

Influence of seasonality on the presence of okadaic acid associated with *Dinophysis* species: A four-year study in Sardinia (Italy)

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Abstract

In Sardinia (Italy), bivalve molluscs production plays an important role in the trade balance. Diarrhoeic shellfish poisoning (DSP), an intoxication caused by the ingestion of bivalve molluscs that have accumulated high levels of Okadaic acid (OA), may represent a serious risk for the public health and a remarkable economic loss for the producers. Aim of this work was to improve knowledge about the repeatability of OA accumulation phenomena in various seasons trying to understand whether or not there was a trend. Also, the interaction between toxic algae and OA accumulation was examined. In this study, data of lipophilic toxins, water temperature and abundance of DSP-producing microalgal species were collected in a four-year period (2015–2018) in coastal production areas of Sardinia. Several episodes of OA positive values (>160 eq µgAO/Kg pe, Reg 853/04) were recorded during the study period in different production areas of Sardinia and in different seasons. A seasonal repeatability of OA accumulation in molluscs was observed in some production areas; moreover, different temporal gaps between the presence of toxic algae and OA accumulation were reported. Toxicity was observed almost exclusively in *Mytilus galloprovincialis* Lamarck (99%), being this matrix the most abundant species bred in Sardinia.

Introduction

Phytoplankton plays a critical role in the marine ecosystem, being primary producers. It collects light energy from the sun and transforms it into chemical energy, making it available for higher trophic levels

(Chavez *et al.*, 2011). The abundance of phytoplankton may increase during its natural seasonal cycle originating algal blooms. They can have a negative impact on marine ecosystems and the resources they provide, including fish kill, economic loss and human diseases (Hallengraeff, 2003; Anderson *et al.*, 2012).

Harmful algal blooms (HABs) have been increasing in geographical distribution (Hallengraeff, 1993; Allen *et al.*, 2006). This trend has been associated with several factors, including introduction of new species with ballast waters, eutrophication of coastal areas, and climate changes (Anderson *et al.*, 2002; Edwards *et al.*, 2006; Hegaret *et al.*, 2008; Haisler *et al.*, 2008; Moore *et al.*, 2008), that include warming of the surface water temperature, reduction in pH due to ocean acidification, increased stratification, alteration of salinity, light field intensity, nutrient flux, and grazing (Fu *et al.*, 2012; Wells *et al.*, 2015). From the presence of potentially toxic harmful algal species (HAS) it is possible to have an accumulation of marine toxins in shellfish, for which toxic and non toxic algae represent the primary source of food. Potential consequent human poisoning caused by consumption of shellfish contaminated with phycotoxins have been reported for decades. Among the algal species potentially producing toxins with range expansion, some are responsible of diarrhoeic shellfish poisoning (DSP), a alimentary intoxication caused by the consumption of shellfish contaminated with diarrhoeic shellfish toxins (DTXs), produced by several species belonging to the phytoplanktonic genus *Dinophysis* and some benthic *Prorocentrum* (Yasumoto *et al.*, 1980; Lee *et al.*, 1989; Boni *et al.*, 1993). The first DSP events were described in Japan in 1970s (Yasumoto *et al.*, 1978). Since then, outbreaks of DSP were reported globally several times. The main DTXs are a group of lipophilic polyester toxins based on Okadaic acid (OA) (Aune *et al.*, 2012), with several analogues, including dinophysitoxin-1 (DTX1), and dinophysitoxin-2 (DTX2) (Bauder *et al.*, 1996). These toxins are inhibitor of serine/threonine protein phosphatases and cause gastrointestinal symptoms in human. Ten species of *Dinophysis* are able to synthesize DTXs (Roguera *et al.*, 2014). Among these, *Dinophysis acuminata* Claparède & Lachmann is the most cosmopolitan species (Pitcher and Calder, 2000; France and Mozetic, 2006). Dinoflagellates producing OA toxin group are carefully monitored in the Mediterranean basin. They may be present during different periods of the year (Naustvoll *et al.*, 2013), or all over the year

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(Tibiriça *et al.*, 2015). They can cause the accumulation of OA group toxins in bivalve molluscs when they are the only or the most abundant source of food in a certain area. OA accumulation may follow different seasonal trends depending on causative toxic algae, as well as on the climate zone of sampling location. So, DSP toxicity events are difficult to predict (Ninčević-Gladan *et al.*, 2008).

