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RESEARCH ARTICLE

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SARS‐CoV‐2 and SARS‐CoV: Virtual screening of potential inhibitors targeting RNA‐dependent RNA polymerase activity (NSP12)

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Abstract

Since the outbreak of severe acute respiratory syndrome (SARS) in 2003, the harm caused by coronaviruses to the world cannot be underestimated. Recently, a novel coronavirus (severe acute respiratory syndrome coronavirus‐2 [SARS‐CoV‐2]) initially found to trigger human severe respiratory illness in Wuhan City of China in 2019, has infected more than six million people worldwide by 21 June 2020, and which has been recognized as a public health emergency of international concern as well. And the virus has spread to more than 200 countries around the world. However, the effective drug has not yet been officially licensed or approved to treat SARS‐Cov‐2 and SARS‐Cov infection. NSP12‐NSP7‐NSP8 complex of SARS-CoV-2 or SARS-CoV, essential for viral replication and transcription, is generally regarded as a potential target to fight against the virus. According to the NSP12‐NSP7‐NSP8 complex (PDB ID: 7BW4) structure of SARS‐CoV‐2 and the NSP12‐NSP7‐NSP8 complex (PDB ID: 6NUR) structure of SARS‐CoV, NSP12‐NSP7 interface model, and NSP12‐NSP8 interface model were established for virtual screening in the present study. Eight compounds (Nilotinib, Saquinavir, Tipranavir, Lonafarnib, Tegobuvir, Olysio, Filibuvir, and Cepharanthine) were selected for binding free energy calculations based on virtual screening and docking scores. All eight compounds can combine well with NSP12‐NSP7‐NSP8 in the crystal structure, providing drug candidates for the treatment and prevention of coronavirus disease 2019 and SARS.

KEYWORDS

antiviral drugs, COVID‐19, drug candidates, NSP12‐NSP7‐NSP8, SARS‐CoV, SARS‐CoV‐2

1 | INTRODUCTION

Since the outbreak of pneumonia caused by a novel coronavirus (severe acute respiratory syndrome coronavirus‐2 [SARS‐CoV‐2]) in Wuhan, China, in December 2019, the world has faced unprecedented challenges in treating the disease caused by this virus.¹⁻³ SARS-CoV-2 is closely related to the SARS virus in Guangdong, China in 2002, both of which belong to the same family of viruses. $4-6$ China has

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implemented strict epidemic prevention and control measures across the country to avoid a larger-scale epidemic.⁷ However, the mortality rate of coronavirus disease 2019 (COVID‐19) caused by SARS‐CoV‐2 is much higher than that of SARS at present.⁴ The virus now presents in more than 200 countries. Even now though the SARS‐CoV no longer appears on a large scale, the infectious power and harmful effects of SARS‐CoV‐2 and SARS‐CoV should not be underestimated. To date, there are no officially licensed or approved drugs against this novel coronavirus and SARS‐CoV. There is an urgent need to find new targets for the development of anti‐SARS‐CoV‐2 and anti‐SARS‐CoV agents.

ORF1a and ORF1b at the 5′‐terminus of the coronavirus (CoV) genomes encode polyprotein 1a and polyprotein 1b, the two proteins could be cleaved into 16 nonstructural proteins (NSPs), which are essential for viral replication and transcription, thus being regarded as a potential virulence factor and a target for $COV^{8,9}$ $COV^{8,9}$ $COV^{8,9}$ Among these NSPs, the NSP12 subunit is the essential RdRp (RNA‐dependent RNA polymerase) of the coronavirus replicative machinery, which was even able to extend a homopolymeric primer‐template substrate by a few dozen nucleotides in vitro. 10,11 10,11 10,11 The 3.1 Å cryo-EM structure of the SARS‐CoV RNA polymerase NSP12 shows that it can bind with its essential cofactors NSP7 and NSP8. 12 12 12 The replication of the SARScoronavirus genome involves two RNA‐dependent RdRps. The first is primer-dependent and associated with the NSP12, whereas the second is catalyzed by NSP8. NSP8 is capable of de novo initiating the replication process and has been proposed to operate as a primase.¹³ In addition, NSP7, a component of the CoV replicase polyprotein, also participates in viral replication processed by binding to NSP12 as another primase. 13 13 13 The NSP12 needs to associate with NSP7 and NSP8 to activate its capability to replicate long RNA.^{[10](#page-11-0)} This elicits us to identify the particularly interesting compound disrupt the binding of NSP7 or NSP8 to NSP12, thus which could be used to inhibit the RdRp activity of NSP12, acting as novel antiviral agents and therapies of SARS‐CoV‐2 and SARS‐CoV.

