

Circular RNA MAT2B Induces Colorectal Cancer Proliferation via Sponging miR-610, Resulting in an Increased E2F1 Expression

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Purpose: Recently, studies have demonstrated that a novel circular RNA (circRNA), circMAT2B, can promote cell proliferation and can thus contribute to the growth and development of hepatocellular carcinoma. However, the precise mechanisms underlying in circMAT2B-induced colorectal cancer (CRC) cell proliferation are not yet fully understood.

Materials and Methods: Quantitative reverse transcription polymerase chain reaction was conducted to evaluate circMAT2B expression in 70 CRC tissues and 70 matched adjacent normal tissues, CRC cell lines and human colonic epithelial cell line (NCM460). The direct interaction between miR-610 and circMAT2B or E2F1 was verified using luciferase reporter assay and biotinylated RNA Pull-down assay. Cell Counting Kit-8, colony formation assay, flow cytometry were utilized to examine the effect of circMAT2B, miR-610 and E2F1 on cell proliferation. Western blot was conducted to evaluate E2F1 expression.

Results: In our study, circMAT2B was found to be upregulated in CRC tissues and cell lines. Furthermore, the silencing of circMAT2B significantly inhibited proliferation. Hence, in order to investigate the mechanism underlying the oncogenic properties of circMAT2B in CRC, a bioinformatics analysis (circular RNA Interactome, <https://circinteractome.nia.nih.gov/>) was performed to screen the putative interacting microRNAs of circMAT2B. miR-610 was identified to be one of the potential targeted miRNAs of circMAT2B. Luciferase reporter and RNA pulldown assay confirmed a direct interaction between circMAT2B and miR-610. Moreover, circMAT2B expression was negatively correlated with the expression of miR-610 in CRC tissues ($r=-0.5131$, $p<0.0001$). Furthermore, we demonstrated circMAT2B upregulated expression levels of the miR-610 target gene E2F1, which is involved in cell proliferation, is overexpression in a broad range of human cancer including CRC. Further studies suggested that E2F1 upregulation could significantly reverse the si-circMAT2B-mediated inhibition of proliferation.

Conclusion: circMAT2B is upregulated in CRC tissues and cell lines. Moreover, circMAT2B promoted CRC proliferation by regulating the miR-610/E2F1 axis, which may serve as a potential therapeutic target for CRC treatment.

Keywords: circMAT2B, miR-610, E2F1, colorectal cancer, proliferation

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Introduction

Among malignant tumors, the incidence of colorectal cancer (CRC) is the second in males and third in females.¹ The preferred treatment for patients with colorectal cancer is surgical resection followed by adjuvant chemotherapy or radiotherapy. However, 30%–50% of patients undergoing curative resection have local and systemic recurrence, which is the leading cause of death in many colorectal cancer patients.²

Therefore, we need to find new therapeutic strategies, especially those that target disease-related molecular.

With the increasing number of genome-wide expression analysis research, a variety of circular RNAs (circRNAs) have gradually been identified, and these circRNAs have altered expression patterns in cancers including CRC.³ circRNAs is a new type of non-coding RNAs, which do not have 5' caps and 3' poly(A) tails, and form a closed circular structure through covalent bonds. Compared with linear RNA, circRNAs are more stable, abundant, conserved due to its special circular structure.^{4,5} Recently, studies have demonstrated that a novel circRNA, circMAT2B, can contribute to cell proliferation, thereby promoting the growth of hepatocellular carcinoma (HCC).⁶ However, the precise mechanisms of circMAT2B-induced CRC cell proliferation are unknown.

As is known, circRNAs can alter its own expression by sponging miRNAs or binding to proteins, resulting in abnormal expression of gene products, which may promote the occurrence of cancer.^{7,8} miR-610, located at 11p14.1, was a newly identified functional miRNA in tumors. It has been reported that the miR-610 inhibited the expression of CCND2 and AKT3, thereby repressing the proliferation of human glioblastoma cells.⁹ Studies have shown that the expression of miR-610 is significantly down-regulated in glioma cells, and miR-610 can block cell proliferation and metastasis by directly targeting MDM2.¹⁰ Reportedly, E2F transcription factor 1 (E2F1) is a member of the E2F family and plays an important role in controlling the cell cycle.¹¹ In addition, E2F1 preferentially binds to the retinoblastoma protein pRB in a cell cycle-dependent manner and mediates both cell proliferation and p53-dependent/independent apoptosis. More and more studies show that this regulatory pathway is abnormal in many human malignancies.¹²

This study showed that the expression of circMAT2B was significantly higher in CRC tissues than that in adjacent tissues. Furthermore, a negative correlation was found between the expression of circMAT2B and miR-610. CircMAT2B downregulates the level of miR-610 target gene E2F1, which mediate cell proliferation, through sponging miR-610, thus promoting CRC progression. Therefore, circMAT2B is considered as a promising target for CRC treatment.

