported using odds ratio (OR) with 95% confidence interval (CI). Univariate and multivariate Cox proportional-hazard regression models were fitted to identify prognostic factors of PFS and OS. Cox regression outcomes are reported using hazard ratio (HR) with 95% CI. Only factors with *P* values <.05 in the univariate analysis were included in the multivariate analysis. We handled missing data by performing multiple imputation analyses in addition to complete case analyses. Multiple imputation analyses for logistic and Cox proportional-hazard regression models were performed after generating 20 imputed data sets using the chained equations method. *P* values <.05 were considered statistically significant. Calculations were obtained using STATA 17 (StataCorp, College Station, TX).

## Results

### Patients' characteristics

A total of 380 patients were identified for this study. After excluding 29 patients who received ibrutinib in combination with other agents (25 patients with Bing-Neel syndrome) and 7 patients with non-IgM lymphoplasmacytic lymphoma, 319 patients with WM were included in the analysis. The patients' baseline characteristics are shown in Table 1. Indications for ibrutinib monotherapy initiation are shown in supplemental Table S1. *CXCR4* mutations were detected in 89 patients, of which 56 carried a nonsense mutation, 31 a frameshift mutation, and 2 had concurrent nonsense and frameshift mutations. Given the small sample size of the latter group, we excluded these patients from subsequent analyses. A higher proportion of platelet count <100 K/uL was observed in patients with vs without *CXCR4* mutations (24% vs 6%; *P* < .001). Moreover, nonsignificantly higher proportion of patients with serum IgM level ≥4000 mg/dl (48% vs

**Table 1. Patients' baseline characteristics**

Characteristic	n (%) or median (range)
Age at WM diagnosis, years	61 (35-91)
Age at ibrutinib initiation, years	68 (40-96)
Age >65 years	190 (60)
Male sex	206 (65)
Hemoglobin level, g/dl	10.3 (4-17)
Hemoglobin <11.5 g/dl	226 (72)
Platelet count, K/uL	211 (9-639)
Platelet count <100 K/uL	42 (13)
Serum IgM level, mg/dl	3400 (88*10 321)
Serum IgM level >4000 mg/dl	129 (40)
Serum IgM level >7000 mg/dl	15 (5)
Serum $\beta$ 2-microglobulin level, mg/l	3.6 (1.4-18.3)
Serum $\beta$ 2-microglobulin level >3 mg/l	164 (65)
Serum albumin level, g/dl	3.7 (2.1-5)
Serum albumin level $\leq$ 3.5 g/dl	92 (30)
BM involvement	60 (5-100)
BM involvement $\geq$ 60%	154 (55)
IPSSWM	
Low	64 (24)
Intermediate	92 (34)
High	112 (42)
MYD88 mutated	265 (97)
CXCR4 mutated	89 (36)
Previously untreated	100 (31)
Previously treated	219 (69)
Median number of prior lines	2 (1-8)
$\geq$ 2 prior lines	115 (53)

\*Six patients with normal serum IgM levels had an IgM monoclonal spike in the serum protein electrophoresis.

37%;  $P = .10$ ) and BM involvement  $\geq$ 60% (64% vs 52%;  $P = .08$ ) were observed in patients with vs without *CXCR4* mutations. Patients' characteristics according to *CXCR4* mutational status are shown in supplemental Table S2. There was a trend toward a lower proportion of hemoglobin  $\leq$ 11.5 g/dl (68% vs 82%;  $P = .10$ ) and BM involvement  $\geq$ 60% (58% vs 79%;  $P = .07$ ) in patients with nonsense vs frameshift *CXCR4* mutations. Patients' characteristics according to *CXCR4* mutation subtypes are shown in supplemental Table S3.

The median follow-up time for the entire cohort was 4.2 years (95% CI, 3.9-4.5). No difference was observed between patients with and without *CXCR4* mutations (4.1 years, 95% CI, 3.6-4.4 vs 3.9 years, 95% CI, 3.3-4.5;  $P = .54$ ).

### Response to ibrutinib monotherapy

At best response, 48 patients (15%) attained MR, 161 (50%) attained PR, 87 (27%) attained VGPR, and 1 (0.3%) attained CR. Overall response was attained in 297 patients (93%), major response in 249 (78%), and deep response in 88 (28%) (supplemental Figure 1A). The rates of major response for patients with and without *CXCR4* mutations were 67% and 86%, respectively

( $P < .001$ ). The rates of deep response for patients with and without *CXCR4* mutations were 16% and 35%, respectively ( $P = .001$ ). Categorical response distribution by *CXCR4* mutational status is shown in supplemental Figure 1B. The rates of major response for patients with nonsense and frameshift *CXCR4* mutations were 59% and 81%, respectively ( $P = .04$ ). The rates of deep response for patients with nonsense and frameshift *CXCR4* mutations were 13% and 23%, respectively ( $P = .22$ ). Categorical response distribution by *CXCR4* mutation subtype is shown in supplemental Figure 1C.

Logistic regression models for major and deep responses are shown in Table 2. Univariate regression analyses showed that hemoglobin level <11.5 g/dl,  $\beta$ 2-microglobulin level >3 mg/l, and BM involvement  $\geq$ 60% were associated with higher odds, while nonsense *CXCR4* mutations were associated with lower odds of attaining major response to ibrutinib monotherapy. The IPSSWM was not predictive of major response ( $P = .40$ ). In the multivariate analysis evaluating these 4 variables, *CXCR4* mutations were independently associated with lower odds of attaining a major response (OR, 0.20; 95% CI, 0.09-0.45;  $P < .001$ ). In addition, hemoglobin <11.5 g/dl was independently associated with higher odds of major response (OR, 2.72; 95% CI, 1.25-5.95;  $P = .01$ ). Univariate analyses showed that hemoglobin  $\leq$ 11.5 g/dl was associated with higher, and *CXCR4* mutations were associated with lower odds of attaining deep response to ibrutinib monotherapy. The IPSSWM was not predictive of deep response ( $P = .29$ ). A multivariate model evaluating hemoglobin and *CXCR4* mutations showed that *CXCR4* mutations were independently associated with lower odds of attaining a deep response (OR, 0.32; 95% CI, 0.15-0.55;  $P = .001$ ). Hemoglobin <11.5 g/dl was independently associated with higher odds of deep response (OR, 2.34; 95% CI, 1.14-4.80;  $P = .02$ ). The multiple imputation analysis did not modify our results (data not shown).