The amino acid sequence alignment revealed that the NSP12 (PDB ID: 7BW4) of SARS‐CoV‐2 shared 96.35% similarity with the NSP12 (PDB ID: 6NUR) of SARS (Figure [1\)](#page-1-0). In addition, comparative analyses of their deduced amino acid sequences revealed that NSP7 and NSP8 of SARS‐CoV‐2 shared 98.8% and 97.5% similarity with that of SARS‐CoV respectively. The similarity of the target structure may also lead to similar antiviral drugs. Therefore, the NSP12 crystal structures of SARS‐CoV‐2 and SARS‐CoV were used as the target proteins. The NSP7 and NSP8 binding pocket of NSP12 were designated as active sites for screening compounds. Computer virtual screening in known drug databases helps to quickly identify potential drug candidates for COVID‐19 prevention and treatment. What's more, previous literature has shown that a number of virtuallyscreened compounds, such as ribavirin, lopinavir, and ritonavir, have proven to be effective in treating COVID-19.^{[14](#page-11-0)-17} Here, through high-throughput screening methods using a pool of 30 000 small molecules, several potential drug candidates were identified for preventing the binding NSP7 or NSP8 to NSP12, suggesting further assessment of the anti‐SARS‐CoV‐2 activity and anti‐SARS‐CoV activity of these compounds respectively in cell culture.

2 | MATERIALS AND METHODS

2.1 | Construction of small molecular ligands

Each sublibrary (Food and Drug Administration [FDA], world‐not‐FDA, investigational-only, <http://zinc.docking.org/substances/subsets/>) was downloaded from the zinc database. Then the model was converted to pdbat format by prepare receptor4.py script with assigning atomic types and atomic charges. All rotatable bonds in the molecule are set to be flexible for flexible docking.

2.2 | Preparation of target proteins

The crystal structure of SARS‐CoV‐2 NSP12 (PDB ID: 7BW4) was used as the target protein of SARS‐CoV‐2. And the crystal structure of SARS NSP12 (PDB ID: 6NUR) was used as the target protein of SARS‐CoV.

2.3 | Molecular docking

Vina1.1.2 was used to perform molecular docking. The docking boxes were set at the NSP12‐NSP7 interface and NSP12‐NSP8 interface (Figure [1](#page-1-0)), respectively. The search exhaustiveness was set as 32, and the number of binding modes was set as 9. Other parameters were set as default. During docking, NSP7 (or NSP8) was removed from the complex and only NSP12 was left as a receptor.

FIGURE 1 The docking boxes of the NSP12-NSP7 interface and NSP12‐NSP8 interface. The yellow part is nsp12. The green part is nsp7. The cyan part is nsp8. The docking box on the left is NSP12‐ NSP7 interface. The docking box on the right is NSP12‐NSP8 interface

TABLE 1 Ten compounds selected from the NSP12-NSP7 interface of SARS-CoV-2

Abbreviations: FDA, Food and Drug Administration; SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2.

