

RESEARCH ARTICLE

Reviving the sound of a 150-year-old insect: The bioacoustics of *Prophalangopsis obscura* (Ensifera: Hagloidea)

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Abstract

Determining the acoustic ecology of extinct or rare species is challenging due to the inability to record their acoustic signals or hearing thresholds. Katydid and their relatives (Orthoptera: Ensifera) offer a model for inferring acoustic ecology of extinct and rare species, due to allometric parameters of their sound production organs. Here, the bioacoustics of the orthopteran *Prophalangopsis obscura* are investigated. This species is one of only eight remaining members of an ancient family with over 90 extinct species that dominated the acoustic landscape of the Jurassic. The species is known from only a single confirmed specimen—the 150-year-old holotype material housed at the London Natural History Museum. Using Laser-Doppler Vibrometry, 3D surface scanning microscopy, and known scaling relationships, it is shown that *P. obscura* produces a pure-tone song at a frequency of ~4.7 kHz. This frequency range is distinct but comparable to the calls of Jurassic relatives, suggesting a limitation of early acoustic signals in insects to sonic frequencies (<20 kHz). The acoustic ecology and importance of this species in understanding ensiferan evolution, is discussed.

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Introduction

Acoustic communication systems have long been a popular model for understanding ecological and evolutionary relationships within and between species. In insects, acoustic systems for both signal generation and recognition have evolved a substantial variety of forms, to facilitate a range of communication functions [1, 2], offering many routes to studying the evolution of acoustic communication. However, for extinct and rare insect species, we are often limited in our abilities to infer details of specific communication systems, as we are unable to record the sounds such species generate or measure their hearing capabilities.

In katydids (or bush-crickets; Orthoptera: Ensifera) and their allies, pure-tone and broad-band sound production has evolved as a key mechanism for mate attraction and conspecific recognition [3–6]. These sounds are produced by tegminal stridulation—the process of moving a hardened scraper on one forewing, against a row of teeth (the file) on the other, producing vibrations on the wing which are then amplified by specialized wing cells (namely the harp

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Abbreviations: f_c , carrier frequency; f_o , resonant frequency; LW, left wing; RW, right wing.

and mirror) [4, 5] to radiate sound. This mechanism of sound production is evolutionarily conserved across a majority of the Ensifera, and its characteristics have been understood since the early 1900s [7, 8]. The retention of this mechanism across a diverse range of taxa, and the increasing ability of state-of-the-art imaging and acoustic technologies, is rapidly allowing researchers to re-visit once inaccessible specimens with novel methodologies to advance our understanding of Ensiferan acoustic communication.

Here we investigate the bioacoustics of *Prophalangopsis obscura* (Walker, 1869) (Ensifera: Prophalangopsidae), an insect belonging to an ancient katydid family of over 90 known species dominant during the Jurassic, with only eight extant members [9]. The genus *Prophalangopsis*, formerly *Tarraga*, has remained monotypic ever since the discovery of *P. obscura*, and thus received considerable interest in relation to the evolutionary history of the Ensifera [10–12]. The enigmatic nature of the type specimen has been compounded by no further male specimens being discovered in over 150 years, and only 2 potential female specimens ever found [13]. In addition, no works have explored the ecology of this species due to their uncertain geographic distribution [13, 14]. Therefore, a thorough study of their acoustic capabilities could improve our understanding of the communication systems and acoustic ecology of *P. obscura* and its long extinct relatives [15–17], and aid in future rediscovery of the species.

Using micro-scanning Laser-Doppler Vibrometry (LDV), we reconstruct the vibration patterns and resonances of the sound production organs (forewings) of the *P. obscura* type specimen. Furthermore, we investigate the morphology of the stridulatory apparatus and tegmina in detail to compliment LDV experiments and infer the likely carrier frequency (f_c) of this species' song over 150 years after specimen preservation. Employing existing validated models, and novel measurements from LDV, we obtain f_c for the acoustic signal of *P. obscura* and use morphological data to calculate acoustic signal structure. Using knowledge of the wing biomechanics of other extant members of this ancient family, we reconstruct the calling song of *P. obscura*, and discuss the importance of this species in understanding the evolution of ensiferan acoustic communication.

Materials and methods

The holotype material

Prophalangopsis obscura (Walker 1869) is a large orthopteran (~10 cm; tegmina wingspan) represented by a single specimen housed at the London Natural History Museum, South Kensington, UK (specimen NHMUK 013806185). Collection details are scarce, with the location information listed only as 'India'. The specimen was originally set in a resting position following collection, but sometime between 1898 and 1939, the specimen was re-mounted with both wings spread [10], a position which remains to this day (Fig 1). At an unknown time after 1939, the left foretibia was lost. The right foretibia, which contains the tympanic ear, remains intact (Fig 1C). Both forewings are present, with the stridulatory (sound producing) organs intact, however the left wing is torn along the apical axis (Fig 1A). In 2005, two female specimens identified as *P. obscura* were located in China, later published by Liu et al. [13]. While male specimens were not identified to confirm the identity of these specimens, they minimally belong to a close relative of *P. obscura*. No permits were required for the described study, which complied with all relevant regulations.

