

REVIEW

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Inflammation in cancer: therapeutic opportunities from new insights

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Abstract

As one part of the innate immune response to external stimuli, chronic inflammation increases the risk of various cancers, and tumor-promoting inflammation is considered one of the enabling characteristics of cancer development. Recently, there has been growing evidence on the role of anti-inflammation therapy in cancer prevention and treatment. And researchers have already achieved several noteworthy outcomes. In the review, we explored the underlying mechanisms by which inflammation affects the occurrence and development of cancer. The pro- or anti-tumor effects of these inflammatory factors such as interleukin, interferon, chemokine, inflammasome, and extracellular matrix are discussed. Since FDA-approved anti-inflammation drugs like aspirin show obvious anti-tumor effects, these drugs have unique advantages due to their relatively fewer side effects with long-term use compared to chemotherapy drugs. The characteristics make them promising candidates for cancer chemoprevention. Overall, this review discusses the role of these inflammatory molecules in carcinogenesis of cancer and new inflammation molecules-directed therapeutic opportunities, ranging from cytokine inhibitors/agonists, inflammasome inhibitors, some inhibitors that have already been or are expected to be applied in clinical practice, as well as recent discoveries of the anti-tumor effect of non-steroidal anti-inflammatory drugs and steroidal anti-inflammatory drugs. The advantages and disadvantages of their application in cancer chemoprevention are also discussed.

Keywords Inflammation, Cytokine, Inflammasome, Cancer prevention and treatment, FDA-approved drugs

Introduction

German physician Rudolf Virchow first reported the relationship between inflammation and tumors in the nineteenth century [1]. Chronic inflammatory processes are a fundamental innate immune response to perturbed tissue homeostasis and affect multiple stages of tumor development, including initiation, promotion, malignant conversion, invasion, and metastasis [2]. It is estimated that 15–20% of tumor-related deaths are linked to inflammation.

Recently, a growing body of evidence from basic research and clinical data indicates that inflammatory molecules and inflammation pathways can promote the occurrence and progression of various tumors. Pro-inflammatory cytokines, like interleukin (IL)–6, IL-1 β , and tumor necrosis factor- α (TNF- α), as well as

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transcription factors, like nuclear factor- κ B (NF- κ B) and signal transducer and activator of transcription 3 (STAT3), are key players in this relationship [3–7]. Encouraged by pro-inflammatory cytokines, NF- κ B and STAT3 can control the expression of target genes, the majority of which are carcinogenic, and increase the ability of cancer cells to survive, proliferate, invade, and spread [8–10]. It is now well-acknowledged that chemicals and processes associated with inflammation can be valuable targets for cancer prevention and treatment.

During the past decades, there has been strong epidemiological evidence showing that nonsteroidal anti-inflammatory drugs (NSAIDs), especially aspirin, have been associated with a reduction in the incidence and mortality of a variety of types of cancer with long-term use, especially colorectal cancer (CRC) [11, 12]. In 2016, the US Preventive Services Task Force (USPSTF) recommended low-dose aspirin for primary prevention of CRC for adults aged 50–59 years [13]. Besides, low-dose aspirin or non-aspirin NSAIDs intake is inversely related to gastric cancer risk based on multiple meta-analyses [14–16]. Recent research based on electronic endoscopy [20] and territory-wide healthcare databases [17] also verified these results. Except for CRC and gastric cancer, aspirin use also reduces the incidence and mortality of endometrial cancer [18], breast cancer [19, 20], esophageal cancer [21], liver cancer [22], and more. Besides, other NSAIDs and steroid anti-inflammatory drugs also have obvious anti-tumor effects. For example, the novel indomethacin derivative CZ-212–3 showed antitumor effects in castration-resistant prostate cancer [23]; indomethacin sensitized death receptor 5 (DR5)-deficient tumor cells to adoptive T-cell therapy [24]; indomethacin-loaded nanocapsules treatment reduced glioblastoma growth in a rat model [25]. These suggest that FDA-approved NSAIDs and steroid anti-inflammatory drugs are promising candidate drugs for cancer prevention and treatment.

In this review, the detailed information on cytokines and their function in tumorigenesis are comprehensively analyzed, and their potential as therapeutic targets for cancer treatment and chemoprevention and their inhibitor/agonists applied in preclinical and clinical studies are also clarified. Uncovering the exact mechanisms of inflammation and inflammatory factors enabled the development of novel, tailored, and highly effective cancer prevention and treatment strategies. We also discussed Food and Drug Administration (FDA)-approved non-anti-tumor drugs that may help prevent and treat cancer-related chronic inflammation, such as aspirin, indomethacin, celecoxib, and other NSAIDs and steroidal anti-inflammatory drugs. The effects of these medicines on proinflammatory cytokines and inflammation-related pathways are addressed, and typical clinical study data

are presented. According to data from the current review, agents that target chronic inflammation may have a wide range of applications in the future for the prevention and treatment of cancer.

The relationship between inflammation and cancer

The process of inflammation

Infection, tissue injury, tissue stress, and malfunction are the main causes of inflammation, and they can cause different physiological reactions and pathological outcomes [26]. The acute inflammatory response is triggered by infection or tissue injury, and the blood components including plasma and leukocytes will be recruited to the site of infection and injury [27]. A chronic inflammatory state ensues if the acute inflammatory response fails to eliminate the pathogen [28]. Except for consistent noxious stimuli like pathogens, other causes of tissue damage such as autoimmune responses or undegradable foreign bodies, can also lead to chronic inflammation [26].

Specialized sensors elicit the mediators of inflammation, which are activated by inducers of inflammation. The mediators, in turn, alter the functional states of tissues and organs (which are the effectors of inflammation). Based on their biological characteristics, the inflammatory mediators can be classified into seven groups, including vasoactive amines, vasoactive peptides, fragments of complement components, lipid mediators, cytokines, chemokines, and proteolytic enzymes.

The cellular and molecular events during inflammation include a series of complex events, from in situ to systemic inflammation throughout the body. When microbes infect the body, an acute inflammatory response occurs, which will be recognized by the innate immune system, and triggered by Toll-like receptors (TLRs) and NOD-like receptors [29]. The initial recognition of infection is mediated by tissue-resident macrophages and mast cells, causing the production of many inflammatory mediators, including chemokines, cytokines, vasoactive amines, and others [30]. If the acute inflammatory response cannot clear the antigen, the inflammation process persists, and chronic inflammation occurs. The different effector classes of the T cells can lead to different inflammatory states [26].

The crosslink between inflammation, the immune system, metabolism and autophagy in cancer

Among the various inflammatory factors that contribute to cancer susceptibility, infection has come to be recognized as a primary cause of inflammation-induced carcinogenesis [2]. During infection, inflammation is a fundamental innate immune response to perturbed tissues homeostasis, and inflammation associated with tumor development is triggered by many immune cells

[31]. The regulatory influence on the immune system determines whether inflammation exerts anti-tumorigenic or pro-tumorigenic effects. Tumor heterogeneity is shaped immunologically and through immunosurveillance by immunity's anti-tumorigenic role. Simultaneously, pro-tumorigenic inflammation fosters cancer by suppressing anti-tumor immunity, modifying the TME to a more permissive state for tumors, and directly influencing cancer and epithelial cells to promote tumors [32]. The factors that determine whether inflammation promotes or suppresses cancer are various. Currently, there are several directions for cancer prevention and treatment by regulating the immune system, including cancer vaccines and armored anti-tumor immune cells, various forms of immunotherapies, anti-tumor antibodies, and biological therapies.

A growing body of research suggests that immune cell functional changes throughout immunological responses are mediated by metabolic reprogramming [33]. These suggest that immune response and metabolic reprogramming are necessary events during inflammation-induced carcinogenesis. Reprogrammed energy production and biosynthesis and epigenetic metabolic reprogramming occur in inflammation responses [34]. The immune system exerts anti-tumor effects through immunosurveillance and immunological sculpting of tumor heterogeneity, while inflammation promotes tumor development by blocking immunity and reshaping the tumor microenvironment by regulating multiple signaling pathways and cellular functions [32].

Eating a diet that meets energy requirements and provides essential nutrients helps maintain a healthy immune system. Both undernutrition and overnutrition are associated with immune dysfunction [35]. The immune system works in conjunction with the digestive system, where immunomodulatory and inflammatory processes are linked to gut bacteria and micronutrients [36]. On one hand, people at risk for deficiencies of immune-enhancing micronutrients, such as the military population, are more susceptible to infection due to their nutritional status [37]. The immune function of soldiers consuming compound nutritional drinks with vitamins and minerals decreased slightly [38]. On the other hand, overeating can lead to the accumulation of excess body fat and obesity, which creates a chronic inflammatory state and worsens immunological deficiencies [36, 39].

Other biological factors, such as aging, also influence immune competence. The immune system undergoes several changes throughout a person's life, beginning with the developing immune responses seen in newborns and children. This progression continues to an optimal immune function typically observed in teenagers and young adults. However, in older adults, the immune

response may decline [40]. Certain lifestyle factors, including age-specific dietary choices, can worsen age-related changes by influencing and modifying immune function, and in some cases, inhibiting it. Supplements designed for different age groups may provide a solid foundation for optimal immune function. Additionally, age significantly affects how the body responds to vaccines [41, 42]. Creating effective vaccine schedules can enhance the advantages of vaccination and increase the immunogenicity of vaccines.

Several transcription factors like NF- κ B are central to immune and inflammatory responses [43]. Metabolic reprogramming occurs in immune cells during inflammatory stages, including energy production biosynthesis reprogramming, and epigenetic reprogramming [33].

Recent studies support the idea that autophagy, a fundamental biological process in mammals, is crucial for regulating inflammation and plays a significant role in cancer related to inflammation. Autophagy is generally considered a protective mechanism that helps prevent the hyperactivation of inflammatory mediators and the malignancy caused by chronic inflammation [44]. Defects in canonical autophagy or mitophagy can lead to necrosis and pathogenic responses, which promote cancer and chronic inflammation [45]. Microorganisms, damaged organelles, and crystals are sources of inflammatory signals, making canonical autophagy an anti-inflammatory process [46]. Autophagy influences immune cells by regulating mitochondrial and endoplasmic reticulum composition. These metabolic changes lead to immunometabolic states that affect macrophage and T cell polarization, ultimately impacting inflammation [47, 48]. The non-canonical autophagy, including LC3-associated phagocytosis (LAP) and its variants, such as LC3-associated endocytosis (LANDO), also utilizes some autophagy machinery and influences inflammation [49]. Research shows that LANDO's long-term clearance of beta-amyloid (Ab) aggregates influences the recycling of Ab receptors and reduces inflammation [50]. When autophagy is inactivated, there is an increased production and secretion of TNF α , type I and type II interferon (IFN), and other inflammatory cytokines [51]. Autophagy deficiency causes uncontrolled innate immune activation, leading to inflammatory diseases like Crohn's disease and an increased risk of colon cancer [52].

Four signaling pathways link autophagy and inflammation in cancer progression: the reactive oxygen species (ROS) signaling pathway, the I κ B kinase (IKK)/NF- κ B signaling axis, the inflammatory cytokine signaling pathway, and the TLR signaling cascade [53]. Autophagy-related molecules like UVRAG, IRGM, ATG7, Pink1, and Park2, as well as several receptors, connect autophagy to inflammation and influence inflammation-induced

tumors. For example, mice with mutated UVRAG show intestinal inflammation and increased risk of colitis-associated cancer through the NLRP3 inflammasome [54]. As a key negative regulator of NLRP3 inflammasome activation [55], immunity-related GTPase M (IRGM) mediates autophagy and enhances cell proliferation in hepatocellular carcinoma with overexpression of AGBL2 [56]. The activation of ATG7 leads to a pro-inflammatory response dependent on NLRP3, which worsens lipotoxicity in insulinoma [57]. Mice lacking Pink1 or Park2 are susceptible to Kras-driven pancreatic cancer due to the accumulation of mitochondrial iron and the subsequent activation of the absent in melanoma 2 (AIM2) inflammasome [58]. FUNDC1 (FUN14 domain-containing 1) is a mitophagy receptor in the mitochondrial membrane that suppresses hepatocellular carcinoma (HCC) initiation by reducing inflammasome activation and inflammation [59].

As our understanding of the relationship between inflammation, the immune system, metabolism and autophagy in cancer deepens, new therapeutic targets that alter inflammation, the immune system, and metabolism are emerging.

Potential therapeutic strategies for cancer prevention and treatment in inflammation-related cancer

Potential inflammatory therapeutic strategies for cancer prevention and treatment

It is becoming evident that the tumor microenvironment, which is mostly controlled by inflammatory cells, plays a crucial role in cancer development by promoting cell division, survival, and migration. Furthermore, certain innate immune system signaling molecules, including chemokines and their receptors for invasion, migration, and metastasis, have been appropriated by tumor cells. This knowledge is fostering new anti-inflammatory therapy approaches to cancer genesis. Blocking antibodies for inflammatory molecules such as TNF- α has shown obvious therapeutic efficacy in some inflammatory diseases and anti-tumor potential. Matrix metalloproteinases (MMPs) inhibitors have been rapidly developed and applied in the clinic, but the outcomes were less than satisfactory, with efficacy reported mainly occurs during early tumor progression [60].

Based on epidemiological data and experimental data, the FDA-approved anti-inflammatory drugs exhibit significant anti-tumor effects, suggesting the potential of these drugs as anti-tumor drugs and the targets of these drugs may act as promising targets for cancer prevention and treatment. The molecular mechanism underlying the anti-tumor effect of aspirin was first discovered in the 1970s, and it was demonstrated that it irreversibly acetylates and inactivates the cyclooxygenase (COX)

enzyme [61]. Since the cancer-promoting effect of COX, many COX inhibitors have been synthesized and have the potential as therapeutic drugs in patients with cancer. For example, flurbiprofen suppressed cell proliferation in colorectal cancer [62], non-small cell lung cancer [63], and others. While the other anti-tumor mechanisms of NSAIDs represented by aspirin and steroidal anti-inflammatory drugs were partly known. Understanding these processes and molecular mechanisms will provide new targets for the prevention and treatment of tumors. For instance, phosphatidylinositol 4,5-bisphosphate 3-kinase catalytic subunit alpha isoform (PIK3CA)/protein kinase B (AKT)/phosphatase and tensin homolog (PTEN), wingless-type MMTV integration site family (Wnt)- β catenin, and NF- κ B pathways are signaling pathways that are significantly activated in cancer, patients with mutations of these pathways showed increased survival after using NSAIDs [64]. Besides, as the important parts of the inflammatory pathway, cytokines, chemokines, growth factors, and components of the extracellular matrix-directed therapy have shown certain effects in anti-tumor treatment. These targets will be summarized in Table 2.

Potential immune therapeutic strategies for cancer prevention and treatment

As an innate immune response to the imbalance of tissue homeostasis, inflammation affects all stages of the occurrence and development of tumors, including regulating aberrant tissue repair, genotoxicity, proliferative responses, invasion, and metastasis. At the same time, tumors modulate the inflammatory environment through the secretion of soluble growth factors and chemo-attractants, causing inflammatory cells to inhibit the anti-tumor T-cell response [2]. Moreover, patients with similar illness stages but different racial/ethnic backgrounds or places of residence may exhibit significantly different tumor mutational burdens due to the consequences of exposure to persistent inflammation, which would change the immune system and the biology of tumors [65]. The complicated interplay of host, tumor, and environmental variables determines immunity and controls the magnitude and timing of the anti-cancer response. Tumors successfully suppress immune responses by activating negative regulatory pathways (also known as checkpoints) linked by acquiring characteristics that allow them to evade detection actively [66–68]. For various malignancies, immunotherapy is proven to be an efficient therapeutic strategy. Although antibodies targeting the immune regulators CTLA4 and programmed cell death ligand 1 (PD-L1)/programmed cell death protein 1 (PD-1) have shown promising therapeutic results, only a small percentage of patients show sustained responses, indicating the need for a more

comprehensive understanding of cancer immunity [69]. A recent study found an interleukin-4 fusion protein (Fc-IL-4), improves the anti-tumor efficacy of immune checkpoint blockade therapies and type 1 immunity-centric adoptive T cell transfer by directly acting on CD8+ T cells and enriching functional terminally exhausted CD8+ T (CD8+ TTE) cells in the tumor, mainly via modification of metabolism mediated by STAT6/mTOR [70].

Differences in inflammation and immune response lead to cancer disparities, including breast cancer, prostate cancer, lung cancer, colorectal cancer, and more. Cancer risk factors such as obesity, affect individuals with African American (AA) and Latina breast cancer more than those with European American (EA) breast cancer. It causes an inflammatory tumor microenvironment in breast tumors and the growth of pro-tumorigenic crown-like structures (CLSs), which comprise dead adipocytes and activated macrophages [71]. Increased numbers of tumor-infiltrating macrophages were found in AA patients [72, 73]. Increased density of CD8+ T cells was found in AA women and AA women with invasive breast cancer [74]. Higher immune content score [75] and difference in the abundance of T-regulatory (Tregs) and T-helper type 2 cells [76] in AA patients compares with EA patients. Exhausted CD8 T-cells also predict the response to anti-PD1 therapy in ER-positive breast cancer women [77]. The immune content in breast tumors also relates to the survival of ER-negative breast cancer patients [78, 79]. Low-grade inflammation and associated immune cells promote prostate cancer progression [80]. Several immune cell types such as CD4+ T cells, natural killer cells (NK) cells, and myeloid-derived suppressor cells and elevated release of IL-6 and IL-8 were associated with worse prognosis of prostate cancer patients [81–85]. The increased expression of PD-L2 was linked to poorer distant metastasis-free survival and inferior radiotherapy outcomes either in isolation or in conjunction with radiation therapy, suggesting that it could be a therapeutic target of interest for prostate cancer [81]. Increased expression of inflammatory factors such as IL-6, vascular endothelial growth factor (VEGF), brain-derived neurotrophic factor (BDNF), Tropomyosin receptor kinase B (TrkB), and higher immune content were found in AA men with prostate cancer [86]. Moreover, pre-clinical studies showed that immune cells such as Tregs, neutrophils, macrophages, and platelets promote prostate cancer metastasis [87–90]. In lung cancer, the circulating plasma levels of inflammation markers such as IL-6, IL-8, interferon gamma (IFN γ), IL-12/IL-23P40, and C-reactive protein are positively associated with lung cancer prognosis [91]. Elevated inflammatory mediators were also related to the risk of lung cancer [92]. IL-15, IL-6, and MCP-4 were identified as potential biomarkers

for early-stage lung cancer [65]. AA CRC has a more immunosuppressive environment than EA CRC patients. AA CRC patients had lower densities of cells labeled with TFNg [91]. Fewer macrophages and CD8+ T-cells, but more B-cells were seen in AA tumors compared to EA colon cancer [93]. Therefore, studying how these differences in the immune environment influence tumor biology and exploring the underlying mechanisms may help guide decisions on target therapy. The increased presence of immune-inflammation characteristics in the tumor helps to develop novel treatment therapeutics due to the reason that not every patient can afford the prohibitive expense of immunotherapy.

Although immune checkpoint inhibitors (ICIs) have significant potential, a variety of inflammatory toxicities known as immune-related adverse events (irAEs) have hindered their effectiveness [94]. ICIs are linked to inflammation and tissue damage in various organs [95, 96]. Inflammation also contributes to resistance against ICIs in cases of high microsatellite instability (MSI-H) colorectal cancer [97]. The reports indicate that reducing inflammation may enhance the effectiveness of immunotherapy. A detailed single-cell analysis of immune cell populations in colitis has identified specific cytokines, chemokines, and surface receptors that could be targeted for therapy. These targets could be beneficial not only for colitis but also for addressing other inflammatory side effects associated with checkpoint blockade, including CXCR6 and CXCL16 [98]. ICIs and IL-6 antagonists may enhance the antitumoral effect while also reducing the risk of serious adverse events [99].

Potential metabolic therapeutic strategies for cancer prevention and treatment

Evidence showed that hypercholesterolemia causes cholesterol to build up in immune cells such as macrophages, which in turn triggers inflammatory reactions. These reactions include increased TLR signaling, activation of inflammasomes, and the spleen and bone marrow generation of neutrophils and monocytes. Activation of TLR signaling causes a reduction in cholesterol efflux at the cellular level, which exacerbates inflammatory reactions and causes further cholesterol buildup [100]. As a key factor that regulates the expression of multiple genes implicated in a wide range of lipid and glucose metabolic pathways, peroxisome proliferator-activated receptor alpha (PPAR α) exhibits marked anti-inflammatory capacities [101, 102]. It plays an important role in various immune cells such as macrophages [103, 104].

As the relationship between metabolic reprogramming and immune cell activation in cancer becomes clearer, novel therapeutic targets are being identified through modifications to immune cells' intracellular metabolism.

Activation of Toll-like receptor (TLR) 4 is a vital factor that induces pro-inflammatory responses and contributes to metabolic syndrome (MetS) [105]. Therefore, TLRs could be promising targets for the treatment of inflammation-associated diseases as well as cancer.

Studies have shown that PPAR α and PPAR γ are closely related to carcinogenesis. On the one hand, loss of PPAR α promotes the development of colon cancer [106]. Overexpression of PPAR α suppressed HCC development [107]. PPAR γ plays an anti-tumorigenic role in CRC [108]. On the other hand, PPAR α facilitates cell proliferation in prostate cancer [109]. PPAR γ acts as an oncogenic role in the development of thyroid carcinoma [110]. Several PPAR-interacting miRNAs, such as miR-506 and miR-27a, have been shown to suppress PPAR α and PPAR γ in cancer cells [111, 112]. Besides, the upregulation of PPARs or PPAR agonists has been reported to be involved in chemotherapy resistance by metabolic reprogramming. For example, up-regulated PPARs or PPAR agonists foster chemotherapy resistance in cisplatin-resistant ovarian cancer [113], hepatocellular carcinoma [114], colorectal cancer [115], and NSCLC [116]. This suggests the potential of PPARs to act as a promising target for cancer treatment and prevention.

Potential autophagic therapeutic strategies for cancer prevention and treatment

Dysregulation of autophagy contributes to inflammation and tumor progression. Enhancing autophagy can boost CTLA-4 expression and Tregs, potentially reducing inflammation and cancer [117]. Inhibiting the autophagy pathway encourages the transformation of macrophages into pro-inflammatory M1 cells, which worsens liver inflammation [118]. Research is focused on small molecules and nanomaterials that modulate autophagy for anti-tumor effects. For example, 3-Methyladenine (3-MA) blocks autophagy, reduces ROS, inactivates pro-inflammatory proteins STAT3 and IL-6, and inhibits the survival of HeLa cells [119]. Combining the induction of autophagy by rapamycin with the activation of TLR4 and TLR9 has shown to have a synergistic effect in enhancing anticancer effectiveness.

Rapamycin, an autophagy activator, demonstrated enhanced anticancer effectiveness when combined with TLR4/9 agonists in melanoma treatment [120]. This indicates the potential of combining immunotherapy with autophagy activators for cancer treatment. Besides, GL-V9, an AMPK activator, prevents colorectal cancer associated with colitis by limiting the NLRP3, nucleotide-binding domain leucine-rich repeat (NLR) and pyrin domain-containing receptor (NLRP3) inflammasome through the process of autophagy [121]. Ergosterol peroxide induces ROS-dependent autophagy and reduces

NLRP3 inflammasome activity, inhibiting lung cancer cell migration and proliferation. However, it is still unclear if autophagy directly suppresses inflammasome activity [122].

Inflammation regulates the hallmarks of cancer

New evidence has strengthened the idea that inflammation plays a crucial role in the development of tumors. Sites of infection, persistent irritation, and inflammation are the origin of many malignancies. It is becoming evident that the tumor microenvironment, which is predominantly regulated by inflammatory cells, plays a crucial role in cancer development by promoting cell division, survival, and migration. It fosters the proliferation of mutant cells and facilitates the spread of malignancy. The hallmarks of inflammation-induced cancer include proliferative and survival signaling promotion, genomic destabilization, angiogenesis promotion, invasion, migration and metastasis induction, and reduced sensitivity of tumors to chemotherapy drugs.

Promoting proliferation and survival signaling

The concept that inflammation enhances cell proliferation has been proposed since 1863, and continuous cell proliferation increases the risk of tumor occurrence [1]. Various inflammatory molecules, as the main component of the tumor microenvironment, contribute to the proliferation of cancer [123]. Chronic inflammation results in the continuous release of pro-inflammatory chemokines and cytokines such as interleukins (ILs) 6 and TNF α , as well as transforming growth factor beta (TGF- β), which promote the survival of tumor cells [124, 125]. Under the stimulation of inflammatory factors such as IL-6 and IL-1 β , NF κ B and STAT3 were activated. Then, the expression of various oncogenic genes was dysregulated, promoting cancer cells' survival and proliferation [126].

Destabilizing genome

As an enabling characteristic of cancer, genome instability is essential for cancer cell evolution [127, 128]. Accurate DNA synthesis and effective DNA repair processes are crucial for maintaining genomic stabilization. Chronic inflammatory processes promote the development of normal cells, their growth, and their transition to malignancy by disrupting cellular homeostasis, producing various DNA damage products, impairing DNA repair pathways, producing an excess of pro-inflammatory cytokines, and lowering the rates at which damaged cells undergo apoptosis [129, 130]. Abundant evidence indicates that inflammation contributes to genomic instability [131–133]. Inflammation generates reactive oxygen and nitrogen species (RONS) [132, 134, 135] and ROS [136] during the inflammatory response, which results

in direct DNA damage. The inflammation-associated DNA damage contributes to colon carcinogenesis in mice [137]. The DNA damage-induced sustained p53 activation contributes to inflammation-associated hepato-carcinogenesis in rats [138].

Promoting angiogenesis

Angiogenesis is recognized as the process of growth and remodeling that transforms an initial vascular system into the intricate branching network that characterizes adult vasculature [139]. Anastomosis, migration, survival, proliferation, and ECM breakdown are all part of this intricate multi-step process [140]. Abnormal vessel growth and function are hallmarks of cancer [141, 142]. Abundant evidence showed that in pathological circumstances such as cancer, angiogenesis, and inflammation are closely related [143]. Various pro-inflammatory chemokines and cytokines released during inflammation are potent activators for endothelial cells (ECs) to attract blood-derived inflammatory cells [144]. Adhesion molecules play a vital role in this process, and they're regulated by various pro-inflammatory mediators [145]. Alternatively, angiogenesis maintains inflammation by providing oxygen and nutrients to meet the metabolic needs of cells at the site of inflammation. The tumor microenvironment is rich in inflammatory molecules such as TNF α , IL-1, IL-6, IL-8, COX-2, and vascular endothelial growth factor (VEGF) [146, 147]. For example, as one of the best-characterized inflammatory mediators in carcinogenesis, TNF α activates NF- κ B [148], which is involved in angiogenesis [149]. VEGF production contributes to angiogenesis, fostering tumor development [150, 151]. Since angiogenesis is essential for the formation and spread of tumors, there were substantial expectations that inhibiting this process would have therapeutic benefits. To demonstrate the utility of anti-angiogenic medicines, more than thirty years and multiple pre-clinical investigations employing a variety of anti-angiogenic strategies were required [152, 153]. As one of the most studied targets in clinical trials, VEGF shows significant biological functions [154]. The first antiangiogenic agent anti-VEGF mAb, has been approved to treat metastasis colorectal cancer combined with chemotherapy [155]. Anti-inflammatory medications have also been demonstrated to have antiangiogenic properties, suggesting that their use may be advantageous for neoplastic therapy.

Inducing invasion, migration, and metastasis

Inflammatory cells and mediators participate in the invasion, migration, and metastasis of malignant cells. Many cells begin to produce chemokines when they are transforming. As a result, they can use chemokines to help migrate to and survive at locations distant from the

original tumor [156–158]. For example, cell migration is influenced by the C-X-C chemokine receptor (CXCR) 4 and its ligand (C-X-C motif) ligand (CXCL) 12 in both healthy and pathological conditions [158]. Malignant cells often express CXCR4 [156]. The degree of CXCR4 expression in primary human malignancy is correlated with lymph node metastasis in colorectal, breast, liver, and esophageal cancer [159–161]. Besides, malignant cells from a variety of tissues also express other functional chemokine receptors, such as CX3Chemokine receptor 1 (CX3CR1), CC-chemokine receptor 1 (CCR1), CCR7, CCR9, CCR10, CXCR1, CXCR2, CXCR3, CXCR5, and CXCR7. These receptors are implicated in organ-specific metastasis [162–167]. For instance, the expression of CCR7 is correlated with lymph node metastasis, and CCR9 is correlated with melanoma metastasis to the small intestine. Many of the receptors are expressed by malignant melanoma cells, which could account for melanomas' high rate of metastasis. Several mechanisms may help malignant cells acquire the ability to express chemokine receptors, including genetic and epigenetic alterations, and autocrine and paracrine extracellular signals. The inflammatory cytokines such as TNF- α , IL-6, and IL-1 β , can enhance the invasive ability of tumor cells in ovarian cancer, breast cancer, and others [168–170]. Higher levels of the cytokine IL-6 are associated with increased risk of distant recurrence of HER2-negative early breast cancer [171]. TGF- β promotes epithelial-mesenchymal transition (EMT) [172, 173] and metastasis [174–178] at a late stage of cancer. Inflammatory macrophages contribute to ovarian cancer cell migration in vitro [179] and ovarian cancer model [180]. Macrophage-deficient mice could develop tumors but can't form pulmonary metastasis in a breast cancer model [181]. In conclusion, malignant cells and invading leukocytes interact with autocrine and paracrine by coordinating chemokines and cytokines. Due to these interactions, malignant cells are more likely to migrate, invade, and survive.

Reducing the sensitivity of tumors to chemotherapeutic drugs, radiotherapy and immunotherapy

Chemotherapy, radiotherapy and immunotherapy are common treatment options utilized for various cancer types. One of the side effects of chemotherapy is producing inflammatory cytokines related to the side effects and drug resistance. Similarly, radiation-induced DNA damage response (DDR) also promotes the release of cytokines and chemokines, which will cause the invasion and metastasis of cancer by triggering the inflammatory process and suppressing the immune function [182, 183]. Resistance has been the significant barrier to effective cancer treatment [184–187]. Inflammatory mediators

such as GM-CSF, IFN- γ , IL-1 β , IL-5, IL-10, and tumor necrosis factor alpha (TNF- α) play crucial roles in the resistance associated with inflammation. For instance, CD4+ T helper 2 (TH2) cells, which are key regulators of inflammatory processes in cancer, can accelerate pulmonary metastasis and promote chemotherapy resistance through the activation of macrophages by IL-4 in breast cancer [188]. Activated downstream of IL-13, the signal transducer and activator of transcription 6 (STAT6) enhances anti-apoptotic pathways, potentially leading to resistance against cytotoxic CD8+ T cells and chemotherapy in breast cancer [189, 190]. NF- κ B upregulates the Wnt family member Wnt16B, which attenuates the effects of cytotoxic chemotherapy in prostate cancer [191]. NF- κ B can also be activated by chemotherapy drugs such as paclitaxel, cisplatin, doxorubicin, and bleomycin [192]. Therefore, inhibition of the inflammatory cytokines pathway and related molecules seems to be an up-and-coming method to improve chemotherapy efficacy. Phytochemicals such as resveratrol, curcumin, genistein, epigallocatechin-3-gallate (EGCG), lycopene, thymoquinone, quercetin, capsaicin, and other phytochemicals, that inhibit the NF- κ B pathway have a preventive effect on drug resistance [193–196]. For example, resveratrol enhances doxorubicin-induced cytotoxicity in chemo-resistant B16 melanoma [197]. Curcumin can sensitize tumors to many chemotherapeutic drugs in various types of cancer, such as colon, breast, liver, lung, ovarian cancer, and others; curcumin also sensitizes tumors to gamma radiation in multiple types of cancer, such as colon cancer, cervical carcinoma, glioma, neuroblastoma, epidermal carcinoma, prostate cancer, and others [198, 199].

Radiation kills cancer cells via various mechanisms, while the dying cells promote the release of several cytokines, such as TGF- β [200]. TGF- β closely relates to radio-resistance by suppressing the immune system and regulating angiogenesis, invasion, and DNA damage [201, 202]. Inhibition of TGF- β RI can reverse the radio-resistance of irradiated cells [203]. In some clinical trials, TGF- β antagonists such as fresolimumab and LY2157299, have been applied to cancer patients undergoing radiotherapy and have obtained some promising results [201]. For example, TGF- β blocking antibody fresolimumab increases the overall survival of metastatic breast cancer patients [204]. These suggest that anti-TGF- β medications and radiation therapy together may improve tumor response and minimize side effects. Additionally, LIF mRNA, which is expressed in immune cells such as T-cells, macrophages, and monocytes, as well as in epithelial carcinoma cells and adjacent stromal cells, promotes radio-resistance in nasopharyngeal carcinoma by activating the mTORC1/p70S6K pathway [205].

The blockade of LIF altered macrophages, slowed tumor progression, and improved the effectiveness of anti-PD-1 therapies in colon cancer [206].

EMT is an important step during resistance [207], IL-6 cytokines can promote EMT by activating transcription factors such as STAT3 and Snail. This suggests that blocking IL-6 family cytokines may enhance the effectiveness of immunotherapy. Drugs that target IL-6 cytokines have been shown to improve the efficacy of immune checkpoint blockade (ICB) and reduce irAEs. Several clinical trials are currently underway to evaluate the combination of anti-IL-6 therapies with immunotherapy in cancer treatment [208]. In animal models of cancer, the upregulation of CD38 is associated with acquired resistance to anti-PD-L1 treatment. CD38 decreases the activity of CD8 T cells and reduces the effectiveness of anti-PD-L1 immunotherapy by contributing to the accumulation of adenosine in tumors [209]. Blocking CD38 made tumors more susceptible to anti-PD-L1 therapy. Moreover, inhibiting CD39's enzymatic activity leads to the depletion of intra-tumoral macrophages, modifies the inflammatory response, boosts T cell proliferation, and overcomes anti-PD-1 resistance in colon cancer [210]. Inhibition of A2A adenosine receptor increases the efficacy of anti-PD-1 and produces T cell-dependent anti-tumor activity in breast and colon cancers [211]. Inhibition of the CD39-CD73-adenosine axis appears to be a promising strategy for overcoming resistance to immunotherapy.

The relationship between inflammatory diseases and cancer

Crohn's disease (CD) and ulcerative colitis (UC) are two types of inflammatory bowel disease (IBD) that involve chronic inflammation of the gastrointestinal tract. Patients with Crohn's disease and ulcerative colitis are at a higher risk of being diagnosed with cancer compared to the general population [212], such as colorectal cancer [213], small-bowel cancer [214], colon cancer [215], and also the skin cancers, lymphomas and cervical abnormalities [216]. CD also increases the incidence of anal cancer and anorectal cancer [217]. The risk of cancer may depend on the severity and duration of inflammation. Pro-inflammatory cytokines such as IL-6, generated during gut inflammation, play a crucial role in cancer development by activating intracellular transcription factors like STAT3 and NF- κ B [218]. Anti-tumor necrosis factor therapy at an intensified dose is the most effective medical treatment for perianal Crohn's disease, as it can lower the risk of anal tumors [219]. Current data indicates that immunosuppression for IBD does not increase the risk of new or recurrent malignancies [220]. The blockade of IL-23 using neutralizing antibodies, which has been tested in clinical trials for Crohn's disease therapy, is

anticipated to be an effective method for treating colon cancer and colorectal cancer. This expectation is based on the regulatory role of IL-23 in carcinogenesis [215].

Aside from IBD, autoimmune diseases like rheumatoid arthritis (RA) and celiac disease are notable. Patients with RA have an increased risk of developing prostate cancer [221], lung cancer [222], nonmelanoma skin cancer (NMSC) [223], and non-Hodgkin's lymphoma [224]. Additionally, RA increases the risk of lymphoid, myeloid, cervical, and oropharyngeal cancers, while decreasing the risk of endometrial and colorectal cancers, according to a study of one million women [225]. Long-term immunological dysregulation, such as altered B cell activation and survival, along with the inflammatory reactions associated with RA development, may further increase the risk of developing cancer [226, 227]. Antagonists of TNF- α , Janus kinase (JAK), and IL-6 are commonly used therapeutics for RA, but their effects on tumor development are mixed. For instance, while TNF inhibitors have been found to increase the risk of NMSC in patients with RA, they can also reduce tumor size and metastasis in cases of colorectal and colon cancer [228, 229]. JAK inhibitors didn't increase the risk of cancer [230, 231]. Celiac disease is an autoimmune disorder that primarily affects the small intestine and is triggered by gluten consumption in genetically predisposed individuals [232]. The development of coeliac disease involves a complex immune response to gluten proteins. Celiac disease also increases the risk of non-Hodgkin lymphoma [233], small bowel carcinoma [234]. Additionally, it raises the mortality rate among cancer patients [235].

