

Delivery of folic acid-modified liposomal curcumin for targeted cervical carcinoma therapy

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Introduction: In this study, novel folic acid (FA)-modified curcumin (CUR) liposomes (LPs) were developed and evaluated for their antitumor activity in vitro and in vivo.

Methods: Characterization of the LPs, including transmission electron microscopy, morphology, particle size, and zeta potential studies, was carried out. Drug entrapment efficiency, drug-loading capacity, and release properties in vitro were tested. The in vitro growth inhibition activity, cellular uptake efficiency, and cell apoptosis of FA-modified CUR LPs were also investigated by a cervical cancer HeLa cell model.

Results: The optimized distearoyl-l-a-phosphatidylethanolamine (DSPE)-PEG₂₀₀₀-FA-LPs/CUR formed spherical vesicles of nanometer sizes and had particle sizes of 112.3±4.6 nm, polydispersity index of 0.19±0.03, and zeta potential of -15.3±1.4 mV. In addition, the EE% and DL% of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR were 87.6% and 7.9%, respectively. Compared with the free drug, FA-modified CUR LPs had sustained-release properties in vitro. In vivo, a strong green fluorescence was observed in the cytoplasmic region after incubation of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR for 2 hrs.

Conclusion: (DSPE)-PEG₂₀₀₀-FA-LPs/CUR showed a superior antiproliferative effect on HeLa cells and had a better antitumor effect in vivo than the non-modified LPs. These results indicated that (DSPE)-PEG₂₀₀₀-FA-LPs/CUR was a promising candidate for antitumor drug delivery.

Keywords: curcumin, folic acid, liposome, release, cell uptake, antiproliferative, antitumor study, HeLa

Introduction

Cervical cancer is the third most commonly diagnosed cancer worldwide and the fourth leading cause of cancer death in women.¹ In China, there are about 132,000 new cases of cervical cancer each year, accounting for 28% of the total number of new cases of cervical cancer worldwide.² At present, the most effective treatment for cervical cancer is achieved through a combination of cisplatin-based chemotherapy with radiation.³ However, the clinical use of this combination treatment is limited because of several serious side effects, such as nephrotoxicity, neurotoxicity, hematological toxicity, chemoresistance, and off-target damage to normal tissues.⁴ In the early stages of the disease, this treatment regimen would result in a patient survival rate of 90%. However, the figure decreases to 17% among patients at advanced stages⁵ due to the presence of chemoradioresistant cells.⁶ Therefore, ongoing efforts are necessary for the development of new, effective therapeutic strategies to enhance chemotherapeutic efficacy and decrease these side effects, maximizing the local response and patient survival.

Curcumin (CUR) is the chief component of the spice turmeric and is derived from the rhizome of the East Indian plant *Curcuma longa*. *Curcuma longa* is a member of the

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Zingiberaceae (ginger) family of botanicals and is a perennial plant native to Southeast Asia.⁷ It has low intrinsic toxicity and a wide range of pharmacological properties, including antitumor, anti-amyloid, antioxidant, and anti-inflammatory effects. Previous studies have shown that free CUR can induce cell cycle arrest and/or apoptosis and inhibit the activation of nuclear factor kappa B in different tumor cell lines.^{8,9} CUR has been associated with regression of premalignant lesions of the bladder, soft palate, gastrointestinal tract, cervix, and skin, with treatment responses in established malignancy.^{10,11} The only factor limiting the use of free CUR in the treatment of cancer is its poor solubility in water, which in turn limits the bioavailability of the system when administered orally. The reason for the treatment failure of CUR is most likely that the drug concentration in the body cannot reach the effective antitumor level. In addition, nonspecific binding capacity also decreases CUR concentration in tumor tissues.

The application of liposomal formulations as carriers for anticancer drug modified by tumor-active-targeted ligand folate has been trialed and described in the literature in recent years.^{12–15} Folate conjugates can bind to the folate receptor (FR) with high affinity and enter the FR-elevated malignant cells by receptor-mediated endocytosis,^{16,17} and meanwhile avoid being endocytosed by those normal cells that express a low level of FRs.¹⁸ Thus, adding folic acid (FA) to the distal ends of liposome (LP)-anchored PEG molecules can further improve tumor cell selectivity and the uptake efficiency of LPs. However, little research has been done on the pharmacodynamics of FA-modified CUR liposomal formulation.

The aim of this study was to develop a novel FA-modified liposomal formulation that could exhibit efficient and stable encapsulation of CUR and therefore mediate its delivery to the solid tumor effectively and specifically. Characterization of the LPs, including transmission electron microscopy (TEM), morphology, particle size, and zeta potential, was carried out. Drug entrapment efficiency, drug-loading capacity, and release properties *in vitro* were tested. The *in vitro* growth inhibition activity, cellular uptake efficiency, and cell apoptosis of FA-modified CUR LPs were also investigated by a cervical cancer HeLa cell model.

