



# Complete Genome Sequences of *Kinneretia* sp. Strain XES5, *Shinella* sp. Strain XGS7, and *Vogesella* sp. Strain XCS3, Isolated from *Xenopus laevis* Skin

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**ABSTRACT** Here, we report the genome sequences of three bacterial isolates, *Kinneretia* sp. strain XES5, *Shinella* sp. strain XGS7, and *Vogesella* sp. strain XCS3, which were cultured from skin of adult female laboratory-bred *Xenopus laevis*.

**X***enopus laevis* (African clawed frog) is a widely used model for gene expression, vertebrate development, regeneration, and disease (1–6). We have isolated a collection of *Xenopus*-associated bacterial isolates as part of research on tadpole regeneration (6). This report presents complete genomes for *Kinneretia* sp. strain XES5, *Shinella* sp. strain XGS7, and *Vogesella* sp. strain XCS3. To date, genomes from only 2 *Kinneretia*, 17 *Shinella*, and 9 *Vogesella* species are available from the RefSeq database; many were isolated from aquatic environments (7–17), consistent with the habitat of *X. laevis*.

Isolates were collected from the skin of captive-bred adult female *X. laevis* frogs from the University of Otago *Xenopus* colony. These *Xenopus* frogs are housed in a mains-water-recirculating aquarium system and fed twice weekly with salmon pellets. Aseptically collected swab samples were plated on Oxoid nutrient agar and incubated at 30°C for 48 h, and colonies were purified by streaking.

Default parameters were used for sequencing and assembly unless otherwise noted. Sequencing quality data are summarized in Table 1. For sequencing of each isolate, single colonies were streaked on a plate. From this plate, approximately 20 individual colonies were picked and placed in lysis buffer. Isolate DNA was then extracted using a Presto Mini genomic DNA (gDNA) bacteria kit. This DNA was used for both Nanopore and Illumina sequencing. Nanopore libraries were prepared with a rapid bar-coding kit (SQK-RBK004; Oxford Nanopore Technologies [ONT]) and sequenced using a GridION system (flow cell R9.4.1). Demultiplexing and base calling used Guppy (v4.3.4). Porechop (v0.2.4) (18) and Filtlong (v0.2.1) (--keep\_percent 95) (19) were used to trim adapters and filter reads.

Illumina libraries were prepared with a Nextera XT kit and sequenced on a MiSeq system using both 1 × 300-bp single-end reads and 2 × 300-bp paired-end reads. Fastp (v0.20.1) (--detect\_adapter\_for\_pe) (20) was used for adapter trimming and quality control (QC).

Reads passing QC were assembled with TruCycler (v0.5.0) (21) using Raven (v1.6.0) (22), Flye (v2.9-b1768) (23), and miniasm (v0.3-r179) (24). All assembled sequences were successfully circularized by TruCycler with the “reconcile” command. Plasmids are defined as circularized extrachromosomal sequences. Assemblies were polished with medaka (v1.4.4; ONT) and Pilon (v1.24) (25). Quality was assessed with CheckM (v1.1.3) (26); all genomes were >99% complete and had <1% contamination.

**Editor** David A. Baltrus, University of Arizona

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The authors declare no conflict of interest.

