

Research article

Open Access

## Ischemic preconditioning reduces the severity of ischemia-reperfusion injury of peripheral nerve in rats

Yusuf Kenan Coban\*<sup>1</sup>, Harun Ciralik<sup>2</sup> and Ergul Belge Kurutas<sup>3</sup>

Address: <sup>1</sup>Dept. Of Plastic Surgery, Sutcuimam University, School of Medicine, Kahramanmaraş, Turkey, <sup>2</sup>Dept. Of Pathology, Sutcuimam University, School of Medicine, Kahramanmaraş, Turkey and <sup>3</sup>Dept. Of Biochemistry, Sutcuimam University, School of Medicine, Kahramanmaraş, Turkey

Email: Yusuf Kenan Coban\* - [cobanyk@ksu.edu.tr](mailto:cobanyk@ksu.edu.tr); Harun Ciralik - [ciralikh@yahoo.com](mailto:ciralikh@yahoo.com); Ergul Belge Kurutas - [ekurutas@hotmail.com](mailto:ekurutas@hotmail.com)

\* Corresponding author

Published: 29 September 2006

Received: 20 February 2006

Accepted: 29 September 2006

*Journal of Brachial Plexus and Peripheral Nerve Injury* 2006, 1:2 doi:10.1186/1749-7221-1-2

This article is available from: <http://www.JBPPNI.com/content/1/1/2>

© 2006 Coban et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background and aim:** Allow for protection of briefly ischemic tissues against the harmful effects of subsequent prolonged ischemia is a phenomenon called as Ischemic Preconditioning (IP). IP has not been studied in ischemia-reperfusion (I/R) model of peripheral nerve before. We aimed to study the effects of acute IP on I/R injury of peripheral nerve in rats.

**Method:** 70 adult male rats were randomly divided into 5 groups in part 1 experimentation and 3 groups in part 2 experimentation. A rat model of severe nerve ischemia which was produced by tying iliac arteries and all identifiable anastomotic vessels with a silk suture (6-0) was used to study the effects of I/R and IP on nerve biochemistry. The suture technique used was a slip-knot technique for rapid release at time of reperfusion in the study. Cytoplasmic vacuolar degeneration was also histopathologically evaluated by light microscopic examination in sciatic nerves of rats at 7th day in part 2 study.

**Results:** 3 hours of Reperfusion resulted in an increase in nerve malondialdehyde levels when compared with ischemia and non-ischemia groups ( $p < 0.001$  and  $p < 0.0001$  respectively). IP had significantly lower nerve MDA levels than 3 h reperfusion group ( $p < 0.001$ ). The differences between ischemic, IP and non-ischemic control groups were not significant ( $p > 0.05$ ). There was also a significant decrease in vacuolar degeneration of sciatic nerves in IP group than I/R group ( $p < 0.05$ ).

**Conclusion:** IP reduces the severity of I/R injury in peripheral nerve as shown by reduced tissue MDA levels at 3 th hour of reperfusion and axonal vacuolization at 7 th postischemic day.

### Background

Ischemia-reperfusion (I/R) causes oxidative injury and ischemic fiber degeneration (IFD), due to injury of the neuron and axon, after enough ischemic times, i.e. 4–5 hours of peripheral nerve ischemia [1,2]. Maximal intercellular adhesion molecule-1 (ICAM-1) expression on endoneurial vessels and polymorphonuclear monocytes

reaches a peak at 24 th hour and macrophages increases nearly four fold at 48–72 hour of reperfusion after a 5 h of near-complete ischemia [3]. All these cells are responsible for demyelination and IFD at prolonged reperfusion after enough ischemic times in peripheral nerves [4,5]. Nerve lipid hydroperoxides reaches greatest levels at 3 hour and a gradual decline follows over the next month

with reperfusion [6]. An aggravated reperfusion injury in Streptozocine induced diabetic rats could be seen with less severe ischemic times [7]. Clinical experience related to I/R injury of peripheral nerve shows that neurologic recovery is possible, if reperfusion starts within 6 hours after ischemia [8].