In addition, we fitted univariate and multivariate models for major and deep response using continuous variables for age at ibrutinib initiation, hemoglobin level, platelet count, serum  $\beta$ 2-microglobulin, albumin, IgM levels, and percentage of BM involvement, in addition to categorical variables for sex, previous treatment, and *CXCR4* mutational status, which confirmed the predictive value of *CXCR4* mutations, especially nonsense *CXCR4* mutations, for lower odds of major and deep responses to ibrutinib monotherapy (supplemental Table S4).

### PFS analysis and risk stratification

At the time of this report, 93 patients (30%) had progressed or died. The median PFS was 6.5 years (95% CI, 6.0-not reached), and the estimated 5-year PFS rate was 60% (95% CI, 51%-68%) (Figure 1A). Univariate and multivariate regression models for PFS are shown in Table 3, top. In the univariate analysis for PFS, platelet count  $\leq$ 100 K/uL and *CXCR4* mutations were associated with worse PFS. The IPSSWM was not prognostic of PFS, as there were no differences between intermediate and low risk ( $P = .19$ ) and high and intermediate risk ( $P = .13$ ). In the multivariate analysis, platelet count  $\leq$ 100 K/uL and *CXCR4* mutations were independent factors associated with worse PFS. There was a poor correlation between platelet count  $\leq$ 100 K/uL and the presence of *CXCR4* mutations ( $r = 0.28$ ). Platelet count  $\leq$ 100 K/uL was associated with HR 2.51 (95% CI, 1.28-4.89;  $P = .007$ ) and *CXCR4* mutations with HR 1.98 (95% CI, 1.17-3.36;  $P = .01$ ). The multiple imputation analysis did not change our results. For patients with platelet count  $\leq$ 100 K/uL and >100 K/uL, median PFS was 4.4 years

**Table 2. Univariate and multivariate logistic regression analyses for major and deep response in 319 patients with WM treated with ibrutinib monotherapy**

Major response Variables	Univariate analysis		Multivariate analysis	
	OR (95% CI)	P	OR (95% CI)	P
Age >65 years	1.53 (0.90-2.61)	.12		
Male sex	1.19 (0.69-2.06)	.53		
Hemoglobin <11.5 g/dl	2.25 (1.28-3.93)	.005	2.72 (1.25-5.95)	.01
Platelet <100 k/ul	0.50 (0.25-2.45)	.05		
β2-microglobulin >3 mg/l	2.41 (1.31-4.43)	.005	1.54 (0.68-3.45)	.30
Albumin <3.5 g/dl	1.69 (0.88-3.24)	.12		
Serum IgM >4000 mg/dl	1.29 (0.74-2.24)	.36		
Serum IgM >7000 mg/dl	0.40 (0.14-1.17)	.09		
Bone marrow >60%	2.03 (1.14-3.62)	.02	2.26 (0.98-5.21)	.06
Previously treated	0.92 (0.52-1.64)	.78		
<i>CXCR4</i> mutated	0.33 (0.17-0.61)	<.001	0.20 (0.09-0.45)	<.001
<i>CXCR4</i> WT	1.00		1.00	
<i>CXCR4</i> NS	0.24 (0.12-0.48)	<.001	0.19 (0.01-0.37)	<.001
<i>CXCR4</i> FS	0.69 (0.25-1.87)	.47	0.40 (0.13-1.23)	.11

