RESEARCH Open Access



The significance of NTR1 expression and its correlation with β -catenin and EGFR in gastric cancer

Zhouyi Zhou¹, Jiaming Xie¹, Ying Cai², Shudong Yang², Ying Chen² and HaoRong Wu^{1*}

Abstract

Background: Several reports indicate the high-affinity receptor of NT (neurotensin), NTR1 (neurotensin receptor 1), in numerous detrimental functions linked to neoplastic progression of several cancer types. Recently, it has also been shown that NTR1 gene is a target of the Wnt/APC oncogenic pathways connected with the β -catenin/Tcf transcriptional complex and NT can stimulate cancer proliferation in an EGFR-dependent mechanism. In this study, we explored NTR1, β -catenin and EGFR expression in gastric cancer. The possible associations of NTR1 expression with clinicopathological factors, prognosis, β -catenin and EGFR were analyzed.

Methods: NTR1, β -catenin and EGFR expression in gastric cancer tissues and the adjacent normal tissues of 210 cases was detected by Immunohistochemistry. The possible associations of NTR1 expression with clinicopathological data, prognosis, β -catenin and EGFR were analyzed.

Results: 1. NTR1 expression in tumor tissues was significantly higher than that in adjacent normal tissues (P < 0.01). 2. Its expression was positively correlated with pathological grade, T stage, N stage and TNM stage and was not correlated with sex, age, tumor size and Lauren's classification. 3. A co-expression of NTR1 and nuclear β-catenin was in 53 (25.2 %) of cases and NTR1 expression was positively correlated with β-catenin nuclear translocation. NTR1 expression was not correlated with EGFR expression, but at a critical value (P = 0.05). 4. By log-rank test, higher expression of NTR1, higher pathological grade, diffusion Lauren's classification and advanced TNM stage showed worse prognosis (P < 0.05). Age, sex, tumor size, β-catenin and EGFR had no prognostic significance. Multivariate Cox analysis showed that NTR1 expression and TNM clinical stage (P < 0.05) were the independent prognostic factors for patients with GC.

Conclusion: By immunohistochemistry, we found that a high expression of NTR1 in GC specimens, which showed a bad prognosis, besides, NTR1 expression was related to invasion and migration of GC. These findings provide new and important information on the progression of GC. This study indicated that NTR1 may play an important role in tumor progression of GC and have its potential to be a predictive biomarker or a therapeutic molecular target in GC. The interaction between NTR1 and β -catenin may participate in the development of GC. However, the relationship between NTR1 and EGFR needs to be further investigated.

Keywords: Gastric cancer, NTR1, β-catenin, EGFR, Clinical pathology, Prognosis

Full list of author information is available at the end of the article



^{*} Correspondence: wuhaorong666@126.com

¹Department of General Surgery, The Second Affiliated Hospital of Soochow University, Suzhou 215004, China

Background

Gastric cancer is a frequent cause of cancer-related death in the world [1]. In China, gastric cancer ranks the third most common cancer [2]. Although the establishment of screening, early diagnosis and curative operation has increased survivals significantly, recurrence and metastasis remains a great challenge for patients with gastric cancer. Gastric cancer development is often associated with a number of molecular abnormalities, including the inactivation of various tumor suppressor genes and/or activation of various oncogenes [3, 4], but the mechanism of the development, invasion and metastasis of gastric cancer is still not clear. Investigations into the molecular alterations in gastric cancer may provide novel insights into the mechanisms responsible for stomach carcinogenesis and lead to the development of biomarkers for the early detection of gastric cancer and the prediction of its prognosis.

Neurotensin (NT) and its cognate receptor (neurotensin receptor 1, NTR1) are neuropeptide-receptor complexes frequently deregulated during the neoplastic process. NT is a 13-amino-acid peptide previously recognized for its distribution along the gastrointestinal tract [5]. The effects of NT are mediated by binding to three neurotensin receptors NTR-1, -2, and -3. The NTR1 and NTR2 are G-protein coupled receptors, while the NTR3 belongs in the sortilin receptor superfamily. The peripheral functions of NT are mainly mediated through its interaction with NTR1, a high affinity receptor coupled to a Gq/G11 protein [6]. Several reports implicate NTR1 in numerous detrimental functions linked to neoplastic progression of several cancer types, including pancreatic, prostate, colon, lung and head and neck cancers [7, 8]. In vivo and in vitro NT effects are abolished by the specific antagonist of NTR1, SR48692, which implies that NTR1 is a major mediator of these transforming actions [9].

