What do we know about the role of IncRNAs in multiple sclerosis?

Viviana Nociti^{1,*}. Massimo Santoro²

https://doi.org/10.4103/1673-5374.306061

Date of acceptance: November 11, 2020

Date of web publication: January 25, 2021

Date of submission: July 5, 2020 Date of decision: August 21, 2020

Abstract

Multiple sclerosis is a chronic, inflammatory and degenerative disease of the central nervous system of unknown aetiology although well-defined evidence supports an autoimmune pathogenesis. So far, the exact mechanisms leading to autoimmune diseases are still only partially understood. We know that genetic, epigenetic, molecular, and cellular factors resulting in pathogenic inflammatory responses are certainly involved. Long non-coding RNAs (IncRNAs) are non-protein coding transcripts longer than 200 nucleotides that play an important role in both innate and acquired immunity, so there is great interest in IncRNAs involved in autoimmune diseases. The research on multiple sclerosis has been enriched with many studies on the molecular role of IncRNAs in the pathogenesis of the disease and their potential application as diagnostic and prognostic biomarkers. In particular, many multiple sclerosis fields of research are based on the identification of IncRNAs as possible biomarkers able to predict the onset of the disease, its activity degree, its progression phase and the response to disease-modifying drugs. Last but not least, studies on IncRNAs can provide a new molecular target for new therapies, missing, so far, a cure for multiple sclerosis. While our knowledge on the role of IncRNA in multiple sclerosis has recently improved, further studies are required to better understand the specific role of IncRNAs in this neurological disease. In this review, we present the most recent studies on molecular characterization of IncRNAs in multiple sclerosis disorder discussing their clinical relevance as biomarkers for diagnosis and treatments.

Key Words: antisense lncRNAs; enhancer lncRNAs; epigenetics; immune system; intergenic lncRNA; intronic lncRNA; multiple sclerosis; sense lncRNAs; single nucleotide polymorphisms

Introduction

Multiple sclerosis (MS), the second leading cause of sustained neurological disability in young people after trauma (Ontaneda et al., 2017), is a chronic, inflammatory and degenerative disease of the central nervous system (CNS) (Naegele et al., 2014) of unknown aetiology. So far, there are two theories on its pathogenesis: the "outside-in" autoimmune hypothesis, for which MS is an exclusive autoimmune inflammatory disease caused by unregulated auto-reactive immune cells that from the periphery go into the CNS parenchyma, attacking various cell types, and the "inside-out" hypothesis, for which MS is a primary degenerative disease in which inflammation is secondary to a release of auto-antigens promoting autoimmunity (Stys and Tsutsui, 2019). Until now, it remains to be determined whether inflammation is primary or secondary to a degenerative process in the brain. Nevertheless, well-defined evidence (as well as the successful use of immunomodulatory drugs in reducing clinical relapses and/or neuroradiological 'activity') demonstrates that an uncontrolled inflammatory response in the CNS is destructive in MS (Thompson et al., 2018). Sustained disability, however, is due to a progressive neurodegenerative process, causing axonal loss and brain atrophy, primary or secondary to the peripheral and compartmentalized inflammation in the CNS (Machado-Santos et al., 2018). To date, no approved therapy has provided marked neuroprotective effects nor antiinflammatory therapies, used in the treatment of the disease, showed efficacy in the progressive phase of MS.

Well-defined evidence showed that the MS pathophysiology is characterized by altered bidirectional interactions among several immune cell types in the periphery and resident cells of the CNS, such as microglia and astrocytes (Li et al., 2018). The MS relapses, occurring in the early phases of the disease, are characterized by the infiltration, into the CNS parenchyma, of pro inflammatory CNS-specific effector T, B and myeloid cells, which are activated and/or regulated in an aberrant way (Dendrou et al., 2015). The altered function of regulatory T (Treg) cells and resistance of CNS-specific effector T cells to Treg cell-mediated regulation could be one possible cause of the neuro-inflammation (Kaskow et al., 2018; Kitz et al., 2018). Furthermore, CNS-resident cells, that secrete many inflammatory mediators, recruit inflammatory cells into the CNS (Dendrou et al., 2015). Therefore, both peripheral and CNS-compartmentalized inflammatory mechanisms contribute to MS pathogenesis (Filippi et al., 2018). In the advanced stages of the disease, infiltrated immune cells into the CNS are few and ongoing CNS-compartmentalized inflammation seems to dominate progressive phases of MS. During this phase, the role of B cells in driving inflammation seems to be prominent, particularly within meningeal inflammation (Magliozzi et al., 2007; Correale et al., 2017).

¹Institute of Neurology, Fondazione Policlinico Universitario 'A. Gemelli' IRCCS, Università Cattolica del Sacro Cuore, Rome, Italy; ²IRCCS Fondazione Don Carlo Gnocchi, Florence, Italy

*Correspondence to: Viviana Nociti, MD, PhD, viviana.nociti@policlinicogemelli.it.

https://orcid.org/0000-0003-3840-6164 (Viviana Nociti)

How to cite this article: Nociti V, Santoro M (2021) What do we know about the role of IncRNAs in multiple sclerosis? Neural Regen Res 16(9):1715-1722.

Until now, the exact mechanisms leading to autoimmune diseases are still only partially understood but we know that genetic, epigenetic, molecular, and cellular factors resulting in pathogenic inflammatory responses driven by self antigenspecific T-cells are certainly involved.

Whole-genome transcriptional analysis have recently shown that the genome, in eukaryotic cells, can be transcribed in numerous types of coding and non-coding RNAs, the letter constituting at least 90% of these RNAs (Djebali et al., 2012). Increasing evidence on non-coding RNAs (ncRNAs) demonstrated that, more than evolutionary "junk genes", they have an important role as regulators of different cellular processes also in many diseases (Sun et al., 2013). ncRNAs are grouped into small ncRNAs (< 200 nucleotides) and long ncRNAs (\geq 200 nucleotides).

Long-ncRNAs (Inc-RNA) have more extensive and complex regulatory mechanisms than small ncRNA (Schmitz et al., 2016). Among their several functions (cell proliferation and differentiation, immune responses, metabolism, and apoptosis), they are very important in human autoimmune diseases playing specialized roles in modulating immune cell differentiation and activation (Sigdel et al., 2015; Wu et al., 2015; Atianand et al., 2017).

Indeed, long non-coding RNAs (lncRNAs) have an important impact on both innate and acquired immunity (Sigdel et al., 2015; Atianand et al., 2017; Zhang et al., 2017).

Innate immune responses are the body's non-specific first line defence against pathogenic microorganisms, by the action of macrophages, dendritic cells, and natural killer cells. LncRNAs may have a critical role in regulating this response to pathogens (Mao et al., 2015; Atianand et al., 2017; Ivanov et al., 2018). Many changes have been found in IncRNA expression in macrophages upon innate immunity activation (Hu et al., 2016; Tong et al., 2016; Yang et al., 2016; Fei et al., 2017; Ye et al., 2018) and in dendritic cells are antigenpresenting cells (Ahmad et al., 2020). LncRNAs also have an important role in regulating the acquired immune responses, a second line of defence against pathogens, producing antigenspecific responses and immunological "memory." Lymphocytes T and B are the main immune cells of the adaptive immune system. T helper cells are also critical in the pathogenesis of several diseases, and overall in autoimmune diseases (Zhu et al., 2010). Important IncRNA expression patterns have been elucidated in T cell function (Atianand MK et al., 2017) and in distinct stages of B cell development (Petri et al., 2015; Brazao et al., 2016; Tayari et al., 2016). LncRNAs also have a key role in cytokine genes regulation but overall the antisense IncRNAs (Atianand et al., 2017).