Allow for protection of briefly ischemic tissues against the harmful effects of subsequent prolonged ischemia is a phenomenon called as Ischemic Preconditioning (IP)[9]. There are two distinct types of protection afforded by this adaptational response, i.e. acute and delayed preconditioning. The factors that initiate the acute and delayed preconditioning responses appear to be similar. However, the protective effects of acute preconditioning are protein synthesis independent, while the effects of delayed preconditioning require protein synthesis. Adaptational responses to I/R injury have been demonstrated in different tissue types [10-14]. IP has not been studied in I/R model of peripheral nerve before. We aimed to study the effects of acute IP on I/R injury of peripheral nerve in rats.

## Materials and methods

### Animals

All animals were obtained from Experimental Research Laboratory of Sutcuimam University School of Medicine. The experimental design was approved by the Ethical committee of KSU. 200–250 g adult male sprague-dawley rats were used in the study. The animals were fed with standard rats diets until the surgical procedures.

We examined I/R induced pathological and biochemical changes along the length of sciatic nerve. Major arteries which supply rat hindlimb were occluded for 4 hours. Reperfusion was accomplished by the release of ties of abdominal aorta and its branches. Nerve pathology and biochemical analysis in sciatic nerve samples of the rats were assessed after 3 hours and 7 days of reperfusion. A total of 70 rats were used in the study. The study was divided into two parts. Part 1 included the biochemical examination of Ischemia, I/R and I/R+IP on sciatic nerves of rats at the early period. Part 2 which consisted of 3 groups aimed to evaluate the histopathological changes in the nerves 7 days after the experimentation. The rats were randomly divided into following groups, 7 rats in each;

#### Part 1: Short time effects of I/R and IP

**Group I-** Normal adult male rats (Non-isch): Non-ischemic group, no intervention was made, simply sciatic nerve samples were taken.

**Group II-** Ischemic group (Ischemic control-0hR): 4 hours of limb ischemia were done and the samples were taken from the sciatic nerves after ischemic insult.

**Group III-** Ischemia-reperfusion group (3hR): 4 hours of ischemia and following 3 hours of reperfusion were done. After I/R of sciatic nerves samples were taken.

**Group IV-** I/R plus ischemic preconditioning group (3hR+IP): Preconditioning (three cycles of 5 minutes of short ischemia with 2 minute's intervals), and then 4 hours of ischemia with 3 hours of reperfusion.

#### Part 2: Long time effects of I/R and IP

**Group 1-** I/R with long duration (7dR): 4 hours of ischemia and 7 days of reperfusion.

**Group 2-** Preconditioning plus I/R with long duration (7dR+IP): The same preconditioning protocol with the group IV, and then, 4 hours of ischemia with following 7 days of reperfusion. In both groups, sciatic nerve samples taken from both limb at 7th day were examined histopathologically.

**Group 3-** Sham operated group: Abdominal aorta and its collaterals were simply exposed under anesthesia, but no intervention was done. Then abdominal incision was primarily closed. At 7th day, sciatic nerve samples were taken as done in the other groups.

### Model of severe nerve ischemia

Our model of severe nerve ischemia was produced by tying of the ilio-lumbar and inferior mesenteric arteries followed by the temporary occlusion of the abdominal aorta and both iliac arteries [15]. We tied off all identifiable anastomotic vessels, including the ilio-lumbar and inferior mesenteric arteries. The aorta and iliac arteries were tied with a silk suture (6-0), using a slip-knot technique for rapid release, when needed. Measurements of the femoral blood pressure (BP) were used to monitor the completeness of the occlusion, and direct inspection of the sciatic nerve epineurial vessels showed that blood flow had been arrested. Sluggish flow was sometimes seen in these vessels several minutes after aortic occlusion, presumably due to partial reestablishment of anastomotic flow.

### Ischemia-reperfusion and ischemic preconditioning model

The rat was anesthetized with intraperitoneal pentobarbital (60 mg/kg) followed by surgery to produce IR. Ischemia was produced by ligating the abdominal aorta, the right iliac artery, the right femoral artery, and all identifiable collateral vessels supplying the sciatic nerves with 6-0 silk sutures. After 3 h of ischemia at 35°C, the ties were released using a slipknot technique for ready release and rapid reperfusion [16]. This procedure was done in IP groups for 3 times before the prolonged ischemia. Sciatic nerves were harvested at 3 hours and 1 week after

ischemia surgery for the MDA measurements and his-topathological studies, respectively.

