Effect of efavirenz-based ART on the pharmacokinetics of rifampicin and its primary metabolite in patients coinfected with TB and HIV

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Objectives: To evaluate the effects of concomitant efavirenz-based ART and genetic polymorphism on the variability in rifampicin and 25-desacetylrifampicin pharmacokinetics.

Patients and methods: Plasma concentrations of rifampicin and 25-desacetylrifampicin from 63 patients coinfected with TB and HIV were analysed by LC-MS/MS followed by non-linear mixed-effects modellina. Patients were aenotyped for SLCO1B1 (463 C>A, 388 A>G, 11187 G>A, rs4149015, 521 T>C and 1436 G>C) and SLCO1B3 (334 T>G).

Results: One-compartment disposition models described the observations adequately. The oral clearances of rifampicin and 25-desacetylrifampicin were 140% and 110% higher, respectively, in patients on concomitant efavirenz-based ART. Rifampicin bioavailability was also lower in patients on concomitant ART. Further, although not included in the final model, a lower relative bioavailability in carriers of WT SLCO1B3 334 T>G compared with carriers of mutations in the genotype was estimated.

Conclusions: The results presented indicate both pre-systemic and systemic induction by efavirenz-based ART affecting rifampicin pharmacokinetics. The described drug-drug interaction has a clinical impact on rifampicin exposure prior to steady state and may impact the early bactericidal activity in patients on efavirenz-based ART.

Introduction

Approximately one-third of all HIV-associated deaths are due to TB, making TB the most common cause of death in HIV-infected individuals.¹ Rifampicin is a key sterilizing drug in the currently used first-line TB therapy. A rifampicin peak plasma concentration (C_{max}) of >8 mg/L has been suggested as a target therapeutic concentration.² The WHO recommends an adjusted daily dose of 10 mg/kg. However, due to the pharmacokinetic variability and exposure-dependent efficacy of rifampicin, doses up to 35 mg/kg are currently being investigated.³ Rifampicin is primarily metabolized to the active metabolite 25-desacetylrifampicin by beta esterases.⁴ Unknown elimination pathways of rifampicin are autoinduced and an up to 2-fold increase in oral clearance has been described at steady state compared with at initiation of antitubercular therapy.⁵ Furthermore, rifampicin undergoes hepatic first-pass extraction and exhibits a non-linear relationship between dose and exposure.^{6,7} In addition, polymorphism in SLCO1B1 genes coding for organic anion-transporting polypeptide 1B1 has been suggested to affect rifampicin exposure.⁸

Efavirenz-based ART is recommended as an alternative firstline treatment for HIV. Efavirenz is a known inducer of metabolic

enzymes.⁹ Hence, there is a risk of drug-drug interactions affecting rifampicin pharmacokinetics during co-administration when concomitantly treating TB and HIV. Although an effect of HIV infection on rifampicin pharmacokinetics has been previously described,¹⁰ HIV treatment has not shown any effect on pharmacokinetic parameters of rifampicin.¹¹⁻¹³ However, in those studies, HIV treatment was initiated at steady state of antitubercular therapy and genetic correlations were not evaluated. Thus, this study aimed to investigate the simultaneous effects of enzymatic polymorphisms and concomitant efavirenz-based ART on the pharmacokinetics of rifampicin and 25-desacetylrifampicin.

Methods

Patients and study design

An open-label observational clinical study was performed in four clinics in Rwanda. Patients were either HIV-treatment naive (arm A) or treated for HIV (arm B) when TB therapy was initiated. Patients in each clinic were offered to participate in the study if inclusion criteria were met. Inclusion criteria were: 21-65 years of age, HIV-antibody positive, TB-drug naive and clinical diagnosis of TB. TB and HIV were managed according to domestic

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guidelines.¹⁴ Patients who were HIV-treatment naive were initiated on ART following 2–8 weeks of TB therapy depending on patient status.