In several studies a weak relationship between cell abundance of *Dinophysis* spp. and toxin levels has been found (Alvez-de Souza *et al.*, 2014). Many factors may explain the difficulty to relate cell abundances and toxicity, e.g. water column stratification (due to interaction between wind and water temperatures) and high variability of the intracellular toxins of *Dinophysis* spp. during blooms (Pizzarro *et al.*, 2009; Roguera *et al.*, 2012; 2014). Filtration rates, accumulation of toxins and availability of alternative food sources for shellfish are factors that may determine the toxic content in bivalves too (Alves-de-Souza *et al.*, 2014), and complicate the prediction of the toxic events based only on cell abundances. Although species of *Dinophysis* form a small fraction of the phytoplankton, they represent a serious threat for the ecosystem and for the human being, due to the production of toxins causing toxic episodes at a very low cell densities (Basti *et al.*, 2015).

The high toxicity of *Dinophysis* spp. at low concentrations is likely attributable to the lipophilic rather than the hydrophilic nature of the toxins (Yasumoto *et al.*, 1978).

The aim of this work was to describe the seasonal variability of lipophilic toxins content in molluscs bred in different production areas of Sardinia (Italy). The presence of toxic algae, such as *Dinophysis* spp. and *Prorocentrum* spp., and the accumulation of lipophilic toxins over the legal limits were studied over a four-year period (2015 – 2018). Investigation of a relationship between OA accumulation and season (seasonality), is extremely important, because it may help all the expert involved in mollusc culture (producers, biologists, sanitary controllers) to plan sampling and controls in order to perform a better analysis of the risk of OA accumulation associated to each producing area.

Materials and Methods

Study area

Sardinia is an island located in the middle of the western Mediterranean Sea, with a surface of 24.090 km² and 1849 km of coasts. Bivalve production and aquaculture play an important role in the economic and social systems. Shellfish farms are located along a surface area of about 1300 ha, both in marine and transitional waters, several of which subject to different human activities such as leisure or industrial activities. Different lagoons have a high trophic status (Padedda *et al.*, 2012) with seasonal variations in nutrient concentration.

Sample collection

Water and mollusc samples were collected from a total of 18 coastal sites (Figure 1) for four years (2015-2018), by the sanitary operators of the Italian National Health System, following protocols of the Sardinian Regional Monitoring Programme, in force since 1992 and periodically updated. For every production area, one to five stations were sampled. Presence of DSTs were determinate from molluscs, whereas the presence and abundance of DSP-producing microalgal species were established in water sampling. Data about water temperature were provided when OA were higher than legal limits.

Sample analysis

Water analysis for the evaluation of the presence and abundance of HAS were performed using Utermöhl's method (1958), in accordance with the EU reference method UNI EN ISO 15204:2006. For the cell count, settling chambers under an inverted

microscope were used (Olympus IX 73). HAS were identified at the species level according to IOC Taxonomic Reference List of Toxic Plankton Algae (<http://www.marinespecies.org/hab/>).

DSP toxin analysis

Shellfish samples collected were analysed to identify Okadaic acid (OA), *Dinophysis* toxins (DTXs), Pectenotoxin group (PTXs), Yessotoxins (YTXs) and Azaspiracid (AZAs) by liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS) (Thermo Scientific TSQ Vantage in 2015 and AB SCIEX QTRAP 4500 from 2016 to 2018) in agreement with the official protocol (AESAN, Vigo, version 5, 2015) governed by the EC Regulation 15/2011. The different phases of extraction and analysis with LC-MS/MS are reported specifically in Bazzoni *et al.* (2018).

