

Supplementary information for:

**A unicellular cyanobacterium relies on sodium energetics to fix N<sub>2</sub>**

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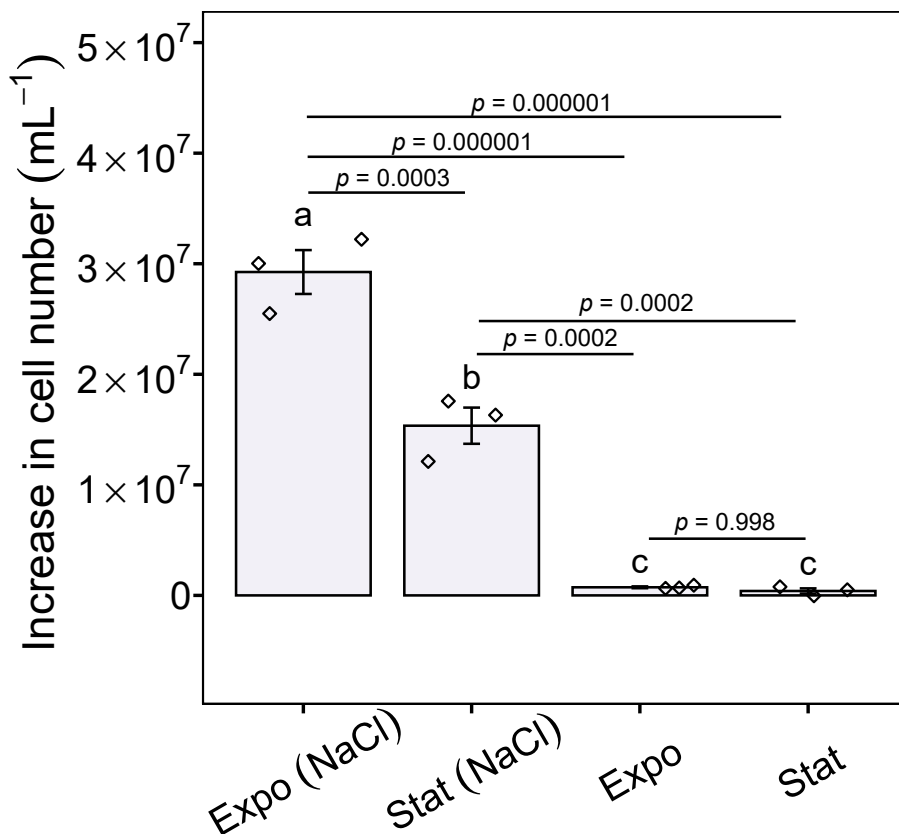
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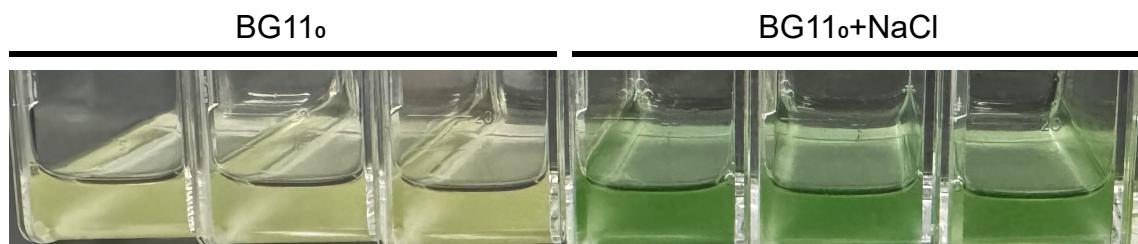
