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## Long non-coding RNA ZEB2-AS1 promotes the proliferation, metastasis and epithelial mesenchymal transition in triplenegative breast cancer by epigenetically activating ZEB2

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#### Abstract

The triple-negative breast cancer is the most malignant type of breast cancer. Its pathogenesis and prognosis remain poor despite the significant advances in breast cancer diagnosis and therapy. Meanwhile, long noncoding RNAs (LncRNAs) play a pivotal role in the progression of malignant tumors. In this study, we found that LncRNA-ZEB2-AS1 was dramatically up-regulated in our breast cancer specimens and cells (MDA231), especially in metastatic tumor specimens and highly invasive cells, and high IncRNA-ZEB2-AS1 expression is associated with clinicopathologic features and short survival of breast cancer patients. LncRNA-ZEB2-AS1 promotes the proliferation and metastasis of MDA231 cells in SCID mice. Thus, it is regarded as an oncogene in triple-negative breast cancer. It is mainly endo-nuclear and situated near ZEB2, positively regulating ZEB2 expression and activating the epithelial mesenchymal transition via the PI3K/Akt/GSK3<sup>β</sup>/Zeb2 signaling pathway. Meanwhile, EGF-induced F-actin polymerization in MDA231 cells can be suppressed by reducing IncRNA-ZEB2-AS1 expression. The migration and invasion of triple-negative breast cancer can be altered through cytoskeleton rearrangement. In summary, we demonstrated that IncRNA-ZEB2-AS1 is an important factor affecting the development of triple-negative breast cancer and thus a potential oncogene target.

#### KEYWORDS

cytoskeleton rearrangement, epithelial mesenchymal transition (EMT), LncRNA-ZEB2-AS1, triple-negative breast cancer

#### 1 | INTRODUCTION

Breast cancer is a malignancy with high death rate; that is, individuals with this disease, especially women, have a five-year overall survival rate of less than 15%.<sup>1</sup> Having high metastasis and recurrence rates, breast cancer is difficult to treat through prophase clinical diagnosis. Although major breakthroughs in clinical treatment methods have been achieved for breast cancer, approximately 100 in 100 000 women aged 55-69 years are expected to contract the disease by 2021.<sup>2</sup> Inhibiting the invasion and metastasis of malignant tumors

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is the most in clinical treatment, and it is also a hot and difficult issue of current research.

Many previous studies confirmed that epithelial mesenchymal transition (EMT) is one of the main mechanisms that cause the dispersion of malignant tumors.<sup>3</sup> EMT triggers a variety of biological changes in normal mammary epithelial cells, which eventually obtains the characteristics of mesenchymal cells. This effect enhances the metastasis capacity, invasiveness and resistance of cancer cells, thereby preventing apoptosis and promoting the production of extracellular matrix components.<sup>4</sup> The occurrence of EMT is related to a variety of molecular mechanisms and signaling pathways.<sup>5</sup> Its main function is the reduction of intercellular adhesion and E-cadherin, mesenchymal vimentin, and N-cadherin expression. Moreover, many transcriptional factors, including snail and ZEB, have crucial roles in EMT-induced processes.<sup>6</sup> The ZEB family (ZEB1 and ZEB2) is closely related to the EMT-related markers of malignant tumors.<sup>7</sup> However, the regulatory role of lncRNA-ZEB2-AS1 in ZEB2 expression in breast cancer remains unreported.

LncRNA is a kind of regulatory RNA that has a transcriptional length of more than 200 nucleotides and has no protein coding ability.<sup>8</sup> LncRNA plays a vital role in malignant tumor development, especially in apoptosis, proliferation, and invasion.<sup>9-11</sup> Abnormal IncRNA expression is observed in malignant tumors, such as gastric cancer, hepatocellular carcinoma (HCC), and glioma.<sup>12-14</sup> Moreover, IncRNA is a competitive endogenous RNA (ceRNAs) that regulates miRNA.<sup>15,16</sup> Despite the significant role IncRNA in malignant tumor development, its regulatory functions and molecular mechanisms remain poorly understood.

In this study, we found that abnormal IncRNA-ZEB2-AS1 expression is associated with patient survival and prognosis and controls ZEB2 expression. In the MDA231 cells, LncRNA-ZEB2-AS1 promoted the proliferation and metastasis of tumor cells and triggered EMT via the PI3K/Akt/GSK3 $\beta$ /Zeb2 signaling pathway and F-actin polymerization.

