PLASTOME ANNOUNCEMENT

OPEN ACCESS

The complete plastid genome sequence of *Chloranthus fortunei* (A. Gray) Solms-Laub. in Chloranthaceae

Jong-Soo Kang^a (D), Bo-Yun Kim^b and Ki-Oug Yoo^a (D)

^aDepartment of Biological Sciences, Kangwon National University, Chuncheon, South Korea; ^bPlant Resources Division, National Institute of Biological Resources, Incheon, South Korea

ABSTRACT

Chloranthus fortunei (A. Gray) Solms-Laub. is a perennial herb in a basal angiosperm family Chloranthaceae. Here, we reported the complete plastid genome of *C. fortunei* using Illumina short-read data. The total genome size was 157,063 bp in length, containing 79 protein-coding genes, 30 tRNA genes, and four rRNA genes. The gene content and order were consistent with previously reported *Chloranthus* plastid genomes. The overall GC content of the *C. fortunei* plastid genome was 39.0%. In the phylogenetic result, genus *Chloranthus* was monophyletic and divided into two subclades: *C. japonicus+C. angustifolius+C. fortunei*, and *C. henryi+C. spicatus+C. erectus*. Our phylogenetic result was consistent with previous phylogenetic studies, and was supported by a previously proposed infrageneric classification of the genus *Chloranthus*.

ARTICLE HISTORY

Received 14 July 2022 Accepted 29 September 2022

Taylor & Francis

Taylor & Francis Group

KEYWORDS

Chloranthus fortunei; Chloranthus; Chloranthaceae; plastid genome

Chloranthus Swartz (Chloranthaceae) consists of two subgenera, subgenus Tricercandra and subg. Chloranthus, based on androecium morphology, such as the extent of splitting in the tripartite lobes (Kong 2000a; Kong and Chen 2000; Kong et al. 2002). Chloranthus fortunei (A. Gray) Solms-Laub. (1869) belongs to subg. Tricercandra, and is distributed in southern parts of China, Korea, and Japan (Kim 2007; Xia and Jérémie 2007). This species has been cultivated as an ornamental herb, and also used for the Chinese folk medicine as a treatment of bone fractures (Ben Cao 1999). Morphologically, C. fortunei is very similar to C. japonicus Siebold which is widely distributed in East Asia (Kim 2007; Xia and Jérémie 2007); however, C. fortunei can be distinguished from the former by the anther position of the androecium, ploidy level, and tripartite and roecium with long longitudinal connections (Kong 2000b; Kim 2007; Xia and Jérémie 2007; Figure 1). Whole plastid genomes have been widely used for molecular phylogenetics, species identifications, and conservation genetics (Burke et al. 2012; Huang et al. 2014; Walker et al. 2014). Here, we report the plastid genome of C. fortunei, which will be useful for the conservation genetic studies of this species as well as phylogenetic reconstructions of Chloranthus and other basal angiosperms.

Leaf material of *C. fortunei* was collected from Ongnyeobong, Geoje-si, Gyeongsangnam-do province of South Korea (latitude 34.8455, longitude 128.6954). The voucher specimen (KWNU91773) has been deposited in the Kangwon National University Herbarium (KWNU; https://

biology.kangwon.ac.kr/, Ki-Oug Yoo, yooko@kangwon.ac.kr). Total genomic DNA was extracted from silica gel dried leaves using the Exgene Plant SV Midi Kit (Geneall Biotechnology, Seoul, South Korea). Paired-end reads of 2×150 bp were generated using an Illumina HiSeq Xten (Theragen Bio Co. Ltd., Suwon, South Korea). A total of 2.26 GB raw reads of 150 bp were generated, of which 146,514 paired-end reads were extracted as plastid genome sequences using a reference genome sequence of the C. japonica plastid genome (KP256024). Using 146,514 reads, the de novo assembly was performed using GetOrganelle pipeline (Jin et al. 2020) with C. japonica plastid genome as a reference, and the assembled contig was manually confirmed using Geneious 7.1 (Biomatters Ltd, Auckland, New Zealand). The initial annotation of the C. fortunei plastid genome was performed using GeSeq (Tillich et al. 2017). After the initial annotation, putative starts, stops, and intron positions were determined by comparison with homologous genes in previously reported Chloranthus plastid genomes. The tRNA genes were annotated using GeSeq and tRNAscan-SE (Schattner et al. 2005). The annotated sequence was deposited in the NCBI GenBank under accession number ON023121, and the circular map of the C. fortunei plastid genome was drawn using the CPGview (http://www.1kmpg.cn/cpgview/).

