Toward a Treatment Normalizing Ovulation Rate in Adolescent Girls With Polycystic Ovary Syndrome

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Adolescent polycystic ovary syndrome (PCOS) is characterized by androgen excess and oligomenorrhea, and commonly driven by hepato-visceral fat excess ("central obesity") ensuing from a mismatch between prenatal and postnatal nutrition, on a background of genetic susceptibility. There is no approved treatment for adolescent PCOS.

We report the pooled results of 2 pilot studies in nonobese girls with PCOS (N = 62, age 15.8 years) that compared the effects of randomized treatment for 1 year, either with an oral estro-progestogen contraceptive (OC), or with a low-dose combination of spironolactone-pioglitazone-metformin (SPIOMET, targeting the excess of ectopic fat).

Auxological and endocrine-metabolic variables (including fasting insulin, androgens, highmolecular-weight adiponectin [HMW-adiponectin], and microRNA [miR]-451a), body composition (dual x-ray absorptiometry) and hepato-visceral fat (magnetic resonance imaging) were assessed on- and posttreatment. Data from menstrual diaries were combined with weekly salivary progesterone measurements to infer ovulation rates during the second and fourth quarter of the posttreatment year.

OC and SPIOMET treatment reduced the androgen excess comparably, and had no differential effects on total-body lean or fat mass. However, SPIOMET was accompanied by more broadly normalizing effects, including on hepato-visceral fat and on circulating insulin, HMW-adiponectin, and miR-451a. On average, there were 3-fold more ovulations post-SPIOMET than post-OC; normovulation was only observed after SPIOMET; anovulation was >10-fold more prevalent post-OC.

Pooled results of randomized studies in nonobese adolescent girls with PCOS indicate that SPIOMET treatment leads to an overall healthier, more insulin-sensitive condition—with less ectopic fat—than OC treatment, and to a more normal posttreatment ovulation rate.

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Key Words: PCOS, ovulation, hepatic fat, visceral fat, metformin, spironolactone, pioglitazone

Abbreviations: BMI, body mass index; cIMT, carotid intima-media thickness; CRP, C-reactive protein; HMW, highmolecular-weight; HOMA-IR, homeostatic model assessment-insulin resistance; miR, microRNA; OC, oral contraceptive; MRI, magnetic resonance imaging; PCOS, polycystic ovary syndrome; SEM, standard error of the mean; SPIOMET, spironolactone-pioglitazone-metformin.

There is no approved treatment for polycystic ovary syndrome (PCOS), a prevalent condition in adolescent girls and young women [1, 2]. Many of these patients are guided into a trajectory that starts with oral contraceptive (OC) treatment, leads into oligo-anovulatory subfertility, then into the use of assisted reproductive techniques, and ultimately into pregnancies with a double-to-triple risk for complications (such as gestational diabetes, preeclampsia, and preterm birth) potentially with lifelong sequelae in the offspring [2].

Evidence is converging toward the insight that adolescent PCOS is frequently driven by hepato-visceral fat excess ("central obesity") ensuing from a mismatch between (rather restrictive) prenatal and (rather abundant) postnatal nutrition, on a background of (epi) genetic susceptibility [3, 4, 5]. This insight has prompted the exploration of an alternative treatment for PCOS consisting of the intake of a low-dose combination of spironolactone (a mixed anti-androgen and anti-mineralocorticoid, also activating brown adipose tissue) [6] with pioglitazone and metformin (2 insulin sensitizers acting through different mechanisms) (SPIOMET) for 1 year. This combination proved to have more normalizing effects than OC treatment, in particular, on ectopic fat excess, insulin sensitivity, and posttreatment ovulation rate [7]. The limited power of the first study (N = 34) prompted the launch of a second study with virtually identical design. Here we report the pooled results of both studies in nonobese girls with PCOS (N = 62).

1. Materials and Methods

A. Study Population & Design

Both pilot studies (ISRCTN29234515 and ISRCTN11062950) had an open-label, randomized, controlled design, and were conducted in the Adolescent Endocrinology Unit of Sant Joan de Déu University Hospital, Barcelona, Spain. Recruitment was biased against overweight/obesity because, in our setting, overweight/obese adolescent girls are primarily referred to the Adolescent Obesity Unit rather than to the Adolescent Endocrinology Unit. In each study, the on-treatment year was followed by a posttreatment year. Study completion rate was 89% (62/71) (Fig. 1, flow chart).