Materials and methods

Materials

CUR was purchased from Changyue Biological Technology Co., Ltd. (Xi'an, China). Distearoyl-L- α -phosphatidylethanolamine (DSPE)-PEG₂₀₀₀-FA was provided by the HuiJia Biopharma Co., Ltd (Xiamen,

China). Soybean phosphatidylcholine (SPC), DSPE-PEG₂₀₀₀, and cholesterol (CHOL) were obtained from Sinopharm Chemical Reagent (Shanghai, China). The human cervical carcinoma HeLa cell line was provided by Second Hospital of Anhui (the Second Hospital of Anhui Medical University) and upon the approval of the ethics committee of the Second Hospital of Anhui Medical University. All other reagents were obtained from Sinopharm Chemical Reagent (Shanghai, China). Methanol and acetonitrile (chromatographic grade) were obtained from Sigma (Aldrich, USA). Deionized water used throughout the research was produced using a Milli-Q System.

Animals

Experiments were carried out on rats weighing 220±20 g and Balb/c mice weighing 20±2 g. The animals were kept in cages in a room at a temperature of 25±2°C, with a 12:12 light–dark cycle. They had free access to food and water. All experiments were carried out in strict accordance with the experimental animal guidelines issued by the National Institutes of Health and approved by the Second Hospital of Anhui Medical University.

Preparation of LPs

FA-modified LPs containing CUR were prepared by the thin-film hydration method as described previously.¹⁹ Briefly, CUR, (DSPE)-PEG₂₀₀₀-FA, SPC, and CHOL (5, 5, 30, and 15, w/w) were dissolved in 5 mL of chloroform to form a mixed solution and then the organic solvent was removed under reduced pressure at 40°C by rotary evaporation to form a thin solid film on the inner walls of the round-bottomed flask. This film was then flushed with nitrogen for 30 mins and stored overnight in a desiccator to remove any traces of chloroform. Glucose and mannitol (1:1, w/w) were dissolved in PBS (pH=7.4). The lipid film was then hydrated with 5 mL of PBS (pH=7.4) at 55°C by rotation (180 rpm×0.5 hr) to form (DSPE)-PEG₂₀₀₀-FA-LPs/CUR (Figure 1). Finally, the (DSPE)-PEG₂₀₀₀-FA-LPs/CUR obtained was freeze-dried into powder. For the preparation of different CUR LPs and DSPE-PEG₂₀₀₀-LPs/CUR, a similar procedure was carried out.

Characterization

In order to determine the optimal parameters relative to the formulation of the (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, drug-loading (DL%), encapsulation efficiency (EE%), particle size, zeta potential, polydispersity index (PDI), and surface

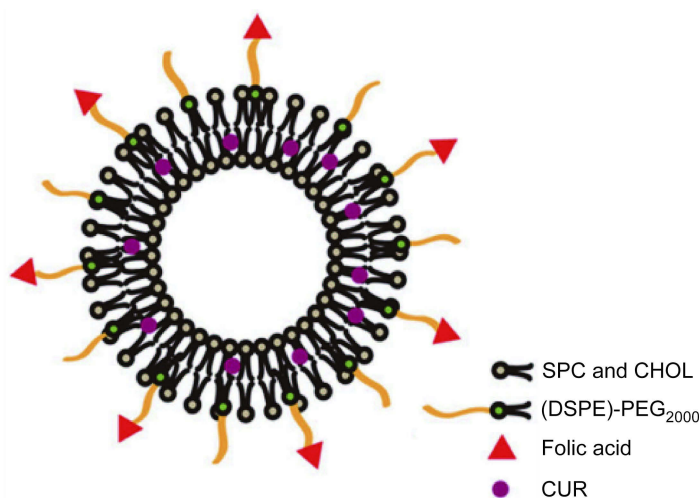


Figure 1 Schematic illustration of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR.

Abbreviations: DSPE, Distearoyl-L- α -phosphatidylethanolamine; SPC, Soybean phosphatidylcholine; CHOL, cholesterol; CUR, Curcumin FA, folic acid; LP, liposome.

morphology of the LPs were measured. Dynamic light scattering (DLS; Zetasizer Nano ZS-90; Malvern Instruments, Malvern, UK) and transmission electron microscope (TEM; Tecnai G2 20; FEI, Memphis, TN, USA) were used for characterizing.

DL% and EE% were calculated as described earlier.²⁰ Firstly, CUR was extracted from LPs (20 mg) with 2 mL 5% glacial acetic acid: acetonitrile (55:45, v/v), and then the extracted solution was properly diluted prior to HPLC analysis. The content of CUR in the LPs was determined by the HPLC method. DL% and EE% were calculated according to Equations (1) and (2), respectively:

$$DL\% = \frac{W_M}{(W_P + W_M)} \times 100\% \quad (1)$$

$$EE\% = \frac{W_M}{W_F} \times 100\% \quad (2)$$

where W_P is the weight of initial feeding polymer, W_M is the weight of drug incorporated in LPs, and W_F is the weight of initial feeding drug.