Materials and Methods

Human Specimens

This study was approved by the Ethics Committee of the First People's Hospital of Taizhou and written informed consents were obtained from all patients. This was conducted in

accordance with the Declaration of Helsinki. The paired tissue samples from the surgical specimens of the First People's Hospital of Taizhou (Taizhou, China). In the study, the samples were stored using liquid nitrogen. Seventy CRC tissues and 70 adjacent normal colorectal tissues were collected.

Cell Lines

Seven human CRC cell lines including NCM460, HT29, HCT116, HCT8, SW480, DLD1, Caco2 were provided by the ATCC (American Type Culture Collection, Manassas, VA, USA). The cells were cultured in 1640 medium with 10% FBS (Gibco, Grand Island, NY, USA) at 37°C in an incubator containing 5% CO₂.

Oligonucleotide Transfection

The scrambled control, miR-610 mimic, miR-610 inhibitor, siRNA targeting circMAT2B and siRNA negative control were all produced by GenePharma Co. (Shanghai, China). The cDNA of E2F1 was subcloned into pcDNA3.1 vector by RiboBio (Guangzhou, China). HCT116 and HT29 cells (1×10^5 cells/well) were seeded in 6-well plates and then transfected with Lipofectamine 3000 from Invitrogen.

Quantitative Reverse Transcription Polymerase Chain Reaction (qPCR)

Using TRIzol reagent from Thermo Fisher Scientific, total RNA was extracted from CRC tissues and cells, then using the reverse transcription reagent kit from Vazyme Biotech (Nanjing, China), the cDNA was generated. In a 20 μ L reaction system, the qPCR was performed according to the instructions. Ten-microliter 2X qPCR SYBR-Green master mix (Vazyme Biotech), 0.4 μ L forward primer (10 μ M), 0.4 μ L reverse primer (10 μ M), 1 μ L cDNA and 0.4 μ L 50X ROX reference dye 1 were included in the 20 μ L reaction system. All specimens were tested in triplicate. All primers used in this study are listed in Table 1. The results were corrected by the expression of β -actin and the $2^{-\Delta\Delta Cq}$ method was used for the analysis of results.¹³

Cell Counting Kit-8 (CCK-8) and Colony Formation Assay

In a 96-well plate, 2×10^3 cells were seeded in each well. After cells and 10 μ L CCK-8 solution (Dojindo Laboratories, Kumamoto, Japan) were incubated at 37°C for 1 hr, the absorbance of the mixed solution at 450 nm was measured. In a 6-well plate, HCT116 and HT29 cells (500 per well) were seeded and cultured for 1–2 weeks. The colonies were

Table 1 Specific Primer for qPCR

Name		Primers(5'→3')
β-actin	Forward	CAGCCATGTACGTTGCTATCCAGG
	Reverse	AGGTCCAGACGCAGGATGGCATG
circMAT2B	Forward	GGAACAAATCCACCTTAC
	Reverse	TTCATTGCCAGACCAG
miR-610	Forward	GCCGAGTGAGCTAAATGTGT
	Reverse	CTCAACTGGTGTCTGTGGA
E2F1	Forward	AGCGGCGCATCTATGACATC
	Reverse	GTCAACCCCTCAAGCCGTC

stained with crystal violet solution and counted with a microscope. All tested were repeated three times.

Flow Cytometry

At 48 hrs after transfection, cells were collected and stained with Annexin V-Alexa Fluor 647 and PI (Fcmacs, Nanjing, China) for 15 mins. The processed samples were tested by Beckman CytOflex flow cytometer. Using CytExpert software, the percentage of apoptosis cells was analyzed.

Luciferase Reporter Assay

CircMAT2B sequences containing wild-type or mutant miR-610 binding sites were synthesized. The sequences were then inserted into psi-CHECK2 vector (Promega, Madison, WI, USA). Then, the wild-type or mutated vector and miR-610 mimics or mimics control were co-transfected into HCT116 and HT29 cells using Lipofectamine 3000 reagent (Invitrogen, Carlsbad, CA, USA), respectively. 48h after transfection, the luciferase activity of the collected cells was detected by the Dual-Luciferase Reporter Assay System from Promega.

Biotinylated RNA Pull-Down Assay

The pull-down assay with biotinylated RNA was operated according to the previous process.^{14,15} In short, in order to pull down miRNAs for circMAT2B, probe-coated beads were generated by co-cultivation of the biotinylated-circMAT2B probe and C-1 magnetic beads (Life Technologies, Carlsbad, CA, USA). Then, the probe-coated beads and sonicated HCT116 or HT29 cells were co-incubated at 4°C overnight, followed by elution and qPCR.

Western Blot

Samples were lysed in radioimmunoprecipitation buffer (Solarbio) to extract total protein. The protein concentration was detected by a bicinchoninic acid protein assay kit. The

extracted total proteins were separated using SDS-PAGE and then transferred onto polyvinylidene fluoride membranes (Merck KGaA, Darmstadt, Germany). The membranes were incubated with primary antibodies, and then with a secondary antibody. Protein detection was performed using an enhanced chemiluminescence substrate kit from Thermo Scientific and ImageJ software.