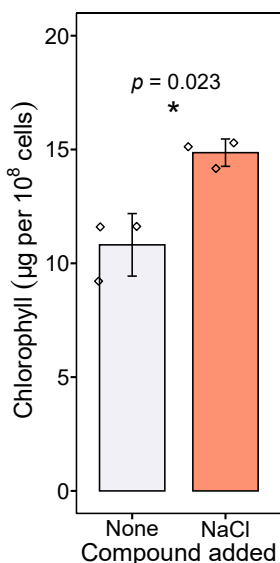
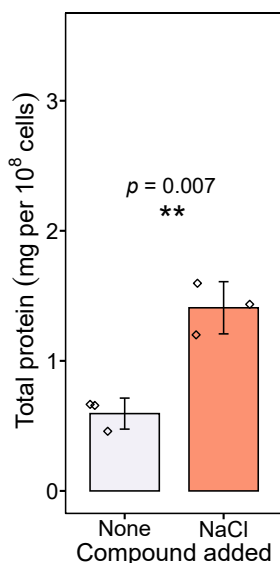
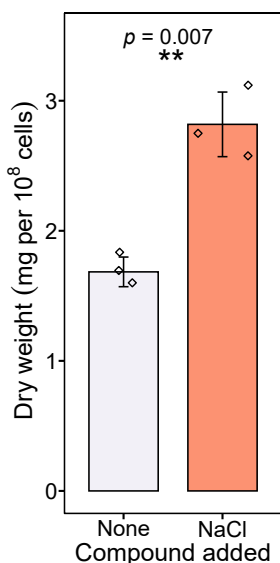
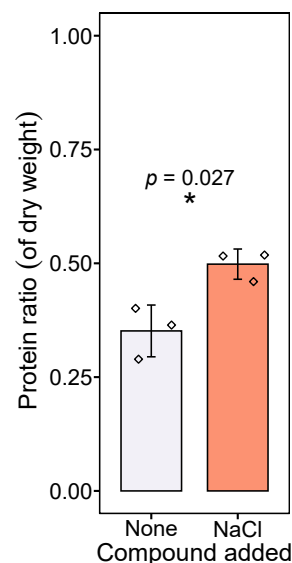
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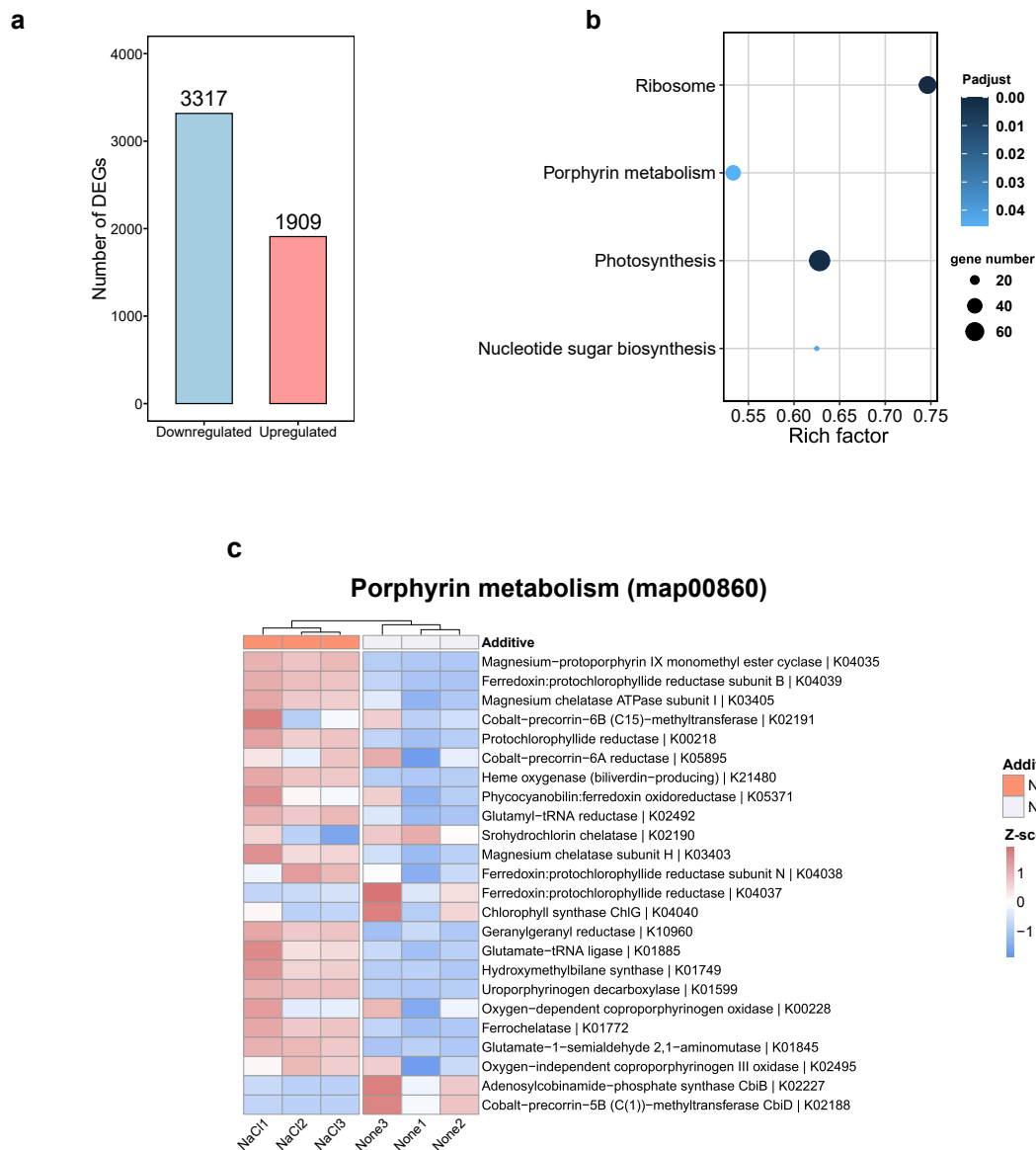
Supplementary figures 1-8