The inclusion criteria were hirsutism (score > 8 on modified Ferriman-Gallwey scale), oligomenorrhea (menstrual intervals > 45 days), gynecological age > 2.0 years, and absence of sexual activity (no need for contraception). Exclusion criteria were 21-hydroxylase deficiency; glucose intolerance or diabetes; evidence of thyroid, liver, or kidney dysfunction; hyperprolactinemia; and prior use of medications affecting gonadal/adrenal function, or carbohydrate/lipid metabolism [7, 8]. Mediterranean diet and regular exercise were recommended to all participating girls; OC treatment consisted of 20 μ g ethinylestradiol plus 100 mg levonorgestrel for 21/28 days, and placebo for 7/28 days; SPIOMET treatment consisted of a low-dose combination of spironolactone 50 mg/day, pioglitazone 7.5 mg/day, and metformin 850 mg/day [7].

Age-matched, healthy girls (N = 52; mean age 16.3 years) recruited from nearby schools served as controls. All had regular menstrual cycles, and none was hirsute or taking medication.

The primary endpoint was posttreatment ovulation rate; secondary outcomes included hirsutism score, fasting insulin, androgens, lipids, high-molecular-weight (HMW) adiponectin, C-reactive protein (CRP), carotid intima-media thickness (cIMT), body composition, and hepato-visceral fat [7]; circulating microRNA (miR)-451a could only be measured in a subset of the participating girls (footnote below Table 1).

Blood sampling in both patients and controls was at all time points performed either in the follicular phase of the menstrual cycle (days 3-7) or after 2 months of amenorrhea; at study start, the ratio of amenorrheic to oligomenorrheic girls was 1 to 7.

B. Assessments

Birth weight, birth length, and body mass index (BMI) (and their Z-scores) were retrieved from medical records. Endocrine-metabolic variables and cIMT were assessed as described



Figure 1. Flow chart for Study 1 and Study 2.

[7, 8]. Homeostatic model assessment-insulin resistance (HOMA-IR) was calculated as [fasting insulin in mU/L] x [fasting glucose in mg/dL]/405. Ovulation rates were inferred by combining data from menstrual diaries and from progesterone concentrations assessed in weekly saliva samples, obtained over 12 weeks in the second quarter and then 12 weeks in the fourth quarter of the posttreatment year [7]. Progesterone was measured by enzymelinked immunosorbent assay (ELISA) (Novatec, Inmundiagnostica, cat# DSNOV25, RRID:AB_2827743) [9]. Circulating miR-451a was measured as described [5], with results expressed in Z-scores, using the data of healthy control girls as reference; circulating miR-451a concentrations are known to be low in adolescent girls with PCOS (average Z-scores between -3 and -4), and to associate negatively with the degree of androgen excess (as judged by circulating testosterone or free androgen index, when the gonadotropic axis is not silenced), with HOMA-IR, and with hepatic and visceral fat; a normalizing rise of circulating miR-451a concentrations in adolescent girls with PCOS can thus point to a normalizing course toward metabolic health, including toward a normal ovulation rate [5]. In a search for a noninvasive, cycle-independent, on-treatment set of markers that allows to anticipate the posttreatment ovulation rate, we tested whether a "metabolic health Z-score" which combines the Z-scores of fasting insulinemia and circulating miR-451a, associated to posttreatment ovulation rate.

Body composition was assessed by dual x-ray absorptiometry with a Lunar Prodigy and Lunar software (version 3.4/3.5, Lunar Corp, Madison, Wisconsin); abdominal fat (subcutaneous and visceral) and hepatic fat were assessed by magnetic resonance imaging (MRI) using a multiple-slice MRI 1.5 Tesla scan (Signa LX Echo Speed Plus Excite, General Electric, Milwaukee, Wisconsin), as described [7, 8].