Statistical Analysis

SPSS 22.0 software was used for statistical analysis, and GraphPad Prism 6.0 was used for producing figures. All experimental data are expressed in the form of mean ± S.D, and calculated by Student's *t*-test as appropriate. Probability (*P*) values <0.05 were considered statistically significant.

Results

Elevated circMAT2B Expression Levels in CRC Tissue and Cell Lines

At the beginning of the study, we tested the expression level of circMAT2B in CRC tissues and cell lines, respectively. Using qPCR, we found that circMAT2B is significantly higher in the 70 cases of CRC (the clinical characteristics of the patients are summarized in Table 2) than in matched normal tissues (Figure 1A). Similar results were observed in CRC cell lines compared with NCM460 human colonic epithelial cells. Statistical analyses showed that CircMAT2B was markedly overexpressed in the HT29, HCT116, HCT8, SW480, DLD1, Caco2 cells, particularly in the HT29 and HCT116 cells (Figure 1B).

circMAT2B Knockdown Inhibits CRC Cells Proliferation

To explore the function of circMAT2B in the CRC process, we assessed the expression of circMAT2B in different CRC cell lines. Hence, we planned to knockdown the expression of circMAT2B in HCT116 and HT29 cell lines. Two siRNA targeting circMAT2B were designed and qPCR showed that these two siRNAs significantly decreased circMAT2B level (Figure 2A). Due to the highest efficiency of interference, si-circMAT2B#1 and si-circMAT2B#2 were used for the subsequent experiments. Silencing circMAT2B significantly inhibited the proliferation of CRC cell lines (Figure 2B-D).

circMAT2B Directly Binds to miR-610 in CRC

More and more studies have confirmed that circRNAs can act as ceRNAs to sponge miRNAs.¹⁶⁻¹⁸ Therefore, to further

Table 2 The Characteristics of the Patients with Colorectal Cancer (CRC)

Characteristics	Total	High Expression	Low Expression	χ^2	P value
Gender					
Male	39	21	18	0.521	0.470
Female	31	14	17		
Age					
≤60	39	16	23	2.837	0.092
>60	31	19	12		
Tumor size					
T1	19	4	15	15.438	0.001
T2	17	7	10		
T3	19	11	8		
T4	15	13	2		
Distant metastasis					
Positive	37	24	13	6.937	0.008
Negative	33	11	22		
Differentiation					
High	23	18	5	14.934	0.001
Moderate	21	11	10		
Poor	26	6	20		
Lymph node metastasis					
Positive	31	20	11	4.690	0.030
Negative	39	15	24		
TMN stages					
I	21	5	16	13.104	0.004
II	18	8	10		
III	18	11	7		
IV	13	11	2		

explore the potential carcinogenic mechanism of circMAT2B in CRC, a bioinformatics analysis (circular RNA Interactome, <https://circinteractome.nia.nih.gov/>) was performed to screen the putative interacting microRNAs of circMAT2B. miR-610

was identified to be one of the potential targeted miRNAs of circMAT2B (Figure 3A). To validate the direct interaction between miR-610 and circMAT2B, a dual-luciferase reporter assay was conducted by co-transfecting the recombinant

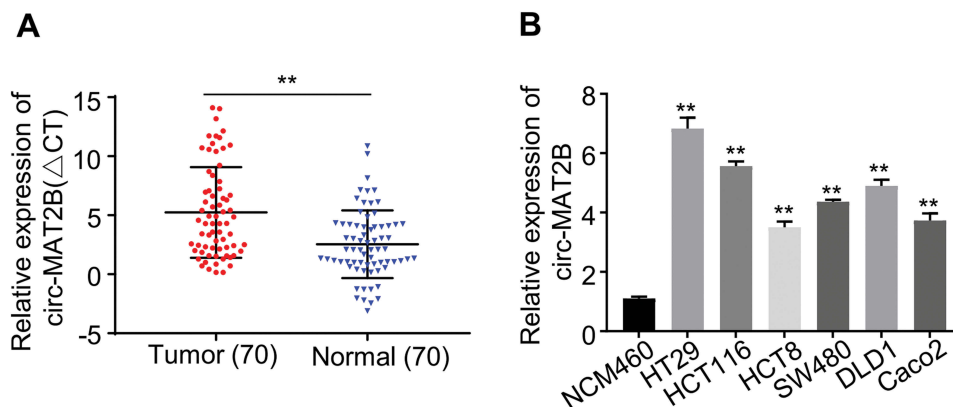


Figure 1 circMAT2B expression levels in CRC tissue and cell lines. **(A)** qPCR for the expression of circMAT2B in CRC tissues. **(B)** The circMAT2B expression in various CRC cell lines examined by qPCR. Data are listed as means ± S.D. of at least three independent experiments. **p<0.01.

