

The European MAPPYACTS Trial: Precision Medicine Program in Pediatric and Adolescent Patients with Recurrent Malignancies



Pablo Berlanga¹, Gaelle Pierron², Ludovic Lacroix³, Mathieu Chicard⁴, Tiphaine Adam de Beaumais⁵, Antonin Marchais⁶, Anne C. Harttrampf¹, Yasmine Iddir^{4,7}, Alicia Larive⁸, Aroa Soriano Fernandez⁹, Imene Hezam¹, Cecile Chevassus⁸, Virginie Bernard¹⁰, Sophie Cotteret³, Jean-Yves Scoazec³, Arnaud Gauthier¹¹, Samuel Abbou¹, Nadege Corradini¹², Nicolas André^{13,14}, Isabelle Aerts¹⁵, Estelle Thebaud¹⁶, Michela Casanova¹⁷, Cormac Owens¹⁸, Raquel Hladun-Alvaro¹⁹, Stefan Michiels⁸, Olivier Delattre^{4,10,15}, Gilles Vassal⁵, Gudrun Schleiermacher^{4,15}, and Birgit Georger^{1,6}

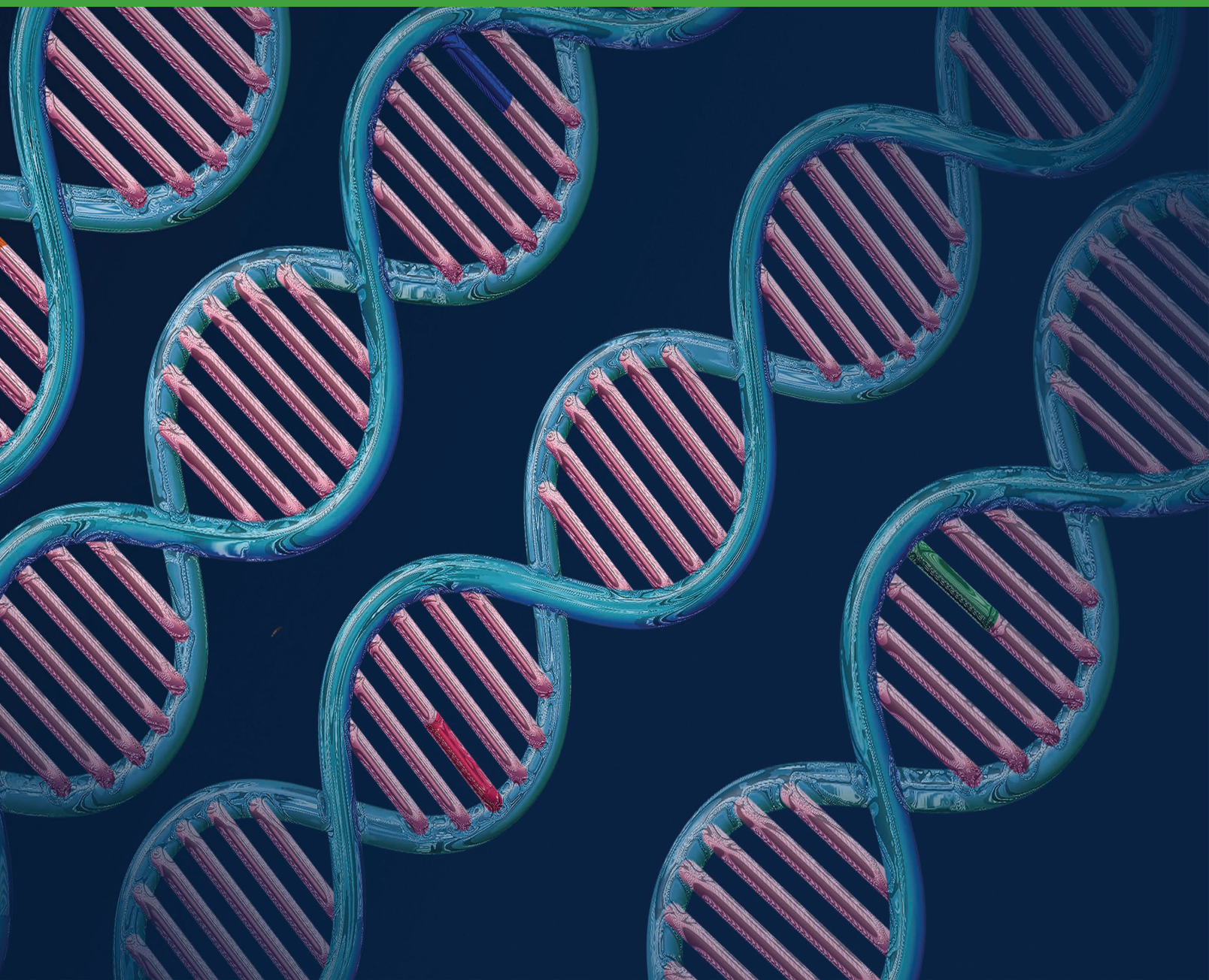
ABSTRACT

MAPPYACTS (NCT02613962) is an international prospective precision medicine trial aiming to define tumor molecular profiles in pediatric patients with recurrent/refractory malignancies in order to suggest the most adapted salvage treatment. From February 2016 to July 2020, 787 patients were included in France, Italy, Ireland, and Spain. At least one genetic alteration leading to a targeted treatment suggestion was identified in 436 patients (69%) with successful sequencing; 10% of these alterations were considered “ready for routine use.” Of 356 patients with follow-up beyond 12 months, 107 (30%) received one or more matched targeted therapies—56% of them within early clinical trials—mainly in the AcSé-ESMART platform trial (NCT02813135). Overall, matched treatment resulted in a 17% objective response rate, and of those patients with ready for routine use alterations, it was 38%. In patients with extracerebral tumors, 76% of actionable alterations detected in tumor tissue were also identified in circulating cell-free DNA (cfDNA).

SIGNIFICANCE: MAPPYACTS underlines the feasibility of molecular profiling at cancer recurrence in children on a multicenter, international level and demonstrates benefit for patients with selected key drivers. The use of cfDNA deserves validation in prospective studies. Our study highlights the need for innovative therapeutic proof-of-concept trials that address the underlying cancer complexity.

¹Department of Pediatric and Adolescent Oncology, Gustave Roussy Cancer Campus, Université Paris-Saclay, Villejuif, France. ²Unité de Génétique Somatique, Service de Génétique, Hospital Group, Institut Curie, Paris, France. ³Department of Pathology and Laboratory Medicine, Translational Research Laboratory and Biobank, AMMICA, INSERM US23/CNRS UMS3655, Gustave Roussy Cancer Campus, Université Paris-Saclay, Villejuif, France. ⁴INSERM U830, Laboratoire de Génétique et Biologie des Cancers, Research Center, PSL Research University, Institut Curie, Paris, France. ⁵Clinical Research Direction, Gustave Roussy Cancer Cam-

pus, Université Paris-Saclay, Villejuif, France. ⁶INSERM U1015, Gustave Roussy Cancer Campus, Université Paris-Saclay, Villejuif, France. ⁷Equipe SiRIC RTOP Recherche Translationnelle en Oncologie Pédiatrique, Institut Curie, Paris, France. ⁸Biostatistics and Epidemiology Unit, Gustave Roussy Cancer Campus, INSERM U1018, CESP, Université Paris-Saclay, Villejuif, France. ⁹Laboratory of Translational Research in Child and Adolescent Cancer, Vall d'Hebron Research Institute (VHIR)-UAB, Barcelona, Spain. ¹⁰Institut Curie Genomics of Excellence (ICGex) Platform, Research Center, Institut Curie, Paris, France. ¹¹Department of Pathology, PSL



Research University, Institut Curie, Paris, France. ¹²Department of Pediatric Oncology, Institut d'Hématologie et d'Oncologie Pédiatrique/Centre Léon Bérard, Lyon, France. ¹³Department of Pediatric Hematology and Oncology, Hôpital de La Timone, AP-HM, Marseille, France. ¹⁴UMR Inserm 1068, CNRS UMR 7258, Aix Marseille Université U105, Marseille Cancer Research Center (CRCM), Marseille, France. ¹⁵SIREDO Oncology Center (Care, Innovation and Research for Children and AYA with Cancer), Institut Curie, PSL Research University, Paris, France. ¹⁶Department of Pediatric Oncology, Centre Hospitalier Universitaire, Nantes, France. ¹⁷Pediatric Oncology Unit, Fondazione IRCCS Istituto Nazionale Tumori, Milano, Italy. ¹⁸Paediatric Haematology/Oncology, Children's Health Ireland, Crumlin, Dublin, Republic of Ireland. ¹⁹Division of Paediatric Haematology and Oncology, Vall d'Hebron Institut de Recerca (VHIR), Vall d'Hebron Hospital Universitari, Vall d'Hebron Barcelona Hospital Campus, Barcelona, Spain.

Note: Supplementary data for this article are available at Cancer Discovery Online (<http://cancerdiscovery.aacrjournals.org/>).

P. Berlanga, G. Pierron, and L. Lacroix are co-first authors.

G. Schleiermacher and B. Geoerger are co-last authors.

Prior presentation: This work was presented in part at the AACR Annual Meeting 2017 in Washington, DC; the AACR Annual Meeting 2019 in Atlanta, GA; the 2019 ASCO Annual Meeting in Chicago, IL; and the 51st Congress of the International Society for Paediatric Oncology (SIOP 2019) in Lyon, France.

Corresponding Author: Birgit Geoerger, Department of Pediatric and Adolescent Oncology, Gustave Roussy Cancer Campus, Université Paris-Saclay, 114 Rue Edouard Vaillant, 94805 Villejuif, France. Phone: 33-1-42-11-46-61; Fax: 33-1-42-11-52-75; E-mail: birgit.geoerger@gustaveroussy.fr

Cancer Discov 2022;12:1266-81

doi: 10.1158/2159-8290.CD-21-1136

This open access article is distributed under Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0 International (CC BY-NC-ND).

©2022 The Authors; Published by the American Association for Cancer Research

INTRODUCTION

Cancer remains the primary cause of disease-related mortality in children and adolescents (1). Comprehensive molecular profiling of tumors through high-throughput technologies identifies molecular targets and predictive biomarkers. Together with improved understanding of tumor biology and development of targeted anticancer agents, these approaches have facilitated therapeutic approaches adapted to cancer molecular profiles. This “cancer precision medicine” approach has now been implemented to guide treatment in patients with advanced malignancy (2–5), including children.

MAPPYACTS (Molecular Profiling for Pediatric and Young Adult Cancer Treatment Stratification; ClinicalTrials.gov identifier: NCT02613962) is a European international prospective precision medicine trial aiming to define the molecular profile of recurrent/refractory malignancies in pediatric and young adult patients in order to suggest the most adapted salvage treatment in the setting of further relapse/progression. The main objective of MAPPYACTS was to identify the proportion of patients who receive matched targeted treatments based on their individual molecular tumor profile at recurrence using whole-exome sequencing (WES) and RNA sequencing (RNA-seq). Secondary objectives were to describe the safety and feasibility of the on-purpose procedure of collecting cancer tissue in a multicentric setting and outcome of patients treated with matched targeted agents. Circulating cell-free DNA (cfDNA) analysis was explored as one main ancillary study to determine the feasibility of noninvasive assessment of tumor-related genomic alterations in patients with extracerebral tumors.

