### MINI REVIEW

# Targeting the PI3K/Akt/mTOR pathway in non-small cell lung cancer (NSCLC)

# Aaron C. Tan 回

Division of Medical Oncology, National Cancer Centre Singapore, Singapore

#### Keywords

Akt pathway; mTOR pathway; non-small cell lung cancer; PI3K pathway; targeted therapy.

#### Correspondence

Aaron C. Tan, Division of Medical Oncology, National Cancer Centre Singapore, 11 Hospital Crescent, 169610, Singapore. Tel: +65 6436 8000 Fax: +65 6225 6283 Email: aaron.tan@singhealth.com.sg

Received: 10 December 2019; Accepted: 7 January 2020.

doi: 10.1111/1759-7714.13328

Thoracic Cancer **11** (2020) 511–518

### Abstract

The traditional classification of lung cancer into small cell lung cancer and nonsmall cell lung cancer (NSCLC) has been transformed with the increased understanding of the molecular alterations and genomic biomarkers that drive the development of lung cancer. Increased activation of the phosphatidylinositol 3-kinase (PI3K)/Akt/mechanistic target of rapamycin (mTOR) pathway leads to numerous hallmarks of cancer and this pathway represents an attractive target for novel anticancer therapies. In NSCLC, the PI3K/Akt/mTOR pathway has been heavily implicated in both tumorigenesis and the progression of disease. A number of specific inhibitors of PI3K, Akt and mTOR are currently under development and in various stages of preclinical investigation and in early phase clinical trials for NSCLC. Early evidence has yielded disappointing results. Clinical trials, however, have been performed on predominantly molecularly unselected populations, and patient enrichment strategies using high-precision predictive biomarkers in future trials will increase the likelihood of success. A greater understanding of the underlying molecular biology including epigenetic alterations is also crucial to allow for the detection of appropriate biomarkers and guide combination approaches.

### Introduction

Lung cancer remains one of the leading causes of cancer death worldwide.1 The traditional classification of lung cancer into small cell lung cancer and non-small cell lung cancer (NSCLC) has been transformed with the increased understanding of the molecular alterations and genomic biomarkers that drive the development of lung cancer. Targeted therapies for epidermal growth factor receptor (EGFR), anaplastic lymphoma kinase (ALK), ROS1 and BRAF have resulted in marked improvements in survival, particularly for patients with advanced disease.<sup>2</sup> Increased activation of the phosphatidylinositol 3-kinase (PI3K)/Akt/ mechanistic target of rapamycin (mTOR) pathway leads to numerous hallmarks of cancer, including acquired growth signal autonomy, inhibition of apoptosis, sustained angiogenesis, increased tissue invasion and metastasis and insensitivity to antigrowth signals. Consequently, this pathway represents an attractive target for novel anticancer therapies.

# Basic biology of the PI3K/Akt/mTOR pathway

The PI3K/Akt/mTOR pathway and signaling cascade is crucial in the regulation of cellular growth and metabolism. The importance of PI3K in cancer was initially described in 1985 after it was implicated in association with polyoma middle-T antigen, which is required for tumorigenesis in animals.<sup>3</sup> Subsequent work has intimately characterized the PI3K signaling pathway, and demonstrated that upregulation of this complex pathway is central in the development of cancer.

PI3Ks are a family of intracellular lipid kinases which phosphorylate the 3'-hydroxyl group of phosphatidylinositol and phosphoinositides.<sup>4</sup> They are divided into three classes (I–III), which each have distinct roles in signal transduction. Class I PI3Ks are divided into class IA PI3Ks that are activated by growth factor receptor tyrosine kinases, and class IB PI3Ks that are activated by G-protein-coupled receptors.<sup>5</sup> Class IA PI3K is a heterodimer consisting of a p85 regulatory subunit and a p110 catalytic

Thoracic Cancer **11** (2020) 511–518 © 2020 The Author. *Thoracic Cancer* published by China Lung Oncology Group and John Wiley & Sons Australia, Ltd **511** This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited. subunit. The p85 regulatory subunit is encoded by the *PIK3R1*, *PIK3R2* and *PIK3R3* genes which encode the p85 $\alpha$ , p85 $\beta$  and p55 $\gamma$  isoforms, respectively, and the p110 catalytic subunit is encoded by the *PIK3CA*, *PIK3CB* and *PIK3CD* genes which encode the p110 $\alpha$ , p110 $\beta$  and p110 $\delta$  isoforms, respectively.<sup>6</sup> Class II PI3Ks consist of a p110-like catalytic subunit only. The *PIK3C2A*, *PIK3C2B* and *PIK3C2G* genes encode the PIK3C2 $\alpha$ , PIK3C2 $\beta$ , PIK3C2 $\gamma$  isoforms, respectively. Class III PI3K consists of a single catalytic member, vacuolar protein sorting 34 (Vps34), which is encoded by the *PIK3C3* gene. Vps34 binds to the adapter protein Vps15, which is encoded by the *PIK3R4* gene.<sup>7</sup>

