**ORIGINAL ARTICLE** 



# Genome-wide association study identifies new loci for albuminuria in the Japanese population

Hiroshi Okuda<sup>1,2,3</sup> • Koji Okamoto<sup>2,3</sup> • Michiaki Abe<sup>1,2,3</sup> • Kota Ishizawa<sup>1,2</sup> • Satoshi Makino<sup>2</sup> • Osamu Tanabe<sup>2,4</sup> • Junichi Sugawara<sup>2</sup> • Atsushi Hozawa<sup>2</sup> • Kozo Tanno<sup>5</sup> • Makoto Sasaki<sup>5</sup> • Gen Tamiya<sup>2,6</sup> • Masayuki Yamamoto<sup>2</sup> • Sadayoshi Ito<sup>2,3</sup> • Tadashi Ishii<sup>1,2</sup>

Received: 19 July 2019 / Accepted: 25 March 2020 © The Author(s) 2021, corrected publication 2021

#### Abstract

**Background** Urinary albumin excretion (UAE) is a risk factor for cardiovascular diseases, metabolic syndrome, chronic kidney disease, etc. Only a few genome-wide association studies (GWAS) for UAE have been conducted in the European population, but not in the Asian population. Here we conducted GWAS and identified several candidate genes harboring single nucleotide polymorphisms (SNPs) responsible for UAE in the Japanese population.

**Methods** We conducted GWAS for UAE in 7805 individuals of Asian ancestry from health-survey data collected by Tohoku Medical Megabank Organization (ToMMo) and Iwate Tohoku Medical Megabank Organization (IMM). The SNP genotype data were obtained with a SNP microarray. After imputation using a haplotype panel consisting of 2000 genome sequencing, 4,962,728 SNP markers were used for the GWAS.

**Results** Eighteen SNPs at 14 loci (*GRM7*, *EXOC1/NMU*, *LPA*, *STEAP1B/RAPGEF5*, *SEMA3D*, *PRKAG2*, *TRIQK*, *SERTM1*, *TPT1-AS1*, *OR5AU1*, *TSHR*, *FMN1/RYR3*, *COPRS*, and *BRD1*) were associated with UAE in the Japanese individuals. A locus with particularly strong associations was observed on *TSHR*, chromosome 14 [rs116622332 (p = 3.99 × 10<sup>-10</sup>)]. **Conclusion** In this study, we successfully identified UAE-associated variant loci in the Japanese population. Further study is required to confirm this association.

Keywords QTL · GWAS · Albuminuria · Genetics · TSHR · Cohort study

# Introduction

Chronic kidney disease (CKD) is one of the most severe global public health problems [1]. The proportion of patients with end-stage renal disease is growing, and thus, the resultant cost poses a big problem in health economics [2]. It is important to diagnose renal failure in the early stages to

Koji Okamoto okamoto5-tky@umin.ac.jp

- <sup>1</sup> Department of Education and Support for Regional Medicine, Tohoku University Hospital, 1-1 Seiryo-machi, Aoba-ku, Sendai, Miyagi 980-8574, Japan
- <sup>2</sup> Tohoku Medical Megabank Organization, Tohoku University, 2-1 Seiryo-machi, Aoba-ku, Sendai, Miyagi 980-8573, Japan
- <sup>3</sup> Department of Nephrology, Endocrinology and Vascular Medicine, Graduate School of Medicine, Tohoku University, 1-1 Seiryo-machi, Aoba-ku, Sendai, Miyagi 980-8574, Japan

prevent disease progression. However, it is very difficult to do so as the typical symptoms of renal failure rarely emerge in the earlier stages. The risks of mortality, myocardial infarction, and progression to kidney failure associated with a particular value of estimated glomerular filtration rate (eGFR) are increased independently in patients with moderate to severe urinary albumin excretion (UAE) [3]. Apart from CKD, UAE is known biomarker of cardiovascular diseases, diabetes mellitus, obesity, hypertension, and all-cause

- <sup>4</sup> Radiation Effects Research Foundation, 5-2 Hijiyama Park, Minami-ku, Hiroshima, Hiroshima 732-0815, Japan
- <sup>5</sup> Iwate Tohoku Medical Megabank Organization, Iwate Medical University, 1-1-1 Idaidori, Yahaba-cho, Shiwa-gun, Iwate 028-3694, Japan
- <sup>6</sup> RIKEN Center for Advanced Intelligence Project Nihonbashi, 1-chome Mitsui Bldg. 15F, 1-4-1 Nihonbashi, Chuo-ku, Tokyo 103-0027, Japan

mortality [4–8]. Even albuminuria of less than 30 mg/gCr (lower than microalbuminuria) is known as a marker of these diseases [7, 8].

While a few genome-wide association studies (GWAS) on UAE have been conducted in individuals with type 2 diabetes mellitus [9] and type 1 diabetes mellitus [10] in European ancestral cohorts [11–13], there is no such GWAS conducted in the Asian population. Here we have conducted GWAS using health-survey data collected in the Tohoku Medical Megabank Organization (ToMMo) and Iwate Tohoku Medical Megabank Organization (IMM) to identify the several candidate genes harboring single nucleotide polymorphisms (SNPs) responsible for UAE in the Japanese population.

### **Material and methods**

#### **Study subjects**

This research was conducted as a part of the residential cohort study of Tohoku Medical Megabank (TMM), a joint organization of the Tohoku University and the Iwate Medical University in Japan, established in 2011 after the Great East Japan Earthquake for creating an advanced medical system.