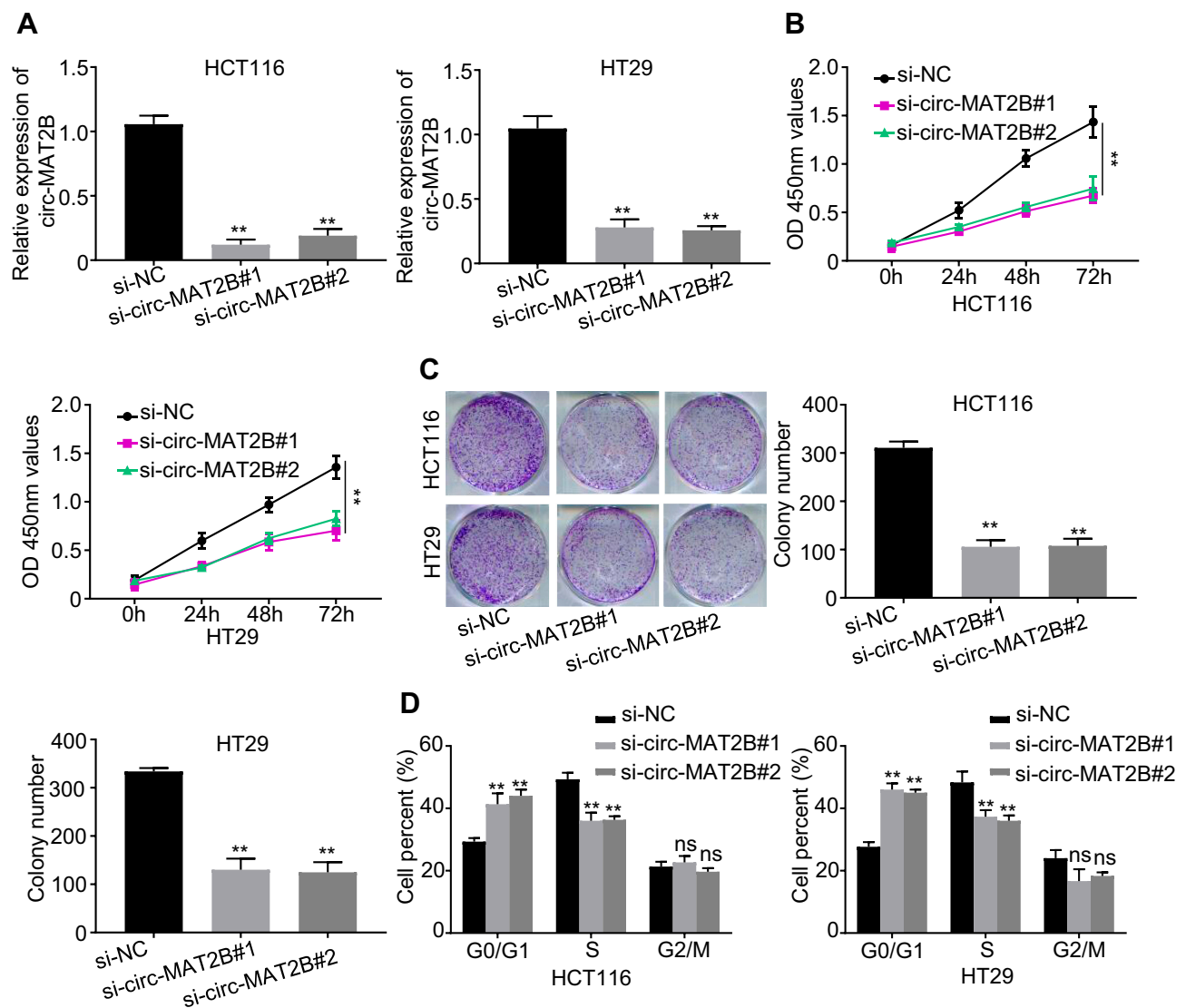


Figure 2 Silencing of circMAT2B inhibits CRC cell proliferation and cell cycle progression in vitro. **(A)** Expression levels of circMAT2B was detected by qPCR in HCT116 and HT29 cells transfected with siRNAs targeting the back-splice junction sequences of circMAT2B or negative control oligonucleotide (si-NC). **(B)** CCK-8 assay showed that the proliferation of HCT116 and HT29 cells was inhibited with circMAT2B silencing. **(C)** Colony formation assays suggested that circMAT2B knockdown inhibited the colony-forming ability of HCT116 and HT29 cells. **(D)** Cell cycle was examined by flow cytometry in HCT116 and HT29 cells with circMAT2B silencing. Data are listed as means \pm S.D. of at least three independent experiments. $**p < 0.01$.

plasmid containing the putative or mutant binding sites (Figure 3A) with miR-610 mimics or its negative control (miR-NC). The results showed that only the luciferase activity of wild-type circMAT2B could be attenuated by the miR-610 mimics (Figure 3A). Furthermore, biotin-coupled circMAT2B pulldown assay confirmed that miR-610 was found in the circMAT2B pulled down pellet compared with the biotin-coupled oligo group (Figure 3B), which indicates that circMAT2B could directly sponge miR-610. In addition, the expression of miR-610 was eminently upregulated when circMAT2B was silenced (Figure 3C). qPCR also confirmed that the expression of miR-610 was significantly lower

in CRC tissues than that in adjacent normal tissues (Figure 3D). In addition, circMAT2B expression in CRC tissues was negatively correlated with the expression of miR-610 ($r = -0.5131$, $p < 0.0001$) (Figure 3E). Taken together, these results demonstrate that circMAT2B directly binds to miR-610 in CRC.