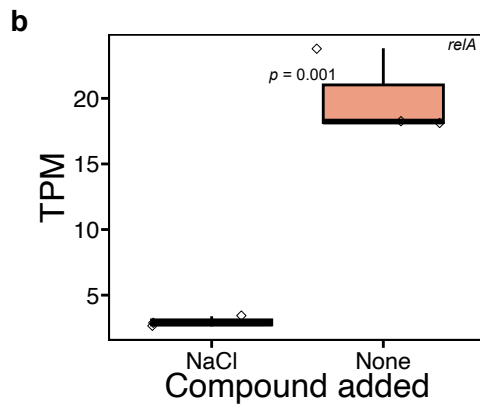
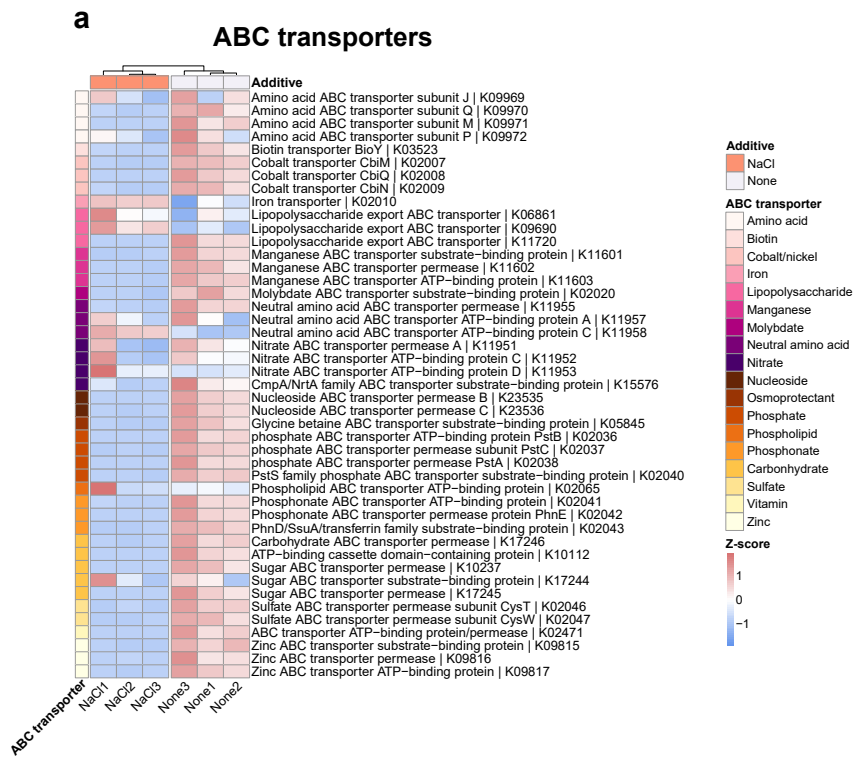
**Supplementary Fig. 1. Population growth over seven days from different growth stages under N deprivation.** Cells were grown in BG11<sub>o</sub> with or without NaCl (18 g/L). Expo (NaCl), exponential phase cells with NaCl; Stat (NaCl), stationary phase cells with NaCl; Expo, exponential phase cells without NaCl; Stat, stationary phase cells without NaCl. Exponential phase cells refer to cultures 7 days post inoculation, whereas stationary phase refers to 20-day-old cultures post inoculation. Notably, N<sub>2</sub>-fixing stationary phase cells grew significantly slower than exponential phase cells in the presence of NaCl. This is because stationary phase cells require more time to recover growth and therefore have a longer lag time. The graph shows the mean  $\pm$  standard deviation ( $n = 3$  biologically independent samples). Different letters on each bar indicate statistical significance ( $p < 0.05$ ) calculated by one-way ANOVA with Tukey's HSD post-hoc analysis over all populations tested. Source data are provided as a Source Data file.