**Neuropathology: edema and axonal vacoullisation**

A sciatic nerve segment at 2 cm long was harvested from each animal. The sciatic nerves were osmicated, dehydrated, infiltrated, and embedded in Spurr's resin. Longitudinal sections of 1.0 cm were stained with hematoxylen eozine. Under 40x magnification, these sections were graded for edema and axonal vacoullisation using previously described methods [17]. The axon may be swollen or shrunken, watery and light, or dark and shrunken. Secondary myelin changes were typically seen, including attenuation, collapse, or break-down. For each section, the vacoullisation and edema were semi-quantitatively graded from 0 to 3 as follows: 0-normal, 1-mild, 2-moderate and 3-severe. No distinction was made as to endoneurial, perivascular, or subperineurial edema. A mean value for each rat was calculated after examination of four sections represented that case.

**Statistics**

The values were expressed as mean ± standart of deviation. The differences between the groups were analysed by using ANOVA. Non-parametric data was evaluated by Mann Whithney-U test. A p value less than 0.05 was considered as significant.

**Results**

**MDA levels in sciatic nerve**

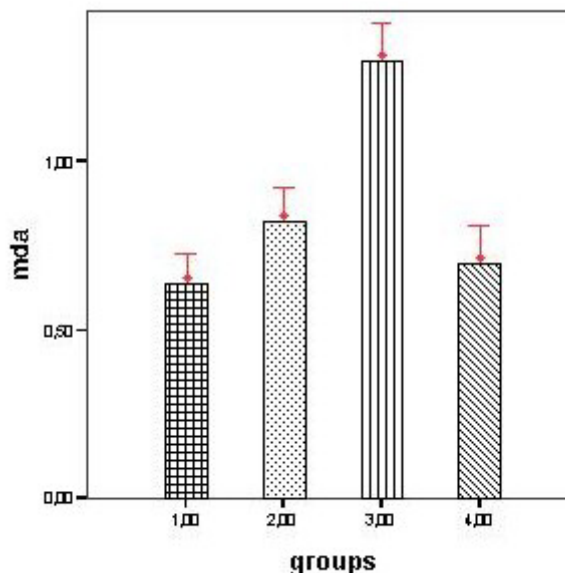
During the occlusion of aorta and iliac arteries, the measurements of femoral blood pressure approximated to zero values in rats of all the groups.

The MDA levels of nerve tissue segments in different groups and nerve vacuollisation degrees is shown in the table 1. Ischemic preconditioning group had significantly lower nerve MDA levels than reperfusion group (p < 0.001). The differences between ischemic, IP and non-ischemic control groups were not significant (p > 0.05), (figure 1).

**Table 1: Non-parametric evaluation scores in part 2 experimentation.**

Parameter	I/R	IP	Sham
Edema	3	2*	0**
Vacuollisation	2	1	0

The scale for the evaluated parameters as follows: normal-0, mild-1, moderate-2, severe-3 for endoneurial edema;no vacuollisation-0, mild vacuollisation-1, massive vacuollisation-2 for axonal vacuollisation. (\* p < 0.05 for I/R versus IP, \*\*p < 0.00001 for I/R versus IP).



**Figure 1**

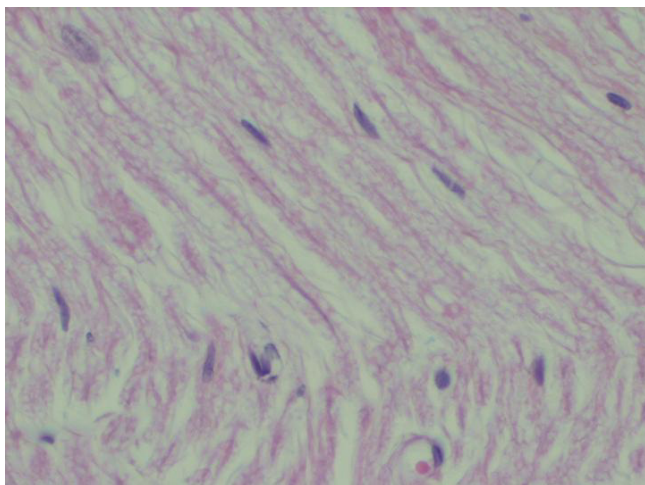
Sciatic nerve MDA (nmol/mg protein) levels in groups. (1.00:non-ischemic controls, 2.00:ischemic preconditioning, 3.00:ischemia-3 h reperfusion, 4.00:ischemia only). Bars show means. Error bars show 95.0% CI of means.