The role of each class of PI3K can be generally categorized into their importance in cell signaling (class I and II) or membrane trafficking (class II and III). A majority of the evidence for the importance of PI3K in human cancer implicates class IA PI3Ks, and specifically the p110a isoform. The presence of PIK3CA gene mutations or amplifications has been found in a diverse range of malignancies.8 In a breast cancer mouse model, inhibition of the p110a isoform led to increased mammary tumorigenesis.9 Preclinical evidence has also identified a modulatory or regulatory role for other class IA isoforms such as p110β and p1108.9,10 Further preclinical data suggests that there exists significant functional redundancy of class IA PI3Ks, and only a small fraction of total class I PI3K activity is required to maintain cell survival and proliferation.11 Inhibition of specific PI3K isoforms, such as p110a, may also lead to the upregulation of alternative bypass pathways such as the ERK pathway. Class IA PI3Ks can be activated by upstream receptor tyrosine kinases and growth factor stimulation. The regulatory subunit of the PI3K binds to the receptor tyrosine kinase and leads to the release of the p110 catalytic subunit, which translocates to the plasma membrane.12 PI3K phosphorylates phosphatidylinositol 4,5-bisphosphate (PIP2), to produce PI(3,4,5)P<sub>3</sub> (PIP3).<sup>13</sup> Phosphate and tensin homolog (PTEN) can regulate this step by dephosphorylating PIP3 to PIP2 and preventing further signal transduction.<sup>14</sup> Activated PIP3 allows for Akt activation via phosphorylation by phosphoinositide-dependent kinase-1 (PDK1), and consequently loss of PTEN is a key mechanism by which cancers increase PI3K signaling.<sup>15</sup> Germline mutations of PTEN as seen in Cowden syndrome also result in high risk of numerous cancers including breast, thyroid, endometrial and genitourinary cancers.<sup>16</sup>

Akt is a member of the AGC (PKA/PKG/PKC) protein kinase family and consists of three homologues, Akt1, Akt2 and Akt3 located at chromosomes 14q32, 19q13 and 1q44, respectively.<sup>17</sup> Akt activation subsequently leads to a number of potential downstream effects. It can result in inhibition of BAD and BAX,

proapoptotic Bcl2 family members. Akt may also phosphorylate Mdm2, which causes downregulation of p53-mediated apoptosis and forkhead transcription factors that produce cell-death promoting proteins.<sup>5</sup> The nuclear factor kappa-light-chain-enhancer of activated B cells (NF $\kappa$ B) transcription factor plays a crucial role in the consequences of PI3K/Akt pathway activation. NFkB regulates gene expression of hundreds of genes which are implicated in apoptosis, cell cycle control, immune modulation, cell survival and cell adhesion and differentiation.<sup>18</sup> Akt prevents negative regulation of NFkB by the IkB family, and in particular IkBa. IkBa removes NFκB from DNA and returns it to the cytoplasm.<sup>19</sup> Another important downstream pathway resulting from Akt activation is activation of the protein kinase, mTOR. This occurs via the phosphorylation of TSC2, which subsequently activates Rheb which then stimulates the mul-

tiprotein complex mTORC1. mTORC1 leads to further downstream activation of the eIF4 complex and subsequent promotion of tumorigenesis, regulation of the cell cycle and inhibition of apoptosis.<sup>4</sup> Another mTOR complex, mTORC2, causes Akt activation through phosphorylation with serine 473.<sup>20</sup>

The importance of upregulation, increased activation and constitutive signaling of the PI3K/Akt/mTOR pathway in cancer is well documented. Somatic mutations that encode for the various components of the signaling cascade and gene amplifications have been demonstrated in numerous different cancers. Mutations in PIK3CA have been identified in up to 36% of hepatocellular cancers, 26% of breast and 26% of colon cancers.<sup>8,21</sup> Smaller rates of PIK3CA mutations are also seen in ovarian, gliomas, gastric and lung cancers.<sup>8,21-23</sup> These mutations are predominantly clustered to two regions in exon 9 and 20, with exon 20 encoding the catalytic domain of  $p110\alpha$ and exon 9 encoding the helical domain of  $p110\alpha$ .<sup>24</sup> PIK3CA amplifications are also common, particularly in squamous cell carcinomas, with amplifications in up to 69% of cervical cancers, 66% of SCC of the lung and 42% of head and neck cancers.<sup>25,26</sup> These amplifications have also been demonstrated in significant proportions of gastric, thyroid, breast, esophageal adenocarcinoma and lung adenocarcinomas.<sup>27-30</sup> PTEN loss of heterozygosity is extremely common in glioblastoma (54%), prostate (35%), breast (23%), melanoma (37%) and gastric (47%) cancers, with PTEN mutations also seen in the same cancers to a lesser extent.<sup>27,31-33</sup> Akt amplifications meanwhile have been identified in head and neck, gastric, pancreatic, ovarian and breast cancers.34,35 Gene mutations in the Akt family which encode Akt1, Akt2 and Akt3 have also been demonstrated, and may lead to constitutive membrane localization of Akt.36

# Importance of the PI3K/Akt/mTOR pathway in NSCLC

In NSCLC, the PI3K/Akt/mTOR pathway has been heavily implicated in both tumorigenesis and the progression of disease. Somatic mutations of PIK3CA and amplifications of PIK3CA are frequently found in patients with NSCLC. A large study of 1144 consecutive NSCLC patients investigated tumor tissue using next generation sequencing (NGS) for PIK3CA mutations.<sup>37</sup> Mutations were identified in 3.7% of patients, with predominance for squamous cell carcinoma (8.9%) compared with adenocarcinoma (2.9%). E545K exon 9 mutations (57.1%) were most common, followed by H1047R exon 20 (16.7%) and E542K exon 9 (14.3%) mutations. Importantly, in a significant proportion of patients (57.1%), this study found coexisting oncogenic mutations in genes encoding for EGFR, BRAF, ALK and KRAS. Other studies have shown similar findings. Yamamoto et al.38 examined 86 NSCLC cell lines and 356 resected NSCLC tumors for PIK3CA mutations in exon 9 or 20 and PIK3CA amplification. Either a mutation or amplification was detected in 12.8% of cell lines and 19.1% of tumors. In this study, PIK3CA amplifications were also more common in patients with squamous cell carcinoma (33.1%) compared with adenocarcinoma (6.2%). The functional importance of PIK3CA mutation or amplification was confirmed with increased Akt activity, and mutations were similarly not mutually exclusive with EGFR or KRAS mutations. Further evidence supports the greater presence of PI3KCA amplification compared with mutation, and predominantly in patients of squamous cell carcinoma histology.39-41