Currently, many MS fields of research are based on the identification of possible biomarkers able to predict the onset of the disease, its activity degree, its progression phase and the response to disease-modifying drugs. Among the various biomarkers analyzed in MS, lncRNAs are of great importance because they are able to regulate the genome expression/ stability and cellular functions such as proliferation, differentiation, apoptosis and development especially in the activation of immune cells both in innate and in adaptive immune system (Esteller, 2011; Wapinski and Chang, 2011). Indeed, it has been demonstrated that unregulation of lncRNAs may have a crucial role in the pathogenesis of autoimmune disorders such as systemic lupus erythematosus, rheumatoid arthritis and psoriasis (Sigdel et al., 2015).

In this review, we present the most recent studies on molecular characterization of lncRNAs in the MS disorder discussing their clinical relevance as biomarkers for diagnosis and treatments.

Search Strategy and Selection Criteria

We have performed a PubMed literature search of articles with search terms including "multiple sclerosis", "pathogenesis", "immune system", "autoimmune diseases", "epigenetic", "non-coding RNA", "long non-coding RNA", "single nucleotide polymorphisms".

Selection criteria included recent articles (2011–2020) on MS pathogenesis, role of lncRNA in autoimmune diseases and recent findings on lncRNAs in MS.

Biogenesis and Function of Long Non-coding RNAs

LncRNAs are a group of non-coding RNA (ncRNAs) transcribed by RNA polymerase II and RNA polymerase III at several loci of the genome (Beermann et al., 2016).

Generally IncRNAs are more than 200 nucleotides in length and exhibit mRNA-like features such as splicing, polyadenylation (poly- A^+) tail and 5' capping (5'cap) but without protein coding capacity (Derrien et al., 2012).

However, recent studies identified same lncRNA with an open reading frame (ORF) and few exons suggesting with a high probability translational ability to encode proteins (Bazin et al., 2017; Wang et al., 2017).

LncRNAs showed a tissue and cell specific expression, so that their levels are influenced by developmental or physiologic and pathologic state (Darren et al., 2012; Ulitsky and Bartel, 2013).

Comparing with mRNA, IncRNAs presented a lower expression in all tissues so that they were initially considered transcriptional *noise* resulting from low RNA polymerase fidelity (Darren et al., 2012). With the advent of highthroughput sequencing such as RNA-sequencing (RNA-seq), several studies showed that IncRNAs are abundant in the human genome and they can regulate the gene expression by DNA, RNA and protein interaction affecting transcriptional, post-transcriptional and translational processes (Atianand and Fitzgerald, 2014).

LncRNAs biogenesis occurs in both the nucleus or the cytoplasm and their cellular localization can influence the regulation mechanism of lncRNAs (**Figure 1**).

In the nucleus the lncRNAs are mainly involved in transcriptional regulation, through epigenetic modulation including histone and chromatin structure alteration that limits DNA accessibility (Tang et al., 2017; **Figure 1**). Instead, in the cytoplasm the lncRNAs are mainly involved in post-transcriptional regulation and post-translational regulation through mechanisms of translation inhibition, alternative splicing regulation and degradation (Tang et al., 2017; **Figure 1**).

LncRNAs classification is based according to their genomic position including proximity to protein-coding genes although several lncRNAs do not fall into this classification but rather turn out to be a combination of these specific characteristics or they cover long genomic sequences (Kung et al., 2013).

The main IncRNA classes are:

1) Intergenic IncRNAs (lincRNAs) represent the largest and most representative group of IncRNAs located among protein coding genes (Xue et al., 2017). By RNA-seq several lincRNAs have been identified in the genome showing a low level of expression (Cabili et al., 2011). They present few exons and can regulate gene transcription in a cell-specific manner by acting either in *cis* or in *trans* regulation (Cabili et al., 2011) (**Figure 1**).

2) Intronic IncRNA (ilncRNA) are entirely derived from the introns of transcript gene in either sense or anti-sense

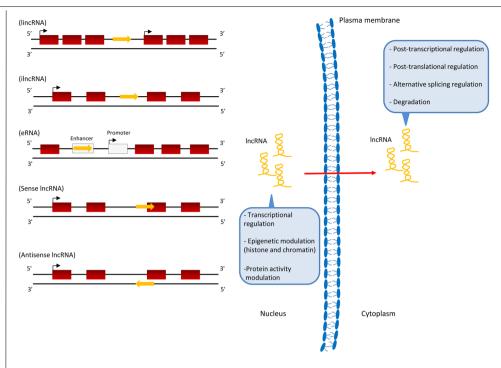


Figure 1 | The biogenesis of IncRNAs involves both the nucleus and the cytoplasm with specific transcriptional and post-transcriptional functions.

The IncRNA classification is based on their transcriptional origin: intergenic IncRNA (lincRNA) transcribed from the genomic region between two genes; intronic IncRNA (ilncRNA) transcribed from the introns: enhancer IncRNAs (eRNAs) transcribed from genomic regions that contain regulative seauences of the gene; sense IncRNAs transcribed from the sense strand of protein-coding gene overlapping partially or completely with intron and exons; anti-sense IncRNAs or natural antisense transcripts (NATs) transcribed from the exons of the protein coding gene on the opposite strand with partial to complete overlapping. IncRNAs: Long non-coding RNAs.

direction. These IncRNAs have been associated with the miRNAs, small nucleolar RNAs (snoRNAs) and circular non-coding RNAs (circRNAs) (Zhang et al., 2013).

The ilncRNA functions and their regulation mechanisms are unclear and relatively unexplored. However, these lncRNAs are transcribed together with their host coding gene, thus they could probably regulate the expression of the host gene (Boivin et al., 2018; **Figure 1**).

3) Enhancer IncRNAs (eRNAs) includes promoter-associated IncRNAs, untranslated region-associated IncRNAs and telomere-associated IncRNAs. Indeed, they are transcripted bi-directionally in both polyadenylated or non-polyadenylated forms from active enhancer genomic regions (Xu et al., 2019). These regions are located regulative sequences of the gene contributing to the transcription start by the binding of transcriptional factors.

In this way, eRNAs can modulate the promoter activity, enhancer interactions and chromatin conformation influencing the transcription of neighboring genes (Chen et al., 2017; Liu et al., 2017; **Figure 1**).

4) Sense IncRNAs are transcribed from the sense strand of protein-coding gene and their sequence overlap, partially or completely, with the entire sequence of a protein coding gene including intron and part of exons (Devaux et al., 2015; **Figure 1**).

5) Anti-sense lncRNAs or natural antisense transcripts are encoded from exons of protein-coding genes in the opposite strand with partial to complete overlapping (Devaux et al., 2015). Anti-sense lncRNAs can modulate neighbouring genes expression through a regulation in *cis* (Magistri et al., 2012). About 70% of coding genes have anti-sense counterparts (Villegas et al., 2015; **Figure 1**).

The IncRNAs have the ability to inhibit or promote the expression of coding genes in tissue or cell-type manner and stage-specific development, suggesting their involvement in the pathogenesis of several diseases such as autoimmune and neurological disorders (Ingwersen et al., 2015; Sigdel et al., 2015).

In the context of autoimmune diseases, the research on MS has been enriched with many studies on the molecular role of lncRNAs in pathogenesis of this disorder and their

potential application in MS such as diagnostic and prognostic biomarkers (Yang et al., 2018; Li et al., 2020).