Tegmina and stridulatory file anatomy

P. obscura possesses a stridulatory file on each forewing. The morphology of each file was imaged using an Alicona InfiniteFocus microscope (Bruker Alicona Imaging, Graz, Austria) at 20x objective magnification, resulting in one composite 3D-image of each file with a vertical

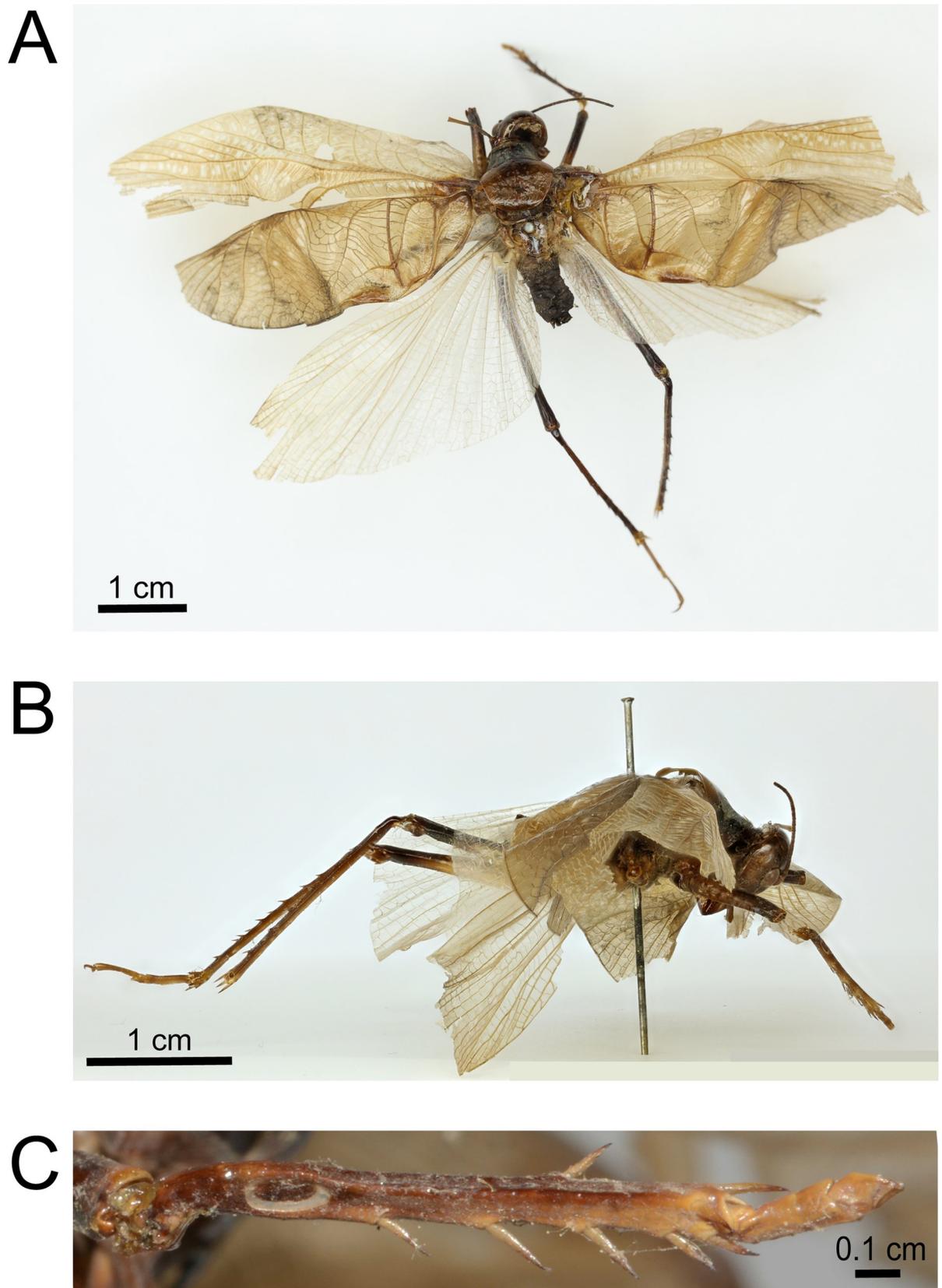


Fig 1. The holotype of *Prophalangopsis obscura* (collected in India, Walker 1869). A, dorsal habitus; B, lateral habitus; C, tympanal organ.

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and horizontal resolution of 0.7 and 7.8 μm , respectively. Using the built-in Alicona software, the length of the stridulatory file was measured, as well as the spacing between stridulatory teeth (inter-tooth distances), and length of each tooth. The inter-tooth distance was measured as the distance between the central tip (cusp) of adjacent teeth.

Forewing resonance and deflection pattern

The resonant frequency (f_o) and deflection patterns of the forewings was measured in the holotype of *P. obscura* using micro-scanning LDV (PSV-500, Polytec GmbH, Waldbronn, Germany), with approximately 1000 measuring points at a sampling frequency of 256 kHz. Acoustic signals for wing excitation were generated by the LDV internal data acquisition board (PCI-4451; National Instruments, Austin, TX, USA), and consisted of broadband periodic chirps ranging from 1 to 60 kHz at 60 dB SPL (re 20 μPa). The signal was amplified by a Pioneer A-400 amplifier (Pioneer, Kawasaki, Japan) and transmitted to a loudspeaker (Vifa, Avisoft Bioacoustics, Glienicke, Germany; flattened frequency response across the whole range) positioned 20 cm in front of the specimen. A reference signal to calculate the transfer function between the wing vibration and the stimulus was recorded using a 1/8" condenser microphone positioned horizontally at the wing plane between the wings (Model 4138-A015, Brüel & Kjaer, Nærum, Denmark). For further details of method, see [18].