#### 2 | MATERIAL AND METHODS

#### 2.1 | Clinical specimens

We obtained breast cancer specimens (BC) and adjacent no tumor (ANT) specimens from the Affiliated Hospital of Weifang Medical University from 2011 to 2016. All the specimens were frozen to -150°C. None of the specimens received radiotherapy or chemotherapy before surgery. Informed consent was obtained from each patient. The study was approved by the Research Ethics Committee of Weifang Medical University.

#### 2.2 | Cell culture

MCF-10A, T47D, MDA-MB-435 (MDA435), MCF-7, and MDA-MB-231(MDA231) were acquired from ATCC (USA). The cells were cultured in minimum essential medium with 10% fetal bovine serum (FBS) at  $37^{\circ}$ C under 5% CO<sub>2</sub> atmosphere.

#### 2.3 | Plasmid construction and cell transfection

MDA231 cells ( $2 \times 10^5$ ) were planted in six-well plates and left overnight. We used Lipofectamine 2000 to transfect the MDA231 cells in accordance with the manufacturer protocol Lnc-ZEB2-AS1-RNAi, and si-NC plasmids were constructed by GeneChem company (shanghai, China) Sequences were provided in Table S1. Stable transfected cells were maintained with 300 µg/mL G418 for 14 days.

#### 2.4 | Wound healing assay

Approximately  $2 \times 10^5$  breast cancer cells were planted onto six-well plates and then left overnight. Wounds were created with a 10 µL pipette tip. Complete medium including 1% FBS was added to the plates with or without EGF. The entire setup was photographed after 36 hours.

#### 2.5 | Cell proliferation assays

For the cell counting kit 8 (CCK-8; Solarbio, Beijing) assays, approximately  $2 \times 10^3$  cells were seeded in a 96-well plate. The proliferation capacities of the cells cultured for 24, 48, 72, 96, and 120 hours were tested by the CCK-8 assays. The cell growth curves were plotted by using the absorbance value at each time point.

#### 2.6 | Transwell assay

MDA231 cells ( $1 \times 10^5$ ) were counted and planted in serum-free essential medium in the upper chamber. Matrigel (Corning) was used for this step. The bottom chamber was added into essential medium with EGF. The cells were incubated for 24 hours at 37°C. The membranes of the MDA231 cells were fixed and stained, photographed, and counted.

#### 2.7 | Western blot

Protein expression was examined as described previously.<sup>17</sup> The following antibodies were employed: ZEB2 (Santa Cruz, 1:500), N-cadherin (Cell Signaling Technology, CST, 1:500), Ecadherin (CST, 1:500), vimentin (CST, 1:1000), GSK3 $\beta$  (CST, 1:500), p-GSK3 $\beta^{Ser9}$  (CST, 1:500), p-Akt <sup>Ser473</sup> (CST, 1:500), Akt (CST, 1:500), p-LIMK<sup>Thr508</sup> (CST, 1:500), p-cofilin<sup>Ser3</sup> (CST, 1:500),  $\beta$ -actin (CST, 1:2000), HRP-conjugated anti-mouse IgG and anti-rabbit IgG antibody (CST, 1:500). All experiments were performed in triplicate.

#### 2.8 | Quantitative real-time PCR

We performed Quantitative real-time PCR (QRT-PCR) to define the relative level of IncRNA-ZEB2-AS1. GAPDH levels were used for normalization. We used TRIzol to extract RNA from fresh breast cancer specimens and cells. Then, cDNA was synthesized with the total RNA by using an M-MLV reverse transcriptase kit (Promega, USA). Relative mRNA expression was normalized through the 2  $^ ^{\Delta\Delta CT}$  method. QRT-PCR was carried out using an Applied Biosystems 7500.

#### 2.9 | Cellular F-actin measurement

MDA231 cells were immobilized with 4% paraformaldehyde with PBS. The cells were then washed three times before they were blocked with a buffer containing goat serum for 45 minutes. The MDA231 cells were stained with Phalloidin (Fluorescein isothiocyanate) FITC for 1 hours, washed three times, and covered with fluorescence decay resistant sealing tablets. Data processing was performed as described previously.<sup>18</sup>

#### 2.10 | Immunofluorescence

The MDA231 cells were fixed with 4% paraformaldehyde for 25 min. The cells were permeabilized with 0.1% Triton X-100 and blocked with goat serum for 45 min at 37°C. The primary antibodies of relation were incubated at 4°C overnight. FITC, Cy3-labeled goat antirabbit IgG secondary antibodies, and DAPI were used.

