

Role of *CENPL*, *DARS2*, and *PAICS* in determining the prognosis of patients with lung adenocarcinoma

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Background: Non-small cell lung cancer (NSCLC) accounts for about 85% of lung cancers, and is the leading cause of tumor-related death. Lung adenocarcinoma (LUAD) is the most prevalent subtype of NSCLC. Although significant progress of LUAD treatment has been made under multimodal strategies, the prognosis of advanced LUAD is still poor due to recurrence and metastasis. There is still a lack of reliable markers to evaluate the LUAD prognosis. This study aims to explore novel biomarkers and construct a prognostic model to predict the prognosis of LUAD patients.

Methods: The Genomic Data Commons-The Cancer Genome Atlas-Lung Adenocarcinoma (GDC-TCGA-LUAD) dataset was downloaded from the University of California, Santa Cruz (UCSC) Xena browser. The GSE72094 and GSE13213 datasets and corresponding clinical information were downloaded from the Gene Expression Omnibus (GEO) database. By analyzing these datasets using DESeq2 R package and Limma R package, differentially expressed genes (DEGs) were found. Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) analyses were used to analyze possible enrichment pathways. A protein-protein interaction (PPI) network was constructed to explore possible relationship among DEGs by using the STRING database. A survival analysis was performed to identify reliable prognostic genes using the Kaplan-Meier method. A multi-omics analysis was performed using the Gene Set Cancer Analysis (GSCA). The Tumor Immune Estimation Score (TIMER) database was used to analyze the association between prognostic genes and drug sensitivity. A multivariate Cox regression was used to identify independent prognostic factors. Next, a nomogram was constructed using the rms R package. Finally, the expressions of aspartyl-tRNA synthetase 2 (DARS2) and phosphoribosyl aminoimidazole carboxylase (PAICS) were detected using immunohistochemistry (IHC).

Results: We screened out 30 DEGs prior to functional enrichment and PPI network analysis revealing potential enrichment pathways and interactions of these DEGs. Then survival analysis revealed the *CENPL*, *DARS2*, and *PAICS* expression was negatively correlated with LUAD prognosis. Additionally, multi-omics

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analysis showed CENPL, DARS2, and PAICS expressions were significantly higher in LUAD tissues than normal tissues. CENPL, DARS2, and PAICS were all up-regulated in late stage and M1 stage. Correlation analysis indicated CENPL, DARS2, and PAICS may not be associated with activation or suppression of immune cells. Drug sensitivity analysis revealed many potentially effective drugs and small molecule compounds. Moreover, we successfully constructed a robust and stable nomogram by combining the DARS2 and PAICS expression with other clinicopathological variables. Finally, IHC results showed DARS2 and PAICS were significantly up-regulated in LUAD.

Conclusions: The *CENPL*, *DARS2*, and *PAICS* expression was negatively correlated with LUAD prognosis. A prognostic model, which integrated *DARS2*, *PAICS*, and other clinicopathological variables, was able to effectively predict LUAD patients prognosis.

Keywords: CENPL; DARS2; PAICS; prognosis; lung adenocarcinoma (LUAD)

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Introduction

Non-small cell lung cancer (NSCLC), which accounts for about 85% of lung cancers, is the leading cause of tumorrelated mortality worldwide (1,2). The proportions of NSCLC histological subtypes vary according to race. Among all NSCLC subtypes, Lung adenocarcinoma (LUAD) accounted for almost 47% of cases in Western patients, while about 55–60% of cases in Chinese patients (3). LUAD is the most prevalent histological subtype in NSCLC, and accounts for 40% of lung cancer cases (4,5). In recent years, significant progress has been made in multimodal treatment strategies, including surgical resection, chemotherapy, radiotherapy, immunotherapy, and molecular targeted therapy.

A study had verified that some independent prognostic factors, including age, gender and histology, can be used to partially predict individual survival of lung cancer patients (6). However, there are still many limitations. For instance, the predictive accuracy of a single independent prognostic factor is limited. Combining multiple independent prognostic factors could improve the accuracy of prediction. To enhance the accuracy of these estimates, Cox proportional hazards models have been widely adopted (7). For instance, a nomogram is a reliable tool that has the function of quantifying risk by combining and clarifying important clinical characteristics of patients (8). By drawing a concise graph of the outcome-risk predictive model, the nomogram derives the risk probability of a specific event, such as lung cancer-specific survival. Multiple studies have confirmed that nomograms have the ability

to predict prognosis in various cancers (9-11). As a 5-year overall survival (OS) rate in advanced LAUD patients remains less than 15% (12), often due to local recurrence and distant metastasis, there is an urgent need to explore novel prognostic biomarkers and construct an effective prognostic model.

