Perspective



Iron chelators or therapeutic modulators of iron overload: Are we anywhere near ideal one?

Iron overload is an important cause of mortality and morbidity in chronic transfusion-dependent states commonly exemplified by transfusion-dependent beta-thalassaemia syndromes [including thalassaemia non-transfusion-dependent (NTD) intermedia] as well as transfusion-dependent myelodysplastic syndromes (MDS). Hence, therapeutic iron chelation is indicated in these conditions¹⁻⁵. There seems to be an increasing interest for different iron chelators and labile iron pool for patients with thalassaemia syndromes (thalassaemia major and thalassaemia intermedia) and transfusion-dependent MDS with respect to their efficacy, tolerability, safety, cost and long-term side effects. Several studies have been published recently on this topic¹⁻⁹. Di Maggio and Maggio⁴ evaluated 10 trials on various iron chelators singly or in combination for iron overloaded transfusion-dependent beta-thalassaemia major. Longitudinal changes in liver iron concentrations using different chelation regimens were discussed well by Vitrano *et al*¹. There is a need for better chelators while search for optimum chelation regimen combining different chelators continues. Adding amlodipine to inhibit iron uptake by heart is another idea³ as heart failure is one of the key serious complications in ironloaded beta-thalassaemia major patients.

Di Maggio and Maggio⁴ reviewed 10 different combination of three iron chelators in the market singly or in combination of two either given simultaneously or alternately in different trials. However, no data were given for trials evaluating three-drug combination or a combination of two oral drugs (deferasirox and deferiprone) though the use of two oral drug combination alternately has been used for hard to chelate patients¹⁰.

Deferiprone as an iron chelator still does not have wide acceptance in the developed countries outside restricted indications in MDS and myocardial iron overload in combination with deferoxamine⁶. Iron deposition in multitransfused beta-thalassaemia patients produces clinical effects in mainly three organ systems, i.e. heart, liver and endocrine glands. Iron deposition-related mortality in transfusion-dependent beta-thalassaemia patients mostly occurs due to cardiac iron overload or iron overload in the liver⁴. In spite of all the intense chelation regimens, a significant number of transfusion-dependent beta-thalassaemia patients develop endocrine deficiency¹¹, showing that present chelation therapy is inadequate particularly with regard to recovery of endocrine gland function. The most effective, deferoxamine has to be given by long hours of subcutaneous injection¹⁰. The other two *i.e.* deferasirox and deferiprone have side effects such as agranulocytosis, hepatitis, renal disturbances, gastric intolerance, retinal and auditory toxicity and olfactory disturbances and these may not produce desired level of tissue iron in iron-overloaded state despite adequately intense therapy in some patients underscoring the need for more effective and safe iron chelators for clinical use^{4,6,10}.

Iron overload has multiple effects on thalassaemia patients. Oxidation damage to red cells due to labile non-transferrin bound iron (NTBI) damages the red cell membrane and shortens transfused red cells lifespan. This increases the transfusion requirement as well as iron overload¹². It has damaging effects on mitochondria¹³ that produce progressive mutation in mitochondrial genes in MDS14 and affect DNA stability in the cell¹⁵. Iron overload also directly suppresses haemopoiesis¹⁶. Although mitochondrial mutation in thalassaemia major has not been studied extensively¹⁷, considering the mechanism of action of NTBI and its ability to cause mutation, mitochondria are likely to be vulnerable to DNA damage in this disease. Hence, we need better iron chelators and should probably start early chelation in these patients.

^{© 2018} Indian Journal of Medical Research, published by Wolters Kluwer - Medknow for Director-General, Indian Council of Medical Research