Over 80,000 almost healthy adult individuals living in the Miyagi and Iwate Prefectures along the Pacific coast of the Tohoku district of northern Japan were recruited from May 2013 to March 2016 for the TMM Project. The participants were of 20-75 years old and completed questionnaires covering a wide range of topics including socio-demographic factors, lifestyle habits, and medical history. Blood and urine tests were performed at baseline survey. The participants living in the Miyagi Prefecture and Iwate Prefecture were recruited by Tohoku University and Iwate Medical University, respectively [14, 15]. We obtained approval from the relevant ethics committees of both the facilities. We obtained written informed consent from each participant when they were enrolled in the TMM cohort study. This study was conducted according to the principals of the Declaration of Helsinki.

From the 10,000 individuals whose data were collected up to 2013, we were able to obtain data of 9,966 individuals after excluding 34 people who withdrew their consent after collection. The data were released as dbToMMo 1.1. Among these 9,966 individuals, 4974 were from the Miyagi prefecture and 4992 were from the Iwate prefecture. Thus, both represented a roughly equal proportion.

#### Sample quality control

Genotyping was performed for 964,193 SNP markers using Illumina's Human Omni Express Exome- 8 version 1.2

BeadChips. Upon conducting quality control of the samples based on the genotyping data, some people were excluded owing to data loss (n=1), genotype defect (low call rate: call rate <0.98, n=5), or close relationship pairs (identity-by-descent estimates, PI\_HAT>3/32, n=2155) [16, 17]. Finally, the data sampled from 7805 individuals passed quality control.

#### Marker quality control

As quality control of the genotyped marker, SNPs with low call rates (<0.95), and low *p* values in the Hardy Weinberg equilibrium (HWE) test (*p* value <  $1.0 \times 10^{-4}$ ), low minor allele frequencies (MAF < 0.01), and low-quality markers among the duplication markers were filtered out. As a result, 595,171 SNPs remained for the downstream analysis.

#### **Genotype imputation**

Genotype imputation was performed using SHAPEIT v2.r837 [18] and IMPUTE2 v2.2.2 [19] software packages with TMM 2KJPN high-quality haplotype reference panel based on the 2049 drafts of the whole genome sequencing and was implemented in the TMM [20]. After genotype imputation, we adjusted the imputation quality (INFO scores) and MAF. The variants with low imputation quality (INFO scores < 0.5) and low minor allele frequency (MAF < 0.03) variants were excluded. Ultimately, 4,962,728 variants were retained for the GWAS.

#### Phenotype

Phenotype information was obtained from the questionnaires covering age, sex, physical measurement including body mass index (BMI), and systolic blood pressure (SBP). For standardizing the blood pressure estimation, we did not use the antihypertensive medication history because systolic blood pressure strongly influences the glomerular pressure and UAE [21].

Urinary Na (UNa), urinary K (UK), urinary creatinine (UCr), urinary albumin (Ualb), serum creatinine (sCre), serum cystatin C (sCysC), hemoglobin A1c (HbA1c), and eGFR were estimated at baseline. We selected eGFR calculated by serum cystatin C (eGFRcys) for the evaluation of renal functional instead of eGFR calculated by serum creatinine (eGFRcre) because serum cystatin C was a better marker of early-stage CKD than serum creatinine [22]. eGFRcys was calculated by the Japanese equation for eGFR from serum cystatin C as follows [23, 24];

eGFRcys mL/min/1.73 m<sup>2</sup> =  $104 \times SCysC^{-1.019}$ × 0.996<sup>age</sup> × 0.929(*if female*) - 8

#### **Statistical analyses**

After performing a standard linear regression analysis of UAE (PLINK version 1.9 software package) for each SNP, we performed GWAS for UAE. We used UAE corrected by creatinine (continuous variable) as a response variable. The analysis was adjusted for the relevant covariates including age, sex, BMI [25], SBP [26], UNa [27], UK [27], HbA1c [26], eGFRcys [28], and the top significant 26 principal components of the genotypes. These are reported confounding factors for albuminuria.

We constructed a Manhattan plot and Quantile–quantile plot (Q-Q plot) to visually evaluate the analysis result. We used the statistical software R with "qqman" package. We constructed Regional plot to evaluate the linkage disequilibrium (LD) structure around the SNPs with Locus Zoom [URL; https://locuszoom.org]. We evaluated the result of the expression Quantitative Trait Loci (eQTL) analysis for some genome-wide significant SNPs by the Genotype Tissue Expression Project (GTEx) portal [29].

#### Results

#### Basic characteristics of the study subjects

The genotype data from 7805 individuals passed quality control and were used for the analysis. The detailed characteristics of the analyzed data are shown in Table 1. There were 60.7% patients of CKD stage 1, 36.6% patients of CDK stage 2, and 2.71% patients of CKD stage  $\geq$  3. In this setting, cystatin C seemed to be more appropriate for kidney function marker. The mean age of the patients was  $61.8 \pm 11.2$  years, and 34.8% of the patients were male. The average systolic blood pressure of these patients was  $127 \pm 17.8$  mm Hg and the median of UAE was 7.4 mg/ gCr [interquartile range (IQR) 8.8]. About one-fourth (24.5%) of the patients were hypertensive [defined as systolic blood pressure > 140 mmHg or diastolic blood pressure > 90 mmHg in accordance with The Japanese Society of Hypertension Guidelines for the Management of Hypertension (JSH 2014)] [30]. In this study, the individuals taking antihypertensive medications were also diagnosed as hypertensive.