In vitro drug release

In vitro release of CUR from the LPs was carried out by the dialysis membrane method. (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, (DSPE)-PEG₂₀₀₀-LPs/CUR, CUR LPs, and free CUR (≈ 5 mg CUR) were suspended in 1 mL aliquots of distilled water and placed in dialysis tubing (8,000–12,000 molecular weight cutoff; Greenbird Inc., Shanghai, China). Dialysis tubes containing (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, (DSPE)-PEG₂₀₀₀-LPs/CUR, CUR LPs, and free CUR were then submerged in 200 mL of PBS (pH 7.4) as the release medium (containing ethanol 20%, v/v) at 37°C and immediately

shaken at a speed of 75 rpm. Subsequently, 0.5 mL aliquots of the release medium were sampled at 0.5, 1, 2, 4, 6, 8, 10, 12, and 24 hrs and immediately replaced with equal volumes of fresh release medium. Finally, the samples were analyzed by HPLC. In the release medium, ethanol was used to provide the sink condition because CUR was insoluble in water.

Stability

The stability of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR after storage at 4°C and 25°C for a period of 3 months was analyzed according to the ICH guidelines. The physical appearance, particle size, zeta potential, EE%, and DL% of the samples were analyzed over the 3-month period with an interval of 1 month.

In vitro anticancer activity assay

The in vitro anticancer activity of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR on human cervical carcinoma HeLa cell line was evaluated by MTT assay. Nontoxic control experiments were carried out using a complete growth culture medium. Briefly, cells were seeded into a 96-well plate at a density of 2×10^4 cells per well. After 24 hrs, the cells were incubated with (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, (DSPE)-PEG₂₀₀₀-LPs/CUR, CUR LPs, and free CUR (from 0.002 μ g/mL to 20 μ g/mL) with the same final concentration for 48 hrs. Blank LPs were added at different concentrations as controls. Cells were then incubated with 20 μ L MTT solution (5 mg/mL in PBS) for 4 hrs before the medium was removed and cells were dissolved with 200 μ L DMSO; the absorbance was 570 and 490 nm as measured by a microplate reader (Thermo Fisher

Scientific). Cell viability (%) was calculated according to the following formula:

$$\text{Cell viability}\% = \frac{(A_{\text{test}} - A_{\text{blank}})}{(A_{\text{control}} - A_{\text{blank}})} \times 100\%$$

A_{test} , A_{control} , and A_{blank} represent the absorbance of cells with different treatments, untreated cells, and blank culture media, respectively. Each experiment was performed six times. Half-maximal inhibitory concentration (IC_{50}) was calculated for all samples by survival analysis using GraphPad Prism 5 software.

Cellular uptake

The cellular internalization of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, (DSPE)-PEG₂₀₀₀-LPs/CUR, CUR LPs, and free CUR was visualized by confocal microscopy using coumarin-6 as a fluorescent probe. HeLa cells were plated in six-well plates at 37°C under 5% CO₂ in folate-free RPMI 1640 medium containing 10% FBS. HeLa cells were inoculated in a cell culture dish with 4 × 10⁵ cells per dish as the initial density. Cells were then incubated with coumarin-6-adsorbed free CUR, (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, (DSPE)-PEG₂₀₀₀-LPs/CUR, and CUR LPs (equivalent to 0.1 µg/mL of coumarin-6). After incubation at 37°C for 2 hrs, the cells were washed three times with cold PBS to remove unbound LPs and then observed under a confocal microscope.

For CUR uptake quantitative estimates, the density of cells inoculated on 24-well plates was 3 × 10⁴. When 70–80% confluence was achieved, cells were incubated with coumarin-6-adsorbed free CUR, (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, (DSPE)-PEG₂₀₀₀-LPs/CUR, and CUR LPs (equivalent to 0.1 µg/mL of coumarin-6). After 2 hrs of culture, cells were washed several times with cold PBS. Subsequently, cells were dissolved by addition of Triton X-100 (0.1%). Fluorescence intensities were measured by a multimode microplate reader at an excitation wavelength of 440 nm and an emission wavelength of 520 nm.