Reconstruction of the sound

To reconstruct the sound of *P. obscura*, several characteristics of the acoustic signal are required. These are (1) the song carrier frequency (f_c), (2) the decay of a single stridulatory tooth strike, (3) the number of oscillations produced during each stridulatory file strike (one full sound pulse), and (4) the timing between stridulatory file strikes. Previous investigations into the morphological parameters of katydid stridulatory apparatus have shown that the best predictors of the f_c are the regions of mechanical displacement of the tegmina (the acoustically active wing cells), and the length of the stridulatory file [7]. In hagloids (Haglidæ and Prophalangopsidæ), which lack a specialised mirror area, it has been suggested that measurements of file length and LDV resonance, or the entire vibrating area, will be better predictors of f_c [7, 8]. To predict f_c , we used existing models [7] to compare the frequency derived from the file length, right tegmen vibrational area, left tegmen vibrational area, and resonance from vibrometry.

Following calculation of the mean f_c predicted by these four techniques, an artificial impulse of a single tooth strike of *P. obscura* was produced at this frequency, including a decay caused by damping. Oscillations of the tegmina mirror cells usually exhibit a free decay of 3–4 ms in species communicating at the determined f_c [3, 11, 19], thus a 4 ms exponential decay was used.

Members of the Prophalangopsidæ have a high stridulatory tooth density and short functional file length, which permits the generation of uniquely pure-tone calls [15, 20, 21], and as pure-tone singing katydids display a 1:1 relation between tooth strikes and the number of oscillations in the song pulse [7], we used f_c , the number of functional stridulatory teeth, and the spacing of the teeth, to infer the pulse structure of the song of *P. obscura*. This was performed using a custom written Matlab code [15] which calculates the instantaneous period for each tooth impact based on the inter-tooth distance measurements. The resulting representative waveform of the acoustic signal of *P. obscura* was further analysed using the Signal Processing Toolbox in Matlab (R2021a, The MathWorks Inc., Natick, USA) with the following spectrogram parameters: FFT size 512, Hamming window, 50% overlap; frequency resolution: 512 Hz, temporal resolution: 0.15 ms.

Results

Tegmina and stridulatory file anatomy

The anatomy of the tegmina (forewings) of *P. obscura* is similar to those observed in both extant and extinct relatives of the Prophalangopsidae. The left wing (LW) and right wing (RW) display stridulatory files that are similar enough to be considered functionally symmetrical. The pattern of tooth distribution is slightly gaussian (Fig 2), suggesting the file could be adapted for sound production during the opening or closing phase of the wings. File length and number of teeth of LW and RW file were 9.60 and 9.99 mm and 134 and 137 teeth, respectively. The inter-tooth distances, tooth lengths, and plectra are symmetrical, suggesting both might have been capable of producing sound pulses (S1 Table).

Forewing resonance and deflection pattern

Despite over 150 years of preservation, it was possible to obtain the deflection (vibratory) pattern of the forewings and f_o in *P. obscura*. An assessment of the regions of the wings theoretically involved in resonant sound production and the displacement of the wings in response to an acoustic stimulus (Fig 3A and 3D) confirmed that the mirror and pre-mirror are the most likely regions for sound production in this species, as with all extant members of this family [21]. Displacement was highest within the mirror area for both the LW (Fig 3B) and RW (Fig 3E). The normalised displacement amplitudes of the mirror area of the LW displayed a peak frequency at 6.3 kHz (Fig 3C). However, despite morphological symmetry of the wings, the RW displayed a peak of 4.8 kHz (Fig 3F).