Homozygous and heterozygous deletions of PTEN have also been shown in lung cancer. Loss of PTEN expression assessed by immunohistochemistry has been demonstrated in up to 24% of 125 resected early stage NSCLC specimens.<sup>42</sup> This may be in part due to increases in promoter methylation. Another series of early stage NSCLC specimens revealed complete loss of PTEN expression in 44% of tumors, reduced level of expression in 29% and normal expression in 27%.43 This study similarly showed methylation of PTEN in 26% of tumors with loss of heterozygosity at microsatellites in chromosome 10q23 occurring in 19% of studied specimens; however, neither was a significant predictor of PTEN protein expression. A retrospective analysis of the phase III FLEX study of chemotherapy in combination with cetuximab in patients with EGFRexpressing advanced NSCLC, showed 35% with negative PTEN expression.44 The presence of PTEN expression is potentially correlated with improved survival. Other studies have also shown total loss of PTEN expression may be more common in squamous cell carcinomas compared with adenocarcinomas.45

Upregulation of the Akt pathway has also been demonstrated in a significant proportion of patients with NSCLC. A study of 110 NSCLC tumors revealed 51% with increased Akt activity determined by immunohistochemistry.46 There was also significant association with Akt activation and increased mTOR and forkhead activity, important downstream targets of Akt. Malanga et al.47 examined 105 resected NSCLC specimens, and identified a somatic mutation of the gene encoding for Akt1 through direct sequencing of PCR products. Two squamous cell carcinoma specimens were found to contain the E17K mutation of Akt1 in exon 4. Another study using a high resolution melting assay, revealed four of 219 NSCLC specimens with an Akt1 mutation.48 Of these, one contained the E17K mutations and was of squamous cell carcinoma histology. The other three tumors showed rare single nucleotide polymorphisms. The relatively rarity of the E17K mutation in NSCLC has been further confirmed in other series.<sup>49</sup> The discordance between levels of Akt overexpression and presence of somatic mutation may indicate coexisting mutations or amplifications that result in Akt activation. This has been demonstrated in preclinical studies in NSCLC cell lines, with Akt activation attributed to loss of PTEN, EGFR or PIK3CA mutation, or HER2 amplification.50

Upregulation of the mTOR pathway has also been illustrated in significant proportions of NSCLC tumors, with increased p-mTOR in up to 90% of patients with adenocarcinoma, 60% of patients with large cell carcinoma and 40% of patients with squamous cell carcinoma.<sup>51-53</sup> Downstream products of mTOR activation, S6K and 4E-BP1 have also been identified in up to 58% and 25% of NSCLC specimens respectively, with a greater predominance in adenocarcinoma.54 There is also a strong correlation of p-S6K and p-mTOR positivity. The importance of mTOR in bronchial adenocarcinoma may be related to its coupling with eIF-4E which functions as an oncogene.55 The presence of mTOR activity may also be a poor prognostic factor in early stage NSCLC. Several studies have demonstrated increased mTOR expression as determined by immunohistochemistry was associated with poor survival.56,57

The importance of the PI3K/Akt/mTOR pathway also extends to its role in tumors with other known activating mutations, such as *EGFR* and *KRAS*. Studies suggest that in patients with an *EGFR* mutation, the Akt/mTOR pathway is constitutively activated in 67% of cases.<sup>58</sup> Additionally, another study revealed 18% of specimens showed positive staining for p-EGFR, p-Akt and p-mTOR, indicating the importance of this signaling cascade.<sup>52</sup> The importance of Akt activation may also relate to the development of resistance to therapy. Preclinical evidence suggests that Akt activation may confer acquired resistance to EGFR

inhibitors in *EGFR* mutant NSCLC.<sup>59</sup> Similarly mTOR activation has been associated with *EGFR* and *KRAS* mutations and may act as a mechanism of resistance to EGFR inhibitors.<sup>60</sup> Given the encompassing downstream signaling effects of the PI3K/Akt/mTOR pathway on the development and progression of malignancy, and its potential influence in response and resistance to standard therapies, it represents an attractive target for anticancer therapy in NSCLC.

## Clinical trials of PI3K/Akt/mTOR inhibitors in NSCLC

A number of specific inhibitors of PI3K, Akt and mTOR are currently under development and in various stages of preclinical investigation and in early phase clinical trials for NSCLC (Table 1). There are several pan-class I PI3K inhibitors, including pictilisib (GDC-0941), PX-866, buparlisib (BKM120), pilaralisib (XL-147) and GNE-317. Pictilisib has been evaluated in phase I trials either alone or in combination with standard chemotherapy. In a phase IA dose-escalation trial of patients with advanced solid tumors, the single-agent maximum tolerated dose was 330 mg, with maculopapular rash as the dose limiting toxicity.83 Pictilisib has been further investigated in a phase IB dose-escalation trial in combination with standard first-line chemotherapy in patients with advanced NSCLC.<sup>61</sup> Patients received pictilisib with either carboplatin and paclitaxel or cisplatin and pemetrexed, with the addition of bevacizumab depending on histology. Of 66 patients, 29 (43.9%) had partial response and 20 (30.9%) had stable disease. This led to the phase II FIGARO study of pictilisib in combination with first-line chemotherapy; however, preliminary data did not reveal any significant progressionfree survival (PFS) or overall survival (OS) benefit.84 Similar findings have been reported in a phase IA/IB trial in a Japanese cohort of patients.<sup>85</sup> PX-866 has been evaluated in combination with docetaxel chemotherapy in a randomized phase II trial of advanced, refractory NSCLC patients.<sup>62</sup> There was no improvement in PFS, response rate or OS with the addition of PX-866. These patients, however, were molecularly unselected. In the phase I trial of PX-866 in combination with docetaxel, there was one NSCLC patient with a PIK3CA mutation who had a prolonged response to continuation PX-866 after cessation of docetaxel.<sup>86</sup> Buparlisib has also been investigated as monotherapy in the BASALT-1 phase II trial of previously treated patients with NSCLC.63 Patients were selected based on identification of PI3K pathway activation. The primary objective for efficacy based on 12-week PFS, was not met. Further studies of buparlisib in combination with chemotherapy were similarly negative,<sup>64</sup> whilst it has also been studied in combination with gefitinib.65 Pilaralisib has also