RESULTS

Patient and Procedure Characteristics

From January 2016 to July 2020, 787 patients were included in 18 centers following informed consent. Figure 1 depicts the detailed study flow. Thirteen patients consented to the MAPPYACTS trial but did not undergo a procedure to obtain cancer tissue and were considered screening failures. Thus, 774 patients underwent a biopsy, surgical tumor resection, and blood or bone marrow sampling as study procedure for cancer tissue collection. The median age at inclusion was 11.6 years (5th–95th percentile range: 2.2; 19.8; range, 0.5–38.5), and 59% were male. Study procedure was performed at a median of the first relapse or progression (range, 1–10). The median time since initial cancer diagnosis was 1.8 years (5th–95th percentile range: 0.4; 9.0; range, 0.1–32.0). Main tumor types were sarcomas (290 patients, 37%), central nervous system (CNS) tumors (216, 28%), other solid tumors (181, 23%), leukemia (54, 7%), and lymphomas (33, 4%; Table 1; Supplementary Table S1).

The 774 patients underwent 833 procedures for cancer tissue sample acquisition. Among patients with solid tumors and lymphomas, the procedure was performed exclusively for MAPPYACTS in 62%. Fifty-three patients (7%) underwent a second procedure and six a third procedure, most due to insufficient material, unsuccessful tumor sequencing, or progression on targeted treatment. Biopsy was the most frequent procedure performed in 55% patients; procedures were done

in 53% from metastatic sites. Sixty-seven procedure-related adverse events were reported in 55 surgical or biopsy procedures (6% of on-purpose and 9% of therapeutic/diagnostic ones), most frequently bleeding/hematoma and pneumothorax. Procedure-related adverse events were grade 1 (17), grade 2 (31), grade 3 (12), and grade 4 (7) in the 833 cases.

Molecular profiling was performed on 695 (84%) samples from 679 (88%) patients. For 138 samples (16%), no sequencing analysis was done, mainly because of low tumor cell content. Seven cases (1.1%) were considered secondary malignancies (all identified by pathology review performed on the sample, which served for sequencing). Six cases were classified as likely radiation-induced, and three were with a family history of cancer (Supplementary Table S2).

Molecular Tumor Board and Clinical Molecular Tumor Board Recommendations

Successful tumor sequencing was seen in 632 samples from 624 patients with WES, RNA-seq, and/or panel sequencing analysis (in three cases; Fig. 1 and Table 1): 91% (628/691) using WES or 90% (550/614) RNA-seq. Lower tumor cellularity and DNA/RNA quality were the main reasons for unsuccessful tumor sequencing.

Profiling coupled with pathology review suggested a revision of the initial diagnosis in 12 patients (1.9%) and in eight cases through the identification of specific gene fusions (Supplementary Table S2). Genetic counseling was recommended for 51 of 674 patients with a potentially relevant germline WES finding (7.6%).

We defined genetic somatic or germline alterations as “potentially actionable” when the detected molecular alteration or affected pathway in the patient’s tumor or germline analysis would be theoretically targetable by an approved or investigational agent, either directly or indirectly in the affected pathway. With this definition, “potentially actionable” therapeutic targets were identified among 436 of 632 (69%) samples in 432 of 624 (69%) patients. Of 1,144 “potentially actionable” findings, 533 were single-nucleotide variants (SNV; 484 somatic, 49 germline), 527 focal copy-number alterations (CNA; 212 amplifications/high-level gains, 315 deletions), 59 targetable gene fusions, and 25 elevated tumor mutational load (Figs. 2 and 3A and B).

One to seven (median: 2) treatment recommendations for targeted agents as single agent or in combination were given per patient involving mainly inhibiting agents of WEE1 ($n = 150$), mTOR ($n = 123$), CDK4/6 ($n = 105$), MEK ($n = 95$), PARP ($n = 64$), BET ($n = 59$), EZH2 ($n = 38$), FGFR ($n = 31$), and PD1/PDL1 ($n = 31$). Suggested targeted agents alone or in combination for the oncogenic alterations are presented in Fig. 4.

To guide treating physicians when prioritizing targets, we used an algorithm based on the ESMO Scale for Clinical Actionability of molecular Targets (ESCAT) evidence scale (27) that we adapted to the pediatric cancer context. It considers biological relevance, clinical evidence, and drug availability within clinical trials of targeting the molecular alteration. Forty-four of the 432 patients with potentially actionable alterations (10%) had a recommendation for the specific treatment at relapse that we considered as “ready for routine use” and for which significant clinical activity had been reported (6–18), that is, gene fusions involving *ALK* ($n = 10$), *BRAF*

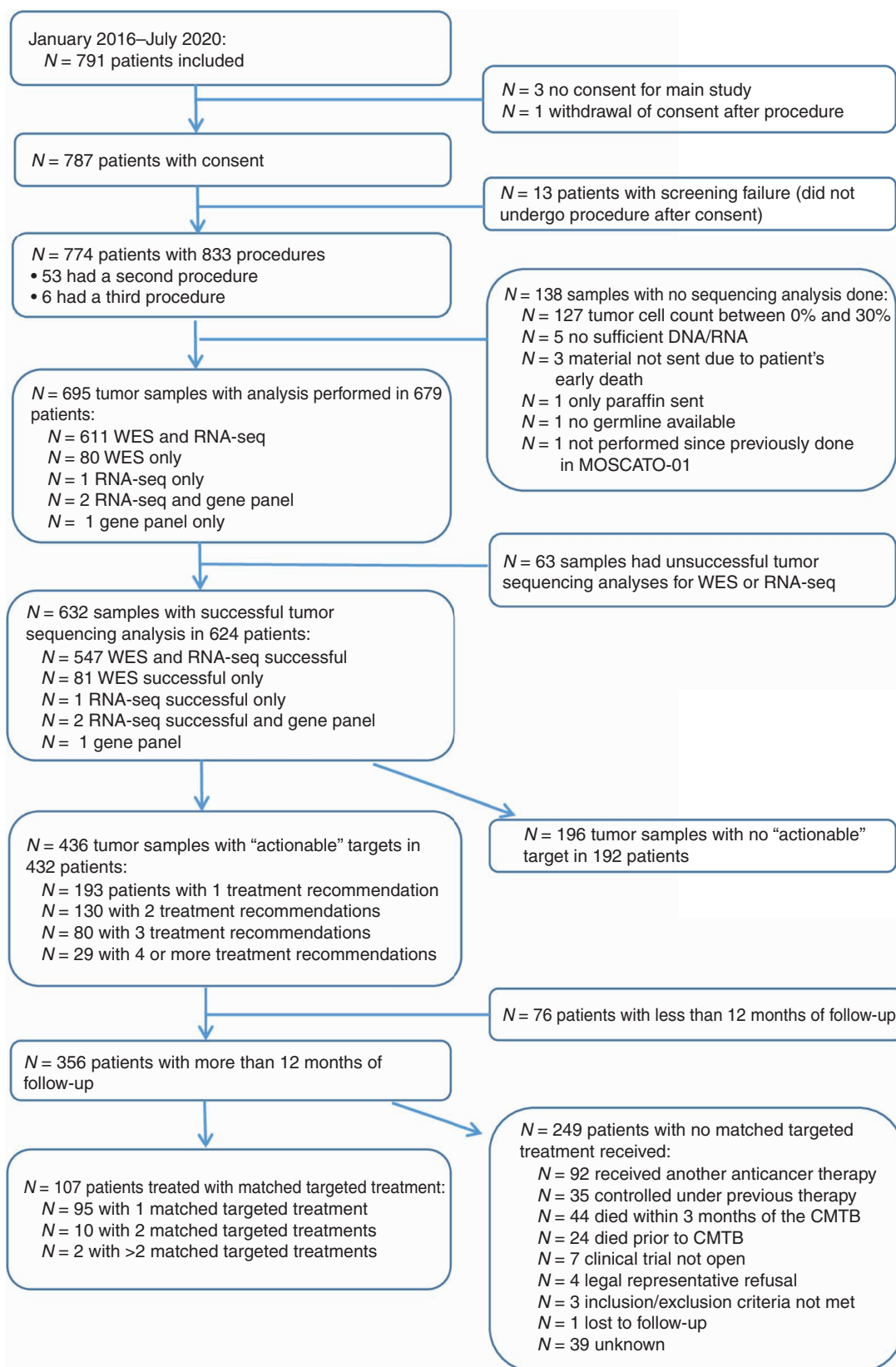


Figure 1. Study flow chart. Paired tumor and germline samples were analyzed in all patients with WES. CMTB, clinical molecular tumor board.

Table 1. Characteristics of patients, sampling study procedures, molecular profiling results, and matched treatments received per tumor types in the MAPPYACTS trial

Diagnoses	Patients n = 774	Study proce- dures n = 833	Successful		Ready for use alteration n = 44 (10%)	Investigational or hypothetical ^a alterations (in genes altered in >3 patients)	≥1 actionable alteration and FU >12 months n = 356 (82%)	Targeted matched treatment n = 107 (30%)
			Study proce- dures n = 833	sequencing analysis n = 632 (74%)				
SARCOMAS	290	317	248	165 (68)	5 (3)		139	39 (23)
Osteosarcoma	79	90	69 (77)	56 (83)	0	TP53, CDKN2A/B, EGFR/ERBB2/3/4, MYC, RBI, TP53, AKT1/2/3, CDK4, IGFLR, MAP2K4, NOTCH1/2/3/4, PTEN	50	14
Ewing sarcoma	71	74	56 (76)	23 (43)	0	CDKN2A/B, TP53	21	2
BCOR or CIC sarcoma	6	6	4 (66)	1 (25)	0		1	0
Other bone sarcoma	1	2	1 (50)	0 (0)	0		0	0
Rhabdomyosarcoma (RMS)	70	77	62 (81)	51 (85)	0	TP53, FGFR1/2/3/4, CDKN2A/B, MYCN, CDK4, PI3KCA	44	14
Alveolar RMS	34	37	30 (81)	22 (76)	0	TP35, MYCN, CDK4	20	10
Embryonal RMS	30	34	27 (79)	25 (96)	0	TP53, FGFR1/2/3/4, CDKN2A/B	20	3
RMS NOS	6	6	5 (83)	4 (80)	0		4	1
Non-RMS soft-tissue sarcoma (NRSTS)	63	68	56 (82)	34 (52)	0	SMARCB1, TP53, CDKN2A/B, NFI	25	9
DSRCT	9	9	7 (78)	1 (12)	0		0	0
MPNST	5	5	5 (100)	5 (100)	0	ROSI-GOPC (1) NFI	5	3
Synovial sarcoma	7	9	6 (67)	1 (17)	0		1	1
Undifferentiated sarcoma	8	9	8 (89)	6 (75)	0	NTRK3-ETV6 (1), NTRK1-LMNA (1)	5	1
Rhabdoid tumor	10	10	9 (90)	9 (100)	0	SMARCB1	5	0
Other NRSTS	24	26	21 (81)	12 (57)	0	ALK-MYH9 (1), COL1A1-PDGFB (1)	9	4
OTHER SOLID TUMORS	181	199	138 (69)	93 (67)	1 (1)		69	20 (21)
Neuroblastoma	104	117	73 (62)	55 (75)	0	MYCN, 11q, ALK, CDKN2A/B, elevated mutational rate, HRAS, TP53	37	8
Carcinoma	29	30	24 (77)	11 (48)	0	RET-CCDC6 (1)	11	2
Wilms tumor	27	29	24 (83)	16 (64)	0	TP53	13	7
Hepatoblastoma	8	8	8 (100)	3 (37)	0	TP53	2	1
Other solid tumors	13	15	9 (62)	8 (89)	0		6	2