The gluten-free diet is the primary treatment for coeliac disease. It helps reduce damage to the small intestinal mucosa caused by repeated gluten ingestion, thus lowering the risk of lymphoproliferative malignancies [236]. Nutritional support, pharmacological agents, and autologous hematopoietic stem-cell transplantation are also recommended for non-responsive type 2 cases [237]. Additionally, utilizing new techniques such as gene silencing (e.g., RNA interference) or gene editing (e.g., CRISPR-Cas9) to reduce immunogenic gluten proteins could enhance the quality of life for individuals on a gluten-free diet [238]. A pilot study indicated that a low-gluten RNA interference wheat line did not provoke an immune response after a short-term oral challenge in patients with treated coeliac disease [239]. These methods may also offer innovative approaches for the primary prevention of celiac disease.

The role of inflammation in certain types of cancer

Several types of tumors are directly related to inflammation. For example, the development of colitis and colitis-associated tumors is closely related to chronic

inflammation [240]. There are several pathogens that play an important role in inflammation-related tumors such as cervical cancer, gastric cancer, and liver cancer. For example, human papillomavirus (HPV) is a major risk factor for the development of cervical cancer, as its proteins are involved, either directly or indirectly, in inflammation that can lead to the onset of this disease [241]. Immune cell infiltration and the increased secretion of inflammatory cytokines are two primary factors in the development of cervical cancer [242].

Helicobacter pylori (*H. pylori*) is classified as a Group I carcinogen and infects over half of the world's population. It has unique properties that allow it to colonize the gastric epithelium in acidic environments. Infections caused by *H. pylori* can lead to chronic gastritis, which may progress to severe gastrointestinal conditions, including gastric cancer [243]. Understanding the complex bacterial virulence mechanisms and their interactions with the host immune system and environmental factors is crucial for comprehending the pathophysiology of *H. pylori* infection.

In China, the hepatitis B virus (HBV) is the primary cause of liver cancer [244]. While liver cancer is primarily caused by hepatitis C virus (HCV) infection in South Korea and Japan [245]. Chronic infections of HBV or HCV, along with factors such as nonalcoholic fatty liver disease (NAFLD) and excessive alcohol consumption, lead to liver injury, chronic inflammation, and ultimately liver cancer [246]. The transformation of inflammatory tumors in the liver is a gradual and dynamic process [247]. The chronic inflammation leads to an altered immune environment, increased secretion of inflammatory factors, and dysfunctional NK cells, ultimately resulting in liver cancer [248].

The role of patient-specific factors in inflammation-related cancer

Various patient-specific factors, such as the microbiome and genetics, play crucial roles in cancer related to inflammation.

The role of microbiome in inflammation-related cancer

The microbiome can influence key characteristics of cancer, including the promotion of harmful local inflammation that contributes to the development of tumors. For instance, *Helicobacter pylori* colonizes the human gastric mucosa, leading to chronic inflammation and resulting in gastric ulcers, which can eventually progress to stomach cancer [243]. Research has shown that *Fusobacterium nucleatum* in colorectal cancer activates NF- κ B, an important modulator of inflammation associated with cancer, through its binding to Toll-like receptors and nucleotide-binding oligomerization domain-like

receptors [249]. Moreover, the carcinogenic potential of gut bacteria results in increased production of IL-6 and TNF, activation of STAT3, and activation of the IL-17–IL-23 pathway [250]. Innate and adaptive host immune responses triggered by microbiota can collaborate to promote the development and spread of malignancies by inducing inflammation that supports cancer growth and increases resistance to cell death.

The composition of microbiota can significantly influence the effectiveness and side effects of cancer treatments, as well as the quality of life after cancer therapy [251]. For example, there is a significant difference in the microbiome composition between responder and non-responder patients undergoing anti-PD-1 immunotherapy [252, 253]. The altered gut microbiome due to antibiotics reduced the effectiveness of ICIs in advanced cancer patients [254]. Moreover, the effectiveness of chemotherapeutic drugs can also be influenced by bacteria, affecting their metabolism and absorption [255]. Probiotics and microbiota transplants may be effective strategies for modifying the microbiome, potentially enhancing cancer therapy responses and improving quality of life. For instance, oral supplementation with *A. muciniphila* or *Bifidobacterium* can boost the effectiveness of PD-1 or PD-L1 immunotherapy in mouse models of melanoma and lung cancer [254, 256]. Importantly, certain probiotic strains, prebiotics like inulin and fructooligosaccharides, and synbiotics can reduce gastrointestinal side effects in cervical cancer patients. They do this by altering key metabolic pathways and inflammation, lowering oncogene activity, and ultimately reducing cancer progression [257]. Microbiome-regulating agents are relatively safe, making them potential adjuvants in clinical cancer treatment and paving the way for personalized therapies. However, more high-quality clinical trials are urgently needed.

The role of genetics in inflammation-related cancer

Cancer was once thought to be solely hereditary, but it is now understood to be influenced by a combination of dysregulated genetic and epigenetic pathways [258]. Genetic alterations are essential for cancer development and the creation of an inflammatory microenvironment, linking inflammation and cancer through an intrinsic pathway [259]. Genetic alterations, including oncogene activation through mutations, chromosomal rearrangements, or amplifications, as well as inactivation of tumor-suppressor genes, contribute significantly to cancer development. Among the most frequently mutated dominant oncogenes in human cancer are myelocytomatosis oncogene (MYC) and rats arcomaviral oncogene homolog (RAS), which induce tumor-promoting inflammatory mediators that play critical roles in tumor growth

and angiogenesis [260, 261]. The Kirsten rats arcomaviral oncogene homolog (KRAS)-G12D mutation promotes immunosuppression and increases resistance to anti-PD-1/PD-L1 immunotherapy in non-small cell lung cancer (NSCLC) [262]. MYC promotes resistance to immunotherapy and IFN γ by downregulating JAK2 [263]. Targeting dysregulated MYC or RAS may enhance immunotherapy effects, as several studies have demonstrated. For instance, inhibiting MYC increased tumor immune cell infiltration and made tumors more responsive to anti-PD1 immunotherapy [264]. OmoMYC, a peptide and mini-protein that inhibits MYC, is currently being developed for evaluation in clinical trials for relapsed or refractory acute myeloid leukemia [265]. Currently, several inhibitors targeting RAS and MYC mutations are in clinical use or undergoing clinical trials. This offers a promising strategy to enhance anti-tumor effects and overcome the drug resistance of immunotherapy. Additionally, tumor-suppressor proteins like von Hippel-Lindau (VHL) and PTEN can regulate the production of inflammatory mediators [266, 267]. PTEN treatment improves anti-PD-1 efficacy in mice and reverses the immune-suppressive phenotype of primary tumor-associated macrophages derived from patients [268]. Administering PTEN externally may serve as a promising cancer immunotherapy strategy.

Targeting inflammatory pathway for cancer chemoprevention

A growing tumor's inflammatory component may involve various leukocyte types, such as neutrophils, dendritic cells, macrophages, eosinophils, mast cells, and lymphocytes. These leukocyte types can produce a wide range of cytokines, as well as cytotoxic mediators like reactive oxygen species, serine and cysteine proteases, MMPs, and membrane-perforating agents, as well as soluble mediators of cell death like TNF- α , interleukins, and IFNs [269]. The detailed information about the inflammatory cells and molecules they secrete are summarized and shown in Fig. 1. And the pro- or anti-tumor effect of these inflammatory molecules are also summarized and shown in Fig. 2 and Fig. 3. TNF α antibodies or inhibitors such as infliximab are undergoing phase 2 clinical trials in breast and renal cell cancer [270]. Chemokine receptor antagonists like bicyclam plerixafor are now in clinical use in non-Hodgkin lymphoma and multiple myeloma [271]. Trabectedin as the cytotoxic drug to tumor-associated macrophages and circulating monocytes, has been used to treat soft-tissue sarcoma and ovarian cancer [272]. Fibroblast plays an important role in tumor development, and inhibitors of the fibroblast growth factor receptor have been studied in various tumors [273].

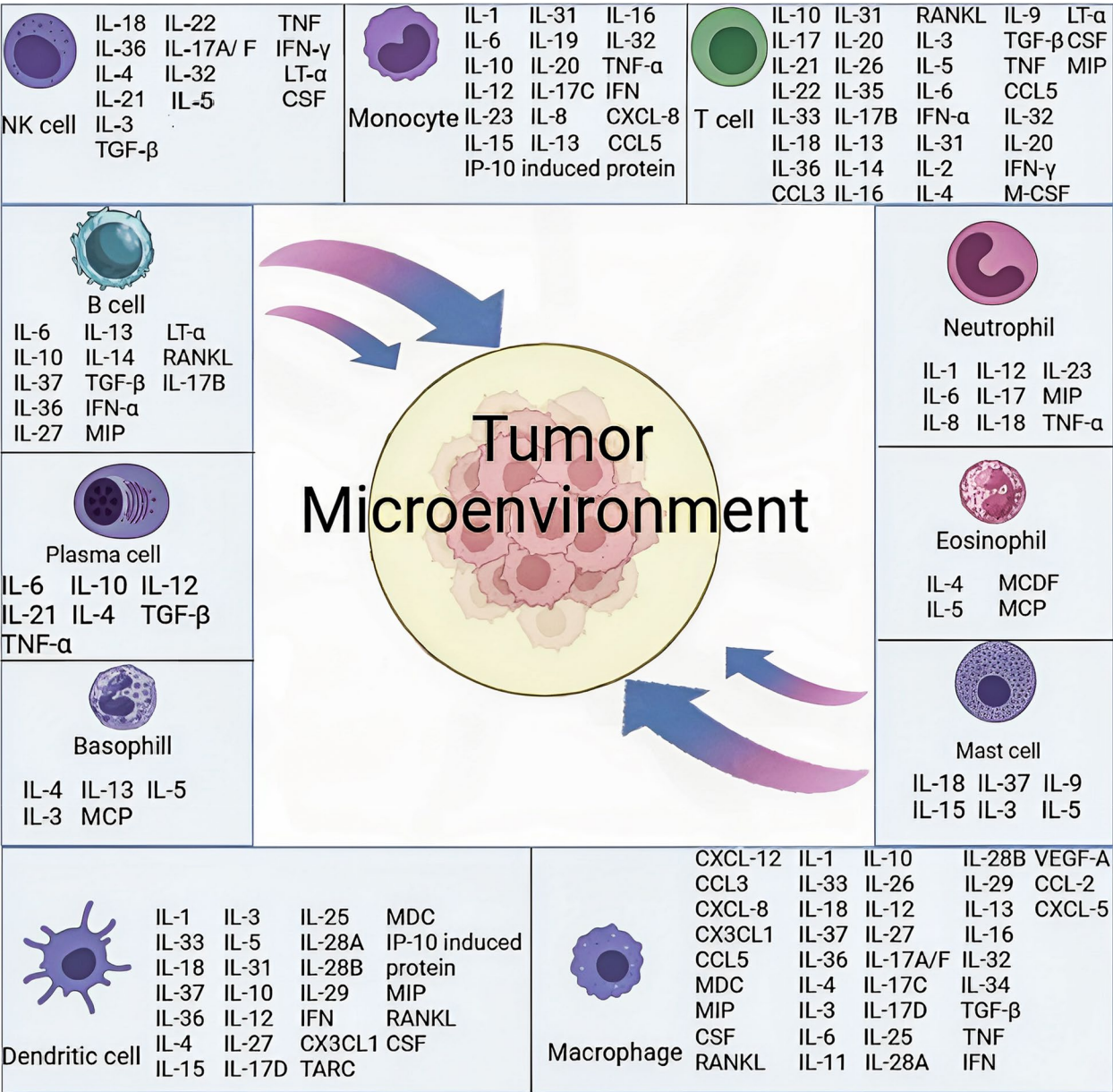


Fig. 1 The detailed information about the inflammatory cells and molecules they secrete. The inflammatory cells include NK cell, monocyte, T cell, neutrophil, eosinophil, mast cell, macrophage, dendritic cell, basophill, plasma cell and B cell. NK cell secretes several kinds of interleukin (IL), interferon (IFN), tumor necrosis factors (TNF), transforming growth factors (TGF), colony-stimulating factor (CSF) and lymphotoxin alpha (LT-α); monocyte secretes several kinds of ILs, IFN, TNF, C-X-C motif ligand (CXCL) and interferon-inducible protein-10 (IP-10) induced protein; T cell secretes several kinds of ILs, IFN, TNF, TGF, macrophage colony-stimulating factor (M-CSF), receptor activator of nuclear factor-kappa B ligand (RANKL), macrophage inflammatory protein (MIP) and LT-α; neutrophil secretes several kinds of ILs, TNF-α and MIP; eosinophil secretes IL-4, IL-5, MCDF and monocyte chemoattractant protein (MCP); mast cells secretes several kinds of ILs; macrophage secretes several kinds of CXCLs and ILs, TGF-β, TNF, IFN, CSF, vascular endothelial growth factor A (VEGF A), macrophage-derived chemokine (MDC) and MIP; dendritic cell secretes macrophage secretes several kinds of ILs, CX3CL1, IFN, RANKL, CSF, thymus and activation-regulated chemokine (TARC), MDC, MIP and IP-10 induced protein; basophill secretes MCP and several kinds of ILs; plasma cell secretes TGF-β, TNF-α and several kinds of ILs; B cell secretes several kinds of ILs, LT-α, RANKL, TGF-β, IFN-α and MIP

Targeting interleukins for cancer chemoprevention

As the means of communication for immune cells and non-immune cells, interleukins play important roles in the occurrence and development of tumors. Their importance as a target and therapeutic agent is demonstrated by the growing number of clinical trials that are presently ongoing. While the results of clinical trials are not satisfactory. Interleukin-6 monoclonal antibodies have been studied in phase 2 and 3 clinical trials in ovarian and renal cancer [274]. Anti-interleukin-6 receptor antibody-like tocilizumab successfully treats cachexia in lung cancer patients [275]. Treated with IL1R antagonists enhances the antitumor effect of gemcitabine and 5-fluorouracil [276]. More details regarding the function of interleukins and underlying mechanisms in cancer are provided in Table 1.

Activating interferons for cancer chemoprevention

As the most effective and widely distributed family of cytokines, IFNs are induced by nucleic acids and related to immunity and vascular. IFNs are composed of type I, II, and III IFNs. Type I IFNs include IFN- α , β , ϵ , κ , and ω ; type II IFNs refer to IFN γ ; type III IFNs include IFN γ 1, IFN γ 2, IFN γ 3 (first called IL-28A, IL-28B, and IL-29) [277]. After stimulation, they are produced and secreted by body cells, such as T lymphocytes, B lymphocytes, macrophages, fibroblasts, and epithelial cells [278]. All IFNs activate the transcription of interferon-stimulated genes through the Janus kinase (JAK)/STAT pathway. Generally, JAK1 and JAK2 are activated by type II IFN signaling to cause STAT1 homodimerization, whereas TYK2 and JAK1 are activated by type I and III IFN signaling to cause STAT1-2 heterodimerization and IFN-stimulated gene (ISG) factor 3 (ISGF3) creation [277]. IFN α and IFN β , which belong to type I interferons, directly control the transcription of over 100 downstream genes, resulting in direct (on cancer cells) and indirect (through immune effector cells and vasculature) effects on tumors [278]. Drug development and patient assessment of interferon-directed therapies have benefited from new understandings of the endogenous and external activation of type I interferons in the tumor and its microenvironment. Modulation of the interferon system may

further reduce cancer morbidity and death when paired with other efficacious cancer treatment methods or with previous observations. Clinical trials were started by the Finnish National Red Cross and Hans Strander and Kari Cantell using partially purified IFN α , which was made from human blood donor leukocytes in Cantell's Helsinki laboratory [279]. Abundant evidence indicated the anti-tumor effects of leukocyte-derived IFN α on various metastasis solid tumors and hematological malignancies [280]. Recombinant IFN α 2 was the first human immunotherapeutic approved by the FDA for cancer [281].

However, exogenous IFN α 2 and IFN β also lead to systemic adverse effects. The discovery of the molecular and cellular aspects of pathways of induction and action of interferon, such as effects of protein products of interferon-stimulated genes (ISGs), cellular actions of interferons, and endogenous nucleic acid-induced pathways, which may provide new opportunities for enhancing the anti-tumor effect of these cytokines. For example, it has been suggested that RNA is a more powerful transcription inducer of interferons regarding RIG-I (retinoic acid-inducible gene I) or Toll-like receptors (TLR3, TLR7, and TLR8). A TLR3 agonist ARNAX facilitates the effect of immunotherapy on patients [282]. TLR 3 and 7 agonists induce a hot triple-negative breast cancer immune environment [283]. TLR7/8-agonist-loaded nanoparticles promote the polarization of tumor-associated macrophages to enhance cancer immunotherapy [284]. Besides, cytoplasmic protein cyclic guanosine monophosphate (GMP)-adenosine monophosphate (AMP) (cGAMP) synthase (cGAS) binds to DNA and initiates the synthesis of cGAMP, activating the stimulator of interferon genes (STING), then activates the endogenous interferon system [279], which provides a fresh understanding of how immune effector cells' endogenous interferon system is activated. And the cGAS-STING pathway plays a fundamental and vital function in identifying immunogenic cancer cells. When the intracellular STING protein is activated, various immunostimulatory molecules are produced, which can lead to the maturation of dendritic cells, the polarization of anti-tumor macrophages, the priming and activation of T cells, the activation of natural killer cells, vascular reprogramming,

(See figure on next page.)

Fig. 2 The the pro-tumor effect of these inflammatory molecules are summarized. **A** Inflammatory molecules which are involved in promoting proliferation, invasion, migration and metastasis are summarized. **B** Inflammatory molecules which contributes to angiogenesis, epithelial-mesenchymal transition, lymph angiogenesis and drug resistance are summarized. **C** Inflammatory molecules which participates in promoting tumor-related inflammation, malignant progress and tumor recurrence, inhibiting cancer cell death, increasing the motility of cancer cells, promoting tumor growth, enhancing radiation resistance and promoting tumorigenesis. **D** Inflammatory molecules which contribute to promoting osteolysis and immunosuppression, enhancing tumor immune tolerance, promoting the occurrence of precancerous lesions, culture promoting tumor microenvironment, promoting immune-escape, enhancing adhesion and promoting the stemness of cancer cells

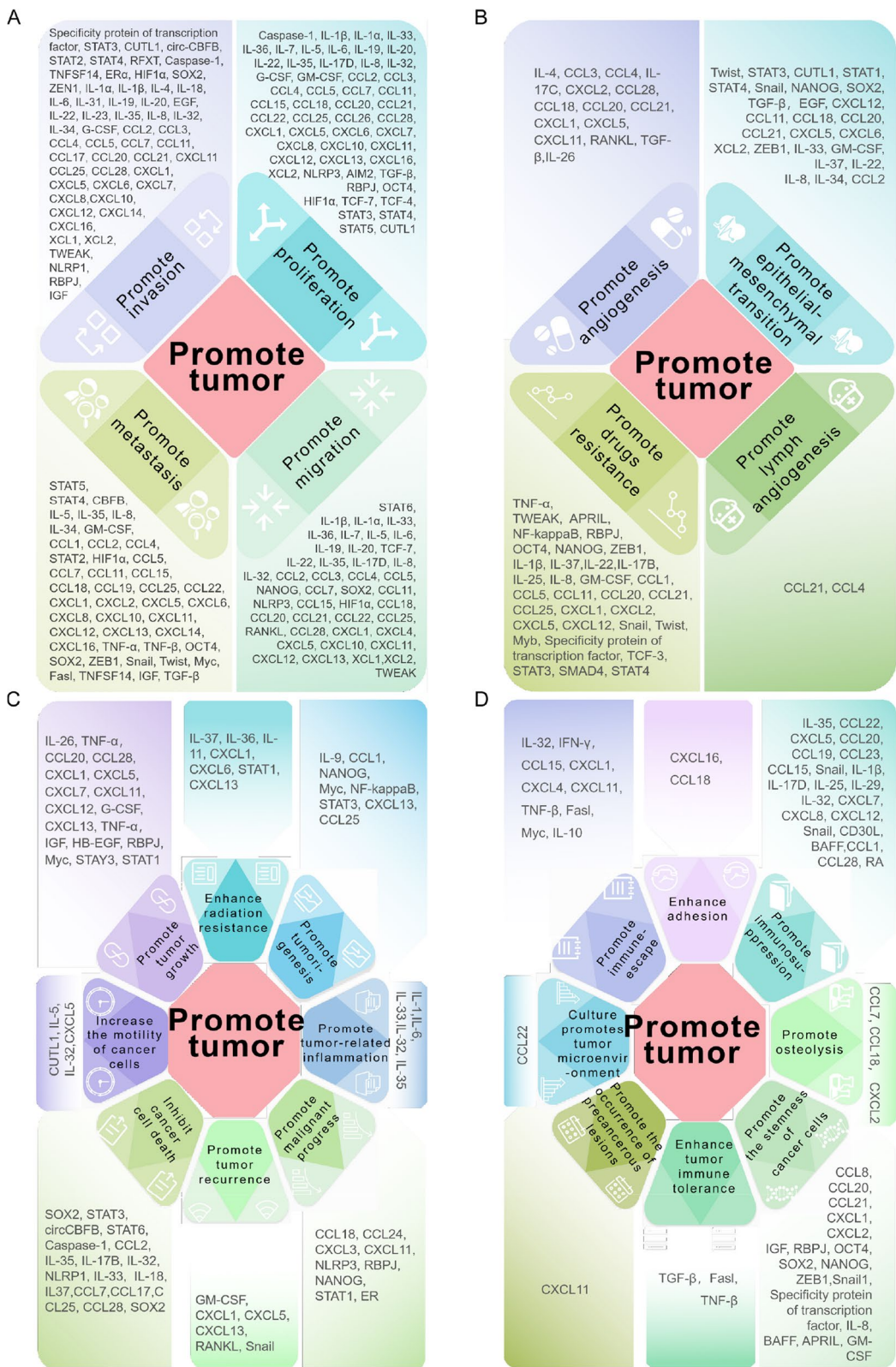


Fig. 2 (See legend on previous page.)

and/or the death of cancer cells. These processes can result in the immune system eliminating tumors and producing anti-tumor immune memory [285]. STING inhibits the reactivation of dormant metastasis in lung adenocarcinoma [286]. STING agonist promotes CAR T cell trafficking and persistence in breast cancer [287]. cGAS-STING-mediated type I interferon signaling enhances the development of stem cell-like CD8⁺T cells by inhibiting Akt activity [288]. The combination of STING agonists with radiotherapy or chemotherapy can enhance the antitumor effect and reduce the side effects caused by conventional treatments [289, 290]. Moreover, co-administering STING agonists with CTLA4 and PD1 antibodies demonstrated a significant survival advantage in a preclinical model of HPV+oral tumors [291]. The combination of the STING agonist DMAXX and CAR T cell therapy significantly increases the number of CAR T cells [287]. Recent studies have improved our understanding of the cGAS-STING pathway in cancer treatment, but prolonged STING activation can promote carcinogenesis. Analysis of the TCGA database shows that several cGAS-encoding genes are significantly upregulated in malignant tissues compared to normal controls, indicating that cGAS-STING signaling may be active in these cancers [292]. Current research indicates that STING can increase the expression of the immune checkpoint indoleamine-2,3-dioxygenase (IDO), which may directly or indirectly reduce T-cell function and numbers, thereby promoting immunological escape [293]. The enzyme cGAS, found in mitochondria, suppresses ferroptosis and promotes the progression of hepatocellular carcinoma [294]. These reports suggest that cGAS may be a potential target for cancer interventions. The inhibitory effects of astin C, a novel STING-specific inhibitor, on Trex1/BMDMs highlight the potential of astin C for cancer treatment [295]. Further research is necessary to evaluate the effectiveness of cGAS-STING pathway inhibition in cancer treatment.

The role of IFNs in cancer is complex, based on the time, cells present, total IFN-I signal levels, and the IFN α/β sub-types mediating the effects, frequently producing different results. Additionally, it's becoming evident that the timing of IFN-I delivery or blockade can

have radically different outcomes, illuminating the complex biology at play. More details regarding the function of TLRs and STING and the role of their agonists in cancer are provided in Table 2 and Table 3 respectively.

Targeting TNFs for cancer chemoprevention

In the late 1970s, macrophages were discovered to create a cytokine called TNF, also known as TNF α , which can inhibit tumor cell growth and cause tumor regression [296]. Lymphotoxin, derived from lymphocytes, has a 50% homologous amino acid sequence and binds to the same receptor with TNF α , it came to be called TNF β [297]. The TNF superfamily, composed of 19 ligands and 29 receptors, participates in multiple biological functions. Since their role in inflammation, apoptosis, proliferation, invasion, angiogenesis, metastasis, immune, and others, TNF superfamily members were promising targets for drug development. Numerous investigations showed TNF superfamily members to be a kind of strong inflammatory cytokines that both stimulate complicated immune responses and have anti-tumor properties. TNF superfamily actions can be both advantageous and dangerous. On the one hand, TNF acts as the major mediator of cancer-related inflammation, and many antagonists against the TNF family and their receptors have been approved by the FDA, and some of these are already undergoing clinical testing. For example, anti-TNF treatment (infliximab) enhanced the effects of chemotherapy in colon cancer treatment [298]. On the other hand, TNF can cause cancer cell death, which makes it a possible cancer therapy. As the first cytokine to be employed for cancer treatment, TNF α has been used in the clinic for the treatment of soft tissue sarcoma [35] and melanoma [36]. However, reducing TNF's toxicity is a major task before TNF can be administered consistently. The advancement of TNF- α therapy in the future will depend on reducing systemic therapy's toxicity and raising TNF- α dosages to increase the direct tumor response. On the other hand, through indirect tumor effects, the use of innovative modes of action may improve safety and efficacy. More details regarding the function of interleukins in cancer are provided in Table 4.

(See figure on next page.)

Fig. 3 The anti-tumor effect of these inflammatory molecules are summarized. **A** Inflammatory molecules which are involved in enhancing radiation efficacy, suppressing migration and malignant progression of tumor, inhibiting angiogenesis, lymph angiogenesis and stemness, promoting synergistic apoptosis and enhancing drug sensitivity are summarized. **B** Inflammatory molecules which contributes to inhibiting inflammation, promoting immunity and tumor cell lysis, preventing immune escape, reducing epithelial-mesenchymal transition and promoting cancer cell apoptosis. **C** Inflammatory molecules which participates in restraining movement, suppressing occurrence, driving cell senescence, inhibiting adhesion and driving non-apoptotic cellular death. **D** Inflammatory molecules which contribute to suppressing invasion, inhibiting metastasis, restraining growth and inhibiting proliferation

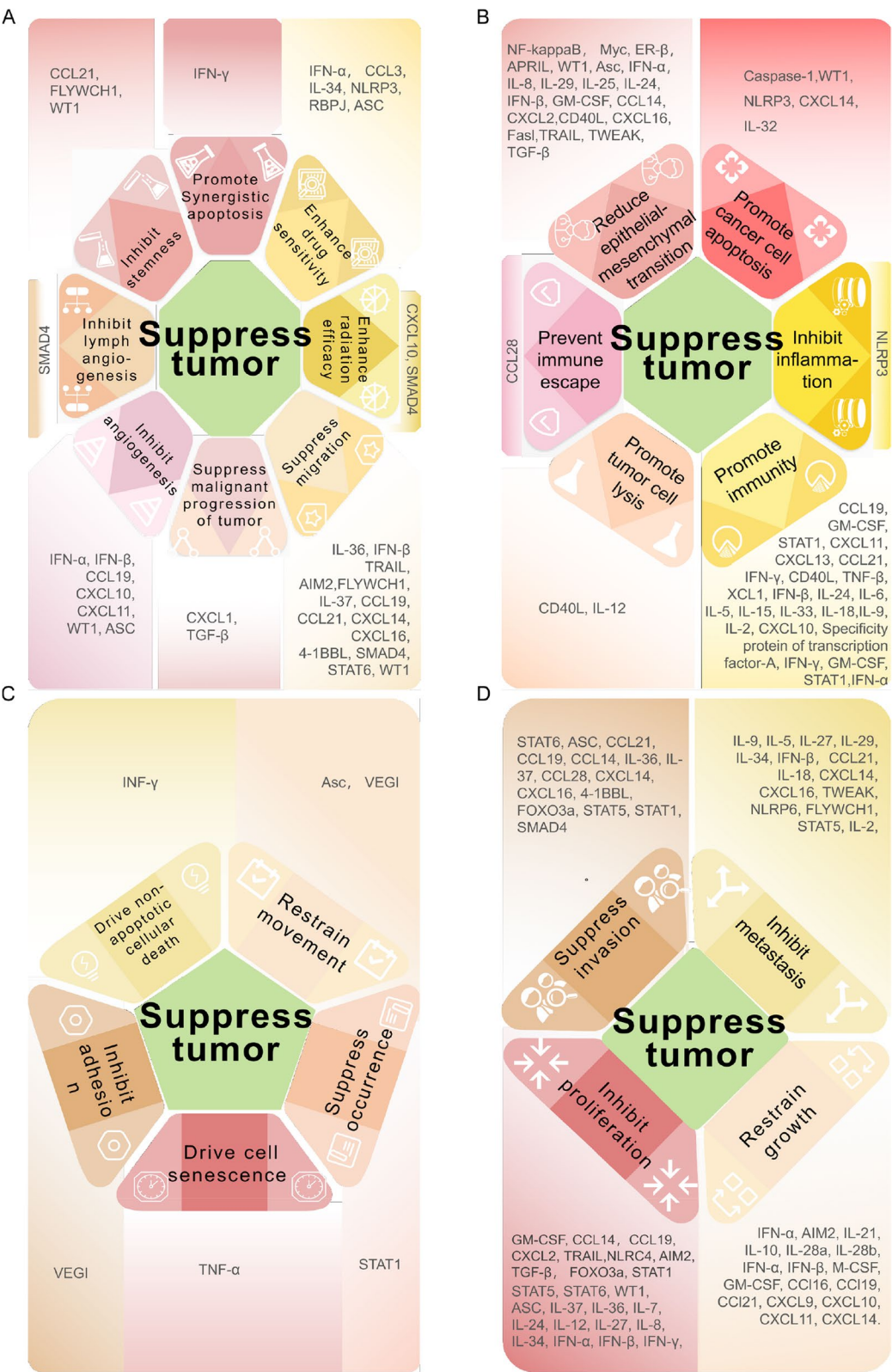


Fig. 3 (See legend on previous page.)