Table 1 PI3K/Akt/mTOR inhibitors evaluated in clinical trials for NSCLC

	Phase of	
	development for	
Drug	NSCLC	References
Pan-PI3K inhibitors		
Pictilisib (GDC-0941)	II	Soria et al. 61
PX-866	II	Levy et al. 62
Buparlisib (BKM120)	II	Vansteenkiste <i>et al</i> . <sup>63</sup> Adjei et al.64 Tan <i>et al</i> . <sup>65</sup>
Pilaralisib (XL-147)	I	Soria et al. <sup>61</sup> Shapiro et al. <sup>66</sup>
Selective PI3K		
inhibitors		
Alpelisib (BYL719)	II	Zhou <i>et al</i> . <sup>67</sup>
INK1117	I.	lartchouk et al. 68
GSK2636771	I.	Mateo et al. <sup>69</sup>
AZD8186	I.	Siu et al.70
SAR260301	I.	Bedard et al. 71
Taselisib	II	Langer et al.72
(GDC-0032)		
Akt inhibitors		
Perifosine	II	Lara et al.73
mTOR inhibitors		
Everolimus	II	Besse <i>et al.</i> <sup>74</sup> Price <i>et al.</i> <sup>75</sup>
Sirolimus	Ш	Moran <i>et al</i> . <sup>76</sup> Blumenthal <i>et al.</i> <sup>77</sup>
Temsirolimus	П	Gandhi <i>et al</i> . <sup>78</sup>
Ridaforolimus	П	Riely et al.79
Dual PI3K/mTOR		-
inhibitors		
BEZ235	П	Herrera et al. <sup>80</sup>
XL765	I.	Cohen <i>et al</i> . <sup>81</sup>
GDC-0980	I	Calvo et al. <sup>82</sup>

been assessed as monotherapy in phase I trials, with one partial response in a patient with NSCLC.<sup>66</sup> Another phase I trial in combination with erlotinib established its safety profile although similarly exhibited only one partial response.<sup>61</sup>

Whilst the pan-class I PI3K inhibitors have shown disappointing efficacy, there are isoform-specific class I PI3K inhibitors also in development. Alpelisib (BYL719) is a potent p110 $\alpha$  inhibitor, and is currently under investigation in a phase II study of patients with advanced NSCLC and a *PIK3CA* mutation or amplification.<sup>67</sup> INK1117 is another p110 $\alpha$ -specific inhibitor in early phase development.<sup>68</sup> Specific p110 $\beta$  inhibitors such as GSK2636771,<sup>69</sup> AZD8186<sup>70</sup> and SAR260301<sup>71</sup> have also been tested in phase I trials in patients with advanced solid tumors including NSCLC. Taselisib (GDC-0032) is a PI3K inhibitor that inhibits the p110 $\alpha$ , p110 $\gamma$  and p110 $\delta$  isoforms, but spares the p110 $\beta$  isoform. It has been evaluated as part of the phase II LUNG-MAP study, in previously treated NSCLC patients with a *PIK3CA* mutation.<sup>72</sup> This substudy was closed for futility after an interim analysis.

Perifosine is an Akt inhibitor that has been investigated in a phase I trial of patients with advanced NSCLC.<sup>87</sup> Of 15 patients evaluated, there was one unconfirmed partial response. The trial was expanded to a phase II trial, the results of which have not yet been reported. MK-2206 is a highly selective Akt inhibitor which was evaluated in a phase II trial, in combination with erlotinib, in NSCLC patients who had previously progressed on erlotinib therapy.<sup>73</sup> Patients were stratified based on *EGFR* mutation status. Median PFS was 4.4 months in *EGFR* mutant patients, and 4.6 months in *EGFR* wild-type patients.

mTOR inhibitors are widely used to prevent transplant rejection, and are approved as anticancer therapy for renal cell carcinoma, neuroendocrine tumors and breast cancer. Everolimus is an mTOR inhibitor that selectively inhibits mTORC1 signaling. It has been assessed in several phase I trials in previously treated advanced NSCLC, either as monotherapy<sup>88</sup> or in combination with pemetrexed chemotherapy.<sup>89</sup> The combination of everolimus and EGFR inhibitors has also been evaluated in phase II trials in NSCLC patients unselected for EGFR mutation status. Everolimus in combination with erlotinib<sup>74</sup> or gefitinib<sup>75</sup> both failed to show significant efficacy to progress to a phase III trial. Sirolimus, another oral mTOR inhibitor, has demonstrated significant toxicity in combination with afatinib;<sup>76</sup> however, showed potential activity in combination with pemetrexed chemotherapy.77 Trials of mTOR inhibitors in molecularly selected patients have also been conducted. Temsirolimus, an intravenous mTOR inhibitor has also been evaluated in a phase II trial in combination with neratinib, an oral HER2 inhibitor, in HER2 mutant advanced NSCLC patients.78 The combination therapy resulted in a 19% response rate. Ridaforolimus has also been investigated in KRAS mutant advanced NSCLC patients in a phase II trial versus placebo, with a two month improvement in PFS.79

Second generation inhibitors have also been developed with dual targeting of both PI3K and mTOR. Importantly, these agents block both mTORC1 and mTORC2 complexes, and have pan-class I PI3K inhibitory activity. BEZ235 is one such agent, currently under evaluation in numerous phase I/II clinical trials, and preclinical evidence suggests strong activity in lung cancer cell lines.<sup>80</sup> XL765 has been tested in a phase I trial in combination with erlotinib with a cohort of patients with NSCLC, and the combination was generally well tolerated.<sup>81</sup> GDC-0980 has also been investigated in combination with chemotherapy in a phase Ib trial with a NSCLC cohort with the trial illustrating an acceptable safety profile.<sup>82</sup> Vistusertib (AZD2014) in combination with paclitaxel demonstrated a

33% response rate in previously treated squamous NSCLC, in the dose escalation arm of the TAX-TORC study. $^{90}$ 

Finally, there are also dual PI3K/mTOR inhibitors that have been developed. Gedatolisib has been evaluated in a phase I trial in combination with chemotherapy (docetaxel or cisplatin) in NSCLC patients or dacomitinib in *EGFR* mutant NSCLC patients.<sup>91</sup> Toxicity profiles were manageable, and ongoing phase I/II trials are underway.