2.4 | Binding free energy calculation

Each simulation system was immersed in a cubic box of TIP3P water with 10Å distance from the solute. The Na⁺ or Cl[−] was applied to neutralize the system. General Amber force field 15 and Amber ff14SB force field were used to parameterizing the ligand and protein respectively. A total of 10 000 steps of minimization with constraints (10 kcal/mol/Å2) on heavy atoms of complex, including 5000 steps of steepest descent minimization and 5000 steps of conjugate gradient minimization, were used to optimize each system. Then each system was heated to 300 K within 0.2 ns followed by 0.1 ns equilibration in NPT ensemble. Finally, 5 ns molecular dynamics (MD) simulation on each system at 300 K was performed. The minimization, heating, and equilibrium are performed with the sander program in Amber18. The 5 ns production run was performed with pmemd.cuda. Based on the 5 ns MD simulation trajectory, binding free energy (ΔG) was calculated with MM/GBSA method according to the following equation: $\Delta G_{\text{cal}} = \Delta$ H-T $\Delta S = \Delta E_{vdw} + \Delta E_{ele} + \Delta G_{eb} + \Delta G_{no}$ -T ΔS , where ΔE_{ele} and Δ_{EVDW} refer to electrostatic and van der Waals energy terms, respectively. $\Delta G_{\rm ph}$ and $\Delta G_{\rm np}$ refer to polar and nonpolar solvation free energies, respectively. Conformational entropy (TΔS) was not calculated for saving time. Besides, the ligands were compared based on the same target, so it is reasonable to ignore the entropy.

3 | RESULTS AND DISCUSSION

3.1 | Docking results of 7496 drugs against NSP12-NSP7 model and NSP12‐NSP8 model of SARS‐CoV‐2 and SARS‐CoV, respectively

The 7964 drugs obtained from the zinc database were screened for molecular docking. Among them, 20 top compounds showed

Abbreviations: FDA, Food and Drug Administration; SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2.

TABLE 3 Thirteen compounds selected from the NSP12‐NSP7 interface of SARS‐CoV

Compounds name	ID	Data	Affinity (kcal/mol)
Lonafarnih	ZINC3950115	Investigational-only	-8.4
Tegobuvir	ZINC100057121	Investigational-only	-8.2
Aromasin	ZINC3973334	FDA	-8
Troglitazone	ZINC968279	World-not-FDA	-8
Hypericin	ZINC3780340	Investigational-only	-8
Nilotinih	ZINC6716957	FDA	-7.9
Cepharanthine	ZINC30726863	World-not-FDA	-7.9
Eltrombopag	ZINC11679756	FDA	-7.8
Tipranavir	ZINC100016058	FDA	-7.8
Saquinavir	ZINC26985532	FDA	-7.8
Indinavir	ZINC22448696	FDA	-7.7
Cobicistat	ZINC85537014	FDA	-7.7
Talmapimod	ZINC34001955	Investigational-only	-7.5

Abbreviations: FDA, Food and Drug Administration; SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2.

the docking score in a range of −7.5 to −8.8 kcal/mol were selected from docking results from the NSP12‐NSP7 interface and NSP12‐NSP8 interface of SARS‐CoV‐2 (Tables [1](#page-2-0) and [2](#page-2-1)). Similarly, 24 top compounds showed the docking score in a range of −7.5 to −8.4 kcal/mol were selected from the NSP12‐NSP7 interface and NSP12‐NSP8 interface of SARS‐CoV (Tables [3](#page-3-0) and [4](#page-3-1)). Among the selected drugs, Tegobuvir can block both the NSP12‐NSP7 interface and the NSP12‐NSP8 interface of two coronaviruses. Cepharanthine can block both the NSP12‐NSP7 interface and the

Abbreviations: FDA, Food and Drug Administration; SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2.

NSP12‐NSP8 interface of SARS‐CoV‐2, as well as the NSP12‐ NSP8 interface of SARS‐CoV. Olysio and Filibuvir both can block the NSP12‐NSP8 interface of two coronaviruses. What is more, Lonafarnib can block the NSP12‐NSP7 interface of two coronaviruses. Further, Nilotinib, Tipranavir, and Saquinavir were all closely associated with antiviral activity.