CNS TUMORS	216	226	184 (82)	125 (56)	28 (22)	103	47 (38)
High-grade glioma	59	64	56 (87)	54 (96)	See subsections mid-line/non-midline	45	13
Midline	24	26	24 (92)	22 (92)	NTRK1-BCAN (1), NTRK2-TLE4/ KANK1/2 (3)	20	13
Non-midline	35	38	32 (84)	32 (100)	BRAF ^{V600E} (4), NTRK2- NACC2/TLE4 (2)	25	14
Low-grade glioma	23	24	17 (71)	17 (88)	BRAF-KIAA1549 (10), NFI germline (2), ROSI-GOPC (1)	12	2
Medulloblastoma	52	53	44 (83)	24 (55)	PTCHI (4) mut/del, TP53/SMO wild-type	20	10
Ependymoma	34	36	30 (83)	7 (23)	0	7	3
Infratentorial	23	25	20 (80)	2 (10)	0	2	1
Supratentorial	11	11	10 (91)	5 (50)	0	5	2
Atypical teratoid rhabdoid tumor	10	11	6 (55)	6 (100)	NTRK3-SPECC1L (1)	6	3
CNS germ cell tumors	7	7	5 (71)	4 (80)	0	3	0
Choroid plexus carcinoma	6	6	5 (83)	4 (80)	0	2	0
Other CNS tumors	25	25	21 (84)	7 (67)	0	5	2
LEUKEMIA	54	57	46 (81)	38 (81)	1 (3)	32	1 (1)
B-acute lymphoblastic leukemia	20	20	18 (90)	15 (83)	0	14	0
T-acute lymphoblastic leukemia	14	15	13 (97)	12 (92)	0	10	1
Acute myeloid leukemia	17	19	12 (63)	9 (69)	IDH1 ^{R132L}	6	0
Other leukemia	3	3	3 (100)	2 (67)	0	2	0
LYMPHOMA	33	34	16 (47)	15 (94)	9 (60)	13	0 (0)
Hodgkin lymphoma	6	6	0 (0)	0 (0)	0	0	0
Anaplastic large-cell lymphoma	15	16	9 (56)	9 (100)	ALK-NPM1/TFG/ TRAF1 (9)	8	0
Other non-Hodgkin lymphoma	12	12	7 (58)	6 (86)	0	5	0

Abbreviations: DSRCT, desmoplastic small round cell tumor; FU, follow-up; MPNST, malignant peripheral nerve sheath tumor; NOS, not otherwise specified.

^aHypothetical evidence-level alterations.

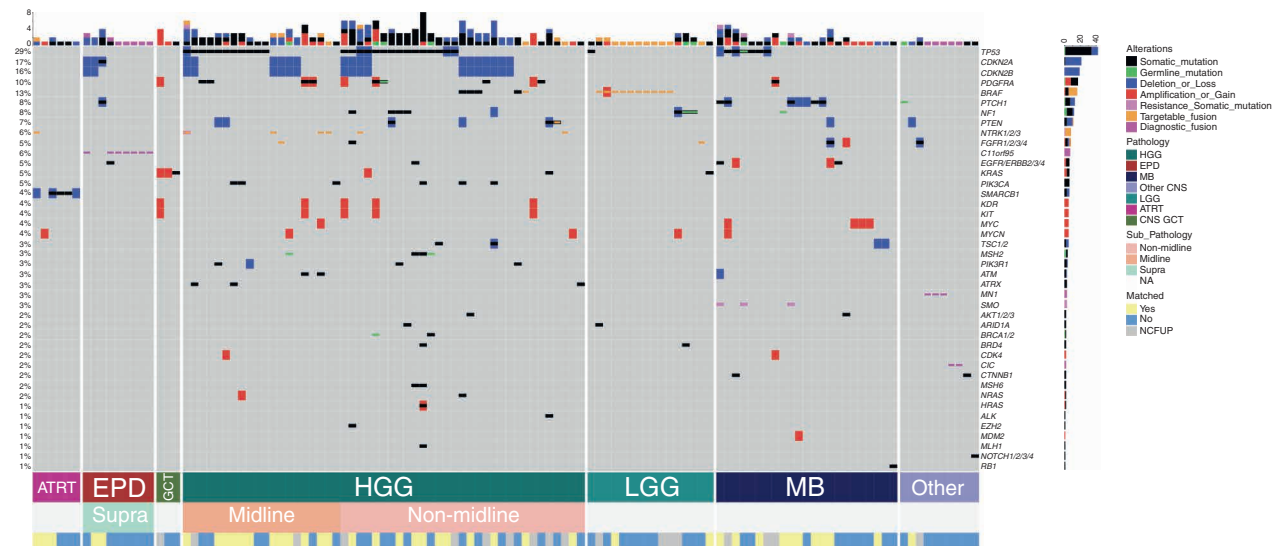


Figure 2. OncoPrint of potentially actionable alterations and canonical fusions in CNS tumors in 118 patients with 119 samples. Tumor types with five or fewer samples are grouped together in “Other.” Only alterations identified five or more times within the whole study are reported here. ATRT, atypical teratoid rhabdoid tumor; EPD, ependymoma; GCT, CNS germ cell tumor; HGG, high-grade glioma; LGG, low-grade glioma; matched, matched therapy received according to recommendations; MB, medulloblastoma; NA, not applicable; NCFUP, follow-up < 12 months; Other, other CNS tumors; subpathology, pathology subtype; supra, supratentorial.

(10), *NTRK1/2/3* (9), *ROS1* (2), *COL1A1/PDGF β* (1), and *RET* (1), pathogenic mutations in *BRAF* (4), *NF1* (2), and *IDH1* (1), or mutation or deletion in *PTCH1* (4) in selected tumor types (Table 1). Nineteen of them (42%) were previously unknown (Supplementary Table S3). Most alterations (909/1,144; 80%) were considered on the “investigational” level, which refers to oncogenic events for which the relevance of matched experimental treatments has been reported with limited activity or is being explored in the clinical setting. These alterations included gene amplifications, fusion, or activating mutations in *MYCN/MYC* (43), *KRAS/NRAS/HRAS* (42), *FGFR1/2/3/4* (27), *PIK3CA/PIK3R1/2* (23), *ALK* (15), and deletions or deleterious mutations in *TP53* (215), *CDK2NA/B* (157), *NF1* (33), *SMARCA4/SMARCB1* (27), *PTEN* (25), and *RBI* (11; Table 1). Ten cases were found with mutations in *NTRK* (3), *SMO* (3), *ALK* (3), and *NRAS* (1) that may be associated with treatment resistance; all except one had previously received the specific NTRK, SMO, ALK, or BRAF/MEK inhibitors, respectively, and benefited from the procedure at treatment failure.

These retained treatment suggestions were then discussed in the clinical molecular tumor board (CMTB) between experts in new drug development, experts in tumor diseases, and the treating physician considering the patient’s history and other treatment options that may be of relevance to the patient.

Patients’ Treatment and Follow-up after CMTB Recommendations

Following CMTB recommendations, patients with a minimum follow-up of 12 months (censored in September 2020) were analyzed. Three hundred fifty-six of the 432 patients with “potentially actionable” alterations had a minimum follow-up of 12 months and 107 (30%) of them had received 122 matched targeted therapies (range, 1–4/patient). Fourteen (11%) therapies were considered “ready for routine use,”

97 (80%) were “investigational,” and 11 (9%) “hypothetical.” Sixty-four (52%) treatments were administered as a single agent, 45 (37%) in combination with chemotherapy, and 13 (11%) combined with another targeted therapy. Sixty-eight (56%) of these treatments were performed within a phase I/II trial, 49 of them (72%) within AcSé-ESMART (NCT02813135).

Of the 249 patients with potentially actionable alterations, main reasons for not receiving the suggested matched treatment were death prior to the CMTB ($n = 24$, 10%) or within the 3 months that followed ($n = 44$, 18%), other nonmatched therapies ($n = 92$, 37%), or effective ongoing therapy ($n = 35$, 14%). Sixteen of the 44 patients (36%) with ready for routine use suggestions had received the recommended matched treatment previously—14 prior to the study procedure and two after the procedure and before the CMTB; in four of them, a second- or next-generation matched agent was administered after the CMTB.

Outcome of Patients Treated with Matched Therapies

Disease response was not evaluable in four patients because matched targeted treatment was received after complete tumor resection, and data were missing for nine patients. In 109 cases with evaluable or measurable disease and response evaluation, 18 were reported to have had partial response [PR; 17% objective response rate (ORR); 95% confidence interval (CI), 10%–25%] and 27 stable disease (SD; 25%), leading to a 41% disease control rate (DCR; 95% CI, 32%–51%). Median treatment duration for the 45 patients with controlled disease was 129 days (range, 58–697 days).

For patients with ready for routine use alterations, ORR was 38% (5/13; 95% CI, 18%–65%); all treatments were administered as single agents. Lower-level evidence recommendations (“investigational” and “hypothetical”) resulted in a 14% ORR (13/96; 95% CI, 32%–51%). The ORR for patients with



Figure 3. OncoPrint of potentially actionable alterations and canonical fusions in sarcoma in 213 patients with 215 samples (A) and other solid tumors in 86 patients with 86 samples (B). Tumor types with five or fewer samples are grouped together in "Other." Only alterations identified five or more times within the whole study are reported here. aRMS, alveolar rhabdomyosarcoma; CA, carcinoma; DSRCT, desmoplastic small round cell tumor; eRMS, embryonal rhabdomyosarcoma; EWS, Ewing sarcoma; matched, matched therapy received according to recommendations; MPNST, malignant peripheral nerve sheath tumor; NA, not applicable; NBL, neuroblastoma; NCFUP, follow-up < 12 months; NOS, not otherwise specified; NRSTS, non-rhabdomyosarcoma soft-tissue sarcoma; OS, osteosarcoma; Other, other solid tumor; RMS, rhabdomyosarcoma; RT, rhabdoid tumor; SS, synovial sarcoma; subpathology, pathology subtype; US, undifferentiated sarcoma; WT, Wilms tumor.

investigational alterations was 14% (12/86; 95% CI, 7%–23%), and for those with hypothetical alterations, it was 10% (1/10; 95% CI, 0%–44%). This study population treated with a matched targeted agent and chemotherapy combination had an ORR of 18% (8/44; 95% CI, 8%–33%), those of single-agent matched targeted treatment of 13% (5/39; 95% CI, 4%–27%), and those of targeted therapy combinations of 0% (0/13; 95% CI, 0%–24%).