# Conclusion

Early evidence for targeted therapies against the PI3K/Akt/ mTOR pathway have yielded disappointing results. Clinical trials, however, have been performed on predominantly molecularly unselected populations, and patient enrichment strategies using high-precision predictive biomarkers in future trials will increase the likelihood of success. A greater understanding of the underlying molecular biology, including epigenetic alterations is also crucial to allow for the detection of appropriate biomarkers and guide combination approaches. Furthermore, innovative and novel clinical trial design will enhance our ability to evaluate novel agents and combinations to account for molecular diversity and ultimately improve patient outcomes.

# Disclosure

The author declares no conflict of interest. A.C.T is the recipient of an International Association for the Study of Lung Cancer (IASLC) Fellowship 2018-2020.

# References

- 1 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018; **68**: 394–424.
- 2 Hirsch FR, Scagliotti GV, Mulshine JL *et al.* Lung cancer: Current therapies and new targeted treatments. *Lancet* 2017; 389: 299–311.
- 3 Whitman M, Kaplan DR, Schaffhausen B, Cantley L, Roberts TM. Association of phosphatidylinositol kinase activity with polyoma middle-T competent for transformation. *Nature* 1985; **315**: 239–42.
- 4 Engelman JA, Luo J, Cantley LC. The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism. *Nat Rev Genet* 2006; 7: 606–19.
- 5 Cantley LC. The phosphoinositide 3-kinase pathway. *Science* 2002; **296**: 1655–7.
- 6 Fruman DA, Meyers RE, Cantley LC. Phosphoinositide kinases. Annu Rev Biochem 1998; 67: 481–507.

- 7 Jean S, Kiger AA. Classes of phosphoinositide 3-kinases at a glance. *J Cell Sci* 2014; **127**: 923–8.
- 8 Samuels Y, Wang Z, Bardelli A *et al.* High frequency of mutations of the PIK3CA gene in human cancers. *Science* 2004; **304**: 554.
- 9 Utermark T, Rao T, Cheng H *et al.* The p110α and p110β isoforms of PI3K play divergent roles in mammary gland development and tumorigenesis. *Genes Dev* 2012; 26: 1573–86.
- 10 Luk SK, Piekorz RP, Nürnberg B, Tony To SS. The catalytic phosphoinositol 3-kinase isoform p110δ is required for glioma cell migration and invasion. *Eur J Cancer* 2012; 48: 149–57.
- Foukas LC, Berenjeno IM, Gray A, Khwaja A, Vanhaesebroeck B. Activity of any class IA PI3K isoform can sustain cell proliferation and survival. *Proc Natl Acad Sci* USA 2010; 107: 11381–6.
- 12 Carpenter CL, Auger KR, Chanudhuri M *et al.* Phosphoinositide 3-kinase is activated by phosphopeptides that bind to the SH2 domains of the 85-kDa subunit. *J Biol Chem* 1993; **268**: 9478–83.
- 13 Alessi DR, James SR, Downes CP *et al.* Characterization of a 3-phosphoinositide-dependent protein kinase which phosphorylates and activates protein kinase Balpha. *Curr Biol* 1997; 7: 261–9.
- 14 Maehama T, Dixon JE. PTEN: A tumour suppressor that functions as a phospholipid phosphatase. *Trends Cell Biol* 1999; 9: 125–8.
- 15 Myers MP, Pass I, Batty IH *et al.* The lipid phosphatase activity of PTEN is critical for its tumor supressor function. *Proc Natl Acad Sci U S A* 1998; **95**: 13513–8.
- 16 Sansal I, Sellers WR. The biology and clinical relevance of the PTEN tumor suppressor pathway. *J Clin Oncol* 2004; 22: 2954–63.
- 17 Murthy SS, Tosolini A, Taguchi T, Testa JR. Mapping of AKT3, encoding a member of the Akt/protein kinase B family, to human and rodent chromosomes by fluorescence in situ hybridization. *Cytogenet Cell Genet* 2000; **88**: 38–40.
- 18 Sonenshein GE. Rel/NF-κB transcription factors and the control of apoptosis. Semin Cancer Biol 1997; 8: 113–9.
- 19 Duronio V. The life of a cell: Apoptosis regulation by the PI3K/PKB pathway. *Biochem J* 2008; 415: 333–44.
- 20 Sarbassov DD, Guertin DA, Ali SM, Sabatini DM. Phosphorylation and regulation of Akt/PKB by the rictormTOR complex. *Science* 2005; **307**: 1098–101.
- 21 Lee JW, Soung YH, Kim SY *et al.* PIK3CA gene is frequently mutated in breast carcinomas and hepatocellular carcinomas. *Oncogene* 2005; 24: 1477–80.
- 22 Campbell IG, Russell SE, Choong DY *et al.* Mutation of the PIK3CA gene in ovarian and breast cancer. *Cancer Res* 2004; 64: 7678–81.
- 23 Hartmann C, Bartels G, Gehlhaar C, Holtkamp N, von Deimling A. PIK3CA mutations in glioblastoma multiforme. *Acta Neuropathol* 2005; 109: 639–42.