Pharmacodynamics

Tumor reduction studies were conducted as reported earlier.²² Female Balb/c mice aged 6–8 weeks were used for the experiment. Tumors were induced by subcutaneous injection of HeLa cells (~5 × 10⁶ cells in 100 µL PBS) in the lower right flank of mice and were allowed to grow for a period of time (approximately 15 days) to a size of approximately 100–150 mm³ as measured by vernier calipers. The animals were then randomly divided into 5

groups of 8 and blank LPs, free CUR, CUR LPs, (DSPE)-PEG₂₀₀₀-LPs/CUR, and (DSPE)-PEG₂₀₀₀-FA-LPs/CUR (equivalent to 25 mg/kg CUR, on alternate days for three weeks) were injected intraperitoneally. The tumor volume was measured every 3 days. Animals were sacrificed after 51 days and the main organs and tumor samples were collected for further analyses. The specimens were fixed with 4% paraformaldehyde and then paraffin embedded for H&E staining and histological examination.

Statistical analysis

All data points were denoted as mean ± SD. A two-tailed *t*-test or a one-way ANOVA was performed, respectively, when comparing two or more groups. Statistical significance was defined as *P* < 0.05. Data were shown as mean ± SD.

Results and discussion

Preparation and characterization

In our current study, we designed FA-mediated stabilized liposomes encapsulating CUR to improve the therapeutic effect of CUR in the treatment of cervical cancer via the combination of active targeting and molecular targeting. A systematic approach was undertaken to develop and characterize preparation methods and liposome formulations in order to control drug leakage and thus maintain optimal drug loading inside the liposomes to increase the delivery of the drug to tumor tissues. CUR was encapsulated into FA-modified and nonmodified LPs using the thin-film hydration method. The obtained physicochemical properties of the formulations are summarized in Table 1. The TEM images and DLS size distribution of the (DSPE)-PEG₂₀₀₀-FA-LPs/CUR are shown in Figure 2. The optimized (DSPE)-PEG₂₀₀₀-FA-LPs/CUR formed spherical vesicles at nanometer sizes and had particle sizes of 112.3 ± 4.6 nm, PDI of 0.19 ± 0.03, and zeta potential of -15.3 ± 1.4 mV. The negative zeta potential might have resulted from the presence of negatively charged lipid DSPE-PEG₂₀₀₀ in the formulation.²¹ In addition, the EE% and DL% of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR were 87.6% and 7.9%, respectively. Under the same loading conditions, ordinary CUR LPs and nonmodified LPs appeared to have higher EE% and DL% and smaller particle sizes; however, no relevant differences were observed in terms of these parameters among the three CUR LPs. In order to determine the influence of the CUR/lipid incubation ratio on loading, several ratios (1:5, 1:9, 1:18, 1:30, w:w) were tested on (DSPE)-PEG₂₀₀₀-FA-LPs/CUR. While a CUR/lipid incubation ratio of 1:9 or lower provided excellent EE% values (>90%), only the

Table 1 Physicochemical properties of CUR LPs; DSPE-PEG₂₀₀₀-LPs/CUR and (DSPE)-PEG₂₀₀₀-FA-LPs/CUR (n=3)

Formulations	Particle size (nm)	Zeta potential (mV)	Polydispersity index	Encapsulation efficiency (%)	Drug loading (%)
CUR LPs	106.8±7.2	+18.1±1.6	0.18	85.4±7.3	8.1±1.4
DSPE-PEG ₂₀₀₀ -LPs/CUR	109.4±5.1	-19.9±1.3	0.17	89.1±4.2	8.4±1.1
(DSPE)-PEG ₂₀₀₀ -FA-LPs/CUR	112.3±4.6	-15.3±1.4	0.19	87.6±6.3	7.9±0.7

Abbreviations: DSPE, Distearoyl-L-α-phosphatidylethanolamine; SPC, Soybean phosphatidylcholine; CHOL, cholesterol; CUR, Curcumin FA, folic acid; LP, liposome.

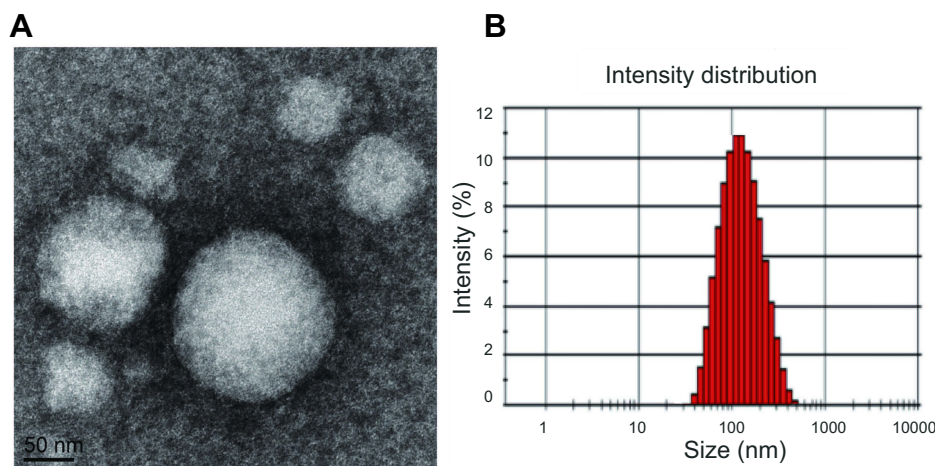


Figure 2 The characterization of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR. **Notes:** (A) The transmission electron microscope images; (B) the size distribution. **Abbreviations:** DSPE, Distearoyl-L-α-phosphatidylethanolamine; CUR, Curcumin FA, folic acid; LP, liposome.

incubation ratio of 1:9 provided the highest DL% among all the formulations.