About 10% of patients had UAE of > 30 mg/gCr, and hence most of the patients had microalbuminuria. The mean age of the UAE positive patients was  $64.4 \pm 8.64$  years and 43.6% of these patients were males. The mean systolic blood pressure of these patients was  $137 \pm 19.9$  mmHg and their average eGFRcys was  $89.5 \pm 23.7$  ml/min/1.73 m<sup>2</sup>. The mean age of the UAE negative patients was  $60.4 \pm 11.5$  years and 33.7% of these patients were male. The mean systolic

Table 1	Demographic	characteristics of	of the study	population

Characteristics	Total $n = 7805$	Ualb $-$ n = 6970	Ualb + $n = 827$
Age, years	$60.8 \pm 11.2$	$60.4 \pm 11.5$	64.8+8.64
Sex. male (%)	2716 (34.8)	2349 (33.7)	360 (43.5)
BMI	23.5 + 3.6	23.4 + 3.52	24.6+3.76
SBP. mmHg	127 + 17.8	126 + 17.2	137 + 19.9
DBP, mmHg	$-75.4 \pm 10.8$	$74.9 \pm 10.6$	$79.7 \pm 11.9$
HTN_treat (%)	211 (2.7)	275 (2.53)	31 (3.79)
HTN_diag (%)	1912 (24.5)	1540 (22.1)	369 (44.6)
Ualb/UCr, mg/gCr (IQR)	7.4 (8.8)*	6.7 (6.2)*	64.7 (99)*
< 30 mg/gCr (%)	6970 (89.3)	. ,	
$\geq$ 30 mg/gCr (%)	827 (10.6)		
UNa, g/l	$3.02 \pm 1.34$	$3.05 \pm 1.28$	$2.91 \pm 1.23$
UK, g/l	$1.63 \pm 1.09$	$1.66 \pm 1.04$	$1.50 \pm 0.91$
HbA1c(NGSP), %	$5.56 \pm 0.59$	$5.52 \pm 0.54$	$5.81 \pm 0.85$
sCre, mg/dl	$0.69 \pm 0.24$	$0.68 \pm 0.15$	$0.75 \pm 0.43$
sCysC, mg/l	$0.77 \pm 0.19$	$0.76 \pm 0.15$	$0.85 \pm 0.15$
eGFRcre, ml/min/1.73 m <sup>2</sup>	78.1±15.6	$78.4 \pm 0.15$	$74.8 \pm 0.15$
eGFRcys, ml/min/1.73 m <sup>2</sup>	$97.4 \pm 21.9$	$98.4 \pm 0.15$	$89.5 \pm 0.15$
CKD stage 1 (%)	4737 (60.7)	4346 (62.4)	390 (47.2)
CKD stage 2 (%)	2857 (36.6)	2499 (35.9)	351 (42.4)
CKD stage 3a (%)	174 (2.23)	112 (1.60)	63 (7.58)
CKD stage 3b (%)	27 (0.35)	12 (0.17)	15 (1.83)
CKD stage 4 (%)	8 (0.10)	1 (0.01)	7 (0.86)
CKD stage 5 (%)	2 (0.03)	1 (0.01)	1 (0.12)

SBP systolic blood pressure, DBP diastolic blood pressure, HTN\_ treat the person treated as hypertension from questionnaire, HTN\_ diag the persons diagnosed based on The Japanese Society of Hypertension Guidelines for the Management of Hypertension (JSH 2014), Ualb/UCr urinary albumin excretion corrected by urinary creatinine, UNa urinary sodium, UK urinary potassium, HbA1c(NGSP) hemoglobin A1c valued as National Glycohemoglobin Standardization Program, sCre serum creatinine, sCysC serum cystatin C, eGFRcre estimated glomerular filtration rate calculated by serum creatinine, eGFRcvs estimated glomerular filtration rate calculated by serum cystatin C, CKD stage 1 eGFRcys  $\geq$  90, CKD stage 2  $\leq$  60 eGFRcys<90, CKD stage 3a 45≤eGFRcys<60, CKD stage 3b 30≤eGFRcys<45, CKD stage 4 15≤eGFRcys<30, CKD stage 5 eGFRcys < 15, Ualb - the group without microalbuminuria nor overt albuminuria, Ualb + the group of microalbuminuria or overt albuminuria, IQR interquartile range

\*Median value

blood pressure of these patients was  $126 \pm 17.2$  mmHg and their average eGFRcys was  $98.4 \pm 21.3$  ml/min/1.73 m<sup>2</sup>.

When we evaluate correlation between each covariant and urinary albumin excretion, there is no significant correlation (Table S1). When we evaluate correlation between each covariant and other covariant, there are weak correlations between age and eGFRcys (correlation factor 0.56), SBP and BMI (correlation factor 0.48), UNa and UK (correlation factor 0.44) (Table S2).

# Genome-wide association study for UAE in the Japanese populations



**Fig. 1** Q-Q plots of GWAS about UAE in the TMM cohort study. The negative logarithm of the observed (*y*-axis) and the expected (*x*-axis) *p*-value was plotted for each SNP (dot), and the red line (y=x) indicates the null hypothesis of no true association. The regression genomic inflation factor ( $\lambda$  score) is 0.987 (SE 6.07×10<sup>-6</sup>) to adequately control the population stratification

 Table 2
 UAE associated SNPs reaching a genome-wide significance

We performed GWAS in 7805 individuals. After genotyping, 595,171 SNPs passed the quality controls and were used for the following imputation analysis. For the imputation analysis, a haplotype reference panel based on the 2049 drafts of the whole genome sequencing was used. Finally, 4,962,728 variants were used for the GWAS. The Q-Q plot is shown in Fig. 1. The genomic inflation factor ( $\lambda$ ) showed 0.987 suggesting that the population substructure should not have any substantial effects on the association analysis [14]. Under these conditions, we obtained 18 genome-wide significant SNPs (Table 2). We constructed a Manhattan plot of this GWAS as shown in Fig. 2.