The genome size of the *C. fortunei* plastid genome was 157,063 bp, including a pair of inverted repeat (IR) regions of 26,102 bp separated by the small single-copy (SSC) region of 18,484 bp, and the large single-copy (LSC) region of

© 2022 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

CONTACT Ki-Oug Yoo 🐼 yooko@kangwon.ac.kr 💽 Department of Biological Sciences, Kangwon National University, Chuncheon, Gangwon-do, South Korea *Present address: Department of Agriculture, Forestry and Bioresources, Plant Genomics & Breeding Institute, College of Agriculture & Life Sciences, Seoul National University, Seoul, South Korea

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Figure 1. Chloranthus fortunei. This species has a tripartite androecium with long longitudinal connections. (A) Habitat, (B) flower, and (C) the specimen deposited in Kangwon National University Herbarium (KWNU) under the voucher no. KWNU91773. The photos of *C. fortunei* in field (A, B) and the voucher specimen (C) were taken and provided by Jong-Soo Kang and Ki-Oug Yoo, respectively.

86,375 bp (Figure 2). The *C. fortunei* plastid genome contained 113 genes, 18 of which were duplicated in the IR region, giving a total of 131 genes. The plastid genome of *C. fortunei* contained 30 distinct tRNAs, seven of which were duplicated in the IR region. Ten protein-coding genes (*atpF, ndhA*, *ndhB*, *petB*, *petD*, *rps12*, *rps16*, *rpl2*, *rpl16*, and *rpoC1*) and six tRNA genes (*trnA-UGC*, *trnG-GCC*, *trnI-GAU*, *trnK-UUU*, *trnL-UAA*, and *trnV-UAC*) contained one intron, while two genes (*clpP*, *ycf3*) contained two introns. A trans-spliced *rps12* gene was divided into two independent transcription units (exon 1, and exons 2–3) as described in previous studies (Hildebrand et al. 1988; Schmitz-Linneweber et al. 2006). The overall GC content was 39.0% in the entire genome, 37.4% in the LSC, 43.2% in the IR, and 34.1% in the SSC regions.

Phylogenetic analysis based on 78 protein-coding genes was performed using representative species from Amborellales in basal angiosperms to Magnoliales in magnoliids, and *Amborella trichopoda* was selected as the outgroup (Figure 3). A total of 69,404 bp was aligned using MAFFT (Katoh and Standley 2013). Maximum-likelihood (ML) analysis was performed using RAxML v. 7.4.2 with 1000 bootstrap replicates and the GTR + I + G model (Stamatakis 2006; Darriba et al. 2012). Our phylogenetic result was consistent with topologies from previous studies in which all families and orders were monophyletic (Angiosperm Phylogeny Group 2016) (Figure 1). Within Chloranthaceae, Sarcandra glabra was sister to the clade of Chloranthus with 100% bootstrap supporting values, and the genus Chloranthus was monophyletic as shown in previous studies (Kong et al. 2002; Zhang et al. 2011). The three species, C. fortunei, C. angustifolius, and C. japonicus of subg. Tricercandra formed a subclade, and the subclade was sister to the other clade of subg. Chloranthus including C. henryi, C. spicatus, and C. erectus with 100% bootstrap supporting values (Figure 3). The pairwise identity of concatenated 78 protein-coding gene sequences within the genus Chloranthus was 99.2%, and those within both two subgenera was 99.5%, respectively.