Results

From 2015 to 2018, a total of 4395 water samples (1715 in 2015, 1040 in 2016, 831 in 2017 and 809 in 2018) and 4732 bivalve molluscs samples (1365 in 2015, 1333 in 2016, 1125 in 2017 and 909 in 2018) were analysed. Non-compliant samples were 61, distributed as follow: 23 in 2015 (2% of total amount of analysed samples), 24 in 2016 (2%), 4 in 2017 (0.4%) and 10 in 2018 (1.1%) (Table 1). All of them were positive only for OA accumulation over the legal limit (160 eq µg AO/Kg e.p.). Positive samples were found in Tortoli (TOR) every considered year, always between February to April (17 times), and always in mussels. The maximum concentration of OA was 1092 µg OA eq/kg e.p. in February 2015. Every OA positive value in

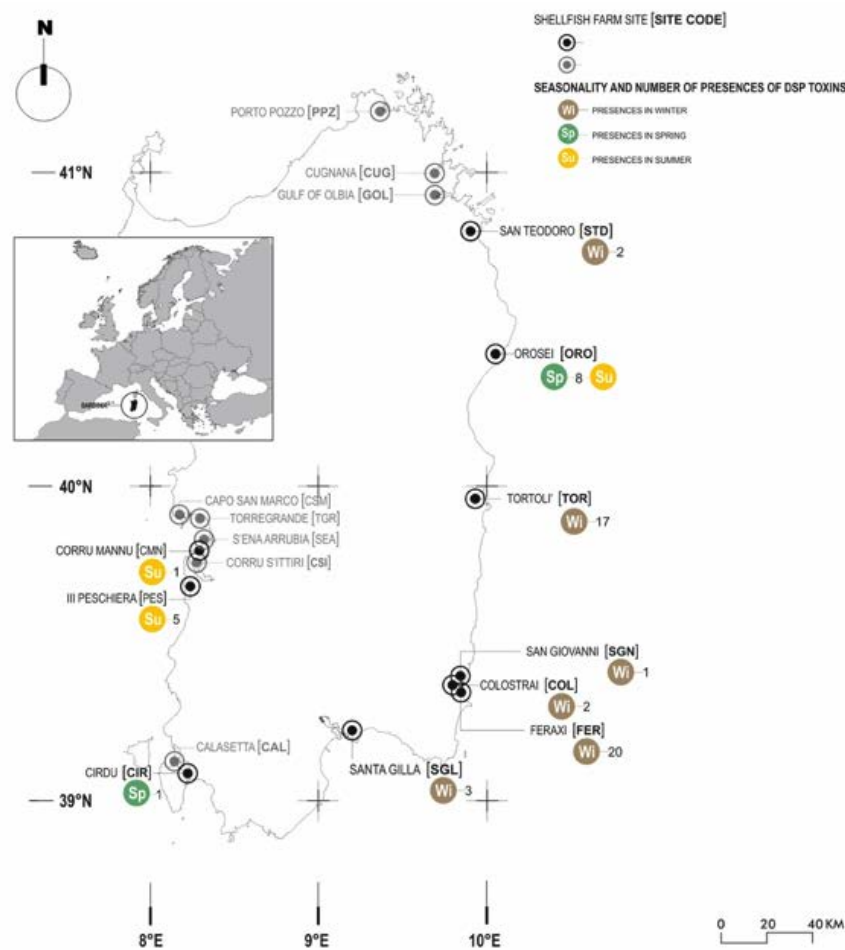


Figure 1. Spatial and seasonal distribution of OA events in different shellfish farming sites from 2015 to 2018.

mussels was observed with the only presence of *D. acuminata*, or less frequently with the contemporary presence of *D. acuminata* and *D. sacculus*. Rarely were detected *D. rotundata* Claparède & Lachmann and *Prorocentrum mexicanum* Osorio Tafall. Unfortunately, in some occasions, no data about the previous or contemporary presence of algae were available. In Orosei (ORO), OA positive values were recorded in 2015, 2016 and 2018 in spring and summer (Table 1). Most times in which OA was higher than the legal limit *D. acuminata* was detected. Twice no algae were detected. In Feraxi (FER) all OA positive values were detected in winter, between 2015 and 2017, with a peak of 1269 µg OA eq/kg e.p. in February 2015. In 2015 most times there was not a contemporary presence of toxic algae and DSP, while in 2016 they were detected together. In few cases no data were available. In San Teodoro (STD), two positive events were detected in January 2016, one with the contemporary presence of *D. acuminata* and one without. Interestingly, the positive events were detected in oysters.