Therefore, we conducted a comprehensive bioinformatic analysis and aimed to explore promising biomarkers and construct a prognostic model for the LUAD. Finally, we successfully established a robust and stable prognostic nomogram by integrating *DARS2*, *PAICS*, and other clinicopathological variables (age, gender, and tumor stage). This promising model may serve as a reference for clinicians and help them to select more effective interventions. We present this article in accordance with the TRIPOD reporting checklist (available at https://tlcr.amegroups.com/ article/view/10.21037/tlcr-24-696/rc).

Methods

Ethics statements

This study was the research of public databases and was a retrospective study. The study protocol was approved by the Ethical Committee of the Affiliated Hospital of Qingdao University (No. QYFY WZLL 28933), and informed consent was obtained from all the participants. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). This study was a single center study (The Affiliated Hospital of Qingdao University). Patients who underwent lobectomy or lung

segmentectomy in our hospital and had pathological invasive adenocarcinoma were included in this study.

Data retrieving and processing

The Genomic Data Commons-The Cancer Genome Atlas-Lung Adenocarcinoma (GDC-TCGA-LUAD) dataset (Data Release 18.0, July 8, 2019) was downloaded from the University of California, Santa Cruz (UCSC) Xena browser. The data type count and Fragments Per Kilobase of transcript per Million mapped reads (FPKM) were selected to extract the "primary solid tumor" and convert it into transcripts per million (TPM) format. "Masked Somatic

Highlight box

Key findings

• This study showed that aspartyl-tRNA synthetase 2 (*DARS2*) and phosphoribosylaminoimidazole carboxylase (*PAICS*) expression were significantly increased in lung adenocarcinoma (LUAD) tissues. Expression of centromere protein L (*CENPL*), *DARS2*, and *PAICS* was negatively correlated with LUAD patients prognosis. We successfully established a prognostic model that integrated *DARS2*, *PAICS*, and other clinicopathological variables (age, gender, and tumor stage) to effectively predict the prognosis of LUAD patients.

What is known and what is new?

- LUAD is the most prevalent histological subtype of non-small cell lung cancer. In recent years, remarkable progress has been made in multimodal treatment strategies. However, the 5-year overall survival rate of advanced LUAD patients remains poor. To accurately estimate the individual survival of LUAD patients, many independent prognostic factors, including age, gender and histology, have been identified.
- This study successfully identified 30 differentially expressed genes that may be associated with the pathogenesis of LUAD. Among these genes, *DARS2* and *PAICS* were identified as prognostic biomarkers. Furthermore, we comprehensively analyzed the expression and prognostic value of *CENPL*, *DARS2*, and *PAICS*. Finally, a prediction model was successfully constructed based on clinical characteristics combined with *DARS2* and *PAICS*.

What is the implication, and what should change now?

• This study comprehensively analyzed the expression and prognostic value of *CENPL*, *DARS2*, and *PAICS* for LUAD. Based on these data, a robust and stable prognostic nomogram was successfully established by integrating *DARS2*, *PAICS*, and other clinicopathological variables (age, gender, and tumor stage). This promising model may serve as a reference for clinicians and help them to select more effective interventions.

Mutation" data were selected as somatic mutation data of LUAD patients. VarScan software (v2.4.0) was used to preprocess the data, and maftools R package (13) was used to visualize the somatic mutation data. At the same time, clinical characteristics of LUAD patients, including age, TNM stage, survival time, and survival status information, were also downloaded from GDC-TCGA-LUAD dataset. Missing clinical information was excluded from the study.

GSE72094 (14) and GSE13213 (15) gene expression data and corresponding clinical information (including survival time and survival state) were downloaded from the Gene Expression Omnibus (GEO) database. Data samples of homo sapiens were used. The GSE72094 chip platform was based on GPL15048. Samples without survival information were removed. Ultimately, 398 surgical specimens were included in this study. The GSE13213 chip platform was based on GPL6480. Samples without survival information were discarded. Ultimately, 117 tumor samples were retained in this study. Limma R package (16) was used to standardize the data.

Identification of DEGs

First, cancer and normal samples were extracted from TCGA dataset. The DESeq2 R package (17) was used for differential analysis of count data, and a log fold change (FC) >1 and an adjusted (adj.) P<0.05 were set as cut-off values. Second, the cancer samples were further divided into early (stage I and II) and late (stage III and IV) stage groups. Again, DESeq2 R package was used for differential analysis, and a logFC >1 and an adj. P<0.05 were used as cut-off values. Third, the GSE72094 and GSE13213 datasets were divided into early and late stage groups based on tumor stage. Limma R package (16) was used for differential analysis, and a logFC >1 and an adj. P<0.05 were chosen as cut-off values. All the statistical P values were bilateral, and a P value <0.05 was considered statistically significant.