3.2 | Docking results of Nilotinib against SARS‐ CoV‐2 NSP12‐NSP7

Nilotinib, a second‐generation small‐molecule tyrosine kinase inhibitor, is widely used in the treatment of chronic myeloid leukemia. 18 Studies have shown that Nilotinib has a potential antiviral effect.^{[19](#page-11-0)} Our docking results showed that Nilotinib was mainly combined with the interface between NSP12 and NSP7 of SARS‐ CoV‐2 through van der Waals potential energy and hydrogen bonds, involving LYS‐411 (Figure [2A](#page-4-0)). Nilotinib could bind to the interface active pockets of the SARS‐CoV‐2 NSP12 and NSP7 (Figure [2B](#page-4-0)). Therefore, Nilotinib can be considered as a candidate drug for treating SARS‐CoV‐2 infection.

3.3 | Docking results of Saquinavir and Tipranavir against SARS‐CoV NSP12‐NSP7

Saquinavir, the first HIV protease inhibitor was introduced into the market in 1995. Tipranavir, a novel nonpeptide protease inhibitor approved for use in patients with resistant strains of HIV. Both of which are safe and generally well-tolerated in HIV-1-infected adults.^{[20,21](#page-11-0)} Our docking results showed that five of the hydrogen bonds involving GLY‐297, PHE‐299, PHE‐325, PHE‐326, and ALA‐ 327 maintained upon the binding of saquinavir with interface

> TABLE 4 Eleven compounds selected from the NSP12‐NSP8 interface of SARS‐CoV

FIGURE 2 The binding model of Nilotinib against SARS‐CoV‐2 NSP12‐NSP7. A, Interactions between Nilotinib (cyan) and associated residues (off‐white) in the interface of the homology model for SARS‐CoV‐2. B, Binding models of Nilotinib (cyan) in the SARS‐CoV‐2 NSP12‐NSP7 protein interface pocket (white surface). Numbers accompanying dashed yellow lines represent the interaction distance (Å). SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2

between SARS‐CoV NSP12 and NSP7 (Figure [3A](#page-4-1)). As for Tipranavir, hydrogen bonds involving PHE‐325 maintained upon the binding of Tipranavir with interface between SARS‐CoV NSP12 and NSP7 (Figure [3C](#page-4-1)). Saquinavir and Tipranavir could bind to the interface active pockets of the SARS‐CoV NSP12 and NSP7 (Figures [3B](#page-4-1) and [3D](#page-4-1)). The previous study showed that saquinavir could bind to the SARS‐CoV‐2 RNA‐dependent RNA polymerase and inhibit the enzyme activity. 22 Our observations further confirm that saquinavir and tipranavir can bind to the NSP12‐NSP7 interface as interfacial blockers, thus making them as candidates for further in vitro evaluation of the anti‐SARS‐CoV activity.

3.4 | Docking results of Lonafarnib against SARS‐ CoV‐2 and SARS‐CoV NSP12‐NSP7

Lonafarnib as a nonpeptidomimetic inhibitor of farnesyltransferase has been used for progeria. $2^{3,24}$ Our docking results showed

FIGURE 3 The binding model of Saquinavir and Tipranavir against SARS‐CoV NSP12‐NSP7. A, Interactions between Saquinavir (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV. B, Binding models of Saquinavir (cyan) in the SARS‐CoV NSP12‐NSP7 protein interface pocket (white surface). C, Interactions between Tipranavir (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV. D, Binding models of Tipranavir (cyan) in the SARS‐CoV NSP12‐NSP7 protein interface pocket (white surface). Numbers accompanying dashed yellow lines represent the interaction distance (Å). SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2

