



Whole-Genome Sequences of 26 Vibrio cholerae Isolates

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The human pathogen *Vibrio cholerae* employs several adaptive mechanisms for environmental persistence, including natural transformation and type VI secretion, creating a reservoir for the spread of disease. Here, we report whole-genome sequences of 26 diverse *V. cholerae* isolates, significantly increasing the sequence diversity of publicly available *V. cholerae* genomes.

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V*ibrio cholerae* is a Gram-negative, facultative anaerobe commonly found in salt water and can cause the fatal diarrheal disease cholera. *V. cholerae* is spread predominantly by fecal contamination of water and food sources in both endemic and epidemic regions (1). Governmental and nongovernmental labs worldwide collect *V. cholerae* isolates for standard bacterial surveillance, and many clinical isolates of *V. cholerae* have already been sequenced (2–4). Clinical samples are often isolated from limited geographic regions and clonally derived with few or no genetic differences (5-8). An expanded characterization of genomes from environmental isolates of *V. cholerae*, which tend to be far more genetically and phenotypically diverse (9), should substantially increase the available sequence diversity of this important human pathogen.

Vibrio spp. are known to encode type VI secretion systems (T6SS), which are often described as bacterial weapons designed

 TABLE 1
 List of V. cholerae strains sequenced in this study

Strain ^a	Location	Source	Yr of isolation	Type VI killing activity ^b	NCBI accession no.
1496-86	United States (LA)	Moore swab	1986	_	MIPC00000000
2523-87	United States (LA)	Moore swab	1974	+	MIPE00000000
VC48	United States (FL)	Oyster	1981	+	MIOT0000000
2633-78	Brazil	Sewage	1978	+	MIPH0000000
857	Bangladesh	Water	1996	+	MIKH0000000
3272-78	United States (MD)	Water	1977	+	MIOZ0000000
TP	United States (CA)	Water	2000	+	MIPK0000000
2559-78	United States (LA)	Crab	1978	+	MIOW0000000
HE46	Haiti (center)	Gray water	2011	+	MIPM0000000
2479-86	United States (LA)	Moore swab	1986	+	MIPB0000000
2497-86	United States (LA)	Moore swab	1987	+	MIPD0000000
2512-86	United States (LA)	Moore swab	1986	+	MIOY0000000
2631-78	United States (LA)	Moore swab	1978	+	MIOX0000000
VC22	United States (FL)	Oyster	1981	+	MIKK0000000
VC53	United States (AL)	Oyster	2009	+	MIOU0000000
VC56	United States (AL)	Oyster	2009	+	MIOV0000000
3568-07	Mexico	Queso fresco	2007	+	MIPL00000000
1074-78	Brazil	Sewage	1978	+	MIPG0000000
3223-74	Guam	Storm drain	1974	+	MIZG0000000
3225-74	Guam	Storm drain	1974	+	MIPF0000000
2740-80	United States (Gulf Coast)	Water	1980	+	MIKI0000000
692-79	United States (LA)	Water	1979	+	MIPA0000000
SIO	United States (CA)	Water	2000	+	MIPJ0000000
C6706	Peru	Patient	1991	_	MIPI0000000
MZO-2	Bangladesh	Patient	2001	_	MIKJ0000000
V52	Sudan	Patient	1968	+	MIPN0000000

^a Strains were isolated from an environmental source, except strains C6706, MZO-2, and V52.

^{*b*} Presence (+) or absence (-) of constitutive type VI killing activity.

to pierce the membranes of adjacent cells and deliver toxic effectors that can lead to lysis of target (prey) cells. In a recent survey, Bernardy et al. (10) noted key differences within a diverse set of isolates for several phenotypes, including chitinase production, contact-dependent killing indicative of T6SS activity, and natural transformation, which can promote horizontal gene transfer. Both clinical and environmental isolates were rarely naturally transformable. In contrast, the majority of environmental, but not clinical, isolates constitutively killed *Escherichia coli* prey. Because different regulatory schemes control the phenotypes tested (11, 12), we sought to better understand the genetics that underlie these diverse *V. cholerae* phenotypes by characterizing wholegenome sequences of 23 environmental and three clinical isolates from Bernardy et al.

All strains were grown overnight in LB medium (Difco) at 37°C, with shaking. Genomic DNA was isolated using a ZR fungal/ bacterial DNA mini prep kit (Zymo Research), and paired-end fragment libraries were constructed using a Nextera XT DNA library preparation kit (Illumina) with a fragment length of 300 bp. Libraries were sequenced by the High Throughput Sequencing Core at Georgia Institute of Technology on an Illumina HiSeq 2500 Rapid platform, producing approximately 280 million 100-bp reads in total. Reads were trimmed using Trimmomatic (13) to remove adapters and bases with a read quality score of <20. Genomes were assembled using SPAdes version 3.5 (14) and annotated using the Rapid Annotation and Subsystem Technology (RAST) web tool provided by the National Microbial Pathogen Data Resource (15-18). T6SS genes were annotated using T6SS Predictor (A. T. Chande et al., unpublished data).

T6SS loci were annotated in all genomes in an effort to characterize the genetic basis of T6SS-mediated killing among diverse environmental *V. cholerae* isolates. All genomes were found to encode the previously characterized large cluster and two auxiliary clusters, which together comprise the canonical T6SS loci. In addition, two previously unreported T6SS loci were discovered in six of the isolates. Numerous examples of novel effector-immunity protein pairs, which function together to catalyze T6SS-mediated killing, were characterized among the set of environmental isolate genomes. Taken together, our genome analysis illuminates the diverse repertoire of genetic mechanisms that underlie T6SSmediated killing in *V. cholerae*.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession numbers listed in Table 1.