NTR1 activation leads to cell proliferation, survival, mobility, and invasiveness in specific cancer cell types via signal transduction through PKC, extracellular signal-regulated kinase 1 and 2, RhoGTPases, NF- κ B, or focal adhesion kinase activation [10–12]. Some studies suggest that NT can stimulate many cancer proliferation in an EGFR-dependent mechanism, including foregut neuroendocrine tumor, prostate, colon and non-small cell lung cancers [13]. Recently, it has been shown that NTR1 gene is a target of the Wnt/APC oncogenic pathways connected with the β -catenin/Tcf transcriptional complex, known to activate genes involved in cell proliferation and transformation [14].

The expression of NTR1 and its relationship with EGFR and β -catenin in gastric cancer have not been reported. In this study, we explored NTR1, β -catenin and EGFR expression in gastric cancer. The possible

associations of NTR1 expression with clinicopathological factors, prognosis, β-catenin and EGFR were analyzed.

Methods

Patients

A total of 210 patients with histologically confirmed gastric adenocarcinoma invading the submucosal layer or deeper were retrospectively included in this study. All patients received the curative gastrectomy with lymph node dissection in our department of Wuxi People's Hospital Affiliated to Nanjing Medical University (Wuxi, China) between January 2011 and December 2011. No patients underwent chemotherapy or radiotherapy prior to surgery. A matched distant non-cancerous sample (5 cm away from the lesion) was also obtained from each patient and used as a control. The clinicopathological characteristics of the patients included age, gender, tumor size, pathological grade, Lauren's classification, TNM stage and lymph node metastasis status.

The median age of the patients was 64 years, ranging from 34 to 90 years. There were 160 males and 50 females (3.2:1). According to tumor differentiation, there were 93 (44.3 %) well and moderate differentiated tumors and 117 (55.7 %) poor and undifferentiated tumors. According to TNM stage, 51 (24.3 %) patients were at T1-2 stage while the left 159 (75.7 %) patients were at T3-4 stage. 157 (74.8 %) patients were positive and 53 (25.2 %) were negative for lymph node metastasis. There were 63 (30.0 %) stage I-II and 147 (70.0 %) stage III-IV patients. Follow-up information, including patients' outcome and the time interval between the date of surgical resection and the date of the cancer-related death, was collected. Those cases lost to follow-up and died from causes other than gastric cancer were regarded as censored data for the analysis of survival. The study was approved by our local ethics committees. Specimens were obtained with informed consent in accordance with the ethical standard of the Helsinki Declaration of 1975, as revised in 2000.

Immunohistochemistry

Paraffin slices were treated according to the EnVision immunohistochemical kit, and results were analyzed using a double-blind method. Two pathologists evaluated scores independently. PBS, instead of the primary antibody, was used as negative control. For NTR1 (polyclonal antibody, abcam, using a concentration of 5 μ g/ml) staining, the degree of expression was categorized as negative (positive staining of <10 % of tumor cells), positive (positive staining of >10 % and <50 % of tumor cells), or strongly positive (positive staining involving >50 % of tumor cells) [15].

According to Aust et al. [16], a semi-quantitative evaluation of β -catenin (monoclonal antibody, MAB-

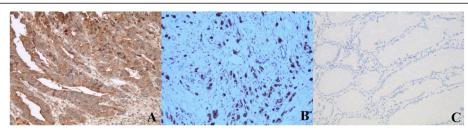


Fig. 1 Expression of NTR1 in gastric cancer tissues (200×). Strongly positive NTR1 expression in intestinal-(**a**) and diffuse–type (**b**) of gastric cancer tissues; negative NTR1 expression in the adjacent normal gastric mucosa (**c**)

259, Fuzhou Maixin Biotech. Co., Ltd, Pre-diluted) nuclear translocation was performed by estimating the percentage of positive nuclei as follows: 0; 1: 0 to 5 %; 2:5 to 25 %; 3: up to 25 %, and a cut-off of >5 % of positive β -catenin nuclei (score 2 or more) was considered significant for nuclear translocation of the protein.