FIGURE 4 The binding model of Lonafarnib against SARS‐CoV‐2 and SARS‐CoV NSP12‐ NSP7. A, Interactions between Lonafarnib (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV‐2. B, Binding models of Lonafarnib (cyan) in the SARS‐CoV‐2 NSP12‐NSP7 protein interface pocket (white surface). C, Interactions between Lonafarnib (cyan) and associated residues (off‐ white) in the interface of the crystal structure for SARS‐CoV. D, Binding models of Lonafarnib (cyan) in the SARS‐CoV NSP12‐NSP7 protein interface pocket (white surface). SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2

that lonafarnib was mainly combined with the interface between NSP12 and NSP7 of SARS‐CoV‐2 and SARS‐CoV through van der Waals potential energy and hydrophobic accumulation, involving PHE‐843, PHE‐440, PHE‐441, PHE‐442 (Figure [4A\)](#page-5-0), as well as PHE‐299, PHE‐727, PHE‐324, PHE‐325, and PHE‐326 (Figure [4C](#page-5-0)). Lonafarnib could both bind to the interface active pockets between the NSP12 and NSP7 of SARS‐CoV‐2 and SARS‐CoV (Figures [4B](#page-5-0) and [4D](#page-5-0)). Therefore, we speculate that Lonafarnib has a potential activity for the treatment of SARS‐Cov‐2 and SARS‐CoV infection.

3.5 | Docking results of Tegobuvir against SARS‐ CoV‐2 and SARS‐CoV

Tegobuvir (GS‐9190) is a specific, covalent inhibitor of the hepatitis C virus (HCV) nonstructural 5B (NS5B) RdRp with demonstrated antiviral activity in patients with genotype 1 chronic HCV infection.25‐[27](#page-11-0) Our docking results showed that the hydrogen bonds involving ALA‐443 and LEU‐329 maintained upon the binding of Tegobuvir and SARS‐CoV‐2 NSP12 interface (Figures [5A](#page-6-0) and [5C](#page-6-0)). Tegobuvir was mainly combined with the interface between SARS‐CoV NSP12 and NSP7 through van der

Waals potential energy and hydrophobic accumulation, involving PHE‐299, PHE‐727, PHE‐324, PHE‐325, and PHE‐326 (Figure [5E](#page-6-0)). Further, van der Waals forces mainly maintain the bond between Tegobuvir and the SARS‐CoV NSP12‐NSP8 interface (Figure [5G](#page-6-0)). Moreover, Tegobuvir could also bind to both of the interface active pockets of the SARS‐CoV‐2 and SARS‐CoV (Figures [5B,D](#page-6-0) and [5F,H\)](#page-6-0). Thus, Tegobuvir could be as a candidate drug against SARS‐CoV‐2 and SARS‐CoV.

3.6 | Docking results of Olysio against SARS‐CoV‐2 and SARS‐CoV NSP12‐NSP8

Olysio, HCV NS3/4A protease inhibitor approved for the treatment of genotype 1 chronic hepatitis C in combination with pegylated in-terferon and ribavirin.^{[28](#page-11-0)} Our docking results showed that the hydrogen bonds and van der Waals forces maintained upon the binding of Olysio and SARS‐CoV‐2 NSP12‐NSP8 interface, involving VAL‐330 and TYR-273 (Figure [6A](#page-7-0)). What is more, the hydrogen bonds involving VAL‐214 and van der Waals forces maintained upon the binding of the Olysio and SARS-CoV NSP12-NSP8 interface (Figure [6C](#page-7-0)). Olysio could also bind to the interface active pockets of the NSP12‐ NSP8 of SARS‐CoV‐2 and SARS‐CoV (Figures [6B](#page-7-0) and [6D](#page-7-0)).