**Histopathologic changes**

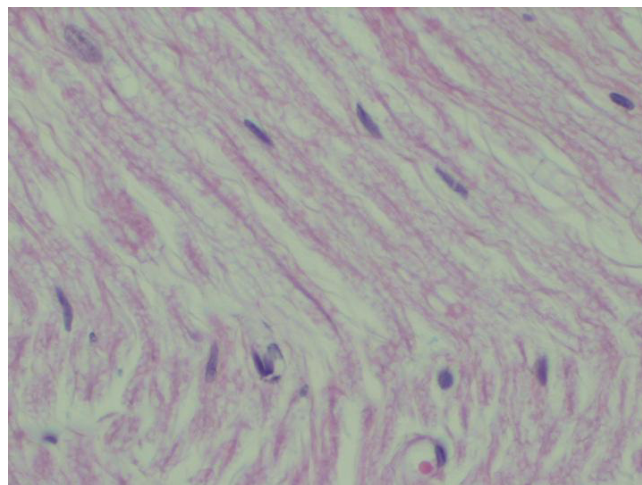
Great cytoplasmic vacuollisation caused by proliferation and dilation of the rough and smooth endoplasmic reticulum and golgi apparatus was observed in I/R and IP groups of part 2 experimentation. The intramyelinic edema within nerve fibers was seen not only in perivascular region, but also, in endoneurial vessels (Figures 2, 3 and 4). IP group had a significantly good histopathologic score than I/R group (p < 0.05). Table 1 shows the scores in the groups.

**Discussion**

Nerve pathology in acute ischemic injury has been delineated in peripheral nerve and reperfusion injury could amplify ischemic pathology. Nerve ischemia plays a major role in the development of pathological alteration in various neuropathies and the effects of ischemia are amplified by reperfusion in various tissues. In nerve tissue, two types of edema is described after I/R; endoneurial edema and intramyelinic edema [17]. Endoneurial edema reflects in blood-nerve barrier and possibly reflects endoneurial events especially severity of IFD. Myelin appears to be particularly susceptible to activated free radicals, activated neutrophils and cytokine formation. Severe ischemia to nerve results in energy rundown followed by conduction failure and fiber degeneration. Inflammatory responses to IR have not only been confined to a few days (up to 7-14

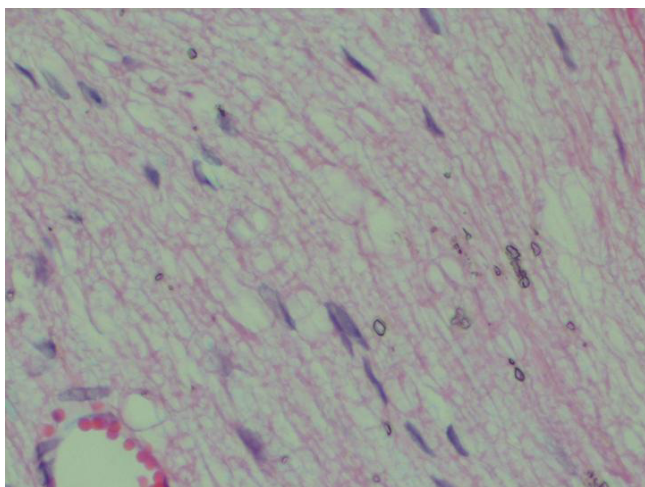


**Figure 2**  
Normal architecture of sciatic nerve of rat is seen (sham group), Hematoxylen esozine 40× magnification.



**Figure 4**  
Mild vacuolisation in axons of sciatic nerve (ischemic preconditioning group, score 2), Hematoxylen esozine 40× magnification.

days) of reperfusion, but also much more extended time (up to 42 days) of reperfusion [18]. Between 7 days and 14 days of reperfusion, the IFD was reported to be the most prominent. Morphological changes of IFD at the light microscopic level occur in concert with endoneurial edema at the 7 and 14 day reperfusion time-points. I/R injury of sciatic nerve has been shown to increase proinflammatory cytokines which are primarily responsible for demyelination after reperfusion in peripheral nerves [19]. Another important indicator of I/R injury of peripheral nerve is Nitric Oxide products which were found to be increased at the first 24 hours of reperfusion in nerve tis-



**Figure 3**  
Increased axonal vacuolisation degeneration is seen at longitudinal section of sciatic nerve (I/R group, score 3), Hematoxylen esozine 40× magnification.

sue and their elevation has been reported to play an essential role in reducing the severity of the I/R injury by inhibiting neutrophil adhesion in postcapillary venules and by decreasing microvascular constriction [20]. In our study, axonal changes at 7th day were evaluated. It has been seen that IP treated group showed less cytoplasmic vacuolisation and edema formation than I/R group ( $p < 0.05$ ). This finding was concomitant to the finding of decreased oxidative injury (i.e. decreased MDA levels in nerve tissue) seen in IP pretreated group.