With respect to the SNPs meeting the significance level, we constructed a regional plot to visually examine LD with the surrounding SNPs (around 1 Mbp) and identified the gene in or around which the SNPs were located. We attached the result of eQTL analyses for genome-wide significant SNPs by GTEx (Fig. 3, Fig S1). Three SNPs were located in *TSHR* on chromosome 14q31 [rs116622332 ( $p = 3.99 \times 10^{-9}$ ), rs199612558 ( $p = 1.00 \times 10^{-9}$ ), and rs17111387 ( $p = 3.42 \times 10^{-8}$ )] (Fig. 3). Two SNPs were located in *GRM7* on chromosome 3p26.1 [rs143146694 ( $p = 2.69 \times 10^{-11}$ , and rs74971332 ( $p = 8.91 \times 10^{-10}$ ) (Fig S1(a)). One SNP was located in *LPA* on chromosome 6q25.3 [rs146871152 ( $p = 7.16 \times 10^{-11}$ )] (Fig S1(b)). One SNP was located in *PRKAG2* on chromosome 7q36.1 [rs118160950 ( $p = 3.43 \times 10^{-8}$ )] (Fig S1(c)). Two SNPs were located

Chr	SNP	Position	Gene(s)	ref	alt	EA	EAF	BETA	SE	INFO	p value	AR <sup>2</sup>
3	rs143146694	6,263,450	GRM7	G	Т	Т	0.035	37.81	5.667	0.931	$2.69 \times 10^{-11}$	0.034
3	rs74971332	7,058,507	GRM7	С	G	G	0.055	26.43	4.307	0.97	$8.91 \times 10^{-10}$	0.034
4	rs75938525	56,659,946	EXOC1/NMU	G	С	С	0.048	31.06	5.029	0.956	$6.93 \times 10^{-10}$	0.034
6	rs146871152	160,984,637	LPA	С	Т	Т	0.034	37.28	5.713	0.97	$7.16 \times 10^{-11}$	0.037
7	rs146418897	22,440,870	STEAP1B/RAPGEF5	Т	С	С	0.033	36.85	5.743	0.941	$1.49 \times 10^{-10}$	0.034
7	rs140221313	84,600,098	SEMA3D	G	А	А	0.038	31.17	5.534	0.934	$1.84 \times 10^{-8}$	0.034
7	rs118160950	151,277,450	PRKAG2	С	Т	Т	0.036	30.48	5.517	0.975	$3.43 \times 10^{-8}$	0.036
8	rs141491217	94,068,096	TRIQK	А	G	G	0.039	33.01	5.84	0.932	$1.63 \times 10^{-8}$	0.035
13	rs79163227	37,208,221	SERTM1	А	G	G	0.056	25.92	4.664	0.941	$2.84 \times 10^{-8}$	0.035
13	rs142317900	45,963,584	TPT1-AS1	G	А	А	0.033	31.13	5.693	0.98	$4.68 \times 10^{-8}$	0.037
13	rs151183316	46,021,543	TPT1-AS1	С	Т	Т	0.035	30.6	5.54	0.981	$3.43 \times 10^{-8}$	0.035
14	chr14:21617499_TCTCA_T	21,617,499	OR5AU1	TCTCA	Т	Т	0.052	28.48	5.126	0.871	$2.87 \times 10^{-8}$	0.034
14	rs116622332	81,506,821	TSHR	Т	С	С	0.046	30.67	4.897	0.98	$3.99 \times 10^{-10}$	0.036
14	rs199612558	81,508,922	TSHR	Т	TA	TA	0.046	29.57	4.835	0.983	$1.00 \times 10^{-9}$	0.037
14	rs17111387	81,515,680	TSHR	С	Т	Т	0.068	22.24	4.025	0.996	$3.42 \times 10^{-8}$	0.037
15	rs140272046	33,494,078	FMN1/RYR3	G	А	А	0.042	33.38	5.886	0.886	$1.47 \times 10^{-8}$	0.034
17	rs148283070	30,129,004	COPRS	А	G	G	0.034	34.15	5.922	0.972	$8.42 \times 10^{-9}$	0.036
22	chr22:49949123_GA_G	49,949,123	BRD1	GA	G	G	0.052	27.83	4.805	0.945	$7.22 \times 10^{-9}$	0.034

*Chr* Chromosome, *SNP* single-nucleotide polymorphism, *Position* Chromosome position (GRCh37/hg19), *Gene* The name of Gene where the SNP is located, *ref* reference allele, *alt* alternative allele, *EA* effective allele, *EAF* effective allele frequency, *BETA* regression coefficient, *SE* Standard error of regression coefficient, *INFO* INFO score,  $AR^2$  Adjusted coefficient of determination. Bold type signifies that the SNPs were located on the gene. Normal type signifies that the SNPs were around the gene

**Fig. 2** Manhattan plot of the GWAS for UAE in the TMM cohort study. The *X*-axis represents the chromosomal positions and the *Y*-axis represents the  $-\log 10 p$ -values. The red horizontal line indicates the genome-wide significance threshold of  $p = 5 \times 10^{-8}$  and the blue horizontal line indicates the genome-wide suggestive threshold of  $p = 5 \times 10^{-5}$ . The name of the genes where the SNPs were located is typed in Manhattan plot



in TPT1-AS1 on chromosome 13q14.13 [rs142317900  $(p=4.68\times10^{-8})$  and rs151183316  $(p=3.43\times10^{-8})$ ] (Fig S1(d)). One SNP was located in EXOC1/NMU on chromosome 4q12 [rs75938525 ( $p = 6.93 \times 10^{-10}$ )] (Fig S1(e)). One SNP was located in STEAP1B/RAPGEF5 on chromosome 7p15.3 [rs146418897 ( $p = 1.49 \times 10^{-10}$ )] (Fig S1(f)). One SNP was located in SEMA3D on chromosome 7q21.11 [rs140221313 ( $p = 1.84 \times 10^{-8}$ )] (Fig S1(g)). One SNP was located in TRIQK on chromosome 8q22.1  $[rs141491217 (p=1.63 \times 10^{-8})]$  (Fig S1(h)). One SNP was located in SERTM1 on chromosome 13q13.3 [rs79163227  $(p=2.84\times10^{-8})$ ] (Fig S1(i)). One SNP was located in OR5AU1 on chromosome 14q11.2 [chr14:21617499\_ TCTCA\_T  $(p = 2.87 \times 10^{-8})$ ] (Fig. 3j). One SNP was located in FMN1/RYR3 on chromosome 15q13.3  $[rs140272046 (p=1.47 \times 10^{-8})]$  (Fig S1(k)). One SNP was located in COPRS on chromosome 17q11.2 [rs148283070  $(p = 8.42 \times 10^{-9})$ ] (Fig S1(1)). One SNP was located in BRD1 on chromosome 22q13.33 [chr22:49949123\_GA\_G  $(p = 7.22 \times 10^{-9})$ ] (Fig S3(m)).