Expression Profile of Long Non-coding RNAs in Multiple Sclerosis

In the last years, many studies have provided evidence of how IncRNAs deregulation is involved in the pathogenesis of MS disease as shown in Table 1. Here, we show the most recent studies on IncRNAs expression in the MS disease considering the biological material [serum, plasma, peripheral blood mononuclear cells (PBMC) and blood] on which the analysis was conducted. Serum represents one of the most accessible biological samples and provides excellent materials for the study of possible disease biomarkers. Recently, Santoro et al., screened 84 IncRNAs, involved in autoimmunity and human inflammatory response, in serum from 8 healthy controls, 16 secondary progressive MS (SPMS), 12 primary progressive MS (PPMS) Italian patients (Santoro et al., 2020). The authors found the up-regulation of taurine up-regulated 1 (TUG1) in SPMS patients, while PPMS patients showed a downregulation of non-protein coding RNA 188 (LRRC75A-AS1) and a significant up-regulation of long intergenic non-protein coding RNA 293 (LINCO0293) and RP11-29G8.3 (Santoro et al., 2020). In-silico analysis with bio-informatics tools to predict the interaction of IncRNAs-miRNAs such as DIANA-LncBase v2 and HMDD v3.0 software (Paraskevopoulou et la., 2016; Huang et la., 2019), identified 21 miRNAs prediction targets of these four IncRNAs that are involved in MS disease (Santoro et al., 2020).

In previous papers, the same authors showed the specific over-expression of three circulating lncRNAs in the serum of 12 relapsing-remitting MS (RRMS) patients: nuclear paraspeckle assembly transcript 1 (*NEAT1*), 7SK small nuclear RNA (*RN7SK RNA*) and *TUG1* (Santoro et al., 2016). These three lncRNAs are involved in specific regulatory functions: *NEAT1* promotes the increase of *CXCL8* expression of the gene encoding interleukin 8 via relocation of SFPQ splicing factor (Imamura et al., 2014), *RN7SK RNA* is involved in regulation of CD4⁺ T lymphocytes (Sung et al., 2006) and *TUG1* is a component of the p53 regulatory network (Rossi et al., 2014).

Although both represent pilot studies given the limited number of samples and need confirmation in larger collection

LncRNAs	Regulation	Patients	Sample	Function	References
TUG1	\uparrow	SPMS	Serum	Involvment in p53 pathway and cell cycle	Santoro et al. (2020)
LRRC75A-AS1	\downarrow	PPMS	Serum	Not determined	Santoro et al. (2020)
LINC00293	\uparrow	PPMS	Serum	Not determined	Santoro et al. (2020)
RP11-29G8.3	\uparrow	PPMS	Serum	Not determined	Santoro et al. (2020)
LincR-Gng2-5' AS	\uparrow	RRMS/SPMS	Serum	Immune regulatory function	Shaker et al. (2020)
LincR-Epas1-3'AS	\checkmark	RRMS/SPMS	Serum	Immune regulatory function	Shaker et al. (2020)
MALAT1	\uparrow	SPMS	Serum	Oncogenic role	Shaker et al. (2019)
InDC	\uparrow	RRMS	Serum	Differentiation and maturation of dendritic cells	Shaker et al. (2019)
NEAT1	\uparrow	RRMS	Serum/Blood	Regulation of CXCL8 expression	Santoro et al. (2016); Dastmalchi et al. (2018)
TUG1	\uparrow	RRMS	Serum/Blood	Involvment in p53 pathway and cell cycle	Santoro et al. (2016); Dastmalchi et al. (2018)
RN7SK RNA	\uparrow	RRMS	Serum	Regulation of CD4 ⁺ T lymphocytes	Santoro et al. (2016)
APOA1-AS	\uparrow	RRMS	Plasma	Negative transcriptional regulator of ApoA1	Ghaiad et al. (2019)
IFNG-AS1	\uparrow	RRMS	Plasma	Transcription/expression of IFN-γ in Th1 cells	Ghaiad et al. (2019)
MEG3a	\downarrow	RRMS	Blood	Autoimmune diseses	Moradi et al. (2020a)
PVT1	\downarrow	RRMS	Blood	Control pf IL-6 release	Eftekharian et al. (2017)
FAS-AS1	\downarrow	RRMS	Blood	Regulation of soluble Fas receptor	Eftekharian et al. (2017)
THRIL	\uparrow	RRMS	Blood	Regulative role in innate immunity	Eftekharian et al. (2017)
IFNG-AS1	\downarrow	RRMS	Blood	Transcription/expression of IFN-γ in Th1 cells	Ganji et al. (2019)
PANDA	\uparrow	RRMS	Blood	p53 protein stabilization	Dastmalchi et al. (2018)
GAS8-AS1	\uparrow	RRMS	Blood	Not determined	Patoughi et al. (2019)
OIP5-AS1	\uparrow	RRMS	Blood	Cell division	Gharesouran et al. (2019)
RP11-530C5.1	\uparrow	RRMS	PBMC	Potential cis-regulatory	Ghoveud et al. (2020)
AL928742.12	\checkmark	RRMS	PBMC	Not determined	Ghoveud et al. (2020)
IFNG-AS1-001	\uparrow	RRMS	PBMC	Transcription/expression of IFN-γ in Th1 cells	Hosseini et al. (2019)
IFNG-AS1-003	\uparrow	RRMS	PBMC	Transcription/expression of IFN-γ in Th1 cells	Hosseini et al. (2019)
AC007278.2	\uparrow	RRMS	PBMC	Regulation of Th1 cell development	Hosseini et al. (2019)
NRON	\checkmark	RRMS/PPMS	PBMC	Nuclear factor repressor of activated T cells	Fenoglio et al. (2018)
TUG1	\checkmark	RRMS/PPMS	PBMC	Involvement in p53 pathway and cell cycle	Fenoglio et al. (2018)
Inc-DDIT4	\uparrow	RRMS	PBMC/CD4 ⁺ T cells	Th17 cell differentiation	Zhang et al. (2018)
Linc-MAF-4	\uparrow	RRMS	PBMC	Th1/Th2 cell differentation	Zhang et al. (2017)
HOTAIR	\uparrow	RRMS	PBMC	Vitamin D and inflammation regulation	Pahlevan Kakhki et al. (202

IFN-y: Interferon gamma; IL-6: interleukin-6; IncRNAs: long non-coding RNAs; PBMCs: peripheral blood mononuclear cells; PPMS: primary progressive multiple sclerosis; RRMS: relapsing-remitting multiple sclerosis; SPMS: secondary progressive multiple sclerosis.

of RRMS, SPMS and PPMS samples, data obtained suggest the potential role of these IncRNAs in progressive MS pathogenesis. Another recent study conducted on serum of 60 healthy controls, 42 RRMS and 18 SPMS Egyptian patients showed an up-regulation of *LincR-Gng2-5' AS* and a downregulation of *LincR-Epas1-3'AS*, two anti-sense intergenic IncRNAs located in T helper 1 (TH1) and in T helper 2 (TH2) cells (Shaker et al., 2020). Furthermore, the unregulation of these two IncRNAs was more marked in SPMS than in RRMS patients with an opposite correlation regarding the Expanded Disability Status Scale (positive for *LincR-Gng2-5'AS* and negative for *LincR-Epas1-3'AS*) (Shaker et al., 2020).

Moreover, in a previous study, Shaker et al. (2019) analyzed the expression levels of metastasis-associated lung adenocarcinoma transcript 1 (*MALAT1*) and InDC IncRNAs in serum from 45 MS patients and 45 controls finding an up-regulation of *MALAT1* in the SPMS subgroup with no significant differences in RRMS patients. Indeed, *Inc-DC* showed an opposed expression: up-regulated in RRMS patients and no changes in SPMS subgroup (Shaker et al., 2019). In both studies, the expression analysis was performed only on some specific IncRNAs making it impossible to know if other IncRNAs are also involved.

Regarding the IncRNAs expression analysis in plasma, we found only one paper by Ghaiad et al., (2019) that determined the expression levels of anti-sense (AS) IncRNAs *APOA1-AS* and *IFNG-AS1* in the plasma of 72 RRMS Egyptian patients (37 during relapse and 35 in remission) and 28 healthy

controls. The authors found a significant up-regulation of *APOA1-AS* during relapse and in remission comparing with healthy controls while ApoA1 and high-density lipoproteins-cholesterol levels were significantly lower in the same phase, together with higher low-density lipoprotein-cholesterol levels (Ghaiad et al., 2019).