Table 1 The role and mechanisms of interleukins in cancer

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|-------------|---|------------------------------|-----------------|--|------------------------|
| IL-37 | IL-1R8, IL-1R5 | Colon cancer | Promote | Promotes colitis-related carcinogenesis through SIGIRR-mediated cytotoxic T cell dysfunction | Wang Z et al., 2022 |
| | | Pancreatic cancer | Promote | Drives gemcitabine resistance through negative feedback signal of IL-37/STAT3/HIF-1 α | Zhao T et al., 2020 |
| | | Skin cancer | Promote | Inhibits tumor immune surveillance by regulating CD103(+) DCs and establishing a relationship between metabolism and immunity | Ceng F et al., 2023 |
| | | Gallbladder cancer | Inhibit | Inhibits the EMT induced by HIF-1 α | Wu T et al., 2018 |
| | | Renal cancer | Inhibit | Inhibits IL-6/STAT3 signal transduction | Jiang Y et al., 2015 |
| | | Cervical cancer | Inhibit | Inhibits proliferation and invasion by suppressing STAT3 | Wang S et al., 2015 |
| | | Lung cancer | Inhibit | Inhibits migration, invasion and proliferation, and promotes apoptosis through IL-6/STAT3 pathway and Bcl-2, NEDD9 and cyclin D1 | Deng Y et al., 2018 |
| | IL-36 α , IL-36 β and IL-36 γ | Oral squamous cell carcinoma | Promote | Stimulates proliferation of cells with high IL-36R expression, and promotes migration of cells with low IL-36R expression | Li Z et al., 2024 |
| | | Colorectal cancer | Promote | Induces proliferation by promoting the expression of different genes involved in the IL-17/IL-23 axis | Baker J et al., 2023 |
| IL-2 | sIL-2R α , IL-2/IL-15R β - γ C, IL-2R α , IL-2/IL-15R β - γ C | Liver cancer | Inhibit | Inhibits the proliferation activity and migration of HCC in vitro | Song Y et al., 2023 |
| | | Gastric cancer | Promote | Mediates the impairment of T cell function | Tsubono M et al., 1990 |
| | | Pancreatic cancer | Inhibit | Inhibits tumor growth by enhancing the immune function of spleen lymphocytes | Zhang J et al., 2009 |
| | | | | Inhibits lymph node metastasis by suppressing the expression of VEGF-D mRNA | Tang R et al., 2009 |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|----------------|---|---------------------------|-----------------|--|------------------------|
| IL-4 | IL-4Ra-γC, IL-4Ra, IL-13Ra1 | Pancreatic cancer | Promote | Promotes cancer progression, invasion and angiogenesis by enhancing the ability of TIM-1-derived cathepsin | Shi J et al., 2021 |
| | | Lung cancer | Promote | Increases cells invasion, migration and vascular remodeling by promoting the differentiation of M0 into M2 macrophages | Zhag Y et al., 2024 |
| | | Liver cancer | Promote | Enhances the radiation resistance by activating ERK pathway | Liu Y et al., 2023 |
| | | prostatic cancer | Promote | Stimulates invasion and migration through AKT/NF-κB pathway | Qv H et al., 2016 |
| IL-7 | IL-7Ra-γC, sIL-7Ra | Breast cancer | Inhibit | Activates CD8 + T cells and stimulates IFNγ- secretion | Yuan C et al., 2014 |
| | | Hepatocellular carcinoma | Inhibit | Reshapes the immune system by improving T cell function and antagonizing immunosuppression network | Zhang S et al., 2024 |
| | | Colon cancer | Inhibit | Amplifies TIL in tumor lesions | Maeurer M et al., 1997 |
| | | Non-small cell lungcancer | Inhibit | Reduces tumor proliferation by changing cell surface molecular expression and enhancing anti-tumor reactivity | Sharma S et al.,1996 |
| IL-9 | IL-9R-γC | Liver cancer | Promote | Increases proliferation by driving the expression ofCCL20 and STAT3; induces the occurrence and metastasis through AKT, β catenin and vimentin | Gerlach K et al., 2019 |
| | | Lung cancer | Promote | Promotes autonomous cells growth, malignant cell transformation and better adhesion through JAK/STAT3 | Gerlach K et al., 2019 |
| | | Bladder cancer | Promote | Enhances tumor growth | Pajulas A et al., 2023 |
| | | | Promote | Promotes tumor immune escape by reducing the cytotoxicity of CD8(+) T cells and NK cells | Zhou Q et al., 2020 |
| IL-15 IL-21 | IL-15, IL15Ra+IL-2, IL-15Rβ-γC IL-21R-γC | Gastric cancer | Inhibit | Enhances the function of CD8(+) T cells | Fang H et al., 2020 |
| | | Breast cancer | Inhibit | Eliminates the metastatic potential bycontrolling extracellular matrix remodeling and cell contractility | Das S et al., 2021 |
| | | Colon cancer | Inhibit | Enhances the cytotoxicity ofTIL | Chen Z et al., 2010 |
| | | Lung cancer | Inhibit | Inhibits tumor growth by increasing the cytotoxicity of NKG2D CAR-NK cells | Zhang Y et al., 2024 |
| IL-3 | IL-3Ra-βC | Pancreatic cancer | Inhibit | Mediates CD4(+) T-eff reaction in IL-3 (-/-) | Zaidi N et al., 2019 |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|-------------|--|-------------------|-----------------|--|------------------------|
| IL-5 | IL-5Ra-βc | Bladder cancer | Promote | Enhances migration and invasion through MMP-9/NF-κB/AP-1 pathway mediated by ERK1/2 | Li O et al., 2013 |
| | | Colon cancer | Promote | Promotes cell growth by enhancing the effect of IGF-II | Makins R et al., 2005 |
| | | Esophageal cancer | Promote | Acts as an PAX2 metastasis effector | Liu P et al., 2015 |
| | | Lung cancer | Inhibit | Blocks metastasis by reducing endothelial barrier permeability through Cd3 T cells | Li F et al., 2020 |
| | | Rectal cancer | Inhibit | Contributes to host defense and releases their toxic granular proteins by activating eosinophils | Tajima K et al., 1998 |
| IL-6 | IL-6Ra-p130 (classic), sIL-6Ra-gp130 (trans) | Pancreatic cancer | Promote | Stimulates the migration and activation via STAT5 signaling | Gitto S et al., 2020 |
| | | Prostatic cancer | Promote | Promotes cell growth through JAK-STAT signaling | Lou W et al., 2000 |
| | | | | Promotes the occurrence by providing Th2 cytokine environment and up-regulating genes related to cell proliferation and angiogenesis | Feurino L et al., 2007 |
| | | Breast cancer | Promote | Inhibits apoptosis and drives cell proliferation and invasion through JAK-STAT signaling | Manore S et al., 2022 |
| | | Ovarian cancer | Promote | Promotes adhesion and invasion through Ras/MEK/ERK and PI3K/AKT pathways | Wang D et al., 2016 |
| IL-11 | IL-11Ra-p130 (classic), sIL-11Ra-gp130 (trans) | Breast cancer | Promote | Promotes the development of BrCAIL-6 by down-regulating HIC1 through paracrine or autocrine signal | Sun X et al., 2020 |
| | | Colorectal cancer | Inhibit | Promotes the activities of macrophage and lymphokine-activated killer cell | Turano M et al., 2021 |
| | | Cervical cancer | Promote | Mediates radiation-resistance through PI3K/AKT signaling pathway | Sun R et al., 2021 |
| | | Pancreatic cancer | Promote | Activates AKT ERK and STAT3 signaling | Jaclyn K et al., 2014 |
| | | Gastric cancer | Promote | Increases the activation of STAT3 | Zhou R et al., 2019 |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|-------------|-----------------------------------|--------------------------|-----------------|--|-------------------------|
| IL-31 | IL-31Ra–OSMRβ | Liver cancer | Promote | Drives hepatocytes to develop into LCSC by obtaining dryness and stimulates their growth and malignant progress | Yuan C et al., 2021 |
| | | Breast cancer | Inhibit | Increases the activity of cytotoxic T cells and decreases the levels of CD4(+) T cells, MDSC and tumor-associated macrophages | Kan Tet al., 2020 |
| IL-10 | IL-10Ra,IL-10Rβ | Lung adenocarcinoma | Promote | Down-regulates STAT1 activity by enhancing the expression of SOCS1 and SOCS3 induced by IFN-γ | Gao Y et al., 2020 |
| | | Gastric cancer | Promote | Drives immunity to escape from microenvironment | Zhang H et al., 2022 |
| | | Carcinoma of colon | Inhibit | Inhibits cell growth by suppressing STAT3 pathway | Won D et al., 2011 |
| IL-19 | IL-20Ra,IL-20Rβ | Salivaryadenocarcinoma | Inhibit | Induces TNF-driven apoptosis | Skrypnyk M et al., 2024 |
| | | Breast cancer | Promote | Enhances tumor development and affects the clinical outcome through JAK/STAT signaling | Sofi Set al., 2023 |
| | | Lung cancer | Promote | Directly promotes tumor proliferation, migration and indirectly provides microenvironment for tumor progress | Chen Y et al., 2013 |
| IL-20 | IL-20Ra,IL-20Rβ, IL-22Ra1,IL-20Rβ | Lung cancer | Promote | Enhances proliferation by stimulating-gIL20Rβ expression and activating JAK1/STAT3 signaling | He Y et al., 2022 |
| | | Oral cancer | Promote | Increases proliferation, migration, ROS production and colony formation by promoting TNF-α,IL-1β, MCP-1, CCR4 and CXCR4 expression through activating STAT3and AKT/JNK/ERK signaling | Xu Y et al., 2012 |
| | | Prostatic cancer | Promote | Increases migration and colony formation by activating p38, ERK1/2, AKT and NF-κB signals | Xu Y et al., 2015 |
| | | Breast cancer | Promote | Enhances proliferation and migration by up-regulating MMP-9, MMP-12, cathepsin k and G | Xu Y et al., 2012 |
| | | Bladder cancer | Promote | Promotes the migration through MMP-9 protein mediated by ERK | Li S et al., 2013 |
| | | Hepatocellular carcinoma | Promote | Promotes tumor progress by inducing TGFβ and MMP-9 expression, and phosphorylating JNK/STAT signaling | Ding W et al., 2018 |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|-------------|--|----------------------------|-----------------|--|-------------------------|
| IL-22 | IL-22Ra1, IL-10Rβ, IL-2Ra2 (also known as IL-22BP) | Lung cancer | Promote | Increases proliferation by activating STAT3 and enhancing the expression of anti-apoptotic B-cell lymphoma 2 | Kobold S et al., 2013 |
| | | | | Promotes tumor cell survival and drug resistance by up-regulating anti-apoptosis proteins | Zhang W et al., 2008 |
| | | Gastric cancer | Promote | Promotes invasion through STAT3 and ERK activation | Fukui H et al., 2014 |
| | | | | Promotes migration and invasion through IL-22R1 /AKT/MMP-9 | Ji Y et al., 2014 |
| | | Breast cancer | Promote | Promotes proliferation in a STAT3-dependent manner | Zhang Y et al., 2020 |
| | | Ovarian cancer | Promote | Promotes tumor development Through STAT3 signaling | Lei B et al., 2018 |
| IL-24 | IL-20Ra,IL-20Rβ, IL-22Ra1, IL-20Rβ | Colorectal cancer | Promote | Promotes proliferation through STAT3 signaling | Wu T et al., 2013 |
| | | Non-small cell lung cancer | Promote | Confers EGFR-TKI resistance through AKT and ERK signaling | Wang X et al., 2019 |
| | | Pancreatic cancer | Promote | Promotes proliferation, invasion and migration by stimulating AKT signal transduction | Wang X et al., 2020 |
| | | Lung cancer | Inhibit | Inhibits proliferation and promotes apoptosis | Qi Q et al., 2014 |
| | | Pancreatic cancer | Inhibit | Induces apoptosis and CTL to kill cancer cells and produces anti-tumor immunity | Xv B et al., 2014 |
| | | Breast cancer | Inhibit | Promotes apoptosis and cell arrest in G2/M phase through PI3K/β-catenin signaling | Deng L et al., 2020 |
| IL-26 | IL-20Ra,IL-10Rβ | Colorectal cancer | Inhibit | Inhibits cell growth by inducing tumor-lysis and apoptosis, and stimulating immunity | Deng L et al., 2020 |
| | | Endometrium cancer | Inhibit | Inhibits proliferation by promoting apoptosis through mitochondrial intrinsic signal pathway | Liao S et al., 2020 |
| | | Non-small cell lung cancer | Promote | Increases net angiogenic activity and tumor growth by promoting CXCR-2 dependent angiogenesis | Numasaki M et al., 2005 |
| | | Breast cancer | Promote | Dephosphorylates and down-regulates EphA3 and phosphorylates endoplasmic reticulum induced by EGFR-TKI via AKT and JNK | Itoh T et al., 2021 |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|---|---|-------------------|-----------------|---|---------------------------|
| IL-12 | IL-12Rβ1, IL-12Rβ2 | Lung cancer | Inhibit | Enhances the cytolytic activity of PBMC on lung cancer cells | Hiraki A et al., 2002 |
| | | Ovarian cancer | Inhibit | Inhibits cell proliferation | Wang J et al., 2006 |
| IL-27 and IL-30 (also known as IL-27 subunit p28) | IL-7Rα (also known as WSX1)–gp130 | | | Promotes the self-renewal of CD133(+) cancer stem cell-like cells | Wang D et al., 2017 |
| | | Oral cancer | Promote | Promotes proliferation promoting by nuclear trans-activation of RelA | Fukuda M et al., 2010 |
| | | Lung cancer | Inhibit | Reduces proliferation and metastasis through miR-935 | Wang T et al., 2019 |
| | | Ovarian cancer | Inhibit | Inhibits proliferation by enhancing STAT3 and inhibiting AKT signaling | Zhang Z et al., 2016 |
| | | Cervical cancer | Inhibit | Restricts angiogenesis by paracrine | Zhang B et al., 2017 |
| | | Prostatic cancer | Inhibit | Inhibits tumor growth and improves the survival rate of patients | Sorrentino C et al., 2019 |
| | | Breast cancer | Promote | Promotes invasion and metastasis | Wang A et al., 2018 |
| IL-35 | IL-12Rβ2–gp130, IL-12Rβ2, IL-12Rβ2 gp130–gp130, IL-27Rα, IL-12Rβ2 | | | Promotes tumor progression by inhibiting proliferation of infiltrating T-conv cells and inducing iTi35 cells | Hao S, et al., 2018 |
| | | Colorectal cancer | Promote | Inhibits proliferation of T cells and may participate in tumor immune tolerance through STAT1 and STAT3 | Ma Y et al., 2016 |
| | | Lung cancer | Promote | Inhibits CD4 T cell-mediated immune response | Hao Y et al., 2022 |
| | | | | Promotes tumor progression by inducing T cell differentiation | Zhou A et al., 2021 |
| | | Prostatic cancer | Promote | Promotes cell proliferation, tumor angiogenesis and limits the antitumor immune response by increasing the ratio of Tregs to MDSC and decreasing the ratio of CD4+ and CD8+ T cells | Zhu J et al., 2020 |
| | | Pancreatic cancer | Promote | Promotes tumor growth by enhancing proliferation and inhibiting apoptosis | Nicholl M et al., 2014 |
| | | Colon cancer | Inhibit | Inhibits migration, invasion, proliferation, colony formation and cancer stem cells by inhibiting β-catenin | Zhang J et al., 2017 |
| | | Lung cancer | Promote | Promotes tumor growth and progresses through the coordination of immune cells (i.e. macrophages) | Ferreira N et al., 2020 |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------------------------|-----------------|------------------------------|--------------------|--|--|
| IL-17B | IL-17RB | Breast cancer | Promote | Promotes tumor resistance to paclitaxel by activating ERK1/2 pathway | Laprevotte E et al., 2017 |
| | | | | Promotes tumor occurrence through NF-κB mediated anti-apoptosis pathway | Huang Z et al., 2014 |
| | | Lung cancer | Promote | Promotes metastasis by activating ERK/β-catenin | Yang Y et al., 2018 |
| | | Pancreatic cancer | Promote | Promotes invasion and the survival of cancer cells through ERK1/2 | Wu H et al., 2015 |
| IL-17C | IL-17RA,IL-17RE | Gastric cancer | Promote | Enhances proliferation and migration through IL17B activated mesenchymal stem cells | Bi Q et al., 2017 |
| | | Lung cancer | Inhibit | Activates IL-17RB/AKT/β-catenin pathway | Bi Q et al., 2016 |
| | | | | Increases the expression of neutrophil chemokine, keratinocyte derived chemokine and macrophage inflammatory protein 2 induced by NTHi and TNF-α | Junnickel Cet al., 2017 |
| | | Colorectal cancer | Promote | Promotes cancer development by improving survival rate | Song X et al., 2014 |
| IL-17D | Unknown | | | Promotes angiogenesis by producing VEGF through STAT3/miR-23a-3p/SEMA6D axis | Li Y et al., 2020 |
| | | Lung cancer | Promote | Promotes tumor progression via p38 MAPK signaling pathway | Li Z et al., 2022 |
| | | Ovarian cancer | Promote | Promotes cell growth by changing of MICA expression level | Zhang J et al., 2014 |
| | | | | Accelerates cell proliferation and enhances migration and invasion by activating NF-κB | Fan Y et al., 2024 |
| IL-25 (also known as IL-17E) | IL-17RA,IL-17RB | Breast cancer | Promote | Exerts immuno-suppressive effect by producing IL-17D | Ruan J et al., 2021 |
| | | Lung cancer | Promote | Promotes cisplatin resistance by increasing the major fornx proteins through activating NF-κB | Shen W et al., 2019 |
| | | Colorectal cancer | Promote | Maintains tumor infiltrating MDSC by promoting IL-C2 | Jou E et al., 2022 |
| | | Breast cancer Lung cancer | Inhibit Inhibit | Activates caspase-mediated apoptosis Inhibits growth and induces apoptosis through STAT1 phosphorylation | Furuta S et al., 2022 Tezuka Y et al., 2012 |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|----------------------------|----------------------------|-------------------|-----------------|---|--------------------------|
| IL-29 | IL-28Rα, IL-10Rβ | Multiple myeloma | Promote | Activates STAT1 and STAT3 | Novak A et al., 2008 |
| | | Pancreatic cancer | Inhibit | Overexpresses P21 and Bax | Balabanov D et al., 2019 |
| | | Cervical cancer | Inhibit | Inhibits proliferation and promotes apoptosis by up-regulating the expression of TRAILR1 | Ha L et al., 2024 |
| IL-8 (also known as CXCL8) | CXCR1, CXCR2 ACKR1/DARC | Skin cancer | Inhibit | Increases MHC class 1, P21 and Rb protein | Romee R et al., 2014 |
| | | Lung cancer | Inhibit | Promotes cell arrest and apoptosis by up-regulating p21 through STAT | Barrera L et al., 2015 |
| | | Gastric cancer | Inhibit | Decreases Bcl-2 and caspase cascade | Gao Z et al., 2014 |
| | | | | Induces a possible NK cell-mediated immune response | Bu X et al., 2014 |
| | | Colorectal cancer | Inhibit | Increases NK and NKT cell activity | Aulino P et al., 2010 |
| | | Esophageal cancer | Inhibit | Increases MHC class 1, P21 and Rb protein | Li Q et al., 2010 |
| | | Breast cancer | Promote | Promotes invasion and migration by promoting Wnt/β-catenin signaling | Mou C et al., 2018 |
| | | Ovarian cancer | Promote | Enhances invasion and migration by promoting EMT | Wang S et al., 2018 |
| | | | | Stimulates cell adhesion and invasion by activating PI3K/AKT and Raf/MEK/ERK signaling and increasing MMP-2 and MMP-9 activity and expression | Niu X et al., 2013 |
| | | Bladder cancer | Promote | Improves drug resistance by maintaining the characteristics of cancer stem cells | Zhu K et al., 2014 |
| | | Renal carcinoma | Promote | Promotes EMT through PKC/ERK signaling | Bi L et al., 2012 |
| | | Pancreatic cancer | Promote | Promotes invasion by regulating MMP-2 activity | Kuwada Y et al., 2003 |
| | | Gastric cancer | Promote | Promotes metastasis through c-Jun and Ets-1 | Chen H et al., 2017 |
| | | Renal carcinoma | Promote | Induces migration by activating AKT signaling through CXCR2 | Bi L et al., 2014 |
| | | Ovarian cancer | Promote | Induces chemo-resistance by increasing the expression of MDRI, Bcl-2, Bcl-xL and XIAP, and activating Raf/MEK/ERK and PI3K/AKT signaling | Niu X et al., 2012 |
| | | Cervical cancer | Promote | Promotes proliferation, invasion and migration by activating ERK and up-regulating MMP-9 | Ye K et al., 2022 |
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Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|---------------------------|----------------------------|-------------------------------|-----------------|--|-------------------------|
| IL-13 | IL-13Rα1, IL-4Rα, IL-13Rα2 | Prostate cancer | Inhibit | Promotes the carcinogenic potential by increasing the expression of IL-RA, IL8RB and ERK and decreasing the expression of NUMB | Jia L et al., 2018 |
| | | Pancreatic cancer | Promote | Promotes proliferation and inhibits apoptosis through STAT3/AKT/NF-κB pathway | Guo Y et al., 2017 |
| | | Colon cancer | Promote | Promotes proliferation by enhancing p44/42 MAPK (ERK1/2) phosphorylation and tyrosine and PI3 kinase activity | Shi J et al., 2021 |
| IL-14α and IL-14β | IL-14R | N/A | Promote | Promotes malignancy by inducing the expression of 11βHSD2 in an IL-13Rα2-dependent manner | Jiang L et al., 2016 |
| | | | | Promotes tumor progression by recruiting immune cells infiltrating into tumors | Richmond J et al., 2014 |
| IL-16 | CD4 | Breast cancer | Promote | Contributes to the implantation of tumor cells into lung parenchyma | Donati K et al., 2017 |
| | | Lung cancer | Promote | Promotes immune escape by developing immuno-suppressive microenvironment | Zhao S et al., 2024 |
| | | Cervical cancer | Promote | Promotes tumor progress by forming a positive regulatory loop with NF-κB/miR-205 | Liu J et al., 2024 |
| IL-32 (also known as Nk4) | Unknown | Pancreatic cancer | Promote | Regulates downstream molecules and promotes invasion | Takagi K et al., 2021 |
| | | Gastric cancer | Promote | Increases invasion | Tsai C et al., 2014 |
| | | Colon cancer | Promote | Creates a favorable environment for tumor growth by up-regulating IL-8, TNF and CCL2 | Catalán V et al., 2017 |
| | Unknown | Esophageal cancer | Promote | Induces polarization of M2 macrophages via FAK/STAT3 pathway | Sun Y et al., 2022 |
| | | Triple negative breast cancer | Promote | Increases migration and invasion through EMT by up-regulating VEGF-STAT3 pathway | Park H et al., 2012 |
| | | Thyroid cancer | Promote | Contributes to cell survival by inducing cytokine IL-8 | Sloot Y et al., 2019 |
| | Unknown | Multiple myeloma | Promote | Produces immuno-suppression and allows tumor growth through NF-κB pathway | Yan H et al., 2019 |
| | | Gastric cancer | Promote | Inhibits autophagy through PI3K/AKT/mTOR signaling | Wang X et al., 2022 |
| | | | | | |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|-------------|----------|---------------------|-----------------|---|-------------------------|
| | | Lung adenocarcinoma | Promote | Promotes migration and invasion by up-regulating NF-κB | Zeng Q et al., 2014 |
| | | Osteosarcoma | Promote | Stimulates invasion and movement by activating AKT and up-regulating MMP13 | Zhou Y et al., 2015 |
| | | Liver cancer | Promote | Inhibits apoptosis and increases cell survival by activating NF-κB and p38/ MAPK pathways | Kang H et al., 2012 |
| | | Gastric cancer | Promote | Promotes tumor progression by increasing metastasis through activating AKT, β-catenin and HIF-1α | Cai C et al., 2014 |
| | | Liver cancer | Promote | Stimulates cell survival and growth by activating and maintaining NF-κB | Han X et al., 2019 |
| | | Lymphoma | Promote | Stimulates cell proliferation by activating MAPK and NF-κB | Hiraku S et al., 2013 |
| | | Colon cancer | Inhibit | Inhibits tumor development by promoting the death signal of TNFR1 | Yun M et al., 2015 |
| | | | | Inhibits dryness and EMT by suppressing STAT3-ZEB1 pathway | Bak Y et al., 2016 |
| | | Colorectal cancer | Inhibit | Enhances TNFα-mediated apoptosis by up-regulating p32-MAPK signaling | Yun M et al., 2023 |
| | | Thyroid cancer | Inhibit | Induces caspase-mediated apoptosis | Heinhuis B et al., 2015 |
| | | Melanoma | Inhibit | Inhibits proliferation and increases apoptosis by up-regulating p21, p53, and TRAILR1 | Nicholl M et al., 2016 |
| | | Pancreatic cancer | Inhibit | Reduces EMT by inhibiting JAK/ST T3 signal and the expression of EMTmarkers and MMP2, 9 and 7 | Bak Y et al., 2016 |
| | | Skin cancer | Inhibit | Improves survival rate by increasing the number of tumor-specific CD8 + T cells | Gruber T et al., 2020 |
| | | Bladdercancer | Inhibit | Inhibits tumor growth by enhancing the cytotoxicity of NK-92 | Wu K et al., 2022 |
| | | Breast cancer | Inhibit | Increases proliferation and decreases apoptosis | Wang S et al., 2015 |
| | | Cervical cancer | Inhibit | Inhibits tumor development by inducing the production of pro-inflammatory cytokines and down- regulating E7 and COX2 through negative feedback loop | Lee S et al., 2011 |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|-------------|---|-------------------|-----------------|---|----------------------------------|
| IL-34 | CSF1R | Mastadenoma | Promote | Promotes epithelial cell transformation | Poudel M et al., 2021 |
| | | Gastric cancer | Promote | Promotes proliferation and EMT | Li C et al., 2022 |
| | | Colon cancer | Inhibit | Inhibits cell proliferation and enhances the susceptibility of cells to oxaliplatin-induced death by suppressing ERK1/2 | Franze E et al., 2018 |
| IL-1α | IL-1R1, IL-1R3 sIL-1R3 | Liver cancer | Inhibit | Inhibits tumor growth and metastasis by inhibiting proliferation and EMT | Tian B et al., 2023 |
| | | Gastric cancer | Promote | Increases the percentage of S phase fraction of cells, stimulates cell proliferation | Furuya Y et al., 2000 |
| | | Breast cancer | Promote | Promotes proliferation, invasion or migration | Qiu J et al., 2021 |
| IL-1β | IL-1R1, IL-1R3 IL-1R2, IL-1R3, sIL-1R2, sIL-1R3 | Gastric cancer | Promote | Promotes invasion by activating NF-κB and MMP-9 expression | Yamanaka, N et al., 2004 |
| | | Ovarian cancer | Promote | Increases expression of IL-1β and may lead to early steps of cancer | Woolery T et al., 2014 |
| | | Cervical cancer | Promote | Promotes proliferation and migration through MEK/ERK signaling pathway | Zhang J et al., 2022 |
| IL-33 | IL-1R3, IL-1R4, sIL-1R4 | Breast cancer | Promote | Induces a cascade reaction of TP63 subtype ΔNP63α signal, and leads to cisplatin resistance | Mendoza-Rodriguez M et al., 2019 |
| | | Osteosarcoma | Promote | Enhances tumor growth by regulating NF-κB signaling and miR-376c/TGFA axis | Liu B et al., 2017 |
| | | Gastric cancer | Promote | Promotes invasion and migration by stimulating the secretion of MMP-3 and IL-6 through ST2-ERK1/2 pathway | Yu X et al., 2015 |
| | | Colorectal cancer | Promote | Promotes cell proliferation through its receptor ST2, and up-regulates COX2 through NF-κB signaling | Li Y et al., 2018 |
| | | Ovarian cancer | Promote | Promotes cell proliferation and inhibits apoptosis by down-regulating p27, Fas and TRAILR1 in vitro | Liu N et al., 2021 |
| | | Breast cancer | Promote | Promotes cell transformation and tumorigenesis | Cui H et al., 2015 |
| | | Lung cancer | Inhibit | Inhibits tumor growth and lung metastasis by promoting proliferation and activation of CD8 T cells and NK cells through NF-κB signaling | Yang K et al., 2022 |
| | | | | | |

Table 1 (continued)

| Interleukin | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|-------------|-----------------------------|-------------------|-----------------|--|------------------------|
| IL-18 | IL-1R5 IL-1R7 IL-18BP | Lung cancer | Promote | Enhances metastasis by down-regulating E-cadherin | Jiang D et al., 2003 |
| | | Pancreatic cancer | Promote | Promotes cell proliferation and movement | Sun Q et al., 2020 |
| | | Colon cancer | Inhibit | Inhibits tumor growth and prolongs the survival by blocking the secretion of TGF-β and IL-4; increasing the secretion of TNF-α, enhancing its cytotoxicity | Chen Z et al., 2010 |
| | | | | Enhances the ability of cancer cells to resist T lymphocytes by up-regulating FasL protein | Zhang W et al., 2002 |
| | | Breast cancer | Inhibit | Inhibits osteolytic bone metastasis | Nakata A et al., 1999 |
| | | Colorectal cancer | Inhibit | Promotes tumor immune surveillance by up-regulating FasL and death ligand | Dupauli J et al., 2015 |

Table 2 The role and mechanisms of Toll-like receptor (TLR) in cancer

| TLR | Expressed cell | Cancer types | Promote/inhibit | Mechanisms | References |
|--------------------------|---|----------------------|-----------------|---|---|
| TLR7 | B lymphocyte, T lymphocyte, neuron | Pancreatic cancer | Promote | Increases cell proliferation and promotes chemo-resistance | Grimmig T et al., 2015 |
| Lmiquimod (TLR7 agonist) | mature dendritic cell, macrophage | Breast cancer | Inhibit | Blocks IL-10 | Yusuf N et al., 2014 |
| | | Basal cell carcinoma | Inhibit | Modulates immune response | Stockfleth E et al., 2003 |
| TLR5 | Epithelial cell, dendritic cell, macrophage, fibroblast | Oral cancer | Promote | Promotes tumor progression | Kaupila J et al., 2013 |
| TLR4 | Endothelial cell, fibroblast, liver cell, macrophage, dendritic cell, epithelial cell | B lymphocyte | | | An |
| | | Oral cancer | Promote | Enhances invasion | Kong Q et al., 2020 |
| | | Cervical cancer | Promote | Promotes proliferation and apoptosis resistance partially through Toll-like receptor 4/NF- κ B pathway | Jiang N et al., 2018 |
| | | Breast cancer | Promote | Promotes tumor progression via TLR4/NF- κ B/STAT3 signaling | Ochi A et al., 2012 |
| | | Skin cancer | Promote | Up-regulates immunosuppressive and pro-inflammatory cytokines and chemokines | Sato Y et al., 2009 |
| | | Lung cancer | Promote | Promotes migration and counterattack of cells by inducing autophagy | Mi-Jeong K et al., 2022 |
| | | Prostate cancer | Promote | Promotes tumor cell activation, proliferation, survival and tumor transformation | Gonzalez-Reyes S et al., 2011; Huang B et al., 2009 |
| | | | | Promotes tumor development by reducing immune function | Engblom C et al., 2016; Ugel S et al., 2015 |
| | | Ovarian cancer | Promote | Contributes to tumor growth through TLR4-MyD88 signaling | Kelly M et al., 2006 |
| | | Colon cancer | Promote | Escapes from immune surveillance by inhibiting the functions of T and NK cells | Huang B et al., 2005 |
| | | Gastric cancer | Promote | Promotes tumor occurrence and progress through NF- κ B pathway | Yue Y et al., 2011 |
| | | Intestinal tumor | Inhibit | Decreases tumors induced by azoxymethane/sodium dextran sulfate | Fukata M et al., 2011 |
| | | | Inhibit | Silence of TLR4 increases metastasis, and TLR4 induces effective cancer antigen-specific cytotoxic T cell immune response | Ahmed A et al., 2013 |
| | | Prostate cancer | Inhibit | Initiates innate immunity to invasive pathogens | Kundu S et al., 2008; Gonzalez-Reyes S et al., 2011; Huang B et al., 2009 |

Table 2 (continued)

| TLR | Expressed cell | Cancer types | Promote/inhibit | Mechanisms | References |
|--|---|-------------------------|-----------------|---|---|
| Coli toxin BCG vaccine (TLR-4 agonist) | | Gastric cancer | Inhibit | Induces exfoliation and autophagy | Galluzzi L et al., 2012 |
| TLR2 | Macrophage, dendritic cell, epithelial cell, fibroblast, endothelial cell, B lymphocyte | Colon cancer | Promote | Promotes proliferation, migration, and invasion through PI3K/AKT and NF- κ B | Wang X et al., 2018 |
| | | Gastric cancer | Promote | Increases proliferation and survival of gastric epithelial cells | Liu Y et al., 2019; West A et al., 2017; Cui L et al., 2021 |
| | | | | Weakens the function of CD8 + lymphocyte | Yang H et al., 2014 |
| | | | | Promotes tumorigenesis independent of inflammation in STAT3-driven cancer | Jenkins B et al., 2012 |
| | | Breast cancer | Promote | Promotes tumor progression and resistance to chemotherapy | Di Lorenzo A et al., 2022 |
| | | Oral cancer | Inhibit | TLR2 deficiency enhances tumor susceptibility by promoting an inflammatory environment | Li B et al., 2024 |
| Polysaccharide Krestin (TLR2 agonist) | | Breast cancer | Inhibit | Has potent anti-tumor effects via stimulating both innate and adaptive immune pathways | Lu H et al., 2011 |
| Bacteria Peptidoglycan (TLR2 agonist) | | Breast Cancer | Promote | Promotes invasion and adhesion by targeting Toll-Like receptor 2 in the cancer cells | Xie W et al., 2010 |
| Coli toxin BCG vaccine (TLR2 agonist) | | Gastric cancer | Inhibit | Induces exfoliation and autophagy | Galluzzi L et al., 2012 |
| TLR8 | Monocyte, macrophage, dendritic cell, neutrophil | Pancreatic cancer | Promote | Increases cell proliferation and promotes chemo-resistance | Grimmig T et al., 2015 |
| TLR9 | Dendritic cell, B lymphocyte, macrophage, fibroblast, epithelial cell | Skin cancer | Promote | Enhances invasion and promotes proliferation through activation of NF- κ B and Cox-2 and secretion of IL-8, IL-1 α (41) and TGF- β (42) | Di J et al., 2009 |
| | | Lung cancer | Promote | Promotes tumor progression | Ren T et al., 2009 |
| TLR3 | Dendritic cell, fibroblast, macrophage, epithelial cell, B lymphocyte | Lung cancer | Promote | Promotes migration and counterattack of cells by inducing autophagy | Mi-Jeong K et al., 2022 |
| | | Oral squamous carcinoma | Inhibit | Promotes apoptosis | Luo Q et al., 2012 |

Targeting colony-stimulating factors for cancer chemoprevention

The ability to produce in vitro colonies of mature myeloid cells from bone marrow precursor cells after the

proliferation and differentiation of these cells was the initial defining characteristic of granulocyte/macrophage colony-stimulating factor (GM-CSF; also known as CSF2), macrophage colony-stimulating factor (M-CSF;

Table 3 The role and mechanisms of stimulator of interferon genes (STING) in cancer

| STING | Cancer types | Promote/Inhibit | Mechanisms | References |
|-------|---------------------------------|-----------------|--|---|
| STING | Ovarian cancer | Promote | Makes cancer-associated fibroblasts sensitive to platinum chemotherapy by inhibiting cGAS-STING pathway | Liu J et al., 2024 |
| | Colorectal cancer | Promote | Promotes proliferation and induces drug resistance by regulating AMPK-mTOR pathway | Yao H et al., 2022 |
| | Colon cancer | Promote | Activates reprogrammed tumor-associated macrophages to M1 phenotype and transforms immune cold peritoneal tumor into T cell inflammatory tumor; STING agonists cooperate with PD-1 and/or COX2 blocker to further inhibit carcinogenesis | Lee S et al., 2021 |
| | Breast cancer | Promote | Down-regulation of STING reduces cell survival rate and increases the sensitivity of genotoxicity treatment in a cell-independent way | Cheradame L et al., 2021 |
| | | | Induces cell survival and immunosuppression by IL-6-mediated STAT3 activation through NF- κ B | Vasiyani H et al., 2022 |
| | Gastric cancer | Promote/Inhibit | Knocking down STING and activating STING with 2'3'-c-GAMP promote polarization of TAMs into pro-inflammatory subtype, and induce apoptosis through IL6R-JAK-IL24 pathway | Miao L et al., 2020 |
| | | Inhibit | Inhibits proliferation, migration and immune escape by activating cGAS-STING/IFN- β | Yuan M et al., 2022 |
| | Pancreatic cancer | Inhibit | Produces type I IFN and activates T cells through CD8 α + DC | Cheng H et al., 2020 |
| | | | Constant stimulation of the cGAS-STING leads to cell death, inhibits tumorigenesis | Gulen M et al., 2017 |
| | | | Has anti-tumor impact on TME by producing type I IFN and priming T cells via CD8 α + DCs | Corrales L et al., 2017 |
| | Colorectal cancer | Inhibit | Weakens the tumorigenesis of colitis-related colorectal cancer by enhancing intestinal epithelial focal death | Gong W et al., 2022 |
| | Breast cancer | Inhibit | Promotes the anti-tumor immune response of tumor-specific CD8 + T cells | Lu Z et al., 2022 |
| | Lewis lung cancer | Inhibit | Activates the anti-tumor immune response of T cells and inhibits tumor progression | Zhang X et al., 2023 |
| | Ovarian cancer | Inhibit | Enhances the anti-tumor activity | Zhang J et al., 2020 |
| | Prostate cancer | Inhibit | Induces immune system rejection and eliminates PCa cells | Alnukhali M et al., 2024 |
| | Skin cancer | Inhibit | Induces antigen-specific reactive T cells by promoting the transcription of type I IFN through activating TBK1 and enhances the phosphorylation of IRF3 or STAT6 | Honda T et al., 2013 Sawada Y et al., 2015 |
| | | | Enhances the anti-tumor effect by combining with DNA damaging agents | Hayman T et al., 2021 Baird J et al., 2018 Liang D et al., 2015 |
| | Squamous cell carcinoma of skin | Inhibit | Promotes the activation of NK cells and DC induced by cetuximab | Lu S et al., 2018 |

Table 3 (continued)

| STING | Cancer types | Promote/Inhibit | Mechanisms | References |
|---------------------------|---------------------------------|-----------------|--|---------------------------|
| | Adult T cell leukemia/ lymphoma | Inhibit | Enhances the formation of IRF3-Bax complex and leads to the apoptosis of adult T-cell leukemia/lymphoma | Bladé J et al., 2010 |
| | Bladder cancer | Inhibit | Activates cytoplasmic pattern recognition receptor and downstream IFN1 pathway | Koti M et al., 2019 |
| DMXAA series (agonist) | Pancreatic cancer | Inhibit | Improves patients' survival rate and anti-tumor immunity by prompting T cells; reduces tumor size by activating cytolytic T cells | Jing W et al., 2019 |
| 3'3'-cGAMP (agonist) | Pancreatic cancer | Inhibit | Reduces metastasis and tumor growth, and promotes anti-tumor immune response | Lu X et al., 2020 |
| ADU-V19 type (agonist) | Pancreatic cancer | Inhibit | Enhances vaccine immunogenicity, vaccine-specific T cells and anti-tumor immune response | Kinlead H et al., 2018 |
| ADU-S100 type (agonist) | Pancreatic cancer | Inhibit | Stimulates immune response by increasing the expression of CXCR3 in T cells | Vonderhaar E et al., 2021 |
| CdGMP series (agonist) | Pancreatic cancer | Inhibit | Increases immune cells by activating APC | Lorkowski M et al., 2021 |
| | | | Activates endogenous tumor-specific lymphocytes and inhibits metastasis by activating APCs | Smith T et al., 2017 |
| IACS-8803 model (agonist) | Pancreatic cancer | Inhibit | Increases lymphoid myeloid population and strengthens checkpoint block | Ager C et al., 2021 |
| c-di-AMP (agonist) | Breast cancer | Inhibit | Induces apoptosis | Vasiyani H et al., 2021 |
| DMXAA or cGAMP (agonist) | Breast cancer | Inhibit | Enhances the therapeutic effect of Th/Tc17 CAR T cells by up-regulating CXCL9 and CXCL10 to promote the infiltration of CAR T cells into tumor tissues | Tian Z et al., 2022 |
| cGAMP (agonist) | Breast cancer | Inhibit | Inhibits tumor growth and prolongs the survival time of pancreatic cancer mouse | DaY et al., 2022 |
| ADU-S100 (agonist) | Prostate cancer | Inhibit | Inhibits tumor progression | Esteves A et al., 2021 |

also known as CSF1), and granulocyte colony-stimulating factor (G-CSF; also known as CSF3). As the main regulators of granulocyte and macrophage populations, CSF can mobilize stem cells to peripheral blood in sufficient quantities for transplantation, speed up the regeneration of protective white blood cells damaged by chemotherapy, boost anticancer immune responses, and possibly contribute to the development of myeloid leukemias [299]. More details regarding the function of colony-stimulating factors in cancer are provided in Table 5.