Stability

The stability of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR was monitored by changes in physical appearance, particle size, zeta potential, EE%, and DL% during storage at 4°C and 25°C (Table 2). The zeta potential exhibited little change over the 3-month storage, showing the stability of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR. The results also indicated that during storage at 4°C or 25°C, no significant variation in physical appearance,

particle size, EE%, and DL% was detected. It appears that the liposomal formulation showed excellent stability during storage, and no drug precipitation or liposome aggregation was observed throughout the 90 days.

In vitro drug release

CUR release in vitro was also investigated using the dialysis method in PBS (pH 7.4) at 37°C for 24 hrs to examine the drug-release properties of LPs (Figure 3). Unlike the rapid release of free CUR, as predicted, three LP types, (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, (DSPE)-PEG₂₀₀₀-LPs/CUR, and

Table 2 Stability of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR after storage at 4°C and 25°C for a period of 3 months (n=3)

Temperature	Time (m)	Physical appearance	Particle size (nm)	Zeta potential (mV)	Polydispersity index	Encapsulation efficiency (%)	Drug loading (%)
25°C	0	Cream yellow	112.3±4.6	-15.3±1.4	0.19	87.6±6.3	7.9±0.7
4°C	1	Cream yellow	112.9±5.4	-15.6±1.3	0.18	86.9±5.9	7.8±1.1
	2	Cream yellow	113.6±4.7	-15.7±1.4	0.20	87.1±5.7	7.8±1.2
	3	Cream yellow	114.6±5.1	-16.5±1.7	0.19	86.5±5.9	7.6±1.2
25°C	1	Cream yellow	113.3±4.2	-16.1±1.4	0.19	86.8±5.7	7.9±1.3
	2	Cream yellow	114.6±4.6	-16.5±1.5	0.21	86.6±6.1	7.7±1.2
	3	Cream yellow	114.9±3.7	-16.3±1.3	0.22	86.2±5.9	7.7±1.1

Abbreviations: DSPE, Distearoyl-L-α-phosphatidylethanolamine; CUR, Curcumin FA, folic acid; LP, liposome; m, months.

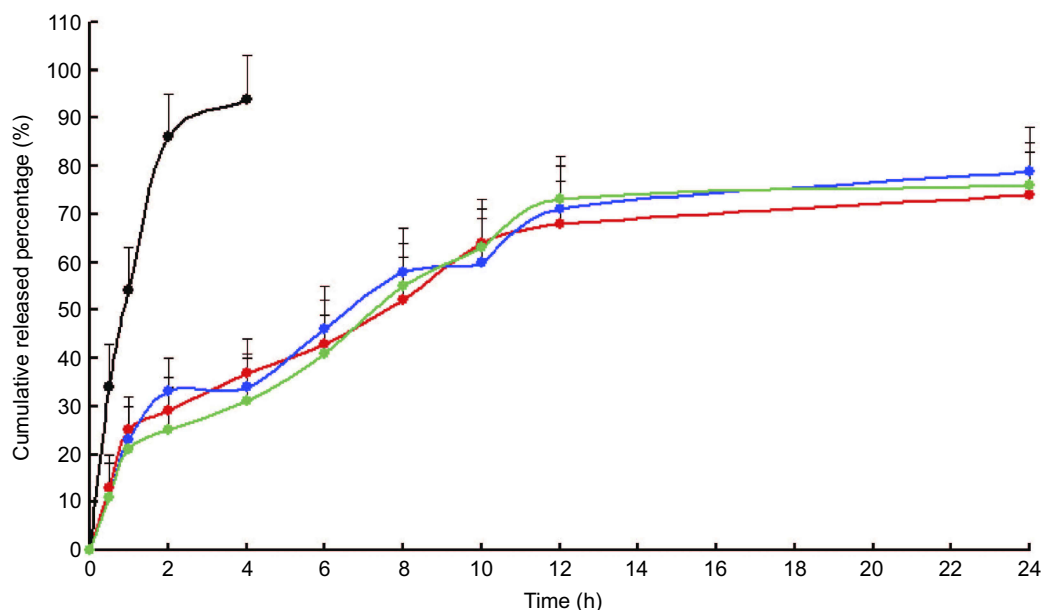


Figure 3 In vitro study of CUR release for free CUR (black), CUR LPs (blue), (DSPE)-PEG₂₀₀₀-LPs/CUR (green), and (DSPE)-PEG₂₀₀₀-FA-LPs/CUR (red) in PBS (pH=7.4) over 24 hrs (n=6).