**a****b****c****d****e**

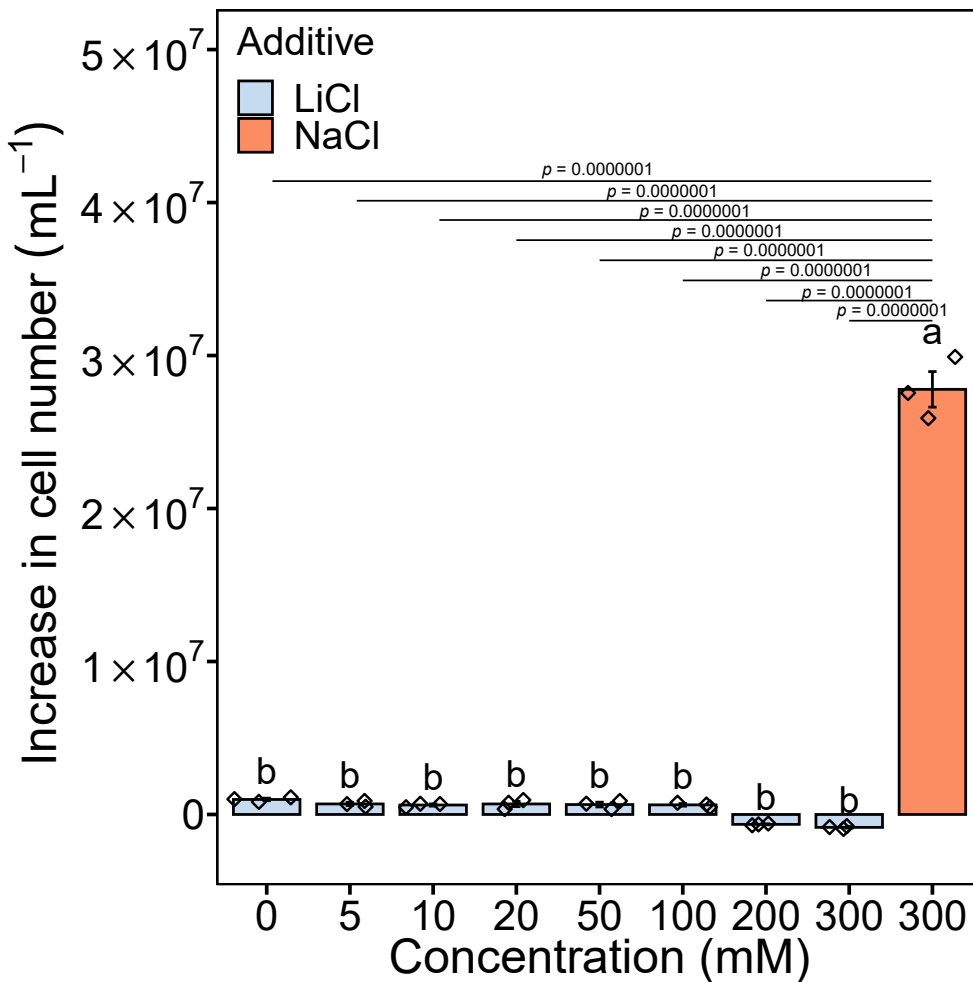
**Supplementary Fig. 2. Images and physiological parameters of cells in the absence or presence of NaCl under diazotrophy.** **a** Images of cells (day 7) grown in BG11<sub>0</sub> or BG11<sub>0</sub> with NaCl (18 g/L) (BG11<sub>0</sub> (NaCl)). Similar results were observed in six independent experiments. **b** Chlorophyll content. **c** Total protein. **d** Dry weight. **e** Protein to dry weight ratio. “None” indicates that no additional substances were added to BG11<sub>0</sub> while “NaCl” denotes the addition of additional NaCl (18 g/L) to BG11<sub>0</sub>. The graphs show the mean  $\pm$  standard deviation ( $n = 3$  biologically independent samples). Corresponding significance markers represent statistical significance calculated by two-tailed Welch’s  $t$  test. Significance: ns (no significance), \* ( $p < 0.05$ ), \*\* ( $p < 0.01$ ), \*\*\* ( $p < 0.001$ ), \*\*\*\* ( $p < 0.0001$ ). Source data are provided as a Source Data file.