FIGURE 5 The binding model of Tegobuvir against SARS ‐CoV ‐2 and SARS ‐CoV. A, Interactions between Tegobuvir (cyan) and associated residues (off ‐white) in the interface of the crystal structure for SARS ‐CoV ‐2. B, Binding models of Tegobuvir (cyan) in the SARS ‐CoV ‐ 2 NSP12 ‐NSP7 protein interface pocket (white surface). C, Interactions between Tegobuvir (cyan) and associated residues (off ‐white) in the interface of the crystal structure for SARS ‐CoV ‐2. D, Binding models of Tegobuvir (cyan) in the SARS ‐CoV ‐2 NSP12 ‐NSP8 protein interface pocket (white surface). E, Interactions between Tegobuvir (cyan) and associated residues (off ‐ white) in the interface of the crystal structure for SARS ‐CoV. F, Binding models of Tegobuvir (cyan) in the SARS ‐CoV NSP12 ‐NSP7 protein interface pocket (white surface). G, Interactions between Tegobuvir (cyan) and associated residues (off ‐ white) in the interface of the crystal structure for SARS ‐CoV. H, Binding models of Tegobuvir (cyan) in the SARS ‐CoV NSP12 ‐NSP8 protein interface pocket (white surface). Numbers accompanying dashed yellow lines represent the interaction distance (Å, C). SARS ‐CoV ‐2, severe acute respiratory syndrome coronavirus ‐ 2

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FIGURE 6 The binding model of Olysio against SARS‐CoV‐2 and SARS‐CoV NSP12‐ NSP8. A, Interactions between Olysio (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV‐2. B, Binding models of Olysio (cyan) in the SARS‐CoV‐2 NSP12‐NSP8 protein interface pocket (white surface). C, Interactions between Olysio (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV‐2. D, Binding models of Olysio (cyan) in the SARS‐ CoV‐2 NSP12‐NSP8 protein interface pocket (white surface). Numbers accompanying dashed yellow lines represent the interaction distance (Å, C). SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2

Thus, based on the present results, Olysio may be considered as a candidate for further in vitro evaluation of anti‐SARS‐CoV‐2 and anti‐SARS‐CoV activity.

3.7 | Docking results of Cepharanthine against SARS‐CoV‐2 and SARS‐CoV

Cepharanthine, an alkaloid tetrandrine isolated from Stephania tetrandra was found to exert strong anticancer, anti-inflammatory, and antioxidant activities. 2^9 In addition, it shows in vitro inhibitory effect on Herpes simplex virus type 1 (HSV-1) infected cells.³⁰ Our docking results showed that Filibuvir was mainly combined with the SARS‐ CoV‐2 NSP12‐NSP7 and NSP12‐NSP8 interface through van der Waals potential energy and hydrophobic accumulation, involving PHE‐843, PHE‐440, PHE‐441, PHE‐442 (Figures [7A](#page-8-0) and [7C\)](#page-8-0). Further, the hydrogen bonds involving ARG‐215 maintained upon the binding of Cepharanthine and SARS‐CoV NSP12‐NSP8 interface, with additionally van der Waals forces (Figure [7E\)](#page-8-0). Cepharanthine could bind to the interface active pockets of the SARS‐CoV‐2 and SARS‐CoV (Figures [7B,7D,7F\)](#page-8-0). The previous study showed Cepharanthine could significantly inhibit the replication of human coronavirus strains OC43[.29](#page-11-0) Taken together, Cepharanthine could be a potential natural

antiviral compound for the prevention and treatment of SARS‐CoV‐2 and SARS‐CoV infection.

3.8 | Docking results of Filibuvir against SARS‐CoV‐ 2 and SARS‐CoV NSP12‐NSP8

Filibuvir is an effective oral non-nucleoside HCV NS5B RdRp inhibitor which exhibits potent antiviral activity against subgenomic HCV replicons in cell culture assays and is a potential treatment of chronic HCV infection. $31,32$ Studies have shown that Filibuvir was well-tolerated and could be considered in combination with other antiviral drugs to achieve better safety and efficacy for chronic $HCV³¹$ Our docking results showed that the hydrogen bonds involving VAL‐330 maintained upon the binding of Cepharanthine and SARS‐CoV‐2 NSP12‐NSP8 interface, with additionally van der Waals forces (Figure [8A](#page-9-0)). Filibuvir was mainly combined with the SARS‐CoV NSP12‐NSP8 interface through van der Waals potential energy (Figure [8C\)](#page-9-0). Filibuvir could bind to the interface active pockets of the SARS‐CoV‐2 and SARS‐CoV NSP12‐NSP8 (Figures [8B](#page-9-0) and [8D](#page-9-0)). Thus, Filibuvir can be considered as a candidate drug for treating SARS‐ CoV‐2 and SARS‐CoV infection, providing evidence for further research.