Iron metabolism is controlled by a master hormone called hepcidin secreted from the liver when liver and tissues sense higher levels of iron in the body. Secretion of hepcidin is negatively regulated by matriptase 2 (also called TMPRSS6) which on activation through a chain of reactions leads to increased transcription and translation of hepcidin. Hepcidin inhibits absorption of iron from intestine and egress of iron from macrophage and other stores by catabolizing the iron transporter ferroportin¹⁸. Hence, matriptase 2 inhibitor or long-acting hepcidin mimetics can cause negative iron balance by preventing its absorption or its transport out of the cell rather than reducing iron overload by chelation. Second-generation TMPRSS6 antisense oligonucleotides were tested in animal model either singly or in combination with other available iron chelators such as deferiprone with good and synergistic reduction of iron overload and limited toxicity¹⁹. This product will now be taken up for phased clinical trials. Similarly, several chemical inhibitors of matriptase 2 have been synthesized and will undergo safety studies²⁰. Small molecule peptides called minihepcidins which are orally bioavailable, have also been synthesized and effectively tested on animals successfully²¹. Small molecules with isoflavone structure (*i.e.*, genistein) have been shown to stimulate hepcidin synthesis from liver²² and can possibly be used to reduce iron levels in the iron-loaded state. Both these classes of compounds tend to reduce hepatic iron overload in preference to iron in other organs and may be especially useful for NTD thalassaemia intermedia where hepatic iron overload is a major challenge. However, no human trial with these compounds has been conducted as yet.

None of the 10 trials described⁴ evaluated serial NTBI or labile free iron serially in their patients. This needs to be included in all future trials.

Hydroxyurea, a commonly used medicine for sickle cell disease and myeloproliferative disorders, was evaluated by us both in transfusion-dependent thalassaemia major and non-transfusion or minimally transfusion-dependent thalassaemia intermedia patients²³⁻²⁵. The reason for considering hydroxyurea as an iron chelator was due to its reported cytotoxicity because of inhibition of ribonucleotide reductase by iron chelation. Its chemical structure has a bidentate iron binding site²⁶.

Hydroxyurea synergistically works with both orally active iron chelators, has very good cardiac iron

removing potential and strongly suppresses NTBI iron levels. It is equally potent in iron removal as deferasirox at a dose level without causing myelosuppression at the dose ranges used. It also reduced iron overload by reducing transfusion requirement in 30 per cent of thalassaemia major patients²⁵. Whether hydroxyurea or some of its future analogues will form another class of iron chelators needs to be seen.

Iron chelation is increasingly being considered for many different conditions such as Plasmodium falciparum infection²⁷ or drug-resistant bacterial infections²⁸ or malignancies²⁹. Drug-resistant bacteria producing carbapenemase and metallobetalactamase (NDM1) can now be countered by an antibiotic where an iron (and other metal) chelating catechol nucleus has been coupled to cephalosporin moiety²⁸. Similarly, iron chelators are being explored to treat various malignancies²⁹. It has been revealed that chelation of iron from proper cellular compartments can alter actions of various signal transduction mechanisms involved in cell growth, mitosis, apoptosis, autophagy and metastasis by involving [phosphatidy] inositol 3-kinase/protein kinase B (PI3-K)/ AKT, c-Jun N-terminal kinase (JNK), extracellular signal-regulated kinase (ERK), p³⁸ mitogen-activated protein kinase (p38MAPK), signal transducer and activator of transcription 3 (STAT3), Wnt (Wingless/ Integrated)] transforming growth factor- β (TGF- β), etc³⁰. Hence, one iron chelator may not fit all the bills. Different iron chelators may remove iron from different compartment of cells or tissues with different efficiency and may have differential toxicity and utility for different diseases. It has been found that efficiency of iron chelation depends on lipid solubility of the drug, but highly lipid-soluble iron chelators also increase the toxicity of the drug as seen in case of moderately lipid-soluble SP-420, a desferrithiocin analogue which was withdrawn from the trial because of nephrotoxicity³¹. Iron chelator has also been used to reduce daunorubicin toxicity on the heart using a heart directed iron chelator dexrazoxane³². However, one of the interesting ways to reduce toxicity and develop efficiency of the iron chelator is by developing metabolically programmable iron chelators³³ where the compound is designed in such a way that after reaching the tissue and chelating iron, the compound is metabolically transformed into more polar compound hence quickly excreted from the body.

It is hoped that many more novel molecules or combination of molecules or repurposing of old molecules singly or in combination for removal of iron with novel mechanism of action or prevention of gastrointestinal absorption of iron will be developed in the future. These may improve iron chelation and overall improvement of health in multitransfused thalassaemia major and other patients in the future.

Financial support & sponsorship: None.

Conflicts of Interest: None.