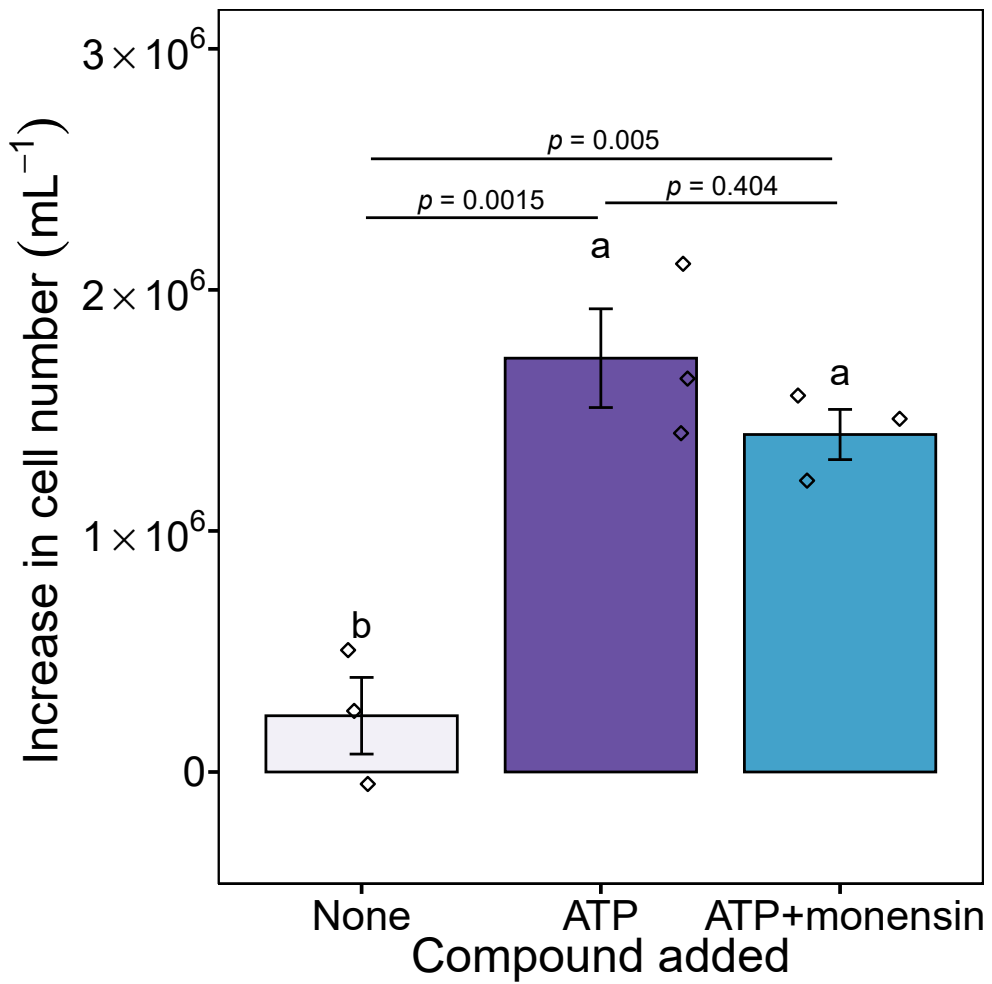
**Supplementary Fig. 3. Transcriptomes and key functional enrichment assay of DEGs.** **a** Number of significantly up- or downregulated DEGs (number of genes denoted at the top). **b** Analysis of enriched KEGG Pathways containing DEGs with a  $P_{\text{adjust}}$  value  $< 0.05$ . The size of the dot reflects the number of genes for each pathway, and the colour reflects the  $p$  value. **c** Heatmap analysis showing the DEGs enriched for porphyrin metabolism (map00860). Column dendrograms show similarity based on Euclidean distance and hierarchical clustering. Gene clusters were determined by k-means clustering with Euclidean distance. KEGG annotations were assigned from the genome annotation. The heatmap colour gradient shows low gene expression (blue) and high gene expression (red). “NaCl” denotes the addition of NaCl (18 g/L) to BG11<sub>o</sub>, while “None” indicates that no additional substances were added to BG11<sub>o</sub>. Source data are provided as a Source Data file.