Rifampicin-based fixed-dose combinations (FDCs) containing rifampicin, isoniazid, pyrazinamide and ethambutol at 150/75/400/275 mg (Svizera, Almere, The Netherlands) were administered to study participants. Weight-band-based doses were used as follows: ≤ 28 kg, one and a half tablets; 29–37 kg, two tablets; 38–54 kg, three tablets; 55–70 kg, four tablets; and ≥ 71 kg, five tablets (in order to administer a rifampicin dose of approximately 10 mg/kg). In addition to the FDCs, eight patients were administered streptomycin.

Ethics

The clinical study was performed in agreement with the Helsinki Declaration and International Conference on Harmonization guidance for Good Clinical Practice. Approval to perform the study was received from the National Ethics Committee of the Ministry of Health in Rwanda (IRB 00001497). Patients provided a signed written informed consent and were informed that participation was voluntary and withdrawal was accepted at any time during the study.

Dose intake and sample collection

Blood samples were collected prior to the first dose of rifampicin and at 1, 2, 3, 4, 6 and 8 h after the first dose. Plasma was harvested by centrifugation of the collected samples and then stored at -30° C for 1 week in clinics before being transferred to -80° C for long-term storage. Due to the observational nature of the study, no instructions regarding food intake in proximity to dose or during sample collection were specified in the study protocol. Blood samples to perform genotyping of SNPs were collected separately.

Bioanalytics

Rifampicin and 25-desacetylrifampicin plasma concentrations were determined by LC-MS/MS.¹⁵ The method was validated according to FDA guidelines¹⁶ at a concentration range of 200–25 000 and 40–5000 ng/mL for rifampicin and 25-desacetylrifampicin, respectively. Intra-day accuracy and precision for rifampicin and 25-desacetylrifampicin were 92%–105% and <11% relative standard deviation, respectively.

Genotyping

A QIAamp DNA Blood Mini Kit was used to perform genomic DNA extraction from blood samples for genotyping of SLCO1B1 (463 C>A, 388 A>G, 11187 G>A, rs4149015, 521 T>C and 1436 G>C) and SLCO1B3 (334 T>G). Genotyping was performed by multiplexed primer extension chemistry of an iPLEX assay with detection of the incorporated allele by MS with a MassARRAY analyser (Agena Bioscience, San Diego, CA, USA).^{17,18} The output data were converted into genotype data using Typer software (Agena Bioscience).

Pharmacokinetic analysis

Modelling was carried out in NONMEM software, version 7.4.3 (ICON Development Solutions, Ellicott City, MD, USA).¹⁹ Models were fitted to observations using first-order conditional estimation with interaction. Observations below the lower limit of quantification (n = 24) were ignored. Perl-Speaks-NONMEM (version 4.8.1) and Pirana (version 2.9.8) were used for model automation and tracking of model development. R (version 3.5.1) with packages mrgsolve (version 0.10.1) and nonmem2R (version 0.2.1) was used for simulations with the final model and to create diagnostic plots, respectively.

A structural model for rifampicin was developed prior to adding 25desacetylrifampicin observations. One- and two-compartment disposition models with first-order absorption and elimination were applied to rifampicin observations. Discrimination between nested models was based on the objective function value (OFV). A difference in OFV is considered χ^2 distributed and a decrease in OFV (Δ OFV) of -3.84 and -10.83 is equivalent to a model improvement at a significance level of P < 0.05 and P < 0.001, respectively. Different absorption models were applied in the model, including lag-time and transit-compartment models with either a fixed or estimated number of transit compartments.²⁰ Allometric scaling by body weight was applied to all clearance and volume parameters by a power of 0.75 and 1, respectively.^{21,22}

One- and two-compartment disposition models were applied to 25desacetylrifampicin observations. Rifampicin was assumed to be eliminated via two different metabolic pathways out of which one resulted in the formation of 25-desacetylrifampicin. A semi-mechanistic model including a liver compartment was evaluated to estimate the intrinsic clearance of rifampicin.¹² The hepatic volume, blood flow and fraction of unbound rifampicin plasma concentrations were then fixed to 2 L, 90 L/h and 0.2, respectively.^{23,24} Inter-individual variability was evaluated on all estimated structural parameters as exponential random effects. Additive, proportional or combined residual errors were assessed separately for rifampicin and 25-desacetylrifampicin.