Kanjaksha Ghosh¹ & Kinjalka Ghosh^{2,*} ¹Surat Raktadan Kendra & Research Centre, Surat 395 002, Gujarat & ²Department of Biochemistry, Tata Memorial Hospital, Mumbai 400 012, Maharashtra, India **For correspondence*: kanjakshaghosh@hotmail.com

Received December 19, 2017

References

- 1. Vitrano A, Sacco M, Rosso R, Quota A, Fiorino D, Oliva E, *et al.* Longitudinal changes in LIC and other parameters in patients receiving different chelation regimens: Data from LICNET. *Eur J Haematol* 2018; *100* : 124-30.
- Porter JB, Elalfy M, Taher A, Aydinok Y, Lee SH, Sutcharitchan P, *et al.* Limitations of serum ferritin to predict liver iron concentration responses to deferasirox therapy in patients with transfusion-dependent thalassaemia. *Eur J Haematol* 2017; 98 : 280-8.
- Eghbali A, Kazemi H, Taherahmadi H, Ghandi Y, Rafiei M, Bagheri B, *et al.* A randomized, controlled study evaluating effects of amlodipine addition to chelators to reduce iron loading in patients with thalassemia major. *Eur J Haematol* 2017; 99: 577-81.
- Di Maggio R, Maggio A. The new era of chelation treatments: Effectiveness and safety of 10 different regimens for controlling iron overloading in thalassaemia major. *Br J Haematol* 2017; 178: 676-88.
- Leitch HA, Parmar A, Wells RA, Chodirker L, Zhu N, Nevill TJ, et al. Overall survival in lower IPSS risk MDS by receipt of iron chelation therapy, adjusting for patient-related factors and measuring from time of first red blood cell transfusion dependence: An MDS-CAN analysis. Br J Haematol 2017; 179: 83-97.
- 6. Killick SB. Iron chelation therapy in low risk myelodysplastic syndrome. *Br J Haematol* 2017; *177* : 375-87.
- Angelucci E, Urru SA, Pilo F, Piperno A. Myelodysplastic syndromes and iron chelation therapy. *Mediterr J Hematol Infect Dis* 2017; 9: e2017021.
- Angelucci E, Cianciulli P, Finelli C, Mecucci C, Voso MT, Tura S, *et al.* Unraveling the mechanisms behind iron overload and ineffective hematopoiesis in myelodysplastic syndromes. *Leuk Res* 2017; *62*: 108-15.

- de Swart L, Reiniers C, Bagguley T, van Marrewijk C, Bowen D, Hellström-Lindberg E, *et al.* Labile plasma iron levels predict survival in patients with lower-risk myelodysplastic syndromes. *Haematologica* 2018; *103* : 69-79.
- Balocco M, Carrara P, Pinto V, Forni GL. Daily alternating deferasirox and deferiprone therapy for "hard-to-chelate" beta-thalassemia major patients. *Am J Hematol* 2010; *85*: 460-1.
- De Sanctis V, Soliman AT, Elsedfy H, Di Maio S, Canatan D, Soliman N, *et al.* Gonadal dysfunction in adult male patients with thalassemia major: An update for clinicians caring for thalassemia. *Expert Rev Hematol* 2017; *10* : 1095-106.
- Rund D, Rachmilewitz E. Beta-thalassemia. N Engl J Med 2005; 353: 1135-46.
- Prá D, Franke SI, Henriques JA, Fenech M. Iron and genome stability: An update. *Mutat Res* 2012; 733 : 92-9.
- Gupta M, Madkaikar M, Rao VB, Mishra A, Govindaraj P, Thangaraj K, *et al.* Mitochondrial DNA variations in myelodysplastic syndrome. *Ann Hematol* 2013; *92*: 871-6.
- Park J, Lee DG, Kim B, Park SJ, Kim JH, Lee SR, *et al.* Iron overload triggers mitochondrial fragmentation via calcineurin-sensitive signals in HT-22 hippocampal neuron cells. *Toxicology* 2015; 337: 39-46.
- Gundabolu K, Chen H, Li H, Shakaladevanapura L, Bhagat TL, Vallumsetla N, *et al*. Inhibition of erythropoiesis by iron overload is mediated through TGFβ signaling. *Blood* 2013; *122*: 2787.
- Khungwanmaythawee K, Sornjai W, Paemanee A, Jaratsittisin J, Fucharoen S, Svasti S, *et al.* Mitochondrial changes in β0-thalassemia/Hb E disease. *PLoS One* 2016; *11*: e0153831.
- Anderson GJ, Frazer DM. Current understanding of iron homeostasis. Am J Clin Nutr 2017; 106: 15598-68.
- Casu C, Aghajan M, Oikonomidou PR, Guo S, Monia BP, Rivella S, *et al.* Combination of tmprss6- ASO and the iron chelator deferiprone improves erythropoiesis and reduces iron overload in a mouse model of beta-thalassemia intermedia. *Haematologica* 2016; *101*: e8-11.
- Roydeva PG, Beckmann AM, Stirnberg M, Cesar J, Kikelj D, Ilaš J, *et al* 3,1-benzothiazines, 1,4-benzodioxines and 1,4-benzoxazines as inhibitors of matriptase-2: Outcome of a focused screening approach. *Pharmaceuticals (Basel)* 2016; 9. pii: E2.
- Ramos E, Ruchala P, Goodnough JB, Kautz L, Preza GC, Nemeth E, *et al.* Minihepcidins prevent iron overload in a hepcidin-deficient mouse model of severe hemochromatosis. *Blood* 2012; *120*: 3829-36.
- Zhen AW, Nguyen NH, Gibert Y, Motola S, Buckett P, Wessling-Resnick M, *et al.* The small molecule, genistein, increases hepcidin expression in human hepatocytes. *Hepatology* 2013; 58 : 1315-25.
- 23. Italia K, Colah R, Ghosh K. Hydroxyurea could be a good clinically relevant iron chelator. *PLoS One* 2013; 8 : e82928.