**Supplementary Fig. 4. Enrichment analysis of ABC transporters and transcript analysis of the stringent response signal-encoding gene *relA*.** **a** Heatmap analysis showing the DEGs enriched for nutrient transporters. Eighteen transporter types have been annotated and compared. Column dendrograms show similarity based on Euclidean distance and hierarchical clustering. Gene clusters were determined by k-means clustering with Euclidean distance. The heatmap colour gradient shows low gene expression (blue) and high gene expression (red). **b** Expression trend of the stringent response signal ppGpp encoding gene *relA*. *relA* was identified as a significantly differentially expressed gene between these two treatments ( $p < 0.05$ , right-tailed Fisher's exact  $t$  test followed by Benjamin-Hochberg (BH) adjustment). Data (TPM) are presented as box plots (lower bound at 25th percentile, centre line at the median, upper bound at 75th percentile) with whiskers at minimum and maximum values. Statistical significance of *relA* was calculated from pairwise TPM comparisons of triplicate samples. "NaCl" indicates the supplementation of additional NaCl (18 g/L) to BG11<sub>o</sub>, while "None" indicates that no additional substances were added to BG11<sub>o</sub>. Source data are provided as a Source Data file.

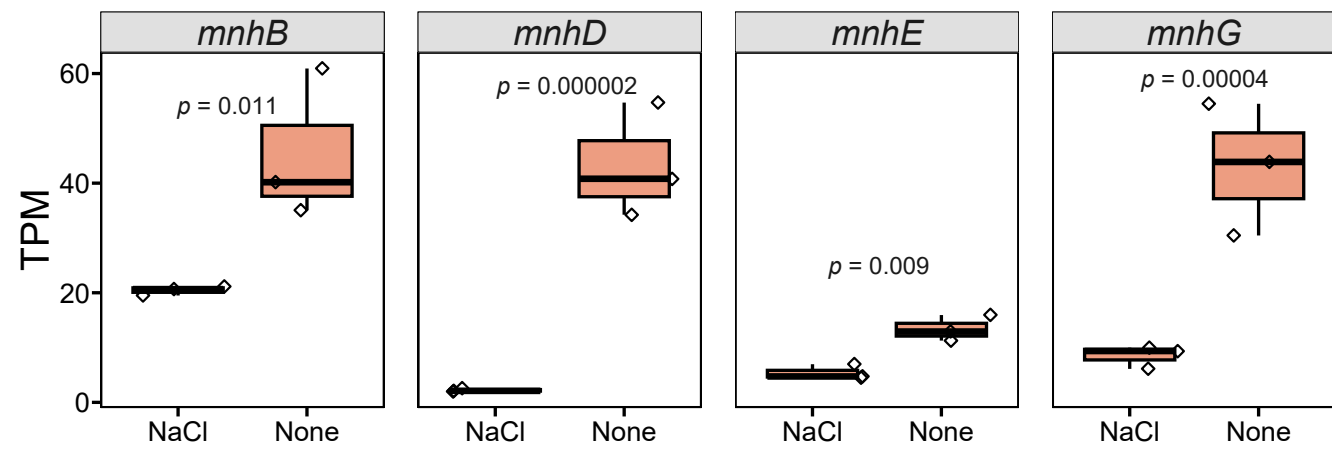


**Supplementary Fig. 5. Population growth over seven days of cells grown in BG11<sub>o</sub> with LiCl.** The increase in cell number of cells grown in BG11<sub>o</sub> with the addition of 0, 5, 10, 20, 50, 100, 200, 300 mM LiCl and 300 mM NaCl was calculated. The graph shows the mean  $\pm$  standard deviation ( $n = 3$  biologically independent samples). Different letters on each bar indicate statistical significance ( $p < 0.05$ ) calculated by one-way ANOVA with Tukey's HSD post-hoc analysis over all populations tested. Source data are provided as a Source Data file.



**Supplementary Fig. 6. Population growth with ATP supplementation in the absence or presence of monensin over three days.** Increase in cell number (3 days after inoculation) of cells grown in BG11<sub>o</sub>, BG11<sub>o</sub> supplemented with 0.1 mM ATP or BG11<sub>o</sub> supplemented with 0.1 mM ATP and 14  $\mu$ M monensin. The graph shows the mean  $\pm$  standard deviation (n = 3 biologically independent samples). Different letters on each bar indicate statistical significance ( $p < 0.05$ ) calculated by ANOVA with Tukey's HSD post-hoc analysis across all populations tested. Source data are provided as a Source Data file.

