



Genome Sequences of Mycobacteriophages Ebony and Holeinone

Jennifer Cook-Easterwood,^a Hailey Gase,^a Chioma Ngene,^b Joanna Katsanos^a

^aDepartment of Biology, Queens University of Charlotte, Charlotte, North Carolina, USA

^bDepartment of Chemistry, Queens University of Charlotte, Charlotte, North Carolina, USA

ABSTRACT Ebony and Holeinone represent the first mycobacteriophages isolated from Charlotte, NC, soil samples, using the host *Mycobacterium smegmatis* strain mc²155. Ebony has a 52,152-bp genome and showed similarity to other phages in the A11 subcluster. Holeinone has a 67,044-bp genome and showed similarity to other phages in the B2 subcluster.

Ebony and Holeinone (Hole In One) were isolated from different soil samples (collected at 35.247° N, 80.931° W and 35.178° N, 80.834° W, respectively) using the bacterial host *Mycobacterium smegmatis* mc²155 as part of the Science Education Alliance-Phage Hunters Advancing Genomic and Evolutionary Science (SEA-PHAGES) program (1). *Mycobacterium smegmatis* mc²155 was provided by the Bacteriophage Institute at University of Pittsburgh and was grown in 7H9 complete liquid medium at 37°C for 5 days. Phages were isolated using enriched isolation and then purified and amplified following the protocols provided in the Howard Hughes Medical Institute (HHMI) SEA-PHAGES Discovery Guide (<https://seaphagesphagediscoveryguide.helpdocsonline.com/home>). Genomic DNA was isolated using a Promega Wizard kit, prepared for sequencing with an NEB Ultra II DNA kit, and run on an MiSeq instrument, yielding at least 1.1 million 150-base single-end reads per genome. Raw reads were assembled with Newbler version 2.7 (default settings) into a single contig for each phage and checked for completeness, accuracy, and genomic termini using Consed version 29 as previously described (2). Ebony had a genome length of 52,152 bp with a fold coverage of 3,187× and a G+C content of 63.8%, with 10-base single-stranded 3' extensions (5'-CGGTCGGTTA-3') (3). Holeinone had a circularly permuted genome with a length of 67,044 bp, a fold coverage of 2,992×, and a G+C content of 68.9%.

Both genomes were annotated using DNA Master (<http://cobamide2.bio.pitt.edu/>), and protein-coding genes were predicted using GLIMMER (4) and GeneMark (5), using the settings described in the HHMI SEA-PHAGES Bioinformatics Guide (<https://seaphagesbioinformatics.helpdocsonline.com/home>). Starterator (<http://seaphages.org/software/>) was used to predict gene starts, and protein function was determined using BLASTp (6) and Phamerator (7). ARAGORN (8) was used to analyze tRNA sequences. Ebony had 98 protein-coding regions, 1 tRNA, and 1 frameshift in the tail assembly chaperone genes. Holeinone had 90 protein-coding regions and no tRNA or frameshifts.

The bacteriophages Ebony and Holeinone represent the first mycobacteriophage species isolated from Charlotte, NC, soil samples. Ebony is part of the A11 subcluster. A Clustal Omega multiple alignment using default settings indicates there is a 98.10% average nucleotide sequence identity (ANI) between Ebony and Et2Brutus and a 97.24% average nucleotide identity (ANI) between Ebony and Mulciber. Holeinone is part of the B2 subcluster, and there is a 98.89% ANI with LizLemon and a 97.18% ANI with Godines.

Citation Cook-Easterwood J, Gase H, Ngene C, Katsanos J. 2019. Genome sequences of mycobacteriophages Ebony and Holeinone. Microbiol Resour Announc 8:e00186-19. <https://doi.org/10.1128/MRA.00186-19>.

Editor John J. Dennehy, Queens College

Copyright © 2019 Cook-Easterwood et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Jennifer Cook-Easterwood, easterwj@queens.edu.

Received 27 February 2019

Accepted 8 May 2019

Published 30 May 2019

TABLE 1 GenBank and SRA accession numbers and genome assembly results

Phage name	GenBank accession no.	SRA accession no.	Avg coverage (×)	Cluster	Genome length (bp)	G+C content (%)	No. of genes
Ebony	MH338236	SRX5352623	3,187	A11	52,152	63.8	98
Holeinone	MG812490	SRX5352622	2,992	B2	67,044	68.9	90

Data availability. GenBank and SRA accession numbers are listed in Table 1.

ACKNOWLEDGMENTS

This work was supported by the Howard Hughes Medical Institute SEA-PHAGES program.

Electron microscopy was performed at the Electron Microscopy Laboratory at Cannon Research Center at Atrium Health. Phage genomes were sequenced at the Pittsburgh Bacteriophage Institute at the University of Pittsburgh. Preliminary genome annotation was performed by the students enrolled in the genetics courses at Queens University of Charlotte during the spring 2017 semester. We thank Daniel A. Russell and Rebecca A. Garlena for comments on the manuscript.

REFERENCES

- Jordan TC, Burnett SH, Carson S, Caruson SM, Clase K, DeJong RJ, Dennehey JJ, Denver DR, Dunbar D, Elgin SCR, Findley AM, Gissendanner CR, Golebiewska UP, Guild N, Hartzog GA, Grillo WH, Hoolowell GP, Hughes LE, Johnson A, King RA, Lewis LO, Rosenzweig WLF, Rubin MR, Saha MS, Sandoz J, Shaffer CD, Taylor B, Louise T, Vazquez E, Ware VC, Barker LP, Bradley KW, Jacobs-Sera D, Pope WH, Russell DA, Cresawn SG, Lopatto D, Bailey CP, Hatfull GF. 2014. A broadly implementable research course in phage discovery and genomics for first-year undergraduate students. *mBio* 5:e01051-13. <https://doi.org/10.1128/mBio.01051-13>.
- Gordon D, Abajian C, Green P. 1998. Consed: a graphical tool for sequence finishing. *Genome Res* 8:195–202. <https://doi.org/10.1101/gr.8.3.195>.
- Russell DA. 2018. Sequencing, assembling, and finishing complete bacteriophage genomes. *Methods Mol Biol* 1681:109–125. https://doi.org/10.1007/978-1-4939-7343-9_9.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
- Besemer J, Borodovsky M. 2005. GeneMark: Web software for gene finding in prokaryotes, eukaryotes, and viruses. *Nucleic Acids Res* 33:W451–W454. <https://doi.org/10.1093/nar/gki487>.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
- Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. *BMC Bioinformatics* 12:395. <https://doi.org/10.1186/1471-2105-12-395>.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.