Gene set enrichment analysis and protein-protein interaction (PPI) network

We used the OmicShare Tools (www.omicshare.com/tools) to conduct Gene Ontology (GO) (18) and Kyoto Encyclopedia of Genes and Genomes (KEGG) (19) enrichment analyses of obtained DEGs. In addition, to explore possible relationships among DEGs, a PPI network of DEGs was constructed through the STRING database (20).

Identification of key genes significantly associated with prognosis

To further assess the prognostic value of identified DEGs and identify reliable prognostic biomarkers, a survival analysis was performed using the TCGA-LUAD, GSE72094, and GSE13213 datasets. The median gene expression was set as cut-off value to compare differences in survival rates. The Kaplan-Meier method was used for survival analysis. And a P value <0.05 was considered statistically significant. Three groups of prognostic genes were intersected to identify key prognostic genes that may be closely related to development of LUAD and thus could serve as prognostic biomarkers. All the statistical P values were bilateral, and a P value <0.05 was considered statistically significant.

Multi-omics analysis of the key prognostic genes

A multi-omics analysis of key prognostic genes was performed using the Gene Set Cancer Analysis (GSCA, http://bioinfo.life.hust.edu.cn/GSCA). This analysis aimed to further investigate the copy number variations (CNVs), single nucleotide variations (SNVs), and methylation changes of key prognostic genes, and explore the prognostic differences and their relationship with transcriptomes. All the statistical P values were bilateral, and a P value <0.05 was considered statistically significant.

Relationship between key prognostic genes and immune infiltration

Using RNA-sequencing data from TCGA, the Tumor Immune Estimation Score (TIMER) (21) (https://cistrome. shinyapps.io/timer/) database was used to estimate the relationship between gene expression and immune cell level of tumor infiltration. TIMER was used to calculate the association between expression of key genes and tumor purity and immune cells, including B cells, CD4⁺T cells, CD8⁺T cells, neutrophil, macrophages, and dendritic cells. All the statistical P values were bilateral, and a P value <0.05 was considered statistically significant.

Relationship between key prognostic genes and drug sensitivity

The data, including messenger RNA (mRNA) expression profiles of genes and pharmacological activity data, were

downloaded from CellMiner database (https://discover.nci. nih.gov/cellminer/) (22). A Spearman correlation analysis was conducted to assess the correlation between expression of key genes and sensitivity of the chemical compounds. A P value <0.05 was considered statistically significant. Additionally, relevant data were obtained from the Genomics of Drug Sensitivity in Cancer (GDSC, https://www. cancerrxgene.org) and the Cancer Therapeutics Response Portal (CTRP, version 2, https://portals.broadinstitute.org/ ctrp/) databases. From the GDSC database, we downloaded gene expression profiles of various tumor cell lines along with their corresponding half-maximal inhibitory concentration (IC₅₀) values for specific drugs to assess drug sensitivity. From the CTRP database, we retrieved area under the curve (AUC) values representing cell viability following drug treatment. To investigate the relationship between key gene expression and drug sensitivity, we first matched the gene expression data with the drug sensitivity data to ensure consistency across cell lines. Subsequently, both gene expression levels and drug sensitivity metrics (IC₅₀ or AUC) were normalized. Spearman's rank correlation analysis was employed to assess the non-parametric correlation between key gene expression levels and drug sensitivity. All statistical analyses were conducted using R software (version 4.0.3). All tests were two-sided, and a P value less than 0.05 was considered statistically significant.

Construction and evaluation of the nomogram and prognostic model

To determine whether key genes were independent prognostic factors of LUAD, univariate and multivariate Cox regression was further performed to analyze key genes and other clinical characteristics (e.g., age, sex, and stage) of the TCGA-LUAD dataset. A P value <0.05 indicated that the factor was an independent prognostic factor of LUAD in multivariate Cox regression. Next, a nomogram was constructed based on optimal multivariate Cox regression analysis to predict 1-, 3-, and 5-year survival probabilities of LUAD patients. The rms R package (https://CRAN. R-project.org/package=rms) was used to construct the nomogram. The time-dependent receiver operating characteristic (ROC) and calibration curves were used to evaluate accuracy and consistency of the model. In addition, as the clinical characteristics of GSE72094 could not be matched with TCGA-LUAD, the GSE13213 dataset was used to test the model, and the time-dependent ROC

Table 1	The chara	cteristics	of LUAD	patients
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Characteristics	Values			
Stage [†]				
IV	27 (5.3)			
1	278 (54.3)			
III	83 (16.2)			
II	124 (24.2)			
OS				
Living	328 (63.8)			
Deceased	186 (36.2)			
Overall survival (months)	21.468 (13.644, 37.15)			
Sex				
Male	239 (46.5)			
Female	275 (53.5)			

Values are presented as n (%) or median (IQR).[†], 2 patients in the database did not have a TNM stage. LUAD, lung adenocarcinoma; OS, overall survival; IQR, interquartile range.

curve was used to evaluate accuracy of the test set. All the statistical P values were bilateral, and a P value <0.05 was considered statistically significant.