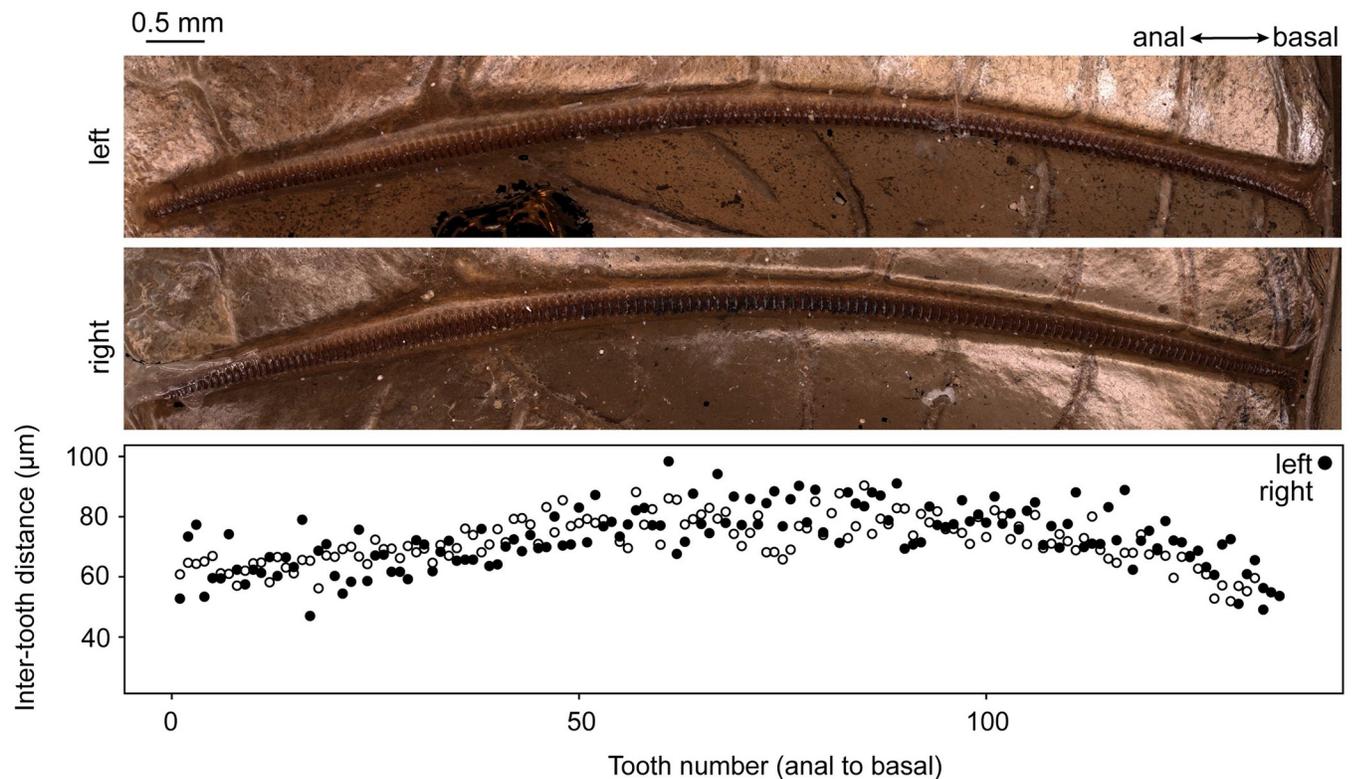


Fig 2. Stridulatory file anatomy and inter-tooth distances in *Prophalangopsis obscura*. Orientation of both files is along the anal (left) to basal (right) axis.

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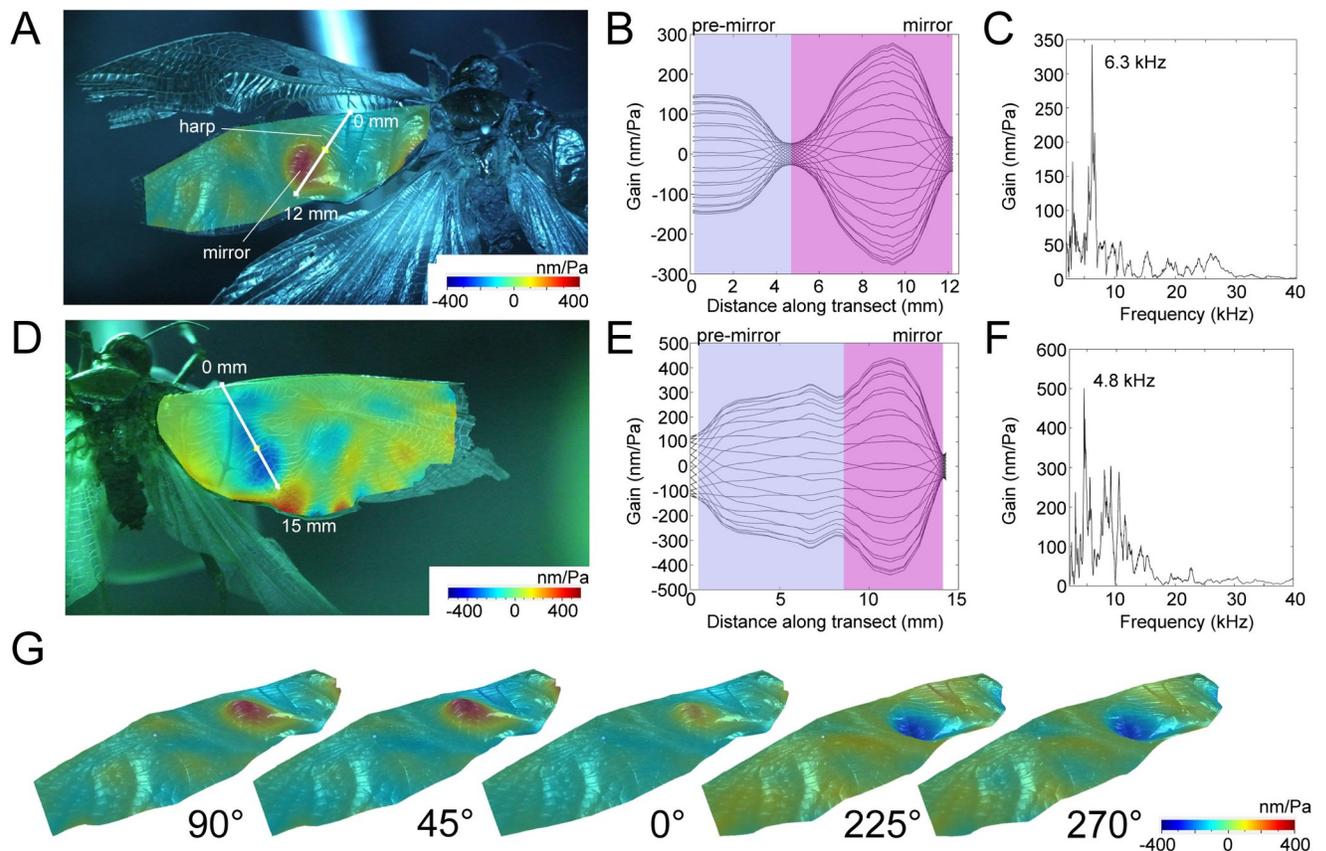