Associated Ancillary Studies

Tumor Mutational Burden

Tumor mutational burden (TMB) was recalculated retrospectively on harmonized raw data in the first 451 enrolled patients with solid malignancies and 452 successfully sequenced WES tumor samples. A median of 0.6 mutations per megabase (mut/Mb; range, 0–195.3 mut/Mb; Fig. 5) was found. Seven patients exhibited >10 mut/Mb [three high-grade glioma (one of them >100 mut/Mb), three neuroblastoma, one medulloblastoma], and three were in the context of a probable, previously unknown constitutional mismatch repair deficiency involving *MSH2*, *MLH1*, and *MSH6* genes. Five tumors displayed microsatellite instability (Supplementary Fig. S1).

cfDNA and Circulating Tumor DNA

To explore the role of liquid biopsies and the possibility of detecting tumor-specific molecular alterations in advanced pediatric diseases, plasma samples at the time of tumor tissue collection were obtained in all cases. Among the first 500 enrolled patients, cfDNA extracted from the plasma of 234 patients with extracerebral solid tumors was analyzed (Supplementary Fig. S2). Nine had technical failure, and 225 were considered for further analysis. cfDNA quantities showed significant differences among different cancer types (ANOVA test: $P = 1.4e-03$), with the highest cfDNA quantities observed in the 43 patients with neuroblastoma compared with the 182 other tumors (Wilcoxon test: $3e-04$; Fig. 6A). The circulating tumor DNA (ctDNA) content in cfDNA, as evaluated by bioinformatics methods, also varied significantly between cancer types (ANOVA test: $P = 2.35e-05$), with highest values in patients with neuroblastoma compared with other disease types: median 35% ctDNA content (range, 0%–95%) versus median 16% (range, 0%–86%), respectively (Wilcoxon test: $1.4e-08$; Fig. 6B).

Among 190 patients for whom tumor WES analysis was deemed successful, cfDNA WES was considered successfully sequenced in 128 of 190 samples (67%) and was more frequent

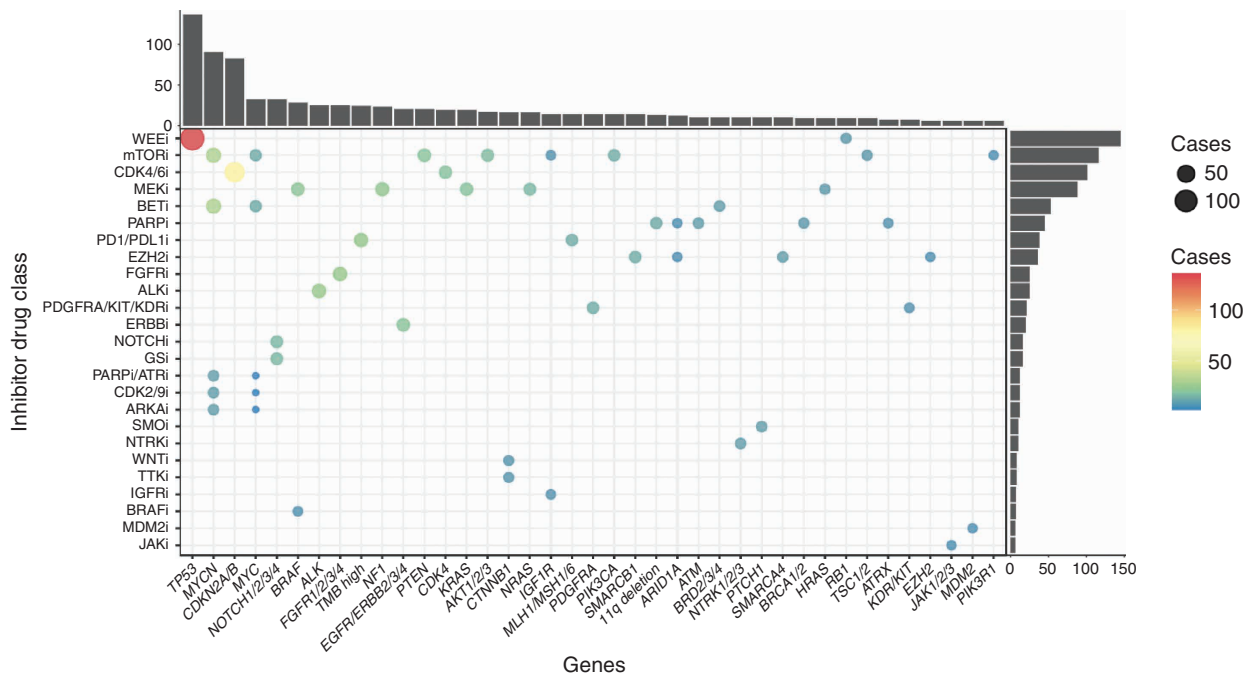


Figure 4. Matched targeted treatment recommendations per genetic alterations (only one alteration by gene and those occurring in more than five cases are represented). GSi, gamma-secretase inhibitor; PD1/PDL1i, programmed cell death protein 1/ligand immune checkpoint inhibitors; TMB, tumor mutational burden.

in patients with metastases than localized disease (66% vs. 47%, respectively, χ^2 P value: 0.04). Among these 128 cfDNA samples, cfDNA WES showed both somatic CNAs and SNVs in 51 cases, whereas in the remaining 77 cases, only somatic SNVs were observed. Detailed analysis of the SNVs indicated that among the 128 cases, a median of 15 somatic SNVs (range, 0–103) were common to plasma and tumor, with a median of six somatic SNVs (range, 0–127) observed only in tumor and a median of one somatic SNV (range, 0–460) only in plasma, with no significant differences between different tumor types (Supplementary Fig. S3). In this patient group (190 patients with extracerebral solid tumors and successful tumor WES analysis), a total of 94 SNVs were considered potentially actionable. Of these, 71 SNVs observed in the tumor were also identified in plasma by cfDNA WES (76%; Fig. 6C and D). Importantly, among 14 of these patients, 35 somatic SNVs in potentially actionable genes were observed by cfDNA WES that were not detected by tumor WES (median per patient 1; range, 1–13). This included, among others, an *ALK* p.Arg1275Gln mutation seen only in the plasma (Fig. 6D).

Concerning the 35 patients for whom tumor WES was either not done ($n = 16$) or unsuccessfully sequenced ($n = 19$), cfDNA WES was considered successful in 11 patients (34%), with a total of 12 SNVs targeting actionable genes identified in five patients.

DISCUSSION

MAPPYACTS reports the feasibility and outcome of an international precision medicine trial in pediatric and young adult patients performing on-purpose cancer tissue collection for molecular characterization of recurrent or refractory

malignancies. The study is the first to explore the role of liquid biopsy for cfDNA analysis in this population.

Consistent with our preliminary monocentric experiences (19, 20) and other reports (19, 21–26), 69% of patients in MAPPYACTS had one or more genomic alterations in their tumor that we considered “potentially actionable” through targeted agents. Importantly, the study procedures proved safe, and 30% of the patients subsequently underwent a matched targeted treatment.

We show that 4% of all defined potentially actionable gene alterations in 10% of patients were considered “ready for routine use,” and the matched single-agent targeted treatment resulted in a 38% ORR in patients receiving this treatment after the CMTB, which was significantly higher than in lower-level evidence treatments (14%). In addition, out of 12 evaluable patients who received such a matched treatment prior to MAPPYACTS inclusion, seven had a PR and two a CR (75% ORR), resulting globally in a 56% ORR (95% CI, 37%–73%) for matched therapies for this highest evidence-level category. This is in line with recent data of the INFORM registry (26). Most of them are identical to our highest-level alterations, albeit we have not considered for example *HRAS/KRAS* and *ALK* mutations because of the limited clinical activity of MEK and ALK inhibitors in patients with these alterations (35–38). Importantly, in half of our patients, the identified high-level evidence gene alterations were not identified in previous routine molecular diagnostics, underlining the importance of more comprehensive molecular profiling in matching rare genomic alterations with available effective targeted therapies. Search for high-level evidence gene alterations should now be part of front-line molecular diagnostics and will allow us to introduce active new targeted agents into standard treatment.

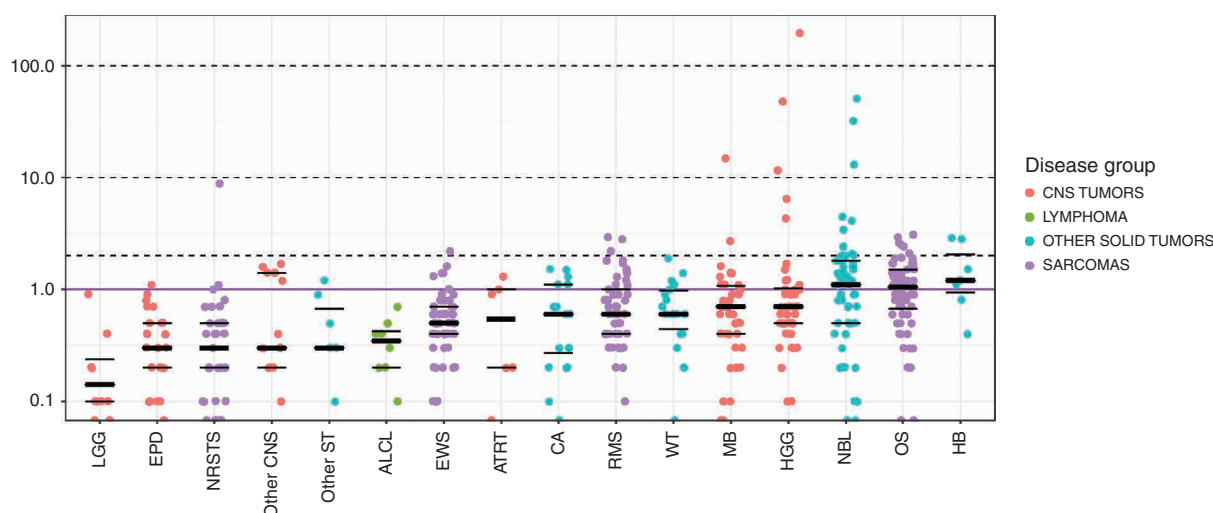


Figure 5. Tumor mutational load in main cancer types (only disease types with more than five cases are presented). Median mutational loads and quartiles 25 and 75 are shown as black solid lines for each tumor type. Samples with 2 to <10 mut/Mb are considered pediatric high, 10 to <100 hypermutators, and >100 ultrahypermutators. ALCL, anaplastic large cell lymphoma; ATRT, atypical teratoid rhabdoid tumor; CA, carcinoma; EPD, ependymoma; EWS, Ewing sarcoma; HB, hepatoblastoma; HGG, high-grade glioma; LGG, low-grade glioma; MB, medulloblastoma; NBL, neuroblastoma; NRSTS, non-RMS soft-tissue sarcoma; OS, osteosarcoma; Other ST, other solid tumors; RMS, rhabdomyosarcoma; WT, Wilms tumor.

An additional direct clinical impact through profiling and pathology review was found in 3.0% of patients with diagnostic modifications, mainly through detection of gene translocations in CNS tumors and sarcomas, or a secondary cancer diagnosis. In the INFORM registry, a change or refinement of diagnosis was identified in 8.2% of cases, which can in part be explained through the inclusion of samples at diagnosis (26). These observations again confirm the importance of performing selected gene DNA analysis as well as RNA-seq ± methylation in diagnostically challenging cases.

Furthermore, we detected mutations known to confer resistance to targeted therapies in 10 of our patients; nine of them had progressed on the specific targeted treatment, highlighting the clinical relevance of considering molecular profiling at targeted treatment failure to define resistance mechanisms and adapt future salvage strategies.

Overall ORR of patients in MAPPYACTS was 17% and 14% for lower-level evidence treatments, thus superior to the reported 4% ORR observed in pediatric phase I/II chemotherapy trials (27) and similar to the outcome in the targeted therapies era (28). The Zero Childhood Cancer Program reported a 31% ORR (11/35 patients); however, half of the patients were included at diagnosis, and the ORR for patients at the time of disease recurrence was not given (29).