**Received** 27 October 2021

**Accepted** 29 November 2021

**Published** 16 December 2021

**TABLE 1** Summary of isolate characteristics and most closely related strains by ANI

Parameter	Data for:		<i>Vogesella</i> sp. strain XCS3
	<i>Kinneretia</i> sp. strain XESS5	<i>Shinella</i> sp. strain XGS7	
Genome size (bp)			
Total	5,810,938	5,248,830	3,889,382
Chromosome	4,026,671	4,025,507	3,724,844
Plasmid(s)	895,176, 733,869, 102,922, 33,339, 18,961	69,886, 18,961	164,538
G+C content (%)	68	64	60
CheckM completeness (%)	99.68	100	99.57
CheckM contamination (%)	0.39	0.58	0.11
No. of protein-coding genes	5,466	4,587	3,570
No. of rRNA operons	65 (3 rRNA operons)	93 (6 rRNA operons)	136 (9 rRNA operons)
Biosample accession no.	<a href="#">SAMN21437801</a>	<a href="#">SAMN21437802</a>	<a href="#">SAMN21437800</a>
SRA accession no.	<a href="#">SRR15901072</a> , <a href="#">SRR15901073</a> , <a href="#">SRR15901074</a>	<a href="#">SRR15901069</a> , <a href="#">SRR15901070</a> , <a href="#">GCA_020535545.1</a>	<a href="#">SRR15901078</a> , <a href="#">SRR15901079</a> <a href="#">GCA_020616155.1</a>
GenBank accession no.			
Landcare accession no.			
No. of Illumina reads	1,074,001	1,807,749	1,028,939
No. of Nanopore reads	63,881	82,808	389,151
Genome coverage (×)			
Illumina	67	98	106
Nanopore	85	107	347
Total	152	205	453
Nanopore read $N_{50}$ (bp)	13,683	15,020	6,172
Most closely related genomes (ANI [%])			
<i>Kinneretia asaccharophila</i> DSM 25082	13.683	<i>Shinella mureinivorans</i> 389 (12) (GenBank accession no. <a href="#">ASM7644403v2</a> ) (90.37),	<i>Vogesella mureinivorans</i> 389 (12) (GenBank accession no. <a href="#">ASM7644403v2</a> ) (90.37),
(7) (GenBank accession no. <a href="#">ASM436240v1</a> ) (89.91), <i>Kinneteria zoogloeooides</i> PQ7 (GenBank accession no. <a href="#">ASM982685v1</a> ) (88.53), <i>Shinella zoogloeooides</i> PQ7 (GenBank accession no. <a href="#">ASM357462v1</a> ) (87.88), <i>Shinella</i> sp. strain PSBB067 (GenBank accession no. <a href="#">ASM784415v2</a> ) (90.18), <i>Vogesella perlicula</i> DS-28 (9) (GenBank accession no. <a href="#">ASM784415v2</a> ) (90.18), <i>Vogesella urethralis</i> YM-1 (37) (GenBank accession no. <a href="#">ASM764404v2</a> ) (88.28), <i>Vogesella</i> sp. strain LIG4 (38) (MG taxon (GenBank accession no. <a href="#">ASM1683914v1</a> ) (87.86), <i>Shinella</i> sp. strain HZN7 (15) (GenBank accession no. <a href="#">ASM165256v1</a> ) (82.86), <i>Vogesella alkaliphila</i> KCTC 32041 (39) (GenBank accession no. <a href="#">ASM1465247v1</a> ) (82.63)			
(81.17)			

Sequences of the 16S rRNA genes were extracted using Barrnap (v0.9) (27) and compared to the NCBI 16S rRNA database (28) using BLASTn (29) to determine the five most closely related genera. Genomes from these genera were downloaded from RefSeq and compared to the isolates using FastANI (v1.32) (30) (Table 1). Genomes were uploaded to GenBank and annotated by PGAP (31, 32).

Currently, few other genomes from these genera (2 *Kinneretia* species, 17 *Shinella* species, and 9 *Vogesella* species) are available from RefSeq, and the ANI values for all three isolates are well below the 96% ANI threshold that is widely used to indicate species boundaries (33). Therefore, these genomes provide a valuable addition to our knowledge of these three genera.

**Data availability.** All sequencing data have been deposited in DDBJ/ENA/GenBank and the SRA under BioProject [PRJNA763310](#). Isolates have been deposited at Manaaki Whenua-Landcare Research (New Zealand). Accession numbers are shown in Table 1.

## ACKNOWLEDGMENT

Funding for the research was provided by the Marsden Fund (project 19-UOO-245).

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