Targeting chemokines for cancer chemoprevention

Chemokines are 8–12 kDa proteins that are released and bind to Gai-protein-coupled seven-transmembrane-spanning receptors (GPCRs), also known as classical chemokine receptors, to control directed cell movement (chemotaxis), adhesion, cell orientation, and cell–cell interactions [300]. Comprising around 50 chemokine

ligands, 20 signaling GPCRs, and 4 ACKRs, the chemokine system is crucial for various pathological processes. Cancer cells, tissue-resident cells, and recruited immune cells that express a wide variety of chemokine ligands and chemokine receptors all influence the process of carcinogenesis. Chemokines govern the invasiveness, proliferation, and stem-like characteristics of tumor cells. They also influence neoangiogenesis, neurogenesis, and fibrogenesis in stem cells [301]. Chemokines play a crucial role in guiding immune cell movement when mounting and subsequently delivering an efficient anti-tumor immune response [300]. Meanwhile, chemokine systems also contribute to pro-tumorigenic immune responses by controlling immune cells' location and cellular interactions in lymphoid organs and the tumor microenvironment (TME). Chemokines have been attractive therapeutic targets because of their role in mediating the recruitment of anti-tumorigenic immune cells and

Table 4 The role and mechanisms of tumor necrosis factors (TNF) in cancer

| TNF | Cancer types | Promote/Inhibit | Mechanisms | References |
|---------------|----------------------------|-----------------|---|------------------------------|
| TNF- α | Gallbladder cancer | Promote | Autocrine mechanisms | Zhu, G et al.,2014 |
| | Cervical cancer | Promote | Increases the expressions of TNF- α | Li, J et al.,2018 |
| | Pancreatic cancer | Promote | Higher expression in the serum of patients with metastatic disease | Karayiannakis, A et al.,2001 |
| | Colorectal cancer | Promote | Increases distant tumor metastasis | Li, Z et al.,2017 |
| | Rectal cancer | Promote | Contributes to distant tumor metastasis | Li, Z et al.,2017 |
| | Breast cancer | Promote | Promotes tumor growth through the positive feedback loop of TNFR1/NF- κ B (and/or p38)/p-STAT3/HBXIP/TNFR1 | Cai, X et al.,2017 |
| | Breast cancer | Inhibit | Shows cytotoxic effects against MCF-7 cells | Ghandadi,M et al.,2017 |
| | Ovarian cancer | Inhibit | Overcomes the resistance of PTX | MIZUTANI,Y et al.,1994 |
| | | | Overcome the resistance of CDDP | MIZUTANI,Y et al.,1993 |
| | Prostate cancer | Inhibit | Inhibits H-3-thymidine uptake by PBMC | Hassan, M et al.,1999 |
| | | | Inactivates the NF- κ B signaling pathway | Wang, M et al.,2020 |
| | Breast cancer | Inhibit | Drives cells to non-apoptotic cellular death via RIP1, activation of JNK and ROS production | MIZUTANI,Y et al.,1993 |
| TNF- β | Non-cardiac gastric cancer | Promote | Elevates induction of cellular death, increases or reduces CXCR4 expressions, decrease BCSCs population | Abdolvand M et al.,2023 |
| | | | Paracrine TNF- α | Ma, G et al.,2013 |
| | Colorectal cancer | Inhibit | The G/A + A/A genotype frequencies were significantly higher in patients with intestinal gastric cancer | Zheng, W et al.,2019 |
| CD40L | Colorectal cancer | Inhibit | Suppresses TNF- β -stimulated NF- κ B signaling | Buhrmann,C et al.,2019 |
| | | | Infects tumor cells and expresses CD40L; have dose-dependent lytic ability against tumor cells | Liu, D et al.,2019 |
| | | | Induces the apoptosis | Pang, X et al.,2017 |
| | Lung cancer | Inhibit | Shows immunogenicity on colon 26/CD40L cells | Wu, L et al.,2010 |
| | | | DC pulsed by the tumor antigens from the reconstitution CD40L enhances its specific immunity capacity | Tian, K et al.,2017 |
| | | | Has direct anti-tumor effects against CD40-positive lung cancers | Xu, W et al.,2015 |
| | | | Activates human DCs to secrete interleukin-12 | Wu, J et al.,2007 |
| | | | Enhances the anti-tumor immunity efficiently | Noguchi, M et al.,2001 |

Table 4 (continued)

| TNF | Cancer types | Promote/Inhibit | Mechanisms | References |
|---|----------------------------|-----------------|---|-------------------------------|
| FasL | Colorectal Cancer | Promote | Causes Duke's stage, lymph node and liver metastasis | Zhang, W et al.,2004 |
| | | | Facilitates hepatic metastasis | Li, S et al.,2003 |
| | Colon cancer | Promote | FasL was strong positive in all lymph node metastases of large intestine cancer | Zhu, Q et al.,2002 |
| | | | Enhances the ability of cancer cells to counterattack T lymphocytes | Zhang, W et al.,2002 |
| | Gastric cancer | Promote | Involves in the pathogenesis and the immune escape and in the degree of differentiation | Pu, W et al.,2003 |
| | Cervical cancer | Promote | Induces TILs apoptosis | Anggraeni, T et al.,2020 |
| CD30L | Non-small cell lung cancer | Inhibit | Abrogates counterattack | Lin, Y et al.,2013 |
| | | Inhibit | Participates in the induction of cell apoptosis | Di, D et al.,2005 |
| | | Inhibit | Increases the expression of PD-L1; promotes the up-regulation of PD-1 expression and inhibits their activation, differentiation and ability to secrete effector cytokines | Wang, X et al.,2020 |
| 4-1BBL | Prostate cancer | Promote | Mediates cancer progression to castration-resistant prostate cancer via enhancing expression and function of AR | Zhu, H et al.,2019 |
| | Colon cancer | Inhibit | Inhibits proliferation, migration and invasion, and retards tumor growth | Ge, Y et al.,2020 |
| | Lung cancer | Inhibit | Decreases cell viability, induces apoptosis and autophagy | Ramos-Gonzalez, M et al.,2024 |
| OX40L | Liver cancer | Inhibit | CD4 + and CD8 + T cells were significantly increased in the OX40L mRNA group | Deng, Z et al.,2022 |
| | Breast cancer | Inhibit | Inhibits cell growth and up-regulates the key immune molecules Ox40L and 4-1BBL | Kaser, E et al.,2022 |
| TNF-related apoptosis-inducing ligand (TRAIL) | Lung cancer | Promote | Inhibits TRAIL-induced apoptosis | Li, H et al.,2021 |
| | | Inhibit | Up-regulates the expression of TRAIL-R1 and TRAIL-R2 | Szliszka, E et al.,2009 |
| | | | Augments the cytotoxic effect of TRAIL | Szliszka, Ewelina et al.,2011 |
| | | | Enhances the cytotoxic and apoptotic effects of TRAIL | Szliszka, E et al.,2012 |
| | | | Inhibits cell proliferation, down-regulates XIAP and modulates tBid and Bax expression | Choi, Y et al.,2014 |
| | | | Suppresses tumor growth | Zhao, Y et al.,2013 |
| | Colon cancer | Inhibit | Up-regulates TRAIL receptor expression, enhances TRAIL-induced cell death partly via O-glycosylation | Semba, M et al.,2022 |
| | Pancreatic cancer | Inhibit | Strengthens the apoptotic signaling pathway | Huang, M et al.,2021 |

Table 4 (continued)

| TNF | Cancer types | Promote/Inhibit | Mechanisms | References |
|----------------|------------------------------|-----------------|--|----------------------------------|
| | Cervical cancer | Inhibit | Enhances TRAIL-induced apoptosis through increasing the expression of TRAIL-R2 | Szliszka, E et al.,2012 |
| | Non-small cell lung cancer | Inhibit | Induces cell death which is dependent on caspase-8 and caspase-3 activation | De Miguel, D et al.,2016 |
| | Ovarian cancer | Inhibit | Enhances TRAIL sensitivity or reverses TRAIL resistance | Liang, R et al.,2020 |
| | Colorectal cancer | Inhibit | Enhances the activation and apoptosis of ROS-dependent caspases 3/7, promotes the induction of the death receptor 5 | Ishaq, M et al.,2015 |
| LIGHT(TNFSF14) | | | Enhances caspase-dependent apoptosis induction via both death receptor- and mitochondrial-mediate apoptosis pathways | Sophonmithprasert, T et al.,2015 |
| | Breast cancer | Inhibit | Induces miR-146a expression and suppresses CXCR4-mediated human breast cancer migration | Wang, D et al.,2013 |
| | Tongue cancer | Promote | Enhances proliferation and migration | Gao, W et al.,2015 |
| RANKL | NSCLC | | Promotes osteolytic bone metastases | Brunetti, G et al.,2020 |
| | Oral squamous cell carcinoma | Promote | Promotes disease recurrence and a cell compartment | Grimm, M et al.,2015 |
| | Cervical cancer | Promote | Recruits Tregs by up-regulating CTSS and enhancing the expression of phosphorylated AKT and mTORC | Wang, Y et al.,2019 |
| | | | Strengthens the dialogue between cells and regulation of IL-8 secretion | Shang, W et al.,2015 |
| | NSCLC | Promote | Activates NF- κ B pathway, increases RANKL and M-CSF expression and induces osteoclastogenesis | Choi, J et al.,2020 |
| | | | Promotes tumor angiogenesis | YU, Z et al.,2009 |
| | Breast cancer | Promote | Promotes migration via the PI3K/AKT-HIF-1 α pathway | Tang, Z et al.,2011 |
| | | | The inhibition of RANKL sensitizes cancer stem cells to denosumab | Cuyas, E et al.,2017 |
| | | | Induces cell migration | Tang, Z et al.,2011 |
| | Gastric cancer | Promote | Induces migration partially through the activation of PI3K and MEK signaling | Wang, Y et al.,2013 |
| | | | Induces cell migration | Wang, Y et al.,2018 |

Table 4 (continued)

| TNF | Cancer types | Promote/Inhibit | Mechanisms | References |
|--------|-------------------|-----------------|--|--------------------------|
| TWEAK | Pancreatic cancer | Promote | TWEAK expression rate was higher than that in chronic pancreatitis and normal pancreatic tissues | Wei, A et al.,2017 |
| | Breast cancer | Promote | Relates to the metastatic ability | Zheng, Y et al.,2008 |
| | Ovarian cancer | Promote | Promotes metastasis via NF-κB pathway activation and VEGF expression | Dai, L et al.,2009 |
| | Colon cancer | Promote | Promotes cell proliferation and infiltration | Zhang, Y et al.,2014 |
| | Ovarian cancer | Inhibit | Enhances cisplatin sensitivity by regulating apoptosis | Ma, N et al.,2013 |
| | | | Promotes macrophage-derived exosomal miR-7 to cell through regulating Dicer | Qiu, X et al.,2018 |
| | | | Activates autophagy and enhances the cisplatin sensitivity | Wang, W et al.,2013 |
| | Colon cancer | Inhibit | Induces apoptosis | Dionne, S et al.,2010 |
| APRIL | Cervical cancer | Inhibit | Promotes cell apoptosis | Wang, D et al.,2010 |
| | Breast cancer | Promote | Mediates breast cancer cell stemness | Pelekanou, V et al.,2018 |
| | Gastric cancer | Promote | Induces cisplatin resistance via activation of the NF-κB pathway | Zhi, X et al.,2015 |
| BAFF | Colorectal cancer | Inhibit | Suppresses cell growth and promotes apoptosis | Wang, J et al.,2010 |
| | Breast cancer | Promote | Mediates cell stemness | Pelekanou, V et al.,2018 |
| | Cervical cancer | Inhibit | Promotes immunosuppression | Ding, J et al.,2023 |
| VEGI | Prostate cancer | Inhibit | Inhibits cellular motility and adhesion | Zhang, N et al.,2009 |
| | Bladder cancer | Inhibit | Inhibits cellular motility and adhesion | Zhang, N et al.,2010 |
| EDA-A2 | Breast cancer | Inhibit | Is down-regulated in breast cancer via promoter methylation | Punj, V et al.,2010 |

supporting their activity within TME. Similarly, inhibiting chemokines that draw in and support immune cells' suppressive roles is an intriguing avenue for future research to enhance treatment outcomes. Additionally, another therapeutic approach being investigated to enhance responses to cancer therapy is the activation or transduction of chemokine receptors on adoptively transferred anti-tumor T cells, which facilitates their ability to enter deeply into the tumor and license their functioning. Significant advancements in our comprehension of the immune system's function during carcinogenesis have resulted in the creation of innovative immunotherapeutic methods for treating diverse tumors, which have substantially aided cancer patients. Immunotherapy continues to be one of the most promising medical advancements of the twenty-first century. More details regarding the function of chemokines in cancer are provided in Table 6.

Targeting inflammasomes for cancer chemoprevention

As the innate immune system receptors and sensors, inflammasomes are multiprotein complexes, which react to recognized indicators of endogenous (linked to cellular damage, ATP, ROS, and DNA) and external (related to infections) stimuli [302]. Inflammasome components include the NACHT, leucine-rich repeat (LRR), and pyrin domain (PYD) domain-containing protein 1 (NLRP1), nucleotide-binding domain leucine-rich repeat (NLR) and pyrin domain-containing receptor (NLRP3) (also known as cryopyrin), NLR family caspase activation and recruitment domain-containing protein 4 (NLRC4), NOD-like receptor family pyrin domain containing 6 (NLRP6), and absent in melanoma 2 (AIM2). Detailed information on the structure of NLRP1, NLRP3, NLRP4, NLRP6, and AIM2 are shown in Fig. 4. Their products like interleukin 1β and interleukin 18, along with the adaptor, apoptosis-associated

speck-like protein containing caspase activation and recruitment domain (ASC) and the effector caspase-1 both play a major role in carcinogenesis [303]. The sensor protein on each type of inflammasome determines whether a form of the inflammasome is present by identifying pathogenic ligands and triggering the assembly of inflammasomes. NLRs convert the biologically inactive pro-IL-1 β and pro-IL-18 into their active forms through caspase-1 [304]. Genetic mutations in NLRP1, NLRP3, NLRC4, and AIM2 are linked with the development of auto-inflammatory diseases, enterocolitis, and cancer [305]. It is commonly known that inflammatory proteins and their byproducts have a role in developing several cancers, such as skin cancer [306], lung cancer [307], and others. Inflammasomes play both protective and detrimental roles in cancer. On one hand, mice that lack NLRP3, ASC, or caspase-1 exhibit increased susceptibility to colitis and to colitis-associated colorectal cancer induced by the chemical colitogen dextran sulfate sodium (DSS) [308, 309]. Infusion of recombinant IL-18 reduces tumor frequency in mice deficient in inflammatory components following azoxymethane (AOM) and DSS treatment [308]. IL-18 plays a role in repairing the epithelial barrier and preventing damage [309]. This may clarify the protective roles of NLRP3 and IL-18 in relation to colitis-associated colorectal cancer. Mice that lack IL-18 are more vulnerable to developing lung metastasis [310]. Mice that were injected daily with recombinant IL-18 for five days exhibited fewer lung metastases [311]. Other NLR sensors, such as NLRP6 and NLRP1b, have also shown protective effects against tumorigenesis. For instance, the NLRP1b inflammasome mediates the secretion of IL-1 β and IL-18 in stromal colon cells, providing protection against colon tumorigenesis [312]. NLRP6 protects against chemical-induced colon cancer by activating caspase-1 and promoting IL-18 production in the intestine [313]. The blockade of ASC promotes the growth of melanoma tumors [314]. On the other hand, in some cases, the activation of the inflammasome can suppress antitumor immunity. For instance, when B16-F10 melanoma cells or RM-1 prostate cancer cells are delivered intravenously (as opposed to subcutaneously) into mice, the activation of NLRP3 is associated with increased lung metastasis. The harmful impact of NLRP3 in melanoma may stem from its inhibitory effect on the activation of NK cells, which are responsible for secreting IFN γ and killing tumor cells. In some cases, inflammasome activation can suppress antitumor immunity. For example, when B16-F10 melanoma or RM-1 prostate cancer cells are delivered intravenously into mice, NLRP3 activation is linked to increased lung metastasis. This detrimental effect may

be due to NLRP3's inhibition of NK cell activation, which is crucial for secreting IFN γ and killing tumor cells [310]. Recombinant IL-18 increases lung metastasis when injected into mice twice a week [311]. The knockdown of the gene encoding ASC suppresses the growth of melanoma xenograft tumors [314]. The types of cells, tissues, and organs involved in an inflammasome significantly influence its characteristics related to tumor promotion and suppression. The biological link between inflammasomes and cancer offers promising opportunities for exploring novel anticancer therapies.

Besides, other studies also point to the tumor-suppressive function of inflammasomes like NLRP3, NLRP1, NLRP6, and Pyrin in the initiation and spread of some malignancies, including colorectal cancer [315]. Recently, some inhibitors and agonists of these inflammasomes exhibit anti-tumor effects. For example, the small molecule OLT1177 suppressed tumor growth by inhibiting NLRP3 [316]. OLT1177 and another NLRP3 inhibitor MCC950 reduced tumor growth in melanoma [317, 318]. Nigericin controls tumor growth by activating NLRP3 in breast cancer and neuroblastoma [319]. Since the multifaceted role of inflammasomes in carcinogenesis, clarifying the mechanism of action in different tumors will provide novel therapeutic approaches for cancer treatment and prevention. Here, we will summarize the promoting and suppressive effects of these inflammasomes and their inhibitors and agonists in cancer initiation and development. A comprehensive list of the role of these inflammasomes in cancer is also shown in Table 7.

The role of NLRP1 in inflammation-mediated carcinogenesis

The receptor NLRP1, the adaptor protein ASC, and the effector protein caspase-1 make up the multi-protein complex known as the NLRP1 inflammasome. NLRP1 is the first identified and currently recognized predominant inflammasome sensor protein in human keratinocytes. NLRP1 has its unique domain, which contains an effector C-terminal caspase recruitment domain (CARD), PYD, a central NBD, an LRP and a function to find domain (FIIND). In the absence of external stimuli, the NBD binds to LRR, inhibiting self-oligomerization and bringing the protein into an inactive state. When cells are exposed to external stimuli, such as viruses, UVB rays, and ribotoxic stress reactions, they bind to the LRR domain, then induce a conformational change in NLRP1 and expose PYD and CARD domains. The downstream proteins which containing PYD and CARD are then mediated, such as the homologous interaction between ASC and caspase-1. Dipeptidyl protease (DPP), anthrax lethal toxin (LT), and parasites can activate NLRP1 [320].

Database analysis indicates that NLRP1 has a distinct expression pattern across various tumors, and patients

Table 5 The role and mechanisms of granulocyte/macrophage colony-stimulating factor (GM-CSF) in cancer

| GM-CSF | Cancer types | Promote/inhibit | Mechanisms | References |
|--------|--|-----------------|--|------------------------------|
| GM-CSF | Colon cancer | Promote | Promotes liver metastasis by down-regulating E- cadherin and up-regulating N- cadherin and MMP2 | Ding X et al., 2018 |
| | | | Makes cells more resistant to cytotoxic drugs through MAPK/ERK signal and EMT-induced transcription factor ZEB1 | Chen Y et al., 2017 |
| | Non small cell cancer | Promote | Promotes carcinogenesis | Oshika Y et al., 1998 |
| | Large cell carcinoma of lung | Promote | Stimulates autocrine tumor through leukemia reaction and obvious eosinophilia | Lammel V et al., 2012 |
| | Breast cancer | Promote | Promotes the tumor-promoting effect of WAT progenitor cells | Reggiani F et al., 2017 |
| | | | The depletion of GM-CSF leads to the decrease of proliferation, invasion and dryness by inhibiting STAT3 phosphorylation and β -catenin signal | Shi H et al., 2020 |
| | | | Promotes metastasis through the positive feedback loop between GM-CSF and CCL18 | Su S et al., 2014 |
| | Gastric cancer | Promote | Promotes chemotherapy induced- CSCs | Xue X et al., 2022 |
| | Non-myeloid carcinoma | Promote | Contributes to cancer recurrence through new angiogenesis | Aliper A et al., 2014 |
| | Lung cancer | Promote | Stimulates the growth or invasion of tumors | Liu Q et al., 2017 |
| | Squamous cell carcinoma of head and neck | Promote | Increases of tumor recurrence or metastasis | Young M et al., 1997 |
| | | | Induces angiogenesis and invasion, and related to immune evasion | Tenhuinink W et al., 2023 |
| | Prostate cancer | Inhibit | Promotes the host immune monitoring of dendritic cells | Bandyopadhyay S et al., 2008 |
| | | | Improves the efficacy of RM-1 prostate cancer cell vaccine | Yin W et al., 2010 |
| | Colon cancer | Inhibit | Regulates immune response | Urduingio R et al., 2013 |
| | Endometrium cancer | Inhibit | Inhibits DMH-induced colon cancer in rats | Dinc S et al., 2007 |
| | | | Inhibits tumor growth by interacting with TGF- β 1 and regulating the expression of TGF- β 1 and TGF- β II receptors | Ripley D et al., 2001 |
| | Bladder cancer | Inhibit | Inhibits tumor growth and regresses established tumors by increasing the number of mature DC and up-regulating the expression of IFN-dependent PD-L1 | Zhang X et al., 2018 |
| | Breast cancer, pancreatic cancer | Inhibit | Enhances the anti-tumor immunity | Antonarakis E et al., 2010 |

Table 5 (continued)

| GM-CSF | Cancer types | Promote/inhibit | Mechanisms | References |
|------------------------------------|--|-----------------|--|--|
| | Squamous cell carcinoma of head and neck | Inhibit | Stimulates the differentiation of dendritic cells, presents tumor antigens and regulates T cell function | Tenhuinink W et al., 2023 |
| | Colon cancer | Inhibit | Regulating immune response | Urdinguio R et al., 2013 |
| | Laryngocarcinoma | Inhibit | Inhibits DMH-induced colon cancer in rats Enhances the immunogenicity of cancer cells, induces proliferation of tumor infiltrating lymphocytes and the tumor-specific cytotoxicity of cytotoxic T lymphocytes | Dinc S et al., 2007 Qiu Z et al., 2001 |
| | Esophageal cancer | Inhibit | Promotes the strong immune response Inhibits proliferation and migration, induces apoptosis and regulates EMT through JAK2-PRMT5 signaling | Miyashita T et al., 2008 Zhang J et al., 2017 |
| | Ovarian cancer | Inhibit | Negatively induces myeloid suppressor cells (MDSC) and promotes tumor progression and metastasis | Zhang Y et al., 2013 |
| | Lung cancer | Inhibit | Inhibits carcinogenesis by being combined with IL-2 Inhibits carcinogenesis by being combined with immunotherapy and IL-18 | Takahashi K et al., 2000 Tian H et al., 2023 |
| | Bladder cancer | Inhibit | Enhances the anti-tumor effect of cisplatin Inhibits tumor growth and leads to a significant increase in CD4(+) , CD8(+) T cells and CD4(+) Foxp3(+) T cells | Luo D et al., 2017 Peng J et al., 2019 |
| | Cervical cancer | Inhibit | Promotes the anti-tumor response by inhibiting the expression of iNOS and COX-2 in a GM-CSFR independent manner Enhances the anti-tumor immune response with nanoparticles loaded with adriamycin and GM-CSF | Jiang N et al., 2015 Zhang X et al., 2023 |
| | Lewis lung cancer | Inhibit | Enhances the anti-tumor immunity with the combination of FasL and GM-CSF | He M et al., 2008 |
| FRG1 (inhibitor) | Breast cancer | Inhibit | Inhibits metastasis by regulating GM-CSF/MEK-ERK axis | Mukherjee B et al., 2022 |
| COX-2 inhibitor (inhibitor) | Lung cancer | Inhibit | Improves the prognosis of lung cancer patients by reducing G-CSF or GM-CSF | Nakata H et al., 2003 |
| Kaempferol and quercetin (agonist) | Prostate cancer | Inhibit | Stimulates the immune response by stimulating the production of GM-CSF, and then lead to DC recruitment to the tumor site | Bandyopadhyay S et al., 2008 |