EGFR (monoclonal antibody, RMA-0554, Fuzhou Maixin Biotech. Co., Ltd, Pre-diluted) reactivity was scored as 0 if there was no membranous reactivity within the tumor, or as 1+, 2+, or 3+ depending on the intensity above the background level [17]. The score of 1+ to 3+ was defined as positive and the score of 0 was defined as negative [18].

Statistical analysis

All data were analyzed using SPSS 18.0. x^2 tests (or Fisher's exact test) were used for samples classified as percentages; The spearman analysis was used to determine the correlation between two variables. Both the Kaplan–Meier method and log-rank test were used for single variant analysis, and a Cox model was used to analyze relationships between survival rates and multiple variables. P < 0.05 was considered statistically significant.

Results

NTR1 expression in tumor tissues was significantly higher than that in adjacent normal tissues and was positively correlated with pathological grade, T stage, N stage and TNM stage

According to the criteria established for immunostaining, 71.0 % (149/210) of tumors were positive for NTR1 staining. Only 28 (13.3 %) cases were positive in adjacent normal gastric mucosa. By way of the x^2 -test, NTR1 expression of tumor tissues was significantly higher (P < 0.01) (Fig. 1).

By way of Spearman analysis, NTR1 expression was positively correlated with pathological grade (Fig. 2), T stage, N stage and TNM stage and was not correlated with sex, age, tumor size and Lauren's classification (Table 1).

NTR1 expression was positively correlated with $\beta\text{-catenin}$ nuclear translocation and was not correlated with EGFR expression

Using immunohistochemical analysis, we observed a coexpression of NTS1 and nuclear β -catenin in 25.2 %(53) of cases and NTR1 expression was positively correlated with β -catenin nuclear translocation (P <0.05) (Figs. 3, 4

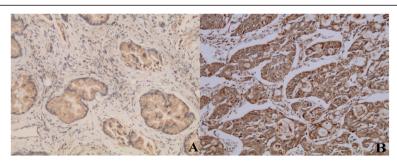


Fig. 2 Expression of NTR1 in intestinal-type of gastric cancer tissues (200x). Positive NTR1 expression in well differentiated intestinal-type of gastric cancer tissues (a); Strongly positive NTR1 expression in poor differentiated intestinal-type of gastric cancer tissues (b)

Table 1 Clinicopathologic features and the expression of NTR1 in 210 gastric cancer patients

ltem	NTR1 Expr			
	Negative	Positive	Strongly positive	<i>p</i> -value
Age				0.302
<60	15	27	23	
≥60	46	53	46	
Sex				0.315
Male	44	61	55	
Female	17	19	14	
Tumor size (cm)				0.291
<5	34	21	31	
≥5	27	59	38	
Pathological grade				0.027
Well and moderate	34	34	25	
Poor and undifferentiated	27	46	44	
Lauren's classification				0.125
Intestinal	32	39	27	
Diffusion	18	20	24	
Mixed	11	21	18	
T stage				0.001
T1-2	26	14	11	
T3-4	35	66	58	
N stage				0.000
Positive	33	66	58	
Negative	28	14	11	
TNM stage				0.011
I- II	28	18	17	
III-IV	33	62	52	

^{*}Spearman analysis

and 5) and was not correlated with EGFR expression (Fig. 6), however at a critical value (P = 0.05) (Table 2).

Higher expression of NTR1, higher pathological grade, diffusion Lauren's classification and TNM stage showed worse prognosis and high NTR1 expression and TNM clinical stage were the independent prognostic factors for patients with GC

We conducted NTR1 survival analysis on 210 cases of gastric cancer with follow-up data. The 1-year and 3-year survival rates of 210 cases were 78.9 % and 68.0 %. Patients with higher NTR1 expression showed a more unfavorable prognosis than those with no expression (P < 0.01) (Fig. 7). By log-rank test, higher expression of NTR1 (P = 0.000), higher pathological grade (P = 0.014), diffusion Lauren's classification (P = 0.004), advanced T stage (P = 0.026), TNM stage (P = 0.000), and N stage (P = 0.000) showed worse prognosis, and age, sex and tumor size had no prognostic significance. Multivariate Cox analysis showed that the following factors were the independent prognostic factors for patients with GC: NTR1 expression and TNM clinical stage (P < 0.05) (Table 3).