Previously protective effects of IP in intestine, liver, myocardium, skeletal muscle and pancreas tissues has been shown [10-14,21,22]. What plays a role in the protective effect of IP is not exactly known, but some putative mechanisms, which are mostly dealt with countering the proinflammatory and proapoptotic effects generated during IR have been put forward [23]. IP has been shown to decrease the formation of hydroxyl radicals during reperfusion [24]. A reduced TNF- $\alpha$  and ICAM-1 mRNA expression seen after IP may account for the inhibitory effects of IP on leukocyte adhesion and ameliorated microcirculatory disturbance after IR in vivo [23-26].

The protective effects of IP against lesions caused by subsequent severe ischemia were primarily described in the heart by Murry et al [9]. Increased antioxidant enzyme activities which may be an indirect indicator of the reduced injury after I/R has been shown in brain ischemic tolerance by IP [27]. However to the best of our knowledge, nobody has studied this phenomenon in peripheral somatic nerve. The beneficial effect of IP in rat sciatic nerve was manifested by a reduction in MDA tissue levels at 3th

hour of reperfusion and ischemic fiber degeneration (IFD) at 7 th postischemic day of reperfusion, in the present study. Lida et al. identified pathologically three phases as follows: phase 1-early reperfusion minimal edema; phase-2 7 th and 14 th day of reperfusion prominent fiber degeneration and endoneurial edema; phase-3 28 th and 42 th days abundant small regenerating fiber clusters, minimal edema [18]. Our observation period is limited to up 7th day, i.e. phase-1. To best of our knowledge, this is the first semiquantitative study that shows an decreased IFD after IR due to the pretreatment with IP. Further studies are needed for understanding that IP may have strategic role in treatment of I/R related peripheral nerve injuries.