C. Statistics & Ethics

Statistical analyses were performed with SPSS 23.0 (IBM, Armonk, New York). Longitudinal changes in quantitative variables between groups were compared by repeated-measures

s With Polycystic C	Adolescent Girls With Polycystic C	Data From Adolescent Girls With Polycystic C
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	-		Eth	inylestradio	l-Levonorg	estrel (N = ;	31)		SP	IOMET (N=	31)	
	Controls (N = 52)	PCOS (N = 62)	$\mathbf{Start}^{\mathbf{a}}$	12 mo	24 mo	Δ 0–12 mo <i>i</i>	A 12–24 mo	${ m Start}^{ m a}$	12 mo	24 mo	Δ 0–12 mo	A 12–24 mo
Birthweight Z-score Age at Menarche	0.2 ± 0.1 12.4 ± 0.1	$-0.6 \pm 0.1^{***}$ 11.6 $\pm 0.1^{***}$	-0.6 ± 0.2 11.6 ± 0.1		::	: :		-0.6 ± 0.1 11.6 ± 0.2	: :	: :	: :	
(yr) Age (yr)	16.3 ± 0.2	15.8 ± 0.2	15.9 ± 0.2	;	:	1	1	15.7 ± 0.2	1	:	:	1
BMI (kg/m ²)	21.3 ± 0.3	$24.2 \pm 0.5^{***}$	24.2 ± 0.7	$24.9 \pm 0.8^{\circ}$	25.1 ± 0.8	0.7 ± 0.3	0.2 ± 0.3	24.2 ± 0.7	23.9 ± 0.7	23.9 ± 0.7	-0.2 ± 0.3^{e}	0.0 ± 0.2
BMI Z-score	0.0 ± 0.1	$0.8 \pm 0.1^{***}$	0.9 ± 0.2	$1.1 \pm 0.2^{\rm b}$	1.2 ± 0.2	0.2 ± 0.1	0.1 ± 0.3	0.8 ± 0.2	0.7 ± 0.2	0.8 ± 0.2	-0.1 ± 0.1^{e}	0.1 ± 0.3
Δ Z-score Birth- weight to BMI	-0.2 ± 0.2	$1.4 \pm 0.2^{***}$	1.5 ± 0.3	1.7 ± 0.3^{b}	1.8 ± 0.3	0.2 ± 0.1	0.1 ± 0.1	1.4 ± 0.3	1.4 ± 0.3	1.4 ± 0.3	0.0 ± 0.1^{e}	0.0 ± 0.1
Waist Circumfer-	74 ± 1	77 ± 1	76 ± 2	$78 \pm 2^{\rm b}$	78 ± 2	2 ± 1	0 ± 1	77 ± 2	$74 \pm 1^{\rm d}$	74 ± 1	-3 ± 0.8^{g}	0 ± 1
ence (cm)		-	-	- - -							5 - 1	
Hirsutism score SHBG (nmol/L)	 63±3	17 ± 1 $30 \pm 2^{***}$	17 ± 1 31 ± 2	$14 \pm 1^{\circ}$ $61 \pm 5^{ m d}$	14 ± 1 32 ± 3^{d}	-3±1 30±4	0 ± 1 -29 ± 5	16 ± 1 30 ± 2	$11 \pm 1^{\circ}$ 32 ± 2	9 ± 1° 39 ± 3°	$-5 \pm 1^{\circ}$ 2 ± 2^{g}	-2 ± 1 7 ± 2^{g}
Testosterone	0.7 ± 0.1	$1.4\pm0.1^{***}$	1.3 ± 0.1	$0.7 \pm 0.1^{\mathrm{d}}$	$1.6\pm0.2^{ m d}$	-0.6 ± 0.1	0.9 ± 0.2	1.5 ± 0.2	$0.8\pm0.1^{\circ}$	1.2 ± 0.2^{c}	-0.7 ± 0.2	0.4 ± 0.2
(nmol/L)		:										