This raises the debate on the definition of “actionable” or “potentially actionable” alterations, which has been used inconsistently throughout the programs. For this clinical research trial, taking into account the low numbers of high-level evidence alterations, we wished not only to limit the report to clearly defined oncogenic driver events with straightforward treatment recommendations but also to describe alterations in genes that could lead to a potential benefit according to preclinical findings or clinical data in adult cancers. Importantly, 96% of our reported “potentially actionable” oncogenic events matched with treatment suggestions were considered at an “investigational” (80%) or “hypothetical”

(16%) evidence level. Albeit these alterations are well-known oncogenic events, their direct targeting, alone or in combination, has not been demonstrated with significant clinical activity, the alteration–treatment match is currently evaluated in clinical trials, or their match has been suggested in preclinical data, respectively. The high frequency of these often multiple alterations, absence of relevant clinical data, and lack of curative options in children and adolescents with relapsed/refractory malignancies led us to run a parallel project to tumor profiling, which explores targeted molecules or combinations in molecularly enriched patient populations. The “Secured Access–European proof-of-concept therapeutic Stratification trial of Molecular Anomalies in Relapsed or refractory Tumors” (AcSé-ESMART) platform trial (NCT02813135) started recruitment in July 2016, is currently open in five countries, and has recruited more than 190 patients, 78% of them referred through the MAPPYACTS trial, in addition to other European profiling programs. The primary objective is to detect signals of activity and potential new biomarkers. Treatment arms in AcSé-ESMART mainly address targeting of alterations in the cell cycle, the PI3K/mTOR pathway, homologous DNA repair, and immune checkpoint (30–32). TP53 alterations were considered investigational in MAPPYACTS based on data in ovarian cancer, suggesting that TP53 deficiency possibly associated with response to WEE1 inhibitors (33), and which we explored in combination with chemotherapy in the trial. The medical need for such clinical trials is reflected by the fact that 56% of the matched treatments in MAPPYACTS patients could be administered within the context of phase I/II clinical trials and 72% of them had been included in AcSé-ESMART. Preliminary data for the ongoing INFORM registry reported 28% (11% in a clinical trial) and the NCI–COG Pediatric Molecular Analysis for Therapeutic Choice (MATCH) trial (NCT03155620) 35% of patients treated with a matching drug (25, 34). The fact that most advanced cancers have no unique key drivers but rather

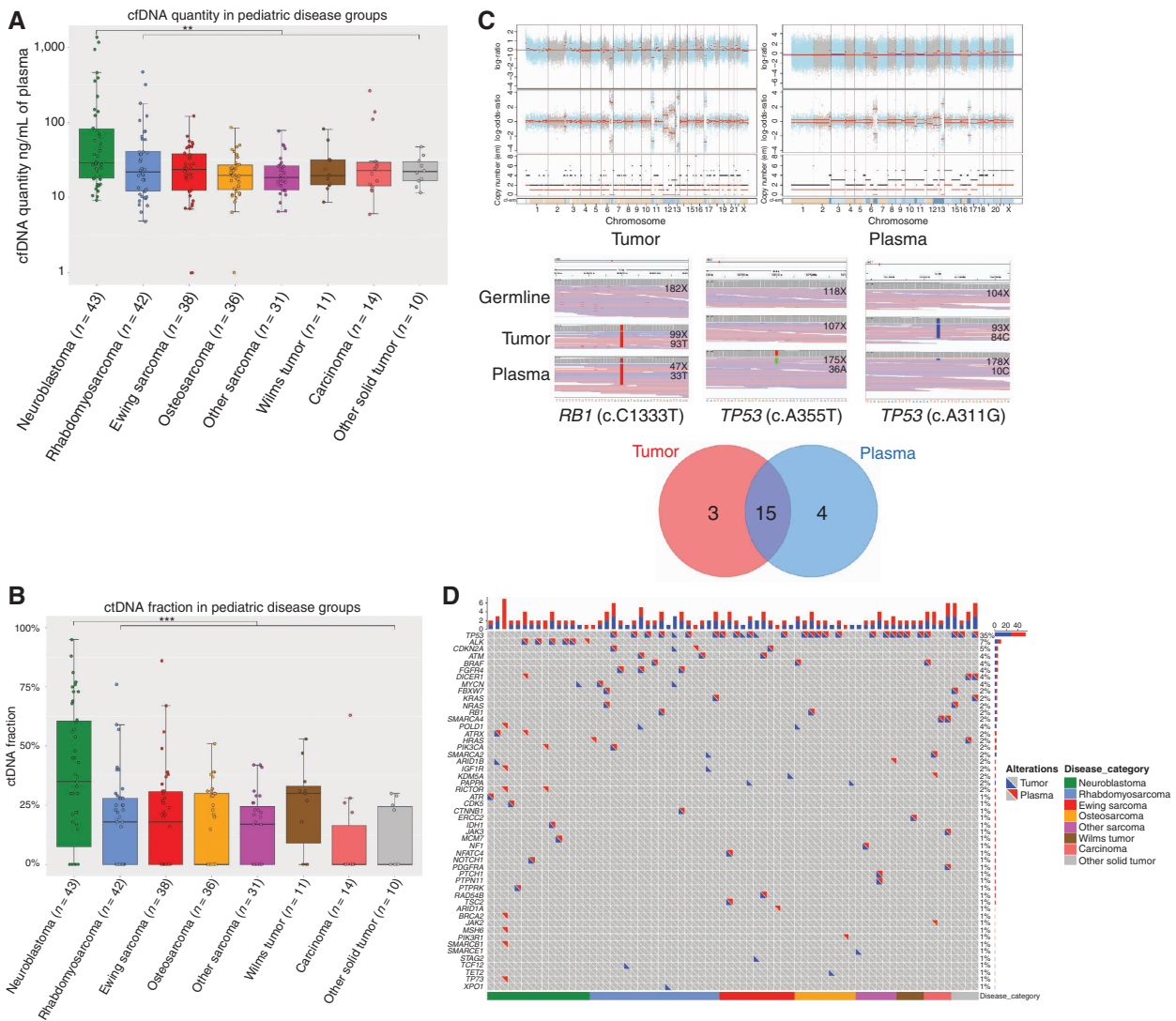


Figure 6. cfDNA and ctDNA ancillary study in 225 patients with extracranial solid tumors. **A**, cfDNA quantity per type of tumor (ng/mL of plasma, log₁₀ scale). **, indicates statistically significant difference (Wilcoxon test: 3e-04). **B**, ctDNA fraction calculated by the Facet tool after WES for each type of tumor. ***, indicates statistically significant difference (Wilcoxon test: 1.4e-08). **C**, Example of a sequencing result for a patient with an alveolar rhabdomyosarcoma. On the top, copy-number profiles generated by the Facet tool, showing the same chromosome alterations identified in the tumor/plasma. Middle, RB1 c.C1333T and TP53 c.A311G mutations were identified in tumor and plasma, whereas TP53 c.A355T was observed only in the plasma. Bottom, Venn diagram of SNVs detected in each type of samples with a good overlap (15 SNVs common between tumor and plasma). **D**, Oncoprint of SNVs in targeting potentially actionable genes detected in tumor (blue) and plasma (red) for 72 cases with both successful tumor and cfDNA analysis (only cases with at least one SNV in an actionable gene are reported here). Histograms on the top indicate the total number of SNVs in actionable genes in each case (blue, tumor; red, plasma); histograms on the right indicate the percentage of cases with alterations in the indicated actionable gene.

multiple oncogenic events in various cancer hallmarks (35) demonstrates the underlying cancer complexity to which current clinical trials are not yet perfectly adapted. Therefore, we wish to stress the importance of detailed comprehensive tumor board discussions for pondering the subjectivity of the treatment suggestions. Such discussions between physician-scientists, experts in new drug development and tumor diseases, as well as the treating physicians must consider the potential relevance of the selected targets based on current knowledge of tumor biology and outcome in clinical trials, the expertise on novel anticancer strategies, availability of the drug, availability of an open clinical trial, as well as other treatment options that may be relevant for the patient.

This consensus proposition must be shared in a comprehensive way with patients and parents so they may decide on the participation of such a treatment option given the uncertainty of their outcome. We insist that whenever possible these experimental treatments should be administered and explored within investigational clinical trials to improve knowledge and optimize safety for these patients. Furthermore, the time point of integration of advanced profiling in each patient's care is critical. MAPPYACTS is to date the only pediatric trial where the tumor sample acquisition intervention was performed within the study. Most of our patients were included at first relapse/progression and received tumor-specific first salvage treatment while waiting for the molecular profiling results.

Although theoretically it would be preferable to treat patients based on the sequencing results of the current relapse, avoiding further temporal tumor evolution, this pragmatic approach was deemed suitable for most patients and, as done in most other programs, with advanced molecular profiling (25, 26, 29).

MAPPYACTS further confirms the incidence of germline findings and cancer predisposition syndromes in 7% to 12% of pediatric patients, consistent with the data of others (19, 26, 36–38). The variation in numbers may relate to different predefined lists of known predisposition genes that were used. Furthermore, our data may suggest that TMB may be higher at disease recurrence than in newly diagnosed patients, as reported previously in a lower number of patients (39). Future comparison of paired primary and relapse tumor samples from the same patients will be necessary to confirm these findings and correlate these with treatment. All findings support the clinical approach of performing tumor profiling at recurrence or treatment failure.

MAPPYACTS is the first study to explore cfDNA analysis as a noninvasive technique to detect actionable alterations in recurrent pediatric and adolescent non-CNS solid tumors. Higher cfDNA quantities, and a higher content of ctDNA in cfDNA, were observed in patients with neuroblastoma compared with other disease types, confirming previous studies of the feasibility of ctDNA studies in these patients with different analytical techniques such as low-coverage whole-genome sequencing, WES, or droplet digital PCR (40). Other processes such as inflammation, systemic infection, or even pregnancy can lead to higher cfDNA content; however, the described pipeline aimed to identify tumor cell-specific genetic alterations specifically. Overall, cfDNA WES led to successful sequencing results in 62% of cases. Unsuccessful sequencing might be linked to low ctDNA content, which might be observed in patients with a low overall tumor burden. Different technical limitations might explain a low sequencing success rate in some cases. Given the approach by cfDNA WES, this study is limited to the detection of SNVs. Importantly, in 77 of 126 cases, although no CNAs could be identified, somatic SNVs could be detected, based on the higher sensitivity of WES for detection of SNVs (threshold for detection: 5% mutated allele fractions) than CNA (threshold: 20%; ref. 41). Furthermore, different capture techniques between the tumor versus plasma analysis hampered CNA analysis. However, the different capture techniques were accounted for in the bioinformatics pipeline, enabling the reliable calling of SNVs, with a detection limit of 1%, as reported previously (41).

Among all detected somatic SNVs in cases with successful cfDNA WES analysis, 57% were seen in both the cfDNA and the tumor, with 31% and 11% specific to the tumor and the cfDNA, respectively, possibly reflecting tumor heterogeneity. Tumor heterogeneity has been widely reported in pediatric cancers, with both spatial heterogeneity of the primary tumor itself, or genetic heterogeneity between the primary and metastatic sites. Based on our data, cfDNA analysis provides an important tool for the exploration of genetic heterogeneity in solid pediatric malignancies at the time of relapse. Altogether, somatic SNVs were detected in all analyzed tumor types, highlighting the importance of cfDNA WES, which might result in a different perspective of the disease. Although the cfDNA WES analysis was performed as an ancillary study and did

not guide treatment recommendations, cfDNA WES identified 76% of all actionable alterations found in the matched solid non-CNS tumors. Our study underlines that in some instances, like neuroblastoma, where osteomedullary relapses are often not successfully sequenced for WES, or in patients without possibility of a new tumor biopsy, cfDNA extracted from plasma might lead to clinically relevant results. Furthermore, recent reports demonstrate the feasibility of cfDNA extracted from CSF for patients with CNS malignancies (42).