Table 6 The role and mechanisms of chemokines in cancer

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------------------|-------------------|-----------------|--|-------------------------|
| CCL1 | CCR8 | Esophageal cancer | Promote | Promotes tumor progression through 40 kDa/Akt target of mammalian rapamycin pathway/proline-rich Akt substrate | Fujikawa M et al., 2021 |
| | | Colorectal cancer | Promote | Promotes chemoresistance through TGF β/NF-κB signaling pathway | Li Z et al., 2018 |
| | | Colorectal cancer | Inhibit | Negatively regulates the progress of liver metastasis | Iwata M et al., 2024 |
| | | Breast cancer | Inhibit | Inhibits tumorigenesis, metastasis and chemotherapy resistance by reducing the binding of H3K27Me3 in p65 and CCL1 promoter regions to recruit Tregs | Xu Y et al., 2017 |
| | | Lung cancer | Inhibit | Inhibits the differentiation of Tregs and the metastasis of lung tumors | Wang M et al., 2022 |
| CCL2 | CCR2 CCR4 CCR5 | Breast cancer | Promote | Promotes cell survival and invasion in vitro | Yao M et al., 2017 |
| | | | | Stimulates stem cell-specific spherical phenotype and CSC self-renewal | Tsuyada A et al., 2012 |
| | | | | Induces proliferation, survival, migration and glycolysis through MET-dependent mechanism | Acevedo D et al., 2022 |
| | | | | Promotes the growth and cell cycle process through SRC and PKC activation | Yao M et al., 2019 |
| | | | | CCL2-mediated matrix interaction drives macrophage polarization to increase tumor occurrence | Archer M et al., 2023 |
| | | Tongue cancer | Promote | Promotes invasion and metastasis through PI3K/AKT pathway | Dong Y et al., 2023 |
| | | Bladder cancer | Promote | LNWAT1 promotes lymphatic metastasis through CCL2-dependent macrophage recruitment | Atala A et al., 2019 |
| | | | | Promotes migration and invasion through PKC activation and tyrosine phosphorylation | Chiu H et al., 2012 |
| | | Ovarian cancer | Promote | Promotes tumor progression through MEK/ERK/ MAP3K19 signaling pathway | Liu W et al., 2023 |
| | | | | Inhibition of CCL2 enhances the treatment efficiency with paclitaxel and carboplatin | Moson F et al., 2014 |
| | | | | Promotes ovarian peritoneal metastasis through p38-MAPK pathway | Yasui H et al., 2020 |
| | | Prostate cancer | Promote | Promotes the migration of prostate cancer | Lin T et al., 2013 |
| | | | | Promotes cell survival by inducing mTOR pathway | Roca H et al., 2009 |
| | | | | Stimulates cell proliferation | Loberg R et al., 2006 |
| | | | | Promotes bone metastasis | Li X et al., 2009 |
| | | | | Inhibiting CCL2 activity can enhance the therapeutic response to taxane therapy | Qian D et al., 2010 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------------------|------------------------------------|-----------------|--|-------------------------|
| CCL3 | CCR1 CCR4 CCR5 | Cervical cancer | Promote | Protects cells from autophagy through phosphatidylinositol 3- kinase /AKT/ survivin pathway | Roca H et al., 2008 |
| | | | | Promotes proliferation, migration, invasion and EMT | Huang T et al., 2020 |
| | | | | Promotes EGFR-TKIs resistant cancer through AKT-EMT pathway | Diao Y et al., 2024 |
| | | Lung cancer | Promote | Plays a key role in tumor promotion by recruiting macrophages and influencing their functions | Zhang J et al., 2018 |
| | | | | Astrocytes promote migration by secreting C–C motif chemokine ligand 2 (CCL2) | Hajal C et al., 2021 |
| | | Colorectal cancer | Promote | CCL2-SOSTM1 positive feedback loop inhibits autophagy to promote chemotherapy resistance | Xu W et al., 2018 |
| | | | | Anti-CCL2 or anti-CCL5 therapy inhibits the growth of cancer | Svensson S et al., 2015 |
| | | Brain tumor | Promote | Mediates the metastasis of dysmucin in ER-negative breast cancer | Nam J et al., 2006 |
| | | | | Promotes cell growth, leads to EMT and promotes cell migration and invasion through PI3K-AKT-mTOR pathway | Luo A et al., 2020 |
| | | Gastric cancer | Promote | Enhances the chemo-sensitivity of docetaxel by triggering the polarization of pro-inflammatory macrophages | Anonymous et al., 2022 |
| | | | | CCL3 –CCRS axis promotes migration and invasion through AKT signaling pathway | Guan BG et al., 2022 |
| | | Hormone dependent mammary gland | Promote | Recombinant Bacteroides fragilis enterotoxin –1 (rBFT-1) promotes proliferation through CCL3-related pathway | Xie XL et al., 2021 |
| | | | | Promotes proliferation, invasion and migration through TRAF6 and NF-κB | Ma XQ et al., 2022 |
| CCL4 | CCR1 CCR2 CCR5 | Oral cancer | Promote | Promotes tumorigenesis by inducing inflammation and angiogenesis, and the recruitment of eosinophils | da Silva J et al., 2017 |
| | | | | Promotes cell migration and invasion through CCR5 binding and phosphorylation of AKT and ERK, thus promoting the progress and poor prognosis of ESCC | Kodama T et al., 2020 |
| | | | | Promotes angiogenesis through the imbalance of miR-374b/VEGF-A axis | Liao Y et al., 2016 |
| | | Esophageal squamous cell carcinoma | Promote | Increases the expression of MMP-2 and enhances the migration ability | Xu C et al., 2013 |
| | | | | CCL3 alone or in combination with anti-PD-1 may be an effective immunotherapy | Wang X et al., 2024 |
| | | Human osteosarcoma | Promote | Promotes proliferation, invasion and migration by targeting VEGF-A signaling pathway | Fu H et al., 2017 |
| | | | | | |
| | | Glioma | Inhibit | | |
| | | | | | |
| | | Endometrium cancer | Promote | | |
| | | | | | |
| | | | | | |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|------------------------------|------------------------------|-----------------|---|----------------------------|
| CCL5 | CCR1 CCR3CCR4 CCR5 | Oral squamous cell carcinoma | Promote | Induces the expression of vascular endothelial growth factor C and lymphangiogenesis through miR-195-3p | Lian M et al., 2018 |
| | | | | Stimulates the expression of angioipoetin –2 and angiogenesis via MEK/ERK/STAT3 | Lu C et al., 2022 |
| | | Human osteosarcoma | Promote | Stimulates migration through the miR-3927-3P/ integrin α v β 3 axis | Tsai H et al., 2022 |
| | | Breast cancer | Promote | Promotes bone metastasis by mediating the interaction between cancer cells and fibroblasts | Sasaki S et al., 2016 |
| | | Breast cancer | Promote | Promotes tumor growth and metastasis | Yao X et al., 2007 |
| | | | | Promotes tumor invasion | Pinilla S et al., 2009 |
| | | Gastric cancer | Promote | Promotes proliferation, invasion and metastasis of gastric cancer cells | Ding H et al., 2016 |
| | | | | Gastric cancer cells use CCL5 derived from CD4 + cells to grow and prevent tumor elimination with CD8 +cells | Sugasawa H et al., 2008 |
| | | | | KLF5 leads to low survival rate and promotes cancer progression by activating CCL5/CCR5 axis | Yang T et al., 2017 |
| | | Prostate cancer | Promote | Promotes the up-regulation of androgen receptor (AR) and leads to enzalutamide resistance by activating AKT | Xiong Z et al., 2024 |
| CCL6 | CCR1 CCR2 CCR5 CCR5 | Lung cancer | Promote | Promotes the migration of human lung cancer cells | Hang C et al., 2009 |
| | | Pancreatic cancer | Promote | Promotes migration and invasion | Singh S et al., 2018 |
| | | Colon cancer | Promote | CCL 5 is involved in cancer progression mediated by tumor-associated dendritic cells through non-coding RNA MALAT-1 | Guan Z et al., 2015 |
| | | N/A | | | |
| | | Gastric cancer | Promote | Inhibition of CCL7 weakens proliferation, migration, invasion and induces apoptosis | Chen M et al., 2023 |
| CCL7 | CCR1 CCR2 | Colon cancer | Promote | Accelerates the early stage of tumor growth and leads to higher lung metastasis rate | Kurzejamska E et al., 2019 |
| | | | | Promotes metastasis through ERK-JNK signaling pathway | Li Y et al., 2016 |
| | | Ovarian cancer | Promote | CCL7-induced invasion needs to express MMP 9 by activating ERK signaling | Zheng M et al., 2021 |
| | | Pancreatic cancer | Promote | Pancreatic stellate cells promote pancreatic cancer invasion through CCL7/CCR5 axis in hypoxic microenvironment | Wu Y et al., 2017 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|--------------|--|-----------------|--|------------------------------|
| CCL8 | CCR1 CCR2 | Lung cancer | Promote | ABCE1 participates in tumor occurrence and progression through CCL7 signaling | Wu Z et al., 2018 |
| | | Lung adenocarci- noma | Promote | LINC01094/SP1/CCL7 Axis promotes macrophage accumulation and tumor cell spread in lung adenocarci- noma | Wu Z et al., 2022 |
| | | Breast cancer | Promote | Promotes metastasis by regulating the tumor-promoting activity of tumor microenvironment and promotes tumor growth by recruiting macrophages | Farmaki E et al., 2020 |
| | | Glioblasto-ma | Promote | CCL8 secreted by tumor-associated macrophages promotes invasion and dryness through ERK1/2 signal | Zang X et al., 2020 |
| | | Colon cancer | Promote | Accelerates tumor progression through CCL-8 /CCR5/mTORC1 axis | Zhou H et al., 2023 |
| CCL9 | CCR1CCR3 | Lung cancer | Promote | Enhances the survival rate of tumor cells in lung before metastasis | Yan H et al., 2015 |
| | | Pancreatic ductal adenocarci-noma | Promote | Carcinogenic Kras enhances pancreatic ADM through its new downstream target molecule CCL9 to start PDAC supply | Liou G et al., 2024 |
| | | Liver cancer | Promote | Recruits MDSC to promote tumor growth in mice with orthotopic liver cancer | Li B et al., 2023 |
| CCL10 | CCR1CCR4 | N/A | | | Levina V et al., 2009 |
| CCL11 | CCR2 CCR3 | Ovarian cancer | Promote | Plays an important role in the proliferation and invasion | Huang W et al., 2010 |
| | | Head and neck cancer | Promote | Cancer-related fibroblasts promote tumor invasion of head and neck cancer through CCL11 and CCR3 signal transduction pathway | Tian M et al., 2016 |
| | | Glioblasto-ma | Promote | Promotes proliferation, migration and invasion | Lin S et al., 2021 |
| | | Non-small cell lung cancer | Promote | Activates AKT and ERK signaling and promotes metastasis through epithelial-mesenchymal transition (EMT) | Polosukhina D et al., 2021 |
| | | Colon cancer | Promote | CCL11 aggravates colitis and inflammation-related colon tumors | Liu Y et al., 2017 |
| | | Breast cancer | Promote | Accelerates tumor growth and induces drug resistance and metastasis | Bekaert S et al., 2021 |
| | | | Inhibit | Asthma-related inflammation promotes lung metastasis through CCL11-CCR3 pathway CCL11 has anti-tumor effect in BRCA | Chen X et al., 2024 |
| | | Anaplastic large cell lymphoma (ALCL) | Promote | Increases cell survival rate and proliferation , induces ERK1/2 phosphorylation, induces the expression of anti-apoptosis proteins Bcl-xL and survivin and enhances tumor growth | Tomomitsu M et al., 2011 |
| | | Pancreatic cancer | Inhibit | Autochemokine-lipolytic signal transduction inhibits CCL11- eosinophil axis to promote tumor progress | Bhattacharyya S et al., 2024 |
| CCL12 | CCR2 | N/A | | | |
| CCL13 | CCR2 CCR3 | N/A | | | |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|--------------|--------------------------|-----------------|--|-------------------------|
| CCL14 | CCR1 CCR5 | Thyroid cancer | Promote | CCL14 may be involved in the recurrence of THCA | Zhang M et al., 2023 |
| | | Myeloma | Promote | Promotes tumor growth and survival signals by activating PI3K/AKT and ERK/MAPK pathways and c-myc | Li Y et al., 2015 |
| | | Pancreatic cancer | Promote | Up-regulates migration and invasion | Messex J et al., 2022 |
| | | Colon cancer | Promote | Myeloid suppressor cells promote invasion through CCL15-CCR1 chemokine axis | Itatani Y et al., 2014 |
| | | | | MDSCs accumulate and invade the primary cancer through CCL15-CCR1 chemokine axis, and promotes tumor progression | Inamoto S et al., 2015 |
| | | | | CCL15 secreted by SMAD4 deficient cells recruited CCR1(+) cells to promote lung metastasis | Yamamoto T et al., 2017 |
| | | | Inhibit | Inhibits proliferation and invasion by inhibiting the formation of M2-like TAM | Li N et al., 2021 |
| | | | | Long-chain noncoding RNA CCL14-AS inhibits invasion and lymph node metastasis by regulating MEPIA | Li M et al., 2023 |
| CCL16 | CCR1 CCR2 | Hepatocellular carcinoma | Promote | Recruits inhibitory monocytes to promote the immune escape and CCL15-CCR1 axis creates a complex tumor-promoting inflammatory microenvironment | Liu L et al., 2019 |
| | | | Inhibit | Inhibits proliferation and promotes apoptosis by inhibiting the activation of Wnt/ β -catenin pathway | Zhu M et al., 2019 |
| | | Hepatocellular carcinoma | Promote | Promotes tumorigenesis by recruiting M2-like tumor-associated macrophages through CCL16-CCR1 axis | Dai Z et al., 2024 |
| | | Breast cancer | Inhibit | Inhibits tumor growth and prevents metastasis | Guiducci C et al., 2004 |
| CCL17 | CCR4-CCR8 | Cervical cancer | Promote | Promotes cell proliferation through JNK and STAT5 signaling pathways | Liu L et al., 2015 |
| | | | | | |
| CCL18 | CCR8 | Colitis-related cancer | Promote | Promotes tumor occurrence by affecting the composition of intestinal microbiota and reducing cell apoptosis | Metzger R et al., 2023 |
| | | Breast cancer | Promote | Induces cytoskeleton aggregation through its receptor and promotes migration | Chen J et al., 2014 |
| | | | | Promotes angiogenesis and tumor progression | Lin L et al., 2015 |
| | | | | Induces migration and invasion through PCAF-dependent acetylation | Song X et al., 2018 |
| | | | | Promotes infiltration and migration through integrin aggregation | Chen J et al., 2014 |
| | | | | Promotes invasion by inhibiting e-cadherin expression mediated by EZH2 | Jia H et al., 2023 |
| | | | | Promotes invasion and metastasis, and cancer passes through Annexin A2 | Zhao C et al., 2024 |
| | | | | Promotes malignant behavior by up-regulating Src/PI3K/Akt signaling mediated by ARF6 | Huang X et al., 2022 |
| | | | | Promotes metastasis by down-regulating miR98 and miR27b | Lin X et al., 2015 |
| | | | | | |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|--------------|--|-----------------|---|--------------------------|
| CCL19 | CCR1 CCR7 | Gastric cancer | Promote | Promotes invasion and migration through ERK1/2/NF-κB signaling pathway | Hou X et al., 2016 |
| | | Human pancreatic ductal adenocarcinoma | Promote | Accelerates the progress of PDAC by promoting epithelial-mesenchymal transformation, invasion and migration | Meng F et al., 2015 |
| | | Oral cancer | Promote | CCL18-NIR1 promotes the growth and metastasis by activating JAK2/STAT3 | Jiang X et al., 2020 |
| | | Non-small cell lung cancer | Promote | Enhances the adhesion of NSCLC cells by activating ELMO1- integrin 1 signal | Shi L et al., 2016 |
| | | Mastocarci-ncma | Promote | CCL18 derived from TAMs plays a key role in promoting breast cancer metastasis through its receptor PITPNM3 | Chen J et al., 2011 |
| | | Lung cancer | Promote | Induces epithelial-mesenchymal transition and enhances the invasion potential | Ploenes T et al., 2013 |
| | | Bladder cancer | Promote | Promotes migration, invasion and EMT by binding CCR8 | Liu X et al., 2019 |
| | | Ovarian cancer | Promote | As a component of ascites, CCL18 plays an important role in tumor cell migration | La D et al., 2016 |
| | | | | Enhances invasion, migration and adhesion in vitro | Zhang W et al., 2013 |
| | | | | Promotes invasion through mTORC2 pathway | Wang Q et al., 2016 |
| | | Prostate cancer | Promote | The up-regulation of CCL18 may be related to the malignant progress of PCa | Chen G et al., 2014 |
| | | Oral squamous cell carcinoma | Promote | Stimulates growth and invasion in an autocrine way through Akt activation | Jiang X et al., 2016 |
| | | Esophageal cancer | Promote | Promotes the malignant progression of tumor by up-regulating the expression of HOTAIR | Wang W et al., 2019 |
| | | Colon cancer | Promote | Promotes the proliferation, migration and invasion of SW620 cells | Lu J et al., 2014 |
| | | Small cell lung cancer | Promote | Relates to metastasis and poor prognosis, promotes tumor progression and metastasis and damages the function of CD8+T cells | Liu Q et al.,2021 |
| | | Gastric cancer | Inhibit | Enhances the immune effect of mice against gastric cancer | Chen Z et al., 2021 |
| | | | | Inhibits proliferation, migration and invasion in CCL-19 /CCR7/AIM2 pathway | Zhou R et al., 2020 |
| | | Colon cancer | Inhibit | Activates the immune system | Liu X et al., 2019 |
| | | | | Inhibits angiogenesis by promoting miR-206 and inhibiting Met/ERK/Elk-1/HIF-1α/VEGF-A pathway | Xu Z et al., 2018 |
| | | Lung cancer | Inhibit | Chemically attracts dendritic cells and T lymphocytes, which has anti-tumor effect | Hillinger S et al., 2003 |
| | | | | Reduces the tumor load through extensive mononuclear infiltration of the tumor | Hillinger S et al., 2006 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|--------------|------------------------------|-----------------|--|--------------------------------|
| CCL20 | CCR6 | Colon cancer | Promote | Inhibits tumor growth by promoting local anti-tumor T cell response | Cheng H et al., 2018 |
| | | | | Induces proliferation and migration through autocrine HGF- α -Met and MSP-MSPR signaling pathways | Nandi B et al., 2021 |
| | | Pancreatic cancer | Promote | Promotes migration, epithelial-mesenchymal transformation and invasion | Liu B et al., 2016 |
| | | Breast cancer | Promote | Reduces the expression of IFN- γ secreted by CD8+ T cells through CCR6 + Tregs | Xu L et al., 2010 |
| | | | | Recruits immature dendritic cells into tumor tissues to impair immune response | Treilleux I et al., 2004 |
| | | | | Promotes migration and invasion | Kim K et al., 2009 |
| | | | | Promotes angiogenesis | Lee S et al., 2017 |
| | | | | Recombinant human CCL20 induces VEGF expression | He H et al., 2017 |
| | | | | Promotes angiogenesis | Marsigliante S et al., 2016 |
| | | | | Recruits macrophages into tumors to promote their growth | Lee SK et al., 2017 |
| CCL21 | CCR1 CCR7 | Colon cancer | Promote | Up-regulates ABCB1 to promote chemical resistance to taxanes | Chen W et al., 2018 |
| | | | | Regulates PMN-MDSCs and promotes dryness through CXCL2-CXCR2 pathway | Zhang R et al., 2023 |
| | | | | Up-regulates P-gp, Bmi-1, Nanog and OCT-4 by up-regulating AKT/GSK-3 β /Snail, and promotes the chemo-therapy resistance and stem cell characteristics | Lu S et al., 2016 |
| | | | | Promotes the chemotherapy resistance and stem cell characteristics of CRC cells | Lu L et al., 2016 |
| | | | | Promotes the migration and proliferation of BC cells | Peng J et al., 2023 |
| | | Breast cancer | Promote | Triggers migration and invasion through ERK and EMT signaling | Zhong G et al., 2017 |
| | | Lung cancer | Promote | COPD promotes tumor progress by enhancing the migration of CCL21-dependent cancer cells | Kuznar-Kaminska B et al., 2016 |
| | | | | Promotes invasion and metastasis by changing the intracellular Ca2+ concentration | Liu J et al., 2012 |
| | | Non-small cell lung cancer | Promote | Promotes EMT and enhances the dryness of OSCC through JAK2/STAT3 signaling pathway | Chen Y et al., 2020 |
| | | Oral squamous cell carcinoma | Promote | Promotes tumor progress by inducing angiogenesis and lymphangiogenesis | Unver N et al., 2021 |
| | | Pancreatic cancer | Promote | CCL21 and SPARCL1 may contribute to the drug resistance of ovarian cancer | Yin F et al., 2013 |
| | | Ovarian cancer | Promote | | |
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Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------------|--|--|--|--------------------------------|
| CCL22 | CCR4 | Colon cancer | Inhibit | Inhibits migration and invasion, and weakens their stem cell-like phenotype | Rong Y et al., 2017 |
| | | Lung cancer | Inhibit | Dome nanocapsules can effectively deliver CCL21 to maintain anti-tumor activity and inhibit tumor growth | Kar U et al., 2011 |
| | | Adenocarci-noma | Inhibit | Inhibits tumor growth and metastasis | Yousefieh N et al., 2009 |
| | | Non-small cell lung cancer | Inhibit | CCL21-DC overcomes drug resistance of immunotherapy and produces systemic tumor-specific immunity | Salehi-Rad R et al., 2023 |
| | | Neuroblast-oma | Inhibit | The new nano-preparation of CCL21 is an effective treatment for neuroblastoma | Poelaert B et al., 2020 |
| | | Oral cancer | Promote | Cultivates pre-tumor environment by promoting cell transformation and Treg infiltration | Huang Y et al., 2019 |
| | | Gastric cancer | Promote | Retinal opacification is a suitable microenvironment for migration, survival and metastasis. The CCL22-CCR4 axis is helpful to this selective permeation process | Cao L et al., 2014 |
| | | Prostate cancer | Promote | CCL17 and CCL22 promote the migration and invasion of prostate cancer cells by enhancing Akt phosphorylation | Maolake Aerken et al., 2017 |
| | | Lung cancer | Promote | RANKL-induced chemokines derived from CCL22/macrophages produced by osteoclasts promote bone metastasis | Nakamura E et al., 2006 |
| | | Squamous cell carcinoma of head and neck | Promote | CCR4/CCL22 promotes lymph node metastasis in head and neck squamous cell carcinoma | Takahiro T et al., 2013 |
| CCL23 | CCR1 | Ovarian cancer | Promote | Promotes the immunosuppression of TME by inducing depleted T cell phenotype | Kamat K et al., 2022 |
| CCL24 | CCR3CCR2B CCR5 | Hepatocellu-lar carcinoma | Promote | Progress of CCL-23 inhibits liver cancer through CCR1/AKT/ESR1 feedback loop | Meng J et al., 2021 |
| | | Breast cancer | Promote | CCL24 leads to HCC malignant tumor through RhoB-VEGFA-VEGFR2 angiogenesis pathway | Jin I et al., 2017 |
| CCL25 | CCR9 | Ovarian cancer | Promote | Promotes invasion by regulating various EMT markers | Zhang Z et al., 2016 |
| | | | Promotes proliferation and up-regulates anti-apoptosis signal transduction | Promotes proliferation and up-regulates anti-apoptosis signal transduction | Johnson-Holiday S et al., 2011 |
| | | | Promote | Contributes to migration and invasion | Johnson S et al., 2010 |
| | | | Promote | Inhibits cisplatin-induced apoptosis and supports the drug resistance | Johnson S et al., 2010 |
| | | | Promote | The expression of CCR9 was positively correlated with tumor size, lymph node metastasis, TNM late stage and overall survival rate | Zhong Y et al., 2015 |
| | | Non-small cell lung cancer | Promote | CCR9/CCL25 interaction induces migration, invasion, anti-apoptosis and tumorigenesis of NSCLC cells | Li B et al., 2015 |
| | | Hepatocellu-lar carcinoma | Promote | Induces the tumorigenesis by activating PI3K/Akt pathway | Li B et al., 2015 |
| | | | | Promotes the migration and invasion of HCC cells by regulating EMT markers | Zhang Z et al., 2016 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------|----------------------------------|-----------------|--|--------------------------------|
| CCL26 | CCR3 | Breast cancer | Promote | CCR9 enhances proliferation and tumorigenicity | Zhang Z et al., 2014 |
| | | | Promote | Activates Akt in PI3K-dependent and FAK-independent ways to promote cisplatin resistance in breast cancer cells | Johnson-Holiday C et al., 2011 |
| | | Pancreatic cancer | Promote | Promotes the invasion of PDAC by activating PI3K/AKT/mTOR pathway | Chen X et al., 2021 |
| | | Colon cancer | Promote | Participates in tumor progression by regulating EMT signaling pathway | Sun A et al., 2022 |
| CCL27 | CCR10 | N/A | | Participates in promotion and invasion by stimulating tumor-associated macrophage infiltration | LAN Q et al., 2018 |
| CCL28 | CCR3 | Breast cancer | Promote | Promotes proliferation and inhibits apoptosis, which may be regulated by Bcl-2 | Lin F et al., 2013 |
| | | | Promote | Promotes tumor progress through ERK/MAPK-mediated anti-apoptosis and metastasis signaling pathway | Yang X et al., 2017 |
| | | Pancreatic ductal adenocarcinoma | Promote | CCL28 blockade can inhibit tumor growth through tumor-cell-internal and external mechanisms | Yan J et al., 2021 |
| | | Ovarian cancer | Promote | Hypoxia induces CCL28 series to recruit Treg cells to promote cancer progression through tumor-specific immune paralysis | Facciabene A et al., 2012 |
| | | Lung adenocarcinoma | Promote | Hypoxia induces CCL28 series to recruit Treg cells to enhance angiogenesis of lung adenocarcinoma | Liu B et al., 2021 |
| | | Hepatocellular carcinoma | Promote | Hypoxia-induced CCL28 series promotes angiogenesis by targeting CCR3 | Huang G et al., 2016 |
| | | | Promote | Hypoxia-induced CCL28 series promotes the recruitment of regulatory T cells and tumor growth | Ren L et al., 2016 |
| | | | Promote | High expression of CCL28 in hypoxic microenvironment promotes migration and invasion | Zhou Y et al., 2013 |
| | | | Inhibit | Transcription activates CCL28, inhibits M2 polarization of macrophages and prevents immune escape | Liu S et al., 2024 |
| | | Oral squamous cell carcinoma | Inhibit | CCL28 series-induced RAR β expression inhibits bone invasion of oral squamous cell carcinoma | Park J et al., 2019 |
| | | | Promote | Overexpression of CXCL1-1 and its receptor CXCR2-2 promotes tumor invasion | Cheng W et al., 2011 |
| | | Gastric cancer | Promote | CXCL1 promotes tumor growth by activating VEGF pathway | Wei Z et al., 2015 |
| | | | | Drives cells into the lymphatic system by activating integrin β 1/FAK/AKT signaling | Wang Z et al., 2017 |
| | | Colon cancer | Promote | Enhances metastasis by cell migration, MMP-7 expression and EMT | Xu T et al., 2018 |
| | | | | | |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------------|----------------------------|-----------------|---|-------------------------|
| CXCL2 | | Hepatocellular carcinoma | Promote | CXCL1 plays a key role in the growth and apoptosis of HCC | Han K et al., 2015 |
| | | Prostate cancer | Inhibit | Inhibits malignant tumor, limits tumor cells from escaping from primary tumor and strengthens growth stagnation | Benelli R et al., 2013 |
| | | Ovarian cancer | Inhibit | MIR-27b-5p may inhibit the progression of ovarian cancer by targeting CXCL1 | Liu C et al., 2020 |
| | | Epithelial ovarian cancer | Promote | CXCL2 plays an important role in platinum tolerance of epithelial ovarian cancer (EOC) | Nie S et al., 2021 |
| | | Colon cancer | Promote | Promotes tumorigenesis through G1-2 and Gq/11, and contributes to CSC characteristics | Chen M et al., 2019 |
| | | | | The adhesion and growth is mediated by CXCL2-CXCR2 signal and αV integrin-dependent adhesion to ECM protein | Lepsenyi M et al., 2021 |
| | | | | ETTL3 promotes lung metastasis by targeting the m6A-Snail-CXCL2 axis to recruit M2-type immunosuppressed macrophages | Ouyang P et al., 2024 |
| | | | | Promotes the infiltration of M2 macrophages and metastasis of tumor cells | Bao Z et al., 2022 |
| | | Non-small cell lung cancer | Promote | CXCL2 contributes to the resistance of ANODINI in NCI-H1975 cells | Lu J et al., 2019 |
| | | Oral squamous cell cancer | Promote | CXCL2 synthesized by oral squamous cell carcinoma is involved in cancer-related bone destruction | Oue E et al., 2012 |
| CXCL3 | CXCR2 | Gastric cancer | Promote | Omental adipocytes trigger GC cells to form an invasive phenotype through CXCL2 secretion, induce angiogenesis, cell growth and metastasis | Natsume M et al., 2020 |
| | | Hepatocellular cancer | Inhibit | Overexpression of CXCL2 inhibits and promotes apoptosis | Ding J et al., 2018 |
| | | Osteosarc-oma | Inhibit | MIR-532-5p plays an anti-tumor role in OS cells by regulating CXCL2 | Ma Y et al., 2020 |
| | | Prostate cancer | Promote | Overexpression of CXCL3 type cancer can enhance the carcinogenic potential of prostate | Gui S et al., 2016 |
| | | Uterine cervix cancer | Promote | Overexpression of CXCL3 promotes the tumorigenic potential of cervix cancer cells pass through MAPK/ERK pathway | Qi Y et al., 2019 |
| | | Pancreatic cancer | Promote | Promotes metastasis through a novel myofibroblast-hijacked cancer escape mechanism | Sun X et al., 2021 |
| | | Oral squamous cell cancer | Promote | Overexpression of CXCL3 affects the malignant behavior through MAPK signaling pathway | Wong J et al., 2021 |
| | | Colon cancer | Promote | Promotes the malignant behavior of tumor cells in an ERK-dependent manner | Cheng Y et al., 2023 |
| | | Colon cancer | Promote | Through the negative immuno-modulatory function of CXCR3, non-platelet derived CXCL4 can be hijacked by cancer cells to escape the host immune system | Deng S et al., 2019 |
| | | Breast cancer | Promote | Specific CXCL4-CXCL12 heterodimers inhibit migration at least partially by competing for CXCR4 receptors | Nguyen K et al., 2021 |
| CXCL5 | CXCR1 CXCR2 | Ovarian cancer | Promote | CXCL5 promotes ovarian cancer and achieves cell proliferation by up-regulating the expression of cyclin D1 | Jian F et al., 2018 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------|-----------------------|-----------------|--|-------------------------------|
| | | Liver cancer | Promote | Increases migration and invasion through autocrine and paracrine mechanisms | Xu X et al., 2014 |
| | | Colon cancer | Promote | Promotes metastasis by activating ERK/Elk-1/Snail and AKT/GSK3β/β-catenin | Zhao J et al., 2017 |
| | | Uterine cervix cancer | Promote | Induces tumor angiogenesis by enhancing the expression of FOXD1 mediated by AKT/NF-κB pathway | Chen C et al., 2019 |
| | | Pancreatic cancer | Promote | Contributes to oncogenic potential of Hela uterine cervix cancer cells | Feng X et al., 2018 |
| | | | | CXCL5 promotes PC cell growth and EMT process | Wang Z et al., 2022 |
| | | Breast cancer | Promote | Necrotic apoptosis before invasion promotes migration and invasion through CXCL5-CXCR2 axis | Ando Y et al., 2020 |
| | | | | DDR1/CXCL5 promotes immune infiltration of Tregs and drives tumor growth and metastasis | Li H et al., 2023 |
| | | | | CXCL5 is sufficient to promote the proliferation and colonization of breast cancer cells in bones | Romero-Moreno, R et al., 2019 |
| | | | | Increases cancer progression through ERK/MSK1/Elk-1/Snail signaling pathway | Xu Y et al., 2013 |
| | | | | A S100A14-CCL2/CXCL5 signal axis drives breast cancer metastasis | Li X et al., 2020 |
| | | Cervical cancer | Promote | Promotes proliferation and migration through ERK signaling pathway and autocrine pathway | Chen S et al., 2021 |
| | | Bladder cancer | Promote | CXCL5 may promote mitomycin resistance by activating EMT and NF-κB pathways | Wang C et al., 2018 |
| | | | | CXCL5 is very important for the growth and progress of bladder tumor | Zheng J et al., 2014 |
| | | | | Promotes migration and invasion through activating PI3K/AKT-induced MMP2/MMP9 up-regulation | Gao Y et al., 2015 |
| | | Prostate cancer | Promote | Enhances cell migration and EMT through early growth response –1/ snail signaling pathway | Guo B et al., 2011 |
| | | | | CXCL5 can promote the growth of LNCaP cells by acting on its own receptor CXCR2 | Qi Y et al., 2014 |
| | | Lung cancer | Promote | Promotes immune escape through autocrine and paracrine mechanisms by up-regulating the chemotaxis of lung cancer and neutrophils | Sun D et al., 2024 |
| | | | | Promotes proliferation and movement by activating MAPK/ERK1/2 and PI3K/AKT | Wang L et al., 2018 |
| | | Gastric cancer | Promote | The interaction between TAMs and gastric cancer cells promotes chemotherapy resistance through CXCL5/PI3K/AKT/mTOR pathway | Su P et al., 2022 |
| | | | | Promotes tumor occurrence by regulating tumor immunosuppression mediated by NF-κB and Wnt/β-catenin signals | Liu L et al., 2020 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------------|---------------------------------|-----------------|--|---------------------------|
| CXCL6 | CXCR1 CXCR2 | Osteosarcoma | Promote | Promotes migration and invasion in autocrine- and paracrine-dependent manners | Dang H et al., 2017 |
| | | Glioblastoma | Promote | Promotes the tumorigenesis and angiogenesis through JAK-STAT/NF- κ B | Mao P et al., 2023 |
| | | Cholangio- carcinoma | Promote | Promotes tumor metastasis and recurrence by recruiting infiltrating neutrophils | Zhou S et al., 2014 |
| | | Non-small cell lung cancer | Promote | A2AR-mediated CXCL5 upregulation on macrophages promotes NSCLC progression via NETosis | Lei Q et al., 2024 |
| | | Lung cancer | Inhibit | Inhibits tumor immunity by regulating PD-1/PD-L1 signal transduction | Xie X et al., 2022 |
| | | Non-small cell lung cancer | Promote | CXCL6 promotes the survival and metastasis of non-small cell lung cancer cells by down-regulating miR-515-5p | Li J et al., 2018 |
| | | Hepatocellular cancer | Promote | Activates IFN- γ /p38 MAPK/NF- κ B signal and promotes EMT and radiation resistance | Li X et al., 2023 |
| | | | | Promotes liver invasion through targeting MMP9 | Zheng Y et al., 2016 |
| | | Esophageal squamous cell cancer | Promote | Enhances the growth and metastasis of ESCC cells in vivo and in vitro | Zheng S et al., 2021 |
| | | Melanoma | Promote | (GCP)-2/CXCL6 induces angiogenesis and promotes tumor growth | Verbeke H et al., 2011 |
| CXCL7 | CXCR1 CXCR2 | Cholangiocarcinoma | Promote | Promotes the proliferation and invasion of cholangiocarcinoma cells | Guo Q et al., 2017 |
| | | Breast cancer | Promote | Promotes invasion, the expression of VEGF-C/D and heparanase | Yu M et al., 2010 |
| | | | | Secretion of CXCL7 by monocytes promotes the progress of breast cancer | Wang Y et al., 2021 |
| | | | | The interaction between breast cancer cells and monocytes promotes tumor progression through CXCL7-mediated signal transduction | Lin S et al., 2021 |
| | | Renal-cell carcinoma | Promote | CXCL7 /CXCR1/2 axis is the key driving factor for the growth of clear cell renal cell carcinoma | Grepin R et al., 2014 |
| | | Pancreatic cancer | Promote | IFN α - induced BST2 + tumor-associated macrophages promote immunosuppression and tumor growth through ERK-CXCL7 signal | Zheng C et al., 2024 |
| | | Triple negative breast cancer | Promote | The multiple positive feed-forward loops of MCT-1/IL-6/IL-6R/CXCL7/PD-L1 axis promotes the metastatic niche and immunosuppressive microenvironment | Aushia T et al., 2024 |
| | | Gastric cancer | Promote | Participates in the immunosuppression microenvironment by inducing PD-L1 (+) macrophages | Lin C et al., 2019 |
| | | Colon cancer | Promote | CXCL8 gene silencing significantly inhibits proliferation and invasion via PI3K/Akt/NF- κ B signaling | Ma J et al., 2015 |
| | | | | CXCL8 upregulates LSECtin through AKT, and promotes proliferation and invasion | Fang S et al., 2022 |
| CXCL8 | CXCR1 CXCR2 | Thyroid cancer | Promote | Promotes the tumor-promoting effect of TC cells | Coperchini F et al., 2024 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|-------------------------|---|-----------------|--|---|
| CXCL9 | CXCR3 CXCR7 | Ovarian cancer and gastric cancer | Promote | Promotes peritoneal metastasis | Awwad O et al., 2018 |
| | | Prostate cancer | Promote | Promotes tumor progress by inhibiting cytokines in T cells | Tan S et al., 2018 |
| | | Ovarian cancer | Inhibit | Inhibits tumor growth | Seitz S et al.,2022 |
| CXCL10 | CXCR2 CXCR3 CXCR4 | Breast cancer | Promote | Promotes proliferation and Tamoxifen-resistant MCF7 cells through AKT pathway | Wu X et al., 2020 |
| | | Gastric cancer | Promote | Induces migration through a novel crosstalk between Cxcr3 and Egfr receptor CXCL10 signal transduction promotes the metastasis of ING4 deficient breast cancer Targeted autophagy promotes T lymphocyte migration by inducing the expression of CXCL10 | Tsutsumi E et al., 2022 Tsutsumi E et al., 2023 Meng Q et al., 2022 |
| | | | | Promotes gastric invasive cancer through PI3K/AKT dependent MMP production | Zhou H et al., 2016 |
| | | Colon cancer | Promote | Enhances metastasis by triggering small GTP enzymes such as RhoA and cdc42 | Wang Z et al., 2021 |
| | | Cervical cancer | Promote | Promotes M2 polarization of macrophages in tumor microenvironment and enhances proliferation, migration and invasion via activating STAT3/NF-κB/CCL2 signal | Li A et al., 2024 |
| | | Ovarian cancer | Inhibit | Promotes CTL activation to inhibit ovarian cancer | Dong M et al., 2024 |
| | | Prostate cancer HER2 positive breast cancer Breast cancer | Inhibit | Enhances the killing effect of T cells and inhibits angiogenesis | Li W et al., 2021 |
| | | | | Inhibits proliferation and reduces PSA production by up-regulating CXCR3 receptor | Nagpal M et al., 2006 |
| | | | | Induces activation of CD8+ T cells to promote effect of immunotherapy on HER2-positive breast cancer | Zhang X et al., 2022 |
| | | Colon cancer Cervical cancer | Inhibit | controlling the self-regulation of CXCL10 and the characteristics of malignant tumor by mediating NF-κB signaling pathway | Jin W et al., 2017 |
| | | | | Inhibits tumor growth, increases CD8+ T cell infiltration and induces tumor blood vessels to normalize by making colorectal cancer cells overexpressing cetuximab | Yan W et al., 2023 |
| | | | | Enhances the radiotherapy effect of HeLa cells through cell cycle redistribution | Yang L et al., 2012 |
| CXCL11 | CXCR3 | Hepatocellular cancer | Promote | Promotes the proliferation and migration of HCC cells through LINC00152/miR-205-5p/CXCL11 axis | Liu G et al., 2022 |
| | | Squamous cell carcinoma of head and neck | Promote | Promotes tumor lymph node metastasis | Wang X et al., 2021 |
| | | Colon adenocarcinoma and rectal adenocarcinoma | Promote | Enhances the infiltration of TAMs into tumor environment, promotes EMT of cancer and promotes tumor metastasis by inducing the expression of TGF-β1 | Liu M et al., 2021 Zeng YJ et al., 2016 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------------|------------------------------|-----------------|---|---------------------------|
| CXCL12 | CXCR4 CXCR7 | Renal cancer | Promote | EP300/CBP promotes proliferation and migration by stabilizing CXCL11 mRNA level | Zeng X et al., 2022 |
| | | Oral cancer | Promote | Promotes tumor angiogenesis | Suyama T et al., 2005 |
| | | | | Promotes tumor growth and immune escape | Wang X et al., 2021 |
| | | | | Promotes early malignant transformation and tumor development | Xia J et al., 2011 |
| | | Cutaneous melanoma | Promote | Promotes the occurrence of precancerous lesions | Wang X et al., 2021 |
| | | | | Induces cytoskeleton remodeling and promotes tumor metastasis | Kawada K et al., 2004 |
| | | Breast cancer | Promote | Activates ERK pathway and enhances tumor invasion | Hwang H et al., 2020 |
| | | Multiple myeloma | Promote | Activates tyrosine kinase, induces MMP-2 and MMP-9 secretion and promotes tumor growth and metastasis | Pellegrino A et al., 2004 |
| | | Ovarian cancer | Promote | The high expression of CXCR3-A in endometriosis inhibits cytotoxic T cells and promotes the occurrence of precancerous lesions | Furuya M et al., 2007 |
| | | Basal cell carcinoma | Promote | Promotes the proliferation of human immortalized keratinocytes and promotes tumor growth | Furuya M et al., 2011 |
| | | Thyroid cancer | Promote | Promotes angiogenesis in metastatic THCA through EGF-EGFR positive feedback loop | Lo B et al., 2010 |
| | | Colon cancer | Promote | Inhibits proliferation, migration, induces apoptosis, and inhibits tumor growth | Liang J et al., 2021 |
| | | | | Down-regulation of CXCL11 inhibits cell growth and EMT | Fallahi P et al., 2018 |
| | | Pancreatic cancer | Promote | Induces invasion and EMT by activating NF-κB signaling pathway | Gao Y et al., 2018 |
| | | Cutaneous melanoma | Inhibit | Recruits immune cells, promotes bone marrow activation, enhances anti-tumor immune response and inhibit tumor growth | Sun L et al., 2019 |
| | | Renal cancer | Inhibit | Has immunosuppressive activity on tumor vasculature and tumor angiogenesis | Harlin H et al., 2009 |
| | | Gastric adenocarcinoma | Inhibit | Activates CXCR3 and up-regulates PD-L1 expression through STAT and PI3K-Akt pathways, thus improving the effectiveness of immunotherapy | Gacci M et al., 2009 |
| | | Lung cancer | Inhibit | Mediates the infiltration of cytotoxic T lymphocytes and inhibit angiogenesis | Zhang C et al., 2018 |
| | | | | Activates Th1, promotes M1 polarization in macrophages and inhibits tumor growth | Verbeke H et al., 2012 |
| | | Bladder Urothelial carcinoma | Inhibit | Induces CD8 T cell infiltration, inhibits angiogenesis and enhances the efficacy of immunotherapy | Pasini F et al., 2014 |
| | | | | Improves the chemosensitivity | Mitsuhashi A et al., 2021 |
| | | Lung cancer | Promote | The destruction of CXCL12 inhibits the growth and migration of lung cancer cells | Zhang Y et al., 2019 |
| | | | | | Imai H et al., 2010 |

Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|----------------|--------------------------------|-----------------|---|-------------------------|
| CXCL13 | CXCR4 CXCR7 | Gastric cancer Colon cancer | Promote | CXCL12 induces lung cancer cell migration by polarized mtDNA redistribution | Ma J et al., 2014 |
| | | | Promote | Promotes distant metastasis by activating CXCR4 | Ishigami S et al., 2007 |
| | | Pancreatic cancer | Promote | Silencing CXCL12 inhibits proliferation, invasion and angiogenesis by down-regulating MAPK/P3K/AP-1 signaling | Ma J et al., 2018 |
| | | | Promote | CXCL12–CXCR4 promotes the proliferation of pancreas and invades cancer cells | Shen B et al., 2014 |
| | | Breast cancer | Promote | Signal transduction through CXCL12–CXCR4 is an important for migration | Nguyen K et al., 2020 |
| | | | Promote | CXCL12-γ was identified as an effective transfer promoter | Lei P et al., 2015 |
| | | Prostate cancer | Promote | Promotes peripheral invasion of prostate cancer | Zhang S et al., 2009 |
| | | Non-small cell lung cancer | Promote | The interaction between cancer-associated fibroblasts and tumor epithelial cells through CXCL12/CXCR4 axis promotes tumor proliferation | Wald O et al., 2011 |
| | | Colon cancer | Promote | Enhances the metastatic potential through PI3K/Akt/mTOR pathway | Ma J et al., 2017 |
| | | Bladder cancer | Promote | Down-regulating CXCR4/CXCL12 axis can reduce cancer growth and metastasis | Song Z et al., 2015 |
| | | | Promote | The combination of CXCR7 and CXCL12 promotes lung metastasis | Wang M et al., 2018 |
| | | Non-small cell lung cancer | Promote | Enhances immune escape by inhibiting autophagy degradation of PDL1 mediated by P62 | Zhang Z et al., 2023 |
| | | Intestinal gastric cancer | Promote | CXCL12–CXCR4 biological axis is involved in regulating the metastasis of non-small cell lung cancer | Phillips R et al., 2003 |
| | | Ovarian cancer | Promote | CXCL12/CXCR7 may be the biological axis of proliferation, invasion and lymph node and liver metastasis | Xin Q et al., 2016 |
| | | Thyroid carcinoma | Promote | Cancer-related fibroblasts induce EMT and cisplatin resistance through CXCL12/CXCR4 axis | Zhang F et al., 2020 |
| | | Ovarian cancer | Promote | CXCL12–CXCR4 biological axis plays an important role in the process of thyroid cancer metastasis | Wu Y et al., 2012 |
| | | Cervical cancer | Inhibit | CXCL12 promotes cell invasion by inhibiting the expression of ARHGAP10 | Luo N et al., 2020 |
| | | Lung cancer | Promote | Inhibits anchorage independent cell growth | Yadav S et al., 2016 |
| | | Colon cancer | Promote | Mediates radiotherapy resistance of lung cancer by activating Akt | Geng S et al., 2020 |
| | | | | Promotes the migration of lung cancer cells | Zhao C et al., 2021 |
| | | | | Plays a key role in carcinogenesis, tumor development, metastasis and recurrence | Qi X et al., 2013 |

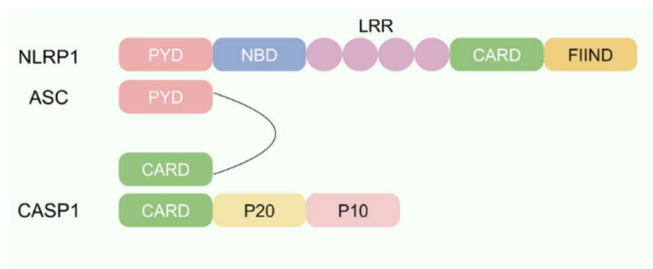
Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|------------|------------------|-------------------------------|-----------------|---|------------------------|
| CXCL 14 | CXCR4 CXCR7 | Prostate cancer | Promote | CXCL13-CXCR5 axis promotes growth, migration and invasion through PI3K/AKT | Zhu Z et al., 2015 |
| | | | | Promotes the occurrence of intestinal tumors by activating epithelial AKT signal | Zhao Q et al., 2021 |
| | | Renal-cell carcinoma | Promote | CXCL13 participates in AR regulating the growth of prostate cancer xenograft in mice | Tian Q et al., 2019 |
| | | Human osteosarcoma | Promote | Promotes proliferation and migration by binding to CXCR5 and activating PI3K/AKT/mTOR signaling | Zheng Z et al., 2019 |
| | | Breast cancer | Promote | Promotes migration through phospholipase C β (PLC beta), protein kinase C α (PKC α), c-Src and nuclear factor- κ B (NF- κ B) | Liu J et al., 2020 |
| | | Breast cancer | Promote | CXCL13 inhibition causes the decrease of tumor growth via CXCR5/ERK signaling | Xu L et al., 2018 |
| | | Breast cancer | Inhibit | Triggers effective anti-tumor immunity by attracting immune cells to infiltrate | Ma Q et al., 2021 |
| | | Cervical cancer | Inhibit | Down-regulation of DNA methylation-dependent CXCL13 may promote tumor occurrence and progress | Ma D et al., 2020 |
| | | Non-small cell lung cancer | Promote | CXCL14 promotes metastasis of non-small cell lung cancer through ACKR2-dependent signaling pathway | Zhang Z et al., 2023 |
| | | Breast cancer | Promote | The CXCL14/ACKR2 pathway is a clinically relevant stimulator of EMT, invasion and metastasis | Sjoberg E et al., 2019 |
| CXCL 15 | CXCR2 | Ovarian cancer | Promote | Up-regulating CXCL14 promotes the proliferation of ovarian cancer cells | Li X et al., 2021 |
| | | Pancreatic cancer | Promote | CXCL14 significantly increases the invasion of pancreatic cancer cells | Wente M et al., 2008 |
| | | Colon cancer | Inhibit | Inhibits the migration, invasion and EMT by inhibiting NF- κ B signal transduction | Cao B et al., 2013 |
| | | Breast cancer | Inhibit | Inhibits cell proliferation and invasion, and weakens the growth and lung metastasis | Gu X et al., 2012 |
| | | Triple negative breast cancer | Inhibit | Inhibits tumor progress by changing the immune characteristics of tumor microenvironment, and it is mediated in a T cell-dependent manner | Gibbs C et al., 2024 |
| | | N/A | | | |
| | | Lung cancer | Promote | Promotes the viability and invasion and leads to lung cancer metastasis | Zhou W et al., 2011 |
| | | | | Promotes proliferation and invasion by regulating NF- κ B pathway | Liang K et al., 2018 |
| | | | | CXCL16-CXCR6 regulates lung cancer cell viability and invasion | Hu W et al., 2014 |
| | | Breast cancer | Promote | Blocking CXCR6-CXCL16/CXCR4-CXCL12 receptor-ligand interaction prevent brain metastasis | Chung B et al., 2017 |
| CXCL 16 | CXCR6 SR-PSOX | Prostate cancer | Promote | CXCL16/CXCR6 may be another independent axis of chemokines for bone metastasis | Zhou W et al., 2010 |
| | | | | | |