Androstenedione (nmol/L)	3.5 ± 0.2	$5.3 \pm 0.3^{***}$	4.8 ± 0.3	2.5 ± 0.2^{d}	5.7 ± 0.6^{d}	-2.3 ± 0.3	3.2 ± 0.5	5.7 ± 0.4	3.5 ± 0.3^{d}	$5.3 \pm 0.6^{\circ}$	-2.2 ± 0.4	1.8 ± 0.6
Free Testosterone	0.0 ± 0.2	$2.9 \pm 0.5^{***}$	2.3 ± 0.5	$0.3 \pm 0.3^{\circ}$	$3.6 \pm 0.8^{\rm d}$	-2.0 ± 0.6	3.3 ± 0.7	3.2 ± 0.9	$0.5 \pm 0.3^{\circ}$	2.0 ± 0.7^{c}	-2.7 ± 0.9	1.5 ± 0.7
Z-score Free Androstene-	0.0 ± 0.2	$1.8 \pm 0.3^{***}$	1.1 ± 0.3	-0.9 ± 0.2^{d}	$2.2\pm0.5^{ m d}$	-2.0 ± 0.3	3.1 ± 0.5	2.2 ± 0.4	0.1 ± 0.3^{d}	$1.8\pm0.6^{\circ}$	-2.1 ± 0.4	1.7 ± 0.6
dione Z-score												
Fasting Insulin (nmol/L/)	49 ± 7	$76 \pm 7^{***}$	83 ± 7	$104 \pm 7^{\rm b}$	76 ± 7^{c}	21 ± 7	-28±7	69 ± 7	$42 \pm 7^{\rm d}$	49 ± 7	-27 ± 7^g	$7 \pm 7^{\rm f}$
HOMA-IR	1.5 ± 0.1	$2.3 \pm 0.2^{***}$	2.6 ± 0.3	3.0 ± 0.3	$2.2 \pm 0.2^{\circ}$	0.4 ± 0.2	-0.8 ± 0.3	2.1 ± 0.2	$1.2 \pm 0.1^{\mathrm{d}}$	1.3 ± 0.2	$-0.9 \pm 0.3^{\rm f}$	$0.1\pm0.2^{\rm f}$
OGTT Mean Gly- cemia Z-score	:	0.2 ± 0.1	0.1 ± 0.1	0.2 ± 0.1	0.1 ± 0.1	0.1 ± 0.1	-0.1 ± 0.1	0.2 ± 0.1	$0.1 \pm 0.1^{\circ}$	0.1 ± 0.1	$-0.1 \pm 0.1^{\rm f}$	0.0 ± 0.1
Mean Insulinemia 7.score	ł	3.2 ± 0.3	3.5 ± 0.4	3.7 ± 0.5	3.1 ± 0.5	0.2 ± 0.5	-0.6 ± 0.5	2.8 ± 0.4	0.6 ± 0.2^{d}	0.6 ± 0.2	-2.2 ± 0.3^{g}	0.0 ± 0.2
ALT (µkat/L)	0.30 ± 0.02	$0.23 \pm 0.02^{***}$	0.23 ± 0.02	$0.32 \pm 0.03^{\circ}$	0.27 ± 0.02	0.09 ± 0.02	-0.05 ± 0.03	0.23 ± 0.02	0.23 ± 0.02	0.23 ± 0.02	$-0.00 \pm 0.02^{\circ}$	-0.00 ± 0.02
GGT (µkat/L)	0.22 ± 0.02	0.21 ± 0.02 0.22 ± 0.02	0.21 ± 0.02 0.22 ± 0.02	0.21 ± 0.02^{d} 0.30 ± 0.02^{d}	0.25 ± 0.02^{b}	0.00 ± 0.02 0.08 ± 0.02	0.00 ± 0.02 - 0.05 ± 0.02	0.22 ± 0.02	0.21 ± 0.02 0.18 ± 0.02	$0.22 \pm 0.02^{\circ}$	-0.01 ± 0.02^{g} -0.04 ± 0.02^{g}	0.04 ± 0.02 0.04 ± 0.02
Triacylglycerol (mmol/L)	0.60 ± 0.03	0.68 ± 0.03	0.66 ± 0.03	0.75 ± 0.05^{b}	$0.64 \pm 0.03^{\circ}$	0.09 ± 0.03	-0.11 ± 0.03	0.70 ± 0.05	0.67 ± 0.05	0.63 ± 0.05	-0.03 ± 0.05 ^e	-0.04 ± 0.03