In Santa Gilla (SGL), there were positive values of OA only in winter 2015; in that occasion no algae were detected. In Cirdu (CIR) there was only a single positive case, in May 2016 in mussels. In that occasion there were no algae. In III Peschiera (PES), there was only one positivity, it was the longest positive period for OA recorded in our study, from June to July 2018. On

that occasion, *D. acuminata* and *D. sacculus* were observed only in June at the onset of positivity, but the presence of toxins in mussels was detected beyond the legal limits until the end of July. Corru Mannu (COR), was the only place in the west coast where the only one positive value, in summer (2018), due to a compresence of *P. mexicanum* (400 cell/l) and *D. acuminata* (900 cell/l).

On a total of 61 OA positive cases, in 29 events (47.5%) we did not detect any algae, in 21 occasions (34.4%) there were algae and OA at the same time and in 11 occasions (18%) the presence of toxic algae was detected previously than OA accumulation over the legal limits.

Further details about the presence of OA above the legal limits, the presence of toxic algae and their species are given in Table 1. Data about water temperature are included.

Discussion

This study, including four years of data collected along all Sardinian mollusc production sites, documented the presence of OA in bivalve molluscs and that of potentially toxic microalgae belonging to *Dinophysis* and *Prorocentrum* genus, involved in the onset of DSP syndrome. Furthermore, the seasonal variability of OA content in mussels and the interaction between toxic algae and OA presence were

investigated. Several cases of OA accumulation over legal limits were recorded, distributed during the four years. In winter, all of them were detected in the Sardinian east coast. Most positive cases in spring and in summer were detected on the south-west and west coasts, except for one case in east coast. No positive value was detected during autumn.

On 18 production areas analysed, 10 were characterised at least by one positive event occurred during the study period. Only in TOR, there were positive events every year, always in winter, with a variety of *Dinophysis* species, and in few occasions *P. mexicanum*. In FER there were positive events in three out of four years, in winter too, with the presence of *D. acuminata* and *D. sacculus*. Relevant is the situation recorded in ORO, where positive events were recorded for three years in spring and in summer. Every time, the algal species recorded were *D. acuminata* and *D. sacculus*.

In the other production areas there were positive cases only occasionally, sometimes with multiple samples in short periods when the legal limits were exceeded, but always limited to a single year. On the basis of these data, a seasonality may be supposed in ORO, FER and TOR (all located in the eastern Sardinian coast) while the other sites showed no seasonal trend. Water temperature values registered every year were almost the same in each site (Table 1), but it is difficult to understand their role of for the

Table 1. Okadaic acid (OA) positivity in bivalve molluscs related to environmental parameter and potentially toxic algae (HAS) in the various seasons from 2015 to 2018.

Producing area	Site code	Year	OA positivities	Total samples analyzed	Positivity period	HAS species	Temperature (°c)	Shellfish product
Cirdu	CIR	2016	1	65	May	No data	20	Mussels
Colostrai	COL	2015	2	12	February – March	No data	12 – 14.2	Mussels
Corru Mannu	CMN	2018	1	94	August	<i>D. acuminata</i> / <i>P. mexicanum</i>	24	Mussels
Feraxi	FER	2015	8	57	February – March	<i>D. acuminata</i>	10 – 14	Mussels
Feraxi	FER	2016	10	57	January – March	<i>D. acuminata</i> / <i>D. sacculus</i>	11 – 14.5	Mussels
Feraxi	FER	2017	2	37	February – March	<i>D. acuminata</i> / <i>D. sacculus</i>	13	Mussels
Orosei	ORO	2015	2	26	April	<i>D. acuminata</i>	21	Mussels
Orosei	ORO	2016	5	26	June – July	<i>D. acuminata</i> / <i>D. sacculus</i>	22 – 28	Mussels – Clams
Orosei	ORO	2018	1	33	May	<i>D. acuminata</i>	18	Clams
San Giovanni	SNG	2015	1	46	February	No data	13	Mussels
Santa Gilla	SGL	2015	3	91	February	No data	10 – 13.3	Mussels
San Teodoro	STD	2016	2	65	January	<i>D. acuminata</i>	14.5	Oysters
Tortoli	TOR	2015	7	256	February – March	<i>D. acuminata</i> / <i>D. sacculus</i> <i>D. rotundata</i> / <i>P. mexicanum</i>	8.7 – 17.2	Mussels
Tortoli	TOR	2016	6	256	March	<i>D. acuminata</i>	11.9 – 12.6	Mussels
Tortoli	TOR	2017	2	202	February	<i>D. acuminata</i>	12.8	Mussels
Tortoli	TOR	2018	2	186	February	<i>D. acuminata</i> / <i>D. sacculus</i>	11 – 11.2	Mussels
Terza Peschiera	PES	2018	5	25	June – July	<i>D. acuminata</i> / <i>D. sacculus</i>	23 – 26	Mussels