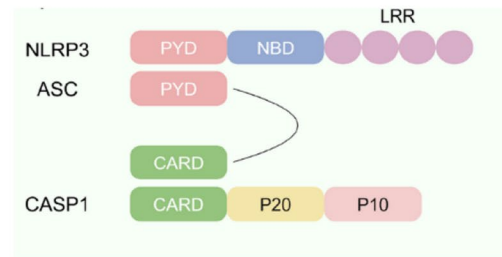
Table 6 (continued)

| chemokines | Receptor | Cancer types | Promote/inhibit | Mechanisms | References |
|--------------|----------------|---------------------------------|-----------------|---|--------------------------|
| CX3C XCL1 | CX3CR1 XCRI | Ovarian cancer | Promote | Maintains the high invasion and migration ability of cells by combining with CXCR6 | Yang Y et al., 2015 |
| | | Colon cancer | Promote | CXCL16/CXCR6 may be involved in the proliferation, invasion and metastasis | Fu Y et al., 2017 |
| | | Gastric cancer | Promote | Promotes tumorigenesis by enhancing ADAM10-dependent CXCL16/CXCR6 axis activation | Han J et al., 2021 |
| | | | | Promotes cell proliferation | Takiguchi G et al., 2016 |
| | | | | Promotes tumor progress through the expression of Ror1 mediated by STAT3 | Ikeda T et al., 2020 |
| | | | | Promotes tumorigenesis via ADAM10-dependent CXCL16/CXCR6 axis and activates Akt and MAPK signaling | Han J et al., 2023 |
| | | Adenocarci-noma | Promote | Enhances migration, invasion and adhesion with endothelial cells | Singh R et al., 2016 |
| | | Thyroid cancer | Promote | Enhances migration and invasion, and changes the phenotype of macrophages into M2 macrophages | Zhao S et al., 2016 |
| | | Breast cancer | Inhibit | Inhibits migration and invasion, and induces apoptosis of breast cancer cells | Fang Y et al., 2014 |
| | | | | Inhibits the migration and invasion of breast cancer cells in vitro | Fang Y et al., 2013 |
| | | Renal cancer | Inhibit | Inhibits migration of renal cell induced by CXCL16 | Gutwein P et al., 2009 |
| | | Colon cancer | Inhibit | CXCL16 can inhibit the metastasis of liver through NKT cells in CRC | Kee J et al., 2013 |
| CX3C XCL1 | CX3CR1 XCRI | | | Inhibits liver metastatic by promoting tumor-associated macrophage α TNF- induced apoptosis | Kee J et al., 2014 |
| | | Breast cancer | Promote | Promotes the proliferation of drug-resistant cells by activating mTOR pathway | Bai Y et al., 2015 |
| | | | | The activation of ERK/HIF-1 α /EMT is involved in migration induced by XCL1 | Do H et al., 2021 |
| | | | | Improves the survival rate by promoting cancer immunity | Zhou W et al., 2020 |
| XCL2 | XCRI | Clear cell renal cell carcinoma | Promote | Inhibit apoptosis and promotes the proliferation, migration, invasion and epithelial-mesenchymal transition | Cao Q et al., 2022 |

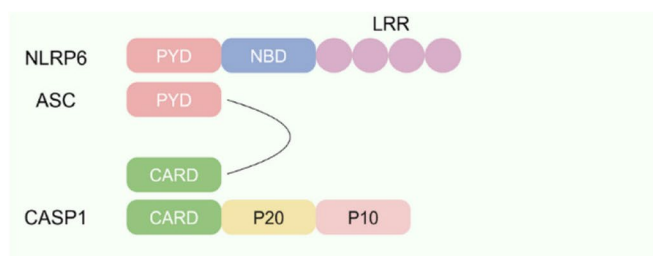
A NLRP1 inflammasome



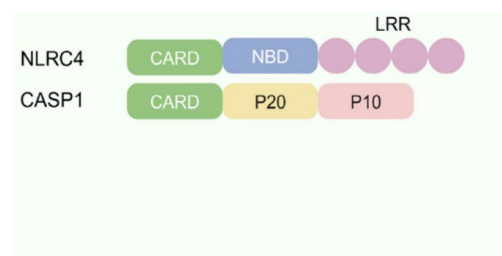
B NLRP3 inflammasome



C NLRP6 inflammasome



D NLRC4 inflammasome



E AIM2 inflammasome

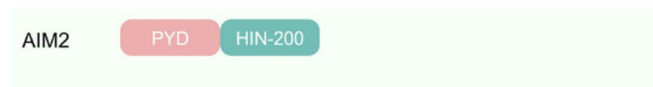


Fig. 4 Nucleotide-binding domain, leucine-rich repeat containing receptors (NLRs) upon activation form a multiprotein complex known as the “inflammasome”. The inflammasome complex consists of an NLR protein, the apoptosis-associated speck-like protein (ASC) (an adaptor protein) and a caspase. The detailed information on the structure of NLRP1, NLRP3, NLRP4, NLRP6, and AIM2 inflammasome are shown. A The Nlrp1 gene encodes an N-terminal PYD domain, a central NBD domain, a C-terminal LRR domain, and a C-terminal FIIND domain. NLRP1 directly recruits procaspase-1 through its CARD domain. B The Nlrp3 gene encodes an N-terminal PYD domain, a central NBD domain, and a C-terminal LRR domain. NLRP3 lacks a CARD domain and therefore, interacts with ASC to recruit procaspase-1. C The Nlrp6 gene encodes an N-terminal PYD domain, a central NBD domain, and a C-terminal LRR domain. NLRP6 is recruited to the “specks” formed by ASC oligomerization, leading to procaspase-1 activation. D The Nlr4 gene encodes an N-terminal CARD domain, a central NBD domain, and C-terminal LRR. The interaction of NLRC4 with ASC is unclear. NLRC4 results in pro-IL-1b and pro-IL-18 processing or caspase-1-dependent pyroptosis via an ASC-dependent mechanism or an ASC-independent mechanism. E The AIM protein consists of an N-terminal PYD domain, mediating homotypic interactions with ASC and a C-terminal HIN-200 domain for DNA binding

with high NLRP1 expression generally have a better prognosis in lung adenocarcinoma (LUAD) and pancreatic adenocarcinoma (PAAD) [321]. The activation of the NLRP1 sensor protects against colitis-associated CRC through mechanisms dependent on effector cytokines [315]. NLRP1 is also positively correlated with an increased risk of prostate cancer [322]. The dysfunctions in the NLRP1 pathway are also linked to skin cancers, including melanoma, Kaposi sarcoma, and squamous cell carcinoma [323]. Increased expression of NLRP1 was significantly associated with immune cell infiltration in gastric cancer [324]. An endogenous thioredoxin (TRX) has been identified as a binder to NLRP1 and inhibits NLRP1

inflammasome activation. This opens opportunities for therapeutic intervention in NLRP1 inflammasome activation in the future.

The role of NLRP3 in inflammation-mediated carcinogenesis

The NLRP3 inflammasome is widely present in immune cells. As the most thoroughly studied inflammasome, NLRP3 inflammasome includes the sensor NLRP3, the adaptor ASC, and the effector caspase-1. NLRP3 consists of three domains, a central nucleotide-binding domain (NBD), a C-terminal leucine-rich repeat (LRR), and an N-terminal PYD. ROS production, ionic flux, mitochondrial dysfunction, and lysosomal damage are the four

main activation mechanisms of NLRP3 inflammasome. Under the stimulation of microbial or endogenous molecules like TLR ligands, NF- κ B is activated, and pro-IL-1 β , pro-IL-18, and NLRP3 are induced. Then, various stimuli like extracellular ATP, glucose, bacterial and virus infection, mitochondrial damage/dysfunction, and more facilitated the maturation of pro-IL-1 β and pro-IL-18, promoting the activation of NLRP3 inflammasome.

NLRP3 and NLRP3 inflammasome members like caspase-1, IL-1 β and IL-18 are potential therapeutic targets due to their role in inflammation-associated diseases and cancer. Recent research reported that NLRP3 inflammasome-related genes were dysregulated in 15 cancers [325]. NLRP3 is overexpressed and activated in several cancers, like non-small cell lung cancer [326], melanoma [327], and more. Patients with cancer have a higher frequency of *Nlrp3* polymorphism, such as pancreatic cancer [328], melanoma [329], and others. They were introduced as a double-edged sword in tumorigenesis. On one hand, the NLRP3 inflammasome promotes tumor formation and metastasis in breast cancer [330], and overexpressed human IL-1 β in mice stomach increases the risk of gastric cancer [331]. NLRP3 affects the adaptive immune system to promote carcinogenesis in pancreatic cancer [332], and pharmacologic blocking of NLRP3 enhances the efficacy of immunotherapy [318]. NLRP3 signaling promotes T cell differentiation into tumor-promoting T cell populations and restricts antitumor T cell immunity [333]. NLRP3-mediated IL-1 β production promotes pancreatic ductal adenocarcinoma by immunosuppression [334]. On the other hand, mice deficient in NLRP3 are hypersusceptible to carcinogen-induced colitis-associated cancer (CAC) [335]. While NLRP3 inhibition via mitophagy prevents CAC, indicating a harmful role of NLRP3 in CAC [336]. The differences in gut microbiota, genetic background, and experimental technique may explain the inconsistent effects of NLRP3 in CAC. Besides, NLRP3 is down-regulated in hepatic cancer tissues [337], and the up-regulation of NLRP3 inhibits hepatic cancer cell growth [338]. The varying functions of NLRP3 in the etiology of cancer present novel prospects and obstacles in comprehending its dual roles of pro- and anti-tumorigenic effects. The tumor microenvironment may have an impact on these many functions by altering NLRP3 activity. Furthermore, it appears that NLRP3 plays distinct roles in the pathophysiology of cancers that originate in particular organs due to its variable expression in different cells and tissues, and elucidating the role and mechanism of NLRP3 in different cancers will contribute to precision therapy.

The role of NLRC4 in inflammation-mediated carcinogenesis

NLRC4 contains three domains: the N-terminal CARD domain, the C-terminal LRR domain, and the NACHT

domain. NACHT domain is composed of the NBD, helical domain 1 (HD1), winged helix domain (WHD), and helical domain 2 (HD2). A functional type IV secretion system (T4SS) for *L. pneumophila* or a functional type III secretion system (T3SS) for *S. Typhimurium*, *S. flexneri*, and *P. aeruginosa* is necessary for NLRC4 inflammasome activation [339]. Regulatory mechanisms also impact NLRC4 activation, mostly comprising transcription control and post-translation changes, specifically phosphorylation and perhaps ubiquitination. Interferon regulatory factor (IRF) 8 induces NLRC4 transcription, and infection with *S. typhimurium* induces NLRC4 phosphorylation at serine 533 [340]. Flagellin also induces NLRC4 phosphorylation but can't activate NLRC4 inflammasome [341]. NLRC4 has been shown to express differently yet variably across various tumor tissues. For example, NLRC4 mRNA level is increased in stomach cancer, glioma, and breast cancer, but it is decreased in colorectal cancer compared with normal adjacent tissues [339]. However, NLRC4 mRNA is almost unchanged in hepatocellular carcinoma [342]. And there is no agreement regarding its role in any form of cancer development, even in the same tumor type. Knocking out NLRC4 promotes tumor formation in colon cancer [343] and melanoma [344]. Higher NLRC4 expression is closely related to poor prognosis in breast cancer [345] and glioma [346]. Activated NLRC4 inflammasome activates IL-1 β , which promotes breast cancer progression by adipocyte-mediated vascular endothelial growth factor A (VEGFA) expression and angiogenesis [345]. Besides, in colitis-associated tumorigenesis, the role of NLRC4 depends on how NAIPs function. NAIPs have protective effects on colon cancer development, which is independent of NLRC4 [347]. Even though there isn't a consensus on the involvement of NLRC4 or NAIP in cancer, the necessity to employ littermate controls in these experiments will improve future research consistency and reproducibility. Furthermore, the discovery of NAIPs' or NLRC4's inflammasome-independent activities in carcinogenesis is intriguing because it might reveal new route targets for the creation of immunotherapies.

The role of NLRP6 in inflammation-mediated carcinogenesis

As a recently identified receptor in the mammalian innate immune system, the NOD-like receptor family pyrin domain containing 6 (NLRP6) was formerly known as PYPAF5 [348]. It comprises N-terminal PYD, central NBD, and C-terminal LRR domain. Following the recognition of PAMP and DAMP, NLRP6 forms the NLRP6 inflammasome by assembling with ASC and Caspase-1. This process facilitates the maturation of pro-IL-1 β and pro-IL-18, as well as gasdermin-D-induced pyroptosis. NLRP6 functions as an inflammasome or

Table 7 The role and mechanisms of inflammasome in cancer

| Inflammasome | Cancer types | Promote/inhibit | Mechanisms | References |
|---|----------------------------------|-----------------|---|---|
| NLRP1 | Breast cancer | Promote | Promotes tumorigenesis and proliferation | Wei, Y et al.,2017 |
| | Prostate cancer | Promote | Enhances tumorigenesis by promoting the maturation and release of pro-inflammatory cytokines IL-1 β | Liang, K et al.,2023 |
| | Metastatic melanoma | Promote | Enhances inflammasome activation and suppresses apoptosis | Zhai, Z et al.,2017 |
| High p62 (NLRP1 inhibitor) | Cutaneous SCC cells | Promote | Suppresses the NLRP1 inflammasome and increases stress resistance | Hennig, P et al.,2022 |
| | Non-melanoma skin cancer | Promote | Lower NLRP1 level is associated with worse clinical outcomes and poorer prognosis | Tan, J et al.,2023 |
| | Triple-negative breast cancer | Inhibit | Contribute to the antiproliferative effects of celecoxib | Arzuk, E et al.,2024 |
| NLRP3 | Colorectal cancer | Promote | Contributes to cell migration and invasion Promotes invasion and migration | Deng, Q et al.,2019 Zhang, L et al.,2022 |
| | Ovarian cancer | Promote | Contributes to the malignant process of DDP-resistant cells | Li, W et al.,2023 |
| 3,4-Methylenedioxy- β -nitrostyrene (MNS) (NLRP3 inhibitor) | Pancreatic ductal adenocarcinoma | Inhibit | Inhibits inflammation and restores immunity | Liu, H et al.,2020 |
| MCC950 (NLRP3 inhibitor) | Pancreatic ductal adenocarcinoma | Inhibit | Inhibits LPS-induced pancreatic adenocarcinoma inflammation | Yaw, A et al.,2020 |
| Vitamin D (NLRP3 inhibitor) | Breast cancer | Inhibit | Mediates the modulation of stemness | Zheng, W et al.,2024 |
| Caffeic Acid Phenethyl Ester (NLRP3 inhibitor) | Colon cancer | Inhibit | Inhibits NLRP3 Inflammasome | Dai, G et al.,2020 |
| RNF20 (NLRP3 inhibitor) | Liver cancer | Inhibit | Reduces cell proliferation and Warburg effect by promoting NLRP3 ubiquitination | Liu, D et al.,2024 |
| Metformin (NLRP3 inhibitor) | Colorectal cancer | Inhibit | Delays tumor progression | Liu, G et al.,2024 |
| Fermented Quercetin (NLRP3 inhibitor) | Colorectal cancer | Inhibit | Decreases resistin-induced chemo-resistance to 5-Fluorouracil | Lee, K et al.,2022 |
| Inhibition of HDAC2 (NLRP3 agonist) | Breast cancer | Inhibit | Sensitizes anti-tumour therapy by promoting NLRP3/GSDMD-mediated pyroptosis | Guan, X et al.,2024 |
| H. pylori CagA (NLRP3 agonist) | Gastric cancer | Promote | Promotes invasion and migration by activating NLRP3 inflammasome pathway | Zhang, X et al.,2021 |
| | Cervical cancer | Promote | Promotes migration, invasion and EMT by regulating macrophage differentiation | Zhou, H et al.,2020 |
| miR-223-3p (NLRP3 inhibitor) | Prostate cancer | Inhibit | Reduces tumor growth and immunosuppression | Zhang, L et al.,2019 |
| miR-22 (NLRP3 agonist) | Prostate cancer | Inhibit | Inhibits PI3K/AKT signaling pathway | Wu, H et al.,2021 |

Table 7 (continued)

| Inflammasome | Cancer types | Promote/inhibit | Mechanisms | References |
|--------------|---------------------------|-----------------|---|---------------------------|
| NLRC4 | Glioma | Promote | Contributes to a poor prognosis | Lim, J et al.,2019 |
| | Lung adenocarcinoma Cells | Promote | Induces apoptosis and immune infiltration | Hu, B et al.,2023 |
| | Melanoma tumor | Promote | Suppresses tumor growth in an inflammasome-independent manner | Janowski, A et al.,2016 |
| | Breast cancer | Inhibit | Weakens the ability of flagellin to inhibit tumor proliferation | Zhang, J et al.,2019 |
| NLRP6 | Colorectal cancer | Promote | Promotes liver metastasis by modulating M-MDSC-induced immunosuppressive microenvironment | Chang, L et al.,2024 |
| | Gastric cancer | Inhibit | Suppresses tumorigenicity | Wang, Q et al.,2018 |
| | | | Suppresses tumor growth via GRP78 ubiquitination | Wang, X et al.,2020 |
| AIM2 | Prostate cancer | Promote | Mediates P14ARF-Mdm2-P53-dependent cellular senescence | Wang, H et al.,2018 |
| | | | IFN-inducible AIM2 protein is a cytosolic DNA sensor in macrophages and keratinocytes | Ponomareva, L et al.,2013 |
| | Colon cancer | Inhibit | Suppresses colon tumorigenesis via DNA-PK and AKT | Wilson, J et al.,2015 |
| | Bladder cancer | Inhibit | Inhibits tumorigenesis and enhances the therapeutic effect | Zhou, H et al.,2022 |
| | Colorectal cancer | Inhibit | Inhibits cell proliferation and migration through suppressing Gli1 | Xu, M et al.,2020 |

a non-inflammasome in a variety of ways. Various factors can regulate the activation of NLRP6. For example, mouse macrophages bind to lipoteichoic acid and stimulate the assembly of the NLRP6 inflammasome [349]; microbial signals of type I IFN and PPAR- γ agonists regulate NLRP6 transcription [350, 351]; microbial metabolites activate or inhibit the NLRP6 inflammasome [352].

NLRP6 gene-deficient mice got more colitis-associated colorectal tumors after AOM/DSS treatment [313]. Elinav’s experiment suggests that the main source of active IL-18 generation through NLRP6 may be intestinal epithelial cells [353], which explains that NLRP6 gene-deficient animals’ serum and colon tissue showed decreasing levels of IL-18 with time but not IL-1. NLRP6 also plays a critical regulatory role in the linked pathogenic changes before HCC. For example, NLRP6 inflammasome and effector protein IL-18 suppress the development of NAFLD/nonalcoholic steatohepatitis (NASH) and metabolic syndrome by regulating intestinal microbiota [354]. NLRP6 overexpression reduces steatosis, inflammation, and fibrosis

and lowers the production of CCL20 in alcoholic hepatitis (AH) animal models [355]. On the contrary, inhibiting the activation of NLRP6 inflammasome may improve liver steatosis in mice [356]. The contradictory conclusions regarding the role of NLRP6 in HCC may be due to the different stages of the disease and the differences in experimental models and methods. Further studies are needed to define the role of NLRP6 in the development and spread of HCC and comprehend the extent to which NLRP6 switches from inhibiting tumor growth to promoting malignancy. Besides, NLRP6 functions as a tumor suppressor factor in gastric cancer (GC) [357] while promoting small cell lung cancer (SCLC) metastasis [307]. Even though there is low or no expression of NLRP6 in other organs except for the colon, liver, and stomach, it’s also related to other organ diseases such as brain injury [358] and acute renal injury [359]. These suggest the potential role of NLRP6 in the carcinogenesis of these organs, and further studies are needed to expand the cognitive boundaries of NLRP6 research.

The role of AIM2 in inflammation-mediated carcinogenesis

As the best-characterized member of the AIM2-like receptors (ALRs), AIM2 contains the N-terminal PYD domain and one or two C-terminal HIN domains (hematopoietic, interferon-inducible, and nuclear localization). The multicomponent AIM2 inflammasome is formed when AIM2 recognizes dsDNA and recruits the adaptor protein ASC, which then activates caspase-1 [360]. AIM2 can be directly activated by transfection of dsDNA into the cytoplasm [361] and DNA derived from the gut microbiota or host-DNA released after intestinal injury [362]. While AIM2 activation is negatively regulated by several factors such as IFI16- β [363], IFN-inducible protein PYD-only protein 3 (POP3) [364], and the viral protein pUL83 released during human cytomegalovirus (HCMV) [365], herpes simplex virus-1 (HSV-1) tegument protein VP22 [366], hepatitis B e-antigen (HBeAg) [367], HCMV IE86 protein [368] and others. Besides, ubiquitinated TRIM11 promotes p62-dependent selective autophagy-induced AIM2 inflammasome degradation following DNA stimulation [369].

AIM2 has bidirectional roles in tumorigenesis in different types of cancer. In hepatic and colon cancer, AIM2 suppresses tumor growth [370], whereas in cutaneous squamous cell carcinoma, it promotes tumor growth [371]. AIM2 contains a site for microsatellite instability, which leads to gene mutations in CRC and inhibits its development [372, 373]. Additionally, it was demonstrated that AIM2 has a role in the pathophysiology of DNA damage caused by chemotherapy, suggesting that medications targeting AIM2 may offer therapeutic advantages during radiation therapy [362]. These indicate that suppressing the activity of AIM2 inflammasome could be investigated due to their role in carcinogenesis. During the past decade, several synthetic inhibitors of AIM2 such as suppressive oligodeoxynucleotides [364], pyrin-containing proteins, and antimicrobial cathelicidin peptides [374] have been discovered, and exploring their anti-tumor effect and underlying mechanisms are promising strategies for cancer prevention and treatment.

The role of inflammasome adaptor ASC in inflammation-mediated carcinogenesis

In human leukemia cells treated with chemotherapeutics drugs, ASC was first identified as a protein that aggregates (or “specks”), which was also known as PYCARD/Target of Methylation-induced Silencing-1 (TMS1). The ASC/TMS1 protein has a 22 kDa structure and comprises two domains: the C-terminal CARD domain and PYD domain [375]. Several immune and normal epithelial cells express ASC/TMS1, which localizes in the nucleus, redistributes in the cytoplasm during activation, and finally aggregates into specks [376].

ASC can be either increased in tumor cells and over-expressed in the myeloid compartment (mostly TAMs) within the tumor microenvironment, or downregulated in malignancies, primarily due to aberrant methylation. On one hand, it is a key adapter molecule of the inflammasome complex, which is responsible for mediating the release of inflammatory cytokines of IL-1 β and IL-18, which were known to have tumor-promoting effects [377–379]. Besides, ASC also exhibits pro-tumor effects through other indirect pathways such as chronic inflammation, macrophage recruitment, IL-17 pathway activation, angiogenesis, and others [378]. On the other hand, ASC was downregulated in several types of cancer, and the tumor-suppressive effect of ASC was supported by the discovery that methylation silences its expression and prevents tumor cells from passing through apoptosis [380]. Besides, the regulatory effect of ASC on thymic stromal lymphopoietin (TSLP) secretion by cancer-associated fibroblasts (CAFs) contributes to improving the overall survival of pancreatic cancer patients [381]. ASC delays UV-induced skin tumorigenesis [382]. ASC inhibits lung cancer suppression via Bcl-2 and pSrc [383]. Restoring ASC expression makes colorectal cancer cells more susceptible to caspase-independent cell death caused by genotoxic stress [380]. In addition, ASC showed dual roles even in the same type of tumor. For example, ASC inhibits tumorigenesis in primary melanoma, while ASC promotes tumorigenesis in metastatic melanoma [314]; mice had fewer tumors when ASC was specifically deficient in myeloid cells, while mice had more tumors when ASC was specifically deficient in keratinocyte cells [384]. These opposing functions of ASC may be due to the tissue or cell-specific expression context, and dissecting the role of ASC in different cancer types and additional research on ASC and its upstream and downstream mediators may improve our comprehensive of molecular processes behind carcinogenesis and contribute to the ASC targeting strategies for cancer treatment and prevention.

Targeting components of the extracellular matrix for cancer chemoprevention

As a highly dynamic structure, the extracellular matrix (ECM) is present in all tissues and constantly undergoing controlled remodeling. ECM comprises large, insoluble proteins mainly formed of separate structural domains with highly conserved sequences and arrangements. These domains are glycosylated and often have sulfated glycosaminoglycan chains, resulting in negative charges [385]. This characteristic of ECM molecules gives them great potential to interact with other charged molecules, like growth factors and chemokines, to affect the accessibility or local concentration of these factors [386]. Since

collagen, the most abundant ECM component, was first identified and characterized in the 1930s.

In chronically inflamed tissues, the ECM fragments or ECM molecules are upregulated, and they're modulated by proteases, especially MMPs, and inflammatory cytokines such as TNF, IFN γ , and TGF β generated by extravasating cells or activated tissue-resident cells. Recent research has found that immune cells can be stimulated by ECM components or fragments that are increased during inflammation, thus sustaining the inflammatory response. On the one hand, ECM can serve a structural role as a barrier or scaffold for cells that invade inflamed tissues. On the other hand, the biophysical characteristics and their biochemical makeup provide immune cells with specific signals that regulate proliferation, migration, apoptosis, adhesion, differentiation, and survival.

Aberrant ECM influences immune cell behavior, such as infiltration and activation, and plays a role in cancer metastasis. For example, activating the collagen receptor DDR1 can enhance macrophage infiltration in atherosclerotic plaques [387]. The size and density of collagen fibrils can influence the migration of immune cells [388]. Although immune cell infiltration is promoted, collagen type I hinders macrophages from effectively killing cancer cells by blocking their polarization and subsequent activation [389]. This highlights the complex characteristics of how ECM deregulation modulates the immune response. Furthermore, breast cancer cells with high levels of the hyaluronan receptor CD44 have a better survival rate compared to those with low CD44 levels [390]. This indicates that hyaluronan and possibly other components of the extracellular matrix support the survival of metastatic cancer cells.

Tenascin-C (TNC) is a glycoprotein that plays a significant role in the ECM and is notably expressed in pathological conditions, particularly in cancer and chronic inflammatory diseases. TNC activates TLR4, which in turn stimulates macrophages and fibroblasts to release pro-inflammatory cytokines such as IL-6, IL-8, and TNF- α [391], and affects the recruitment of immune cells [392]. Higher TNC expression was observed in melanoma cells from advanced tumors and metastases, while early melanoma tumors and normal melanocytes displayed weak or absent TNC expression [393]. Tenascin-C is located in the bone endosteum and contributes to the development of prostate bone metastases [394]. TNC deficiency in human breast cancer cells significantly decreases metastasis to the lungs and bones in xenograft models [395].

Dysregulated ECM remodeling leads to many diseases, including cancer. As one of the major components of cancer, the ECM plays various crucial roles in signaling

regulation, microenvironment modulation, and mechanical support. The ECM shows multifaceted effects in regulating multiple hallmarks, including angiogenesis, tumor progression, immune response, and cancer cell migration of cancer. Under specific circumstances, the ECM restrains malignant tumor progression [396]. However, the ECM also promotes tumor progression [389]. Collagen IV over-expression boosts cell survival of lung cancer [397]. The ECM also facilitates the metastasis of cancer cells [398]. The miR-29 over-expression inhibits metastasis by regulating gene expression in ECM remodeling in breast cancer [399]. This suggests that blocking the ECM molecules may be therapeutically beneficial for cancer prevention and treatment. The ECM components including tenascin C, endostatin, tumstatin, canstatin, arresten, and hexastatin also have pro- and anti-angiogenic functions. For example, tenascin C promotes cancer cell proliferation and induces angiogenesis [400]. Arresten suppresses the angiogenesis of HUVECs by blocking PI3K/Akt signaling pathway [401]. The ECM plays a vital role in regulating immune responses. For example, the ECM niche in the spleen promotes the differentiation and survival of marginal zone B cells to support antibody production [402]. Besides, matrikines regulate immune cell behaviors such as the interaction of immune cells with cancer cells [403], the recruitment and activation of immune cells [404], and more. Here, we will summarize the role of the ECM components including ECM proteins, ECM fragments, the proteases that cleave the ECM, the MMPs that degrade the ECM, and others, the underlying mechanisms are also illustrated. A deeper comprehension of the biological activities of the ECM will provide intriguing opportunities for therapeutic intervention of cancer. More details regarding the function of ECM proteins and fragments in cancer are provided in Table 8.

The chemo-preventive effects of FDA-approved anti-inflammatory drugs in cancer chemoprevention

Long-term users of aspirin or other NSAIDs had a decreased incidence of cancer, which offers encouraging opportunities for cancer chemoprevention. Aspirin can reduce the risk of tumor occurrence [405], prevent colorectal cancer [406], inhibit metastasis of melanoma and breast cancer [407], and more. Besides, steroidal anti-inflammatory drugs such as dexamethasone, suppress tumor growth of non-small cell lung cancer [408], enhance gynecologic cancer chemotherapy [409], and more. These FDA-approved anti-inflammation drugs have well-established safety profiles, making them potentially valuable tools for cancer chemoprevention. Clarifying the anti-tumor effects and underlying mechanisms of the

anti-inflammation drugs will provide novel options and targets for cancer prevention and treatment. Here, we will summarize the effects and underlying mechanisms of FDA-approved anti-inflammatory drugs, and the advantages and disadvantages of these drugs' application in cancer chemoprevention will be also be discussed.

The anti-tumor effect of non-steroidal anti-inflammatory drugs

The chemo-preventive effect of NSAIDs in cancer was proposed in 2002 [410]. Epidemiological studies have shown a consistent 40–50% reduction in the risk of developing CRC by taking NSAIDs [411]. The epidemiological evidence supporting the efficacy of aspirin for the prevention of CRC is substantial [412]. As a classic NSAID, the role of aspirin in cancer chemoprevention has been well-studied. For example, aspirin reduces the risk of breast cancer [413] and pancreatic cancer [414], reduces mortality in endometrial cancer patients [18] and colon cancer [415], inhibits tumor progression of HCC [22], and pancreatic cancer [416] and others. Except for aspirin, other NSAIDs like indomethacin, acetaminophen, ibuprofen, meloxicam, and others also exhibit anti-tumor effects in various cancers. For example, indomethacin inhibits cell proliferation and invasion in colorectal cancer [417] and breast cancer [418], induces apoptosis of colon cancer [419], and suppresses angiogenesis of colon cancer [420] and others. A comprehensive list of the anti-tumor properties of NSAIDs is shown in Table 9.

The anti-tumor effect of steroidal anti-inflammatory drugs

Unlike NSAIDs, the anti-tumor effect of steroidal anti-inflammatory drugs is controversial. As a commonly used class of steroid anti-inflammatory drugs, on the one side, glucocorticoids (GCs) such as dexamethasone as an adjunctive treatment strategy for lymphoma in the clinic, they can reduce tumor size and improve patient's tolerance to chemotherapy [421]. About other solid tumors, GCs reduce treatment-related toxicity after surgery or radiotherapy in brain tumors [422] and reduce the risk of lung cancer [423]. On the other side, GCs favor pancreatic cancer progression [424], promote metastasis of breast cancer [425], induce resistance in prostate cancer [426], and others. A comprehensive list of the anti-tumor properties of steroidal anti-inflammatory drugs is shown in Table 10.

The outcome of clinical application of anti-inflammation drugs in cancer chemoprevention

Although cytokines are important regulators of both innate and adaptive anticancer immunity, their extreme toxicity, pleiotropy, and poor drug-like qualities make

them challenging to utilize as treatments [427, 428]. Developments in immune checkpoint blocking, protein engineering, and receptor biology and biochemistry drive rekindled interest in developing cytokine therapies [429, 430]. Clinical prospects have emerged from engineering strategies targeted at disentangling cytokines' pro- and anti-tumorigenic characteristics, which have shown encouraging preclinical outcomes [431]. Cytokines are the first effective cancer immunotherapy to provide humans with long-lasting anti-tumor immunity. Recombinant interferon (IFN α 2b) and IL-2 are approved cytokine therapeutics for cancer clinical treatment [432].

Other cytokines such as IL-4, IL-6, IL-7, IL-9, IL-10, IL-12, IL-15, IL-18, IL-21, IL-2R, IL-4R, IL-5R, IFN α , IFN β , IFN γ , TNF α and more have also been applied in clinical trials. For example, in a phase I/II study (NCT00000769), IL-4 at the dose of 1.5 microg/kg/day is poorly tolerated in patients with advanced AIDS-related Kaposi's sarcoma (KS) and didn't show an effective role in KS [433]. The anti-tumor effect of IL-4 in patients is ambiguous, and more clinical trials are recruiting or undergoing in other cancers such as recurrent malignant astrocytoma (NCT00003842) and ER+ breast cancer (NCT05967884). In phase II clinical trial (NCT00433446), a monoclonal antibody against IL-6 (CNT0328, also called siltuximab) was applied to pre-treated patients with castration-resistant prostate cancer, CNT0328 resulted in a prostate-specific antigen (PSA) response rate of 3.8% and a response evaluation criteria in solid tumors (RECIST) stable disease rate of 23% [434]. In a phase 2 randomized study (NCT00911859), CNT0328 at 8.3 and 11 mg/kg doses were well tolerated, while CNT0328 didn't significantly improve the complete response (CR) and long-term outcomes in patients of multiple myeloma [435].