Abbreviations: DSPE, Distearoyl-L- α -phosphatidylethanolamine; CUR, Curcumin FA, folic acid; LP, liposome.

CUR LPs, exhibited similar gradual release properties under pH 7.4. Over time, all formulations of CUR LPs were released much more slowly than free drug. After 2 hrs, the cumulative release was about 86% in the free drug (CUR) and 30% in LPs. A biphasic release was observed in this case with a rapid release of about 30% in 2 hrs followed by sustained drug release of about 70% over 12 hrs. It suggested that in comparison with the free drug, (DSPE)-PEG₂₀₀₀-FA-LPs/CUR have sustained release properties. However, there was no significant difference among the three LPs in the release curve.

The (DSPE)-PEG₂₀₀₀-FA-LPs/CUR was surface-coated with the hydrophilic polymer PEG, which formed a hydrophilic barrier to prevent the accumulation of LPs and to improve their physical stability. In addition, phospholipid, the main ingredient used in this experiment, is a corrosive substance. Because of the poor water solubility of the drug, it is difficult for it to penetrate into the matrix of the LPs. The sustained release of the drug might mainly contribute to the corrosion rates of the LPs prepared with these phospholipids. After calculation, the in vitro drug-release kinetic model of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR in release medium was found to fit well with the Higuchi equation: $Q=9.523t^{1/2}-1.231$ ($r=0.993$). Thus, it was speculated that the sustained-release property of LPs could assist in decreasing drug

leakage before drug-loaded systems arrived at the targeted tumors and cells.

In vitro anticancer activity assay

For anticancer effects in vitro, HeLa cells were treated with either (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, (DSPE)-PEG₂₀₀₀-LPs/CUR, CUR LPs, or free CUR at different concentrations of the drug. Cells were then subjected to an MTT assay. The cell viability profile is presented in Figure 4. As shown, a significant reduction in cell viability was observed in the (DSPE)-PEG₂₀₀₀-FA-LPs/CUR group, while the cytotoxicity of cells was clearly lower in the free CUR and (DSPE)-PEG₂₀₀₀-LPs/CUR groups. The negative control was represented by blank LPs, for which cell survival was 100%. From this result, we found that 1) the carrier material used in this study was safe and nontoxic; 2) as the CUR concentration increased, greater cell death was observed; 3) (DSPE)-PEG₂₀₀₀-FA-LPs/CUR showed a superior antiproliferative effect on HeLa cells, which may be expected to demonstrate obvious antitumor effect in vivo. After calculation, the IC₅₀ value of the (DSPE)-PEG₂₀₀₀-FA-LPs/CUR was found to be 0.45 μ g/mL compared with a value of 0.82 μ g/mL for (DSPE)-PEG₂₀₀₀-LPs/CUR, 1.47 μ g/mL for free CUR, and 1.21 μ g/mL for CUR LPs (Figure 4). Blank LPs were tested as controls and did not induce significant cell death (data not shown).

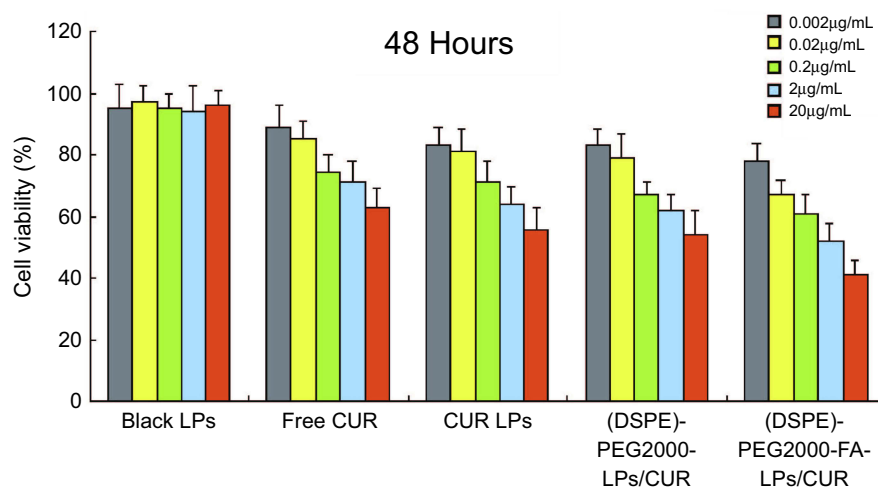


Figure 4 The cytotoxicity study of blank LPs, free CUR, and CUR LPs, (DSPE)-PEG₂₀₀₀-LPs/CUR and (DSPE)-PEG₂₀₀₀-FA-LPs/CUR in HeLa cells (n=3).