FIGURE 7 The binding model of Cepharanthine against SARS‐CoV‐2 and SARS‐ CoV. A, Interactions between Cepharanthine (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV‐2. B, Binding models of Cepharanthine (cyan) in the SARS‐CoV‐2 NSP12‐NSP7 protein interface pocket (white surface). C, Interactions between Cepharanthine (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV‐2. D, Binding models of Cepharanthine (cyan) in the SARS‐CoV‐2 NSP12‐ NSP8 protein interface pocket (white surface). E, Interactions between Cepharanthine (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV. F, Binding models of Cepharanthine (cyan) in the SARS‐CoV NSP12‐NSP8 protein interface pocket (white surface). SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2

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3.9 | Binding free energy calculated by MM/GBSA

Through the simulation trajectory of 5 ns MD simulations, we calculated the binding free energy of eight drugs by MM/GBSA method. The calculated binding free energies of Nilotinib, Cepharanthine, Lonafarnib, Tegobuvir for the NSP12‐NSP7 of SARS‐CoV‐2 were −22.3412 ± 2.4994, −22.1316 ± 2.1664, −25.5364 ± 3.0488, −24.1066 ± 2.5504 kcal/mol, respectively, which highlighted Lonafarnib as the most active one (Table [5\)](#page-9-1). The interaction of van der Waals forces contributed more than the electrostatic interaction for Nilotinib, Cepharanthine, and Tegobuvir, indicating that van der Waals force is the main driving force for the combination of the three drugs. What is more, the calculated binding free energies of Olysio, Cepharanthine, Tegobuvir, and Filibuvir for the

NSP12‐NSP8 of SARS‐CoV‐2 were −29.2605 ± 3.6317, −29.8408 ± 3.3839, −26.7327 ± 5.0249, −28.1876 ± 2.3322 kcal/mol, respectively, indicating Tegobuvir with the strongest binding free energy (Table [6\)](#page-9-2).

The calculated binding free energies of Saquinavir, Tipranavir, Tegobuvir, and Lonafarnib for the NSP12‐NSP7 of SARS‐CoV were −23.0269 ± 4.4383, −12.1704 ± 3.2929, −24.4461 ± 3.5461, −16.8649 ± 1.8442 kcal/mol, respectively, which highlighted Tegobuvir as the most active one (Table [7\)](#page-10-0). The interaction of van der Waals forces contributed more than the electrostatic interaction for Tipranavir, Tegobuvir, and Lonafarnib indicating that van der Waals force is the main driving force for the combination of the three drugs. In addition, the calculated binding free energies of Olysio, Tegobuvir, Filibuvir, and Cepharanthine for the NSP12‐ NSP8 of SARS‐CoV were −28.5431 ± 3.4974, −23.8177 ± 2.8786,

FIGURE 8 The binding model of Filibuvir against SARS‐CoV‐2 and SARS‐CoV NSP12‐ NSP8. A, Interactions between Filibuvir (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV‐2. B, Binding models of Filibuvir (cyan) in the SARS‐ CoV‐2 NSP12‐NSP8 protein interface pocket (white surface). C, Interactions between Filibuvir (cyan) and associated residues (off‐white) in the interface of the crystal structure for SARS‐CoV. D, Binding models of Filibuvir (cyan) in the SARS‐ CoV NSP12‐NSP8 protein interface pocket (white surface). Numbers accompanying dashed yellow lines represent the interaction distance (Å). SARS‐ CoV‐2, severe acute respiratory syndrome coronavirus‐2

TABLE 5 The calculated binding energies of ligand to the interface of SARS‐CoV‐2 NSP12‐NSP7

Abbreviation: SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2.