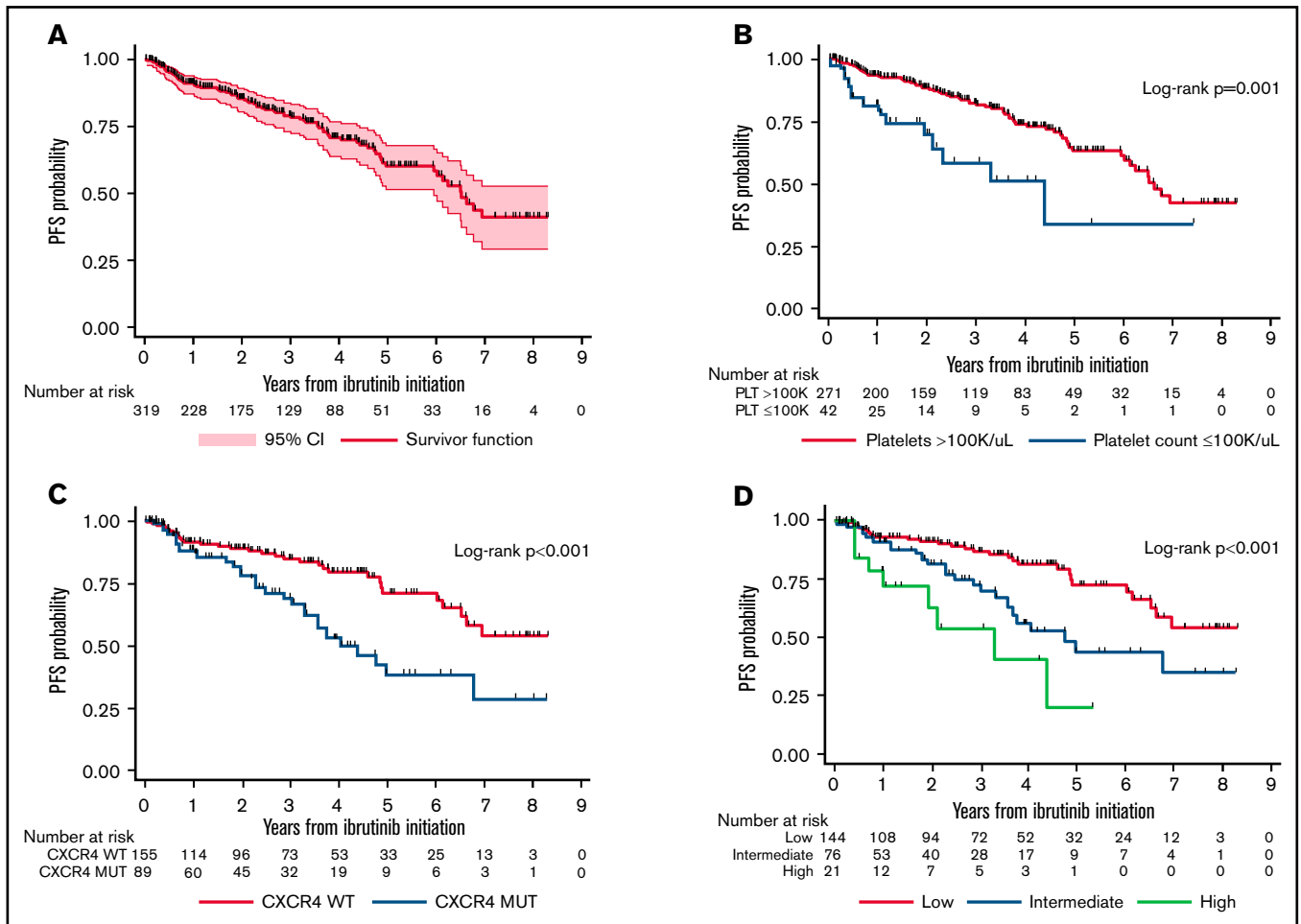
Deep response Variables	Univariate analysis		Multivariate analysis	
	OR (95% CI)	P	OR (95% CI)	P
Age >65 years	1.04 (0.63-1.71)	.88		
Male sex	1.08 (0.65-1.82)	.76		
Hemoglobin <11.5 g/dl	1.94 (1.07-3.53)	.03	2.34 (1.14-4.80)	.02
Platelet <100 k/ul	1.03 (0.50-2.11)	.94		
β2-microglobulin >3 mg/l	1.46 (0.81-2.64)	.21		
Albumin <3.5 g/dl	1.70 (1.01-2.88)	.05		
Serum IgM >4000 mg/dl	0.60 (0.36-1.01)	.05		
Serum IgM >7000 mg/dl	0.64 (0.18-2.34)	.50		
Bone marrow >60%	1.65 (0.96-2.83)	.07		
Previously treated	1.31 (0.76-2.25)	.33		
<i>CXCR4</i> mutated	0.34 (0.18-0.66)	.001	0.32 (0.15-0.55)	.001
<i>CXCR4</i> WT	1.00		1.00	
<i>CXCR4</i> NS	0.26 (0.11-0.61)	.002	0.25 (0.11-0.60)	.002
<i>CXCR4</i> FS	0.53 (0.21-1.31)	.17	0.47 (0.19-1.16)	.11

MYD88 mutational status was not included as sample size for *MYD88* wild-type status was small (n = 8). FS, frameshift; NS, nonsense; WT, wild type.

(95% CI, 0.9-not reached) and 6.6 years (95% CI, 6.0-not reached), respectively, and the estimated 5-year PFS rates were 34% (95% CI, 8%-63%) and 63% (95% CI, 54%-71%), respectively (log-rank  $P = .001$ ) (Figure 1B). For patients with and without *CXCR4* mutations, median PFS was 4.4 years (95% CI, 3.3-6.8) and not reached, respectively, and the estimated 5-year PFS rates were 39% (95% CI, 23%-54%) and 71% (95% CI, 59%-80%), respectively (log-rank  $P < .001$ ) (Figure 1C). There was no statistically significant difference in PFS between patients with nonsense and frameshift *CXCR4* mutations ( $P = .10$ ).

We then evaluated a PFS scoring system in WM patients on ibrutinib monotherapy using platelet count  $\leq 100$  K/uL and *CXCR4* mutations as adverse prognostic factors. This analysis included 241 WM patients with available data; 144 patients (60%) had no

adverse factors, 76 patients (32%) had 1 adverse factor, and 21 patients (9%) had 2 adverse factors (Table 4). Patients with 0, 1, and 2 adverse factors had median PFS not reached, 4.8 years (95% CI, 3.6-not evaluable) and 3.3 years (95% CI, 1.0-not evaluable), respectively, and estimated 5-year PFS rates of 72% (95% CI, 60%-82%), 44% (95% CI, 27%-60%), and 20% (95% CI, 1%-55%), respectively (log-rank  $P < .001$ ) (Figure 1D). In a univariate Cox proportional-hazard regression model, patients with 1 adverse factor had an HR of 2.21 (95% CI, 1.29-3.80;  $P = .004$ ), and patients with 2 adverse factors had an HR of 4.70 (95% CI, 2.18-10.1;  $P < .001$ ) when compared with patients without adverse factors. Patients with 2 adverse factors had an HR of 2.33 (95% CI, 1.07-5.07;  $P = .03$ ) compared with patients with 1 adverse factor. The multiple imputation analysis did not alter our results.



**Figure 1.** Progression-free survival (PFS) estimates in 319 patients with WM treated with ibrutinib monotherapy, for the entire cohort (A), according to platelet count (B), according to *CXCR4* mutational status (C), and according to the proposed PFS scoring system (D).

The proposed PFS scoring system was independently significant after adjusting for age, sex, serum IgM level, hemoglobin level, serum  $\beta$ 2-microglobulin level, serum albumin level, BM involvement, and prior therapy status ( $P < .05$  in all instances). PFS estimates according to the proposed PFS scoring system stratified by age, serum IgM level, hemoglobin level, and prior therapy status are shown in supplemental Figure S2.