Abbreviations: DSPE, Distearoyl-L- α -phosphatidylethanolamine; CUR, Curcumin FA, folic acid; LP, liposome.

Cellular uptake

Free CUR, CUR LPs, (DSPE)-PEG₂₀₀₀-LPs/CUR, and (DSPE)-PEG₂₀₀₀-FA-LPs/CUR were labeled by a fluorescent probe. As shown in Figure 5, the nucleus displayed green fluorescence, which was stained by coumarin-6. A strong green fluorescence was observed in the perinuclear region of HeLa cells treated with (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, indicating that a sufficient amount of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR entered the cytoplasm. In addition, the fluorescence intensity of the (DSPE)-PEG₂₀₀₀-LPs/CUR group was

weaker than that of the (DSPE)-PEG₂₀₀₀-FA-LPs/CUR group.

Moreover, HeLa cells pretreated with free CUR and CUR LPs also exhibited mild green fluorescence, likely attributable to weak ability to cross cells. The cellular uptake ratio of the LPs was detected by a multimode microplate reader to be $5.7 \pm 1.6\%$, $16.5 \pm 4.9\%$, $38.4 \pm 4.4\%$, and $66.4 \pm 6.2\%$ for free CUR, CUR LPs, (DSPE)-PEG₂₀₀₀-LPs/CUR, and (DSPE)-PEG₂₀₀₀-FA-LPs/CUR-treated HeLa cells, respectively. These results demonstrated that the FA-modified strategy can facilitate the high-efficiency uptake of CUR by HeLa cells.

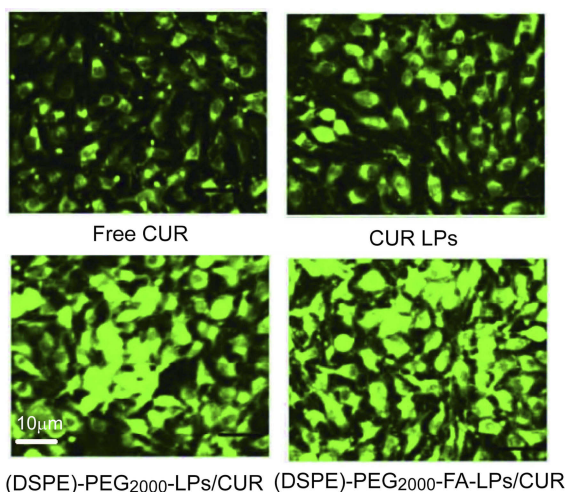


Figure 5 Uptake of Free CUR, CUR LPs, (DSPE)-PEG₂₀₀₀-LPs/CUR, and (DSPE)-PEG₂₀₀₀-FA-LPs/CUR by HeLa cells. Fluorescence intensities were measured by a multimode microplate reader at an excitation wavelength of 440 nm and an emission wavelength of 520 nm.

Abbreviations: DSPE, Distearoyl-L- α -phosphatidylethanolamine; CUR, Curcumin FA, folic acid; LP, liposome.

Pharmacodynamics

To assess the in vivo antitumor efficacy of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, human cervical cancer xenograft mice were employed. As shown in Figure 6, all the treatment groups were better able to inhibit tumor growth and decrease tumor size compared with the control group. At the end of the experiment, tumors were found to be more sensitive to (DSPE)-PEG₂₀₀₀-FA-LPs/CUR at a volume of $77.3 \pm 56.5 \text{ mm}^3$, compared to that of the control group ($987.6 \pm 89.4 \text{ mm}^3$), free CUR ($634.3 \pm 67.4 \text{ mm}^3$), CUR LPs ($432.7 \pm 52.6 \text{ mm}^3$), and (DSPE)-PEG₂₀₀₀-LPs/CUR ($232.6 \pm 57.2 \text{ mm}^3$). The results indicated a significant effect on the reduction rate of tumor volume for (DSPE)-PEG₂₀₀₀-FA-LPs/CUR, which was statistically higher than other groups. The results mainly concerned the following aspects: 1) the effect of EPR (confirmed by another biodistribution study; data not shown); 2) enhancing intratumoral retention of CUR in terms of (DSPE)-PEG₂₀₀₀-FA-LPs/CUR;