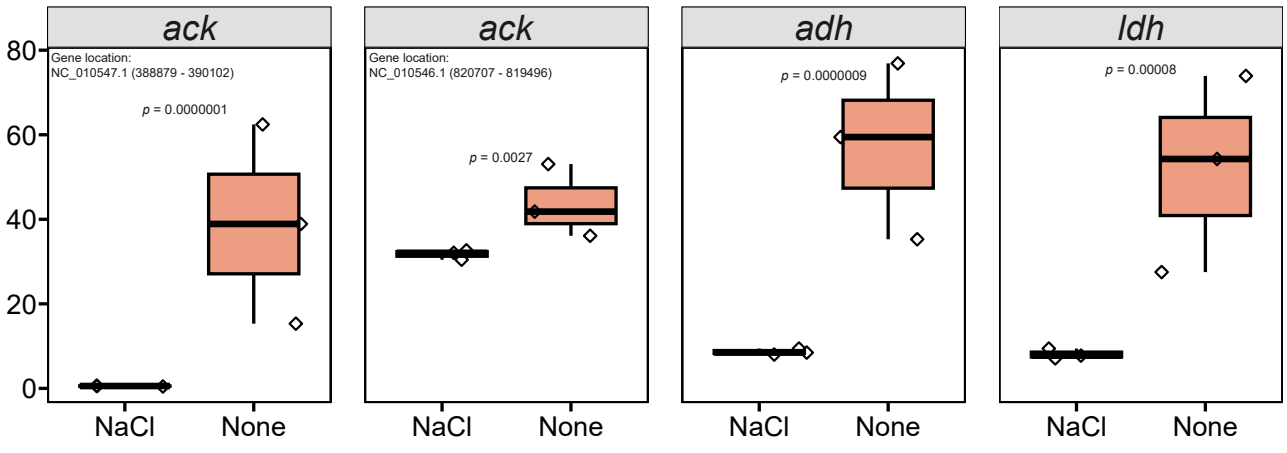




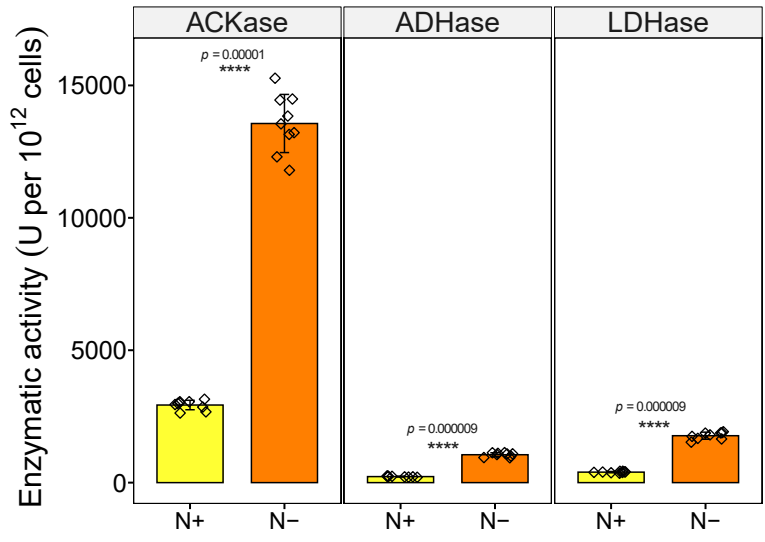
**Supplementary Fig. 7. Expression trend of Na<sup>+</sup>/H<sup>+</sup> antiporter encoding genes.** All four Na<sup>+</sup>/H<sup>+</sup> antiporter subunit encoding genes (subunit B (*mnhB*, K05566, KEGG), subunit D (*mnhD*, K05568, KEGG), subunit E (*mnhE*, K05569, KEGG), subunit G (*mnhG*, K05571, KEGG)) were identified as significantly differentially expressed genes between these two treatments ( $p < 0.05$ , right-tailed Fisher's exact  $t$  test followed by Benjamin-Hochberg (BH) adjustment). Data (TPM) are presented as box plots (lower bound at 25th percentile, centre line at the median, upper bound at 75th percentile) with whiskers at minimum and maximum values. Statistical significance of four genes was calculated from pairwise TPM comparisons of triplicate samples (grown in BG11<sub>o</sub>) of NaCl deprivation treatment (None) compared to the NaCl supplementation (18 g/L) treatment (NaCl). Source data are provided as a Source Data file.

**a**

TPM



**b**



**Supplementary Fig. 8. Transcript analysis of genes involved in fermentation and enzymatic activities of enzymes.