An additional ancillary research study in MAPPYACTS was the development of patient-derived xenografts (PDX). During the 4 years of the study, nine participating research laboratories have established 131 xenografts (43). The characterization of the PDX and their comparison with the primary tumors are ongoing. They represent a valuable tool for future research studies, and most of them have been shared with the Innovative Medicines Initiative (IMI2) ITCC-P4 project (<https://www.itccp4.eu>).

The MAPPYACTS trial was a dynamic process, undergoing continuous adaptation since commencing in 2016. CMTB interpretations and recommendations were made to the best of our knowledge at a given date, representing one limitation of the trial. Evidence levels for most targets will change based on current scientific knowledge, which is rapidly evolving and informed by ongoing research. Various and multiple oncogenic events that are not unique drivers in the diseases contribute to the complexity of cancers, and their biological relevance as well as their therapeutic targeting is mostly undefined. In addition, a limited understanding of the function of gene variants of unknown significance and a lack of insight about posttranslational and immunologic contexts are major limits of current precision medicine programs.

Our study further highlights these observations. The sequencing efforts on pediatric tumors at relapse underscore the low proportion of high-level evidence or “ready for routine use” alterations. This strongly underlines that high-throughput molecular profiling together with matched targeted therapies, innovative clinical trials, development of new technologies that allow cancer characterization beyond the genetic level, development of combination strategies, and analysis of the personal immune response of each patient are needed to overcome the current limitations.

In 2020, very high-throughput sequencing using WGS 80×, WES 150×, and RNA-seq has been introduced into the standard of care at relapse or treatment failure for children and adults in France through the Plan France Médecine Génomique 2025 (PFMG2025), launched by the Haute Autorité de Santé (HAS) and supported by the French Ministry of Health within the Plan Cancer 3. However, we believe that high-throughput molecular profiling and a better understanding of tumor biology, leading to matched targeted combination treatments, linking to clinical interventional studies, and defining novel technologies and innovative treatment strategies, will be needed to improve the survival of most high-risk patients. This will be explored within the MAPPYACTS 2 study that will complement the PFMG2025 project and include the validation of liquid biopsies and cfDNA analyses as well as studies on the health economic impact and psychosocial outcomes related to these analyses and treatments. In conclusion, MAPPYACTS underlines the feasibility of molecular profiling

at the time of pediatric cancer recurrence on a multicenter international level. Although selected high evidence-level alterations should be part of the initial diagnostic workup, cancer complexity justifies the continuous efforts and introduction of high-throughput sequencing and treatment recommendations as a standard of care for high-risk cancers. MAPPYACTS has identified future innovative diagnostic and treatment strategies, including the encouraging results of cfDNA analysis in solid extracerebral tumors, which are deserving of validation in further prospective studies.

METHODS

Study Population

Eligible patients were aged ≥ 6 months with recurrent/refractory solid tumors or leukemia, ≤ 18 years at cancer diagnosis, independent of the age at the time of disease recurrence, and potentially eligible for an early-phase clinical trial. In the setting of solid tumors, the lesion had to be accessible for biopsy or surgical resection. Patients were required to have adequate performance status, bone marrow, and organ function. Written informed consent was signed by the patient or parents/legal representatives, and assent of the minor child according to local laws to perform the procedure and molecular analysis of the tumor and blood sample, as well as optional ancillary studies, was obtained. Detailed inclusion/exclusion criteria are available in Supplementary Methods.

Study Design and Procedure

The MAPPYACTS trial recruited from February 2016 until July 2020 in 15 French centers, Italy (from October 2017), Ireland (May 2018), and Spain (September 2019). Tumor biopsy or surgery of the recurrent/refractory malignancy, or bone marrow/blood sample if leukemia, was performed following written informed consent and inclusion in order to collect cancer tissue for molecular profiling specifically for this study. In parallel to the procedure, a blood sample for constitutional DNA extraction and cfDNA was collected. Molecular analyses were performed in the French National Cancer Institute (INCa)-labeled next-generation sequencing platforms Gustave Roussy, Institut Curie, and Centro Nacional de Análisis Genómico Barcelona. In the setting of unsuccessful tumor sample sequencing or progression during targeted treatment (Amendment 5), a new procedure could be offered and performed after new written consent was obtained.

The trial was approved by independent ethics committees and national medical authorities and conducted according to the principles of the Declaration of Helsinki.

Adverse Event Reporting Related to the Study Procedure

All adverse events definitely/probably/possibly related to the MAPPYACTS study procedure were assessed by the local investigator and reported according to the Common Terminology Criteria for Adverse Events (CTCAE v4.0).

Sample Analysis

Tumor cellularity in specimens from the sample used for nucleic acid extraction was determined by an experienced pathologist; those with $\geq 30\%$ tumor cellularity were processed. Tumor DNA, RNA, and germline DNA from whole blood samples were extracted using the AllPrep DNA/RNA Mini Kit and DNeasy Blood and Tissue Kit.

WES

Sequencing libraries were constructed according to standard procedures from 600 ng of tumor and paired constitutional DNA. WES was captured using the Agilent SureSelect V5 (50 Mb), Clinical Research Exome (54 Mb) kit, SureSelect XT human All exon CRE

version 1 or 2, or Twist Human Core Exome Enrichment System. Sequencing of subsequent libraries was performed using Illumina sequencers (NextSeq 500 or HiSeq 2000/2500/4000) in 75-bp paired-end mode, aiming for a mean depth of coverage of 100 \times .

Bioinformatics processing was based on Illumina Pipeline (CASAVA1.8) or in-house pipelines consisting in alignment of sequencing reads on the hg19 human genome (Build37), determination of structural variants, CNAs, and variant detection, as well as determination of their functional impact (19, 41). In case of low cellularity, gene panel sequencing could be performed (19).

RNA-seq

Libraries were prepared with TruSeq Stranded mRNA kit, PolyA mRNA capture with oligo dT beads 1 mg total RNA, fragmentation to approximately 400 bp, cDNA double-strand synthesis, and ligation of adaptors, library amplification, and sequencing (2, 19). For the optimized detection of potential fusion transcripts, an in-house designed metacaller approach was used (details on sequencing and bioinformatics in Supplementary Methods; refs. 16–20).

Mutational Tumor Load

Sequencing library quality was estimated with fastqc and fastqscreen. Reads were mapped with BWA (v0.7.17 with parameters: -M -A 2 -E) onto the human reference genome assembly hg19/GRCh37. SNVs and small indels were called using GATK3 (Indel Realigner, Base Recalibrator), samtools (fixmate, markdup, mpileup), and Varscan (v2.3.9) from paired normal/tumors bam files. Variant annotation was performed with ANNOVAR using the public database released on November 7, 2019, from the 1000 Genomes Project, Exome Aggregation Consortium (ExAC), NHLBI-ESP project, and Kaviar. Functional prediction of variants was performed using the data set dbnsfp30a. Questionable somatic variants observed in less than three reads, with an allele frequency lower than 0.05, described in the 1000 Genomes and ExAC databases with a frequency higher than 0.05%, or nonexonic variants were excluded.

Somatic coding mutations were filtered according to their enrichment in the tumor samples compared with the paired normal samples as reported previously (30). Mutational load was calculated as the number of nonsynonymous somatic variants divided by the total length of targeted regions by the exome capture kits with a minimum coverage of 10 \times .

cfDNA

A blood sample for study of cfDNA was collected on EDTA following inclusion in the MAPPYACTS study (mean delay between biopsy of the tumor and blood sample 3.4 days; range, 0–42 days). For patients with non-CNS solid tumors, cfDNA extracted from plasma was quantified using Qubit HS DNA assays for WES at 100 \times (41). ctDNA content in cfDNA was calculated using the Facets tools. cfDNA WES successful sequencing analysis was defined as a WES profile resulting in any CNA and/or at least three somatic SNVs also seen in the tumor (Supplementary Methods).

Molecular Abnormality Reporting and Treatment Decision

Sequencing results were reviewed by a molecular geneticist, summarized in a report, and discussed with the physician-scientist core team during weekly MTBs. A successfully sequenced tumor sample was considered when WES identified somatic CNAs or somatic SNVs, when filtering on matched germline, or when RNA-seq identified tumor cell-specific fusion transcripts. SNVs were retained if pathogenic or likely pathogenic. For tumor suppressor genes, homozygous loss-of-function variants and focal deletions were considered; in genes like *TP53* and *ATM*, heterozygous loss-of-function was also reported. Furthermore, heterozygous chr11q deletions encompassing the homologous repair genes *ATM*, *CHK1*, *MRE11A*, and *H2AFX*

were also considered. For oncogenes, focal amplifications and high-level gains were retained. Germline findings were reported in cancer predisposition genes, as published by Zhang and colleagues (38).

In order not to restrict reporting to only a few recurrent molecular events and to provide treatment recommendations for the majority of patients, a wide definition of molecular alterations highlighted in the MTBs was applied. The term “potentially actionable” refers to a detected molecular alteration or affected pathway in the patient’s tumor and/or germline analysis that theoretically would be targetable by an approved or investigational agent either directly or indirectly in the affected pathway. Prioritization was based on the clinical evidence of targeting the molecular alteration and considering accessibility of the treatment. Alterations were considered “ready for routine use” if the referred alteration–drug match had been associated with antitumor activity, defined as >30% ORR, or improved outcome in clinical trials in a given tumor type or similar indication (6, 8–14, 16, 18, 44–46). Oncogenic events for which the relevance of experimental treatments was reported with limited activity and/or is being addressed in clinical trials were considered “investigational,” and those for which only preclinical data exist “hypothetical.” Known solvent front, gatekeeper, or xDFG mutations were classified as “resistance” mutations.

MTB results were then discussed with the treating physician in weekly CMTBs, and recommendations were summarized in a CMTB report that ranked therapeutic options according to available evidence of the retained molecular alterations but included other treatments relevant for the patient at this disease stage. Germline abnormalities were discussed in consenting patients/families; oncogenetic counseling was suggested in case of clear pathogenicity.

Outcome Assessments

For patients receiving recommended matched targeted therapy, tumor response assessed every 6 to 8 weeks from treatment start, according to the standard in each tumor entity [RECIST 1.1, Response Assessment in Neuro-Oncology (RANO), International Neuroblastoma Response Criteria (INRC); refs. 47–49], was recorded. ORR was defined as the proportion of patients who achieved a PR or CR and DCR as ORR and SD.

Data Sharing Agreement

Sequencing data and basic clinical annotations have been deposited in European Genome-phenome Archive (EGA; hosted by the EBI and CRG) with the data set accession code EGAS00001005935. Further information about EGA can be found on <https://ega-archive.org> (“The European Genome-phenome Archive of human data consented for biomedical research”; <http://www.nature.com/ng/journal/v47/n7/full/ng.3312.html>). Additional, more detailed clinical data from the clinical trial can be requested by completing the data request form for Gustave Roussy clinical trials (<https://redcap.gustaveroussy.fr/redcap/surveys/?s=DYDTLPE4AM>) for validation by the trial steering committee and the sponsor prior to transfer of detailed clinical data.