Covariates were tested by stepwise inclusion followed by stepwise elimination. Covariates were included and retained in the model at a significance level of 0.05 and 0.001, respectively. Continuous covariates tested were ALT, AST, age, serum creatinine, creatinine clearance as determined by the Cockcroft and Gault equation²⁵ and CD4 cell count. The continuous covariates were centred on their respective medians. Categorical covariates tested were sex, concomitant HIV therapy (study arm), study site and SLCO1B1 (463 C>A, 388 A>G, 11187 G>A, rs4149015, 521 T>C and 1436 G>C) and SLCO1B3 (334 T>G) genotypes. Individuals with missing genotypes (n = 7) were categorized as a separate group in the initial screening of genotype covariates. Three approaches were evaluated for statistically significant covariates: including patients with missing genotypes as a separate category; assigning all patients with missing genotypes to the most common genotype; or using mixture models to assign patients with missing genotypes to different groups.²⁶ Differences in parameter estimates, OFV and plausibility of the estimated covariate effect were evaluated.

The final model was evaluated by parameter plausibility, goodness-offit plots and a prediction corrected visual predictive check (pcVPC) (n = 1000). A bootstrap (n = 200) was performed to calculate parameter precisions and CIs of the final model.

Stochastic simulations using the final model were performed to evaluate rifampicin exposure following a standard dose (10 mg/kg) and a high dose (35 mg/kg) depending on included covariates. Fixed parameters from a previously described model were used to account for the rifampicin non-linear increase in exposure with higher dose when evaluating highdose rifampicin.⁵ The non-linear increase in exposure with higher dose was applied on the bioavailability (F) in the model according to the following equation:

$$F = 1 + \frac{0.504 \times (\text{dose} - 450)}{67 + (\text{dose} - 450)}$$

where 0.504 is the maximal increase in F and 67 (mg) is the increase in dose above 450 mg corresponding to 50% of the maximal increase in F.

Results

Pharmacokinetic analysis

In total, 874 concentration-time observations were included in the present analysis (raw data are show in Figure 1). Out of the 63 patients included, 56 were genotyped. Patient characteristics are



Figure 1. Plasma concentration-time profiles of rifampicin and its metabolite 25-desacetylrifampicin in patients coinfected with HIV and TB following the first dose of rifampicin. Patients were either on concomitant efavirenz-based ART (+ART) or HIV-treatment naive (No ART).

Table 1	L. Dem	ographics	of	adult	Rwandan	coinfe	ted	TB/HIV	patients
concurr	ently tr	eated for	oot	h diseo	ases or onl	y treate	d fo	r TB	

	Concurrent HIV treatment ^a	HIV-treatment naive
Number of patients	23	40
Age (years)	40 (26–57)	38 (21–52)
Weight (kg)	48 (35–65)	50 (30–68)
Serum creatinine (µmol/L)	71 (44–159)	66 (35–159)
Creatinine clearance (mL/min)	81 (30–155)	84 (38–155)
AST (U/mL)	33 (11-248)	34 (11–131)
ALT (U/mL)	36 (9–126)	30 (5-101)
CD4 cell count (cells/mm ³)	230 (21–716)	240 (6–524)
Sex (female/male)	10/13	16/24
Rifampicin dose (mg/day)	300 (n=1); 450	300 (n=4); 450
	(n=13); 600 (n=9)	(n=22); 600 (n=14

Continuous data given as median (range).

Categorical data given as counts.