Discussion

NT has been shown to exert numerous oncogenic effects involved in tumor growth and metastatic spread. These effects are mostly mediated by NTR1, making NTR1 an actor in cancer-progression [7, 19]. Recently, it was found that the expression of NTR1 was significantly correlated to an increase in the number of tumors when sporadic cancer was generated in mouse models by inflammation [20]. Vias et al. [21] found that long-term anti-androgen treatment of LNCaP cells produced a sub-line exhibiting up-regulated expression of NT and NTRs, which increased the proliferation rate, accelerated cell cycle progression, and increased invasiveness. NTR1 expression was also found at very high levels in the human androgen-independent PC3 cell line. In pancreas, NT binding sites were found specifically in pancreatic cancer but not in normal pancreas and chronic

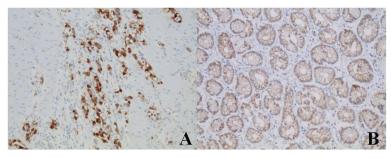


Fig. 3 β-catenin expression in gastric cancer and adjacent normal gastric mucosa (200×). Nuclear β-catenin expression in gastric cancer (**a**) and nomal β-catenin membrane staining in the adjacent normal gastric mucosa (**b**)



Fig. 4 Expression of NTR1 and β-catenin in intestinal-type of gastric cancer tissues (200×). HE staining image (**a**), IHC staining of NTR1 (**b**) and nuclear β-catenin expression (**c**) in the same region of the same intestinal-type cases. ×200

pancreatitis [22]. Moderate to strong expression of NTR1 in colonic adenomas and adenocarcinomas suggest that increased NTR1 expression may be an early event during colonic tumorigenesis in colonic adenocarcinomas [23]. Using immunohistochemical analysis, we found NTR1 protein was up-regulated obviously in 71 % of the patients than that in adjacent normal tissues and was correlated with clinicopathological factors and prognosis, therefore speculating NTR1 gene over-expression may be involved in the pathogenesis of GC.

Besides the contribution of NTR1 expression in tumor progression and aggressive behavior [23], it was also identified as a prognosis marker in colon, breast, lung, and head and neck carcinomas and malignant pleural mesothelioma [19, 24]. In pancreatic cancer, the NTR1 mRNA levels were higher in advanced tumor stage (stages III and IV) than early tumor stage (stages I and II) [25]. In breast invasive carcinomas, the high expression of NTR1 was associated with the SBR grade, the size of the tumor, and the number of metastatic lymph nodes [26]. In colonic adenocarcinomas, adenocarcinomas that infiltrated into and beyond the muscularis propria showed a higher intensity of NTR1 expression compared with tumors that were localized to the mucosa or submucosa. In some cases, infiltrating margins and foci of lymphovascular invasion

showed a higher intensity of expression than the main mass of the tumor [23]. In head and neck squamous cell carcinomas, NT and NTR1 mRNA high levels were significantly correlated with higher rates of distant metastasis as well as with the survival rate [8]. Experimental tumor xenografts generated by NT- and NTR1silenced human lung cancer cells revealed that NT enhanced primary tumor growth and production of massive nodal metastasis via autocrine and paracrine regulation loops [15]. Investigating the biological role of NTR1 expression in GC by IHC, we found that NTR1 expression was positively correlated with histologic grade, TNM stage and lymph node metastasis in GC. Univariate analysis showed that the following factors were significantly related to postoperative survival in GC: NTR1 expression and TNM clinical stage. Our results suggest that NTR1 is a potential marker and/or a pejorative mediator of gastric cancer progression associated with poor prognosis. Lymph node metastasis is usually a reliable prognostic indicator, but skip metastases and micrometastases easily lead to missed diagnosis in routine pathology work. Tumor stage is a prognostic factor, however it is relatively hysteretic for prognosis. Therefore, high expression of NTR1 could lead to clinical attention even without lymph node metastasis. Currently, further studies are needed on molecular mechanism of NTR1 promoting invasion and metastasis

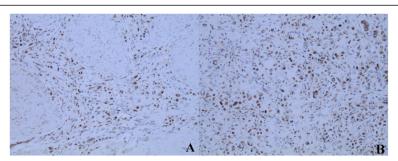


Fig. 5 Expression of NTR1 and β -catenin in diffuse-type of gastric cancer tissues (200x). IHC staining of NTR1 and β -catenin in the same case of the diffuse-type cases. ×200

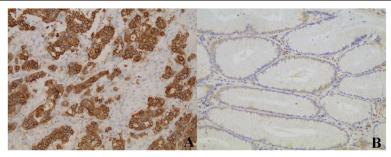


Fig. 6 EGFR expression in gastric cancer and adjacent normal gastric mucosa. Positive EGFR expression (score 3+) in gastric cancer (**a**) and negative EGFR expression in the adjacent normal gastric mucosa (**b**)

and investigations of whether NTR1 could be used as a target for novel therapeutic approaches in GC.