## References

- Lida H, Scheichel AM, Wang Y, Schmelzer JD, Low PA: **Schwan cell is a target in ischemia-reperfusion injury to peripheral nerve.** *Muscle Nerve* 2004, **30(6)**:761-6.
- Nagamatsu M, Schmelzer JD, Zollman PS, Smithson IL, Nickander KK, Low PA: **Ischemic reperfusion causes lipid peroxidation and fiber degeneration.** *Muscle nerve* 1996, **19(1)**:37-47.
- Nukada H, McMorran PD, Shimizu J: **Acute inflammatory demyelination in reperfusion nerve injury.** *Ann Neurol* 2000, **47(1)**:71-79.
- Nukada H, Anderson GM, McMorran PD: **Reperfusion nerve injury pathology due to reflow and prolonged ischemia.** *J Peripher Nerv Syst* 1997, **2(1)**:60-69.
- Carden DL, Granger DN: **Pathophysiology of ischemia-reperfusion injury.** *J Pathol* 2000, **190(3)**:255-66.
- Nagatsu M, Scelzer JD, Zollman PS, Smithson IL, Nickander KK, Low PA: **Ischemic reperfusion causes lipid peroxidation and fiber degeneration.** *Muscle Nerve* 1996, **19**:37-47.
- Nukada H, Lynch CD, McMorran PD: **Aggravated reperfusion injury in STZ-diabetic nerve.** *J Peripheral Nerv Syst* 2002, **7**:37-43.
- Meghoo CAL, Gonzalez EA, Tyroch AH, Wohltmann CD: **Complete occlusion after blunt injury to the abdominal aorta.** *J Trauma-Injury Infection and Critical Care* 2003, **55**:795-799.
- Murry CE, Jennings RB, Reimer KA: **Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium.** *Circulation* 1986, **74**:1124-1136.
- Mallick IH, Yang W, Winsletand MC, Seifalian AM: **Protective effects of ischemic preconditioning on the intestinal mucosal microcirculation following ischemia-reperfusion of the intestine.** *Microcirculation* 2005, **12**:615-25.
- Lee WYSM: **Ischemic preconditioning protects post-ischemic oxidative damage to mitochondria in rat liver.** *Shock* 2005, **24**:370-5.
- Morihina M, Hasebe N, Erdene B, Sumitomo K, Matsusaka T, Izawa K, Fujino T, Fukuzawa J, Kikuchi K: **Ischemic Preconditioning Enhances Scavenging Activity of Reactive Oxygen Species and Diminishes Transmural Difference of Infarct Size.** *Am J Physiol Heart Circ Physiol* . 2005 Jul 22; [Epub ahead of print]
- Dembinski A, Warzecha Z, Ceranowicz P, Tomaszewska R, Dembinski M, Pabianczyk M, Stachura J, Konturek S: **Ischemic preconditioning reduces the severity of ischemia/reperfusion-induced pancreatitis.** *Eur J Pharmacol* 2003, **473**:207-16.
- Marian CF, Jigo LP, Ionac M: **Ischemic preconditioning of free muscle flaps: An experimental study.** *Microsurgery* 2005, **25**:524-31.
- Low PA, Tuck RR: **Effects of changes of blood pressure, respiratory acidosis and hypoxia on blood flow in the sciatic nerve of the rat.** *J Physiol* 1984, **347**:513-24.
- Wang Y, Schmelzer JD, Scheichel A, Lida H, Low PA: **Ischemia-reperfusion injury of peripheral nerve in experimental diabetic neuropathy.** *J of The Neurological Sciences* 2004, **227**:101-107.
- Nukada H, McMorran PD: **Perivascular demyelination and intramyelinic oedema in reperfusion nerve injury.** *J Anat* 1994, **185**:259-66.
- Lida H, Schmelzer JD, Schmeichel AM, Wang Y, Low PA: **Peripheral nerve ischemia: Reperfusion injury and fiber degeneration.** *Experimental Neurology* 2003, **184**:997-1002.
- Mitsui Y, Okamoto K, Martin DP, Schmelzer JD, Low PA: **The expression of proinflammatory cytokine mRNA in the sciatic-tibial nerve of ischemia-reperfusion injury.** *Brain Res* 1999, **844**:192-95.
- Milcan A, Arslan E, Bagdatlioglu OT, Bagdatlioglu C, Polat G, Kanik A, Talas DU, Kuyurtar F: **The effect of alprostadil on ischemia-reperfusion injury of peripheral nerve in rats.** *Pharmacol Res* 2004, **49**:67-72.
- Barrier A, Olaya N, Chiappini F, Roser F, Scatton O, Artus C, Franc B, Dudoit S, Flahault A: **Ischemic preconditioning modulates the expression of several genes leading to the overproduction of iL-R, iNOS and Bcl-2 in a human model of liver ischemia-reperfusion.** *FASEB J* 2005, **19**:1617-26.
- Raphad J, Drenger B, Rivo J, Brenshtein E, Chvios M, Gozal Y: **Ischemic preconditioning decreases the reperfusion related formation of hydroxyl radicals in a rabbit model of regional myocardial ischemia and reperfusion: the role of K (ATP) channels.** *Free Radical Res* 2005, **39**:747-54.
- Danielisova V, Nemethova M, Gottlieb M, Burada J: **Changes of endogenous antioxidant enzymes during ischemic tolerance acquisition.** *Neurochem Res* 2005, **30**:559-65.
- Zahler S, Kupatt C, Becker BF: **Endothelial preconditioning by transient oxidative stress reduces inflammatory responses of cultured endothelial cells to TNF-alpha.** *FASEB J* 2000, **14**:555-64.
- Hung LM, Wei W, Hsueh YJ, Chu WK, Wei FC: **Ischemic preconditioning ameliorates microcirculatory disturbance through downregulation of TNF-alpha production in a rat cremaster muscle model.** *J Biomed Sci* 2004, **11**:773-80.
- Funaki H, Shimizu K, Harada S, Tsuyama H, Fushida S, Tani T, Miwa K: **Essential role for nuclear factor (kappa) B in ischemic preconditioning for ischemia-reperfusion injury of the Mouse liver.** *Transplantation* 2002, **74**:551-556.
- Danielisova V, Nemethova M, Gottlieb M, Burada J: **Changes of endogenous antioxidant enzymes during ischemic tolerance acquisition.** *Neurochem Res* 2005, **30**:559-65.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