^aHIV treatment includes efavirenz, lamivudine and zidovudine or tenofovir.

summarized in Table 1 and genotype frequencies are summarized in Table 2. The dispositions of rifampicin and 25-desacetylrifampicin were most adequately described by one-compartment models. The absorption was described using a transit compartment model (n = 1) with an absorption rate constant equal to the transit rate. A pcVPC to illustrate the predictive properties of the model is shown in Figure 2. Whereas the observed median rifampicin concentrations were in the upper range of its 95% CI, the model overall described the data adequately.

The oral clearance of rifampicin was 2.4-fold higher in patients on concomitant treatment with efavirenz-based ART (Δ OFV = -12.9) compared with patients who were ART naive. The AUC_{25-desacetylrifampicin}/AUC_{rifampicin} ratio (non-compartmental analysis) was higher in patients on concomitant ART (0.14) compared with the patients who were ART naive (0.11) (*P* = 0.008, Wilcoxon test). Adding an effect of ART on 25-desacetylrifampicin clearance (2.1-fold increase) further reduced OFV by -13.0.

The relative bioavailability was significantly lower in patients on concomitant ART ($\Delta OFV = -24.8$). Inter-individual variability in bioavailability was reduced from 114% to 82% when the effects of ART were added. Additionally, carriers of WT SLCO1B3 334 T>G had a lower relative bioavailability (-0.33) compared with carriers of the heterozygous (T/G) or homozygous (G/G) mutation ($\Delta OFV = -7.8$). A mixture model predicted all individuals with missing genotypes to be carriers of the SLCO1B3 334 T>G WT variant. Estimating separate effects for T/G carriers and G/G carriers did not result in a different OFV. Therefore, a combined effect for carriers of a mutation in the SNP was estimated. However, the effect of SLCO1B3 was excluded from the final model due to inadequate statistical significance. Parameter estimates and precisions of the final model are shown in Table 3.

Simulations

The final model was used to simulate exposures (AUC_{0-24h} and C_{max}) following the currently used dose (10 mg/kg) and high dose (35 mg/kg) of rifampicin. Simulations were stratified on whether

Table 2. Distribution of SLCO1B1 and SLCO1B3 SNPs in Rwandan patients coinfected with TB and HIV ($n\,{=}\,56^{\circ})$

Genotype	Allele	Number (%)
SLCO1B1		
463 C>A	C/C	53 (95)
	C/A	3 (5)
388 A>G	A/A	3 (5)
	A/G	15 (27)
	G/G	38 (68)
11187 G>A	G/G	48 (86)
	G/A	8 (14)
rs4149032	T/T	27 (48)
	T/C	21 (38)
	C/C	8 (14)
521 T>C	T/T	46 (82)
	T/C	10 (18)
1436 G>C	G/G	51 (91)
	G/C	5 (9)
SLCO1B3		
334 T>G	T/T	24 (43)
	T/G	19 (34)
	G/G	13 (23)

^aNumber of individuals with missing genotypes = 7.

the patients were on concomitant ART or ART naive. The simulations predicted the majority of patients administered doses of 10 mg/kg in both groups to have a $C_{\rm max}$ below the value suggested to achieve an adequate therapeutic effect.² In patients on efavirenz-based ART, 35 mg/kg doses resulted in $C_{\rm max}$ values above the recommended threshold for a large proportion of the simulated patients. Figure 3 depicts simulated plasma concentration-time profiles following a 10 mg/kg or 35 mg/kg dose, depending on administration of efavirenz-based ART.

Simulations of exposure for a typical individual weighing 50 kg (the median weight in the dataset) predicted a mean AUC_{0-24h} of 76 mg·h/L in ART-naive patients and 19 mg·h/L in patients on ART for doses of 10 mg/kg (Table 4). For doses of 35 mg/kg, an AUC_{0-24h} of 455 mg·h/L and 106 mg·h/L was predicted in ART-naive patients and patients on ART, respectively.