- 24 Samuels Y, Velculescu VE. Oncogenic mutations of PIK3CA in human cancers. *Cell Cycle* 2004; **3**: 1221–4.
- 25 Ma YY, Wei SJ, Lin YC *et al.* PIK3CA as an oncogene in cervical cancer. *Oncogene* 2000; **19**: 2739–44.
- 26 Massion PP, Kuo WL, Stokoe D *et al*. Genomic copy number analysis of non-small cell lung cancer using array comparative genomic hybridization: Implications of the phosphatidylinositol 3-kinase pathway. *Cancer Res* 2002; 62: 3636–40.
- 27 Byun DS, Cho K, Ryu BK *et al.* Frequent monoallelic deletion of PTEN and its reciprocal associatioin with PIK3CA amplification in gastric carcinoma. *Int J Cancer* 2003; **104**: 318–27.
- 28 Wu G, Mambo E, Guo Z *et al.* Uncommon mutation, but common amplifications, of the PIK3CA gene in thyroid tumors. *J Clin Endocrinol Metab* 2005; **90**: 4688–93.
- 29 Wu G, Xing M, Mambo E *et al.* Somatic mutation and gain of copy number of PIK3CA in human breast cancer. *Breast Cancer Res* 2005; 7: R609–16.
- 30 Miller CT, Moy JR, Lin L *et al*. Gene amplification in esophageal adenocarcinomas and Barrett's with high-grade dysplasia. *Clin Cancer Res* 2003; **9**: 4819–25.
- 31 Wang SI, Puc J, Li J *et al.* Somatic mutations of PTEN in glioblastoma multiforme. *Cancer Res* 1997; **57**: 4183–6.
- 32 Wang SI, Parsons R, Ittmann M. Homozygous deletion of the PTEN tumor suppressor gene in a subset of prostate adenocarcinomas. *Clin Cancer Res* 1998; **4**: 811–5.
- 33 Celebi JT, Shendrik I, Silvers D, Peacocke M. Identification of PTEN mutations in metastatic melanoma specimens. *J Med Genet* 2000; **37**: 653–7.
- 34 Bellacosa A, de Feo D, Godwin AK *et al.* Molecular alterations of the AKT2 oncogene in ovarian and breast carcinomas. *Int J Cancer* 1995; **64**: 280–5.
- 35 Staal SP. Molecular cloning of the akt oncogene and its human homologues AKT1 and AKT2: Amplification of AKT1 in a primary human gastric adenocarcinoma. *Proc Natl Acad Sci U S A* 1987; **84**: 5034–7.
- 36 Carpten JD, Faber AL, Horn C *et al*. A transforming mutation in the pleckstrin homology domain of AKT1 in cancer. *Nature* 2007; **448**: 439–44.
- 37 Scheffler M, Bos M, Gardizi M *et al.* PIK3CA mutations in non-small cell lung cancer (NSCLC): Genetic heterogeneity, prognostic impact and incidence of prior malignancies. *Oncotarget* 2015; **6**: 1315–26.
- 38 Yamamoto H, Shigematsu H, Nomura M *et al.* PIK3CA mutations and copy number gains in human lung cancers. *Cancer Res* 2008; **68**: 6913–21.
- 39 Ji M, Guan H, Gao C, Shi B, Hou P. Highly frequent promoter methylation and PIK3CA amplification in nonsmall cell lung cancer (NSCLC). *BMC Cancer* 2011; 11: 147.
- 40 Okudela K, Suzuki M, Kageyama S *et al.* PIK3CA mutation and amplification in human lung cancer. *Pathol Int* 2007; 57: 664–71.

- 41 Kawano O, Sasaki H, Endo K *et al.* PIK3CA mutation status in Japanese lung cancer patients. *Lung Cancer* 2006; **54**: 209–15.
- 42 Soria JC, Lee HY, Lee JI *et al.* Lack of PTEN expression in non-small cell lung cancer could be related to promoter methylation. *Clin Cancer Res* 2002; **8**: 1178–84.
- 43 Marsit CJ, Zheng S, Aldape K *et al.* PTEN expression in non-small-cell lung cancer: Evaluating its relation to tumor characteristics, allelic loss, and epigenetic alteration. *Hum Pathol* 2005; **36**: 768–76.
- 44 O'Byrne KJ, Gatzemeier U, Bondarenko I *et al.* Molecular biomarkers in non-small-cell lung cancer: A retrospective analysis of data from the phase 3 FLEX study. *Lancet Oncol* 2011; **12**: 795–805.
- 45 Spoerke JM, O'Brien C, Huw L *et al.* Phosphoinositide 3-kinase (PI3K) pathway alterations are associated with histologic subtypes and are predictive of sensitivity to PI3K inhibitors in lung cancer preclinical models. *Clin Cancer Res* 2012; **18**: 6771–83.
- 46 Balsara BR, Pei J, Mitsuuchi Y *et al.* Frequent activation of AKT in non-small cell lung carcinomas and preneoplastic bronchial lesions. *Carcinogenesis* 2004; 25: 2053–9.
- 47 Malanga D, Scrima M, De Marco C *et al.* Activating E17K mutation in the gene encoding the protein kinase AKT1 in a subset of squamous cell carcinoma of the lung. *Cell Cycle* 2008; 7: 665–9.
- 48 Do H, Solomon B, Mitchell PL, Fox SB, Dobrovic A. Detection of the transforming AKT1 mutation E17K in nonsmall cell lung cancer by high resolution melting. *BMC Res Notes* 2008; 1: 14.
- 49 Bleeker FE, Felicioni L, Buttitta F *et al.* AKT1(E17K) in human solid tumours. *Oncogene* 2008; **27**: 5648–50.
- 50 Guo Y, Du J, Kwiatkowski DJ. Molecular dissection of AKT activation in lung cancer cell lines. *Mol Cancer Res* 2013; 11: 282–93.
- 51 Dobashi Y, Suzuki S, Matsubara H, Kimura M, Endo S, Ooi A. Critical and diverse involvement of Akt/mammalian target of rapamycin signaling in human lung carcinomas. *Cancer* 2009; **115**: 107–18.
- 52 Dobashi Y, Suzuki S, Kimura M *et al.* Paradigm of kinasedriven pathway downstream of epidermal growth factor receptor/Akt in human lung carcinomas. *Hum Pathol* 2011; 42: 214–26.
- 53 Hiramatsu M, Ninomiya H, Inamura K *et al.* Activation status of receptor tyrosine kinase downstream pathways in primary lung adenocarcinoma with reference of KRAS and EGFR mutations. *Lung Cancer* 2010; **70**: 94–102.
- 54 Dobashi Y, Watanabe Y, Miwa C, Suzuki S, Koyama S. Mammalian target of rapamycin: A central node of complex signaling cascades. *Int J Clin Exp Pathol* 2011; 4: 476–95.
- 55 Mamane Y, Petroulakis E, Rong L, Yoshida K, Ler LW, Sonenberg N. eIF4E-from translation to transformation. *Oncogene* 2004; **23**: 3172–9.
- 56 Gately K, Al-Alao B, Dhillon T *et al.* Overexpression of the mammalian target of rapamycin (mTOR) and angioinvasion