Fig 3. Forewing resonance in *Prophalangopsis obscura*. (A) Displacement map of the LW; (B) Deflection pattern of the white profile line in A; (C) Frequency spectrum of the left mirror; (D) Displacement map of the RW; (E) Deflection pattern of the profile line in D; (F) Frequency spectrum of the right mirror; (G) Angled view of the left forewing displacement pattern at 4.8 kHz.

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Reconstruction of the sound

Using f_0 , stridulatory file length, and vibrational areas of the tegmina resulting from LDV deflection measurements, f_c was calculated (Table 1). Based on phylogenetically controlled linear models of several measurement parameters [7], we believe the f_c to be $\sim 4.7 \pm 0.05$ kHz (Table 1). The measurements of inter-tooth distances and f_c allowed the calculation of a time vector of a single sound pulse of the species' acoustic signal (For more details of the song reconstruction method, see [15]).

The Matlab script for sound pulse reconstruction [15] revealed that the structure of a single call pulse (Fig 4) is similar to that of fossil relatives of the same family [15], but differs in

Table 1. Model measurement parameters for calculation of the likely carrier frequency (f_c) of *Prophalangopsis obscura*.

Measurement parameter	Measurement (x)	Slope (m)	Intercept (c)	f_c (kHz)
File length (mm)	9.6	-0.97	3.74	4.693
RW vibrating area (mm ²)	45.31	-0.62	3.91	4.691
LW vibrating area (mm ²)	39.05	-0.54	3.53	4.716
LDV resonance (kHz)				4.800
Average				4.725

For all estimates of f_c : $\ln(f_c) = m * \ln(x) + c$, where \ln = natural logarithm.

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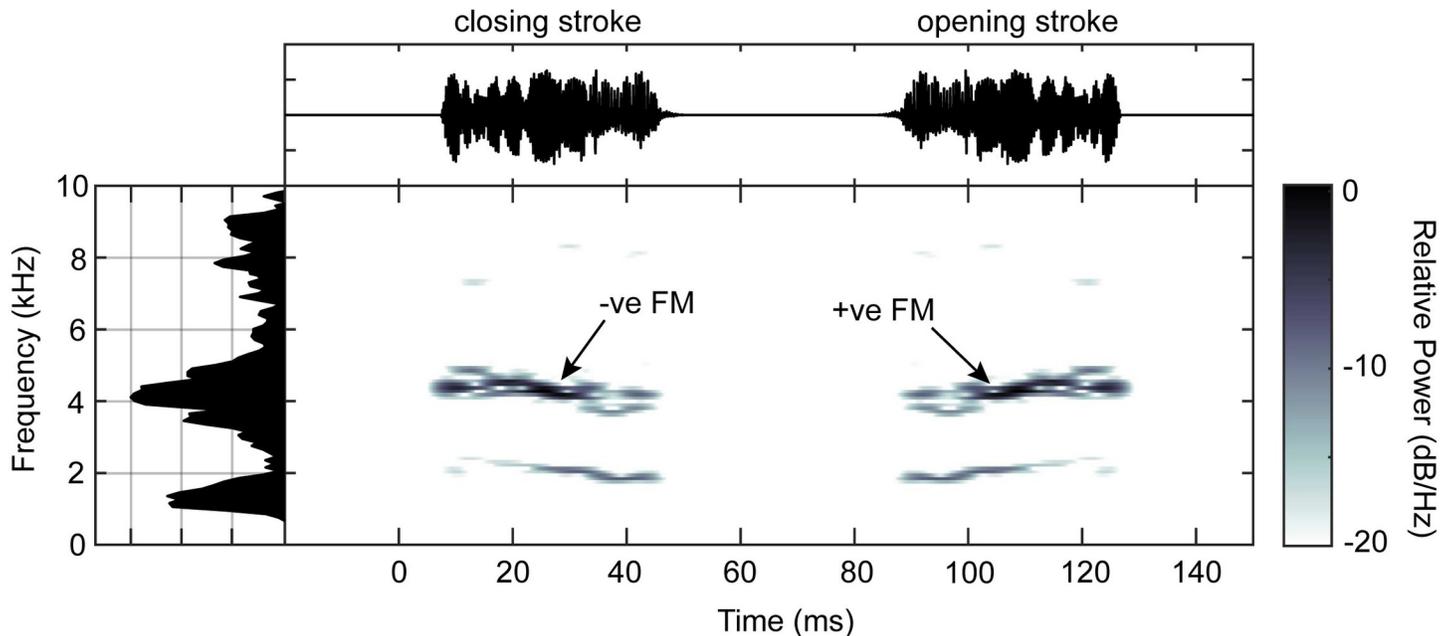


Fig 4. Reconstruction and spectral analysis of a diplo-syllable containing two pulses of the sound of *Prophalangopsis obscura*. Waveform of two chirps (top), with spectrogram below and frequency spectrum on the left marginal axis. The 2nd chirp is an artificial reversal of the 1st chirp, to demonstrate that frequency modulation (FM) will differ depending on whether sound is produced during the opening or closing wing stroke.