Moreover, the expression levels of *IFNG-AS1* were also significantly higher in RRMS patients during relapse and in remission comparing with healthy controls (Ghaiad et al., 2019).

This study provides evidence for diagnostic and prognostic role of *APOA1-AS* and *IFNG-AS1* in RRMS patients although further analysis is needed for validation in a larger cohort of MS patients.

Whole blood is also often used to evaluate IncRNA expression levels. Moradi et al. (2020a) analyzed in the blood of 10 controls and 20 Iranian RRMS patients, the expression profile of *MEG3a*, *AC000061.1_201*, and *AC007182.6*, three IncRNAs involvement in the pathogenesis of human autoimmune diseases. The authors found a down-regulation only of *MEG3a* that negatively correlate with Expanded Disability Status Scale. Additionally, analysis of receiver operating characteristic curve showed the ability of this IncRNA to discriminate between RRMS and healthy individuals suggesting *MEG3a* (Moradi et al., 2020a). Even if *MEG3a* is indicated as a potential diagnostic biomarker to distinguish MS patients, this study was conducted on a small cohort of samples (10 controls vs. 20 RRMS patients) and further experiments are needed to validate these results in a large number of individuals considering also the different stages of the MS disease.

Another blood study conducted on Iranian RRMS (n = 50) patients versus 50 healthy controls showed a downregulation of Fas cell surface death receptor- antisense 1 (FAS-AS1) and plasmacytoma variant translocation 1 (PVT1) with an up-regulation of $TNF-\alpha$ and heterogeneous nuclear ribonucleoprotein L (THRIL) and (Eftekharian et al., 2017). Even if these three lncRNAs were involved in innate immunity apoptosis regulation during lymphocyte development and immune responses (Li et al., 2014; Aune and Spurlock, 2016; Austin et al., 2017), the authors do not find a significant correlation between the expression levels and clinical feature of RRMS thus the role of these lncRNAs in MS pathogenesis remains to be clarified.

Always in the blood of Iranian RRMS (n = 50) patients, Ganji et al. (2019) found a down-regulation of *GSTT1-AS1* and *IFNG-AS1* IncRNAs with an up-regulation of their coding targets *TNF* and *IFNG*. Indeed, *GSTT1-AS1* (or *IncRNA-CD244*) is involved in the inhibition regulation of *IFNG* and tumor necrosis factor (*TNF*) genes (Wang et al., 2015), while *IFNG-AS1* together with T-bet is able to regulate the transcription of *IFNG* (Aune et al., 2016). These data disagree with the results obtained by Ghaiad that found an up-regulation of *IFNG-AS1* in RRMS patients.

This discordance is probably related to the different biological source (plasma *versus* blood) chosen for the analysis, ethnic group (Egyptian *versus* Iranian) or the phase of disease and drug treatment at the time of MS patient recruitment.

Furthermore, some studies conducted on whole blood have shown that the expression levels of some InRNAs can be modulated by age and gender.

Indeed, Dastmalchi et al. (2018) detected an over-expression of *NEAT1*, *TUG1* and P21 associated ncRNA DNA damage activated (*PANDA*) in blood samples of 50 RRMS patients. In particular, the expression of *NEAT1* and *TUG1* was inversely correlated with age at onset and disease duration in female patients while *PANDA* showed a more prominent overexpression in male patients (Dastmalchi et al., 2018). These data indicated a gender-dependent role of *NEAT1 TUG1* and *PANDA* in the pathogenesis of MS suggesting that the presence of a sex-determined factor, hormones and drugs, can regulate the expression of these lncRNAs (Dastmalchi et al., 2018).

Patoughi et al. (2019) found in the blood of RRMS (n = 50) versus healthy controls (n = 50), an up-regulation of the antisense RNA 1 growth arrest specific 8 (*GAS8-AS1*) with higher levels of expression in male patients.

Moreover, Gharesouran et al. (2019) detected high levels of OIP5-AS1 expression in the blood of RRMS (n = 50) patients, and these data are particularly evident in men less than 30 years old. With the increasing age of patients, the expression of this lncRNA seems to influence the incidence and onset of the disease suggesting a regulative role in the cell division process (Gharesouran et al., 2019).

Regarding the analysis of lncRNAs on cells, many studies have been carried out on PBMC isolated from blood.

Indeed, comparing 50 RRMS patients versus 25 healthy controls, Ghoveud et al. (2020) found in PBMC an upregulation of *RP11-530C5.1* and a down-regulation of *AL928742.12* IncRNAs. Although the results obtained with the receiver operating characteristic analysis suggested potential biomarker roles of both these IncRNAs, the study needs validation in a larger sample of both healthy controls and MS patients at different stages of the disease. Hosseini et al., found in the PBMC of 50 RRMS (25 in relapsing phase and 25 in the remitting phase treated with interferon beta) high expression of *AC007278.2* and *IFNG-AS1-001* IncRNAs in relapsing phase MS patients compared with the healthy controls while *IFNG-AS1-003* IncRNA was elevated in MS patients in the remitting phase comparing with those in relapsing phase (Hosseini et al., 2019). In association with MS disease, *AC007278.2* turns out to be correlated with the expression of *IL18R1* and *IL18RAP* genes that encode the α and β chains of the heterodimeric IL-18 receptor involved in Th1 cell development (Hosseini et al., 2019). Indeed, *IFNG-AS1-001* is correlated with *IFNG* expression suggesting this lncRNA as a potential target for MS treatment (Hosseini et al., 2019).

Another study on PBMC from cohort of Italian population (27 RRMS, 13 PPMS and 31 healthy controls) showed a significant down-regulation of *MALAT1*, *MEG9*, *NRON*, *ANRIL*, *TUG1*, *XIST*, *SOX2OT*, *GOMAFU*, *HULC*, *BACE-1AS* IncRNAs (Fenoglio et al., 2018).

The validation analysis conducted on an independent Belgian cohort composed by 17 RRMS, 7 PPMS and 23 healthy controls showed a down-regulation of only *NRON* and *TUG1* (Fenoglio et al., 2018). The data concerning the low levels of *TUG1* expression were opposite to those of Dastmalchi et al. (2018) and Santoro et al. (2016) which instead showed an up-regulation. Probably, this discrepancy could be related to the biological sample used for the analysis (blood and serum *versus* PBMC), different phase of the disease and pharmacological treatment.

Besides analyzing the levels of expression, some lncRNAs have been characterized by their involvement in regulatory functions.

Indeed, transfection experiments for over-expression of *Inc-DDIT4*, that found up-regulated in both PBMCs and CD4⁺ T cells of RRMS patients, in naive CD4⁺ T cells showed an inhibition of Th17 cell differentiation through the regulation of DNA-damage-inducible transcript 4 (DDIT4) expression resulting in the modulation of DDIT4/mTOR pathway (Zhang et al. 2018).

These data suggest a possible role of *lnc-DDIT4* in the pathogenesis of MS through the regulation Th17 cell differentiation. Moreover, the over-expression of *Linc-MAF-4*, other lncRNA found up-regulated in PBMC from RRMS patients, into naive $CD4^+T$ cells induced Th1-cell differentiation inhibiting Th2-cell differentiation by the action of transcription factor MAF (Zhang et al., 2017).

On the other hand, a down-regulation of *linc-MAF-4* has an opposite effect with inhibition of Th1 cells differentiation and the development of Th2 cells (Zhang et al., 2017). These data suggest an important role of *linc-MAF-4* in the pathogenesis of MS disease.