Immunobistochemistry

Samples were obtained from LUAD patients. Both paired tumor and adjacent normal lung tissues were collected from each patient. All samples were resected and immediately placed in liquid nitrogen. Four-um-thick sections were deparaffinized and rehydrated, and endogenous peroxidases were blocked with 3% hydrogen peroxide at room temperature. The sections were then incubated with 3% bovine serum albumin (GC305010, Wuhan Servicebio Technology Co., Ltd., Wuhan, China). Rabbit polyclonal anti-PAICS antibody (12967-1-ap, Proteintech) and rabbit polyclonal anti-DARS2 (13807-1-ap, Proteintech) were each incubated in a 1:200 dilution of phosphate-buffered saline (PBS) at 4 °C overnight, and then incubated with a horseradish peroxidase (HRP)-coupled anti-rabbit polymer, after which diaminobenzidine detection was performed (G1212, Wuhan Servicebio Technology Co., Ltd.). The sections were counterstained with hematoxylin. Both PAICS and DARS2 immunoreactivity in the tumor and adjacent normal lung tissues was evaluated with light microscope. At last, the differences of PAICS and DARS2 protein expression between normal lung tissues and LUAD specimens were quantified and significantly analyzed

Statistical analysis

The statistical analysis was performed using R software (https://www.r-project.org/, version 4.1.1). For comparisons of two groups of continuous variables, the independent Student's *t*-test was used to estimate statistical significance of the normally distributed variables, and the Mann-Whitney U test (Wilcoxon rank-sum test) was used to analyze differences between non-normally distributed variables. All the statistical P values were bilateral, and a P value <0.05 was considered statistically significant.

Results

Patient characteristics

In this study, the characteristics of LUAD patients were shown in *Table 1*. Two hundred and seventy-eight patients diagnosed with stage I LUAD were identified. One hundred and twenty-four patients diagnosed with stage II LUAD were identified. Eighty-three patients diagnosed with stage III LUAD were identified. Twenty-seven patients diagnosed with stage IV LUAD were identified.

Identification of DEGs

First, the count data of TCGA-LUAD were standardized using the DESeq2 package. After standardization, a principal component analysis (PCA) of normal and tumor samples was conducted and results showed that there was a significant difference between both groups (Figure 1A). The PCA also showed that there was no discrimination between early and late stage groups (Figure 1B). Analyzing DEGs of normal and tumor samples of TCGA-LUAD, as well as early and late stage groups of TCGA-LUAD, GSE13213, and GSE72094, identified the following 30 DEGs: OTUD1, GINS1, PAICS, GPD1, PPP1R15A, SIRPB1, TRPV2, AGRP, CENPL, PRAM1, DARS2, GART, ILF2, NCF2, ALOX5AP, CYP27A1, NELL1, CD300LF, HK3, RTN1, MNDA, CNTNAP2, PLLP, TUBA4A, CD37, ENHO, EVX1, FAM180A, FHAD1, and NME9 (Figure 1C). We used heat maps to display 30 DEGs in TCGA-LUAD, GSE72094, and GSE13213 datasets (Figure 1D-1F).

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Figure 1 Identification of differentially expressed genes. (A) PCA of normal and tumor samples from TCGA-LUAD. (B) PCA of early and late stage group from TCGA-LUAD. (C) Venn diagram of intersecting DEGs from normal and tumor samples from TCGA-LUAD, early and late stage groups from TCGA-LUAD, early and late stage groups from GSE13213 and GSE72094. (D) Heatmap of 30 DEGs in TCGA-LUAD dataset. (E) Heatmap of 30 DEGs in the GSE13213 dataset. (F) Heatmap of 30 DEGs in the GSE72094 dataset. TCGA-LUAD, The Cancer Genome Atlas-Lung Adenocarcinoma; PCA, principal component analysis; DEGs, differentially expressed genes.