Discussion

A population pharmacokinetic model for rifampicin and 25-desacetylrifampicin was developed. The model was used to evaluate the effects of concomitant ART and genetic polymorphism on the pharmacokinetic parameters of rifampicin and its major metabolite. Rifampicin oral clearance was more than 2-fold higher in patients on concomitant ART compared with patients who were ART naive. Mechanistically, the findings could be explained by induction of hepatic metabolic enzymes by ART. Interestingly, the higher rifampicin clearance in patients on concomitant ART is close to the approximately 2-fold increase in rifampicin clearance after autoinduction observed in other studies.^{5,12} Previous studies have suggested that rifampicin concentrations are similar between patients on concomitant efavirenz-based ART and patients who are ART naive. If rifampicin and efavirenz-based ART induce the same pathway, the autoinduction could have masked such an interaction in previous studies since plasma concentrations have been measured at steady state of antitubercular therapy.^{11–13}

Polymorphism in SLCO1B1 has previously been suggested to affect rifampicin pharmacokinetics.^{8,27} In the present cohort, an effect of SLCO1B3 334 T>G on relative bioavailability was observed. The covariate was not included in the final model due to inadequate statistical significance and lack of prior knowledge on the effect of SLCO1B3 polymorphism on rifampicin pharmacokinetics. However, if the effect is confirmed, the relatively large difference in exposure may make SLCO a suitable factor for individualized therapy. Food intake affects rifampicin exposure and may partly have contributed to some of the variability in bioavailability.^{28,29} However, since food intake was not documented in the clinical study, potential effects of food were not evaluated in the present analysis.

The lower bioavailability in patients on ART is similar to the decrease in rifampicin bioavailability (from 93% to 68%) during the first week of rifampicin administration reported by Loos et al.⁶ The authors suggested a time-dependent increase in pre-hepatic metabolism of rifampicin, which was maximized after 1 week. In the present model, pre-hepatic and hepatic first-pass effects were estimated as total relative bioavailability, which may contribute to the estimated higher effect of ART on biogvailability compared with pre-hepatic autoinduction by rifampicin.⁶ A semi-mechanistic model including a hepatic compartment to separate pre-hepatic bioavailability from the first-pass effect was evaluated. However, for such a model, 25-desacetylrifampicin would have to be assumed to be eliminated either via a hepatic route or via a nonhepatic route since two separate pathways would be unidentifiable. No such assumptions are required for an empirical model, which was therefore considered more suitable since esterases responsible for 25-desacetylrifampicin formation are present in both the liver and the blood.

Efavirenz induces CYP3A4 and P-glycoprotein, both of which may affect the bioavailability of drugs.³⁰ The turnover half-life of P-glycoprotein has been estimated to be 5–17 h, in contrast to the 70 h turnover half-life of CYP3A4.^{31,32} The reduction in bioavailability maximized after 1 week in the study by Loos *et al.*⁶ may plausibly be caused by induction of P-glycoprotein due to its shorter turnover half-life. Hence, the reduced bioavailability of rifampicin in our study could be explained by intestinal P-glycoprotein, which is fully induced following 1 week of efavirenz-based ART. P-glycoprotein may further be saturated at higher doses of rifampicin. Such a hypothesis is supported by higher serum rifampicin concentrations following inhibition of intestinal P-glycoprotein.³³

Simulations predicted an average AUC_{0-24h} of 76 mg·h/L following the first dose of rifampicin in ART-naive patients. The prediction is well in line with a recent meta-analysis of rifampicin pharmacokinetics averaging a first-dose AUC of 72.6 mg·h/L.³⁴ Further, the simulations predicted an average AUC_{0-24h} of 19 mg·h/L in patients on concomitant ART. Such an exposure is at the lower end of the rifampicin steady-state AUCs presented by the metaanalysis and suggests a 75% reduction in rifampicin exposure in patients on efavirenz-based ART. In the simulations of 35 mg/kg doses a previously described non-linear increase in bioavailability with higher dose was included.⁵ Despite inclusion of such nonlinearity, the model predicted a 45% lower AUC in patients on ART



Figure 2. pcVPC (n = 1000) for the final population pharmacokinetic model of rifampicin and 25-desacetylrifampicin stratified by concomitant ART. Circles are plasma concentration-time observations, the continuous line and the broken lines are the median and the 90th percentiles of the observed data, respectively, and the shaded areas represent the 95% CIs of the 5th, 50th and 95th percentiles predicted by the model.