circMAT2B Increases the Expression of Oncogene E2F1 by Sponging miR-610 in CRC

There is increasing evidence that miR-610 acts on multiple oncogenes simultaneously. Therefore, we explored that circMAT2B promotes tumor proliferation by regulating the

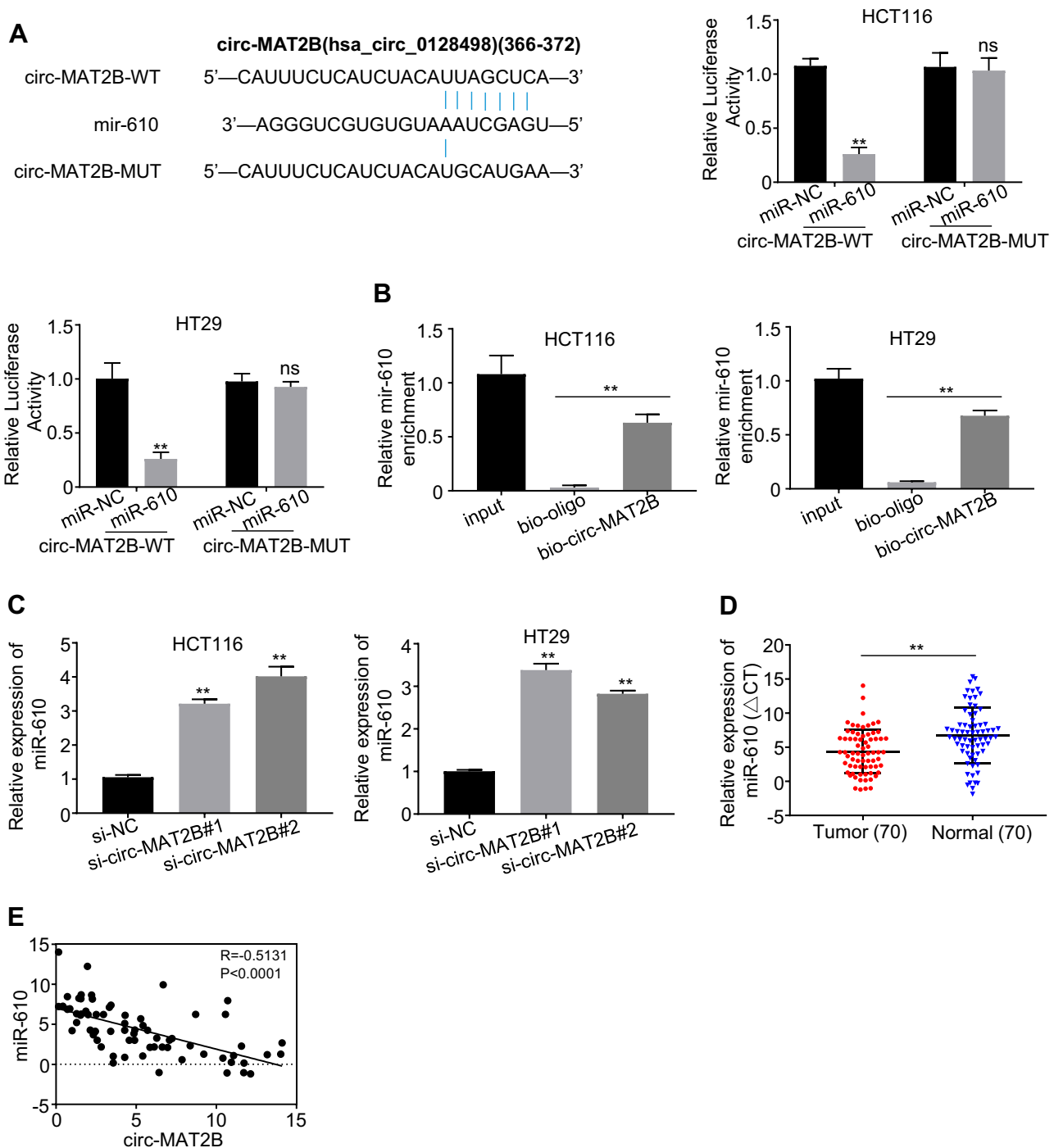


Figure 3 circMAT2B was directly targeted by miR-610. **(A)** Bioinformatics analysis methods were conducted to predict the putative binding sites of miR-610 in circMAT2B. Dual-luciferase reporter assay was applied to validate the interaction between miR-610 and circMAT2B. **(B)** miR-610 was pulled down and enriched with circMAT2B probe and then detected by qPCR in HCT116 and HT29 cells. **(C)** qPCR analysis of miR-610 expression in HCT116 and HT29 cells transfected with siRNAs targeting the back-splice junction sequences of circMAT2B or negative control oligonucleotide (si-NC). **(D)** qPCR analysis of miR-610 expression in CRC tissues (n=70). **(E)** The Pearson's correlation coefficients were used to evaluate the correlation between circMAT2B and miR-610 in CRC tissues (n = 70) ($r = -0.5131$, $p < 0.0001$). Data were represented as means \pm S.D. of at least three independent experiments. ** $p < 0.01$.