Ethical approval

This study complies with relevant institutional, national, and international guidelines and legislations. According to the national and



Figure 2. The map of the *Chloranthus fortunei* plastid genome. The circular map of the *C. fortunei* plastome was drawn using the CPGview program. The map consists of six circles and information about each circle is as follows: (from the center) the first circle indicates repeat distribution. The second circle indicates the tandem repeats with short bars. The third circle indicates the microsatellite sequences with short bars. The fourth circle indicates the size of LSC, SSC, and IR regions. The fifth circle indicates the GC content. The sixth circle indicates the genes having different colors based on their functions.

local legislations, no specific permission was required for collecting the species in this study, and Ki-Oug Yoo identified and deposited the voucher specimen in the Kangwon National University Herbarium (KWNU).

Author contributions

Ki-Oug Yoo and Bo-Yun Kim planned and designed the research. Jong-Soo Kang and Ki-Oug Yoo collected the plant material. Jong-Soo Kang and Bo-Yun Kim performed experiments. Jong-Soo Kang performed analysis and interpretation of data. Jong-Soo Kang and Ki-Oug Yoo wrote the first draft of the manuscript, and all authors revised and approved the final manuscript. All authors agree to be accountable for all aspects of the work.

Disclosure statement

No potential competing interest was reported by the authors.

Funding

This work was supported by the National Institute of Biological Resources (NIBR) Grant funded by the Korea Government [No. NIBR202212101].

ORCID

Jong-Soo Kang () http://orcid.org/0000-0002-0172-0021 Ki-Oug Yoo () http://orcid.org/0000-0002-2676-6878

Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI (https://www.ncbi.nlm.nih.gov/) under the accession no. ON023121. The associated BioProject, SRA, and Bio-Sample numbers are PRJNA816642, SRR18360190, and SAMN26686909, respectively.



Figure 3. Phylogenetic tree based on 78 protein-coding genes using the ML method. Asterisk indicates newly reported plastid genome in this study. Bootstrap values are shown near the nodes. The following sequences were used: *Myristica fragrans* MN495963 (Cai et al. 2021), *Magnolia sieboldii* MN990583 (Wang et al. 2020), *Cinnamomum longipetiolatum* MN698965 (Zheng et al. 2019), *Calycanthus chinensis* MG561304 (Chen et al. 2019), *Piper kadsura* KT223569 (Lee et al. 2016), *Saururus chinensis* MN263891 (Jin et al. 2019), *Drimys granadensis* DQ887676 (Cai et al. 2006), *Chloranthus fortunei* ON023121 (this study), *Chloranthus angustifolius* MW581013 (Zhang unpublished), *Chloranthus japonicus* KP256024 (Sun et al. 2016), *Chloranthus erectus* MH394412 (Zeng et al. 2018), *Chloranthus spicatus* EF380352 (Hansen et al. 2007), *Chloranthus henryi* MK922064 (Liu et al. 2019), *Sacuradra glabra* MH939147 (Wang et al. 2020), *Schisandra sphenanthera* MK193856 (Wei et al. 2020), *Illicium anisatum* KY085919 (Zhang and Handy unpublished), *Cabomba aquatica* MG720559 (Gruenstaeudl et al. 2018), *Nymphaea colorata* MT107631 (Sun et al. 2021), and *Amborella trichopoda* AJ506156 (Goremykin et al. 2003).