Authors’ Disclosures

P. Berlanga reports grants from Imagine for Margo, Institut National du Cancer, Fondation ARC, Fédération Enfants et Santé, Société Française de lutte contre les Cancers et les Leucémies de l’Enfant et l’adolescent (SFCE), and Dell during the conduct of the study, as well as other support from EUSA Pharma, Bayer, and Bristol Myers Squibb outside the submitted work. G. Pierron reports grants from the Annenberg Foundation, Association Hubert Gouin Enfance et Cancer, Association Meghanora, Fédération Enfants Cancers et Santé, and Imagine for Margo during the conduct of the study. Y. Iddir reports grants from the Annenberg Foundation, Association Hubert Gouin Enfance et Cancer, Association Meghanora, Fédération Enfants Cancers et Santé, and Imagine for Margo during the conduct of the study. A. Soriano Fernandez reports grants from

Fundación FERO and the Rotary Clubs Barcelona Eixample, Barcelona Diagonal, Santa Coloma de Gramanet, München-Blutenburg, Deutschland Gemeindienst e.V. and others from Barcelona and province, and the AECC (Spanish Association Against Cancer) during the conduct of the study. N. André reports grants, nonfinancial support, and other support from Bristol Myers Squibb; other support from Pierre Fabre and Akina Pharmaceuticals; and nonfinancial support and other support from Bayer outside the submitted work. I. Aerts reports grants from the Annenberg Foundation, Association Hubert Gouin Enfance et Cancer, Association Meghanora, Fédération Enfants Cancers et Santé, and Imagine for Margo during the conduct of the study. M. Casanova reports advisory roles for Roche, Bayer, AstraZeneca, Servier, and Pfizer. S. Michiels reports personal fees from statistical advice to IDDI, Janssen, Amaris, and Roche and is a data and safety committee monitoring member of clinical trials for Sensorion, Biophytis, Servier, and Yuhan outside the submitted work. G. Vassal reports grants from the French NCI, Fondation ARC, and Imagine for Margo during the conduct of the study and has provided advice on pediatric oncology drug development to AstraZeneca, Bayer, Bristol Myers Squibb, Celgene, Hutchison-Medi Pharma, Ipsen, Lilly, Merck, Novartis, Pfizer, and Roche/Genentech (did not accept personal remuneration). G. Schleiermacher reports grants from the Annenberg Foundation, Association Hubert Gouin Enfance et Cancer, Association Meghanora, Fédération Enfants Cancers et Santé, and Imagine for Margo during the conduct of the study, as well as grants and nonfinancial support from Bristol Myers Squibb, grants from Pfizer, MSDavenir, Roche, INCa PRTK, Imagine for Margo, Fédération Enfants Cancers et Santé, Association Hubert Gouin Enfance et Cancer, the Annenberg Foundation, ARC Fondation ARC pour la recherche sur le cancer, and Ligue Nationale contre le Cancer outside the submitted work. B. Georger reports grants from Imagine for Margo, Institut National du Cancer, Fondation ARC, Fédération Enfants et Santé, Société Française de lutte contre les Cancers et les Leucémies de l’Enfant et l’adolescent (SFCE), and Dell during the conduct of the study, as well as other support from AstraZeneca and Boehringer Ingelheim outside the submitted work. No disclosures were reported by the other authors.

Authors’ Contributions

P. Berlanga: Conceptualization, data curation, formal analysis, validation, investigation, methodology, writing—original draft, writing—review and editing. **G. Pierron:** Conceptualization, data curation, formal analysis, validation, investigation, methodology, writing—review and editing. **L. Lacroix:** Conceptualization, data curation, formal analysis, validation, investigation, methodology, writing—review and editing. **M. Chicard:** Data curation, formal analysis, validation, investigation, methodology, writing—original draft, writing—review and editing. **T. Adam de Beaumais:** Data curation, formal analysis, validation, investigation, project administration, writing—review and editing. **A. Marchais:** Data curation, software, formal analysis, validation, investigation, writing—review and editing. **A.C. Harttrampf:** Data curation, validation, investigation, writing—review and editing. **Y. Iddir:** Data curation, software, formal analysis, validation, investigation, writing—original draft, writing—review and editing. **A. Larive:** Data curation, software, methodology, writing—review and editing. **A. Soriano Fernandez:** Data curation, software, formal analysis, validation, investigation, writing—review and editing. **I. Hezam:** Conceptualization, supervision, project administration, writing—review and editing. **C. Chevassus:** Data curation, software, formal analysis, visualization, methodology, project administration, writing—review and editing. **V. Bernard:** Data curation, investigation, writing—review and editing. **S. Cotteret:** Data curation, software, formal analysis, validation, investigation, methodology, writing—review and editing. **J.-Y. Scoazec:** Data curation, formal analysis, validation, investigation, methodology, writing—review and editing. **A. Gauthier:**

Data curation, software, formal analysis, investigation, writing–review and editing. **S. Abbou:** Data curation, software, writing–review and editing. **N. Corradini:** Data curation, software, methodology, writing–review and editing. **N. André:** Data curation, validation, investigation, writing–review and editing. **I. Aerts:** Data curation, validation, investigation, writing–review and editing. **E. Thebaud:** Data curation, validation, writing–review and editing. **M. Casanova:** Data curation, validation, writing–review and editing. **C. Owens:** Data curation, validation, writing–review and editing. **R. Hladun-Alvaro:** Data curation, investigation, writing–review and editing. **S. Michiels:** Conceptualization, data curation, software, formal analysis, supervision, validation, investigation, methodology, writing–review and editing. **O. Delattre:** Data curation, investigation, writing–review and editing. **G. Vassal:** Conceptualization, resources, data curation, software, supervision, methodology, writing–review and editing. **G. Schleiermacher:** Conceptualization, resources, data curation, software, formal analysis, supervision, funding acquisition, validation, investigation, visualization, methodology, writing–original draft, writing–review and editing. **B. Geoerger:** Conceptualization, resources, data curation, software, formal analysis, supervision, funding acquisition, validation, investigation, visualization, methodology, writing–original draft, writing–review and editing.

Acknowledgments

We are grateful to all patients and their parents who participated in the trial as well as to the treating clinical teams of the 18 hospitals and their referring centers (Gustave Roussy, Villejuif, France; Institut Curie, Paris, France; Centre Léon Bérard, Lyon, France; Hôpital La Timone, Marseille, France; CHU Nantes, France; Centre Oscar Lambret, Lille, France; CHU Bordeaux, France; Hôpital Trousseau, Paris, France; Strasbourg, France; CHU Nancy, France; CHU Toulouse, France; Children’s Health Ireland, Crumlin, Ireland; CHU Angers, France; Hôpital Robert Debré, Paris, France; Fondazione IRCCS Istituto Nazionale dei Tumori di Milano, Italy; Vall d’Hebron Barcelona Hospital Campus, Spain; Hôpital Jeanne de Flandre, Lille, France; Hospices Civils de Lyon, France). We thank the teams of the surgery and interventional radiology departments, local pathology departments, biobanks, and biology platforms at Gustave Roussy (Catherine Richon, Helene Rocheteau, Zsofia Balogh, Noémie Pata-Merci, Adeline Perez, Marie Xiberras, Mathias Marques, Marc Deloger, and Yannick Boursin), Institut Curie (Megan Bouvet, Stelly Ballet, Camille Benoist, Eleonore, Frouin, Eve Lapouble, and Nathalie Clement), and Vall d’Hebron Institut de Recerca (VHIR), Vall d’Hebron Hospital Universitari, Barcelona (Soledad Gallego, Gabriela Guillén, Sergio López, Marta Garrido, Alexandra Navarro, Ainara Magdaleno, and Ivo Gut and the CNAG team), as well as the national cosponsoring teams and the sponsoring clinical team of Gustave Roussy (Delphine Vuillier-Le Goff, Thibaud Motreff, and Salim Laghouati). We are highly thankful to all of the clinicians and researchers who contributed to the MTB discussions, particularly Jacques Grill, Hélène Cavé, Hélène Lapillonne, Jessica Zucman-Rossi, Eric Le Touzé, Eric Pasmant, and Josh Waterfall. We are highly grateful to all funders. This study was supported by grants from Institut National du Cancer (INCa) through the PHRC “INCa-DGOS_8519” MERRI, Fondation ARC, Association Imagine for Margo, Fédération Enfants et Santé, the Société Française de lutte contre les Cancers et les leucémies de l’Enfant et l’adolescent (SFCE), and Dell. ctDNA analyses performed at Institut Curie were supported by grants from the Annenberg Foundation, Association Hubert Gouin Enfance et Cancer, and Association Meghanora. Analyses performed at Vall d’Hebron Research Institute/Centre Nacional d’Anàlisi Genòmica were supported by grants from Fundació FERO and the Rotary Clubs Barcelona Eixample, Barcelona Diagonal, Santa Coloma de Gramanet, München-Blutenburg, Deutschland Gemeindienst e.V. and others from Barcelona and province. B. Geoerger is supported by the “Parrainage médecin-chercheur” of Gustave Roussy.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received August 20, 2021; revised November 3, 2021; accepted February 7, 2022; published first March 16, 2022.

REFERENCES

- Gatta G, Botta L, Rossi S, Aareleid T, Bielska-Lasota M, Clavel J, et al. Childhood cancer survival in Europe 1999–2007: results of EUROCARE-5—a population-based study. *Lancet Oncol* 2014;15:35–47.
- Massard C, Michiels S, Ferté C, Le Deley MC, Lacroix L, Hollebecque A, et al. High-throughput genomics and clinical outcome in hard-to-treat advanced cancers: results of the MOSCATO 01 trial. *Cancer Discov* 2017;7:586–95.
- Le Tourneau C, Delord JP, Gonçalves A, Gavaille C, Dubot C, Isambert N, et al. Molecularly targeted therapy based on tumour molecular profiling versus conventional therapy for advanced cancer (SHIVA): a multicentre, open-label, proof-of-concept, randomised, controlled phase 2 trial. *Lancet Oncol* 2015;16:1324–34.
- Stockley TL, Oza AM, Berman HK, Leighl NB, Knox JJ, Shepherd FA, et al. Molecular profiling of advanced solid tumors and patient outcomes with genotype-matched clinical trials: the Princess Margaret IMPACT/COMPACT trial. *Genome Med* 2016;8:109.
- Trédan O, Wang Q, Pissaloux D, Cassier P, de la Fouchardière A, Fayette J, et al. Molecular screening program to select molecular-based recommended therapies for metastatic cancer patients: analysis from the ProfILER trial. *Ann Oncol* 2019;30:757–65.
- Mossé YP, Voss SD, Lim MS, Rolland D, Minard CG, Fox E, et al. Targeting ALK with crizotinib in pediatric anaplastic large cell lymphoma and inflammatory myofibroblastic tumor: a Children’s Oncology Group Study. *J Clin Oncol* 2017;35:3215–21.
- Geoerger B, Schulte J, Zwaan CM, Casanova M, Fischer M, Moreno L, et al. Phase I study of ceritinib in pediatric patients (Pts) with malignancies harboring a genetic alteration in ALK (ALK+): safety, pharmacokinetic (PK), and efficacy results. *J Clin Oncol* 33, 2015 (suppl; abstr 10005).
- Hargrave DR, Bouffet E, Tabori U, Broniscer A, Cohen KJ, Hansford JR, et al. Efficacy and safety of dabrafenib in pediatric patients with BRAF V600 mutation-positive relapsed or refractory low-grade glioma: results from a phase I/IIa study. *Clin Cancer Res* 2019;25:7303–11.
- Hargrave DR, Moreno L, Broniscer A, Bouffet E, Aerts I, Andre N, et al. Dabrafenib in pediatric patients with BRAF V600-positive high-grade glioma (HGG). *J Clin Oncol* 36, 2018 (suppl; abstr 10505).
- Fangusaro J, Onar-Thomas A, Young Poussaint T, Wu S, Ligon AH, Lindeman N, et al. Selumetinib in paediatric patients with BRAF-aberrant or neurofibromatosis type 1-associated recurrent, refractory, or progressive low-grade glioma: a multicentre, phase 2 trial. *Lancet Oncol* 2019;20:1011–22.
- Hong DS, DuBois SG, Kummar S, Farago AF, Albert CM, Rohrberg KS, et al. Larotrectinib in patients with TRK fusion-positive solid tumours: a pooled analysis of three phase 1/2 clinical trials. *Lancet Oncol* 2020;21:531–40.
- Robinson GW, Gajjar AJ, Gauvain KM, Basu EM, Macy ME, Maese LD, et al. Phase 1/1B trial to assess the activity of entrectinib in children and adolescents with recurrent or refractory solid tumors including central nervous system (CNS) tumors. *J Clin Oncol* 37, 2019 (suppl; abstr 10009).
- Drilon AE, DuBois SG, Farago AF, Geoerger B, Grilley-Olson JE, Hong DS, et al. Activity of larotrectinib in TRK fusion cancer patients with brain metastases or primary central nervous system tumors. *J Clin Oncol* 37, 2019 (suppl; abstr 2006).
- Kieran MW, Chisholm J, Casanova M, Brandes AA, Aerts I, Bouffet E, et al. Phase I study of oral sonidegib (LDE225) in pediatric brain and solid tumors and a phase II study in children and adults with relapsed medulloblastoma. *Neuro Oncol* 2017;19:1542–52.
- Robinson GW, Orr BA, Wu G, Gururangan S, Lin T, Qaddoumi I, et al. Vismodegib exerts targeted efficacy against recurrent sonic hedgehog-