expression of miR-610 targeting oncogenes. Expected targets were determined using the bioinformatics tool miRanda. The analysis revealed that E2F1 3'-UTR has

miR-610 binding sites (Figure 4A). To confirm that miR-610 regulated E2F1, a luciferase-based reporter gene plasmid with WT or mutated 3'-UTR sequences for E2F1 was

constructed. The luciferase activity of E2F1 was significantly reduced when cells were co-transfection with miR-610 mimics and reporter plasmids. However, there was no evident effect on the luciferase activity in cells transfected with miR-610 mimics and mutated vectors (Figure 4A). In HCT116 and HT29 cells with circMAT2B knockdown, transcription of E2F1 was significantly decreased, and restored by using miR-610 inhibitor (Figure 4B). The above data indicated that miR-610 reduces the mRNA expression of E2F1 by targeting its 3'-UTR. Consistent with transcription data, Western blot analysis showed that E2F1 was regulated by circMAT2B and miR-610 in CRC cell lines (Figure 4C). Furthermore, qPCR also showed that E2F1 expression was significantly higher in CRC tissues than that in adjacent

normal tissues (Figure 4D). Moreover, the expression of E2F1 was positively correlated with circMAT2B expression in CRC tissues ($r=-0.5131$, $p<0.0001$) (Figure 4E). These results confirmed that E2F1 expression is regulated by miR-610 bound to its 3'UTR, and miR-610 is sponged by circMAT2B.

Silencing of miR-610 or Overexpression of E2F1 Remarkably Rescues the Decreased Proliferative Ability of CRC Cells Caused by circMAT2B Silencing

We further examined whether circMAT2B regulates the expression of oncogene E2F1 by sponging miR-610.