Functional enrichment analysis and PPI network construction

The GO results showed that DEGs were mainly enriched in BPs and MFs (*Figure 2A*). The top three GO enrichment results were as follows: GO0004637: phosphoribosylamineglycine ligase activity; GO0004638: PAICS activity; and GO0004639: phosphoribosylaminoimidazole succinocarboxamide synthase activity. KEGG analysis showed that the top three enriched pathways were: ko00524: neomycin, kanamycin and gentamicin biosynthesis, ko04380: osteoclast differentiation; and ko00230: purine metabolism (*Figure 2B*).

In the PPI network, a total of 30 nodes were connected. There were also a total of 9 edges, and the average node rate was 0.6. The P value of the PPI enrichment analysis was 0.01 (Figure 2C).

Identification of key genes significantly associated with prognosis

The results of prognostic analysis revealed common prognostic genes in the three datasets: *CENPL*, *DARS2* and *PAICS (Figure 3A*). Expression levels of CENPL, DARS2, and PAICS were negatively correlated with prognosis of LUAD patients in the GSE13213 and GSE72094 datasets (*Figure 3B-3G*). In TCGA-LUAD dataset, OS, progressionfree survival (PFS) and disease-specific survival (DSS) were selected for survival analysis. In the survival analysis of OS and DSS, a high expression of CENPL, DARS2, and PAICS indicated a poorer prognosis. However, in the survival analysis of PFS, only a high expression of DARS2



Figure 2 Functional enrichment analysis and PPI network construction. (A) GO enrichment analysis of 30 DEGs. (B) KEGG enrichment analysis of 30 DEGs. (C) PPI network of DEGs. The cyan line stands for "from curated databases". The magenta line stands for "experimentally determined". The green line stands for "gene neighborhood". The red line stands for "gene fusions". The blue line stands for "gene co-occurrence". The yellow-green line stands for "textmining". The black line stands for "co-expression". The purple line stands for "protein homology". PPI, protein-protein interaction; GO, Gene Ontology; DEGs, differentially expressed genes; KEGG, Kyoto Encyclopedia of Genes and Genomes.

indicated a poorer prognosis. (Figure 4A-4I).

Multi-omics analysis of key prognostic genes

To further understand the characteristics and potential mechanisms of these key prognostic genes in the pathogenesis of LUAD, a multi-omics analysis was performed. In pan-cancer expression profiles, expression levels of *CENPL* and *PAICS* were increased in all tumor tissues, except for thyroid cancer tissues. Expression of *DARS2* was also significantly up-regulated in all tumor tissues, except for thyroid cancer, renal clear cell carcinoma, and renal papillary carcinoma tissues. Expression of *CENPL*, *DARS2*, and *PAICS* was significantly higher in tumor tissues

of LUAD and lung squamous cell carcinoma compared to normal lung tissues (*Figure 5A-5C*). Furthermore, *CENPL*, *DARS2*, and *PAICS* were all highly expressed in late stage groups (*Figure 5D-5F*) compared with early stage, especially in patients with stage IV disease (*Figure 5G*).

To comprehensively analyze their characteristics of *CENPL*, *DARS2* and *PAICS*, changes in their genome levels were analyzed and revealed that the mutation sites of all three genes were rare and missense mutations (*Figure 6A-6C*).

CNV analysis showed that a greater proportion of copy number amplification (including heterozygous and pure sum amplification) was observed in *CENPL* and *DARS2* compared to *PAICS*, while the proportion of copy number amplification and deletion was similar in *PAICS* (*Figure 6D-6F*).



Figure 3 Identification of key genes significantly associated with prognosis. (A) Venn diagram of DEGs associated with prognosis in TCGA-LUAD, GSE72094, and GSE13213 datasets, among which three intersecting genes were identified; *CENPL*, *DARS2* and *PAICS*; (B,D,F) Kaplan-Meier OS curves based on *CENPL*, *DARS2*, and *PAICS* expression for LUAD patients in the GSE13213 dataset. (C,E,G) Kaplan-Meier OS curve based on *CENPL*, *DARS2*, and *PAICS* expression for LUAD patients in the GSE72094 dataset. *CENPL*, centromere protein L; *DARS2*, aspartyl-tRNA synthetase 2; *PAICS*, phosphoribosylaminoimidazole carboxylase; TCGA-LUAD, The Cancer Genome Atlas-Lung Adenocarcinoma; DEGs, differentially expressed genes; OS, overall survival.



Figure 4 Survival analysis of *CENPL*, *DARS2*, and *PAICS* expression in TCGA-LUAD dataset. (A-C) Kaplan-Meier OS curves based on *CENPL*, *DARS2*, and *PAICS* expression; (D-F) Kaplan-Meier PFS curves based on *CENPL*, *DARS2*, and *PAICS* expression; (G-I) Kaplan-Meier DSS curves based on *CENPL*, *DARS2*, and *PAICS*, and *PAICS*, and *PAICS*, aspartyl-tRNA synthetase 2; *PAICS*, phosphoribosylaminoimidazole carboxylase; exp, expression; TCGA-LUAD, The Cancer Genome Atlas-Lung Adenocarcinoma; PFS, progression-free survival; DSS, disease-specific survival.