Table 2. Toxic algal abundances and temporal gaps between the presence of toxic algae (Harmful algal species-Diarrhoetic shellfish poisoning, HAS-DSP) and OA accumulation.

Sampling station	Positive test data	OA values ($\mu\text{gao Eq/Kg E.P.}$)	HAS sampling data	HAS present	HAS abundance (Cells/L)	Temporal gap HAS-DSP	Water temperature
Feraxi 1	03/02/2015	1269	13/01/2015	<i>Dinophysis acuminata</i>	<120	3 weeks earlier	10°C
Feraxi 1	10/02/2015	263.1	10/02/2015	None	<120		12°C
Feraxi 1	19/02/2015	516.2	19/02/2015	None	<120		11.5°C
Feraxi 1	23/02/2015	738.9	23/02/2015	None	<120		11.5°C
Feraxi 1	26/02/2015	317	Data non available				
Feraxi 1	09/03/2015	424.5	04/03/2015	<i>Dinophysis acuminata</i>	<120	5 days earlier	10°C
Feraxi 1	19/03/2015	383.1	Data non available				
Feraxi 1	23/03/2015	257.2	Data non available				
Santa Gilla 2	03/02/2015	1480	03/02/2015	None	<120		10°C
Santa Gilla 2	19/02/2015	704.8	19/02/2015	None	<120		13.3°C
Santa Gilla 3	19/02/2015	498.8	19/02/2015	None	<120		13.3°C
San Giovanni 4	23/02/2015	618.9	23/02/2015	None	<120		13°C
Colostrai	19/02/2015	341.3	19/02/2015	None	<120		12°C
Colostrai	09/03/2015	470.4	09/03/2015	None	<120		14.2°C
Orosei 1	27/04/2015	184	27/04/2015	<i>Dinophysis acuminata</i>	1560	Contemporary presence	21°C
Orosei 4	27/04/2015	205	27/04/2015	<i>Dinophysis acuminata</i>	1680	Contemporary presence	21°C
Tortoli 1	11/02/2015	192.5	11/02/2015	<i>Dinophysis acuminata</i>	1800	Contemporary presence	8.7°C
Tortoli 2	11/02/2015	1092	11/02/2015	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i>	142662 37895	Contemporary presence	8.7°C
Tortoli 3	26/02/2015	188.2	26/02/2015	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i>	14400 400	Contemporary presence	10°C
Tortoli 3	11/03/2015	301.3	11/03/2015	<i>Dinophysis acuminata</i>	2400	Contemporary presence	11.3°C
Tortoli 3	18/03/2015	326.5	Data non available				11.3°C
Tortoli 3	23/03/2015	181.7	Data non available				12°C
Tortoli 3	15/04/2015	210.2	15/04/2015	<i>Dinophysis acuminata</i>	9400	Contemporary presence	17.2°C
Feraxi 1	07/01/2016	312	07/01/2016	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i>	1040 120	Contemporary presence	14.5°C
Feraxi 1	13/01/2016	1086	13/01/2016	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i>	1720 <120	Contemporary presence	14.5°C
Feraxi 1	20/01/2016	790	20/01/2016	None	<120		11°C
Feraxi 2	20/01/2016	660	12/01/2016	<i>Dinophysis acuminata</i>	120	8 days earlier	11°C
Feraxi 1	03/02/2016	547	03/02/2016	<i>Dinophysis acuminata</i>	<120	Contemporary presence	14°C
Feraxi 1	10/02/2016	778	10/02/2016	<i>Dinophysis acuminata</i>	<120	Contemporary presence	14°C
Feraxi 1	17/02/2016	315	Data non available				13°C
Feraxi 1	19/02/2016	273	Data non available				13°C
Feraxi 1	24/02/2016	385	24/02/2016	<i>Dinophysis acuminata</i>	<120	Contemporary presence	14°C
Feraxi 1	04/03/2016	317	04/03/2016	<i>Dinophysis acuminata</i>	<120	Contemporary presence	13°C
San Teodoro 1	13/01/2016	173±62	13/01/2016	<i>Dinophysis acuminata</i>	120	Contemporary presence	14.5°C
San Teodoro 2	13/01/2016	161±58	13/01/2016	None	<120		14.5°C
Tortoli 1	07/03/2016	247±89	01/03/2016	<i>Dinophysis acuminata</i> <i>Dinophysis rotundata</i>	2200 200	6 days earlier	12°C
Tortoli 2	07/03/2016		01/03/2016	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i>	1800 200	6 days earlier	12.4°C
Tortoli 3	07/03/2016	252±91	01/03/2016	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i> <i>Dinophysis rotundata</i> <i>Prorocentrum mexicum</i>	3800 200 200 400	6 days earlier	11.9°C
Tortoli 1	09/03/2016	204±83	Data non available				12.6°C
Tortoli 2	09/03/2016	299±115	Data non available				12.6°C
Tortoli 3	09/03/2016	200±82	Data non available				12.4°C
Cirdu	23/05/2016	675	23/05/2016	None	<120		20°C