Finally, Pahlevan Kakhki et al. (2017) analyzed in the PBMC from 52 RRMS the expression levels of HOX transcript antisense intergenic RNA (*HOTAIR*) and anti-sense lncRNA of the INK4 locus (*ANRIL*), both lncRNAs are involved in the inflammatory disorders. The authors correlated the expression levels of these two lncRNAs with the serum levels of vitamin D considered that can regulate the expression of lncRNAs (Riege et al., 2017).

Their data showed that MS patients at base line (before vitamin D treatment) had higher expression levels of *HOTAIR* but not *ANRIL* IncRNAs compared with the healthy controls.

After vitamin D treatment, *HOTAIR* levels returned to those found in the healthy controls (Pahlevan Kakhki et al., 2017). These data suggest that vitamin D could modulate inflammation through the regulation of *HOTAIR*, however further studies are needed to clarify this issue.

In conclusion, a large number of studies have been carried out on the molecular analysis of lncRNAs and their role in the pathogenesis of MS. Unfortunately, the data produced are still limited and inconclusive. In fact, there are some important critical issues to consider: i) the number of samples with both MS patients and controls used for the lncRNAs analysis is limited and needs validation in a larger study sample; ii) the expression levels of lncRNAs is influenced by several factors such as the biological sample chosen for the analysis (serum, plasma, blood and PBMC), pharmacological treatment, stage of MS disease, age and sex; iii) an appropriate normalization method based on the stable expression of reference gene across different sample groups and tissue in order to avoid introducing a significant error in the quantification of lncRNA levels.

Single Nucleotide Polymorphisms in Multiple Sclerosis-Related Long Non-coding RNAs

LncRNAs perform their regulatory function through the secondary and tertiary structures they can adopt. The presence of single nucleotide polymorphisms (SNPs) inside the lncRNAs can alter these structures influencing their expression levels and functions (Hangauer et al., 2013). Indeed, several bio-informatic tools have been developed to predict lncRNA structures based on the presence of a specific SNP (Gong et al., 2015; Ren et al., 2018).

Moreover, some SNPs are involved in the modulation of IncRNA alternative splicing by regulating exon skipping and production of specific isoforms that could present a different binding affinity for their target modifying the downstream events (Aguilo et al., 2016; Chowdhury et al., 2017).

Based on these premises, several studies analyzed the association of some SNPs present within the IncRNAs sequence with the risk of MS.

Bahrami et al. (2020) analyzed the SNPs association with RRMS susceptibility on blood samples from Iranian population (300 RRMS patients *vs.* 300 healthy controls) of two IncRNAs involved in oxidative stress and autophagy: *TRPM2-AS* (rs933151) and *HNF1A-AS1* (rs7953249). The authors found that SNP rs933151 was statistically associated with RRMS risk and T allele of this SNP was statistically poorly represented in RRMS patients compared with healthy controls. Instead, the rs7953249 was not associated with RRMS susceptibility (Bahrami et al., 2020).

The expression analysis on both IncRNAs has not been conducted in MS patients; therefore, it is not possible to understand whether this SNP may have a role in the modulation of *TRPM2-AS* and *HNF1A-AS1* levels.

Other IncRNAs studied for a possible association between SNP and MS risk were *HOTAIR* and *ANRIL*. Previous data showed that only *HOTAIR* had higher expression levels in the MS patients, which can be modulated through vitamin D treatment (Pahlevan Kakhki et al., 2017)

Their data showed that MS patients at base line (before vitamin D treatment) but not *ANRIL* lncRNAs compared with the healthy controls.

Three SNPs (rs12826786, rs1899663 and rs4759314) of *HOTAIR* were genotyped by Taheri and colleagues in blood from 403 Iranian RRMS patients *versus* 420 healthy controls (Taheri et al., 2020). The authors found a significant association with MS risk only for rs4759314 SNP (Taheri et al., 2020). The high expression levels of *HOTAIR* found in MS patients suggest an important role of this SNP in the regulation of the *HOTAIR* expression.

On the other hand, Rezazadeh et al. (2018) also evaluated the association of rs1333045, rs4977574, rs1333048, and

rs10757278 SNPs of *ANRIL* and MS risk in blood from 410 Iranian RRMS patients and 410 healthy controls. The authors found protective effect of CCGG and TAAA haplotypes against MS (rs1333045, rs1333048, rs4977574, and rs10757278 respectively), while TAGG and CCGA haplotypes were significantly associated with MS risk (Rezazadeh et al., 2018) even if the expression levels of *ANRIL* IncRNA are not altered in RRMS patients (Pahlevan Kakhki et al., 2017).

Moreover, *MALAT1* was another lncRNA with an up-regulation in the SPMS and no significant differences in RRMS patients (Shaker et al., 2019). Indeed, also the analysis of two *MALAT1* (rs619586 and rs3200401) SNPs conducted by Eftekharian et al. (2019) on blood from 428 Iranian RRMS patients *versus* 505 healthy controls showed a low association with MS risk only for rs619586 polymorphism.

Finally, a significant association with RRMS was found for the rs55829688 and rs2067079 polymorphisms in growth arrest-specific 5 (*GAS5*) (Eftekharian et al., 2019; Moradi et al., 2020b).

In agreement with previous data, the up-regulation of *GAS5* was found in whole blood of MS patients suggesting an important regulatory role of IncRNA *GAS5* genetic variant in the pathogenesis of MS (Mayama et al., 2016).

All these data suggest a possible role of the SNPs in the regulation of lncRNAs even if the possible molecular mechanism that underlies the impact of SNPs on lncRNA structure and function remains unknown.

In general, the analysis of SNPs should also be extended to other ethnic groups as most of the published studies have been conducted on an Iranian population.

Furthermore, each SNP should be associated with the analysis of IncRNA expression, splicing and secondary structure in the same population and in the same patient.

It is necessary to adopt an approach that analyzes the functional differences of the lncRNA alleles and their ability to regulate expression of downstream genes.

Concluding Remarks

Until now, the exact mechanisms leading to diseases with an autoimmune pathogenesis, such MS, are still not entirely understood, but we know that genetic, epigenetic, molecular, and cellular factors resulting in pathogenic inflammatory responses are certainly involved. LncRNAs have, among their several functions (cell proliferation and differentiation, metabolism, and apoptosis), an important impact on both innate and acquired immunity, so there is great interest on IncRNAs involved in autoimmune diseases. About MS, the research has been enriched with many studies on the molecular role of IncRNAs in pathogenesis of the disease and their potential application such as diagnostic and prognostic biomarkers. In particular, many MS fields of research are based on the identification of IncRNAs as possible biomarkers able to predict the onset of the disease, its activity degree, its progression phase and the response to disease modifying drugs. Last but not least, studies on IncRNAs in MS can provide new molecular target for novel therapies, missing, so far, a cure for the disease.

Expert opinion: while our knowledge on the role of IncRNA in MS has recently improved, we know that this knowledge is at the dawn and further studies are required to better understand the specific role of IncRNAs in MS and in other autoimmune diseases. Among the various biomarkers analyzed in MS, IncRNAs are of great importance because they are able to regulate the genome expression/stability and cellular functions especially in the activation of immune cells both in innate and in adaptive immune system. So far, there are still several deep-rooted problems regarding the IncRNAs function in autoimmune diseases. Why are IncRNAs abnormally expressed in autoimmune diseases? What are the specific mechanisms underlying this abnormal expression? Have changes in IncRNAs got a causal role in disease activity and/or in disease progression? Assumed that IncRNAs play important roles as regulators of several biological processes, is it possible that this indirect unregulation is linked to autoimmune disease pathogenesis? Further studies are needed to answer the aforementioned and many other questions on this new and relevant topic for MS and other autoimmune diseases.

Author contributions: Study design, manuscript drafting and revision: VN and MS; collection of clinical and diagnostic data: VN; collection of molecular genetic data: MS. Both authors approved the final version of the paper.