EGFR transactivation by NT/NTR1 complex has been observed in several cell lines. In PC3 prostate cancer cells, NT activates proliferation through EGFR transactivation in a PKC (protein kinase C, PKC)dependent pathway [27, 28]. In the colonic HCT116 cells, NT/NTR1 induces a PKC-dependent ERK phosphorylation and an EGFR metalloproteinase-mediated transactivation, however, in colonic HT-29 cells, the EGFR tyrosine kinase inhibitor, gefitinib, blocks NTSstimulated phosphorylation of both ERK and Akt, indicating the transactivation of EGFR independently of PKC. The activation of Akt is only partly inhibited by gefitinib, suggesting an multiple mechanism to EGFR transactivation in a partially redundant manner [29]. The mechanism of NT-induced EGFR transactivation is still not clearly elucidated. The release of EGFR ligands-like (TGF-α, Hb-EGF, or amphiregulin), as pro-ligand, by NT has been proposed. These ligands are released by proteolytic cleavage involving enzymes of the metalloproteinase family [30-32]. Once released, these ligands bind to EGFR and activate the downstream signaling cascades of EGFR activation [27]. These findings suggest a cooperative relationship between the neurotensinergic system and EGFR pathway.

Table 2 Correlation between NTR1 and β-catenin nuclear translocation and EGFR expression in gastric cancer

Item	NTR1 Expre	NTR1 Expression intensity			
	Negative	Positive	Strongly positive		
Nuclear ß-catenin expression					
Negative	42	60	36		
Positive	19	20	33		
EGFR expression				0.050	
Negative (score 0)	29	24	21		
Positive (score 1-3+)	32	56	48		

^{*}Spearman analysis

However, Massa F et al. observed that although both NT and EGF enhance colonic epithelial cells growth, the intracellular pathways involved in these effects are quite independent and NT is unable to transactivate EGFR in two cell lines, HT29 and HCT116 [33], in contrast to results obtained in other colonic and prostatic cancer cell lines [27, 34]. These distinct pathways could be the consequence of the co-expression of NTR1 and NTR3 that are shown to be involved in complex for the signaling of NT [6]. Olszewski-Hamilton U. et al. [35] revealed dependence of relative expression of NTR1 and EGFR on cell density and extracellular pH in human pancreatic cancer cell lines. They suggested that downregulation of EGFR at higher cell densities concomitant with upregulation of NTR1 seems to indicate mutual exclusive roles for these receptors. EGFR may be important in the initial growth of pancreatic tumor cells and replaced by increased expression of other growth factor receptors, like NTR1, during metastatic dissemination. Taken together, these observations indicate that the cellular pathways leading to cell proliferation by NT and EGF are more complex in some cancer cell lines and that the ways to develop tools to decrease tumor growth remain more complicated than expected from previous studies. Using immunohistochemical analysis, we found that NTR1 expression was not correlated with EGFR expression, however at a critical value (P = 0.05). We think that, in addition to the limited number of cases, this phenomenon may be due to the heterogeneity in gastric cancer. NTR1 expression may be correlated with EGFR expression in certain types, while not in some other types, thus needing to be further confirmed by cell gene technology.

Some other mechanisms may exist, recently NTR1 gene expression induced by the accumulation of β -catenin in human colonic adenomas has been found. In support of this hypothesis, analysis of the regulatory sequences in the NTR1 gene revealed the presence of a consensus T cell factor (Tcf) binding site potentially linking the activation of the NTR1 and APC (adenomatous polyposis coli)/b-catenin pathway.