A correlation analysis between CNV and mRNA expression levels of the transcriptome data showed that CNV of *DARS2* was significantly positively correlated with its mRNA expression (*Figure 6G-61*). However, CNV prognostic analysis of *CENPL*, *DARS2*, and *PAICS* showed that there was no significant prognostic difference between amplified, deleted, and wild type mutations (Table S1). In addition, significant differences were observed in methylation of all three genes between LUAD and normal lung tissues. However, there was no significant difference in all three genes' prognosis (Tables S2,S3).

The relationship between key prognostic genes and immune infiltration

Expression levels of *CENPL*, *DARS2*, and *PAICS* were not significantly correlated with tumor purity, B cells, $CD4^+ T$ cells, $CD8^+ T$ cells, macrophages, neutrophil, or dendritic cells (*Figure 7A-7C*). Subsequently, when analyzing the relationship between CNVs of *CENPL*, *DARS2*, and *PAICS*,



Figure 5 Expression of *CENPL*, *DARS2*, and *PAICS* in pan-cancer transcriptome and early and advanced stage LUAD patient data. (A-C) *CENPL*, *DARS2*, and *PAICS* expression of pan-cancer transcriptome data in TCGA-LUAD dataset. The red box stands for the primary tumor. The purple box stands for the metastatic tumor. The blue box stands for the normal tumor. (D-F) *CENPL*, *DARS2*, and *PAICS* expression of early and advanced stage LUAD patients in TCGA-LUAD, GSE13213, and GSE72094 datasets. (G) *CENPL*, *DARS2*, and *PAICS* expression of M0 and M1 patients in TCGA-LUAD dataset. *, P<0.05; **, P<0.01; ***, P<0.001. TCGA-LUAD, The Cancer Genome Atlas-Lung Adenocarcinoma; *CENPL*, centromere protein L; *DARS2*, aspartyl-tRNA synthetase 2; *PAICS*, phosphoribosylaminoimidazole carboxylase.

and immune cell infiltration, we found that immune cell infiltration tended to be decreased in CNV amplified types (Figure S1A-S1C).

Relationship between key prognostic genes and drug sensitivity

As the CENPL, DARS2, and PAICS genes were elevated in the late stage LUAD patients, especially in stage IV patients, but not associated with immune cells, we combined drug and small molecule compound data from the GDSC and CTRP datasets. Drug sensitivity of CENPL, DARS2, and PAICS in the GCP, GDSC, and CTRP datasets was analyzed by cellMiner. In the GCP dataset, we found that CENPL was positively correlated with calusterone, nelarabine, fenretinide, and rapamycin, DARS2 was positively correlated with vorinostat, fulvestrant, parthenolide, and allopurinol, and PAICS was positively correlated with chelerythrine, amonafide, pyrazoloacridine, and nelarabine (Figure 8A-8C). In the GDSC dataset, we found that CENPL and PAICS were positively correlated with 17-AAG, afatinib, and gefitinib, while CENPL and PAICS were negatively correlated with BIX02189, BMS345541, and BX-912 (Figure 8D). In the CTRP dataset, Results showed that the expression of CENPL, DARS2, and PAICS was negatively correlated with the response of most drugs, including BI-2536, CD-437, and CHM-1 (Figure 8E).



Figure 6 SNP and CNV analysis of CENPL, DARS2, and PAICS. (A-C) Mutation sites of CENPL, DARS2, and PAICS in TCGA-LUAD dataset; (D-F) CNV analysis of CENPL, DARS2, and PAICS in TCGA-LUAD dataset; (G-I) Spearman correlation between CENPL, DARS2, and PAICS CNVs and mRNA expression in TCGA-LUAD dataset. CENPL, centromere protein L; DARS2, aspartyl-tRNA synthetase 2; PAICS, phosphoribosylaminoimidazole carboxylase; SNP, single nucleotide polymorphism; CNV, copy number variation; FDR, false discovery rate; RSEM, RNA-Seq by Expectation-Maximization; LUAD, lung adenocarcinoma; TCGA-LUAD, The Cancer Genome Atlas-Lung Adenocarcinoma.