Continue to the next page.

Table 2. Continued from previous page.

Sampling station	Positive test data	OA values ($\mu\text{gao Eq /Kg E.P.}$)	HAS sampling data	HAS present	HAS abundance (Cells/L)	Temporal gap HAS-DSP	Water temperature
Orosei 4	15/06/2016	607	19/05/2016	<i>Dinophysis acuminata</i>	800	>3 weeks earlier	22°C
Orosei 6	29/06/2016	1066	19/06/2016	<i>Dinophysis acuminata</i>	1200	10 days earlier	22°C
Orosei 6	04/07/2016	371	04/07/2016	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i>	2200 400	Contemporary presence	28°C
Orosei 6	19/07/2016	299	19/07/2016	None	<120		25°C
Orosei 6	25/07/2016	180±75	25/07/2016	None	<120		25°C
Tortoli 1	27/02/2017	167±70	27/02/2017	<i>Dinophysis acuminata</i>	1000	Contemporary presence	12.8°C
Tortoli 2	27/02/2017	165±69	27/02/2017	<i>Dinophysis acuminata</i>	800	Contemporary presence	12.8°C
Feraxi 1	28/02/2017	204±83	21/02/2017	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i>	1000 400	7 days earlier	
Feraxi 1	09/03/2017	180±75	Data non available				13°C
Tortoli 2	05/02/2018	174	05/02/2018	<i>Dinophysis acuminata</i>	500	Contemporary presence	11.2°C
Tortoli 3	05/02/2018	161	05/02/2018	<i>Dinophysis sacculus</i>	100	Contemporary presence	11°C
Orosei 4	16/05/2018	404	07/05/2018	<i>Dinophysis acuminata</i>	800	9 days earlier	18°C
III Peschiera 1	07/06/2018	547	07/06/2018	<i>Dinophysis acuminata</i> <i>Dinophysis sacculus</i>	18320 2720		
III Peschiera 1	12/06/2018	405	Data non available				25°C
III Peschiera 1	19/06/2018	377	Data non available				25°C
III Peschiera 1	25/06/2018	399	Data non available				24°C
III Peschiera 1	28/06/2018	216	Data non available				26°C
III Peschiera 1	02/07/2018	270	Data non available				26°C
Corru Mannu 2	28/08/2018	178	23/08/2018	<i>Dinophysis acuminata</i>	900	5 days earlier	24°C

seasonality, since data of water temperature all over the years, where OA didn't exceed legal limits, were not available to make a comparison. In the same coast, there were other sites where seasonality was not detected (among them, there were places where only a positive case was detected). Therefore, seasonality may not be related to particular features of the eastern coast, or if it may be related, is not the only factor that determine it. Toxicity was observed almost exclusively in *Mytilus galloprovincialis* Lamark (99%), being this matrix the most abundant species bred in Sardinia. The HAS analysis reported that *D. acuminata* was present in most of the cases in which OA exceed legal limits and data about algal presence were available. No evident differences were detected in algal presence from one year to another where there was seasonality, neither related to the different months when OA acid positive events occurred.