# Discussion

We performed GWAS for UAE in the Japanese general population and identified 18 SNPs, of which, 17 were not reported in any previous report. rs118160950 was already reported as a SNP related to UAE by GWAS performed in the European ancestry [31]. In the previous reports of GWAS for UAE, the study subjects were not a general cohort but consisted of diabetes patients, heart failure patients, or pregnant women with hypertension. In addition, the study subjects were mainly European or African American but not Asians. Our GWAS has profound significance among the Japanese general population.

In previous studies, rs10795433 [9] and rs1801239 [13] were reported as the significant SNPs associated with UAE. They are located on the *CUBN* gene locus. *CUBN* encodes

cubilin protein acting as a receptor for vitamin B12-intrinsic factor complexes. It was hypothesized that the SNPs found in *CUBN* on chromosome 10 were significantly  $(p=1.0 \times 10^{-11})$  involved in UAE. However, these loci were not found to be significantly associated in our study. The reasons seem to be related to the difference in the studied population, because the reported significant covariates were not replicated in our population. We showed evidence of genetic differences between the Europeans and the East Asians. The other possibility is that these SNPs on *CUBN* are associated with UAE only in the diseased condition. There may be big differences in the mechanism between pathophysiological albuminuria and physiological albuminuria.

When we evaluated the functional class of 18 SNPs, 6 SNPs (rs74971332, rs146871152, rs118160950, rs116622332, rs199612558, and rs17111387) were intronic, 11 SNPs (rs143146694, rs75938525, rs146418897, rs140221313, rs141491217, rs79163227, rs151183316, chr14:21617499\_TCTCA\_T, rs140272046, rs148283070, and chr22:49949123\_GA\_G) were intergenic, and 1 SNP (rs142317900) was on the non-coding exons. No SNP was located on the coding exons. These candidates may possibly affect the factors regulating the transcription of the genes encoding the proteins involved in UAE.

In our study, the SNPs located in *TSHR* showed strong peaks. *TSHR* encodes TSHR (thyroid stimulating hormone receptor), and the TSH receptor is a member of the G protein-coupled receptor superfamily of integral membrane proteins which is coupled to the Gs protein [32, 33]. TSHR expresses mainly on the surface of the thyroid follicular cells and contributes to thyroid hormone secretion. Several pathways are proposed for kidney injury and proteinuria mediated by thyroid dysfunction [34, 35]. In hyperthyroid-ism, intra-glomerular hypertension, consequent hyperfiltration, increased production of free radicals, and increased renin–angiotensin–aldosterone system are risk factors for albuminuria. In hypothyroidism, GFR and tubular transport capacity are reduced. Hypothyroidism also results in



**Fig. 3** Association signals around the significant loci in *TSHR* locus. The upper panel is association signals around significant loci. The *X*-axis represents chromosomal positions (GRC37/hg19) and the *Y*-axis represents  $-\log_10 p$  -values. The lead variant is shown in purple. Colors represent the degree of LD ( $r^2$ ) between each variant and the lead variant. The LD ( $r^2$ ) was calculated based on the combined dataset of TMM subjects. The lower panels represent the Single-tis-

increased glomerular capillary permeability to proteins directly causing proteinuria [36]. The other possibility is that TSHR could have direct effects on albumin re-uptake on the tubular cells of the kidney. TSHR is also expressed in other tissue, for e.g. adipose tissues and fibroblasts. It is known that a small amount of TSHR exists in the kidneys sue eQTL analyses, where the target was mostly expressed. The data were from GTEx (V8). The X-axis is represents chromosomal positions (GRC38/hg38) and the Y-axis represents  $-\log 10$  eQTL p -values. The X-axis between the upper and the lower panel is adjusted by calculating with hgLiftOver [https://genome.ucsc.edu/cgi-bin/hgLiftOver].

and mainly in the tubules [37–39]. We evaluated the expression of TSHR by immunohistochemistry in the renal biopsy specimens. We found a weak expression of TSHR in the kidney mainly in the tubules. A significant association between the staining level of TSHR and proteinuria was not detected (data not shown) and further study should be required to prove this concept.

PRKAG2 coding 5'-AMP-activated protein kinase subunit gamma-2. AMP-activated protein kinase (AMPK) is a heterotrimeric protein composed of a catalytic alpha subunit, a noncatalytic beta subunit, and a noncatalytic regulatory gamma subunit [40]. AMPK is an important energysensing enzyme that monitors the cellular energy status and functions by inactivating the key enzymes involved in regulating the de novo biosynthesis of fatty acids and cholesterol. Mutations in this gene have been associated with WPW (Wolff-Parkinson-White) syndrome [41, 42], familial hypertrophic cardiomyopathy [43, 44], and enlarged kidneys [45]. Studies in transgenic mice indicate that these mutations cause glycogen storage disease of the heart [46]. Several other hereditary glycogen storage diseases present with renal pathologies, such as renal tubular dysfunction [47]. *PRKAG2* did not indicate any stronger significance in our GWAS. However, we may consider that renal tubular dysfunction induced by glycol storage can affect the UAE.