Construction and evaluation of the nomogram and prognostic model

The univariable Cox analysis showed that *DARS2*, *PAICS* and tumor stage were determined to be independent prognostic factors of LUAD (P<0.01). On the contrary, CENPL was not a prognostic factor. These results were confirmed in the multivariable Cox analysis (P<0.05) (*Figure 9A*). Second, a prognostic nomogram was established integrating age, gender, *DARS2*, *PAICS*, and tumor stage (*Figure 9B*). The 1-, 3-, and 5-year areas under the curve (AUCs) of the training set were 0.749, 0.704, and 0.698, respectively. The 1-, 3-, and 5-year AUCs of the test set were 0.931, 0.813, and 0.746, respectively (*Figure 9C*). In addition, calibration curves of the 1-, 3-, and 5-year survival rates showed an optimal agreement between nomogram prediction and actual observations (*Figure 9D*).

Immunobistochemistry

Immunohistochemistry analysis of LUAD histological

samples showed a higher expression of DARS2 and PAICS in malignant samples compared with normal lung tissue (*Figure 10*) (the independent Student's *t*-test, P<0.05).

Discussion

In the past decades, major breakthroughs have been made in LUAD treatment. However, the long-term clinical outcomes of advanced LUAD patients have remained poorer than expected. Due to delayed diagnosis, high recurrence rate, and drug resistance, the 5-year OS rate of advanced LUAD patients remains less than 15% (12). This undermines an unmet need for effective prognostic biomarkers.

In the present study, we comprehensively analyzed the expression and prognostic value of *CENPL*, *DARS2*, and *PAICS* in LUAD patients in multiple databases using bioinformatic analyses. These results suggested that *CENPL*, *DARS2*, and *PAICS* might be oncogenes, and their genes are amplified to increase the host gene expression and



Figure 7 Immune cell infiltration analysis of *CENPL*, *DARS2*, and *PAICS* based on the TIMER database. (A) Correlation between *CENPL* and tumor purity, B cells, CD4⁺ T cells, CD8⁺ T cells, macrophages, neutrophil, and dendritic cells. (B) Correlation between *DARS2* and tumor purity, B cells, CD4⁺ T cells, CD8⁺ T cells, macrophages, neutrophil, and dendritic cells. (C) Correlation between *PAICS* and tumor purity, B cells, CD4⁺ T cells, CD8⁺ T cells, macrophages, neutrophil, and dendritic cells. (C) Correlation between *PAICS* and tumor purity, B cells, CD4⁺ T cells, CD8⁺ T cells, macrophages, neutrophil, and dendritic cells. (C) Correlation between *PAICS* and tumor purity, B cells, CD4⁺ T cells, CD8⁺ T cells, macrophages, neutrophil, and dendritic cells. *CENPL*, centromere protein L; *DARS2*, aspartyl-tRNA synthetase 2; *PAICS*, phosphoribosylaminoimidazole carboxylase; TPM, transcripts per million.

promote tumor progression and metastasis. In addition, we successfully constructed a prediction model for OS based on LUAD clinical characteristics, *DARS2*, and *PAICS*. This model is promising and may serve as a reference for clinicians and help them to make better clinical decisions when treating LUAD patients.

CENPL is a member of the centromere protein (*CENP*) family and plays important roles in regulating cell division. *CENPs* are crucial members of the centromere and kinetochore, which determine the separation of chromosomes during mitosis and meiosis (23). A previous study confirmed that several *CENPs* are highly expressed in LUAD tissues compared to normal lung tissues and have significant prognostic value (24).

DARS2, which is encoded by the class-II aminoacyltRNA synthetase family gene, is a mitochondrial enzyme that specifically aminoacylates aspartyl-tRNA (25). *DARS2* is located in chromosome 1q25.1, which mainly exists in the liver, spinal cord and brain stem (26-28). A previous study showed that the expression of *DARS2* was significantly upregulated in LUAD tissues, and *DARS2* plays a role in LUAD by targeting the ERK/c-Myc signaling pathway (29). *PAICS* is a *de novo* purine metabolic enzyme that generates N-succinocarboxyamide-5-aminoimidazole ribonucleotide via using 5-aminoimidazole ribonucleotide (AIR) by adenylosuccinate lyase. Previous research has shown that *PAICS* is significantly overexpressed in LUAD tissues and is a putative prognostic biomarker of LUAD (30). In addition, *PAICS* has also been shown to play an oncogenic role in EGFR wild-type NSCLC and represent a potential therapeutic target (31).

In recent years, there has been remarkable progress in treatment modes for lung cancer. Currently, molecular targeted therapy and immunotherapy are receiving more and more attention. Immunotherapy is rapidly gaining popularity in the armamentarium of treatment modalities for lung cancer. The most recent and very promising development is the use of neoadjuvant chemo-immunotherapy in resectable NSCLC (32). Our results suggested that all three genes may not be associated with the activation or suppression of immune cells. However, we have successfully discovered a variety of sensitive drugs associated with *CENPL*, *DARS2*, and *PAICS*. The mechanism of these genes needs to be further studied in the future.