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frequency and duration (Fig 4). The duration of a single pulse was found to be 42 ms (Fig 4), which is very close to the predicted pulse duration from functional file length using an existing model (40.78 ms; Montealegre-Z et al. 2017). Surprisingly, a slight frequency modulation in each chirp of the call was observed (Fig 4). Extant species display similar modulations as a result of changing velocity over the course of each wing stroke. The first predictions of how crickets produced their sounds looked at tooth distribution to infer whether sounds are produced during opening or closing of the wings [22]. As the frequency of the sound is a function of tooth strikes per time period, any changes to wing velocity over the course of one wing stroke will cause frequency modulation in the sound. Looking at the almost gaussian distribution of seemingly functional teeth in *P. obscura*, we cannot confirm whether this species is able to stridulate during the opening or closing of the wings, or both. The final reconstruction of the sound (Fig 4; S1 Audio) therefore consists of a putative diplo-syllable containing two pulses with every other chirp artificially reversed, to leave this element of the reconstruction open for future interpretations.

Discussion

Using LDV techniques, we were able to obtain the deflection pattern of the tegmina of *P. obscura* and use information on tegmina and stridulatory file anatomy to reconstruct the song of the 150-year-old preserved museum specimen. The anatomy of the tegmina and stridulatory file display similarities to both fossil and extant prophalangopsids [15, 16, 21], and this similarity is also represented in the frequency and structure of the song (Fig 4), although f_c here is lower than that of related fossil species [15]. The resonant frequency (f_o) of the tegmina provided by the LDV recordings matched the expected frequency from the models, and we were able to obtain the area of deflection, which was also used to calculate the potential calling song frequency (Table 1). Despite the matching frequency information provided by the right

tegmen, the left tegmen did not predict a similar f_c . We believe this discrepancy is due to a tear down the apical axis of the LW, given the similarities of the frequency predicted by the models to the resonance of the right tegmen (Table 1). In many singing ensiferans, the LW is found to be a better predictor of f_c [7], however in the prophalangopsidae, it is known that the wings are functionally symmetrical [7], so we can be confident that the RW resonance is representative of the LW resonance. The retention of resonance in the RW may seem surprising, as insect cuticles become stiffer as they desiccate over time [23], and thus we may expect such stiffening to result in changes to resonance. However, in this case, we believe that the topology of the wings plays more of an important role in f_c , and due to the size of the tegmina, the effect of drying is not so pronounced. The thickness and area of the tegmina dictate the resonant properties of the musical areas of the wing, and larger musical areas display less variation in frequency response with changes to thickness [24]. For example, in the gryllid *Tarbinskiellus portentosus* with a harp size of $\sim 25 \text{ mm}^2$, tegmina thickness would need to decrease by more than 30% before thickness would begin to greatly modify resonance [24]. Thus, for a large species like *P. obscura* which has a harp size of $\sim 50 \text{ mm}^2$, even a significant change in tegmina thickness from desiccation would be unlikely to result in large changes to resonance, explaining why the resonance is here maintained. Nonetheless, further studies into the effects of wing thickness and tissue desiccation on tegmina resonance across orthopterans would offer a rich dataset for future works to calculate taxon-specific frequency changes over time, increasing the information we can obtain from dry museum specimens.

Just like the other extant members of this family, and unlike modern katydid (Tettigoniidae), *P. obscura* is likely capable of using both wings for singing, with both tegmina possessing symmetrical stridulatory files, plectra, and acoustically functional areas [15, 16, 21]. The mirror region of the tegmina displayed the greatest deflection, and the pattern of deflection followed that of extant relatives [21]. However, as suggested by Zeuner [10], the tegmina are not as specialised for sound production as other closely related extant species such as the great Grig *Cyphoderris monstrosa*. The size and function of the wings is one of the key features of *P. obscura* that separates it from the other extant prophalangopsids and resembles the specimens of the fossil record [10, 15]. While all other extant prophalangopsids (e.g. *Cyphoderris* spp.) are flightless and use their wings exclusively for sound production and mate attraction/gifting [25], *P. obscura* has wings potentially large enough for short or sustained flight, resembling both the extinct prophalangopsids and many tettigoniids.

Reduced flight is a well-established evolutionary mechanism to reduce or avoid predation by aerial predators, and in particular, bats [26]. The other extant species in this family, all of whom have lost the ability to fly, exhibit novel anti-predator defences, namely ultrasonic sound production organs [27], which likely evolved to act as a deterrent to a new host of predators they now face after switching to a terrestrial lifestyle. Such anti-predator adaptations are not present in *P. obscura*, nor are any other morphological adaptations associated with predation by bats such as enlarged cuticular spines [26]. We may predict therefore that this species lives in a region with reduced selection pressure from ultrasonic aerial predators, allowing it to retain the Jurassic form even after the emergence of echolocating bats [28]. Similarly, low frequency calling songs such as that of *P. obscura* are indicative of reduced pressures from eavesdropping predators, as low frequency sounds travel larger distances and could give away the location of the signaller [29]. Tettigoniids regularly preyed upon by bats benefit from the increased attenuation of ultrasonic conspecific signals by a reduced detection range by eavesdropping predators [26]. However, it should be noted that correlating call frequency to ecology in such a manner does not consider other factors which will be driving call frequency evolution [26, 29].