Table 3. Parameter estimates of the fina	pharmacokinetic model of rifampicin in	adult patients coinfected with TB and HIV
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Parameter	Population mean (%RSE)	95% CI	% IIV (%RSE)
CL _{RIF} (L/h)	5.80 (12.9)	4.52 to 7.61	28.6 (34.5)
Fm	0.71 (16.5)	0.45 to 0.89	-
V _{RIF} (L)	48.0 (11.0)	38.6 to 58.6	53.3 (41.0)
MTT (h)	1.65 (18.0)	1.2 to 2.5	139.5 (25.5)
CL _{DERIE} (L/h)	29.6 (15.2)	21.2 to 37.8	-
Effect of ART on CL _{RIF}	+1.44 (39.6)	0.75 to 3.1	-
Effect of ART on CL _{DERIE}	+1.09 (44.3)	0.49 to 2.5	-
Relative F	1 fix	-	80.9 (23.1)
Effect of ART on F	-0.42 (42.7)	-0.003 to -0.61	_
Residual errors			
proportional, RIF (%)	41.7 (7.69)	34.5 to 46.3	-
proportional, DERIF (%)	36.5 (7.83)	31.0 to 41.9	-
additive. RIF (umol/L)	0.001 fix	-	-
additive, DERIF (µmol/L)	0.001 fix	-	-

 CL_{RIF} , rifampicin oral clearance; F_m , fraction of rifampicin clearance forming 25-desacetylrifampicin; V_{RIF} , oral volume of distribution for rifampicin; MTT, mean transit time; CL_{DERIF} , clearance of 25-desacetylrifampicin; F, bioavailability; IIV, inter-individual variability; RSE, relative standard error.



Figure 3. Simulated rifampicin plasma concentration-time profiles after the first dose (90% of the simulated data, n = 1000) in patients on concomitant ART (+ART) or ART naive. The continuous line is the mean plasma concentration, the shaded area is the 95% range of the simulated data and the broken line is the therapeutic threshold (8 mg/L).

Table 4. Model predictions and clinically observed rifampicin AUC_{0-24h} (mg·h/L) at steady state after a standard dose and a high dose

	Rifampi	cin dose
	10 mg/kg	35 mg/kg
Model prediction ^a		
first dose	76.1	455.0
ART induced	19.3	106.4
Meta-analysis ³⁴		
first dose	72.6	-
steady state	38.7	194.6

Data are given as geometric means.

^aPrediction based on 1000 simulations of a typical individual weighing 50 kg.

compared with the average steady-state AUC observed in clinical trials. In addition to saturation in rifampicin clearance, the underprediction of AUC may indicate saturation of the effect on pre-hepatic bioavailability. Thus, the effect of efavirenz-based ART may be proportionally smaller for higher doses of rifampicin. A study of the interaction at higher doses of rifampicin would give further answer to a potentially non-linear effect on prehepatic induction.

Simulations of 10 mg/kg doses indicated that the standard treatment results in too-low exposure of rifampicin. A dose of 35 mg/kg would result in a $C_{\rm max}$ above the recommended threshold for the majority of patients. Studies have shown that short-term (up to 12 weeks) treatment with 35 mg/kg rifampicin is safe^{3,35} and ongoing clinical trials will further give answer to the tolerability of high-dose rifampicin. The results presented here indicate that the standard-dose regimen is inadequate in patients

coinfected with TB and HIV and that higher doses of rifampicin are crucial in the studied population.