Figure 8 Relationship between key prognostic genes and drug sensitivity. (A-C) Drug sensitivity analysis of *CENPL*, *DARS2*, and *PAICS* in GCP dataset. (D) Drug sensitivity analysis of *CENPL*, *DARS2*, and *PAICS* in the GDSC dataset. (E) Drug sensitivity analysis of *CENPL*, *DARS2*, and *PAICS* in the CTRP dataset. *CENPL*, centromere protein L; *DARS2*, aspartyl-tRNA synthetase 2; *PAICS*, phosphoribosylaminoimidazole carboxylase; GDSC, Genomics of Drug Sensitivity in Cancer; CTRP, Cancer Therapeutics Response Portal; FDR, false discovery rate.

In this study, we found CENPL, DARS2, and PAICS are predictive biomarkers in LUAD patients. This study did not only analyzed the molecular differences between early and late stage LUAD at the transcriptome level, but also explored the differential expression of CENPL, DARS2, and PAICS, and related differences in tumor microenvironment by conducting both genomics and immune infiltration analyses. The combined application of multi-omics and multi-analysis methods could greatly improve the accuracy of predicting marker molecules. However, this study also had some limitations. The data in this study were derived from public databases and multiple datasets were jointly used to improve the accuracy of the results. Although we have performed an in-depth analysis based on the available data set, it is undeniable that the limitation of the database may affect the accuracy of the results. Therefore, we expect that more real-world studies or prospective clinical trials could improve the above data and conduct a more comprehensive analysis to further verify the results of this study. To properly translate these biomarkers into clinically



Figure 9 Construction and validation of a prognostic model for LUAD patients. (A) Univariable and multivariable Cox analyses were performed by combining expression of *CENPL*, *DARS2*, and *PAICS* with other clinical parameters (age, gender, and tumor stage). (B) Nomogram for predicting the 1-, 3-, and 5-year OS probabilities of LUAD patients. (C) Time-dependent ROC curve of the training set in TCGA-LUAD dataset and the test set in the GSE13213 dataset. (D) Calibration plot of the established nomogram for predicting probabilities of 1-, 3-, and 5-year OS. HR, hazard ratio; CI, confidence interval; *CENPL*, centromere protein L; *DARS2*, aspartyl-tRNA synthetase 2; *PAICS*, phosphoribosylaminoimidazole carboxylase; TCGA-LUAD, The Cancer Genome Atlas-Lung Adenocarcinoma; TPR, true positive rate; FPR, false positive rate; AUC, area under the curve; OS, overall survival.

usable molecular markers, it is necessary to expand the sample and adopt prospective experimental methods to validate our findings.

Conclusions

In conclusion, we successfully identified 30 DEGs that may

be associated with the pathogenesis of LUAD. Among these genes, *DARS2* and *PAICS* were identified as biomarkers that could predict the LUAD prognosis. We comprehensively analyzed the expression and prognostic value of *CENPL*, *DARS2*, and *PAICS* for LUAD. Additionally, a prediction model was successfully constructed based on clinical characteristics combined with *DARS2* and *PAICS*.



Figure 10 IHC for PAICS and DARS2 protein expression in resected LUAD specimens and normal tissues. (A) Representative images for PAICS protein expression in normal lung tissue. (B) Representative images for PAICS protein expression in LUAD specimen. (C) Analysis results of PAICS protein expression between normal lung tissues and LUAD specimens, the data represent the mean \pm SD of three independent experiments. *, P<0.05. (D) Representative images for DARS2 protein expression between normal lung tissues for DARS2 protein expression in LUAD specimen. (F) Analysis results of DARS2 protein expression between normal lung tissues and LUAD specimens, the data representative images for DARS2 protein expression in LUAD specimen. (F) Analysis results of DARS2 protein expression between normal lung tissues and LUAD specimens, the data represent the mean \pm SD of three independent experiments. *, P<0.05. LUAD, lung adenocarcinoma; PAICS, phosphoribosylaminoimidazole carboxylase; DARS2, aspartyl-tRNA synthetase 2; IHC, immunohistochemistry; SD, standard deviation.

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Footnote

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Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at https://tlcr.amegroups.com/article/view/10.21037/tlcr-24-696/coif). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study protocol was approved by the Ethical Committee of the Affiliated Hospital of Qingdao University (No. QYFY WZLL 28933), and informed consent was obtained from all the participants.

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