 $^{\rm a}$ ΔE_{vdw} = van der Waals energy terms; ΔE_{ele} = electrostatic energy; ΔG_{gb} = polar solvation free energy; ΔG_{np} = nonpolar solvation free energy; ΔG_{cal} = final estimated binding free energy calculated from the above terms (kcal/mol).

Abbreviation: SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2.

 $^{\rm a}$ ΔE_{vdw} = van der Waals energy terms; ΔE_{ele} = electrostatic energy; ΔG_{gb} = polar solvation free energy; ΔG_{np} = nonpolar solvation free energy; ΔG_{cal} = final estimated binding free energy calculated from the above terms (kcal/mol).

TABLE 6 The calculated binding energies of ligand to the interface of SARS‐CoV‐2 NSP12‐NSP8

TABLE 7 The calculated binding energies of ligand to the interface of SARS‐CoV NSP12‐NSP7

Abbreviation: SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2.

 $^{\rm a}\Delta E_{\rm vdw}$ = van der Waals energy terms; $\Delta E_{\rm ele}$ = electrostatic energy; $\Delta G_{\rm gb}$ = polar solvation free energy; ΔG_{no} = nonpolar solvation free energy; ΔG_{cal} = final estimated binding free energy calculated from the above terms (kcal/mol).

TABLE 8 The calculated binding energies of ligand to the interface of SARS‐CoV NSP12‐NSP8

Abbreviation: SARS‐CoV‐2, severe acute respiratory syndrome coronavirus‐2.

 a ΔE_{vdw} = van der Waals energy terms; ΔE_{ele} = electrostatic energy; ΔG_{gb} = polar solvation free energy; ΔG_{np} = nonpolar solvation free energy; ΔG_{cal} = final estimated binding free energy calculated from the above terms (kcal/mol).

−30.0087 ± 2.6150, −27.9163 ± 3.1346 kcal/mol, respectively (Table [8\)](#page-10-1), indicating Filibuvir with the strongest binding free energy.

4 | CONCLUSION

The indiscriminate spread of the coronavirus poses a threat to human health around the world. So far, no drug has been officially approved to treat SARS‐CoV‐2 and SARS‐CoV infection. NSP12‐NSP7‐NSP8 complex is a potential target to fight against the virus. SARS‐CoV‐2 and SARS‐CoV have extremely high similarity to the target of this complex. As for a highly active NSP12 polymerase complex, viral cofactors NSP7 and NSP8 are essential, suggesting that the particularly interesting compounds could disrupt the binding of NSP7 or NSP8 to NSP12. Thus, this study carried out high‐throughput drug screening for such interfacial active pockets. Subsequently, 44 compounds were selected for further evaluation based on virtual screening and docking scores, leading eight compounds for the calculation of binding free energy. Among them, Lonafarnib, Tegobuvir, Olysio, Filibuvir, and Cepharanthine can simultaneously be used as candidate drugs for the treatment of SARS‐CoV‐2 and SARS‐CoV infection, making them possible for broad‐spectrum antiviral drugs. According to published literature, all of them have antiviral activity. Therefore, this study suggests that eight compounds can be tested in vitro for their anti‐SARS‐CoV‐2 or anti‐SARS‐CoV effect, providing more choices for clinical treatment and prevention of SARS‐CoV‐2 and SARS‐CoV infection.

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CONFLICT OF INTERESTS

The authors declare that there are no conflict of interests.

AUTHOR CONTRIBUTIONS

RZ, YT, JX, and HX designed the study. RZ and LC performed most studies, but a few were carried out by GY and HZ. All authors contributed thoughts and advice. YT, JX, and HX did a literature search. YT and JX collected the data. RZ. and LC analyzed and interpreted the data. RZ wrote the text, and the other authors contributed to the final text presentation. All authors have proved the submission.

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