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REFERENCES

- 1. Finkelstein RA. 1996. Cholera, *Vibrio cholerae* O1 and O139, and other pathogenic vibrios, chap. 24. *In* Baron S (ed), Medical microbiology, 4th ed. University of Texas Medical Branch at Galveston, Galveston, TX.
- CDC. 2008. Summary of human Vibrio cases reported to CDC, 2008. http: //www.cdc.gov/nationalsurveillance/pdfs/jackson_vibrio_cste2008 _final .pdf.
- Benson DA, Karsch-Mizrachi I, Lipman DJ, Ostell J, Wheeler DL. 2005. GenBank. Nucleic Acids Res 33:D34–D38. http://dx.doi.org/10.1093/nar/ gki063.
- Pruitt KD, Tatusova T, Maglott DR. 2007. NCBI reference sequences (RefSeq): a curated non-redundant sequence database of genomes, transcripts and proteins. Nucleic Acids Res 35:D61–D65. http://dx.doi.org/ 10.1093/nar/gkl842.
- Chatterjee S, Ghosh K, Raychoudhuri A, Chowdhury G, Bhattacharya MK, Mukhopadhyay AK, Ramamurthy T, Bhattacharya SK, Klose KE, Nandy RK. 2009. Incidence, virulence factors, and clonality among clinical strains of non-O1, non-O139 *Vibrio cholerae* isolates from hospitalized diarrheal patients in Kolkata, India. J Clin Microbiol 47:1087–1095. http://dx.doi.org/10.1128/JCM.02026-08.
- Horwood PF, Collins D, Jonduo MH, Rosewell A, Dutta SR, Dagina R, Ropa B, Siba PM, Greenhill AR. 2011. Clonal origins of *Vibrio cholerae* O1 El Tor strains, Papua New Guinea, 2009–2011. Emerg Infect Dis 17: 2063–2065. http://dx.doi.org/10.3201/eid1711.110782.
- Ali A, Chen Y, Johnson JA, Redden E, Mayette Y, Rashid MH, Stine OC, Morris JG. 2011. Recent clonal origin of cholera in Haiti. Emerg Infect Dis 17:699–701. http://dx.doi.org/10.3201/eid1704.101973.
- Bakhshi B, Mahmoudi-Aznaveh A, Salimi-Khorashad A. 2015. Clonal dissemination of a single *Vibrio cholerae* O1 Biotype El Tor strain in Sistan-Baluchestan province of Iran during 2013. Curr Microbiol 71: 163–169. http://dx.doi.org/10.1007/s00284-015-0806-x.
- Chun J, Grim CJ, Hasan NA, Lee JH, Choi SY, Haley BJ, Taviani E, Jeon YS, Kim DW, Lee JH, Brettin TS, Bruce DC, Challacombe JF, Detter JC, Han CS, Munk AC, Chertkov O, Meincke L, Saunders E, Walters RA, Huq A, Nair GB, Colwell RR. 2009. Comparative genomics reveals mechanism for short-term and long-term clonal transitions in pandemic *Vibrio cholerae*. Proc Natl Acad Sci U S A 106:15442–15447. http://dx.doi.org/ 10.1073/pnas.0907787106.
- Bernardy EE, Turnsek MA, Wilson SK, Tarr CL, Hammer BK. 2016. Diversity of clinical and environmental isolates of *Vibrio cholerae* in natural transformation and contact-dependent bacterial killing indicative of type VI secretion system activity. Appl Environ Microbiol 82:2833–2842. http://dx.doi.org/10.1128/AEM.00351-16.
- Watve SS, Thomas J, Hammer BK. 2015. CytR is a global positive regulator of competence, type VI secretion, and chitinases in *Vibrio cholerae*. PLoS One 10:e0138834. http://dx.doi.org/10.1371/journal.pone.0138834.
- Metzger LC, Stutzmann S, Scrignari T, Van der Henst C, Matthey N, Blokesch M. 2016. Independent regulation of type VI secretion in *Vibrio cholerae* by TfoX and TfoY. Cell Rep 15:951–958. http://dx.doi.org/ 10.1016/j.celrep.2016.03.092.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. http:// dx.doi.org/10.1093/bioinformatics/btu170.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to singlecell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/ cmb.2012.0021.
- 15. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/ 1471-2164-9-75.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F,

Stevens R. 2014. The SEED and the Rapid annotation of microbial genomes using subsystems technology (RAST). Nucleic Acids Res 42: D206–D214. http://dx.doi.org/10.1093/nar/gkt1226.

17. Wattam AR, Abraham D, Dalay O, Disz TL, Driscoll T, Gabbard JL, Gillespie JJ, Gough R, Hix D, Kenyon R, Machi D, Mao C, Nordberg EK, Olson R, Overbeek R, Pusch GD, Shukla M, Schulman J, Stevens RL, Sullivan DE, Vonstein V, Warren A, Will R, Wilson MJ, Yoo HS, Zhang C, Zhang Y, Sobral BW. 2014. PATRIC, the bacterial bioinformatics database and analysis resource. Nucleic Acids Res 42:D581–D591. http://dx.doi.org/10.1093/nar/gkt1099.

 Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. http://dx.doi.org/10.1038/srep08365.