We used the GTEx database to examine eQTL of signiicant variants and suggestive variants in each locus. Unfortunately, no eQTL was found in the lead significant variants (Table S3). Then we extended the candidates for including suggestive variants whose p value was less than  $1.0 \times 10^{-5}$ . and also that has strong LD against each significant SNP  $(r^2 > 0.2)$ . There are significant eQTL of NMNAT1P1 pseudo-gene on TSHR gene locus in rs17111387, rs74771569, rs78176261, which three SNPs had strong LD against rs116622332 (the lead variant in Chromosome 14). Though NMNAT1P1 itself is a pseudo-gene, many significant eQTL variants are also shared with TSHR and NMNAT1P1. Additionally, about the genes around every lead variant, we can find the consistency between the peak of Manhattan plot and the peak of eQTL information about TSHR in thyroid (Fig. 3). That means there is a probability that rs116622332 allele on chromosome 14, affects TSHR and NMNAT1P1 expression in thyroid. eQTL analysis identified that rs77317344, which is in strong LD with rs14237900 (the lead variant in Chromosome 13), affects the expression of COG3. We cannot find other significant information in eQTL analysis about the other genes.

There are some limitations to our study. First, in this study, replication is lacking. Replication studies in other Japanese cohorts and/or other populations are required. Second, many of the individuals analyzed in our study were affected by the Great East Japan Earthquake of 2011. We should consider mental disturbance and stress caused by this big disaster as a confounding factor. To conclude, we investigated the UAE associated SNPs in the Japanese population after adjusting for age, gender, hypertension, and impaired glucose tolerance. The 18 identified SNPs were uncovered to show a statistically significant effect on the UAE. There

are limited studies evaluating the association with other candidate genes that we detected. The functional and biological roles exerted by each of the SNPs/genes are required to be elucidated in further studies.

**Electronic supplementary material** The online version of this article (https://doi.org/10.1007/s10157-020-01884-x) contains supplementary material, which is available to authorized users.

Acknowledgements This study was supported by the TMM Project (Special Account for Reconstruction from the Great East Japan Earthquake) from the Ministry of Education, Culture, Sports, Science and Technology (MEXT) and the Japan Agency for Medical Research and Development (AMED). The authors sincerely express thanks to all the patients and volunteers who were enrolled in the ToMMo and IMM in the disaster-struck areas affected by the Great East Japan Earthquake. We thank the members of ToMMo and IMM, including GMRCs, office and administrative personnel, and software engineers, for their assistance in the projects. The full list of the members is available at https:// www.megabank.tohoku.ac.jp/english/a190601/ for ToMMo and https://

**Data availability** The datasets analyzed in this study are not open to the public for ethical reasons but are available upon request after approval of the Ethical Committee of Tohoku University, the Ethical Committee of Iwate Medical University, and the Materials and Information Distribution Review Committee of the TMM Project.

#### Compliance with ethical standards

**Conflict of interest** The author(s) received no specific funding for this work.

Human and animal rights The study was conducted in accordance with the guidelines written in the Declaration of Helsinki. We obtained approval from the relevant ethics committees at Tohoku Medical. Megabank Organization. The assignment number is 2018-4-034.

**Informed consent** Informed consent was obtained from all individual participants included in the study.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

#### References

 Collaborators GMaCoD. Global, regional, and national life expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980–2015: a systematic analysis for the Global Burden of Disease Study 2015. Lancet. 2016;388(10053):1459–544. doi: 10.1016/S0140-6736(16)31012-1.