References

- Angiosperm Phylogeny Group. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. Bot J Linn Soc. 181:1–20.
- Ben Cao ZB. 1999. Editorial committee of the administration bureau of traditional Chinese medicine. Chinese Materia Medica (Zhonghua Benchao). Vol. 3. Shanghai, China: Shanghai Science & Technology Press; p. 449–450.
- Burke SV, Grennan CP, Duvall MR. 2012. Plastome sequences of two New World bamboos-Arundinaria gigantea and Cryptochloa strictiflora (Poaceae)-extend phylogenomic understanding of Bambusoideae. Am J Bot. 99(12):1951–1961.
- Cai CN, Ma H, Ci XQ, Conran JG, Li J. 2021. Comparative phylogenetic analyses of Chinese *Horsfieldia* (Myristicaceae) using complete chloroplast genome sequences. J Syst Evol. 59(3):504–514.
- Cai Z, Penaflor C, Kuehl JV, Leebens-Mack J, Carlson JE, dePamphilis CW, Boore JL, Jansen RK. 2006. Complete plastid genome sequences of *Drimys, Liriodendron,* and *Piper*: implications for the phylogenetic relationships of magnoliids. BMC Evol Biol. 6:77.
- Chen X, Yang J, Zhang H, Bai R, Zhang X, Bai G, Dai P, Zhao G. 2019. The complete chloroplast genome of *Calycanthus chinensis*, an endangered species endemic to China. Conserv Genet Resour. 11(1):55–58.
- Darriba D, Taboada GL, Doallo R, Posada D. 2012. jModelTest 2: more models, new heuristics and parallel computing. Nat Methods. 9(8):772.

- Goremykin VV, Hirsch-Ernst KI, Wölfl S, Hellwig FH. 2003. Analysis of the *Amborella trichopoda* chloroplast genome sequence suggests that *Amborella* is not a basal angiosperm. Mol Biol Evol. 20(9):1499–1505.
- Gruenstaeudl M, Gerschler N, Borsch T. 2018. Bioinformatic workflows for generating complete plastid genome sequences—an example from *Cabomba* (Cabombaceae) in the context of the phylogenomic analysis of the water-lily clade. Life. 8(3):25.
- Hansen DR, Dastidar SG, Cai Z, Penaflor C, Kuehl JV, Boore JL, Jansen RK. 2007. Phylogenetic and evolutionary implications of complete chloroplast genome sequences of four early-diverging angiosperms: *Buxus* (Buxaceae), *Chloranthus* (Chloranthaceae), *Dioscorea* (Dioscoreaceae), and *Illicium* (Schisandraceae). Mol Phylogenet Evol. 45(2):547–563.
- Hildebrand M, Hallick RB, Passavant CW, Bourque DP. 1988. Trans-splicing in chloroplasts: the rps 12 loci of Nicotiana tabacum. Proc Natl Acad Sci USA. 85(2):372–376.
- Huang H, Shi C, Liu Y, Mao SY, Gao LZ. 2014. Thirteen *Camellia* chloroplast genome sequences determined by high-throughput sequencing: genome structure and phylogenetic relationships. BMC Evol Biol. 14: 151.
- Jin JJ, Yu WB, Yang JB, Song Y, dePamphilis CW, Yi TS, Li DZ. 2020. GetOrganelle: a fast and versatile toolkit for accurate *de novo* assembly of organelle genomes. Genome Biol. 21(1):241.
- Jin L, Yang J, Liu C, He M, Yan H. 2019. Characterization of the complete plastome of medicinal plant *Saururus chinensis* (Saururaceae). Mitochondrial DNA Part B. 4(2):3206–3207.

- Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol. 30(4):772–780.
- Kim CH. 2007. Chloranthaceae. In: Park CW, editors. The genera of vascular plants of Korea. Seoul, South Korea: Academy Publishing Co.; p. 148–149.
- Kong HZ. 2000a. Systematics of the genus *Chloranthus* Swartz (Chloranthaceae) [Ph.D. dissertation]. Beijing: Institute of Botany, Chinese Academy of Sciences.
- Kong HZ. 2000b. Karyotypes of *Sarcandra* Gardn. and *Chloranthus* Swartz (Chloranthaceae) from China. Bot J Linn Soc. 133(3):327–342.
- Kong HZ, Chen ZD. 2000. Phylogeny in *Chloranthus* Swartz (Chloranthaceae) inferred from sequence analysis of nrDNA ITS region. Acta Bot Sin. 42: 762–764.
- Kong HZ, Chen ZD, Lu AM. 2002. Phylogeny of Chloranthus (Chloranthaceae) based on nuclear ribosomal ITS and plastid trnL-F sequence data. Am J Bot. 89(6):940–946.
- Lee JH, Choi IS, Choi BH, Yang S, Choi G. 2016. The complete plastid genome of *Piper kadsura* (Piperaceae), an East Asian woody vine. Mitochondrial DNA Part A. 27(5):3555–3556.
- Liu X, Liao X, Liu Z, Lan S. 2019. Complete chloroplast genome of *Chloranthus henryi* (Chloranthaceae). Mitochondrial DNA Part B. 4(2): 2964–2965.
- Schattner P, Brooks AN, Lowe TM. 2005. The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs. Nucleic Acids Res. 33(Web Server issue):W686–W689.
- Schmitz-Linneweber C, Williams-Carrier RE, Williams-Voelker PM, Kroeger TS, Vichas A, Barkan A. 2006. A pentatricopeptide repeat protein facilitates the *trans*-splicing of the maize chloroplast *rps12* pre-mRNA. Plant Cell. 18(10):2650–2663.
- Stamatakis A. 2006. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics. 22(21):2688–2690.

- Sun C, Chen F, Teng N, Xu Y, Dai Z. 2021. Comparative analysis of the complete chloroplast genome of seven *Nymphaea* species. Aquat Bot. 170:103353.
- Sun J, Zhang G, Li Y, Chen Y, Zhang X, Tang Z, Wu H. 2016. The complete chloroplast genome sequence of *Chloranthus japonicus*. Mitochondrial DNA Part A. 27(5):3202–3204.
- Tillich M, Lehwark P, Pellizzer T, Ulbricht-Jones ES, Fischer A, Bock R, Greiner S. 2017. GeSeq- versatile and accurate annotation of organelle genomes. Nucleic Acids Res. 45(W1):W6–W11.
- Walker JF, Zanis MJ, Emery NC. 2014. Comparative analysis of complete chloroplast genome sequence and inversion variation in *Lasthenia burkei* (Madieae, Asteraceae). Am J Bot. 101(4):722–729.
- Wang W, Zou P, Liu G, Dai S. 2020. Characterization of the complete chloroplast genome sequence of *Sarcandra glabra* (Chloranthales). Mitochondrial DNA Part B. 5(1):864–865.
- Wang YB, Liu BB, Nie ZL, Chen HF, Chen FJ, Figlar RB, Wen J. 2020. Major clades and a revised classification of *Magnolia* and Magnoliaceae based on whole plastid genome sequences via genome skimming. J Syst Evol. 58(5):673–695.
- Wei XP, Li HJ, Che P, Guo HJ, Zhang BG, Liu HT, Qi YD. 2020. Comparing chloroplast genomes of traditional Chinese herbs Schisandra sphenanthera and S. chinensis. Chin Herb Med. 12(3):247–256.
- Xia N, Jérémie J. 2007. Chloranthaceae Blume. In: Wu ZY, Raven PH, Hong DY, editors. Flora of China. Vol. 4. Beijing: Science Press; p. 132–138.
- Zeng CX, Hollingsworth PM, Yang J, He ZS, Zhang ZR, Li DZ, Yang JB. 2018. Genome skimming herbarium specimens for DNA barcoding and phylogenomics. Plant Methods. 14:43.
- Zheng Y, Luo Y, Li Y, Wang Y. 2019. The complete chloroplast genome sequence of *Cinnamomum longipetiolatum*. Mitochondrial DNA Part B. 5(1):198–199.
- Zhang Q, Antonelli A, Feild TS, Kong H-Z. 2011. Revisiting taxonomy, morphological evolution, and fossil calibration strategies in Chloranthaceae. J Syst Evol. 49(4):315–329.