Table 1. Continued

			Eth	inylestradic	ol-Levonorg	jestrel (N =	31)		$^{\mathrm{SP}}$	IOMET (N=	31)	
	(N = 52)	$\Gamma = 62$)	$\operatorname{Start}^{\operatorname{a}}$	12 mo	24 mo	Δ 0–12 mo	A 12–24 mo	$\mathbf{Start}^{\mathrm{a}}$	12 mo	24 mo	Δ 0–12 mo	Δ 12–24 mo
LDL-cholesterol	2.2 ± 0.1	2.3 ± 0.1	2.3 ± 0.1	2.7 ± 0.1^{d}	$2.2 \pm 0.1^{\mathrm{d}}$	0.4 ± 0.1	-0.5 ± 0.1	2.2 ± 0.1	2.2 ± 0.1	2.0 ± 0.1^{d}	$0.0 \pm 0.1^{\mathrm{f}}$	-0.2 ± 0.1^{e}
HDL-cholesterol	1.4 ± 0.1	1.3 ± 0.1	1.3 ± 0.1	1.3 ± 0.1	1.4 ± 0.1	0.0 ± 0.1	0.1 ± 0.1	1.3 ± 0.1	$1.4 \pm 0.1^{\circ}$	$1.3\pm0.1^{\mathrm{b}}$	$0.1 \pm 0.1^{\mathrm{e}}$	$-0.1 \pm 0.1^{\mathrm{f}}$
(mmol/L) HMW-adiponectin	9.3 ± 0.8	$6.8\pm0.6^*$	6.5 ± 0.6	$8.9 \pm 1.3^{\mathrm{b}}$	8.6 ± 0.8	2.6 ± 1.1	-0.3 ± 1.5	7.1 ± 0.9	$17.1 \pm 2.6^{\mathrm{d}}$	$10.3 \pm 1.5^{\circ}$	$10.0 \pm 2.1^{\mathrm{f}}$	$-7 \pm 2^{\mathrm{e}}$
(mg/L) C-Reactive Protein (nmol/L)	6.7 ± 0.9	$14.3 \pm 1.9^{***}$	11.4 ± 1.9	$24.8\pm3.8^{\circ}$	18.1 ± 3.8	13.4 ± 3.8	-6.7 ± 5.7	17.1 ± 3.8	$6.7 \pm 0.9^{\circ}$	6.7 ± 0.9	-10.4 ± 3.8^{g}	0.0 ± 0.9
Carotid IMT (mm) Systolic Rlood Pres-	 113 + 1	$.37 \pm .00$	$.37 \pm .01$ 113 + 2	$.37 \pm .01$	$.36 \pm .01^{\rm b}$ 112 + 2	$00 \pm .00$	$01 \pm .01$ 3 + 2	$.37 \pm 0.01$	$.35 \pm 0.01^{d}$ 112 + 1 ^b	$.35 \pm 0.01$	02 ± 0.01^{e} 4 + 2^{e}	$.00 \pm .01$ 2 + 2
sure (mmHg) Diastolic Blood	70 ± 1	72 ± 1	71 ± 1	74 ± 1^{b}	73 ± 1	00 1 1 1 1 1	- - - - - - - - - - - - - - - - - - -	73 ± 1	71 ± 1	70 ± 1	$-2 \pm 1^{\circ}$	-1+1
Pressure (mmHg) miR-451a Z-score) 0.00 ± 0.28 ·	$-3.57 \pm 0.11^{***}$	-3.75 ± 0.12	-3.31 ± 0.12	-3.59 ± 0.16	:	ł	-3.32 ± 0.19	0.37 ± 0.31 .	$-1.05 \pm 0.43^{\circ}$	I	1
DXA BMD (g/cm^2)	: :	1.19 ± 0.01 35 6 + 0 6	1.18 ± 0.02 35.7 ± 0.8	1.19 ± 0.02 36.4 ± 0.0	$1.20 \pm 0.02^{\rm b}$ 36.5 ± 0.0	0.01 ± 0.01	0.01 ± 0.01	1.19 ± 0.02 $35 5 \pm 0.0$	1.19 ± 0.02 35.6 ± 0.8	$1.21 \pm 0.02^{\rm b}$ 36.1 ± 0.8	0.00 ± 0.01	0.02 ± 0.01
Fat Mass (Kg)	:	22.1 ± 1.0	21.8 ± 1.4	$23.2 \pm 1.5^{\circ}$	23.4 ± 1.6	1.4 ± 0.5	0.2 ± 0.6	22.4 ± 1.6	22.5 ± 1.4	22.1 ± 1.7	0.1 ± 0.8	-0.4 ± 0.6
Abd MRI Subc Fat	94 ± 9	$174 \pm 14^{***}$	169 ± 18	184 ± 19	180 ± 20	15 ± 9	-4 ± 13	179 ± 21	171 ± 19	167 ± 23	-8±11	-4 ± 9
(cm ²) Visceral Fat (cm ²) Liver Fat (%)	28 ± 1 10 ± 1	$43 \pm 2^{***}$ $17 \pm 1^{***}$	41 ± 3 17 ± 1	45 ± 4 19 ± 1	39 ± 3 17 ± 1^{b}	$\begin{array}{c} 4\pm3\\ 2\pm1\end{array}$	-6 ± 3 -2 ± 2	$\begin{array}{c} 44\pm3\\ 18\pm1\end{array}$	$35 \pm 2^{\mathrm{b}}$ $10 \pm 1^{\mathrm{d}}$	36 ± 3 10 ± 1	$-9 \pm 4^{\mathrm{f}}$ -8 $\pm 1^{\mathrm{g}}$	$\begin{array}{c} 1\pm 2\\ 0\pm 1\end{array}$
Values are mean \pm ; Abbreviations: Abd molecular-weight a tolerance test; SHB #miR-451a (controls " on significant diffe $P < 0.05$, $^{e}P \leq 0.01$; $P < 0.05$, $^{*}P \leq 0.01$, $*P < 0.05$, $^{**}P \leq 0.01$	SEM. MRI, abdomi diponectin; H G, sex hormo G, sex hormo i, n = 13; OC ε rences betwee and ^d P \leq 0.001 be and ^{**} P \leq 0.	nal magnetic r IOMA-IR, hom ne-binding glo at start, n = 12 en randomized 1 within subgr tween subgrou 001 between a	esonance ims teostasis moc bulin; ; SPIOMET s subgroups a oups for 0-12 n tps for 0-12 n the for st st	aging; BMD, ł lel assessmer at start, n = 9 t start 2 mo & 12-24 ch tart and conti	one mineral it - insulin r ; OC at 12 m mo change (<i>i</i> ange (Δ) col group	density; BM esistance; IN io, n = 25; SF	I, body mass AT, intima-m TOMET at 1:	index; DXA, c edia thicknes 2 mo, $n = 24$;	dual x-ray ab ss; miR-451a OC at 24 mo	sorptiometry , microRNA- , n = 15; SPIG	; HMW adipo 451a; OGTT, OMET at 24 n	aectin, high- oral glucose 10, n = 16)