- subgroup medulloblastoma: results from phase II pediatric brain tumor consortium studies PBTC-025B and PBTC-032. *J Clin Oncol* 2015;33:2646–54.
16. Vassal G, Faivre L, Geoerger B, Plantaz D, Auvrignon A, Coze C, et al. Crizotinib in children and adolescents with advanced ROS1, MET, or ALK-rearranged cancer: results of the AcSé phase II trial. *J Clin Oncol* 34, 2016 (suppl; abstr 11509).
 17. Gerdemann U, Lee YA, Henry D, Smith S, Ortiz MV, Rothenberg SM, et al. First experience of LOXO-292 in the management of pediatric patients with RET-altered cancers. *J Clin Oncol* 37, 2019 (suppl; abstr 10045).
 18. DiNardo CD, Stein EM, de Botton S, Roboz GJ, Altman JK, Mims AS, et al. Durable remissions with ivosidenib in IDH1-mutated relapsed or refractory AML. *N Engl J Med* 2018;378:2386–98.
 19. Harttrampf AC, Lacroix L, Deloger M, Deschamps F, Puget S, Auger N, et al. Molecular screening for cancer treatment optimization (MOSCATO-01) in pediatric patients: a single-institutional prospective molecular stratification trial. *Clin Cancer Res* 2017;23:6101–12.
 20. Pincez T, Clément N, Lapouble E, Pierron G, Kamal M, Bieche I, et al. Feasibility and clinical integration of molecular profiling for target identification in pediatric solid tumors. *Pediatr Blood Cancer* 2017;64:e26365.
 21. Mody RJ, Prensner JR, Everett J, Parsons DW, Chinnaiyan AM. Precision medicine in pediatric oncology: lessons learned and next steps. *Pediatr Blood Cancer* 2017;64:e26288.
 22. Harris MH, DuBois SG, Glade Bender JL, Kim A, Crompton BD, Parker E, et al. Multicenter feasibility study of tumor molecular profiling to inform therapeutic decisions in advanced pediatric solid tumors: the individualized cancer therapy (iCat) study. *JAMA Oncol* 2016;2:608.
 23. Worst BC, van Tilburg CM, Balasubramanian GP, Fiesel P, Witt R, Freitag A, et al. Next-generation personalised medicine for high-risk paediatric cancer patients – the INFORM pilot study. *Eur J Cancer* 2016;65:91–101.
 24. Khater F, Vairy S, Langlois S, Dumoucel S, Sontag T, St-Onge P, et al. Molecular profiling of hard-to-treat childhood and adolescent cancers. *JAMA Netw Open* 2019;2:e192906.
 25. Parsons DW, Janeway KA, Patton D, Coffey B, Williams PM, Hamilton SR, et al. Identification of targetable molecular alterations in the NCI-COG Pediatric MATCH trial. *J Clin Oncol* 37, 2019 (suppl; abstr 10011).
 26. van Tilburg CM, Pfaff E, Pajtlér KW, Langenberg KPS, Fiesel P, Jones BC, et al. The pediatric precision oncology INFORM registry: clinical outcome and benefit for patients with very high-evidence targets. *Cancer Discov* 2021;11:2764–79.
 27. Kim A, Fox E, Warren K, Blaney SM, Berg SL, Adamson PC, et al. Characteristics and outcome of pediatric patients enrolled in phase I oncology trials. *Oncologist* 2008;13:679–89.
 28. Cohen JW, Akshintala S, Kane E, Gnanapragasam H, Widemann BC, Steinberg SM, et al. A systematic review of pediatric phase I trials in oncology: toxicity and outcomes in the era of targeted therapies. *Oncologist* 2020;25:532–40.
 29. Wong M, Mayoh C, Lau LMS, Khuong-Quang D-A, Pinese M, Kumar A, et al. Whole genome, transcriptome and methylome profiling enhances actionable target discovery in high-risk pediatric cancer. *Nat Med* 2020;26:1742–53.
 30. Pasqualini C, Rubino J, Brard C, Cassard L, André N, Rondof W, et al. Phase II and biomarker study of programmed cell death protein 1 inhibitor nivolumab and metronomic cyclophosphamide in paediatric relapsed/refractory solid tumours: arm G of AcSé-ESMART, a trial of the European Innovative Therapies for Children with Cancer Consortium. *Eur J Cancer* 2021;150:53–62.
 31. Bautista F, Paoletti X, Rubino J, Brard C, Rezai K, Nebchi S, et al. Phase I or II study of ribociclib in combination with topotecan-temozolomide or everolimus in children with advanced malignancies: arms A and B of the AcSé-ESMART Trial. *J Clin Oncol* 2021;39:3546–60.
 32. Morscher RJ, Brard C, Berlanga P, Marshall LV, André N, Rubino J, et al. First in child phase I/II study of the dual mTORC1/2 inhibitor vistusertib (AZD2014) as monotherapy and in combination with topotecan-temozolomide in children with advanced malignancies: Arms E and F of the AcSé-ESMART trial. *Eur J Cancer* 2021;157:268–77.
 33. Leijen S, van Geel RMJM, Pavlick AC, Tibes R, Rosen L, Razak ARA, et al. Phase I study evaluating WEE1 inhibitor AZD1775 as monotherapy and in combination with gemcitabine, cisplatin, or carboplatin in patients with advanced solid tumors. *J Clin Oncol* 2016;34:4371–80.
 34. Van Tilburg CM, Pfaff E, Pajtlér KW, Langenberg K, Fiesel P, Jones BC, et al. The pediatric precision oncology study INFORM: clinical outcome and benefit for molecular subgroups. *J Clin Oncol* 38, 2020 (suppl; abstr LBA10503).
 35. Hanahan D. Hallmarks of cancer: new dimensions. *Cancer Discov* 2022;12:31–46.
 36. Parsons DW, Roy A, Yang Y, Wang T, Scollon S, Bergstrom K, et al. Diagnostic yield of clinical tumor and germline whole-exome sequencing for children with solid tumors. *JAMA Oncol* 2016;2:616.
 37. Mody RJ, Wu YM, Lonigro RJ, Cao X, Roychowdhury S, Vats P, et al. Integrative clinical sequencing in the management of refractory or relapsed cancer in youth. *JAMA* 2015;314:913.
 38. Zhang J, Walsh MF, Wu G, Edmonson MN, Gruber TA, Easton J, et al. Germline mutations in predisposition genes in pediatric cancer. *N Engl J Med* 2015;373:2336–46.
 39. ICGC PedBrain-Seq Project, ICGC MMML-Seq Project, Gröbner SN, Worst BC, Weischenfeldt J, Buchhalter J, et al. The landscape of genomic alterations across childhood cancers. *Nature* 2018;555:321–7.
 40. Chicard M, Boyault S, Colmet Daage L, Richer W, Gentien D, Pierron G, et al. Genomic copy number profiling using circulating free tumor DNA highlights heterogeneity in neuroblastoma. *Clin Cancer Res* 2016;22:5564–73.
 41. Chicard M, Colmet-Daage L, Clement N, Danzon A, Bohec M, Bernard V, et al. Whole-exome sequencing of cell-free DNA reveals temporo-spatial heterogeneity and identifies treatment-resistant clones in neuroblastoma. *Clin Cancer Res* 2018;24:939–49.
 42. Escudero L, Llorc A, Arias A, Diaz-Navarro A, Martínez-Ricarte F, Rubio-Perez C, et al. Circulating tumour DNA from the cerebrospinal fluid allows the characterisation and monitoring of medulloblastoma. *Nat Commun* 2020;11:5376.
 43. Marques Da Costa ME, Daudigeos-Dubus E, Surdez D, Marchais A, Dessen P, Scoazec JY, et al. Patient-derived xenografts (PDX) development in MAPPYACTS - a pediatric precision cancer medicine trial in relapsed and refractory tumors [abstract]. In: Proceedings of the 51st Congress of the International Society of Paediatric Oncology (SIOP 2019); 2019 Oct 23–26; Lyon, France. Geneva (Switzerland): SIOP; 2019. Abstract nr SIOP19-1241 (oral presentation).
 44. Schulte JH, Moreno L, Ziegler DS, Marshall LV, Zwaan CM, Irwin M, et al. Final analysis of phase I study of ceritinib in pediatric patients with malignancies harboring activated anaplastic lymphoma kinase (ALK). *J Clin Oncol* 38, 2020 (suppl; abstr 10505).
 45. Ortiz MV, Gerdemann U, Raju SG, Henry D, Smith S, Rothenberg SM, et al. Activity of the highly specific RET inhibitor selpercatinib (LOXO-292) in pediatric patients with tumors harboring RET gene alterations. *JCO Precis Oncol* 2020;4:341–7.
 46. Navarrete-Dechent C, Mori S, Barker CA, Dickson MA, Nehal KS. Imatinib treatment for locally advanced or metastatic dermatofibrosarcoma protuberans: a systematic review. *JAMA Dermatol* 2019;155:361.
 47. Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, et al. New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur J Cancer* 2009;45:228–47.
 48. Park JR, Bagatell R, Cohn SL, Pearson AD, Villablanca JG, Berthold F, et al. Revisions to the International Neuroblastoma Response Criteria: a consensus statement from the National Cancer Institute Clinical Trials Macdonald Meeting. *J Clin Oncol* 2017;35:2580–7.
 49. Wen PY, Macdonald DR, Reardon DA, Cloughesy TF, Sorensen AG, Galanis E, et al. Updated response assessment criteria for high-grade gliomas: Response Assessment in Neuro-Oncology Working Group. *J Clin Oncol* 2010;28:1963–72.