### OS survival analysis

Fifty-three patients (16%) in our cohort have died. Causes of death are listed in supplemental Table S5. The median OS was not reached, and the estimated 5-year OS rate was 75% (95% CI, 67%-81%) (Figure 2A). There was no statistically significant difference in OS between patients with and without *CXCR4* mutations ( $P = .27$ ) or between patients with nonsense and frameshift *CXCR4* mutations ( $P = .91$ ). Univariate and multivariate Cox proportional-hazard regression models for OS are shown in Table 3. In the univariate regression analysis, age  $>65$  years, hemoglobin level  $<11.5$  g/dl, platelet count  $<100$  K/uL, serum  $\beta$ 2-microglobulin  $>3$  mg/l, and serum albumin level  $<3.5$  g/dl were associated with a worse OS. In the multivariate analysis, age  $>65$  years was the only independent factor associated with a worse OS (HR, 3.15; 95% CI, 1.41-7.04;  $P = .005$ ). The multiple imputation

analysis did not change our results. For patients age  $>65$  years (median age 73, range 66 to 96) and  $\leq 65$  years (median age 59, range 40 to 65), the median OS was 6.9 years (95% CI, 5.2-not evaluable) and not reached, and the estimated 5-year OS rates were 63% (95% CI, 50% to 73%) and 87% (95% CI, 78% to 93%), respectively (Figure 2B). Given the strong prognostic value of age in OS, which could have masked the prognostic value of other factors, we fitted a separate multivariate model including the variables associated with worse OS in the univariate analysis but excluding age. In this model, platelet count  $\leq 100$  K/uL (HR, 2.21; 95% CI, 1.01-4.85;  $P = .047$ ) and serum albumin level  $<3.5$  g/dl (HR, 2.06; 95% CI, 1.08-3.93;  $P = .03$ ) were independently associated with a worse OS.

We then evaluated the IPSSWM as a prognostic tool in 89 patients with WM who received ibrutinib as primary therapy and had IPSSWM data available, of which 8 patients (9%) have died. The median follow-up for these patients was 3 years (95% CI, 2-3.5). The 3-year OS rates for patients with low-, intermediate-, and high-risk disease were 92% (54% to 99%), 96% (73% to 99%) and 87% (71% to 94%), respectively ( $P = .74$ ) (Figure 2C). When compared with patients with low-risk disease, patients with intermediate- and high-risk disease had HR of death of 1.73 (95% CI, 0.16-19.1;  $P = .65$ ) and 2.24 (95% CI, 0.26-19.2;  $P = .46$ ), respectively. High-risk

**Table 3. Univariate and multivariate Cox proportional-hazard regression analyses for PFS and OS in 319 patients with WM treated with ibrutinib monotherapy**

PFS Variables	Univariate analysis		Multivariate analysis	
	HR (95% CI)	P	HR (95% CI)	P
Age >65 years	1.21 (0.77-1.89)	.41		
Male sex	1.09 (0.67-1.78)	.72		
Hemoglobin <11.5 g/dl	1.40 (0.84-2.32)	.19		
Platelet <100 k/ul	2.55 (1.42-4.59)	.002	2.51 (1.28-4.89)	.007
β2-microglobulin >3 mg/l	1.48 (0.84-2.61)	.17		
Albumin <3.5 g/dl	1.57 (0.97-2.57)	.07		
Serum IgM >4000 mg/dl	0.90 (0.58-1.41)	.64		
Serum IgM >7000 mg/dl	1.90 (0.76-4.73)	.17		
Bone marrow >60%	0.75 (0.47-1.22)	.25		
Previously treated	1.40 (0.77-2.53)	.27		
<i>CXCR4</i> mutated	2.31 (1.41-3.79)	.001	1.98 (1.17-3.36)	.01
<i>CXCR4</i> WT	1.00			
<i>CXCR4</i> NS	2.93 (1.71-5.00)	<.001	2.56 (1.54-4.50)	.001
<i>CXCR4</i> FS	1.49 (0.68-3.23)	.32	1.22 (0.55-2.74)	.63

OS Variables	Univariate analysis		Multivariate analysis	
	HR (95% CI)	P	HR (95% CI)	P
Age >65 years	3.03 (1.61-5.70)	.001	3.15 (1.41-7.04)	.005
Male sex	1.55 (0.81-2.95)	.18		
Hemoglobin <11.5 g/dl	2.36 (1.15-4.84)	.02	2.00 (0.81-4.93)	.13
Platelet <100 k/ul	3.10 (1.61-5.97)	.001	2.05 (0.94-4.48)	.07
β2-microglobulin >3 mg/l	3.01 (1.27-7.18)	.01	1.69 (0.69-4.14)	.25
Albumin <3.5 g/dl	2.40 (1.37-4.24)	.002	1.65 (0.85-3.19)	.14
Serum IgM >4000 mg/dl	1.39 (0.81-2.39)	.23		
Serum IgM >7000 mg/dl	2.07 (0.74-5.77)	.17		
Bone marrow >60%	0.87 (0.49-1.54)	.63		
Previously treated	1.46 (0.67-3.15)	.34		
<i>CXCR4</i> mutated	1.37 (0.74-2.53)	.31		
<i>CXCR4</i> WT	1.00			
<i>CXCR4</i> NS	1.42 (0.71-2.85)	.32		
<i>CXCR4</i> FS	1.49 (0.61-3.64)	.39		

MYD88 mutational status was not included as the sample size for *MYD88* wild-type status was small (n = 8). See Table 2 for definitions.

patients had HR 1.29 (95% CI, 0.25-6.67;  $P = .76$ ) vs intermediate-risk patients. The multiple imputation analysis did not change our results. The proposed PFS scoring system was not prognostic of OS

**Table 4. Proposed prognostic score for PFS for patients with WM treated with ibrutinib monotherapy**

Variables	HR (95% CI)	P
<i>CXCR4</i> mutations	2.51 (1.28-4.89)	.001
Platelet count <100 K/ul	1.98 (1.17-3.36)	.07