general linear model. Differences in longitudinal changes between groups were tested by the interaction term among between- and within-subject effects. P < 0.05 was considered significant. Data are presented as mean ± standard error of the mean (SEM).

The studies were conducted after approval by the Institutional Review Board of Sant Joan de Déu Hospital, after written informed consent by the parents, and after assent by each participating girl.

2. Results

Table 1 summarizes the pooled results, which indicate that SPIOMET treatment was accompanied by more broadly normalizing effects than OC, including for waist circumference, circulating insulin, HMW-adiponectin and CRP, cIMT, as well as on visceral and hepatic fat (Fig. 2).

Table 2 shows that there were a mean 3-fold and a median 5-fold more ovulations after SPIOMET than OC; normovulation (as judged by 5 or 6 ovulations over 24 weeks) was only observed after SPIOMET; anovulation (as judged by 0 or 1 ovulation over 24 weeks) was > 10-fold more frequent after OC. Menstrual regularity after SPIOMET (90%) was only 2-fold more prevalent than after OC (42%), thus underestimated the difference in ovulation rates.

Fig. 3 illustrates that the randomized treatments led to marked differences in on-treatment metabolic health (as judged by combined Z-scores of fasting insulin and miR-451a) and in posttreatment ovulation rate, both of which were more normalized after SPIOMET.

3. Discussion

Pooled data corroborated SPIOMET as a combination treatment that is accompanied by more normalization of the endocrine-metabolic status, and is followed by markedly more ovulations than OC in nonobese adolescent girls with PCOS. The consistency of the ovulation rates across the posttreatment year suggests that the lower ovulation rates after OC are attributable to persistence of the underpinning PCOS pathophysiology rather than to residual inhibition of the gonadotropic axis. In healthy young women, ovulatory function is known to recover within 3 months after stopping OC treatment [10, 11].

Ectopic adiposity and insulin resistance failed to improve during standard treatment with OC. In contrast, SPIOMET treatment was accompanied by a loss of hepato-visceral fat excess and by a normalization of insulin sensitivity (as judged by HOMA-IR, and by the insulin response to an oral glucose load), both of which were maintained during the



Figure 2. Hepatic fat content (by magnetic resonance imaging) in nonobese adolescent girls with PCOS who were randomized to receive either an oral contraceptive (OC; N = 31; red circles) for 12 months, or a low-dose combination of spironolactone-pioglitazone-metformin (SPIOMET; N = 31; blue circles) for 12 months; subsequently, both subgroups were untreated for 12 months. Body weight did not change in either subgroup. The dotted line indicates the average level in healthy control girls of similar age. Results are expressed as mean \pm SEM. P < 0.0001 for on-treatment change between subgroups.