- Italia K, Chandrakala S, Ghosh K, Colah R. Can hydroxyurea serve as a free radical scavenger and reduce iron overload in β-thalassemia patients? *Free Radic Res* 2016; *50* : 959-65.
- Italia KY, Jijina FF, Merchant R, Panjwani S, Nadkarni AH, Sawant PM, *et al.* Effect of hydroxyurea on the transfusion requirements in patients with severe HbE-β-thalassaemia: A genotypic and phenotypic study. *J Clin Pathol* 2010; *63* : 147-50.
- Italia K, Colah R, Ghosh K. Experimental animal model to study iron overload and iron chelation and review of other such models. *Blood Cells Mol Dis* 2015; 55: 194-9.
- 27. Mohanty D, Ghosh K, Pathare AV, Karnad D. Deferiprone (L1) as an adjuvant therapy for *Plasmodium falciparum* malaria. *Indian J Med Res* 2002; *115* : 17-21.
- Falagas ME, Skalidis T, Vardakas KZ, Legakis NJ; Hellenic Cefiderocol Study Group. Activity of cefiderocol (S-649266) against carbapenem-resistant Gram-negative bacteria collected from inpatients in Greek hospitals. *J Antimicrob Chemother* 2017; 72: 1704-8.

- 29. Corcé V, Gouin SG, Renaud S, Gaboriau F, Deniaud D. Recent advances in cancer treatment by iron chelators. *Bioorg Med Chem Lett* 2016; *26* : 251-6.
- Lui GYL, Kovacevic Z, Richardson V, Merlot AM, Kalinowski DS, Richardson DR, *et al.* Targeting cancer by binding iron: Dissecting cellular signaling pathways. *Oncotarget* 2015; 6: 18748-79.
- Taher AT, Saliba AN, Kuo KH, Giardina PJ, Cohen AR, Neufeld EJ, *et al.* Safety and pharmacokinetics of the oral iron chelator SP-420 in β-thalassemia. *Am J Hematol* 2017; *92* : 1356-61.
- 32. Schloemer NJ, Brickler M, Hoffmann R, Pan A, Simpson P, McFadden V, et al. Administration of dexrazoxane improves cardiac indices in children and young adults with acute myeloid leukemia (AML) while maintaining survival outcomes. J Pediatr Hematol Oncol 2017; 39: e254-8.
- Bergeron RJ, Bharti N, McManis JS, Wiegand J. Metabolically programmed iron chelators. *Bioorg Med Chem* 2015; 23 : 5954-71.

372