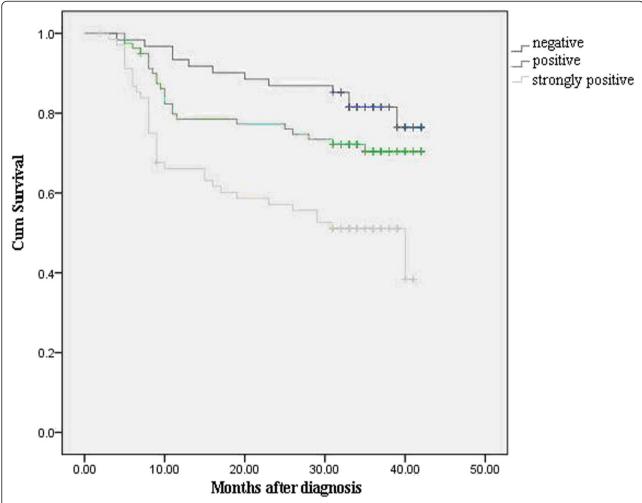


Fig. 7 Overall survival curves of patients with gastric cancer according to the immunostaining results of NTR1. Higher expression of NTR1 (P = 0.000) showed worse prognosis

It has been demonstrated that inhibitors of GSK-3 β (protein kinase involved in the phosphorylation of β -catenin and its degradation) which cause the significant accumulation of β -catenin, upregulate the level of NTR1 transcription [36]. During inflammatory bowel disease-related oncogenesis, two pathways of NTR1 overexpression existed: one triggered by NT overexpression, and a second associated with β -catenin nuclear accumulation, both being not mutually exclusive [37]. Similar results have been obtained

in other cancers such as lung, prostate, and breast cancers [38, 39]. In our study, we found that NTR1 expression was associated with nuclear β -catenin expression, suggesting that the interaction between NTR1 and β -catenin may participate in the development of GC. What needs to be further confirmed is whether β -catenin upregulate the level of NTR1 expression or upregulated NTR1 expression potentiate β -catenin nuclear translocation and thus activate β -catenin mediated signaling pathway.

Table 3 Multiple factor Cox model regression variable form of gastric cancer

ltem	В	SE	Wald	df	Sig. ^a	Exp (B)	95.0 % CI for Exp (B)	
							Lower	Upper
NTR1 Expressin Intensity	0.444	0.186	5.686	1	0.017	1.559	1.082	2.245
N stage	3.097	1.122	7.624	1	0.006	22.139	2.457	199.522

^aCox model analysis

Conclusion

By immunohistochemistry, we found the association between NTR1 and GC specimens, in which patients with high NTR1 expression have a bad prognosis. The relationship between NTR1 and invasion and migration of GC was revealed. These findings provide new and important information on the progression of GC. This study indicated that NTR1 may play an important role in tumor progression of GC and have its potential as a predictive biomarker and a therapeutic molecular target in GC. The interaction between NTR1 and β -catenin may participate in the development of GC, while the relationship between NTR1 and EGFR needing to be further investigated.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

ZYZ and HRW designed the experiment. ZYZ and JMX analyzed the data and wrote the manuscript. YC carried out the immunohistochemical examination. YC and SDY evaluated clinicopathological data. All authors read and approved the final manuscript.

Authors' information

Zhouyi Zhou, Working in Wuxi People's Hospital Affiliated to Nanjing Medical University.

Acknowledgements

We gratefully acknowledge financial support from scientific research fund of Department of Pathology, Wuxi People's Hospital Affiliated to Nanjing Medical University.

The authors thank Qiu Rao for assistance in designing the experiment.

Author details

¹Department of General Surgery, The Second Affiliated Hospital of Soochow University, Suzhou 215004, China. ²Department of Pathology, Wuxi People's Hospital Affiliated to Nanjing Medical University, Wuxi 214023, China.