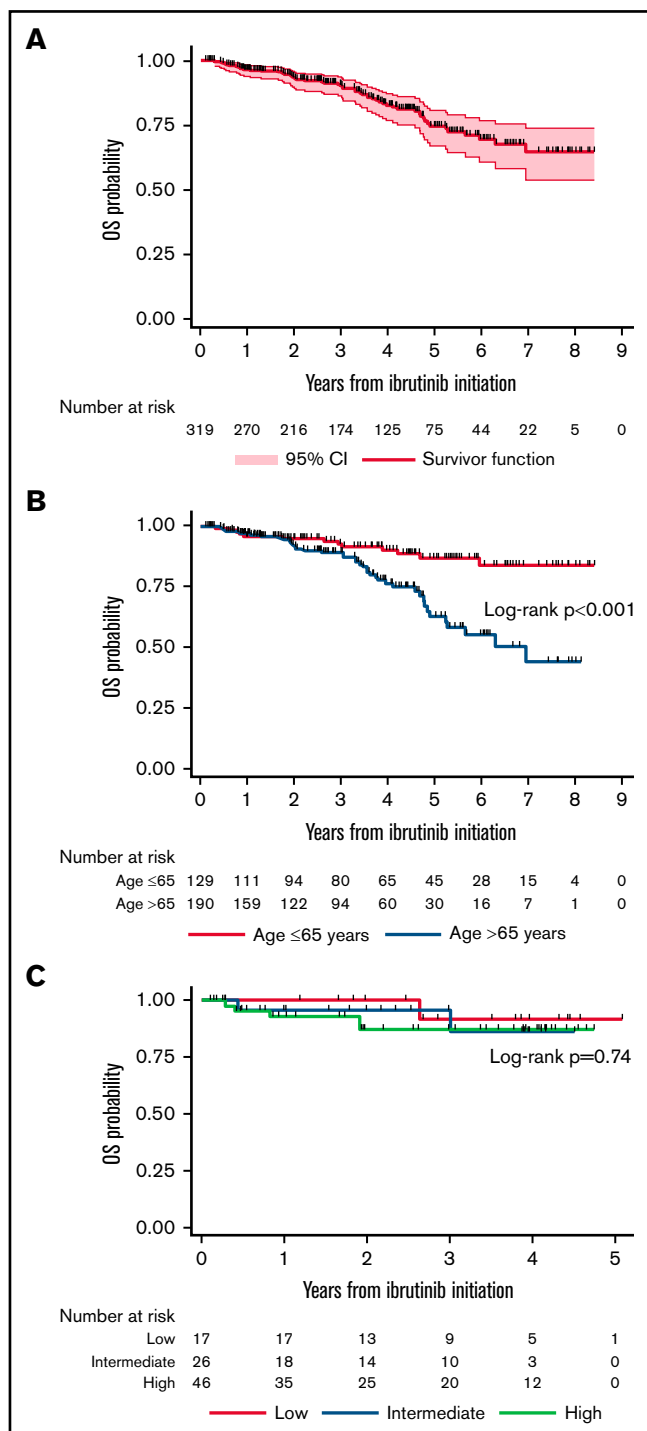
Stratum	Score	n (%)	Failed	5-y PFS rate, % (95% CI)	HR (95% CI)
Low	0	144 (60)	28	72 (60-82)	1.0 (Ref)
Intermediate	1	76 (32)	25	44 (27-60)	2.2 (1.3-3.8)
High	2	21 (9)	9	20 (1-55)	4.7 (2.2-10.1)

in patients who received ibrutinib as primary therapy ( $P = .54$  for the entire model).

In addition, we fitted univariate and multivariate models for PFS and OS using continuous variables for age at ibrutinib initiation, hemoglobin level, platelet count, serum β2-microglobulin, albumin, IgM levels, and percentage of BM involvement, in addition to categorical variables for sex, previous treatment, and *CXCR4* mutational status, which confirmed the adverse prognostic value of *CXCR4* mutations, especially nonsense *CXCR4* mutations, in PFS but no effect in OS on ibrutinib monotherapy (supplemental Table S6).

## Discussion

In the present study, we aimed at evaluating factors predictive of response and prognostic of survival in WM patients treated with



**Figure 2.** Overall survival estimates in 319 patients with WM treated with ibrutinib monotherapy, for the entire cohort (A), according to age (B), and according to the IPSSWM in patients who received ibrutinib as primary therapy (C).

ibrutinib monotherapy. To the best of our knowledge, this is the largest effort to study these factors in WM patients treated with BTK inhibitors.

Our study showed having a *CXCR4* mutation was the only independent factor associated with lower odds of attaining deeper responses on ibrutinib monotherapy. In multivariate models adjusted

for relevant clinical factors, the odds of a major response were 80% lower, and the odds of a deep response were 65% lower in patients with than in patients without *CXCR4* mutations. Our findings are consistent with published experience in patients with WM treated with ibrutinib on and off clinical trials.<sup>9,10,12,13,22</sup> In the recently published ASPEN study, in which patients with WM were randomized to ibrutinib or zanubrutinib, approximately 10% of patients with *CXCR4* mutations attained a VGPR while the rate of VGPR in patients without *CXCR4* mutations was 20% to 30%.<sup>23</sup> Furthermore, in the INNOVATE study, which randomized patients with WM to ibrutinib plus rituximab and placebo plus rituximab, the VGPR rate or better to ibrutinib plus rituximab was lower in patients with *CXCR4* mutations (19% and 34%, respectively). Therefore, *CXCR4* mutations associate with lower rates of deep response in patients with WM treated with ibrutinib (alone and in combination with rituximab) or with other covalent BTK inhibitors.

Platelet count  $\leq 100$  K/uL and *CXCR4* mutations were independently associated with worse PFS. We proposed a risk stratification scoring system for PFS in which patients with no risk factors (low risk), 1 risk factor (intermediate risk), and 2 risk factors (high risk) had 5-year PFS rates of 72%, 45%, and 19%, respectively. Thrombocytopenia has previously been reported as an adverse prognostic factor for OS in patients with WM,<sup>21</sup> but thrombocytopenia has not previously been reported as an adverse marker for PFS on ibrutinib. Thrombocytopenia could represent either a higher BM burden of disease or a BM microenvironment with a specific cytokine profile that portends a mechanism of resistance to the ibrutinib effect.