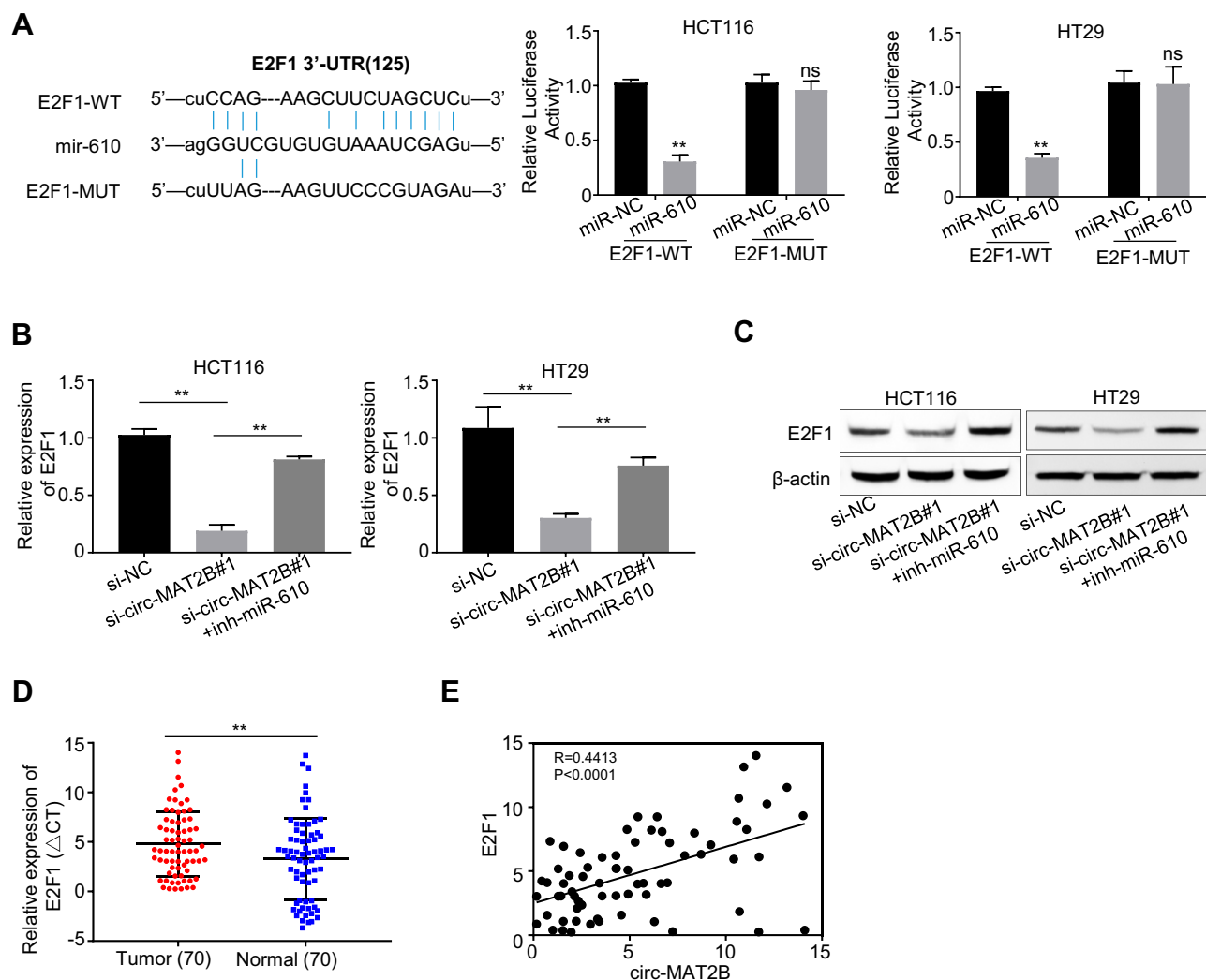


Figure 4 circMAT2B increases the expression of oncogene E2F1 by sponging miR-610 in CRC. (A) miR-610 and its putative binding sequence in the 3'-UTR of E2F1. The luciferase activity of E2F1 was evidently inhibited by miR-610. (B) qPCR analysis of the mRNA expression of E2F1 in HCT116 and HT29 cells. (C) Western blot analysis of the protein expression of E2F1 in HCT116 and HT29 cells. (D) qPCR analysis of E2F1 expression in CRC tissues (n=70). (E) The Pearson's correlation coefficients were used to evaluate the correlation between circMAT2B and E2F1 in CRC tissues (n = 70) ($r = 0.4413$, $p < 0.0001$). Data were represented as means \pm S.D. of at least three independent experiments. ** $p < 0.01$.

First, the HCT116 and HT29 cells were transfected with si-NC, si-circMAT2B#1, si-circMAT2B#1+inh-miR-610 and si-circMAT2B#1+E2F1. Next, CCK-8 assays, colony formation assays, and flow cytometry were used for the analysis of CRC proliferation. The data showed that the silencing of miR-610 or overexpression of E2F1 reversed the inhibiting effect of circMAT2B silencing on cell proliferation (Figure 5A-C) in CRC.

Discussion

For the past 30 years, circRNAs have been considered as noncoding RNAs (ncRNAs) and ignored as transcriptional noise of eukaryotes.^{19,20} However, due to the rapid development of bioinformatics, the widespread expression of circRNAs has been found in a variety of cells and

species.²¹ Moreover, more and more scientists recognize that circRNAs may play a vital role in the development of cancer and affect the hallmarks of cancer.^{16-18,22-25} However, the specific function of circRNAs in the development of CRC is still unclear. In this study, qPCR detected the expression of circMAT2B in 70 pairs of CRC and adjacent tissues, and the results showed that circMAT2B was significantly higher in CRC tissues. Moreover, negative correlations were also found between the expression of circMAT2B and miR-610. CircMAT2B downregulates the level of miR-610 target gene E2F1, which mediate cell proliferation, through sponging miR-610, thus promoting CRC progression. These results indicated that circMAT2B might be related to the progression of CRC.