#### 2.11 | Animal studies

The Server Combined Immune-deficiency (SCID) mice were provided by the Animal Care and Use Committee of Wei Fang Medical University. Si-NC/MDA231 and MDA231/si-ZEB2-AS1 cells  $(2 \times 10^6)$  were injected into the oxter of each female SCID mouse (n = 10). When the xenografts became evident, tumor volume was

#### 2.12 | Statistical analysis

Data were analyzed by SPSS v16.0. All values were expressed as mean  $\pm$  SD. The results were analyzed with T test or ANOVA. X<sup>2</sup>-test was used for the analysis of the connection between IncRNA-ZEB2-AS1 and the clinicopathologic features. *P* < 0.05 was considered significant in all cases

#### 3 | RESULTS

#### 3.1 | Up-regulation of IncRNA-ZEB2-AS1 in breast cancer specimens and cells was related to clinicopathologic features and decreased survival of breast cancer patients

To determine the biological function of IncRNA-ZEB2-AS1 in the tumorigenesis of triple-negative breast cancer, we detected IncRNA-ZEB2-AS1 expression levels in 98 paired of BC specimens and ANT specimens by qRT-PCR. The results revealed that IncRNA-ZEB2-AS1 was more markedly up-regulated in breast cancer specimens than paired ANTs (Figure 1A). We further detected expressed IncRNA-ZEB2-AS1 with or without lymph node metastasis in breast cancer specimens (Figure 1B). Meanwhile, we measured the expression level of IncRNA-ZEB2-AS1 in breast cancer cells (T47D, MDA231, MCF-7, and MDA435) and normal mammary epithelial cells (MCF-10A). The data revealed that IncRNA-ZEB2-AS1 expression was markedly



**FIGURE 1** The differentially expressed of IncRNA-ZEB2-AS1 in breast cancer specimens and cells. A, Expression of IncRNA-ZEB2-AS1 in 98 pairs of BC and ANT specimens. B, Differentially expressed IncRNA-ZEB2-AS1 in breast cancer specimens with and without lymph node metastasis. C, LncRNA-ZEB2-AS1 detected in MCF-7, T47D, MDA231, MDA435 and MCF-10A cells. D, LncRNA-ZEB2-AS1 localization in the nucleus and cytoplasm was tested. E, Kaplan-Meier method analysis (log-rank test) was carried out for the verification of relationship between abnormal IncRNA-ZEB2-AS1 expression and poor prognosis (\*, *P* < 0.05).

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up-regulated in breast cancer cells compared with MCF-10A cells. The expression levels of highly invasive MDA231 cells and MDA435 were higher IncRNA-ZEB2-AS1, and those of invasive cells T47D and MCF-7 were lower than IncRNA-ZEB2-AS1 (Figure 1C). LncRNA is considered to have different biological functions in different locations of the cells. Meanwhile, we first detected the distribution of IncRNA-ZEB2-AS1 in the MDA231 cells. LncRNA-ZEB2-AS1 was mainly located in the nucleus was minimally expressed in the cytoplasm (Figure 1D). Additionally, breast cancer patients with high IncRNA-ZEB2-AS1 levels have shorter survival rates than those with low levels (Figure 1E). The relationship between IncRNA-ZEB2-AS1 expression level and the clinicopathologic features of breast cancer patients was evaluated. LncRNA-ZEB2-AS1 expression is highly associated with tumor differentiation, lymph node metastasis, and distant metastasis in breast cancer but not associated with age or tumor size (Table 1). The results indicated that IncRNA-ZEB2-AS1 plays an oncogenic role and is associated with clinicopathologic features and decreased survival of breast cancer patients.