Besides, several clinical trials have examined the combination of anti-inflammatory agents and ICIs. For instance, a phase 2 trial study (NCT03631407) investigated the combination of vicriviroc (a CCL5 antagonist) administered at doses of 150 or 250 mg alongside pembrolizumab at 200 mg. This combination demonstrated limited antitumor activity in patients with advanced or metastatic microsatellite stable (MSS) or mismatch repair-deficient (dMMR) CRC. However, the toxicity associated with this combination treatment was manageable [436]. In another phase 2 trial study (NCT03396952), the efficacy of high-dose aspirin in combination with ICIs is similar to that of ICI alone [437]. In a phase Ib/II trial study (NCT02807844), the combination of lacnotuzumab (CSF-1 inhibitor) and spartalizumab (PD-1 inhibitor) did not meet the gating criteria for efficacy [438]. Numerous clinical trials are currently recruiting participants or ongoing. For

Table 8 The role and mechanisms of extracellular matrix (ECM) and related proteins in cancer

| ECM proteins | Cancer types | Promote/inhibit | Mechanisms | References |
|---------------|--|-----------------|--|---------------------------|
| Collagen | Non-small cell lung cancer | Promote | Regulates overall survival and cell differentiation | Fang, S et al., 2019 |
| | Lung cancer | Promote | Collagen XXIII is highly expressed in lung cancer | Spivey, K et al., 2010 |
| | Esophageal squamous cell carcinoma | Promote | Regulates overall survival and cell differentiation | Fang, S et al., 2019 |
| | | | Drives invasion through a YAP-centered transduction loop | Khalil, A et al., 2024 |
| | Breast cancer | Promote | Promotes tumor progression and metastasis | Xiong, G et al., 2014 |
| | Hepatocellular carcinoma | Promote | Promotes cell proliferation, adhesion, invasion, metastasis and angiogenesis | Zhang, J et al., 2022 |
| | Lung cancer | Promote | Promotes metastasis, invasion and anoikis resistance | Zhang, H et al., 2018; |
| | | | Promotes cell growth in a three- dimensional culture system | Li, J et al., 2014 |
| | Pancreatic cancer | Promote | Stimulates proliferation, migration, and inhibits apoptosis via autocrine loop | Ohlund, D et al., 2013; |
| | | | Promotes the malignant phenotype of pancreatic cancer cells | Armstrong, T et al., 2004 |
| | | | Promotes metastasis by activating c-Jun NH2-terminal kinase 1 and up-regulating N-cadherin expression | Shintani, Y et al., 2006 |
| | Prostate cancer | Promote | Lactate supports ECM production to sustain metastatic behavior | Ippolito, L et al., 2024 |
| | | | Correlates with recurrence and distant metastasis | Banyard, J et al., 2007 |
| | Head and neck squamous cell carcinoma | Promote | Promotes cancer cell invasion | Chen, Yin-Q et al., 2019 |
| | Ovarian cancer | Promote | Influences proliferation | Sarwar, M et al., 2022 |
| | | | Increases proliferation | Fogg, K et al., 2020 |
| | Liver cancer | Promote | Promotes inflammatory response and proliferation | Shen, X et al., 2024 |
| | Gastric cancer | Promote | Promotes fibroblast collagen synthesis | NAITO, Y et al., 1984 |
| | Squamous cell carcinoma, prostate cancer, colorectal cancer, lung cancer | Promote | Collagen remodeling along cancer progression and provides a novel opportunity for cancer diagnosis and treatment | Song, K et al., 2022 |
| | Non-small cell lung cancer | Inhibit | Inhibits CAF-mediated collagen remodeling, cell migration and invasion | Zeltz, C et al., 2022 |
| Proteoglycans | Colorectal cancer | Inhibit | Inhibits cell differentiation and promotes a stem cell-like phenotype | Kirkland, S et al., 2009 |
| | Breast cancer | Inhibit | Regulates adhesion and migration after the collagen is mineralized | Choi, S et al., 2019 |
| | Brain cancer | Promote | Modulates migration, cell adhesion, tumor invasion, and neurite outgrowth | Yan, Z et al., 2020 |
| | | | Promotes receptor tyrosine kinase signaling and progression | Wade, A et al., 2013 |
| | Liver cancer | Promote | Interacts with many growth factors and relates to tumor invasion | Baghy, K et al., 2016 |
| | Breast cancer | Promote | Restores FGF-2 mitogenic activity | Delehedde, M et al., 1996 |
| | Esophageal squamous cell carcinoma | Promote | Regulates cell survival, invasion and metastasis | Li J et al., 2019 |
| | Pancreatic cancer | Promote | Stimulates cell growth | Zvibel, I et al., 2001 |

Table 8 (continued)

| ECM proteins | Cancer types | Promote/inhibit | Mechanisms | References |
|---------------------------------|--|-----------------|---|----------------------------|
| testican-1 | Colorectal cancer | Promote | Promotes cell proliferation and invasion through PI3K/AKT pathway | Zhao P et.al, 2016 |
| testican-1 | Gastric cancer | Promote | Induces EMT | Kim HP et.al, 2014 |
| lumican | Pancreatic cancer | Inhibit | High expression correlates with favorable survival after surgery | Li X et.al, 2014 |
| | Breast, hepatocellular, lung and prostate cancer , melanoma and multiple myeloma | Inhibit | Inhibits heparanase and growth factor-growth factor receptor active complex formation, sequesters growth factors in ECM, inhibits cell proliferation, metastasis and angiogenesis | Koo, C et al., 2008 |
| syndecan-1, endostatin, decorin | Esophageal squamous cell carcinoma | Inhibit | Regulates cell survival, invasion,metastasis and angiogenesis | Ji C et.al, 2015 |
| Glycoproteins | Colorectal cancer | Promote | Decreases the sensitivity of doxorubicin | Hotta, T et al., 1999 |
| | Breast cancer | Promote | Relates with doxorubicin and vincristine resistance | SANFILIPPO, O et al., 1991 |
| | NSCLC | Promote | Associates with both performance status and lymph node metastasis | Yildirim, A et al., 2007 |
| | Bladder cancer | Promote | Promotes cell proliferation, migration, and invasion and relates to the poor differentiation and recurrence | Zhang, Y et al., 2017 |
| | Colorectal cancer | Promote | Associates with clinic pathologic features and shorter overall survival | Wei, F et al., 2019 |
| | Breast cancer | Inhibit | Modulates multi-drug resistance via inhibiting P-glycoprotein efflux | Tripathi, A et al., 2016 |
| | Breast cancer | Inhibit | Suppresses cell proliferation and migration | Pan, P et al., 2012 |
| | Erythroleukemia, breast and epidermoid cancer | Inhibit | Inhibits p-glycoprotein, reverses drug resistance and restores intracellular drug accumulation | Pan Q, et al., 2005 |
| | Ovarian cancer | Inhibit | Inhibits the growth of ovarian cancer cell by inducing apoptosis | Liu M et al., 2007 |
| Endostatin | | | Inhibits angiogenesis by regulating bcl-2/bax and induces apoptosis | Liu M et al., 2006 |
| | Non-small cell lung cancer | Inhibit | Inhibits cell proliferation by inducing HMGB1 | Meng F et al., 2019 |
| | Lung cancer | | Inhibits tumor growth when combined with PD-1 inhibitor | Fu S et al., 2023 |
| | Colon cancer | Inhibit | Inhibits growth and metastasis when combined with SU6668 and 5-FU | Du Z et al., 2003 |
| | | | Inhibits the progress of chemically induced colon tumor | Li W et al., 2005 |
| | Bladder cancer | Inhibit | Inhibits tumor growth by regulating MMP, VEGF and inducing apoptosis | Du Z et al., 2003 |
| | | | Inhibits angiogenesis | Wu T et al., 2020 |
| | Liver cancer | Inhibit | Inhibits tumor growth when combined with IL12 | Wang X et al., 2005 |
| | Breast cancer | Inhibit | Inhibits tumor growth by inhibiting angiogenesis and increasing apoptosis | Liby K et al., 2003 |
| | Lung cancer | Inhibit | Inhibits growth and induces apoptosis by down-regulating Bcl-2 expression | Cheng X et al., 2008 |
| | | | Inhibits proliferation and angiogenesis, and suppresses tumor growth | Luo X et al., 2014 |
| | Oral squamous cell carcinoma | Inhibit | Delays tumor growth and lymphatic metastasis | Chung I et al., 2008 |
| Tumstatin | Breast cancer | Inhibit | Inhibits cell growth of breast cancer | Zhong Q et al., 2011 |
| | Ovarian cancer | Inhibit | Promotes apoptosis | Wang M et al., 2011 |

Table 8 (continued)

| ECM proteins | Cancer types | Promote/inhibit | Mechanisms | References |
|--------------|--|-----------------|--|-------------------------|
| Canstatin | Laryngocarcinoma | Inhibit | Inhibits tumor growth by reducing angiogenesis and angiogenic factors | Zhang G et al., 2008 |
| | Bladder cancer | Inhibit | Induces apoptosis through mitochondrial apoptosis pathway | Wang L et al., 2016 |
| | Liver cancer | Inhibit | Inhibits cell proliferation and induces apoptosis | Li Z et al., 2011 |
| | Cervical cancer | Inhibit | Inhibits cell proliferation and induces apoptosis through PTEN/PI3K/Akt signaling when combined with Ginkgo biloba extract Rg3 | Yi T et al., 2020 |
| | Prostate cancer | Inhibit | Inhibits tumor growth by inhibiting angiogenesis and angiogenic factors | Zhang G et al., 2007 |
| | Hypervascular hepatocellular carcinoma | Inhibit | Inhibits cell proliferation and tumor growth through apoptosis and anti-angiogenesis effects | Zhang X et al., 2011 |
| | Glioma | Inhibit | Inhibits vascular endothelial cells | Li C et al., 2017 |
| | Colon cancer, Lewis lung cancer | Inhibit | Inhibits proliferation and migration by down-regulating stem cell maintenance factors | Yu W et al., 2021 |
| | Non-small cell lung cancer | Inhibit | Inhibits tumor growth by inhibiting angiogenesis and enhancing apoptosis when being combined with gemcitabine | Yao B et al., 2005 |
| | Gastric cancer | Inhibit | Enhances the sensitivity of cells to cisplatin; promotes apoptosis and inhibits proliferation by inactivating Akt and ERK pathways | Wang W et al., 2010 |
| | Hepatocellular carcinoma | Inhibit | Inhibits proliferation, induces apoptosis and inhibits tumor growth | Li Y et al., 2009 |
| | Non-small cell lung cancer | Inhibit | Inhibits proliferation and metastasis by inducing apoptosis through anoikis and PTEN/Akt pathway | He Y et al., 2010 |
| | Melanoma | Inhibit | Inhibits tumor growth by inhibiting angiogenesis | Goto T et al., 2008 |
| | Colon cancer | Inhibit | Induces apoptosis, inhibits proliferation, enhances the sensitivity of cells to cisplatin by inactivating AKT pathway | Wang W et al., 2015 |
| | Lewis lung cancer | Inhibit | Inhibits tumor progression by triggering intracellular transduction pathway | Sylvie B et al., 2004 |
| | Gastric cancer | Inhibit | Delays tumor growth without obvious adverse reactions | Wang L et al., 2009 |
| | Pancreatic cancer | Inhibit | Inhibits metastasis, angiogenesis and tumor growth by mediating the expression of somatostatin | Lu W et al., 2011 |
| | Glioma | Inhibit | Induces apoptosis through the mitochondrial apoptotic pathway | Xing Y et al., 2019 |
| | Liver cancer | Inhibit | Delays tumor growth by inhibiting angiogenesis | He X et al., 2006 |
| Canstatin | Glioma | Inhibit | Inhibits tumor growth by inhibiting the formation of VM-like structure | Ma Y et al., 2021 |
| | Liver cancer | Inhibit | Inhibits tumor growth and angiogenesis by reducing the expression of Flk-1 | Qi M et al., 2009 |
| | Primary oral squamous cell carcinoma | Inhibit | Inhibits tumor growth | Hwang-Bo J et al., 2010 |
| Canstatin | Esophageal cancer | Inhibit | Inhibits angiogenesis and tumor growth by down-regulating Flk-1 | Zheng X et al., 2009 |

Table 8 (continued)

| ECM proteins | Cancer types | Promote/inhibit | Mechanisms | References |
|--------------|-----------------------------------|-----------------|--|-----------------------------|
| Arresten | Hepatocellular carcinoma | Inhibit | Inhibits proliferation, migration and adhesion, promotes apoptosis and inhibits tumor growth when combined with arsenic trioxide | Zhang F et al., 2020 |
| | Pancreatic cancer | Inhibit | Decreases microvessel density and increases apoptosis; has synergistic effects of oncolytic therapy and antiangiogenic therapy in a CRAd-Cans form | He X et al., 2009 |
| | Lewis lung cancer | Inhibit | Inhibits tumor growth and metastasis | Lu W et al., 2006 |
| | Ovarian cancer | Inhibit | Delays tumor growth by inhibiting angiogenesis | Zhu B et al., 2009 |
| | Colon cancer | Inhibit | Inhibits metastasis by suppressing angiogenesis | Long M et al., 2008 |
| | Gastric adenocarcinoma | Inhibit | Inhibits the proliferation of tumor vascular endothelial cells | Lu C et al., 2005 |
| | Lung cancer | Inhibit | Treats cancer in a CRAd-arresten-TRAIL form | Li S et al., 2015 |
| Hexastatin | Squamous cell carcinoma | Inhibit | Inhibits invasion by suppressing collagen-derived angiogenesis | Mari A et al., 2012 |
| | Melanoma and lung cancer | Inhibit | Inhibits tumor growth and suppresses cell proliferation of melanoma | Wen L et al., 2009 |
| Versican | Lymph node negative breast cancer | Promote | Promotes local invasion and metastasis by tumor remodeling of extracellular matrix through increasing versican deposition | Ricciardelli C et al., 2002 |
| | Breast cancer | Promote | Promotes tumor progression and metastasis | dos Reis D et al., 2019 |
| | | | Enhances the self-renewal of breast cancer through EGFR/AKT/GSK-3 β (S9P) signaling, and endows it with resistance to chemotherapy drugs | Du W et al., 2013 |
| | | | Enhances bone metastasis by promoting migration, invasion and survival of cells | Du W et al., 2023 |
| | | | Promotes invasion, enhances cell viability, proliferation, migration and local tumor growth, enhances vascular endothelial proliferation, migration and angiogenesis | Yee A et al., 2007 |
| | Cervical cancer | Promote | Promotes invasion and metastasis through EGFR signaling | Du W et al., 2010 |
| | | | Enhances local invasion and decreases CD8 positive T cells | Gorter A et al., 2010 |
| | Ovarian cancer | Promote | Promotes tumor growth | Voutilainen K et al., 2003 |
| | | | Increases metastasis by obtaining matrix around HA/versican cells | Miranda P et al., 2011 |
| | | | Changes tumor microenvironment and promotes tumor cancer invasion | Yeung, T et al., 2011 |
| | | | Promotes the metastasis and spread of clinical prostate cancer | Ricciardelli C et al., 2007 |
| | Hepatocellular carcinoma | Promote | Promotes tumor development by regulating miRNA activity | Fang L et al., 2013 |
| | Gastric cancer | Promote | Promotes proliferation and metastasis by activating EGFR-PI3K-AKT pathway | Zhang Y et al., 2020 |
| | | | Promotes the progress of gastric cancer caused by IL-11 | Zhang Z et al., 2012 |

Table 8 (continued)

| ECM proteins | Cancer types | Promote/inhibit | Mechanisms | References |
|--------------|-------------------------------|-----------------|--|--------------------------------|
| Tenascin | Skin cancer Melanoma | Promote | Promotes proliferation, migration and invasion by overexpressing VCAN | Zhai L et al., 2012 |
| | | Promote | Promotes tumor development | Kunisada M et al., 2012 |
| | | Promote | Promotes proliferation and migration, inhibits adhesion to type I collagen, laminin and fibronectin | Hernandez D et al., 2011 |
| | Prostate cancer | Inhibit | Reduces tumorigenicity | Miquel-Serra L et al., 2005 |
| | | Promote | Is up-regulated in cancer tissue and relates to glucose uptake, lactic acid production and glycolytic enzyme expression | Qian Y et al., 2022 |
| | Breast cancer | Promote | Promotes tumor progression, enhances adhesion and colony formation through integrin $\alpha 9 \beta 1$ | Martin R et al., 2017 |
| | | | Supports the tumor initiation ability through transfer niche | Hassan F et al., 2016 |
| | | | Up-regulates the expression of growth-related genes, increases cell migration, mitosis and growth factor-dependent endothelial cell germination and elongation | Jones P et al., 2000 |
| | | | Promotes tumor progress by immobilizing infiltrating T lymphocytes through CXCL12 | Jones F et al., 2000 |
| | | | Contributes to the invasion behavior of tumor cells | Scherberich A et al., 2005 |
| | | | Promotes therapy-resistant metastasis | Insua-Rodriguez J et al., 2018 |
| | | | Increases invasion by promoting Tenascin C inclusion in extracellular vesicles | Campos A et al., 2023 |
| | | | Mediates lung metastasis | Taraseviciute A et al., 2006 |
| | | | Promotes invasion and proliferation | Alcock R et al., 2005 |
| | | | Promotes cell proliferation | Swierczynski S et al., 2011 |
| | Colon cancer | Promote | Promotes invasion and proliferation | Alcock R et al., 2005 |
| | | | Promotes tumorigenesis by activating matrix fibroblasts based on 1- integrin activation | Fujita M et al., 2019 |
| | | | Promotes invasion through EMT regulation and acts as a specific new indicator | Takahashi Y et al., 2013 |
| | | | Promotes EMT-like changes and proliferation and leads to poor prognosis | Yang Z et al., 2018 |
| | | | Drives tumor progression and participates in CSC characteristics through HH signaling | Yang Z et al., 2020 |
| | Gastric cancer | Promote | Inhibits the angiogenesis simulation by suppressing ERK-triggered EMT | Xing K et al., 2021 |
| | Oral squamous cell carcinoma | Promote | Promotes metastasis by enhancing the immunosuppressive lymphatic matrix through CCL21/CCR7 signaling | Caroline S et al., 2020 |
| | Triple negative breast cancer | Promote | Promotes the resistance to T cell-mediated cytotoxicity by blocking the degradation of Tenin C | Li Z et al., 2020 |
| | Endometrium cancer | Promote | Promotes tumor progress by immobilizing infiltrating T lymphocytes through CXCL12 | Murdamoothoo D et al., 1996 |
| | Ovarian cancer | Promote | Promotes patients' survival and supports spheroids formation and tumor progression | Roders A et al., 2024 |

Table 8 (continued)

| ECM proteins | Cancer types | Promote/inhibit | Mechanisms | References |
|--------------|--|-----------------|---|--|
| | Glioblastoma | Promote | Promotes invasion of glioblastoma Induces excessive proliferation of glioblastoma cells | Hirata E et al., 2009 Fujita M et al., 2019 |
| | Laryngocarcinoma | Promote | Promotes proliferation and migration in an autocrine way | Toshimichi Y et al., 1999 |
| | Osteosarcoma | Promote | Promotes distant metastasis of osteosarcoma | Tanaka M et al., 2000 |
| | Pancreatic ductal adenocarcinoma | Promote | Hedgehog signaling stimulates Tenascin C to promote invasion through Annexin A2 | Foley K et al., 2017 |
| | Squamous cell carcinoma of head and neck | Promote | Migration of cancer cells is dependent on tenascin-C expression | Thomas C et al., 2016 |
| | Esophageal squamous cell carcinoma | Promote | Enhances the dry-like characteristics of cancer and promotes EMT-like changes through Akt/HIF1α axis | Yang Z et al., 2019 |
| | Neuroendocrine tumor | Promote | Down-regulation of DKK1 is an important mechanism for TNC to enhance tumor progression by providing tumor microenvironment that promotes angiogenesis | Falk S et al., 2014 |
| | Pancreatic cancer | Promote | Suppresses apoptosis through activation of ERK/NF-κB pathway | Shi M et al., 2015 |
| | | | Promotes tumor progress | Chen J et al., 2009 |
| | | | Promotes the diffusion and metastasis by affecting the proliferation, migration and adhesion | Berchtold S et al., 2011 |
| | Bladder cancer | Promote | Mediates malignant behavior through syndecan-4 and NF-κB signaling | Guan Z et al., 2022 |
| Matrikines | Melanoma | Promote | Up-regulates migration and promotes chemotaxis, mitosis and metastasis | Tran K et al., 2005 |

instance, siltuximab, an IL-6 inhibitor, is being tested to prevent severe irAEs during the rechallenge of ICIs in patients with advanced cancer (NCT06470971). Additionally, drugs used in chemokine modulation therapy, such as celecoxib, recombinant interferon alfa-2b, and rintatolimod, have been combined with pembrolizumab in the treatment of metastatic triple-negative breast cancer (NCT03599453). Further investigation is needed to fully evaluate the clinical utility and identify the patient subgroups that are most likely to benefit from the combination of anti-inflammatory agents with ICIs. More information about the clinical trials of these cytokines or their antibody or their derivatives is summarized in Table 11.

The application of artificial intelligence (AI) in inflammation-related cancer for cancer chemoprevention

In recent years, AI has gained significant popularity. With the advancement of deep learning and the availability of extensive imaging databases, AI-based computer-aided diagnostic (CAD) systems—which incorporate machine

learning (ML), deep learning (DL), and artificial neural networks (ANN)—are increasingly being used to standardize and improve the evaluation of medical imaging.

AI models have been used to identify inflammatory and immune cells in cancer tissues. For instance, deep learning techniques have been employed to detect tumor-infiltrating lymphocytes (TILs), which could serve as a potential prognostic marker for testicular germ cell tumors [439]. Deep convolutional neural networks based on supervised learning have been used to quantify the biomarkers of immune cells in the lung cancer microenvironment [440]. TILs densities and spatial structures can be analyzed using deep learning on pathology images, providing insights into the tumor-immune microenvironment [441]. Deep learning supervised by antibodies was utilized to quantify tumor-infiltrating immune cells, an emerging prognostic biomarker in breast cancer samples [442]. AI-based pathology served as a biomarker for progression-free survival in patients treated with atezolizumab and bevacizumab for hepatocellular carcinoma [443]. Identifying PLA2G1B, a gene crucial for lipid metabolism and inflammation, suggests it may serve as a

Table 9 Mechanisms underlying anti-tumor properties of non-steroidal anti-inflammatory drugs

| Drugs | Cancer types | Mechanisms | References |
|-------------------------|--------------------|---|-----------------------------|
| Salicylates | Colon cancer | Inhibits AKT and ERK phosphorylation, decreases CYR61 expression and blocks TGF- β 1-induced migration | Han, S et al., 2016 |
| | | Induces apoptosis through TGF- β 1/Smad2 pathway | Du, C et al., 2020 |
| Aspirin | Prostate cancer | Decreases ERK1/2 activity and cyclin D1 expression | Gao, Q et al., 2006 |
| | Lung cancer | Blockades growth by suppressing PD-L1 through regulating the transcriptional coactivator of TAZ | Zhang, Y et al., 2020 |
| | Esophageal cancer | Induces apoptosis by inhibiting COX-2 enzymatic activity | Du, C et al., 2020 |
| | Pancreatic cancer | Decreases the expression of cyclin D1 and inactivates GSK-3/3 but not the p38 MAPK pathway | Nakabayashi, R et al., 2022 |
| | Bladder cancer | Identifies GSK3B, CDC20, TPX2, AURKA and CCNE1 as potential therapeutic targets | Li, X et al., 2022 |
| | Ovarian cancer | Inhibits cell viability by blocking phosphorylation of AKT and ERK activated by EGF | Cho, M et al., 2013 |
| | Cervical cancer | Induces apoptosis and inhibits proliferation via suppressing ErbB2 downstream cell survival signaling pathways | Sun, Z et al., 2023; |
| | | Induces apoptosis and inhibits proliferation | Wang, B et al., 2014 |
| Lysine Acetylsalicylate | Endometrial cancer | Improves survival outcomes of patients | Matsuo, K et al., 2016 |
| | Colon cancer | Inhibits cell proliferation | Wang, B et al., 2007 |
| Acetic Acid | Colon cancer | Down-regulates CDK2 and CDK4 and up-regulates p21/WAF1/PIC1 | Xu, M et al., 2005 |
| | | Regulates the focal complexes formation and attenuates EGF-mediated Ca ²⁺ influx | Guo, Y et al., 2016 |
| Indomethacin | Gastric cancer | Induces apoptosis through AKT/GSK3 β /NAG-1 pathway | Pang, R et al., 2011 |
| | Endometrial cancer | Reduces platelet aggregation and TxB2 level when combined with leukiniferon | Mistakopulo, N et al., 1992 |
| | Laryngeal cancer | Inhibits cell proliferation and viability and decreases LPS-induced cell iNOS activity | Zhang, L et al., 2008 |
| | Lung cancer | Inhibits tumor growth and promotes apoptosis when combined with cisplatin | Gao, J et al., 2013 |
| Diclofenac | Colon cancer | Inhibits PI3K/AKT axis through dephosphorylating PTEN, PDK, AKT | Arisan, E et al., 2018 |
| | | Promotes apoptosis | Arisan, E et al., 2018 |
| | Breast cancer | Enhances the anti-proliferative and apoptotic effects when combined with diclofenac, piperine, and D-limonene than each individually used | Sankar, S et al., 2023 |
| | Pancreatic cancer | Blocks cancer cell proliferation | Choi, S et al., 2022 |

Table 9 (continued)

| Drugs | Cancer types | Mechanisms | References |
|-------------------|---|---|---|
| Propionic Acid | | | |
| Ibuprofen | Colon, breast, cervical, gastric, lung cancer, and head and neck cancer | Reduces HDACs and histone demethylase (KDM6A/B) expression, and suppresses gene expression via a COX2-dependent way | Shen, W et al., 2020 |
| | Colon cancer | Suppresses nuclear translocation of β -catenin Induces apoptosis when combined with TRAIL Reverse the effect of ADMA in vitro and in vivo by blocking COX -2 and providing an arginine source | Ouyang, N et al., 2013 Todo, M et al., 2013 Ahmetaj-Shala, B et al., 2017 |
| | Non-small cell lung cancer. breast cancer | Inhibits cell proliferation and metastasis by decreasing survivin expression level and increasing E-cadherin expression level | Zhu, L et al., 2015 |
| | Prostate cancer | Induces apoptosis and oxidative stress via mediating pro-apoptotic signaling by regulating ceramide synthesis when combined with EGCG | Kim, M et al., 2007; |
| | Lung cancer | Induces apoptosis Enhances sensitivity to cisplatin by enhancing apoptosis at several stages of the mitochondrial cascade | Andrews, J et al., 2002 Endo, H et al., 2014 |
| | Liver cancer | Regulates β -catenin signaling and downstream target gene transcription | Ma, J et al., 2009 |
| Ketoprofen | Breast cancer | Induces apoptosis through intrinsic pathway and diminishes the phosphorylation of JAK2 and STAT Induces apoptosis and inhibits autophagy through the extrinsic apoptotic pathway and inhibition of the JAK/STAT signaling Inhibits the migration and invasion through PI3K/AKT/mTOR signaling | Noori, S et al., 2021 Patra, I et al., 2023 Nan, Z et al., 2017 |
| | Gastric cancer | Inhibits cell proliferation Enhances anti-proliferative effects on cells rich in progesterone receptors | Akrami, H et al., 2018 Saha, D et al., 2001 |
| | Colon cancer | Inhibits proliferation through prostaglandin H synthase and prostaglandin production Induces apoptosis through inhibition of PUM1 | Sánchez, T et al., 1999 Gor, R et al., 2023 |
| | Colon cancer and melanoma | Exhibits high cytostatic activity | Buzharevski, A et al., 2019 |
| | Leukemia and Ovarian Cancer | Diminishes cell viability through cAMP/PKA signaling through inhibiting PDE1 and CaM | Afshari, H et al., 2023 |
| | Colon and cervix cancer | Inhibits cell proliferation through transcription factor NF- κ B | Damjanovic, I et al., 2015 |
| Loxoprofen sodium | Lung cancer | Inhibits tumor growth, while didn't affect cell proliferation and viability, decreases intratumoral vessel density, suppresses both intratumoral and systemic VEGF protein and inhibits tubular formation by suppressing VEGF; decreases the plasma VEGF level in patients | Kanda, A et al., 2003 |

Table 9 (continued)

| Drugs | Cancer types | Mechanisms | References |
|------------------------|--------------------------------|--|------------------------------------|
| Naproxen | Prostate and Breast cancer | Shows high anticancer activity, but less toxic against healthy cells | Pedro-Hernandez, L et al., 2022 |
| | Bladder cancer | Inhibits the COX-2 enzyme | Han, M et al., 2022 |
| | | Induces apoptosis and inhibits AKT phosphorylation | Kim, M et al., 2014 |
| | Bladder, colon cancer | Prolongs tumor latency and inhibits tumor growth | Grubbs, C et al., 2009 |
| | Colon cancer | Shows chemo-preventive activity partly due to the inhibition of the transcriptional activation of TCF4 | Pedro-Hernandez, L et al., 2022 |
| | Liver cancer | Induces apoptosis and cell arrest in the G2/M phase | Lavasani, R et al., 2023 |
| Phospho-naproxen (P-N) | Colon cancer | Inhibits NF- κ B activity in a concentration-dependent manner | Patel, L et al., 2023 |
| Oxicam | | | |
| Meloxicam | Bladder cancer | Increases cytotoxicity of sunitinib malate through DNA damage | Arantes-Rodrigues, R et al., 2013 |
| | Gastric cancer | Inhibits proliferation through the down-regulation of COX-2 expression | Li, J et al., 2009 |
| | Liver cancer | Inhibits cell proliferation, reduces microvessel density and up-regulates the expression of Bax when combined with anti-angiogenic therapy | Jiang, X et al., 2010 |
| | Colon cancer | Inhibits proliferation and migration by up-regulating the expression of PTEN | Zhou, M et al., 2016 |
| | Prostate cancer | Prolongs tumor latency and inhibits tumor growth | Montejo, C et al., 2010 |
| | Lung and colon cancer | Increases the intracellular accumulation of doxorubicin and enhances doxorubicin-induced cytotoxicity by inhibiting MRP1 and MRP4 | Chen, S et al., 2016 |
| | Colon adenocarcinoma cancer | Meloxicam-loaded NPs shows cytotoxic effects on cells | Sengel-Turk, C et al., 2012 |
| | Liver cancer | Inhibits tumor progression and enhances the sensitivity of immuno-therapy via the microRNA-200/PD-L1 pathway | Montejo, C et al., 2010 |
| Piroxicam | Prostate cancer | Shows cytotoxic effects and induces apoptosis | Kisla, C et al., 2023 |
| | Breast cancer | Kills cells and induces apoptosis | Peng, X et al., 2012 |
| | Oral cancer | Selectively inhibits cell growth via regulating the S phase of the cell cycle | Ding, HM et al., 2003 |
| Fenamic Acid | | | |
| Mefenamic acid | Colon cancer and breast cancer | Shows anticancer activity against cancer cells | Castillo-Rodriguez, I et al., 2023 |
| | Prostate cancer | Prolongs tumor latency and inhibits tumor growth | Melnikov, V et al., 2021 |
| | Lung cancer | Overcomes drug resistance by controlling biological function of AKR1C | Shiiba, M et al., 2017 |
| | Liver cancer | Blocks PARP-1 cleavage activity and protects against MEF-induced apoptotic cell death | Woo, D et al., 2004 |

Table 9 (continued)

| Drugs | Cancer types | Mechanisms | References |
|----------------------------------|--------------------------------|---|--------------------------------|
| Aryl Acetic Acid Sulindac | Colon cancer | Increases the nuclear level of activated aryl hydrocarbon receptor and mediates CYP expression | Ciolino, H et al., 2008 |
| | | Prolongs tumor latency and inhibits tumor growth | Ohishi, T et.al., 2002 |
| | | Induces apoptosis | Shi, J et al., 2009 |
| | | Down-regulates Sp proteins by up-regulating the Sp-repressor ZBTB10 | Li, X et al., 2016 |
| | | Decrease Fas Ligand expression and induces apoptosis of lymphocyte | Wu, Z et al., 2005 |
| | | Negatively regulates the function of VDAC1 and VDAC2 | Aono, Y et al., 2018 |
| | Pharyngeal cancer | Induces apoptosis and inhibits the Wnt/ β -catenin signaling | Tai, W et al., 2014 |
| | Lung cancer | Increases VEGF-R2 and decreases ADAMTS1 levels | Agdas, F et al., 2021 |
| | | Augments apoptotic activity and intracellular ROS production when combined with simvastatin | Kim, Y et al., 2015 |
| | Liver, colon and breast cancer | Suppresses β -catenin signaling | Han, A et al., 2008 |
| Formic Acid Meclofenamic acid | Prostate cancer | Increases tumor latency and inhibits tumor growth | Kim, C et al., 2005 |
| | | Inhibits cell proliferation | Deng, X et al., 2015 |
| | | Suppresses proliferation, induces apoptosis and reduces angiogenesis | Wang, X et al., 2008 |
| Non-acid Nimesulide | Gastric cancer | Inhibits tumor aggression, increases fibrosis, reduces cell proliferation and tumor vascularity | Delgado-Enciso, I et al., 2015 |
| | | Attenuates insulin-like growth factor 1-induced Akt activation when combined with simvastatin | Sekine, Y et al., 2018 |
| | | Inhibits the activation of COX-2 and influences cell cycle | Xu, M et al., 2003 |
| | | Induces apoptosis | Zhang, Y et al., 2010 |
| | | Enhances gammadelta T cell-mediated killing effect | Liu, J et al., 2010 |
| | | Down-regulates the protein level of P-STAT3, CyclinD1 and Bcl-2 | Chen, M et al., 2008 |
| | | Inhibits cell growth and induces apoptosis by enhancing Bax-to-Bcl-2 ratio and decreases COX-2 mRNA level | Zhang, L et al., 2005 |
| | | Blocks the activation of protein kinase B | Hu, G et al., 2004 |

Table 9 (continued)

| Drugs | Cancer types | Mechanisms | References |
|-------------------|---|--|------------------------|
| Nabumetone | Liver cancer | Induces apoptosis by up-regulating c-myc expression, down-regulating Bcl-2 expression and activating caspase-3 | Song, J et al., 2002 |
| | | Exhibits lethal effect when combined with mDRA-6 | Liu, Y et al., 2008 |
| | Colon cancer | Blocks cell cycle progression and down-regulates the release of VEGF | Fei, S et al., 2006 |
| | | Inhibits COX-2 activity and PGE2 synthesis and blocks the progression of cell cycle | Chen, G et al., 2005 |
| | Breast cancer | Inhibits cell growth which overexpressing COX-2 protein, and up-regulates the expression of E-cadherin | Liu, W et al., 2004 |
| | | Reverses multi-drug resistance by down-regulating P-170 and GST-pi expression | Guo, Y et al., 2008 |
| | | Inhibits IFN-γ-induced PD-L1 surface expression | Liang, M et al., 2009 |
| | | Enhances chemotherapy sensitivity and stimulates apoptosis | Tian, B et al., 2010 |
| | Small cell lung cancer cell | Inhibits cell proliferation | Liu, Y et al., 2006 |
| | Endometrial cancer | Induces apoptosis by activating caspase-3 | Ozalp, SS et al., 2012 |
| | Ladder cancer | Inhibits the growth of T24 cell | Wang, Q et al., 2007 |
| | Pancreatic cancer | Enhances TRAIL-induced apoptosis by promoting clustering of DR5 | Vunnam, N et al., 2023 |
| | Breast and ovarian cancer | Induces apoptosis and inhibits cell growth by enhancing PTEN expression level | Chu, M et al., 2018 |
| | | Induces apoptosis | Jaragh, A et al., 2022 |
| | Head and neck cancer | Inhibits the expression of Ets-1 and Ets-2 | Lamm, W et al., 2005 |
| Esophageal cancer | Inhibits cell proliferation and VEGF expression when combined with 5-FU | Li, M et al., 2010 | |
| Colorectal cancer | Improves selectivity through HA/CD44 receptor interactions | Jian, Y et al., 2017 | |
| Celecoxib | Colon cancer | Induces glycogen synthase kinase-3β | Roy, H et al., 1999 |
| | Intestinal tumorigenesis | Induces apoptosis and down-regulates Bcl-2 | Roy, H et al., 2001 |
| Etoricoxib | | | |
| Celecoxib | Pancreatic cancer | Attenuates invasion and migration | Gu, Z et al., 2015 |
| | Breast cancer | Enhances the anti-tumor effect by down-regulating expressions of VEGF and MMP-2 when combined with p65miRNA | Wang, L et al., 2012 |
| | | Inhibits cell growth | Li, Y et al., 2008 |

Table 9 (continued)

| Drugs | Cancer types | Mechanisms | References |
|------------------|-------------------------------|--|---|
| | Gallbladder cancer | Inhibits cell growth by inducing apoptosis | Wang, Y et al., 2005 |
| | Non-small cell lung cancer | Triggers ER stress and induces apoptosis through both non-classical caspase-4 and GRP78 | Kim, B et al., 2017 |
| | Colon cancer | Suppresses Wnt/ β -catenin signaling but not COX-2 | Egashira, I et al., 2017 |
| | Cervical cancer | Induces the various phosphorylation sites of p53 and activates p53-PUMA pathway Down-regulates the expression of caveolin-1 and inhibits the activation of downstream signaling molecules when combined with Lovastatin | Liu, H et al., 2008 Chen, L et al., 2016 |
| Etoricoxib | Colon cancer | Modulates the Delta Psi(M) | Tanwar, L et al., 2010 |
| Parecoxib Sodium | Thyroid cancer | Improves functional neck lymph node dissection | Peng, Y et al., 2013 |
| | Colon cancer | Provides good postoperative analgesic effect | Xie, H et al., 2011 |
| Rofecoxib | Breast cancer | Inhibits EMT and metastasis by down-regulating β -catenin | Wang, C et al., 2022 |
| | | Inhibits proliferation and migration by up-regulating the expression of microRNA-199a/b-3p and blocks cell cycle when combined with adriamycin | Wang, J et al., 2018 |
| | Gastric cancer | Enhances anti-proliferation effect when combined with octreotide | Liu, C et al., 2004 |
| | Pancreatic cancer | Improves inhibitory effects when combined with octreotide | Zhou, X et al., 2003 |
| | | Inhibits not only ERK-1/ ERK-2 and c-Fos expressions but also AP-1 binding activity | Zhou, X et al., 2005 |
| | Triple-negative breast cancer | Changes gene expression which favors cell cycle arrest | Tseng, W et al., 2002 |
| | | Induces apoptosis by suppressing the expression of Bcl-2 family | Zhao, Y et al., 2016 |
| | NSCLC | Increases apoptosis | Alam, M et al., 2007 |
| | Gastric cancer | Sensitizes chemotherapeutic effect of various anticancer agents | Zhu, F et al., 2010 |
| | | Prolongs tumor latency and inhibits tumor growth | Zhu, F et al., 2007 |
| | Lung cancer | Inhibit tumor growth and prevents recurrences | Tanaka, T et al., 2005 |
| | Colorectal cancer | Negatively regulates angiogenesis | Fenwick, S et al., 2023 |
| | Cervical cancer | Prolongs tumor latency and inhibits tumor growth | Jung, Y et al., 2009 |

Table 9 (continued)

| Drugs | Cancer types | Mechanisms | References |
|-----------|--------------|--|----------------------|
| Imrecoxib | Lung cancer | Up-regulates PTEN and down-regulates cortactin | Wang, L et al., 2017 |
| | | Inhibits tumor growth and lymph node metastasis via down-regulating ezrin and up-regulating E-cadherin when combined with lobaphatin | Wang, D et al., 2013 |
| | Colon cancer | Induces apoptosis by regulating the expression of Survivin and Caspase-3, and enhances the anti-tumor effect of oxaliplatin | Wang, X et al., 2020 |

Table 10 Mechanisms underlying anti-tumor properties of steroidal anti-inflammatory drugs

| Drug | Cancer Types | Mechanisms | References |
|----------------|---------------------------------------|--|---------------------------|
| Hydrocortisone | Breast cancer | Reduces colony formation, inhibits migration, induces cell cycle arrest in the SubG1 phase, induces apoptosis via caspase-3, inhibits migration and down-regulates Bcl-2 | Msalbi, D et al., 2023 |
| | Colon cancer | Entrap docetaxel and complexed with anti-tumor plasmid DNA for enhanced killing of cancer cells | Sridharan, K et al., 2021 |
| | Melanomas, sarcomas, and colon tumors | Inhibit cell viability selectively in colon cancer cells | KERN, D et al., 1984 |
| Prednisone | Non-small cell lung cancer | Induces a shutdown of bypass RTK signaling and inhibits key resistance signals | Gong, K et al., 2021 |

preventive marker for lung cancer through bioinformatics and machine learning methods [444].