Conflicts of interest: *The authors declare no conflicts of interest.* **Financial support:** *None.*

Copyright license agreement: The Copyright License Agreement has been signed by both authors before publication.

Plagiarism check: Checked twice by iThenticate.

Peer review: Externally peer reviewed.

Open access statement: This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

Open peer reviewer: Taskın Duman, Mustafa Kemal University, Turkey.

References

- Aguilo F, Di Cecilia S, Walsh MJ (2016) Long non-coding RNA ANRIL and polycomb in human cancers and cardiovascular disease. Curr Top Microbiol Immunol 394:29-39.
- Ahmad I, Valverde A, Ahmad F, Naqvi AR (2020) Long noncoding RNA in myeloid and lymphoid cell differentiation, polarization and function. Cells 22;9:269.
- Atianand MK, Caffrey DR, Fitzgerald KA (2017) Immunobiology of long noncoding RNAs. Ann Rev Immunol 35:177-198.
- Atianand MK, Fitzgerald KA, (2014) Long non-coding RNAs and control of gene expression in the immune system. Trends Mol Med 20:623-631.
- Aune TM, Crooke PS, Spurlock CF (2016) Long noncoding RNAs in T lymphocytes. J Leukoc Biol 99:31-44.

Aune TM, Spurlock CF (2016) Long non-coding RNAs in innate and adaptive immunity. Virus Res 212:146-160.

- Austin PJ, Tsitsiou E, Boardman C, Jones SW, Lindsay MA, Adcock IM, Chung KF, Perry MM (2017) Transcriptional profiling identifies the long noncoding RNA plasmacytoma variant translocation (PVT1) as a novel regulator of the asthmatic phenotype in human airway smooth muscle. J Allergy Clin Immunol 139:780-789.
- Bahrami T, Taheri M, Omrani MD, Karimipoor M (2020) Associations between genomic variants in IncRNA-TRPM2-AS and IncRNA-HNF1A-AS1 genes and risk of multiple sclerosis. J Mol Neurosci 70:1050-1055.
- Bazin J, Baerenfaller K, Gosai SJ, Gregory BD, Crespi M, Bailey-Serres J (2017) Global analysis of ribosome-associated noncoding RNAs unveils new modes of translational regulation. Proc Natl Acad Sci USA 114: E10018-10027.
- Beermann J, Piccoli MT, Viereck J, Thum T (2016) Non-coding RNAs in development and disease: background, mechanisms, and therapeutic approaches. Physiol Rev 96:1297-1325.
- Boivin V, Deschamps-Francoeur G, Scott MS (2018) Protein coding genes as hosts for noncoding RNA expression. Semin Cell Dev Biol 75:3-12.
- Brazao TF, Johnson JS, Muller J, Heger A, Ponting CP, Tybulewicz VL (2016) Long non-coding RNAs in B cell development and activation. Blood 128:e10-e-19.
- Cabili MN, Trapnell C, Go L, Koziol M, Tazon-Vega B, Regev A, Rinn JL (2011) Integrative annotation of human large intergenic noncoding RNAs reveals global properties and specific subclasses. Genes Dev 25:1915-1927.
- Chen H, Du G, Song X, Li L (2017) Non-coding transcripts from enhancers: new insights into enhancer activity and gene expression regulation. Genom Proteom Bioinform 15:201-207.
- Chowdhury IH, Narra HP, Sahni A, Khanipov K, Schroeder CLC, Patel J, Fofanov Y, Sahni SK (2017) Expression profiling of long noncoding RNA splice variants in human microvascular endothelial cells: lipopolysaccharide effects in vitro. Mediators Inflamm 2017:1-18.
- Correale J, Gaitán MI, Ysrraelit MC, Fiol MP (2017) Progressive multiple sclerosis: from pathogenic mechanisms to treatment. Brain 140:527-546.

- Dastmalchi R, Ghafouri-Fard S, Davood Omrani M, Mazdeh M, Sayad A, Taheri M (2018) Dysregulation of long non-coding RNA profile in peripheral blood of multiple sclerosis patients. Mult Scler Relat Disord 25:219-226.
- Dendrou CA, Fugger L, Friese MA (2015) Immunopathology of multiple sclerosis. Nat Rev Immunol 15:545-558.
- Derrien T, Johnson R, Bussotti G, Tanzer A, Djebali S, Tilgner H, Guernec G, Martin D, Merkel A, Knowles DG, Lagarde J, Veeravalli L, Ruan X, Ruan Y, Lassmann T, Carninci P, Brown JB, Lipovich L, Gonzalez JM, Thomas M, et al. (2012) The GENCODE v7 catalog of human long non coding RNAs analysis of their gene structure, evolution, and epressiom. Genome Res 22:1755-1789.
- Devaux Y, Zangrando J, Schroen B, Creemers EE, Pedrazzini T, Chang CP, Dorn GW 2nd, Thum T, Heymans S, Cardiolinc network (2015) Long noncoding RNAs in cardiac development and ageing. Nat Rev Cardiol 12:415-425.
- Djebali S, Davis CA, Merkel A, Dobin A, Lassmann T, Mortazavi A, Tanzer A, Lagarde J, Lin W, Schlesinger F, Xue C, Marinov GK, Khatun J, Williams BA, Zaleski C, Rozowsky J, Röder M, Kokocinski F, Abdelhamid RF, Alioto T, et al. (2012) Landscape of transcription in human cells. Nature 489:101-108.
- Eftekharian MM, Ghafouri-Fard S, Soudyab M, Omrani MD, Rahimi M, Sayad A, Komaki A, Mazdeh M, Taheri M (2017) Expression analysis of long noncoding RNAs in the blood of multiple sclerosis patients. J Mol Neurosci 63:333-341.
- Esteller M (2011) Non-coding RNAs in human disease. Nat Rev Genet 12:861-874.
- Fei T, Chen Y, Xiao T, Li W, Cato L, Zhang P, Cotter MB, Bowden M, Lis RT, Zhao SG, Wu O, Feng FY, Loda M, He H, Liu XS, Brown M (2017) Genomewide CRISPR screen identifies HNRNPL as a prostate cancer dependency regulating RNA splicing. Proc Natl Acad Sci U S A 114:E5207-5215.
- Fenoglio C, Oldoni É, Serpente M, De Riz MA, Arcaro M, D'Anca M, Pietroboni AM, Calvi A, Lecchi E, Goris A, Mallants K, Dubois B, Comi C, Cantello R, Scarpini E, Galimberti D (2018) LncRNAs expression profile in peripheral blood mononuclear cells from multiple sclerosis patients. J Neuroimmunol 324:129-135.
- Filippi M, Bar-Or A, Piehl F, Preziosa P, Solari A, Vukusic S, Rocca MA (2018) Multiple sclerosis. Nat Rev Dis Primers 4:43.
- Ganji M, Sayad A, Omrani MD, Arsang-Jang S, Mazdeh M, Taheri M (2019) Expression analysis of long non-coding RNAs and their target genes in multiple sclerosis patients. Neurol Sci 40:801-811.
- Ghaiad HR, Elmazny AN, Nooh MM, El-Sawalhi MM, Shaheen AA (2019) Long noncoding RNAs APOA1-AS, IFNG-AS1, RMRP and their related biomolecules in Egyptian patients with relapsing-remitting multiple sclerosis: relation to disease activity and patient disability. J Adv Res 21:141-150.
- Gharesouran J, Taheri M, Sayad A, Mazdeh M, Davood Omrani M (2019) Integrative analysis of OIP5-AS1/HUR1 to discover new potential biomarkers and therapeutic targets in multiple sclerosis. J Cell Physiol 234:17351-17360.
- Ghoveud E, Teimuri S, Vatandoost J, Hosseini A, Ghaedi K, Etemadifar M, Nasr Esfahani MH, Megraw TL (2020) Potential biomarker and therapeutic IncRNAs in multiple sclerosis through targeting memory B cells. Neuromolecular Med 22:111-120.
- Gong J, Liu W, Zhang J, Miao X, Guo AY (2015) IncRNA SNP: a database of SNPs in IncRNAs and their potential functions in human and mouse. Nucleic Acids Res 43:D181- D186.
- Hangauer MJ, Vaughn IW, McManus MT (2013) Pervasive transcription of the human genome produces thousands of previously unidentified long intergenic noncoding RNAs. PLoS Genet 9:e1003569.
- Hosseini A, Teimuri S, Ehsani M, Rasa SMM, Etemadifar M, Nasr Esfahani MH, Megraw TL, Ghaedi K (2019) LncRNAs associated with multiple sclerosis expressed in the Th1 cell lineage. J Cell Physiol 234:22153-22162.
- Hu G, Gong AY, Wang Y, Ma S, Chen X, Chen J, Su CJ, Shibata A, Strauss-Soukup JK, Drescher KM, Chen XM (2016) LincRNA-Cox2 promotes late inflammatory gene transcription in macrophages through modulating SWI/ SNF-mediated chromatin remodeling. J Immunol 196:2799-2808.
- Huang Z, Shi J, Gao Y, Cui C, Zhang S, Li J, Zhou Y, Cui Q (2019) HMDD v3.0: a database for experimentally supported human microRNA-disease associations. Nucleic Acids Res 47: D1013-D1017.
- Imamura K, Imamachi N, Akizuki G, Kumakura M, Kawaguchi A, Nagata K, Kato A, Kawaguchi Y, Sato H, Yoneda M, Kai C, Yada T, Suzuki Y, Yamada T, Ozawa T, Kaneki K, Inoue T, Kobayashi M, Kodama T, Wada Y, et al. (2014) Long noncoding RNA NEAT1-dependent SFPQ relocation from promoter region to paraspeckle mediates IL8 expression upon immune stimuli. Mol Cell 53:393-406.
- Ingwersen J, Menge T, Wingerath B, Kaya D, Graf J, Pro zoro vski T, Keller A, Back es C, Beier M, Scheffler M, Dehmel T, Kieseier BC, Hartung HP, Küry P, Aktas O (2015) Natalizumab restores aberrant miRNA expression profile in multiple sclerosis and reveals a critical role for miR-20b. Ann Clin Transl Neurol 2:43-55.
- Ivanov S, Merlin J, Lee MKS, Murphy AJ, Guinamard RR (2018) Biology and function of adipose tissue macrophages, dendritic cells and B cells. Atherosclerosis 271:102-110.