*CXCR4* mutations have been previously associated with shorter PFS in WM patients treated with ibrutinib monotherapy.<sup>22</sup> In the INNOVATE study that evaluated ibrutinib plus rituximab combination therapy, there were no detectable differences in 30-month PFS rates between patients with and without *CXCR4* mutations.<sup>24</sup> However, at the 50-month follow-up update, the PFS rate appeared numerically higher in patients without *CXCR4* mutations at approximately 80% vs approximately 62% in patients with *CXCR4* mutations.<sup>25</sup> It is possible that the addition of rituximab would partially revert the adverse impact of *CXCR4* mutations when combined with ibrutinib in patients with WM. However, other interventions might be needed to optimize the treatment of patients with WM who harbor *CXCR4* mutations.

Preclinical studies in WM cell lines from our group showed that frameshift and nonsense *CXCR4* mutations were associated with a reduction of *CXCR4* receptor internalization following stimulation with the *CXCR4* ligand CXCL12.<sup>26</sup> Upon ibrutinib exposure, cell death induction was decreased in *CXCR4* mutation-harboring cells thought to be mediated by AKT and ERK activation. *CXCR4* internalization and subsequent cell death were partially regained by exposing these cells to *CXCR4* antagonists. Our findings support targeting *CXCR4* as a valid therapeutic mechanism in WM. A phase 1/2 study evaluated ibrutinib in combination with the anti-*CXCR4* monoclonal antibody ulocuplumab in 13 patients with WM and *CXCR4* mutations; 9 patients were previously untreated, and 7 had a nonsense mutation.<sup>27</sup> Major responses were attained in 100% of the patients with a median time to major response of 1.2 months, and the 2-year PFS rate was 90%. By comparison, ibrutinib therapy is associated with major response rates of 60%, a median time to major response of 4 to 6 months, and 2-year PFS rates of 30% to 70% in patients with WM and *CXCR4* mutations.<sup>12,22,25</sup> NCT04274738 is a phase 1 study evaluating

ibrutinib in combination with the small-molecule CXCR4 inhibitor mavorixafor.<sup>28</sup> The study is currently accruing participants in centers in the United States and Europe.

Based on population-based studies, the survival of patients with WM has improved in recent decades.<sup>29-31</sup> This improvement is likely multifactorial and associated with improved supportive therapies, increased patient disease-related education levels, higher medical practitioner awareness, and the advent of novel treatment options. The IPSSWM included age >65 years, hemoglobin level  $\leq 11.5$  g/dl, platelet count  $\leq 100$  K/uL, serum  $\beta$ -2-microglobulin level >3 mg/l, and monoclonal IgM level >7000 mg/dl and divided patients with low, intermediate, and high risk of overall mortality, though it was developed in a cohort of patients with WM with a low rate of exposure to nonchemotherapeutic approaches.<sup>21</sup> The IPSSWM was later validated in patients treated with rituximab-containing regimens.<sup>32</sup> However, current data do not support a prognostic value of the IPSSWM in patients treated with BTK inhibitors. The INNOVATE study did not show a prognostic value for PFS in patients with WM treated with ibrutinib plus rituximab, but OS was not evaluated.<sup>24</sup> Our study showed that the IPSSWM may be prognostic of OS in patients with WM treated with ibrutinib. However, the main driver of the prognosis appears to be the age at ibrutinib initiation, as age >65 years was the only adverse prognostic factor in a multivariate model. Additionally, the IPSSWM was not prognostic of OS in patients who received ibrutinib as primary therapy. However, the sample size was small, and the follow-up was relatively short. A larger sample size and longer follow-up are needed to better evaluate the prognostic value of the IPSSWM in patients with WM treated with BTK inhibitors as primary therapy.

Our study identified that a hemoglobin level  $\leq 11.5$  g/dl was associated with higher rates of major and deep responses to ibrutinib monotherapy, independent of CXCR4 mutational status. The rationale behind this finding is not well understood but may be associated with the role of CXCL13 in the biology of WM. CXCL13 is a chemokine produced by WM cells and can act as a chemotactic for mast cells into the BM microenvironment.<sup>33,34</sup> In a prior study from our group, high serum CXCL13 levels were inversely correlated with hemoglobin levels in patients with WM and directly associated with higher response rates to ibrutinib monotherapy.<sup>35</sup> The role of CXCL13 in WM needs further investigation.

Our study is not without limitations. All the patients included were evaluated, at least at one point in time, at a clinical center for national reference. However, the patients' characteristics from our cohort are representative of the general population of patients with WM, with a median age at treatment initiation of 68 years and a male-to-female ratio of 1.9:1. Given the retrospective nature of our study, variable proportions of missing data are expected. To minimize the bias introduced by missing data, we performed multiple imputation analyses in addition to complete-case analysis. In all cases, the analyses of

imputed data did not alter our results or resulted in stronger associations than the ones estimated by the complete-case analyses. Finally, the high sensitivity CXCR4 mutational testing performed at our institution, which combines NGS assays for all mutations, PCR assays for nonsense, and Sanger sequencing for frameshift mutations, might not be replicated elsewhere. However, our study permitted an evaluation of the true biological effect of CXCR4 mutations on ibrutinib outcomes. As previously reported, NGS can be associated with a high rate of false-negative results in patients with WM, especially in patients with a low tumor burden in the BM.<sup>36</sup> Standardization of CXCR4 mutational testing is warranted.

The US FDA recently approved zanubrutinib, a second-generation covalent BTK inhibitor for the treatment of patients with WM.<sup>37</sup> Other covalent BTK inhibitors, such as acalabrutinib and tirabrutinib, have also shown efficacy in WM.<sup>38,39</sup> Finally, noncovalent BTK inhibitors, such as pirtobrutinib, are under clinical evaluation in WM.<sup>40</sup> We believe the present scoring system could serve to predict responses and prognosticate PFS in patients with WM treated with zanubrutinib. Additional studies are needed, however, to confirm the prognostic value of our proposed score in patients with WM receiving other covalent and noncovalent BTK inhibitors.

We present a robust prognostic tool for PFS in patients with WM treated with ibrutinib, which could serve to guide prognostic discussions between practitioners, patients, and family members.