Table 2. Posttreatment Ovulation Results in Adolescent Girls With Polycystic Ovary Syndrome Who Were Randomized to Receive an Oral Contraceptive (OC) or Low-Dose Spironolactone + Pioglitazone + Metformin (SPIOMET) for 12 Months, and Were Subsequently Followed for 12 Months Without Treatment. Ovulations Were Assessed Twice Over 12 Weeks, for a Total of 24 Weeks: Between the Study Timepoints of 15 to 18 months (posttreatment months 3-6) and 21 to 24 months (posttreatment months 9-12)

		OC N = 31		S	SPIOMET N = 31	
	15-18 mo (12 wk)	21-24 mo (12 wk)	Total (24 wk)	15-18 mo (12 wk)	21-24 mo (12 wk)	Total (24 wk)
Mean number of ovulations ± SEM	0.8 ± 0.1	0.8 ± 0.1	1.6 ± 0.2	$2.3 \pm 0.2^{\#}$	$2.2 \pm 0.2^{\#}$	$4.5 \pm 0.3^{\#}$
Median number of ovulations (interquartile range)	1 (0-1)	1 (0-1)	1 (1-3)	$3(2-3)^{\#}$	$2(2-3)^{\#}$	$5(3-6)^{\#}$
Normo-ovulatory fraction (%) 5 or 6 ovulations /24 wk			0			$62^{\#}$
Oligo-ovulatory fraction (%)			47			35
An-ovulatory fraction (%) 0 or 1 ovulation /24 wk			53			3#

 $^{\#}P < 0.0001$ between subgroups



Figure 3. Randomized treatment of adolescent girls with PCOS, either with an oral contraceptive (OC) or with a low-dose combination of spironolactone-pioglitazone-metformin (SPIOMET) for 12 months, results in an on-treatment difference of metabolic health (N = 22 *vs* 24) and in a posttreatment difference of ovulation rate (N = 30 *vs* 29), so that this combined metabolic-reproductive outcome is markedly to the advantage of SPIOMET. Metabolic health Z-score was calculated by subtracting the Z-score of fasting insulin from the Z-score of circulating miR-451a after 12 months on treatment. Posttreatment number of ovulations over 6 months was inferred by combining data from menstrual diaries and weekly progesterone measurements in saliva over 12 + 12 weeks, between posttreatment months 3 to 6 and 9 to 12. Body weight did not change in either subgroup. The breadth and height of the boxes represent the ranges from -1 SD to +1 SD, respectively, for metabolic health Z-score and ovulation number. *** P < 0.0001.

posttreatment year, via mechanisms that remain to be identified. The downward normalization of liver fat on SPIOMET may partly relate to the upward normalization of circulating miR-451a, which reduces the expression of thyroid hormone responsive spot 14 (*THRSP*), the key gene driving liver steatosis [12, 13].

The present findings corroborate the concept that insulin resistance reflects ectopic lipid accumulation, particularly in the liver, and that it precedes the development of disorders such as type 2 diabetes and nonalcoholic fatty liver disease [14]. Increased hepatic fat and insulin resistance are prevalent findings in both nonobese and obese adolescents with PCOS,

and seem to relate to the underpinning PCOS pathophysiology rather than to testosterone concentrations [15, 16]. Targeting a reduction in androgen levels may thus not be the best choice to normalize the entire PCOS phenotype and to address subsequent comorbidities. The diverging effects of OC and SPIOMET on insulin resistance and ectopic fat (Fig. 3) may herald diverging influences on subsequent risk for PCOS-associated disorders such as an-ovulatory subfertility, gestational diabetes, and/or type 2 diabetes.

The present results remain to be further confirmed in larger and more diverse PCOS populations, including in girls with obesity, with different ethnic and developmental backgrounds, and with other environmental exposures. In addition, SPIOMET's capacity to reduce an excess of liver fat while total body weight remains virtually unchanged (Fig. 2), remains to be tested beyond PCOS settings, in older age ranges, and in a cascade of fatty liver diseases, including nonalcoholic steatohepatitis.

In conclusion, pooled results in nonobese adolescent girls with PCOS confirmed SPIOMET as a treatment that attenuates insulin resistance, reduces ectopic adiposity, and is followed by a more normal ovulation rate than OC.