Another aspect analysed in this paper was the relationship between the presence of toxic algae and the accumulation of OA in bivalve molluscs. Specifically, most of OA positive cases were preceded by the presence of HAS. Sometimes the elapsed period of time between the appearance of HAS and the accumulation of OA in bivalve molluscs beyond the limit allowed by law

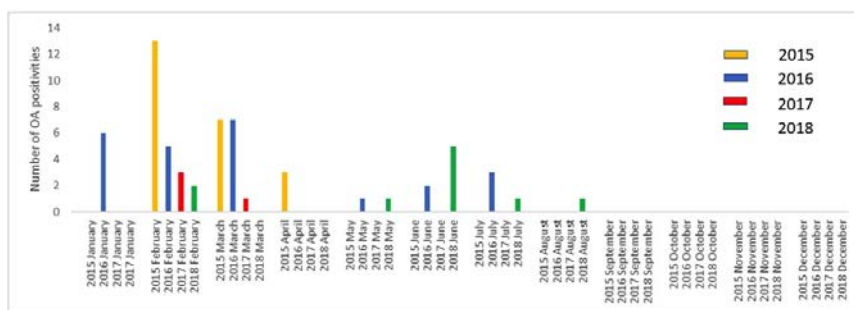


Figure 2. Monthly distribution of OA positivity events between 2015 to 2018.

was high (Table 2). We noticed this temporal gap in 18% of positive cases. In two occasion, in FER in winter 2015 and in ORO in spring 2016, the gap was remarkable, more than three weeks between the detection of *D. acuminata* and the accumulation of OA in mussels. The other temporal gaps were on average of six days. Moreover, data showed that cell abundances of HAS is not related to the amount of OA in molluscs. Another interesting aspect is that there was not a relationship between the HAS abundances and the temporal gap between algal presence and OA accumulation over the legal limits (Table 2). This discrepancy may be associated with

the spatial distribution of dinoflagellates and the fact that some genera are not toxic. In fact, HAS might not be uniformly distributed in water, but organised in spots. Moreover, the molluscs may eat a different HAS amount depending on their percentage on the total amount of algae available (Philips *et al.*, 2011).

In other events, no toxic algae were detected, neither before nor contemporary at OA accumulation, maybe because the temporality of sampling and the presence of toxic algae did not coincide. In some episodes, OA accumulation over legal limits without algal presence may be explained considering the fact that HAS may cause

OA accumulation for several weeks after their disappearance in water. For example, in FER 2015, OA accumulation was detected for the first time three weeks after the algal presence, but OA accumulation was still present after more than six weeks after HAS disappearance (Table 2; Figure 2). In SGL, during 2015 no algae were detected in each episode of OA accumulation and no data about previous HAS were available (since they were collected only concurrently with OA accumulation over the legal limits). But considering dynamics in FER and data in literature (Bazzoni, *et al.*, 2018), we may hypothesise that algae were present even many weeks before OA accumulation.

Conclusions

This study highlighted a seasonal repeatability of OA accumulation in mussels in some areas of Sardinia during the four years analysed. The timing of the seasonal presence of OA changed likely in relation with the variation of local geomorphology and geographic features, and water temperature. Moreover, the presence of OA above the legal limits in some seasons, could be linked to the interaction between HAS presence/absence with environmental parameters such as salinity of sea water, nutrients and particular sea currents that could influence the production of OA. Despite the fact that HAS are thoroughly studied, their presence is still difficult to predict (and therefore a consequent OA production), because it is determined by a complex set of factors. Further studies are necessary to evaluate these factors in order to better understand the seasonality in the production areas in which it was detected. This would allow to the operators in this field to plan a risk map of the areas where OA accumulation is not random but occurs with seasonal trend.

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