A recent study showed no alteration of rifampicin pharmacokinetics by efavirenz-based ART at steady state.¹¹ Since no effect was observed at steady state, efavirenz-based ART plausibly affects rifampicin exposure equivalently to rifampicin autoinduction for both prehepatic and hepatic metabolism. The early bactericidal effect of rifampicin increases with dose and is exposure dependent.³⁶ Moreover, potentially shorter TB therapy with higher rifampicin doses has been proposed.³⁷ Whereas rifampicin bioavailability is autoinduced after the first week of TB therapy, autoinduction of rifampicin clearance is maximized after a few weeks. Patients on efavirenz-based ART will thus have a lower exposure to rifampicin for at least a few weeks compared with patients not yet initiated on ART. The initially lower rifampicin exposure in patients already on concomitant efavirenz-based ART at the initiation of TB therapy may have a significant impact on the early bactericidal effect of rifampicin. Therefore, potential differences in clinical outcome or time to sputum conversion due to the interaction described in the present study require evaluation.

In addition to an effect on rifampicin clearance, the clearance of 25-desacetylrifampicin was significantly higher in patients on concomitant ART. The effect on rifampicin clearance was higher than the effect on 25-desacetylrifampicin clearance. The results suggest that efavirenz-based ART induces both the formation and elimination of the metabolite resulting in a higher AUC_{25-desacetylri-} fampicin.0-8h/AUCrifampicin.0-8h ratio in patients on ART. Rifampicin and efavirenz induce metabolic enzymes through activation of the nuclear pregnane X receptor and constitutive androstane receptor, respectively.^{38,39} In addition, rifampicin has been shown to moderately induce beta esterases in vitro.⁴⁰ The results presented here indicate an induction of beta esterases by efavirenz. Although the efficacy contribution of 25-desacetylrifampicin is considered low, the protein binding of 25-desacetylrifampicin is not known, whereas the protein binding of rifampicin is approximately 80%.²⁴ If the protein binding of 25-desacetylrifampicin is lower, the efficacy contribution may be higher than previously suggested, assuming that only unbound drug or metabolite can have an effect. A 30% change in AUC_{25-desacetylrifampicin}/AUC_{rifampicin} is unlikely to be of clinical relevance, considering the low exposure to 25-desacetylrifampicin. However, the combined lower exposure to both rifampicin and 25-desacetylrifampicin may effect early bactericidal activity.

The present study had some limitations. Firstly, the model was developed on data from a standard-dose regimen and the changes in rifampicin pharmacokinetics following doses of 35 mg/ kg are based on estimates from a previous model in TB patients. Secondly, the ART consisted of efavirenz, lamivudine and zidovudine/tenofovir and the drug responsible for the effect on rifampicin pharmacokinetics could not be identified. However, efavirenz is probably the cause of the drug-drug interaction due to its inductive properties. Thirdly, mechanistic conclusions regarding the change in metabolite/drug exposure ratio were not supported by the data. Nevertheless, induction of both 25-desacetylrifampicin formation and elimination is plausible.

In conclusion, a population pharmacokinetic model for rifampicin and its major metabolite 25-desacetylrifampicin in patients coinfected with HIV and TB was developed. Concomitant treatment with efavirenz-based ART resulted in a significant decrease in rifampicin and 25-desacetylrifampicin exposure. The described drug-drug interaction could potentially affect the time course of TB elimination due to a lower early bactericidal activity of rifampicin. Such an effect may increase the risk of resistance development.

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Transparency declarations

None to declare.

Author contributions

J.S., A.Ä. and M.A. wrote the original manuscript. J.S., A.Ä. and M.A. designed the research. J.S. performed the research. J.S., A.Ä. and M.A. analysed the data. E.B. and M.A. wrote the clinical study protocol. E.B. was the clinical study coordinator.

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