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L.I. had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Additional Information

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References

^{1.} Witchel SF, Oberfield SE, Peña AS. Polycystic ovary syndrome: pathophysiology, presentation, and treatment with emphasis on adolescent girls. *J Endocr Soc.* 2019;**3**(8):1545-1573.

^{2.} Ibáñez L, Oberfield SE, Witchel S, Auchus RJ, Chang RJ, Codner E, Dabadghao P, Darendeliler F, Elbarbary NS, Gambineri A, Garcia Rudaz C, Hoeger KM, López-Bermejo A, Ong K, Peña AS, Reinehr T, Santoro N, Tena-Sempere M, Tao R, Yildiz BO, Alkhayyat H, Deeb A, Joel D, Horikawa R, de Zegher F, Lee PA. An International Consortium Update: pathophysiology, diagnosis, and treatment of polycystic ovarian syndrome in adolescence. *Horm Res Paediatr*. 2017;88(6):371-395.

de Zegher F, Reinehr T, Malpique R, Darendeliler F, López-Bermejo A, Ibáñez L. Reduced prenatal weight gain and/or augmented postnatal weight gain precede polycystic ovary syndrome in adolescent girls. *Obesity*. 2017;25(9):1486-1489.

- de Zegher F, López-Bermejo A, Ibáñez L. Central obesity, faster maturation, and 'PCOS' in girls. Trends Endocrinol Metab. 2018;29(12):815-818.
- Díaz M, Bassols J, López-Bermejo A, de Zegher F, Ibáñez L. Low circulating levels of miR-451a in girls with polycystic ovary syndrome: different effects of randomized treatments. J Clin Endocrinol Metab. 2019 Nov 15;pii:dgz204.
- Thuzar M, Law WP, Dimeski G, et al. Mineralocorticoid antagonism enhances brown adipose tissue function in humans: A randomized placebo-controlled cross-over study. *Diabetes Obes Metab* 2019;21:509-516.
- 7. Ibáñez L, del Río L, Díaz M, et al. Normalizing ovulation rate by preferential reduction of hepatovisceral fat in adolescent girls with polycystic ovary syndrome. J Adolesc Health. 2017;61(4):446-453.
- Malpique R, Sánchez-Infantes D, Garcia-Beltran C, et al. Towards a circulating marker of hepatovisceral fat excess: S100A4 in adolescent girls with polycystic ovary syndrome - Evidence from randomized clinical trials. *Pediatr Obes* 2019;14(5):e12500.
- 9. Progesterone saliva ELISA kit; Novatec, Inmundiagnostica, cat# DSNOV25, RRID:AB_2827743, https://antibodyregistry.org/search.php?q=AB_2827743.
- Rice-Wray E, Correu S, Gorodovsky J, Esquivel J, Goldzieher JW. Return of ovulation after discontinuance of oral contraceptives. *Fertil Steril* 1967;18(2):212-218.
- Wiegratz I, Mittmann K, Dietrich H, Zimmermann T, Kuhl H. Fertility after discontinuation of treatment with an oral contraceptive containing 30 microg of ethinyl estradiol and 2 mg of dienogest. *Fertil Steril* 2006;85(6):1812-1819.
- 12. Chella Krishnan K, Kurt Z, Barrere-Cain R, et al. Integration of multi-omics data from mouse diversity panel highlights mitochondrial dysfunction in non-alcoholic fatty liver disease. *Cell Syst.* 2018;6(1):103-115.e7.
- 13. Zeng N, Huang R, Li N, et al. MiR-451a attenuates free fatty acids-mediated hepatocyte steatosis by targeting the thyroid hormone responsive spot 14 gene. *Mol Cell Endocrinol*. 2018;474:260-271.
- Samuel VT, Shulman GI. Nonalcoholic fatty liver disease, insulin resistance, and ceramides. N Engl J Med 2019;381(19):1866-1869.
- Cree-Green M, Bergman BC, Coe GV, et al. Hepatic steatosis is common in adolescents with obesity and PCOS and relates to de novo lipogenesis but insulin resistance. *Obesity (Silver Spring)*. 2016;24(11):2399-2406.
- Cree-Green M, Rahat H, Newcomer BR, et al. Insulin resistance, hyperinsulinemia, and mitochondria dysfunction in nonobese girls with polycystic ovarian syndrome. J Endocr Soc. 2017;1(7):931-944.