Unfortunately, further inferences on natural history remain challenging as the precise origins of the type specimen remain obscure. Previous literature on the specimen references a

wide geographic area broadly synonymous with the extent of the former British India at the time of collection (e.g. Hindustan, E. Indies). The combined historical evidence suggests that the specimen was collected in northern India, although it is at present not possible to give a more precise location. If the female specimen described in Liu et al. [13] are confirmed to be *P. obscura* and not a closely related species, then the known range may be extended from northern India to include Tibet, a region certainly too cold to support an abundance of echolocating bats. Further collections from this area to confirm the association between males and females, and to investigate the local composition of potential predators, would be very valuable.

Following this song reconstruction, it may be plausible to deploy autonomous recording units (ARUs) into potential field sites and use signal detection algorithms to aid in the rediscovery of this species [30, 31]. We hope that in time, further specimens of *Prophalangopsis obscura* are located, to record the true song of this elusive species, and to validate the accuracy of the predictions presented here.

Supporting information

S1 Table. Morphological characters of the tegmina stridulatory files of *P. obscura*.
(DOCX)

S1 Audio. The reconstructed calling song of *Prophalangopsis obscura*.
(WAV)

S1 Fig. Visual reconstruction of *Prophalangopsis obscura* on a tree branch in a temperate montane habitat. Illustrated by CW.
(JPG)

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Formal analysis: Charlie Woodrow.

Funding acquisition: Fernando Montealegre-Z.

Investigation: Charlie Woodrow, Ed Baker, Thorin Jonsson, Fernando Montealegre-Z.

Methodology: Charlie Woodrow, Thorin Jonsson, Fernando Montealegre-Z.

Resources: Ed Baker, Thorin Jonsson.

Supervision: Fernando Montealegre-Z.

Visualization: Charlie Woodrow.

Writing – original draft: Charlie Woodrow.

Writing – review & editing: Ed Baker, Thorin Jonsson, Fernando Montealegre-Z.

References

1. Drosopoulos S, Claridge MF. *Insect Sounds and Communication: Physiology, Behaviour, Ecology and Evolution*. CRC Press, Taylor and Francis; 2006. <https://doi.org/10.1201/9781420039337>
2. Yack JE. The structure and function of auditory chordotonal organs in insects. *Microsc Res Tech*. 2004; 63: 315–337. <https://doi.org/10.1002/jemt.20051> PMID: 15252876
3. Gwynne DT. Phylogeny of the Ensifera (Orthoptera): A Hypothesis Supporting Multiple Origins of Acoustical Signalling, Complex Spermatophores and Maternal Care in Crickets, Katydid, and Weta. *J Orthoptera Res*. 1995; 4: 203–218. <https://doi.org/10.2307/3503478>
4. Bennet-Clark HC. Wing resonances in the Australian field cricket *Teleogryllus oceanicus*. *J Exp Biol*. 2003; 206: 1479–1496. <https://doi.org/10.1242/jeb.00281> PMID: 12654887
5. Montealegre-Z F, Mason AC. The mechanics of sound production in *Panacanthus pallicornis* (Orthoptera: Tettigoniidae: Conocephalinae): The stridulatory motor patterns. *J Exp Biol*. 2005; 208: 1219–1237. <https://doi.org/10.1242/jeb.01526> PMID: 15781883
6. Morris GK. Song in Arthropods. In: Davey KG, editor. *Encyclopedia of Reproduction*, Vol 4. Academic Press; 1999. pp. 508–517.
7. Montealegre-Z F, Ogden J, Jonsson T, Soulsbury CD. Morphological determinants of signal carrier frequency in katydids (Orthoptera): a comparative analysis using biophysical evidence of wing vibration. *J Evol Biol*. 2017; 30: 2068–2078. <https://doi.org/10.1111/jeb.13179> PMID: 28921699
8. Chivers BD, Jonsson T, Soulsbury CD, Montealegre-Z F. Structural biomechanics determine spectral purity of bush-cricket calls. *Biol Lett*. 2017; 13: 0–5. <https://doi.org/10.1098/rsbl.2017.0573> PMID: 29187608
9. Cigliano, M.M., Braun, H., Eades, D.C., Otte D. Orthoptera Species File Version 5.0/5.0. 2021 [cited 20 Dec 2019]. Available: <http://orthoptera.speciesfile.org>
10. Zeuner FE. *Fossil Orthoptera Ensifera*. British Museum of Natural History; 1939.
11. Desutter-Grandcolas L. Phylogeny and the evolution of acoustic communication in extant Ensifera (Insecta, Orthoptera). *Zool Scr*. 2003; 32: 525–561.
12. Song H, Amédégnato C, Cigliano MM, Desutter-Grandcolas L, Heads SW, Huang Y, et al. 300 million years of diversification: Elucidating the patterns of orthopteran evolution based on comprehensive taxon and gene sampling. *Cladistics*. 2015; 31: 621–651. <https://doi.org/10.1111/cla.12116> PMID: 34753270
13. Liu X, Zhou M, Bi W, Tang L. New data on taxonomy of recent Prophalangopsidae (Orthoptera: Hagloidea). *Zootaxa*. 2009; 62: 53–62.
14. Béthoux O. Grylloptera—A unique origin of the stridulatory file in katydids, crickets, and their kin (Archaeorthoptera). *Arthropod Syst Phylogeny*. 2012; 70: 43–68.
15. Gu J-J, Montealegre-Z F, Robert D, Engel MS, Qiao G-X, Ren D. Wing stridulation in a Jurassic katydid (Insecta, Orthoptera) produced low-pitched musical calls to attract females. *Proc Natl Acad Sci U S A*. 2012; 109: 3868–3873. <https://doi.org/10.1073/pnas.1118372109> PMID: 22315416
16. Gu AJ, Qiao G, Ren D. Revision and new taxa of fossil Prophalangopsidae (Orthoptera: Ensifera) Published by: Orthopterists' Society Stable URL: <http://www.jstor.org/stable/20789566> REFERENCES Linked references are available on JSTOR for this article: You may need to log. *J Orthoptera Res*. 2010; 19: 41–56.
17. Gorochov A V. New data on taxonomy and evolution of fossil and Recent Prophalangopsidae (Orthoptera: Hagloidea). *Acta Zool cracoviensia*. 2003; 46: 117–127.
18. Montealegre-Z F, Windmill JFC, Morris GK, Robert D. Mechanical phase shifters for coherent acoustic radiation in the stridulating wings of crickets: The plectrum mechanism. *J Exp Biol*. 2009; 212: 257–269. <https://doi.org/10.1242/jeb.022731> PMID: 19112145
19. Field LH. Structure and evolution of stridulatory mechanisms in New Zealand wetas (Orthoptera: Stenopelmatidae). *Int J Insect Morphol Embryol*. 1993; 22: 163–183. [https://doi.org/10.1016/0020-7322\(93\)90008-O](https://doi.org/10.1016/0020-7322(93)90008-O)
20. Morris GK, DeLuca PA, Norton M, Mason AC. Calling-song function in male haglids (Orthoptera: Haglidae, Cyphoderris). *Can J Zool*. 2002; 80: 271–285. <https://doi.org/10.1139/z02-003>
21. Chivers BD, Béthoux O, Sarria-S FA, Jonsson T, Mason AC, Montealegre-Z F. Functional morphology of tegmina-based stridulation in the relict species *Cyphoderris monstrosa* (Orthoptera: Ensifera: Prophalangopsidae). *J Exp Biol*. 2017; 220: 1112–1121. <https://doi.org/10.1242/jeb.153106> PMID: 28082619
22. Lutz FE, Hicks WR. An analysis by movietone of a cricket's chirp (*Gryllus assimilis*). *Am Museum Novit*. 1930; 1–14.