- Ozieh MN, Bishu KG, Dismuke CE, Egede LE. Trends in healthcare expenditure in United States adults with chronic kidney disease: 2002–2011. BMC Health Serv Res. 2017;17(1):368. https:// doi.org/10.1186/s12913-017-2303-3 (Epub 2017/05/22).
- Hemmelgarn BR, Manns BJ, Lloyd A, James MT, Klarenbach S, Quinn RR, et al. Relation between kidney function, proteinuria, and adverse outcomes. JAMA. 2010;303(5):423–9. https://doi.org/ 10.1001/jama.2010.39.
- Gerstein HC, Mann JF, Yi Q, Zinman B, Dinneen SF, Hoogwerf B, et al. Albuminuria and risk of cardiovascular events, death, and heart failure in diabetic and nondiabetic individuals. JAMA. 2001;286(4):421–6.
- Brantsma AH, Bakker SJ, Hillege HL, de Zeeuw D, de Jong PE, Gansevoort RT, et al. Urinary albumin excretion and its relation with C-reactive protein and the metabolic syndrome in the prediction of type 2 diabetes. Diabetes Care. 2005;28(10):2525–30.
- Matsushita K, Coresh J, Sang Y, Chalmers J, Fox C, Guallar E, et al. Estimated glomerular filtration rate and albuminuria for prediction of cardiovascular outcomes: a collaborative meta-analysis of individual participant data. Lancet Diabetes Endocrinol. 2015;3(7):514–25. https://doi.org/10.1016/S2213-8587(15) 00040-6 (Epub 2015/05/28).
- Hillege HL, Fidler V, Diercks GF, van Gilst WH, de Zeeuw D, van Veldhuisen DJ, et al. Urinary albumin excretion predicts cardiovascular and noncardiovascular mortality in general population. Circulation. 2002;106(14):1777–822.
- Klausen K, Borch-Johnsen K, Feldt-Rasmussen B, Jensen G, Clausen P, Scharling H, et al. Very low levels of microalbuminuria are associated with increased risk of coronary heart disease and death independently of renal function, hypertension, and diabetes. Circulation. 2004;110(1):32–5. https://doi.org/10.1161/01.CIR. 0000133312.96477.48 (Epub 2004/06/21).
- Teumer A, Tin A, Sorice R, Gorski M, Yeo NC, Chu AY, et al. Genome-wide association studies identify genetic loci associated with albuminuria in diabetes. Diabetes. 2016;65(3):803–17. https://doi.org/10.2337/db15-1313 (Epub 2015/12/02).
- Sandholm N, Forsblom C, Mäkinen VP, McKnight AJ, Osterholm AM, He B, et al. Genome-wide association study of urinary albumin excretion rate in patients with type 1 diabetes. Diabetologia. 2014;57(6):1143–53. https://doi.org/10.1007/s00125-014-3202-3 (Epub 2014/03/05).
- Hwang SJ, Yang Q, Meigs JB, Pearce EN, Fox CS. A genomewide association for kidney function and endocrine-related traits in the NHLBI's Framingham Heart Study. BMC Med Genet. 2007;8(Suppl 1):S10. https://doi.org/10.1186/1471-2350-8-S1-S10 (Epub 2007/09/19).
- Ellis JW, Chen MH, Foster MC, Liu CT, Larson MG, de Boer I, et al. Validated SNPs for eGFR and their associations with albuminuria. Hum Mol Genet. 2012;21(14):3293–8. https://doi.org/ 10.1093/hmg/dds138 (Epub 2012/04/05).
- Böger CA, Chen MH, Tin A, Olden M, Köttgen A, de Boer IH, et al. CUBN is a gene locus for albuminuria. J Am Soc Nephrol. 2011;22(3):555–70. https://doi.org/10.1681/ASN.2010060598.
- Hachiya T, Komaki S, Hasegawa Y, Ohmomo H, Tanno K, Hozawa A, et al. Genome-wide meta-analysis in Japanese populations identifies novel variants at the TMC6-TMC8 and SIX3-SIX2 loci associated with HbA. Sci Rep. 2017;7(1):16147. https://doi. org/10.1038/s41598-017-16493-0 (Epub 2017/11/23).
- Kuriyama S, Yaegashi N, Nagami F, Arai T, Kawaguchi Y, Osumi N, et al. The Tohoku Medical Megabank Project: Design and Mission. J Epidemiol. 2016;26(9):493–511. https://doi.org/10.2188/jea.JE20150268 (Epub 2016/07/02).
- 16. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a tool set for whole-genome association

and population-based linkage analyses. Am J Hum Genet. 2007;81(3):559–75. https://doi.org/10.1086/519795 (Epub 2007/07/25).

- Tanaka F, Yamamoto K, Suzuki S, Inoue H, Tsurumaru M, Kajiyama Y, et al. Strong interaction between the effects of alcohol consumption and smoking on oesophageal squamous cell carcinoma among individuals with ADH1B and/or ALDH2 risk alleles. Gut. 2010;59(11):1457–64. https://doi.org/10.1136/gut.2009.205724 (Epub 2010/09/09).
- Delaneau O, Howie B, Cox AJ, Zagury JF, Marchini J. Haplotype estimation using sequencing reads. Am J Hum Genet. 2013;93(4):687–96. https://doi.org/10.1016/j.ajhg.2013.09.002.
- Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. PLoS Genet. 2009;5(6):e1000529. https://doi. org/10.1371/journal.pgen.1000529 (Epub 2009/06/19).
- Nagasaki M, Yasuda J, Katsuoka F, Nariai N, Kojima K, Kawai Y, et al. Rare variant discovery by deep whole-genome sequencing of 1,070 Japanese individuals. Nat Commun. 2015;6:8018. https:// doi.org/10.1038/ncomms9018 (Epub 2015/08/21).
- Oliveras A, Armario P, Martell-Clarós N, Ruilope LM, de la Sierra A, Registry SSoH-RH. Urinary albumin excretion is associated with nocturnal systolic blood pressure in resistant hypertensives. Hypertension. 2011;57(3):556–60. https://doi.org/10.1161/ HYPERTENSIONAHA.110.165563 (Epub 2011/01/10).
- Dharnidharka VR, Kwon C, Stevens G. Serum cystatin C is superior to serum creatinine as a marker of kidney function: a metaanalysis. Am J Kidney Dis. 2002;40(2):221–6. https://doi.org/10. 1053/ajkd.2002.34487.
- Horio M, Imai E, Yasuda Y, Watanabe T, Matsuo S, GFR CDtJEfE. GFR estimation using standardized serum cystatin C in Japan. Am J Kidney Dis. 2013;61(2):197–203. https://doi.org/ 10.1053/j.ajkd.2012.07.007 (Epub 2012/08/11).
- Matsuo S, Imai E, Horio M, Yasuda Y, Tomita K, Nitta K, et al. Revised equations for estimated GFR from serum creatinine in Japan. Am J Kidney Dis. 2009;53(6):982–92. https://doi.org/10. 1053/j.ajkd.2008.12.034 (Epub 2009/04/01).
- Liu X, Liu Y, Chen Y, Li Y, Shao X, Liang Y, et al. Body mass index (BMI) is associated with microalbuminuria in Chinese hypertensive patients. Int J Environ Res Public Health. 2015;12(2):1998–2008. https://doi.org/10.3390/ijerph120201998 (Epub 2015/02/10).
- Araki S, Haneda M, Sugimoto T, Isono M, Isshiki K, Kashiwagi A, et al. Factors associated with frequent remission of microalbuminuria in patients with type 2 diabetes. Diabetes. 2005;54(10):2983–7. https://doi.org/10.2337/diabetes.54.10.2983.
- 27. Aaron KJ, Campbell RC, Judd SE, Sanders PW, Muntner P. Association of dietary sodium and potassium intakes with albuminuria in normal-weight, overweight, and obese participants in the Reasons for Geographic and Racial Differences in Stroke (REGARDS) Study. Am J Clin Nutr. 2011;94(4):1071–8. https:// doi.org/10.3945/ajcn.111.013094 (Epub 2011/08/31).
- Bansal N, Zelnick LR, Alonso A, Benjamin EJ, de Boer IH, Deo R, et al. eGFR and albuminuria in relation to risk of incident atrial fibrillation: a meta-analysis of the jackson heart study, the Multi-Ethnic study of atherosclerosis, and the cardiovascular health study. Clin J Am Soc Nephrol. 2017;12(9):1386–98. https://doi. org/10.2215/CJN.01860217 (Epub 2017/08/10).
- Carithers LJ, Ardlie K, Barcus M, Branton PA, Britton A, Buia SA, et al. A novel approach to high-quality postmortem tissue procurement: The GTEx project. Biopreserv Biobank. 2015;13(5):311–9. https://doi.org/10.1089/bio.2015.0032.
- Shimamoto K, Ando K, Fujita T, Hasebe N, Higaki J, Horiuchi M, et al. The Japanese Society of Hypertension Guidelines for the Management of Hypertension (JSH 2014). Hypertens Res. 2014;37(4):253–390. https://doi.org/10.1038/hr.2014.20.