- Kaskow BJ, Baecher-Allan C (2018) Effector T cells in multiple sclerosis. Cold Spring Harb Perspect Med 8:a029025.
- Kitz A, Singer E, Hafler D (2018) Regulatory T cells: from discovery to autoimmunity. Cold Spring Harb Perspect Med 8:a029041.
- Kung JTY, Colognori D, Lee JT (2013) Long noncoding RNAs: Past, present, and future. Genetics 193:651-669.
- Li QW, Lei W, Chen C, Guo W (2020) Recent advances of long noncoding RNAs involved in the development of multiple sclerosis. Chin J Nat Med 18:36-46.
- Li R, Patterson K, Bar-Or A (2018) Reassessing the contributions of B cells in multiple sclerosis. Nat Rev Immunol 19:696-707.
- Li Z, Chao TC, Chang KY, Lin N, Patil VS, Shimizu C, Head SR, Burns JC, Rana TM (2014) The long noncoding RNA THRIL regulates TNFalpha expression through its interaction with hnRNPL. Proc Natl Acad Sci U S A 111:1002-1007.
- Liu F (2017) Enhancer-derived RNA: a primer. Genom Proteom Bioinform 15:196-200.
- Machado-Santos J, Saji E, Tröscher AR, Paunovic M, Liblau R, Gabriely G, Bien CG, Bauer J, Lassmann H (2018) The compartmentalized inflammatory response in the multiple sclerosis brain is composed of tissue-resident CD8+ T lymphocytes and B cells. Brain 141:2066-2208.
- Magistri M, Faghihi MA, Laurent GS, Wahlestedt C (2012) Regulation of chromatin structure by long noncoding RNAs: Focus on natural antisense transcripts. Trends Genet 28:389-396.
- Magliozzi R, Howell O, Vora A, Serafini B, Nicholas R, Puopolo M, Reynolds R, Aloisi F (2007) Meningeal B-cell follicles in secondary progressive multiple sclerosis associate with early onset of disease and severe cortical pathology. Brain 130:1089-1104.
- Mahdi Eftekharian M, Noroozi R, Komaki A, Mazdeh M, Taheri M, Ghafouri-Fard S (2019) GAS5 genomic variants and risk of multiple sclerosis. Neurosci Lett 701:54-57.
- Mao AP, Shen J, Zuo Z (2015) Expression and regulation of long noncoding RNAs in TLR4 signaling in mouse macrophages. BMC Genomics 16:45.
- Mayama T, Marr A, Kino T (2016) Differential expression of glucocorticoid receptor noncoding RNA repressor Gas5 in autoimmune and inflammatory diseases. Horm Metab Res 48:550-557.
- Moradi A, Rahimi Naiini M, Yazdanpanahi N, Tabatabaeian H, Nabatchian F, Baghi M, Azadeh M, Ghaedi K (2020a) Evaluation of the expression levels of three long non-coding RNAs in multiple sclerosis. Cell J 22:165-170.
- Moradi M, Gharesouran J, Ghafouri-Fard S, Noroozi R, Talebian S, Taheri M, Rezazadeh M (2020b) Role of NR3C1 and GAS5 genes polymorphisms in multiple sclerosis. Int J Neurosci 130:407-412.
- Naegele M, Martin R (2014) The good and the bad of neuroinflammation in multiple sclerosis. Handb Clin Neurol 122:59-87.
- Ontaneda D, Thompson AJ, Fox RJ, Cohen JA (2017) Progressive multiple sclerosis: prospects for disease therapy, repair, and restoration of function. Lancet 389:1357-1366.
- Pahlevan Kakhki M, Nikravesh A, Shirvani Farsani Z, Sahraian MA, Behmanesh M (2018) HOTAIR but not ANRIL long non-coding RNA contributes to the pathogenesis of multiple sclerosis. Immunology 153:479-487.
- Paraskevopoulou MD, Vlachos IS, Karagkouni D, Georgakilas G, Kanellos I, Vergoulis T, Zagganas K, Tsanakas P, Floros E, Dalamagas T, Hatzigeorgiou AG (2016) DIANA-LncBase v2: indexing microRNA targets on non-coding transcripts. Nucleic Acids Res 44: D231-238.
- Patoughi M, Ghafouri-Fard S, Arsang-Jang S, Taheri M (2019) GAS8 and its naturally occurring antisense RNA as biomarkers in multiple sclerosis. Immunobiology 224:560-564.
- Petri A, Dybkaer K, Bogsted M, Thrue CA, Hagedorn PH, Schmitz A, Støve Bødker J, Johnsen HE, Kauppinen S (2015) Long noncoding RNA expression during human B-cell development. PLoS One 10:e0138236.
- Ren C, An G, Zhao C, Ouyang Z, Bo X, Shu W (2018). Lnc2Catlas: an atlas of long noncoding RNAs associated with risk of cancers. Sci Rep 8:1909.
- Rezazadeh M, Gharesouran J, Moradi M, Noroozi R, Omrani MD, Taheri M, Ghafouri-Fard S (2018) Association Study of ANRIL Genetic Variants and Multiple Sclerosis. J Mol Neurosci 65:54-59.
- Riege K, Hölzer M, Klassert TE, Barth E, Bräuer J, Collatz M, Hufsky F, Mostajo N, Stock M, Vogel B, Slevogt H, Marza M (2017) Massive effect on IncRNAs in human monocytes during fungal and bacterial infections and in response to vitamins A and D. Sci Rep 7:40598.
- Rossi S, Motta C, Studer V, Macchiarulo G, Volpe E, Barbieri F, Ruocco G, Buttari F, FinardiA, Mancino R,Weiss S, Battistini L,Martino G, Furlan R, Drulovic J, Centonze D (2014) Interleukin-1β causes excitotoxic neurodegeneration and multiple sclerosis disease progression by activating the apoptotic protein p 53. Mol Neurodegener 9:56.
- Santoro M, Nociti V, Lucchini M, De Fino C, Losavio FA, Mirabella M (2016) Expression profile of long non-coding RNAs in serum of patients with multiple sclerosis. J Mol Neurosci 59:18-23.
- Santoro M, Nociti V, Lucchini M, Loiodice M, Centofanti F, Botta A, Losavio FA, De Fino C, Mirabella M (2020) A pilot study of IncRNAs expression profile in serum of progressive multiple sclerosis patients. Eur Rev Med Pharmacol Sci 24:3267-3273.
- Schmitz SU, Grote P, Herrmann BG (2016) Mechanisms of long noncoding RNA function in development and disease. CMLS 73:2491-2509.