** **a** *ack* encoding acetate kinase (K00925, KEGG) (two *ack* genes were annotated at different locations of the genome), *adh* encoding alcohol dehydrogenase (K00001, KEGG), *ldh* encoding lactate dehydrogenase (K00016, KEGG). The locations of two *ack* genes are NC\_010547.1 (388879 - 390102) and NC\_010546.1 (820707 - 819496) in the NCBI database. Data (TPM) are presented as box plots (lower bound at 25th percentile, centre line at the median, upper bound at 75th percentile) with whiskers at minimum and maximum values. All three genes were identified as significantly differentially expressed genes between these two treatments ( $p < 0.05$ , right-tailed Fisher's exact  $t$  test followed by Benjamin-Hochberg (BH) adjustment). Statistical significance of genes analysed was calculated from pairwise TPM comparisons of triplicate samples of NaCl deprivation treatment (None) compared to the NaCl addition treatment (NaCl). **b** Enzymatic activities of ACKase, ADHase and LDHase. "N+" indicates BG11 with NaCl (18 g/L) while "N-" denotes BG11<sub>0</sub> with NaCl (18 g/L). The graphs show the mean  $\pm$  standard deviation ( $n = 9$  biologically independent samples). Corresponding significance markers represent statistical significance calculated by two-tailed Welch's  $t$  test. Significance: ns (no significance),  $^*(p < 0.05)$ ,  $^{**}(p < 0.01)$ ,  $^{***}(p < 0.001)$ ,  $^{****}(p < 0.0001)$ . Source data are provided as a Source Data file.