Received: 20 April 2015 Accepted: 9 July 2015 Published online: 28 July 2015

References

- Blum MA, Takashi T, Suzuki A, Ajani JA. Management of localized gastric cancer. J Surg Oncol. 2013;107:265–70.
- Lin Y, Ueda J, Kikuchi S, Totsuka Y, Wei WQ, Qiao YL, et al. Comparative epidemiology of gastric cancer between Japan and China. World J Gastroenterol. 2011;17:4421–8.
- Tahara E. Molecular aspects of invasion and metastasis of stomach cancer. Verh Dtsch Ges Pathol. 2000;84:43–9.
- 4. Sud R, Wells D, Talbot IC, Delhanty JD. Genetic alterations in gastric cancers from British patients. Cancer Genet Cytogenet. 2001;126:111–9.
- Vita N, Laurent P, Lefort S, Chalon P, Dumont X, Kaghad M, et al. Cloning and expression of a complementary DNA encoding a high affinity human neurotensin receptor. FEBS Lett. 1993;317:139–42.
- Martin S, Navarro V, Vincent JP, Mazella J. Neurotensin receptor-1 and -3 complex modulates the cellular signaling of neurotensin in the HT29 cell line. Gastroenterology. 2002;123:1135–43.
- Thomas RP, Hellmich MR, Townsend Jr CM, Evers BM. Role of gastrointestinal hormones in the proliferation of normal and neoplastic tissues. Endocr Rev. 2003;24:571–99.
- Shimizu S, Tsukada J, Sugimoto T, Kikkawa N, Sasaki K, Chazono H, et al. Identification of a novel therapeutic target for head and neck squamous cell carcinomas: a role for the neurotensin-neurotensin receptor 1 oncogenic signaling pathway. Int J Cancer. 2008;123:1816–23.

- Gully D, Canton M, Boigegrain R, Jeanjean F, Molimard JC, Poncelet M, et al. Biochemical and pharmacological profile of a potent and selective nonpeptide antagonist of the neurotensin receptor. Proc Natl Acad Sci U S A. 1993:90:65–9.
- Zhao D, Pothoulakis C. Rho GTPases as therapeutic targets for the treatment of inflammatory diseases. Expert Opin Ther Targets. 2003;7:583–92.
- Leyton J, Garcia-Marin L, Jensen RT, Moody TW. Neurotensin causes tyrosine phosphorylation of focal adhesion kinase in lung cancer cells. Eur J Pharmacol. 2002;442:179–86.
- Ehlers RA, Zhang Y, Hellmich MR, Evers BM. Neurotensin-mediated activation of MAPK pathways and AP-1 binding in the human pancreatic cancer cell line, MIA PaCa-2. Biochem Biophys Res Commun. 2000;269:704–8.
- Moody TW, Chan DC, Mantey SA, Moreno P, Jensen RT. SR48692 inhibits non-small cell lung cancer proliferation in an EGF receptor-dependent manner. Life Sci. 2014;100:25–34.
- Souazé F, Viardot-Foucault V, Roullet N, Toy-Miou-Leong M, Gompel A, Bruyneel E, et al. Neurotensin receptor 1 gene activation by the Tcf/beta-catenin pathway is an early event in human colonic adenomas. Carcinogenesis. 2006;27:708–16.
- Alifano M, Souazé F, Dupouy S, Camilleri-Broët S, Younes M, Ahmed-Zaïd SM, et al. Neurotensin receptor 1 determines the outcome of non-small cell lung cancer. Clin Cancer Res. 2010;16:4401–10.
- Aust DE, Terdiman JP, Willenbucher RF, Chew K, Ferrell L, Florendo C, et al. Altered distribution of beta-catenin, and its binding proteins E-cadherin and APC, in ulcerative colitis-related colorectal cancers. Mod Pathol. 2001;14:29–39.
- Kim MA, Lee HS, Lee HE, Jeon YK, Yang HK, Kim WH. EGFR in gastric carcinomas: prognostic significance of protein overexpression and high gene copy number. Histopathology. 2008;52:738–46.
- Atmaca A, Werner D, Pauligk C, Steinmetz K, Wirtz R, Altmannsberger HM, et al. The prognostic impact of epidermal growth factor receptor in patients with metastatic gastric cancer. BMC Cancer. 2012;12:524.
- Dupouy S, Mourra N, Doan VK, Gompel A, Alifano M, Forgez P.
 The potential use of the neurotensin high affinity receptor 1 as a biomarker for cancer progression and as a component of personalized medicine in selective cancers. Biochimie. 2011:93:1369–78.
- Bugni JM, Rabadi LA, Jubbal K, Karagiannides I, Lawson G, Pothoulakis C.
 The neurotensin receptor-1 promotes tumor development in a
 sporadic but not an inflammation-associated mouse model of colon cancer.
 Int J Cancer. 2012;130:1798–805.
- Vias M, Burtt G, Culig Z, Veerakumarasivam A, Neal DE, Mills IG. A role for neurotensin in bicalutamide resistant prostate cancer cells. Prostate. 2007;67:190–202.
- Wang JG, Li NN, Li HN, Cui L, Wang P. Pancreatic cancer bears overexpression of neurotensin and neurotensin receptor subtype-1 and SR 48692 counteracts neurotensin induced cell proliferation in human pancreatic ductal carcinoma cell line PANC-1. Neuropeptides. 2011;45:151–6.
- Gui X, Guzman G, Dobner PR, Kadkol SS. Increased neurotensin receptor-1 expression during progression of colonic adenocarcinoma. Peptides. 2008;29:1609–15.
- Alifano M, Loi M, Camilleri-Broet S, Dupouy S, Régnard JF, Forgez P. Neurotensin expression and outcome of malignant pleural mesothelioma. Biochimie. 2010;92:164–70.
- Wang L, Friess H, Zhu Z, Graber H, Zimmermann A, Korc M, et al. Neurotensin receptor-1 mRNA analysis in normal pancreas and pancreatic disease. Clin Cancer Res. 2000:6:566–71.
- Dupouy S, Viardot-Foucault V, Alifano M, Souazé F, Plu-Bureau G, Chaouat M, et al. The neurotensin receptor-1 pathway contributes to human ductal breast cancer progression. PLoS One. 2009;4:e4223.
- Hassan S, Dobner PR, Carraway RE. Involvement of MAP-kinase, Pl3-kinase and EGF-receptor in the stimulatory effect of Neurotensin on DNA synthesis in PC3 cells. Regul Pept. 2004;120:155–66.
- Amorino GP, Deeble PD, Parsons SJ. Neurotensin stimulates mitogenesis of prostate cancer cells through a novel c-Src/Stat5b pathway. Oncogene. 2007;26:745–56.
- 29. Müller KM, Tveteraas IH, Aasrum M, Ødegård J, Dawood M, Dajani O, et al. Role of protein kinase C and epidermal growth factor receptor signalling in growth stimulation by neurotensin in colon carcinoma cells. BMC Cancer. 2011;11:421.