- Shaker OG, Golam RM, Ayoub S, Daker LI, Elguaad MKA, Said ES, Khalil MAF (2020) Correlation between LincR-Gng2-5'and LincR-Epas1-3'as with the severity of multiple sclerosis in Egyptian patients. Int J Neurosci 130:515-521.
- Shaker OG, Mahmoud RH, Abdelaleem OO, Ibrahem EG, Mohamed AA, Zaki OM, Abdelghaffar NK, Ahmed TI, Hemeda NF, Ahmed NA, Mansour DF (2019) LncRNAs, MALAT1 and Inc-DC as potential biomarkers for multiple sclerosis diagnosis. Biosci Rep 39: BSR20181335.
- Sigdel KR, Cheng A, Wang Y, Duan L, Zhang Y (2015) The emerging functions of long noncoding RNA in immune cells: autoimmune diseases. J Immunol Res 2015:848790.
- Stys PK, Tsutsui S (2019) Recent advances in understanding multiple sclerosis. F1000Res 8:F1000 Faculty Rev-2100.
- Sun L, Goff LA, Trapnell C, Alexander R, Lo KA, Hacisuleyman E, Sauvageau M, Tazon-Vega B, Kelley DR, Hendrickson DG, Yuan B, Kellis M, Lodish HF, Rinn JL (2013) Long noncoding RNAs regulate adipogenesis. Proc Natl Acad Sci U S A 110:3387-3392.
- Sung TL, Rice AP (2006) Effects of prostratin on cyclin T1/P-TEFb function and the gene expression profile in primary resting CD4+ T cells. Retrovirology 3:66.
- Taheri M, Noroozi R, Sadeghpour S, Omrani MD, Ghafouri-Fard S (2020) The rs4759314 SNP within Hotair IncRNA is associated with risk of multiple sclerosis. Mult Scler Relat Disord 40:101986.
- Tang YJ, Zhou T, Yu X, Xue Z, Shen N (2017) The role of long non-coding RNAs in rheumatic diseases. Nat Rev Rheumatol 13:657-669.
- Tayari MM, Winkle M, Kortman G, Sietzema J, de Jong D, Terpstra M, Mestdagh P, Kroese FG, Visser L, Diepstra A, Kok K, van den Berg A, Kluiver J (2016) Long noncoding RNA expression profiling in normal B-cell subsets and hodgkin lymphoma reveals Hodgkin and Reed-Sternberg cell-specific long noncoding RNAs. Am J Pathol 186:2462-2472.
- Thompson, AJ, Baranzini, SE, Geurts J, Hemmer B, Ciccarelli O (2018) Multiple sclerosis. Lancet 391:1622-1636.
- Tong Q, Gong AY, Zhang XT, Lin C, Ma S, Chen J, Hu G, Chen X (2016) LincRNA-Cox2 modulates TNFalpha- induced transcription of Il12b gene in intestinal epithelial cells through regulation of Mi-2/NuRD-mediated epigenetic histone modifications. FASEB J 30:1187-1197.
- Ulitsky I and Bartel DP (2013) lincRNAs: Genomics, evolution, and mechanisms. Cell 154:26-46.
- Villegas VE, Zaphiropoulos PG (2015) Neighboring gene regulation by antisense long non-coding RNAs. Int J Mol Sci 16:3251-3266.
- Wang H, Wang Y, Xie S, Liu Y, Xie Z (2017) Global and cell-type specific properties of lincRNAs with ribosome occupancy. Nucleic Acids Res 45:2786-2796.
- Wang Y, Zhong H, Xie X, Chen CY, Huang D, Shen L, Zhang H, Chen ZW, Zeng G (2015) Long noncoding RNA derived from CD244 signaling epigenetically controls CD8+ T-cell immune responses in tuberculosis infection. Proc Natl Acad Sci 112:E3883-3892.
- Wapinski O, Chang HY (2011) Long noncoding RNAs and human disease. Trends Cell Biol 21:354-361.
- Wu GC, Pan HF, Leng RX, Wang DG, Li XP, Li XM, Ye DQ (2015) Emerging role of long noncoding RNAs in autoimmune diseases. Autoimmun Rev 14:798-805.
- Xu F, Jin L, Jin Y, Nie Z, Zheng H (2019) Long noncoding RNAs in autoimmune diseases. J Biomed Mater Res A 107:468-475.
- Xue M, Zhuo Y, Shan B (2017) MicroRNAs, long noncoding RNAs, and their functions in human disease. Methods Mol Biol 1617:1-25.
- Yang X, Wu Y, Zhang B, Ni B (2018) Noncoding RNAs in multiple sclerosis. Clin Epigenetics 10:149.
- Yang X, Yang J, Wang J, Wen Q, Wang H, He J, Hu S, He W, Du X, Liu S, Ma L (2016) Microarray analysis of long noncoding RNA and mRNA expression profiles in human macrophages infected with mycobacterium tuberculosis. Sci Rep 6:38963.
- Ye Y, Xu Y, Lai Y, He W, Li Y, Wang R, Luo X, Chen R, Chen T (2018) Long non-coding RNA cox-2 prevents immune evasion and metastasis of hepatocellular carcinoma by altering M1/M2 macrophage polarization. J Cell Biochem 119:2951-2963.
- Zhang F, Liu G, Li D, Wei C, Hao J (2018) DDIT4 and associated IncDDIT4 modulate Th17 differentiation through the DDIT4/TSC/mTOR pathway. J Immunol 200:1618-1626.
- Zhang F, Liu G, Wei C, Gao C, Hao J (2017) Linc-MAF-4 regulates Th1/Th2 differentiation and is associated with the pathogenesis of multiple sclerosis by targeting MAF. FASEB J 31:519-525.
- Zhang Q, Chen L, Cui S, Li Y, Zhao Q, Cao W, Lai S, Yin S, Zuo Z, Ren J (2017) Expression and regulation of long noncoding RNAs during the osteogenic differentiation of periodontal ligament stem cells in the inflammatory microenvironment. Sci Rep 7:13991.
- Zhang Y, Zhang XO, Chen T, Xiang JF, Yin QF, Xing YH, Zhu S, Yang L, Chen LL (2013) Circular intronic long noncoding RNAs. Mol Cell 51:792-806.
- Zhu J, Yamane H, Paul WE (2010) Differentiation of effector CD4 T cell populations. Ann Rev Immunol 28:445-489.

P-Reviewer: Duman T; C-Editors: Zhao M, Qiu Y; T-Editor: Jia Y