Artificial intelligence has recently emerged as a powerful and promising tool for developing anti-tumor medications more quickly, affordably, and effectively [445]. Due to the crucial role of TLR4 in pro-inflammatory responses, and considering the costly, time-consuming, and labor-intensive nature of traditional drug design approaches, novel TLR4 modulators identified through artificial intelligence and computer-assisted drug design show great promise and have demonstrated positive results [446]. Due to their high selectivity and low toxicity, anti-inflammatory peptides (AIPs) have shown greater therapeutic potential against inflammatory diseases compared to small compounds. Additionally, machine learning plays a crucial role in predicting peptides [447].

Artificial intelligence solutions for endoscopy-based cancer evaluation hold great promise for the future. However, the application of these models in clinical practice faces several challenges, including the need for more robust validation studies and overcoming regulatory hurdles. At the same time, AI facilitates data-driven decision-making, helping to expedite drug discovery and development while reducing failure rates. Additionally, AI-powered precision medicine allows doctors to tailor early therapies to the individual needs of each patient.

Open questions/Future perspective

It is now evident that inflammatory cells and related inflammatory pathways significantly influence tumor growth. However, several issues still need to be addressed. 1) In the absence of an external carcinogenic agent, can inflammation itself lead to neoplasia? While various studies have suggested that inflammation can cause cancer without the presence of exogenous carcinogens, evidence shows that more DNA mutations were observed in a mouse model of bowel inflammation lacking IL-10 when exposed to external carcinogens [448], and more powerful proof is required to make such a disclosure. 2) Due to the dual role of inflammatory molecules, such as inflammasomes and interleukins, in cancer development and progression, it is important to consider the question about whether we can suppress tumor growth by shifting the balance between “bad” inflammation and “good” inflammation? The adaptive, humoral, and innate immune systems all play integral roles in the complex relationship between local and systemic inflammation and tumor growth. Therefore, triggering an effective anti-tumor adaptive immune response is crucial and requires careful attention. Recently, the use of bromodomain and extra-terminal domain (BET) protein inhibitors (iBET) has been reported to induce cell death and reduce the aggressiveness of oral squamous cell carcinoma (OSCC) [449]. iBET protects mice from

Table 11 Summary of pharmacological strategies direct targeting inflammatory molecules for cancer therapy in clinical trials (information was obtained from <https://www.clinicaltrials.gov/>)

| Identifier | Phase | Drug | Cancer types | Treatment | Status | References |
|-------------|--------------------|-----------------|---|---|-------------------------|---------------------------|
| NCT01943422 | Phase 1 | IFN α 2b | Advanced melanoma | Combined BRAF inhibitor Vemurafenib and high-dose interferon α -2b | Terminated | N/A |
| NCT00913913 | Phase 2 | IFN α 2b | Metastatic renal cell carcinoma | Bevacizumab, autologous tumor/DC Vaccine, IL-2 and IFN α -2b | Completed | N/A |
| NCT00871533 | Early Phase 1 | IFN α 2b | Melanoma | IFN α 2b/ PEG- IFN α 2b | Terminated With Results | N/A |
| NCT05870475 | Phase 2 | IFN α 2b | Hydroxyurea-resistant/intolerant PV | Pegylated interferon α -2b in combination with Ruxolitinib | Recruiting | N/A |
| NCT02339324 | Phase 1 | IFN α 2b | Locally/regionally advanced/recurrent melanoma | Standard-dose interferon α -2b (HD) and anti-PD1 monoclonal antibody | Completed | N/A |
| NCT05756166 | Phase 1 Phase 2 | IFN α 2b | Metastatic or unresectable triple negative breast cancer | Rintatolimod, celecoxib and interferon α -2b with pembrolizumab | Recruiting | N/A |
| NCT02086721 | Phase 1 | IL-2 | Oligometastatic solid tumor | Combining L19-IL2 with SABR | Completed | Relinde, I et al., 2020 |
| NCT00590824 | Phase 2 | IL-2 | Resectable recurrent stage III or stage IV melanoma | Pilot hu14.18-IL2 | Completed | Albertini, M et al., 2018 |
| NCT05829057 | Phase 1 | IL-2 | Bladder, esophagus, liver, ovarian and small-cell lung cancer | The IIT study of evaluation of P-IL-2 | Recruiting | N/A |
| NCT05471271 | Not Applicable | IL-2 | Metastatic solid tumor | IL-2 | Recruiting | N/A |
| NCT05307874 | Phase 1 Phase 2 | IL-2 | Advanced solid tumors | ICT01 plus low dose SC IL-2 | Recruiting | N/A |
| NCT02306954 | Phase 2 | IL-2 | Metastatic renal cancer | High dose interleukin-2 (IL-2) and stereotactic body radiation (SBRT) | Active, not recruiting | Steven, K et al., 2012 |
| NCT00100906 | Phase 2 | IL-2 | Metastatic renal cell cancer | Sequential ATRA then IL-2 for modulation | Completed | N/A |
| NCT00003842 | Phase 1 | IL-4 | Recurrent malignant astrocytoma | IL-4(38-37)-PE38KDEL immunotoxin | Unknown Status | N/A |
| NCT00923910 | Phase 1 Phase 2 | IL-4 | Cancers of the blood | WT1 peptide-pulsed dendritic cells, donor lymphocytes, IL-4 | Completed | N/A |
| NCT00039052 | Phase 1 | IL-4 | Recurrent or metastatic kidney cancer, non-small cell lung cancer, or breast cancer | Intravenous interleukin-4 PE38KDEL cytotoxin | Completed | Yan, L et al., 2018 |
| NCT02858895 | Phase 2 | IL-4 | Recurrent or progressive glioblastoma | MDNA55 (IL-4) | Completed | N/A |
| NCT00001564 | Phase 2 | IL-4 | Recurrent pediatric sarcomas | Tumor-specific peptide vaccination and IL-2 with or without autologous T cell transplantation | Completed | N/A |
| NCT02494206 | Not Applicable | IL-4 | Breast cancer related upper extremity lymphedema | QBX258 (IL-4) | Completed | Mehra, B et al., 2021 |
| NCT00000769 | Phase 1 | IL-4 | AIDS and Kaposi's sarcoma | Interleukin-4 (IL-4) | Completed | N/A |
| NCT04253080 | Not Applicable | IL-4 | Cutaneous melanoma | IL-4 | Recruiting | N/A |
| NCT00014677 | Phase 2 | IL-4 | Recurrent glioblastoma multiforme | NBI-3001 | Completed | N/A |
| NCT04903197 | Phase 1 | IL-4 | Non-hodgkin lymphoma | VAY736 | Recruiting | N/A |
| NCT06191887 | Phase 1 | IL-4 | Relapsed or refractory B-cell hematologic malignancies | Bendamustine | Recruiting | N/A |
| NCT01368107 | Phase 2 | IL-7 | Metastatic breast cancer | IL-7 | Completed | N/A |
| NCT06204991 | Phase 1 | IL-7 | Locally advanced or metastatic melanoma | ADP-TILIL7 | Not yet recruiting | N/A |
| NCT06221553 | Phase 1 | IL-7 | Diffuse intrinsic pontine glioma | IL-7Ra | Recruiting | N/A |

Table 11 (continued)

| Identifier | Phase | Drug | Cancer types | Treatment | Status | References |
|-------------|--------------------|-------|---|---|-------------------------|------------------------|
| NCT03932565 | Phase 1 | IL-7 | Malignant solid tumors | Fourth-generation CAR-T | Unknown status | N/A |
| NCT03941769 | Phase 1 Phase 2 | IL-7 | Acute myeloid leukemia, myelodysplastic syndrome, chronic myeloid leukemia, or myeloproliferative disease after a haploidentical or cord blood stem cell transplant | 2018-0674—IL-7 | Completed | N/A |
| NCT03198546 | Phase 1 | IL-7 | Cancer with GPC3 expression | GPC3 and/or TGFβ | Recruiting | N/A |
| NCT05659628 | Phase 1 | IL-7 | RR-DLBCL | CD19 CAR-T | Recruiting | N/A |
| NCT05378464 | Phase 1 | IL-7 | Metastatic HER2+ breast cancer | Adoptive T cell therapy following HER2-pulsed dendritic cell vaccine & pepinemab /trastuzumab | Recruiting | N/A |
| NCT04381741 | Phase 1 | IL-7 | Relapsed or refractory diffuse large B cell lymphoma | CD19 CAR-T expressing IL7 and CCL19 combined with PD1 mAb | Enrolling by invitation | N/A |
| NCT04588038 | Phase 1 | IL-7 | Recurrent squamous cell carcinoma of head and neck | NT-17 (IL-7) | Recruiting | N/A |
| NCT05465954 | Phase 2 | IL-7 | Recurrent glioblastoma | Efneptakin alfa and Pembrolizumab | Recruiting | N/A |
| NCT03513952 | Phase 2 | IL-7 | Locally advanced, inoperable, or metastatic urothelial carcinoma | Atezolizumab and CYT107 | Completed | N/A |
| NCT00091338 | Phase 1 | IL-7 | Metastatic melanoma | Interleukin-7 and vaccine | Completed | N/A |
| NCT03239392 | Phase 1 Phase 2 | IL-9 | LGL leukemia or refractory CTCL | IV BNZ-1 | Completed | N/A |
| NCT01035697 | Observational | IL-9 | Cerebral palsy | IL-9 | Completed | N/A |
| NCT05991583 | Phase 1 Phase 2 | IL-10 | Advanced malignant tumors | IBB0979 | Recruiting | N/A |
| NCT05396339 | Phase 1 Phase 2 | IL-10 | Advanced malignant solid tumors | IAE0972 | Recruiting | N/A |
| NCT06468358 | Phase 1 Phase 2 | IL-10 | Advanced or metastatic solid tumors | LB1410 in Combination With LB4330 | Recruiting | N/A |
| NCT05704985 | Phase 1 | IL-10 | Locally advanced or metastatic EGFR+ tumors | Using DK210 | Recruiting | N/A |
| NCT02009449 | Phase 1 | IL-10 | Advanced solid tumors(VY) | LY3500518 | Active, not recruiting | N/A |
| NCT00433446 | Phase 2 | IL-6 | Metastatic prostate cancer | S0354, Anti-IL-6 chimeric monoclonal Antibody | Completed | N/A |
| NCT00841191 | Phase 1 Phase 2 | IL-6 | Solid tumors | Siltuximab (CNTO 328) | Completed | N/A |
| NCT05704634 | Phase 1 | IL-6 | Non-small cell lung cancer | IL6-receptor antibody Sarilumab in combination with anti-PD1 antibody Cemiplimab | Recruiting | N/A |
| NCT01219010 | Phase 1 | IL-6 | Undetermined significance, smoldering multiple myeloma, or indolent multiple myeloma | Siltuximab | Completed | Thomas, S et al., 2014 |
| NCT00955812 | Phase 1 | IL-6 | Solid tumors | STAT3 inhibitor | Completed | N/A |
| NCT04641910 | Observational | IL-6 | Acute myeloid leukemia | FLT3 ligand plasma concentration kinetic profile and IL-6 concentration | Recruiting | N/A |
| NCT01417546 | Phase 1 | IL-12 | Solid tumors | NHS-IL12 | Completed | Nicole, J et al., 2023 |
| NCT02483312 | Phase 1 | IL-12 | Acute myelogenous leukemia | IL-12 | Recruiting | N/A |
| NCT04471987 | Phase 1 | IL-12 | Advanced solid tumor, metastatic solid tumor | IL12-L19 | Recruiting | N/A |

Table 11 (continued)

| Identifier | Phase | Drug | Cancer types | Treatment | Status | References |
|-------------|--------------------|-------|--|---|--------------------|-------------------------|
| NCT04708470 | Phase 1 Phase 2 | IL-12 | Advanced cancers including HPV-associated malignancies, small bowel, and colon cancers | Bintrafusp alfa, NHS-IL12, Entinostat | Recruiting | N/A |
| NCT00005604 | Phase 1 | IL-12 | Advanced solid tumors | Interleukin-12 plus interleukin-2 | Completed | N/A |
| NCT02531425 | Phase 1 | IL-12 | TNBC | IT-pIL12-EP | Completed | N/A |
| NCT00015977 | Phase 2 | IL-12 | Metastatic prostate cancer | Vaccine therapy plus interleukin-12 | Completed | N/A |
| NCT00004074 | Phase 1 | IL-12 | Cancer that has high levels of HER2/Neu | Interleukin-12 and Trastuzumab | Completed | N/A |
| NCT00323206 | Not Applicable | IL-12 | Malignant melanoma | Phase I trial of intratumoral pIL-12 electroporation | Completed | N/A |
| NCT00406939 | Phase 1 | IL-12 | Prostate cancer | IL-12 | Completed | N/A |
| NCT03281382 | Phase 1 | IL-12 | Metastatic pancreatic cancer | Interleukin 12 | Completed | Barton, K et al., 2021 |
| NCT02960594 | Phase 1 | IL-12 | Solid tumors at high risk of relapse | hTERT immunotherapy alone or in combination with IL-12 DNA | Completed | N/A |
| NCT00016289 | Phase 2 | IL-12 | Ovarian epithelial cancer or primary peritoneal cancer | Interleukin-12 | Completed | N/A |
| NCT01440816 | Phase 2 | IL-12 | Merkel cell cancer | IL-12 | Completed | N/A |
| NCT05756907 | Phase 1 Phase 2 | IL-12 | Platinum-resistant ovarian cancer | Combination of SON-1010 (IL12-FHAB) and Atezolizumab | Recruiting | N/A |
| NCT05788926 | Phase 1 | IL-12 | Metastatic non-small cell lung cancer | TG6050 | Recruiting | N/A |
| NCT01118052 | Phase 2 | IL-12 | Persistent or recurrent ovarian epithelial cancer, fallopian tube cancer, or primary peritoneal cancer | EGEN-001 | Completed | Alvarez, R et al., 2014 |
| NCT00028535 | Phase 1 | IL-12 | Solid tumors | Interleukin-12 | Completed | N/A |
| NCT02395822 | Phase 2 | IL-15 | Relapsed or refractory AML | Haplo NK With SQ IL-15 | Completed | N/A |
| NCT01021059 | Phase 1 | IL-15 | Refractory metastatic malignant melanoma and metastatic renal cell cancer | Intravenous recombinant human IL-15 | Completed | N/A |
| NCT02689453 | Phase 1 | IL-15 | Refractory or relapsed chronic and acute adult T-cell leukemia | Subcutaneous recombinant human IL-15 (s.c. rhIL-15) and Alemtuzumab | Completed | N/A |
| NCT03669172 | Phase 1 Phase 2 | IL-15 | Acute leukemia | IL-15 | Completed | N/A |
| NCT05110742 | Phase 1 Phase 2 | IL-15 | Relapsed/refractory hematological malignances | CD5 CAR engineered IL15-transduced | Recruiting | N/A |
| NCT00107718 | Phase 2 | IL-18 | Previously untreated metastatic melanoma | SB-485232 | Completed | N/A |
| NCT00659178 | Phase 1 | IL-18 | Advanced stage epithelial ovarian cancer | SB-485232 (Interleukin 18) | Completed | N/A |
| NCT01768338 | Phase 1 | IL-18 | Lymphoma | Recombinant human IL-18 and ofatumumab | Completed | N/A |
| NCT02277392 | Phase 1 | IL-18 | Recurrent ovarian, fallopian tube or primary peritoneal cancer | Recombinant human interleukin-18 (Sb-485232) combined with adoptive transfer of vaccine-primed CD3/CD28-costimulated autologous T-cells | Withdrawn | N/A |
| NCT05914376 | Phase 1 | IL-21 | Advanced tumors | Recombinant human IL-21 oncolytic vaccinia virus injection (hv01) | Recruiting | N/A |
| NCT06198296 | Phase 1 | IL-21 | GPC3-positive solid tumors | IL-15 and IL-21 armored GPC3-CAR T cells | Not yet recruiting | N/A |
| NCT00617253 | Phase 2 | IL-21 | Stage IV renal cell carcinoma | Recombinant human IL-21 (rIL-21) and sunitinib | Completed | N/A |

Table 11 (continued)

| Identifier | Phase | Drug | Cancer types | Treatment | Status | References |
|-------------|--------------------|-------|--|---|--------------------|------------|
| NCT00336986 | Phase 2 | IL-21 | Metastatic melanoma | IL-21 | Completed | N/A |
| NCT02809092 | Phase 1 Phase 2 | IL-21 | Acute myeloid leukemia | Interleukin-21 | Unknown status | N/A |
| NCT04715191 | Phase 1 | IL-21 | Pediatric solid tumors | Interleukin-15 and -21 armored glypican-3-specific chimeric antigen receptor | Not yet recruiting | N/A |
| NCT01629758 | Phase 1 | IL-21 | Solid tumors | IL-21/anti-PD-1 combination | Completed | N/A |
| NCT01489059 | Phase 1 | IL-21 | Melanoma | IL-21/ipilimumab combination | Completed | N/A |
| NCT00514085 | Phase 2 | IL-21 | Metastatic or recurrent malignant melanoma | Interleukin-21 | Completed | N/A |
| NCT04220684 | Phase 1 | IL-21 | Relapsed/refractory acute myeloid leukemia | IL-21 | Recruiting | N/A |
| NCT04729543 | Phase 1 Phase 2 | IL-21 | Melanoma and head and neck cancer (MC2TCR) | MAGE-C2/HLA-A2 TCRT cells | Recruiting | N/A |
| NCT00095108 | Phase 1 | IL-21 | Metastatic malignant melanoma and metastatic kidney cancer | Interleukin-21 | Completed | N/A |
| NCT00523380 | Phase 2 | IL-21 | Ovarian cancer | Recombinant interleukin-21 | Completed | N/A |
| NCT02858895 | Phase 2 | IL-4R | Recurrent or progressive glioblastoma | MDNA55 | Completed | N/A |
| NCT00019032 | Phase 1 | IL-2R | Chronic lymphocytic leukemia | Monoclonal antibody | Completed | N/A |
| NCT00076180 | Phase 1 | IL-2R | T-Cell large granular lymphocytic leukemia | Hu-Mik-beta1 | Completed | N/A |
| NCT00001249 | Phase 1 | IL-2R | Cutaneous T-Cell lymphoma (CTCL) and adult T-Cell leukemia (ATL) | Yttrium-90 radiolabeled anti-Tac | Completed | N/A |
| NCT00072969 | Phase 2 | IL-2R | Cytopenia of myelodysplastic syndrome (MDS) | Recombinant humanized anti-IL-2 receptor Antibody (Dacilizumab) versus Antithymocyte globulin (ATG) | Completed | N/A |
| NCT05200559 | Phase 1 Phase 2 | IL-2R | Recurrent or metastatic solid tumors | E7777 combined with pembrolizumab | Recruiting | N/A |
| NCT00001567 | Phase 2 | IL-2R | Hairy cell leukemia | Roferon-A | Completed | N/A |
| NCT05699811 | Phase 1 Phase 2 | IFNα | Locally advanced/metastatic solid tumors | IFNα | Recruiting | N/A |
| NCT02151448 | Phase 1 Phase 2 | IFNα | Peritoneal surface malignancies | αDC1 vaccine + chemokine modulatory regimen (CKM) | Completed | N/A |
| NCT02923466 | Phase 1 | IFNβ | Refractory solid tumors | Ph1 administration of VSV-IFNβ-NIS monotherapy and in combination with avelumab | Completed | N/A |
| NCT03647163 | Phase 1 Phase 2 | IFNβ | Select solid tumors | Systemic VSV-IFNβ-NIS in Combination with checkpoint inhibitor therapy | Completed | N/A |
| NCT04291105 | Phase 2 | IFNβ | Select solid tumors | Voyager V1 | Recruiting | N/A |
| NCT05076760 | Phase 1 | IFNβ | Solid tumors including non-small cell lung cancer | MEM-288 oncolytic virus alone and in combination with nivolumab | Recruiting | N/A |
| NCT04695327 | Phase 1 | TNFα | Oncolytic adenovirus TILT-123 | TNFα and IL-2 | Recruiting | N/A |
| NCT02076620 | Phase 1 | TNFα | Advanced solid tumours | L19 TNFα | Completed | N/A |
| NCT03259230 | Observational | IFNγ | Malignancy-associated hemophagocytic lymphohistiocytosis (M-HLH) | Interferon γ and other inflammatory mediators | Completed | N/A |

Table 11 (continued)

| Identifier | Phase | Drug | Cancer types | Treatment | Status | References |
|-------------|--------------------|--------------|---|--|---|--------------------------|
| NCT06344052 | Phase 2 | IFN γ | Locally advanced basal cell carcinoma | SP-002 with vismodegib | Recruiting | N/A |
| NCT06430788 | Phase 2 | IFN γ | Pediatric aplastic anemia | Enapalumab | Recruiting | N/A |
| NCT06052839 | Phase 2 | IFN γ | Recurrent/metastatic HNSCC | Pulsed dose chemotherapy plus pembrolizumab | Recruiting | N/A |
| NCT00004016 | Phase 1 | IFN γ | Recurrent or metastatic melanoma or other solid tumors | Interferon γ | Completed | N/A |
| NCT02614456 | Phase 1 | IFN γ | Advanced solid tumors | Combination of interferon- γ and nivolumab | Completed | Zibelman, M et al., 2023 |
| NCT01957709 | Early Phase 1 | IFN γ | Soft tissue sarcoma | Recombinant interferon γ | Terminated (Enough samples were collected for data analysis) | N/A |
| NCT00059878 | Phase 2 | IFN γ | Aspergillosis or other fungal infections | Voriconazole with or without interferon γ | Completed | N/A |
| NCT00004032 | Phase 1 | IFN γ | Refractory epithelial ovarian cancer | Tumor vaccine and interferon γ | Completed | N/A |
| NCT02197169 | Phase 1 | IFN γ | Recurrent glioblastoma or gliosarcoma brain tumors (TARGET-I) | DNX-2401 With Interferon Gamma (IFN- γ) | Completed | N/A |
| NCT06371482 | Phase 2 | IFN γ | Limited stage small cell lung cancer (Camel-01) | Durvalumab combined with chemoradiotherapy | Recruiting | N/A |
| NCT05156541 | Phase 3 | IFN γ | Anogenital warts (ING-HPV-1) | Drug Ingaron (Interferon- γ) | Completed | N/A |
| NCT00786643 | Phase 2 | IFN γ | Metastatic colorectal carcinoma | Interferon γ | Completed | N/A |
| NCT03112590 | Phase 1 Phase 2 | IFN γ | HER-2 positive breast cancer | Interferon γ | Completed | N/A |
| NCT00616720 | Phase 2 | IFN γ | Multiple myeloma | Interferon γ or aldesleukin and vaccine | Completed | N/A |
| NCT02550678 | Phase 1 Phase 2 | IFN γ | Low-risk nodular basal cell carcinoma (ASN-002-001) | ASN-002, 5-FU | Completed | N/A |
| NCT03063632 | Phase 2 | IFN γ | Mycosis Fungoides and Sézary Syndrome and Advanced Synovial Sarcoma | Combination of two experimental drugs MK-3475 (Pembrolizumab) and interferon- γ | Completed | N/A |
| NCT00070187 | Phase 2 Phase 3 | IFN γ | Refractory or relapsed Hodgkin's lymphoma | Cyclosporine, Interferon γ , and Interleukin-2 after high-dose myeloablative chemotherapy | Completed | N/A |
| NCT04628338 | Early Phase 1 | IFN γ | Acute myeloid leukemia and myelodysplastic syndrome | IFN- γ | Completed | N/A |

pancreatic inflammation caused by LPS and the resulting cytokine storm, while also inhibiting cell proliferation in pancreatic cancer [450]. Another promising approach is to restore a balanced host response by normalizing the inflammatory network. This involves reducing high levels of pro-inflammatory cytokines and other tumor-promoting characteristics of infiltrating cells while increasing the levels of anti-inflammatory cytokines.

3) Do sex steroid hormones play a role in the interplay between inflammation and cancer? The occurrence of tumors varies between genders due to differing sex hormones. Analyzing the interaction between these hormones and inflammation in cancer development could enhance the treatment of hormone-related cancers, such as breast, lung, ovarian, cervical, and prostate cancers. Recent studies have shown that Treg cells, which play a crucial role in managing immune responses and reducing tissue inflammation, are more abundant in male visceral adipose tissue (VAT) compared to females. These differences in Treg cell populations are influenced by the tissue environment and shaped by sex hormones, which help to decrease inflammation in adipose tissue [451]. Considering the widespread use of selective androgen receptor modulators and selective estrogen receptor modulators like tamoxifen, understanding the interaction between sex hormones, inflammation, and cancer will significantly influence the clinical treatment of cancer.

Discussion

Cancer is still the second leading cause of death after cardiovascular disease in the world, the main reason is that about 50% of cancers are diagnosed at a late stage. The limited efficacy of existing treatment methods and the high failure rate in drug development make the situation more complex, and there is an urgent need for a better understanding of disease mechanisms, the development of early detection technology, and cancer chemoprevention strategies. Here, we systematically analyzed the role of inflammation and its related molecules in tumors. We expect that a substantial amount of information and intricacies continually uncovered in the area will eventually be condensed into a few key principles that regulate the molecular and cellular processes underpinning inflammation that promote tumor growth.

The role of inflammatory molecules in cancer has attracted the researchers' attention during the past decades. New insights into the pro- or anti-tumor effect in the tumor and its microenvironment have given impetus to drug discovery and patient evaluation of inflammation-directed strategies. For example, as the most effective and widely distributed family of cytokines, IFN-Is, or type I interferons, are essential for launching a successful anti-tumor response. Antigen-presenting cells require

interferon-alpha to trigger T cell responses; additionally, IFN-Is directly boost CD8+ T cell activity and cytotoxicity, stimulate CD4+ Th1 cell development, augment cytotoxicity from natural killer cells, and limit regulatory T cells. However, IFN-Is induce the expression of negative regulatory molecules, which can reduce immunological responses and induce fatigue, thus facilitating the growth of tumors. Besides, other inflammatory molecules such as interleukins, tumor necrosis factors, colony-stimulating factors, chemokines, and inflammasomes also showed multifaceted effects in cancer. The two-side effect may depend on the cancer type, time, cells present, total IFN-I signal levels, and others. Therefore, further studies are needed to dissect the dual role of these inflammatory molecules in different cancer models, which will enable the identification of novel molecular and immunological targets and lead to the development of novel therapeutic strategies. Recently, researchers created a portable smart blue-light controlled (PSLC) gadget based on optogenetic technology. The findings demonstrate that blue light can efficiently control pro-inflammatory cytokine expression in both in vitro and in vivo settings, which offers a unique strategy for cytokine therapy [452].

FDA-approved non-anticancer drugs- "old medicine", have obvious advantages in anti-tumor treatment—"new use", because FDA-approved drug applications could better avoid numerous obstacles and uncertainties in every step of converting drugs into clinical applications. This suggests that the application of non-anticancer drugs may be a promising strategy for cancer chemoprevention. The evident role of inflammation in cancer development and progression prompted the application of anti-inflammatory medications as a therapeutic strategy. While the application of anti-inflammatory drugs in clinical anti-tumor therapy still needs to overcome several obstacles.

1) Targeted delivery to the cancerous cell. Although the anti-inflammatory medications now approved by the FDA are undoubtedly effective, their off-target effects and toxicities make them less desirable options for cancer therapy when taken at the current dosages and frequency. Rather, to improve drug targetability and reduce off-target adverse effects, recent developments in nanotechnology have facilitated a paradigm shift away from traditional anti-inflammatory medications and toward anti-inflammatory nano-therapeutics in cancer therapy. However, the field of anti-inflammatory nanomedicines is still in its infancy with little commercial application, and there is a great need to consider and identify the issues with nano-inflammatory therapies to aid in the practical clinical translation of commercially available anti-inflammatory nanotherapeutics.

2) Tumor cells have different or even opposite responses to these anti-inflammatory drugs. Abundant evidence indicates that

anti-inflammation drugs are promising candidates for preventing carcinogenesis and cancer recurrence because of their availability and relatively low occurrence of side effects compared to other chemotherapeutic drugs. Nevertheless, due to the paucity of information and the complexity of carcinogenesis, these anti-inflammation drugs showed different or even inverse effects. Well-designed, long-term clinical trials are required to ascertain the clinical application potential of these medications, and additional trials are required to investigate the doses, kinds, and length of response of these drugs. 3) The side effects after long-term use. Although the FDA-approved anti-inflammatory drugs have far fewer side effects than chemotherapeutics, some anti-inflammatory drugs, such as aspirin, also showed obvious side effects after long-term use. The main side effects of NSAIDs are cardiovascular (CV) and renal adverse effects. While the main side effects of seroidal anti-inflammatory drugs such as glucocorticoids (GRs) are diabetes, glaucoma, and suppression of the hypothalamic–pituitary–adrenal axis, among others. NSAIDs cause cell death by directly targeting mitochondria, while NSAIDs have been shown to enhance mitochondrial health in dose-dependent ways [453], which suggesting that future research should focus on comparing equipotent dosages of these drugs. Additionally, it's important to assess the alleviation of symptoms. There are ongoing efforts to develop selective GR agonists (SEGRAs) with the hypothesis that they are safer than traditional glucocorticoids. To support this hypothesis, appropriate in vitro and in vivo studies are needed to provide reliable experimental results. 4) Patients continue to experience other thromboembolic events despite aspirin therapy, which is known as aspirin resistance (AR). Besides, anti-inflammatory drugs may lead to a degree of drug resistance. For example, dexamethasone can increase the resistance of human tumor cells to ionizing radiation and chemotherapy [454].

Hence, larger-scale adoption of the chemoprevention strategy is likely to require improved identification of individuals for whom the protective benefits outweigh the harms. Such a precision medicine approach may emerge through further clarification of these anti-inflammatory drugs' mechanism of action.

Abbreviations

| | | | |
|----------------|--|---------------|--|
| IL | Interleukin | PD-L1 | Programmed cell death ligand 1 |
| TNF- α | Tumor necrosis factor alpha | LAP | LC3-associated phagocytosis |
| NF- κ B | Nuclear factor- κ B | LANDO | LC3-associated endocytosis |
| STAT3 | Signal transducer and activator of transcription 3 | Ab | Beta-amyloid |
| NSAIDs | Nonsteroidal anti-inflammatory drugs | IFN | Interferon |
| CRC | Colorectal cancer | ROS | Reactive oxygen species |
| USPSTF | US Preventive Services Task Force | IKK | I κ B kinase |
| DR5 | Death receptor 5 | IRGM | Immunity-related GTPase M |
| FDA | Food and Drug Administration | AIM 2 | Absent in melanoma 2 |
| TLRs | Toll-like receptors | FUNDC1 | FUN14 domain-containing 1 |
| PD-1 | Programmed cell death protein 1 | MMPs | Matrix metalloproteinases |
| | | COX | Cyclooxygenase |
| | | PIK3CA | Phosphatidylinositol 4,5-bisphosphate 3-kinase catalytic subunit alpha isoform |
| | | AKT | Protein kinase B |
| | | PTEN | Phosphatase and tensin homolog |
| | | Wnt | Wingless-type MMTV integration site family |
| | | Fc-IL-4 | Interleukin-4 fusion protein |
| | | AA | African American |
| | | EA | European American |
| | | CLSs | Crown-like structures |
| | | Tregs | T regulatory cells |
| | | NK cells | Natural killer cells |
| | | ICIs | Checkpoint inhibitors |
| | | irAEs | Immune-related adverse events |
| | | MSI-H | High microsatellite instability |
| | | PPAR α | Peroxisome proliferator-activated receptor alpha |
| | | MetS | Metabolic syndrome |
| | | 3-MA | Methyladenine |
| | | TLR | Toll-like receptor |
| | | NLRP3 | Nucleotide-binding domain leucine-rich repeat (NLR) and pyrin domain-containing receptor |
| | | TGF- β | Transforming growth factor beta |
| | | ECs | Endothelial cells |
| | | VEGF | Vascular endothelial growth factor |
| | | BDNF | Brain-derived neurotrophic factor |
| | | TrkB | Tropomyosin receptor kinase B |
| | | CXCR | C-X-C chemokine receptor |
| | | CXCL | (C-X-C motif) ligand |
| | | CX3CR1 | CX3Chemokine receptor 1 |
| | | CCR1 | CC-chemokine receptor 1 |
| | | EMT | Epithelial-mesenchymal transition |
| | | DDR | DNA damage response |
| | | IFN- γ | Interferon gamma |
| | | STAT6 | Signal transducer and activator of transcription 6 |
| | | EGCG | Epigallocatechin-3-gallate |
| | | ICB | Immune checkpoint blockade |
| | | CD | Crohn's disease |
| | | UC | Ulcerative colitis |
| | | IBD | Inflammatory bowel disease |
| | | RA | Rheumatoid arthritis |
| | | NMSC | Nonmelanoma skin cancer |
| | | HPV | Human papillomavirus |
| | | H. pylori | Helicobacter pylori |
| | | HBV | Hepatitis B virus |
| | | HCV | Hepatitis C virus |
| | | NAFLD | Nonalcoholic fatty liver disease |
| | | MYC | Myelocytomatosis oncogene |
| | | RAS | Rats arcomaviral oncogene homolog |
| | | KRAS | Kristen rats arcomaviral oncogene homolog |
| | | NSCLC | Non-small cell lung cancer |
| | | VHL | Von Hippel-Lindau |
| | | JAK | Janus kinase |
| | | ISGF3 | IFN-stimulated gene (ISG) factor 3 |
| | | RIG-I | Retinoic acid-inducible gene I |
| | | cGAMP | Cyclic guanosine monophosphate (GMP)-adenosine monophosphate (AMP) |
| | | cGAS | Cyclic GMP-AMP synthase |
| | | STING | Stimulator of interferon genes |
| | | IOD | Indoleamine-2,3-dioxygenase |
| | | GM-CSF | Granulocyte/macrophage colony-stimulating factor |

| | |
|--------|--|
| M-CSF | Macrophage colony-stimulating factor |
| G-CSF | Granulocyte colony-stimulating factor |
| GPCRs | Gai-protein-coupled seven-transmembrane-spanning receptor |
| TME | Tumor microenvironment |
| LRR | Leucine rich repeat |
| PYD | Pyrin domain |
| NLRP1 | Pyrin domain (PYD) domain-containing protein 1 |
| NLRC4 | NLR family caspase activation and recruitment domain-containing protein 4 |
| NLRP6 | NOD-like receptor family pyrin domain containing 6 |
| ASC | Apoptosis-associated speck-like protein containing caspase activation and recruitment domain |
| DSS | Dextran sulfate sodium |
| AOM | Azoxymethane |
| CARD | C-terminal caspase recruitment domain |
| FIIND | Function to find domain |
| DPP | Dipeptidyl protease |
| LT | Lethal toxin |
| LUAD | Lung adenocarcinoma |
| PAAD | Pancreatic adenocarcinoma |
| TRX | Thioredoxin |
| NBD | Nucleotide-binding domain |
| HD1 | Helical domain 1 |
| WHD | Winged helix domain (WHD) |
| HD2 | Helical domain 2 |
| T4SS | Type IV secretion system |
| T3SS | Type III secretion system |
| IRF | Interferon regulatory factor |
| NASH | Nonalcoholic steatohepatitis |
| AH | Alcoholic hepatitis |
| GC | Gastric cancer |
| SCLC | Small cell lung cancer |
| ALRs | AIM2-like receptors |
| POP3 | PYD-only protein 3 |
| HCMV | Human cytomegalovirus |
| HSV-1 | Herpes simplex virus-1 |
| HbeAG | Hepatitis B e-antigen |
| TMS1 | Target of methylation-induced silencing-1 |
| TSLP | Thymic stromal lymphopoietin |
| CAFs | Cancer-associated fibroblasts |
| ECM | Extracellular matrix |
| TNC | Tenascin-C |
| PSA | Prostate-specific antigen |
| RECIST | Response evaluation criteria in solid tumors |
| CR | Complete response |
| MSS | Metastatic microsatellite stable |
| dMMR | Mismatch repair-deficient |
| AI | Artificial intelligence |
| CAD | Computer-aided diagnostic |
| ML | Machine learning |
| DL | Deep learning |
| ANN | Artificial neural networks |
| TILs | Tumor-infiltrating lymphocytes |
| AIPs | Anti-inflammatory peptides |
| BET | Bromodomain and extra-terminal domain |
| iBET | Bromodomain and extra-terminal domain (BET) protein inhibitors |
| OSCC | Oral squamous cell carcinoma |
| VAT | Visceral adipose tissue |
| PSLC | Portable smart blue-light controlled |
| CV | Cardiovascular |
| GRs | Glucocorticoids |
| SEGRAs | Selective GR agonists |
| AR | Aspirin resistance |

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Authors' contributions

All authors have contributed to the article and approved its publication. Y.X., F.L., designed and wrote the manuscript. Y.W. and Y.Z., prepared the figures.

Y.J. and Q.W., modified the language. Z.D. and K.L., reviewed and edited the manuscript.

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Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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