## Authorship

Contribution: J.J.C. designed the study, performed the analysis, and wrote the initial draft of the manuscript; J.J.C., S.R.S., C.A.F., C.R.L., T.P.W., and S.P.T. provided clinical care to the patients; J.J.C., J.N.G., S.R.S., and K.M. collected the data; M.L.G., A.K., X.L., M.M., N.T., Z.R.H., and C.J.P. ran the genomic studies; J.J.C., J.N.G., S.R.S., A.R.B., and S.P.T. analyzed the data; and all the authors approved the final version of the manuscript.

Conflict-of-interest disclosure: J.J.C. received research funds and/or honoraria from Abbvie, BeiGene, Janssen, Millennium, Pharmacyclics, Roche, and TG Therapeutics. A.R.B. received research funds and/or honoraria from Adaptive, Beigene, CSL Behring, Genzyme, Karyopharm, Pharmacyclics, and Sanofi. S.P.T. received research funds and/or honoraria from Beigene, BMS, Eli Lilly, Janssen, Pharmacyclics, and X4 Pharmaceuticals. All other authors declare no competing financial interests.

ORCID profiles: J.J.C., 0000-0001-9490-7532; S.R.S., 0000-0002-0075-6735; J.N.G., 0000-0001-6717-7669; Z.R.H., 0000-0002-1689-1691; A.R.B., 0000-0002-3868-9267; S.P.T., 0000-0001-6393-6154.

Correspondence: Jorge J. Castillo, 450 Brookline Ave, Mayer 221, Boston, MA 02215; e-mail: jorgej\_castillo@dfci.harvard.edu.

## References

1. Swerdlow SH, Cook JR, Sohani AR, et al. Lymphoplasmacytic lymphoma. In: Swerdlow SH, Campo E, Harris NL, et al, eds. WHO Classification of Tumours of Hematopoietic and Lymphoid Tissues. Lyon: IARC; 2017:232-235
2. Hunter ZR, Xu L, Yang G, et al. The genomic landscape of Waldenstrom macroglobulinemia is characterized by highly recurring MYD88 and WHIM-like CXCR4 mutations, and small somatic deletions associated with B-cell lymphomagenesis. *Blood*. 2014;123(11):1637-1646.



3. Treon SP, Xu L, Yang G, et al. MYD88 L265P somatic mutation in Waldenström's macroglobulinemia. *N Engl J Med*. 2012;367(9):826-833.
4. Yang G, Zhou Y, Liu X, et al. A mutation in MYD88 (L265P) supports the survival of lymphoplasmacytic cells by activation of Bruton tyrosine kinase in Waldenström macroglobulinemia. *Blood*. 2013;122(7):1222-1232.
5. Cao Y, Hunter ZR, Liu X, et al. CXCR4 WHIM-like frameshift and nonsense mutations promote ibrutinib resistance but do not supplant MYD88(L265P)-directed survival signalling in Waldenström macroglobulinaemia cells. *Br J Haematol*. 2015;168(5):701-707.
6. Treon SP, Cao Y, Xu L, Yang G, Liu X, Hunter ZR. Somatic mutations in MYD88 and CXCR4 are determinants of clinical presentation and overall survival in Waldenström macroglobulinemia. *Blood*. 2014;123(18):2791-2796.
7. Schmidt J, Federmann B, Schindler N, et al. MYD88 L265P and CXCR4 mutations in lymphoplasmacytic lymphoma identify cases with high disease activity. *Br J Haematol*. 2015;169(6):795-803.
8. Poulain S, Roumier C, Venet-Caillault A, et al. Genomic landscape of CXCR4 mutations in Waldenström macroglobulinemia. *Clin Cancer Res*. 2016;22(6):1480-1488.
9. Castillo JJ, Xu L, Gustine JN, et al. CXCR4 mutation subtypes impact response and survival outcomes in patients with Waldenström macroglobulinaemia treated with ibrutinib. *Br J Haematol*. 2019;187(3):356-363.
10. Treon SP, Tripsas CK, Meid K, et al. Ibrutinib in previously treated Waldenström's macroglobulinemia. *N Engl J Med*. 2015;372(15):1430-1440.
11. Dimopoulos MA, Trotman J, Tedeschi A, et al; iNOVATE Study Group and the European Consortium for Waldenström's Macroglobulinemia. Ibrutinib for patients with rituximab-refractory Waldenström's macroglobulinaemia (iNOVATE): an open-label substudy of an international, multi-centre, phase 3 trial. *Lancet Oncol*. 2017;18(2):241-250.
12. Treon SP, Gustine J, Meid K, et al. Ibrutinib monotherapy in symptomatic, treatment-naïve patients with Waldenström macroglobulinemia. *J Clin Oncol*. 2018;36(27):2755-2761.
13. Castillo JJ, Gustine JN, Meid K, et al. Response and survival outcomes to ibrutinib monotherapy for patients with Waldenström macroglobulinemia on and off clinical trials. *HemaSphere*. 2020;4(3):e363.
14. Abeykoon JP, Zanwar S, Ansell SM, et al. Ibrutinib monotherapy outside of clinical trial setting in Waldenström macroglobulinaemia: practice patterns, toxicities and outcomes. *Br J Haematol*. 2020;188(3):394-403.
15. Owen RG, Treon SP, Al-Katib A, et al. Clinicopathological definition of Waldenström's macroglobulinemia: consensus panel recommendations from the Second International Workshop on Waldenström's Macroglobulinemia. *Semin Oncol*. 2003;30(2):110-115.
16. Kyle RA, Treon SP, Alexanian R, et al. Prognostic markers and criteria to initiate therapy in Waldenström's macroglobulinemia: consensus panel recommendations from the Second International Workshop on Waldenström's Macroglobulinemia. *Semin Oncol*. 2003;30(2):116-120.
17. Owen RG, Kyle RA, Stone MJ, et al; Vllth International Workshop on Waldenström macroglobulinaemia. Response assessment in Waldenström macroglobulinaemia: update from the Vllth International Workshop. *Br J Haematol*. 2013;160(2):171-176.
18. Kluk MJ, Lindsley RC, Aster JC, et al. Validation and implementation of a custom next-generation sequencing clinical assay for hematologic malignancies. *J Mol Diagn*. 2016;18(4):507-515.
19. Xu L, Hunter ZR, Tsakmaklis N, et al. Clonal architecture of CXCR4 WHIM-like mutations in Waldenström macroglobulinaemia. *Br J Haematol*. 2016;172(5):735-744.
20. Xu L, Hunter ZR, Yang G, et al. MYD88 L265P in Waldenström macroglobulinemia, immunoglobulin M monoclonal gammopathy, and other B-cell lymphoproliferative disorders using conventional and quantitative allele-specific polymerase chain reaction [published correction appears in *Blood*. 2013;121(26):5259]. *Blood*. 2013;121(11):2051-2058.
21. Morel P, Duhamel A, Gobbi P, et al. International prognostic scoring system for Waldenström macroglobulinemia. *Blood*. 2009;113(18):4163-4170.
22. Treon SP, Meid K, Gustine J, et al. Long-term follow-up of ibrutinib monotherapy in symptomatic, previously treated patients With Waldenström macroglobulinemia. *J Clin Oncol*. 2021;39(6):565-575.
23. Tam CS, Opat S, D'Sa S, et al. A randomized phase 3 trial of zanubrutinib vs ibrutinib in symptomatic Waldenström macroglobulinemia: the ASPEN study. *Blood*. 2020;136(18):2038-2050.
24. Dimopoulos MA, Tedeschi A, Trotman J, et al; iNOVATE Study Group and the European Consortium for Waldenström's Macroglobulinemia. Phase 3 trial of ibrutinib plus rituximab in Waldenström's macroglobulinemia. *N Engl J Med*. 2018;378(25):2399-2410.
25. Buske C, Tedeschi A, Trotman J, et al. Five-year follow-up of ibrutinib plus rituximab vs placebo plus rituximab for Waldenström's macroglobulinemia: final analysis from the randomized phase 3 iNOVATETM Study. *Blood*. 2020;136(suppl 1):24-26.
26. Cao Y, Hunter ZR, Liu X, et al. The WHIM-like CXCR4(S338X) somatic mutation activates AKT and ERK, and promotes resistance to ibrutinib and other agents used in the treatment of Waldenström's macroglobulinemia. *Leukemia*. 2015;29(1):169-176.
27. Treon SP, Meid K, Hunter ZR, et al. Phase 1 study of ibrutinib and the CXCR4 antagonist ulocuplumab in CXCR4-mutated Waldenström macroglobulinemia. *Blood*. 2021;138(17):1535-1539.
28. Treon S, Buske C, Thomas S, et al. Preliminary clinical data from a phase 1B study of mavoxixafor and ibrutinib in patients with Waldenström macroglobulinemia with MYD88 and CXCR4 mutations. *Hematol Oncol*. 2021;39(suppl 2):Abstract 373.
29. Castillo JJ, Olszewski AJ, Cronin AM, Hunter ZR, Treon SP. Survival trends in Waldenström macroglobulinemia: an analysis of the Surveillance, Epidemiology and End Results database. *Blood*. 2014;123(25):3999-4000.
30. Castillo JJ, Olszewski AJ, Kanan S, Meid K, Hunter ZR, Treon SP. Overall survival and competing risks of death in patients with Waldenström macroglobulinaemia: an analysis of the Surveillance, Epidemiology and End Results database. *Br J Haematol*. 2015;169(1):81-89.