are poor prognostic factors in early stage NSCLC: A verification study. *Lung Cancer* 2012; **75**: 217–22.

- 57 Dhillon T, Mauri FA, Bellezza G et al. Overexpression of the mammalian target of rapamycin: A novel biomarker for poor survival in resected early stage non-small cell lung cancer. J Thorac Oncol 2010; 5: 314–9.
- 58 Dobashi Y, Koyama S, Kanai Y, Tetsuka K. Kinase-driven pathways of EGFR in lung carcinomas: Perspectives on targeting therapy. *Front Biosci (Landmark Ed)* 2011; 16: 1714–32.
- 59 Jacobsen K, Bertran-Alamillo J, Molina MA et al. Convergent Akt activation drives acquired EGFR inhibitor resistance in lung cancer. Nat Commun 2017; 8: 410.
- 60 Conde E, Angulo B, Tang M *et al.* Molecular context of the EGFR mutations: Evidence for the activation of mTOR/S6K signaling. *Clin Cancer Res* 2006; **12**: 710–7.
- 61 Soria JC, Adjei AA, Bahleda R *et al.* A phase IB doseescalation study of the safety and pharmacokinetics of pictilisib in combination with either paclitaxel and carboplatin (with or without bevacizumab) or pemetrexed and cisplatin (with or without bevacizumab) in patients with advanced non-small cell lung cancer. *Eur J Cancer* 2017; **86**: 186–96.
- 62 Levy B, Spira A, Becker D *et al*. A randomized, phase 2 trial of Docetaxel with or without PX-866, an irreversible oral phosphatidylinositol 3-kinase inhibitor, in patients with relapsed or metastatic non-small-cell lung cancer. *J Thorac Oncol* 2014; **9**: 1031–5.
- 63 Vansteenkiste JF, Canon JL, De Braud F *et al.* Safety and efficacy of Buparlisib (BKM120) in patients with PI3K pathway-activated non-small cell lung cancer: Results from the phase II BASALT-1 study. *J Thorac Oncol* 2015; **10**: 1319–27.
- 64 Adjei AA, Bennouna J, Leighl NB *et al.* Safety and efficacy of buparlisib (BKM120) and chemotherapy in advanced, squamous non-small cell lung cancer (sqNSCLC): Results from the phase Ib/II BASALT-2 and BASALT-3 studies. *J Clin Oncol* 2016; **34**: e20522–2.
- 65 Tan DSW, Lim KH, Tai WM *et al.* A phase Ib safety and tolerability study of a pan class I PI3K inhibitor buparlisib (BKM120) and gefitinib (gef) in EGFR TKI-resistant NSCLC. *J Clin Oncol* 2013; **31**: 8107–7.
- 66 Shapiro GI, Rodon J, Bedell C *et al.* Phase I safety, pharmacokinetic, and pharmacodynamic study of SAR245408 (XL147), an oral pan-class I PI3K inhibitor, in patients with advanced solid tumors. *Clin Cancer Res* 2014; 20: 233–45.
- 67 Zhou Q, Zhang XC, Peng B *et al.* A phase II cluster study of single agent AUY922, BYL719, INC280, LDK378, and MEK162 in Chinese patients with advanced non-small cell lung cancer (NSCLC). *J Clin Oncol* 2014; **32**: TPS8122–2.
- 68 Iartchouk N, Koenig E, Shah P *et al.* Abstract A169: Assessment of genotype-correlated sensitivity to the investigational PI3Kα selective inhibitor MLN1117 in preclinical models. *Mol Cancer Ther* 2013; **12**: A169–9.