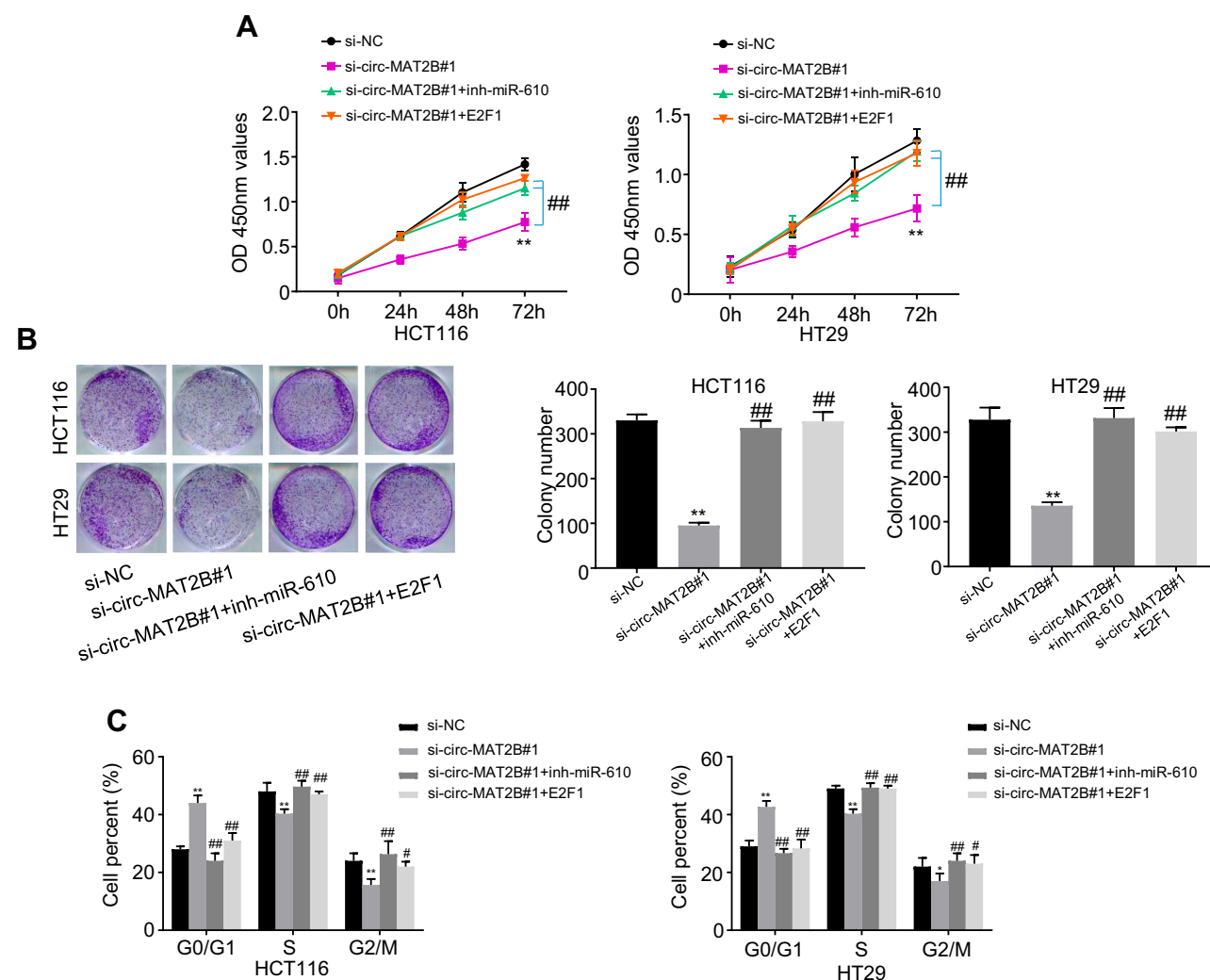


Figure 5 circMAT2B regulates cell proliferation in CRC via miR-610 and E2F1. (A) Proliferation in treated HCT116 and HT29 cells was detected by CCK-8 assays. (B) Colony-forming abilities in treated HCT116 and HT29 cells was detected by colony formation assays. (C) Cell cycle progression was examined by flow cytometry in HCT116 and HT29 cells with circMAT2B silencing. Data are listed as means \pm S.D. of at least three independent experiments. * $p < 0.05$, ** $p < 0.01$ (vs si-NC group); # $p < 0.05$, ### $p < 0.01$ (vs si-circMAT2B#1 group).

Research has shown that ubiquitin-like modifier-activating enzyme 2 (UBA2) expression may enhance CRC cell proliferation. Furthermore, the expression of UBA2 may be associated with p53/MDM2/p21 and PI3K/AKT signaling pathways.²⁶ Another study confirmed that miR-203a inhibits CRC proliferation by targeting RING-finger protein 6 (RNF6), offering novel insights into the regulatory network of miR-203a-modulated cell cycle and proliferation, and suggest that miR-203a was a potential therapeutic target in CRC treatment.²⁷ To explore the diagnostic values of circMAT2B in CRC, qPCR was used to measure circMAT2B expression in 70 pairs of CRC and matched adjacent tissues. The results confirmed that compared with matched adjacent tissues, the mRNA level of circMAT2B was significantly increased in CRC tissues. Moreover, the functional studies in CRC cell lines showed that circMAT2B promotes CRC cell proliferation, suggesting that circMAT2B might be related to the development of CRC and could become a promising therapeutic target for CRC. Consistent with this, a previous study has shown a significant increase in circMAT2B expression in HCC tissues and cell lines, and increased circMAT2B indicated poor prognosis, suggesting its tumor-promoting effect.⁶ The underlying mechanisms of circRNA function in cancer development and progression have not been clarified.²⁸ The most widely reported function of circRNAs is as miRNA sponges in the circRNA–miRNA–mRNA axes.^{29,30} In order to study the mechanism of the oncogenic properties of circMAT2B in CRC, the target miRNAs of circMAT2B was predicted by circular RNA Interactome software. The results suggested that circMAT2B contains a putative binding site for miR-610 based on the bioinformatics analysis. Furthermore, we confirmed that circMAT2B could sequester and inhibit miR-610 activity by sponging endogenous miR-610, which targets the oncogene E2F1, which is related to cell proliferation, and is overexpression in a variety of human cancer including CRC.^{31–33} Further studies showed that the down-regulation of miR-610 and the upregulation of E2F1 could effectively reverse the circMAT2B-mediated inhibition of proliferation in HCT116 and HT29 cells. Overall, our research confirmed that circMAT2B promoted CRC proliferation by sponging miR-610, which targets the oncogene E2F1.

In conclusion, oncogenic circMAT2B was identified, and the role and mechanism in CRC proliferation was investigated. All the above data showed that circMAT2B expression was significantly upregulated in CRC tissues

and cell lines. Moreover, circMAT2B could be used as an effective non-invasive biomarker for CRC detection. CircMAT2B accelerated CRC proliferation by regulating the miR-610/E2F1 axis, which may serve as a promising therapeutic target for CRC treatment.

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Disclosure

The authors declare that there is no conflict of interest.

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