- Köttgen A, Pattaro C, Böger CA, Fuchsberger C, Olden M, Glazer NL, et al. New loci associated with kidney function and chronic kidney disease. Nat Genet. 2010;42(5):376–84. https://doi.org/10. 1038/ng.568 (Epub 2010/04/11).
- Farid NR, Szkudlinski MW. Minireview: structural and functional evolution of the thyrotropin receptor. Endocrinology. 2004;145(9):4048–57. https://doi.org/10.1210/en.2004-0437 (Epub 2004/07/01).
- Calebiro D, Nikolaev VO, Lohse MJ. Imaging of persistent cAMP signaling by internalized G protein-coupled receptors. J Mol Endocrinol. 2010;45(1):1–8. https://doi.org/10.1677/JME-10-0014 (Epub 2010/04/08).
- Iglesias P, Díez JJ. Thyroid dysfunction and kidney disease. Eur J Endocrinol. 2009;160(4):503–15. https://doi.org/10.1530/EJE-08-0837 (Epub 2008/12/18).
- Vargas F, Moreno JM, Rodríguez-Gómez I, Wangensteen R, Osuna A, Alvarez-Guerra M, et al. Vascular and renal function in experimental thyroid disorders. Eur J Endocrinol. 2006;154(2):197–21212. https://doi.org/10.1530/eje.1.02093.
- 36. Wheatley T, Edwards OM. Mild hypothyroidism and oedema: evidence for increased capillary permeability to protein. Clin Endocrinol (Oxf). 1983;18(6):627–35.
- Williams GR. Extrathyroidal expression of TSH receptor. Ann Endocrinol (Paris). 2011;72(2):68–73. https://doi.org/10.1016/j. ando.2011.03.006 (Epub 2011/04/20).
- Dutton CM, Joba W, Spitzweg C, Heufelder AE, Bahn RS. Thyrotropin receptor expression in adrenal, kidney, and thymus. Thyroid. 1997;7(6):879–84. https://doi.org/10.1089/thy.1997.7.879.
- Sellitti DF, Akamizu T, Doi SQ, Kim GH, Kariyil JT, Kopchik JJ, et al. Renal expression of two 'thyroid-specific' genes: thyrotropin receptor and thyroglobulin. Exp Nephrol. 2000;8(4–5):235–43. https://doi.org/10.1159/000020674.
- Cheung PC, Salt IP, Davies SP, Hardie DG, Carling D. Characterization of AMP-activated protein kinase gamma-subunit isoforms and their role in AMP binding. Biochem J. 2000;346(Pt 3):659–69.
- Vaughan CJ, Hom Y, Okin DA, McDermott DA, Lerman BB, Basson CT. Molecular genetic analysis of PRKAG2 in sporadic

Wolff-Parkinson-White syndrome. J Cardiovasc Electrophysiol. 2003;14(3):263–8.

- Gollob MH, Green MS, Tang AS, Gollob T, Karibe A, Ali Hassan AS, et al. Identification of a gene responsible for familial Wolff-Parkinson-White syndrome. N Engl J Med. 2001;344(24):1823– 31. https://doi.org/10.1056/NEJM200106143442403.
- 43. Blair E, Redwood C, Ashrafian H, Oliveira M, Broxholme J, Kerr B, et al. Mutations in the gamma(2) subunit of AMP-activated protein kinase cause familial hypertrophic cardiomyopathy: evidence for the central role of energy compromise in disease pathogenesis. Hum Mol Genet. 2001;10(11):1215–20.
- Gollob MH. Modulating phenotypic expression of the PRKAG2 cardiac syndrome. Circulation. 2008;117(2):134–5. https://doi. org/10.1161/CIRCULATIONAHA.107.747345.
- 45. Burwinkel B, Scott JW, Bührer C, van Landeghem FK, Cox GF, Wilson CJ, et al. Fatal congenital heart glycogenosis caused by a recurrent activating R531Q mutation in the gamma 2-subunit of AMP-activated protein kinase (PRKAG2), not by phosphorylase kinase deficiency. Am J Hum Genet. 2005;76(6):1034–49. https:// doi.org/10.1086/430840 (Epub 2005/05/02).
- Arad M, Benson DW, Perez-Atayde AR, McKenna WJ, Sparks EA, Kanter RJ, et al. Constitutively active AMP kinase mutations cause glycogen storage disease mimicking hypertrophic cardiomyopathy. J Clin Invest. 2002;109(3):357–62. https://doi.org/10. 1172/JCI14571.
- Ozen H. Glycogen storage diseases: new perspectives. World J Gastroenterol. 2007;13(18):2541–53.

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.