23. Klocke D, Schmitz H. Water as a major modulator of the mechanical properties of insect cuticle. *Acta Biomater.* 2011; 7: 2935–2942. <https://doi.org/10.1016/j.actbio.2011.04.004> PMID: 21515418
24. Godthi V, Balakrishnan R, Pratap R. The mechanics of acoustic signal evolution in field crickets. *J Exp Biol.* 2022; 225. <https://doi.org/10.1242/jeb.243374> PMID: 35258611
25. Judge KA, de Luca PA, Morris GK. Food limitation causes female haglids to mate more often. *Can J Zool.* 2011; 89: 992–998. <https://doi.org/10.1139/z11-078>
26. Morris GK, Mason AC, Wall P, Belwood JJ. High ultrasonic and tremulation signals in neotropical katydids (Orthoptera: Tettigoniidae). *J Zool.* 1994; 233: 129–163. <https://doi.org/10.1111/j.1469-7998.1994.tb05266.x>
27. Woodrow C, Judge KA, Pulver C, Jonsson T, Montealegre-Z F. The Ander's organ: a mechanism for anti-predator ultrasound in a relict orthopteran. *J Exp Biol.* 2021; 224: jeb237289. <https://doi.org/10.1242/jeb.237289> PMID: 33443038
28. Teeling EC, Dool S, Springer MS. Phylogenies, fossils and functional genes: the evolution of echolocation in bats. In: Gunnell GF, Simmons NB, editors. *Evolutionary History of Bats: Fossils, Molecules and Morphology.* Cambridge University Press; 2012. pp. 1–22.
29. Heller K - G. Acoustic signalling in palaeotropical bushcrickets (Orthoptera: Tettigoniodea: Pseudophyllidae): does predation pressure by eavesdropping enemies differ in the Palaeo- and Neotropics? *J Zool.* 1995; 237: 469–485. <https://doi.org/10.1111/j.1469-7998.1995.tb02775.x>
30. Pérez-Granados C, Bustillo-de la Rosa D, Gómez-Catasús J, Barrero A, Abril-Colón I, Traba J. Autonomous recording units as effective tool for monitoring of the rare and patchily distributed Dupont's Lark *Chersophilus duponti*. *Ardea.* 2018; 106: 139–146. <https://doi.org/10.5253/arde.v106i2.a6>
31. Schroeder KM, McRae SB. Automated auditory detection of a rare, secretive marsh bird with infrequent and acoustically indistinct vocalizations. *Ibis (Lond 1859).* 2020; 162: 1033–1046. <https://doi.org/10.1111/ibi.12805>