31. Kristinsson SY, Eloranta S, Dickman PW, et al. Patterns of survival in lymphoplasmacytic lymphoma/Waldenström macroglobulinemia: a population-based study of 1,555 patients diagnosed in Sweden from 1980 to 2005. *Am J Hematol*. 2013;88(1):60-65.
32. Kastritis E, Morel P, Duhamel A, et al. A revised international prognostic score system for Waldenström's macroglobulinemia. *Leukemia*. 2019;33(11):2654-2661.
33. Hunter ZR, Xu L, Yang G, et al. Transcriptome sequencing reveals a profile that corresponds to genomic variants in Waldenström macroglobulinemia. *Blood*. 2016;128(6):827-838.
34. Tripodo C, Gri G, Piccaluga PP, et al. Mast cells and Th17 cells contribute to the lymphoma-associated pro-inflammatory microenvironment of angioimmunoblastic T-cell lymphoma. *Am J Pathol*. 2010;177(2):792-802.
35. Vos JM, Tsakmaklis N, Patterson CJ, et al. CXCL13 levels are elevated in patients with Waldenström macroglobulinemia, and are predictive of major response to ibrutinib. *Haematologica*. 2017;102(11):e452-e455.
36. Gustine JN, Xu L, Yang G, et al. Bone marrow involvement and subclonal diversity impairs detection of mutated CXCR4 by diagnostic next-generation sequencing in Waldenström macroglobulinaemia. *Br J Haematol*. 2021;194(4):730-733.
37. U. S. Food & Drug Administration. FDA approves zanubrutinib for Waldenström's macroglobulinemia. Available at: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-zanubrutinib-waldenstroms-macroglobulinemia>. Accessed 8 January 2022.
38. Owen RG, McCarthy H, Rule S, et al. Acalabrutinib monotherapy in patients with Waldenström macroglobulinemia: a single-arm, multicentre, phase 2 study. *Lancet Haematol*. 2020;7(2):e112-e121.
39. Sekiguchi N, Rai S, Munakata W, et al. A multicenter, open-label, phase II study of tirabrutinib (ONO/GS-4059) in patients with Waldenström's macroglobulinemia [published correction appears in *Cancer Sci*. 2021;112(4):1669]. *Cancer Sci*. 2020;111(9):3327-3337.
40. Mato AR, Shah NN, Jurczak W, et al. Pirtobrutinib in relapsed or refractory B-cell malignancies (BRUIN): a phase 1/2 study. *Lancet*. 2021;397(10277):892-901.