- 69 Mateo J, Ganji G, Lemech C *et al.* A first-time-in-human study of GSK2636771, a phosphoinositide 3 kinase beta-selective inhibitor, in patients with advanced solid tumors. *Clin Cancer Res* 2017; **23**: 5981–92.
- 70 Siu LL, De Bono J, Wisinski KB *et al.* Abstract CT329: Phase I study of the PI3Kβ/δ inhibitor AZD8186 in patients with advanced castration resistant prostate cancer, triple negative breast cancer, squamous non-small cell lung cancer or PTEN deficient solid tumors: Update from dose-finding. *Cancer Res* 2015; **75**: CT329–9.
- 71 Bedard PL, Davies MA, Kopetz S *et al.* First-in-human trial of the PI3Kß-selective inhibitor SAR260301 in patients with advanced solid tumors. *Cancer* 2018; **124**: 315–24. https:// doi.org/10.1002/cncr.31044.
- 72 Langer CJ, Redman MW, Wade JL 3rd *et al.* SWOG S1400B (NCT02785913), a phase II study of GDC-0032 (Taselisib) for previously treated PI3K-positive patients with stage IV squamous cell lung cancer (lung-MAP sub-study). *J Thorac Oncol* 2019; **14**: 1839–46.
- 73 Lara PN, Longmate J, Mack PC *et al.* Phase II study of the AKT inhibitor MK-2206 plus Erlotinib in patients with advanced non-small cell lung cancer who previously progressed on Erlotinib. *Clin Cancer Res* 2015; **21**: 4321–6.
- 74 Besse B, Leighl N, Bennouna J *et al.* Phase II study of everolimus-erlotinib in previously treated patients with advanced non-small-cell lung cancer. *Ann Oncol* 2014; 25: 409–15.
- 75 Price KA, Azzoli CG, Krug LM *et al.* Phase II trial of gefitinib and everolimus in advanced non-small cell lung cancer. *J Thorac Oncol* 2010; **5**: 1623–9.
- 76 Moran T, Palmero R, Provencio M *et al.* A phase Ib trial of continuous once-daily oral afatinib plus sirolimus in patients with epidermal growth factor receptor mutation-positive non-small cell lung cancer and/or disease progression following prior erlotinib or gefitinib. *Lung Cancer* 2017; **108**: 154–60.
- 77 Blumenthal GM, Ballas MS, Bernstein W et al. A phase I/II trial of pemetrexed and sirolimus in advanced NSCLC. J Clin Oncol 2010; 28: 7600–0.
- 78 Gandhi L, Besse B, Mazieres J et al. MA04.02 Neratinib ± Temsirolimus in HER2-mutant lung cancers: An international, randomized phase II study. J Thorac Oncol 2017; 12: S358–9.
- 79 Riely GJ, Brahmer JR, Planchard D *et al.* A randomized discontinuation phase II trial of ridaforolimus in non-small cell lung cancer (NSCLC) patients with KRAS mutations. *J Clin Oncol* 2012; **30**: 7531–1.
- 80 Herrera VA, Zeindl-Eberhart E, Jung A, Huber RM, Bergner A. The dual PI3K/mTOR inhibitor BEZ235 is effective in lung cancer cell lines. *Anticancer Res* 2011; 31: 849–54.
- 81 Cohen RB, Janne PA, Engelman JA *et al.* A phase I safety and pharmacokinetic (PK) study of PI3K/TORC1/TORC2

inhibitor XL765 (SAR245409) in combination with erlotinib (E) in patients (pts) with advanced solid tumors. *J Clin Oncol* 2010; **28**: 3015–5.

- 82 Calvo E, Boni V, Heist RS *et al.* 1230PDPHASE 1B study of oral dual-PI3K/MTOR inhibitor GDC-0980 in combination with carboplatin (carbo)/paclitaxel (PAC) ± bevacizumab (BEV) and cisplatin (cis)/pemetrexed (PEM) in patients (PTS) with advanced solid tumors and NSCLC. *Ann Oncol* 2014; **25**: iv429–30.
- 83 Sarker D, Ang JE, Baird R *et al.* First-in-human phase I study of Pictilisib (GDC-0941), a potent pan-class I phosphatidylinositol-3-kinase (PI3K) inhibitor, in patients with advanced solid tumors. *Clin Cancer Res* 2015; 21: 77–86.
- 84 Chen X, Chang CW, Spoerke JM *et al.* Low-pass wholegenome sequencing of circulating cell-free DNA demonstrates dynamic changes in genomic copy number in a squamous lung cancer clinical cohort. *Clin Cancer Res* 2019; 25: 2254–63.
- 85 Yamamoto N, Fujiwara Y, Tamura K *et al.* Phase Ia/Ib study of the pan-class I PI3K inhibitor pictilisib (GDC-0941) administered as a single agent in Japanese patients with solid tumors and in combination in Japanese patients with nonsquamous non-small cell lung cancer. *Invest New Drugs* 2017; **35**: 37–46.
- 86 Bowles DW, Ma WW, Senzer N *et al.* A multicenter phase 1 study of PX-866 in combination with docetaxel in patients with advanced solid tumours. *Br J Cancer* 2013; **109**: 1085–92.
- 87 Henderson I, Spigel DR, Nemunaitis JJ *et al.* A phase
  1 study of weekly, divided dose perifosine in patients (pts)
  with non-small cell lung cancer (NSCLC). *J Clin Oncol* 2006;
  24: 13063–3.
- 88 Soria JC, Shepherd FA, Douillard JY *et al.* Efficacy of everolimus (RAD001) in patients with advanced NSCLC previously treated with chemotherapy alone or with chemotherapy and EGFR inhibitors. *Ann Oncol* 2009; 20: 1674–81.
- 89 Vansteenkiste J, Solomon B, Boyer M *et al.* Everolimus in combination with pemetrexed in patients with advanced non-small cell lung cancer previously treated with chemotherapy: A phase I study using a novel, adaptive Bayesian dose-escalation model. *J Thorac Oncol* 2011; **6**: 2120–9.
- 90 Krebs M, Spicer J, Steele N *et al.* P3.02c-003 TAX-TORC: The novel combination of weekly paclitaxel and the dual mTORC1/2 inhibitor AZD2014 for the treatment of squamous NSCLC. *J Thorac Oncol* 2017; **12**: S1272–3.
- 91 Wainberg ZA, Shapiro G, Curigliano G *et al.* Phase I study of the PI3K/mTOR inhibitor gedatolisib (PF-05212384) in combination with docetaxel, cisplatin, and dacomitinib. *J Clin Oncol* 2016; **34**: 2566–6.