- 30. Sanderson MP, Dempsey PJ, Dunbar AJ. Control of ErbB signaling through metalloprotease mediated ectodomain shedding of EGF-like factors. Growth Factors. 2006;24:121–36.
- Zhao D, Zhan Y, Zeng H, Koon HW, Moyer MP, Pothoulakis C. Neurotensin stimulates expression of early growth response gene-1 and EGF receptor through MAP kinase activation in human colonic epithelial cells. Int J Cancer. 2007;120:1652–6.
- Kataoka H. EGFR ligands and their signaling scissors, ADAMs, as new molecular targets for anticancer treatments. J Dermatol Sci. 2009;56:148–53.
- 33. Massa F, Tormo A, Béraud-Dufour S, Coppola T, Mazella J. Neurotensininduced Erk1/2 phosphorylation and growth of human colonic cancer cells are independent from growth factors receptors activation. Biochem Biophys Res Commun. 2011;414:118–22.
- Zhao D, Zhan Y, Koon HW, Zeng H, Keates S, Moyer MP, et al. Metalloproteinase-dependent transforming growth factor-alpha release mediates neurotensin-stimulated MAP kinase activation in human colonic epithelial cells. J Biol Chem. 2004;279:43547–54.
- Olszewski-Hamilton U, Hamilton G. Dependence of relative expression of NTR1 and EGFR on cell density and extracellular pH in human pancreatic cancer cell lines. Cancers (Basel). 2011;3:182–97.
- Wang X, Jackson LN, Johnson SM, Wang Q, Evers BM. Suppression of neurotensin receptor type 1 expression and function by histone deacetylase inhibitors in human colorectal cancers. Mol Cancer Ther. 2010;9:2389–98.
- Bossard C, Souazé F, Jarry A, Bezieau S, Mosnier JF, Forgez P, et al.
 Over-expression of neurotensin high-affinity receptor 1 (NTS1) in relation with its ligand neurotensin (NT) and nuclear beta-catenin in inflammatory bowel disease-related oncogenesis. Peptides. 2007;28:2030–5.
- Chesire DR, Ewing CM, Gage WR, Isaacs WB. In vitro evidence for complex modes of nuclear beta-catenin signaling during prostate growth and tumorigenesis. Oncogene. 2002;21:2679–94.
- Turashvili G, Bouchal J, Burkadze G, Kolar Z. Wnt signaling pathway in mammary gland development and carcinogenesis. Pathobiology. 2006;73:213–23.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