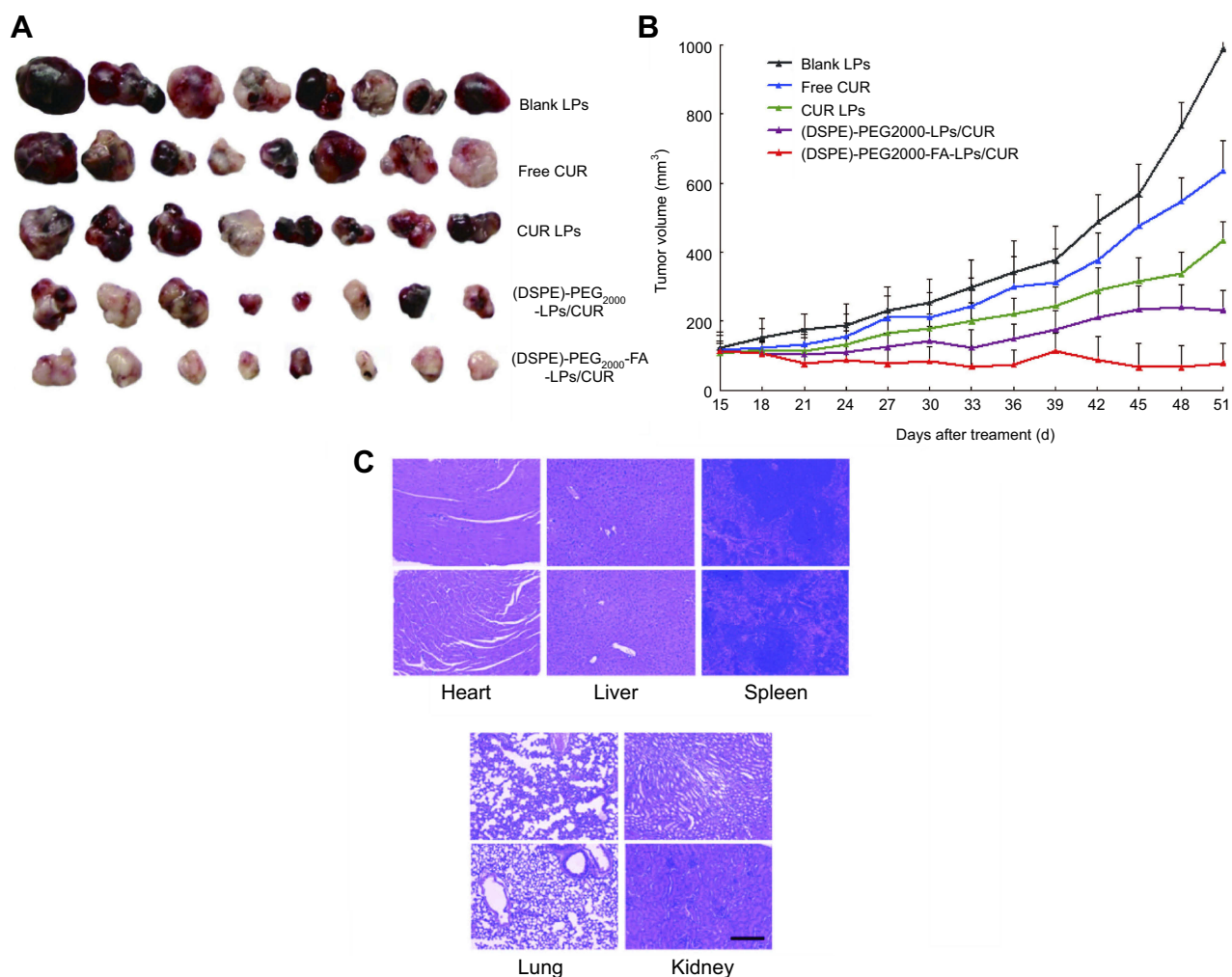


Figure 6 The antitumor activity of blank LPs, free CUR, CUR LPs, (DSPE)-PEG₂₀₀₀-LPs/CUR, and (DSPE)-PEG₂₀₀₀-FA-LPs/CUR on HeLa tumor-bearing mice. **(A)** Representative images of mice bearing HeLa xenograft tumors, after 51 days of treatment. **(B)** Tumor volume growth curves. **(C)** Histopathological studies of heart, liver, spleen, lung, and kidney.

Abbreviations: DSPE, Distearoyl-L-a-phosphatidylethanolamine; CUR, Curcumin FA, folic acid; LP, liposome.

3) receptor ligand-mediated targeted delivery. H&E staining was used to determine whether there was organ damage. The results showed that there were no obvious lesions in heart, liver, spleen, lung, and kidney of the two groups, suggesting that (DSPE)-PEG₂₀₀₀-FA-LPs/CUR had no obvious acute toxicity in vivo (Figure 6C).

We prepared a delivery system of FA-mediated liposomes with encapsulated CUR in this study. According to the results, the drug delivery system in our study was in an appropriate nanoscale size range, exhibited specific targets to tumor cells, and induced more cell death and cell apoptosis. These results suggested that (DSPE)-PEG₂₀₀₀-FA-LPs/CUR possessed the basic characteristics of targeted delivery systems and had the potency to improve the antitumor effects of CUR.

Disclosure

The authors report no conflicts of interest in regard to this work.

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