#### 3.2 | Knockdown of IncRNA-ZEB2-AS1 suppressed the proliferation and invasion of MDA231 cells

The knockdown of lncRNA-ZEB2-AS1 in breast cancer cells implies that lncRNA-ZEB2-AS1 have important roles in breast cancer progression. To further investigate whether lncRNA-ZEB2-AS1 is connected to the occurrence of breast cancer, we researched the function of lncRNA-ZEB2-AS1 in vitro. As shown in Figure 2A, lncRNA-ZEB2-AS1-RNAi-1 (si-ZEB2-AS1) showed better interference efficiency than the negative control group (si-NC). Thus, was selected for further research. In this research, the ability of cell proliferation was examined via CCK-8 assays after the knockdown of lncRNA-ZEB2-AS1 in the MDA231 cells. The results revealed that the rate MDA231 cell proliferation was considerably reduced after the knockdown of lncRNA-ZEB2-AS1 (Figure 2B). Moreover, colonies formed by MDA231 cells were reduced (Figure 2C). To investigate the mechanisms underlying invasion suppression after reduction of lncRNA-ZEB2-AS1, we further tested the effect of lncRNA-ZEB2-AS1 on the invasion and migration ability of the MDA231 cells. The assay results of transwell and wound healing showed that the migration and invasion of MDA231 cells can be inhibited by reducing lncRNA-ZEB2-AS1 (Figure 2D,E). Overall, these results suggested that the decrease of lncRNA-ZEB2-AS1 expression led to the suppression of the proliferation and invasion of MDA231 cells.

# 3.3 | Knockdown of IncRNA-ZEB2-AS1 led to the suppression of the tumorigenesis and metastasis of MDA231 cells in SCID mice

We further evaluated whether knocking down IncRNA-ZEB2-AS1 has an effect on the tumorigenesis and metastasis of MDA231 cells in SCID mice. We recorded changes in the volumes of MDA231/ si-ZEB2-AS1 and si-NC/MDA231 group cells during tumorigenesis. Compared with si-NC/MDA231 group, MDA231/si-ZEB2-AS1 group grew more slowly. The final tumor volume in the MDA231/si-ZEB2-AS1 group was smaller than that in the si-NC/MDA231 group (Figure 3A). Meanwhile, the final tumor weight in the former was lighter than that in the si-NC/MDA231 group (Figure 3B). Changes in the metastasis capabilities of the MDA231 cells were detected after the stable transfection of si-ZEB2-AS1 group showed significantly reduced tumor nodules (Figure 3C). The H&E stains revealed that

		ZEB2-AS1 expression		
Characteristics	Total (n = 98)	Low (n = 49)	High (n = 49)	P value
Age (years)				
≤50	51	26	25	0.500
≥51	47	23	24	
Tumor size (cm)				
≤5 cm	42	19	23	0.270
>5 cm	56	30	26	
Tumor differentiation				
L	21	16	5	0.013
11	48	23	25	
III	29	10	19	
Lymph node metastasis				
Yes	51	18	33	0.002
No	47	31	16	
Distant metastasis				
Yes	45	16	29	0.007
No	53	33	20	

**TABLE 1**Correlation betweenIncRNA-ZEB2-AS1 expression andclinicopathological features of breastcancer patients



FIGURE 2 Knockdown of IncRNA-ZEB2-AS1 suppressed the proliferation and invasion of MDA231 cells. A, Depletion of IncRNA-ZEB2-AS1 in MDA231 cells by IncRNA-ZEB1-AS1-RNAi. B, Cells proliferation activity was detected via CCK-8 assays. C, Left: Colony formation assay results showed that the depletion of IncRNA ZEB2-AS1 led to the remarkable decrease of MDA231/si-ZEB2-AS1 content. Right: Quantification of colony numbers. D, Left: Invasion of MDA231 cells analyzed with transwell chambers (scale bars = 100 μm). Right: Quantification of penetrating cells. E, Left: Wound healing assay (scale bars = 200 μm). Right: Migration distance (\*, P < 0.05).

the MDA231/si-ZEB2-AS1 group had significantly reduced pulmonary metastatic foci (Figure 3D).

#### 3.4 | Knockdown of IncRNA-ZEB2-AS1 led to the down-regulation of ZEB2 expression through PI3K/Akt/GSK3β/Zeb2 signaling pathway and inhibited EMT

The upstream antisense transcription may affect corresponding gene expression.<sup>19</sup> LncRNA-ZEB2-AS1 is an antisense lncRNA. We hypothesized that the antisense transcript IncRNA-ZEB2-AS1 modulates ZEB2 in breast cancer. LncRNA-ZEB2-AS1 regulated the ZEB2 expression in the MDA231 cells. Then, we monitored the MDA231/ si-ZEB2-AS1 cells to observe the effects of IncRNA-ZEB2-AS1 on ZEB2. ZEB2 was remarkably decreased in the MDA231/si-ZEB2-AS1 cells (Figure 4A). We then tested the expression level of ZEB2 mRNA. A remarkable positive correlation between LncRNA-ZEB2-AS1 and ZEB2 mRNA was observed at the protein level (Figure 4B). Given that ZEB2 is a crucial transcription factor modulating EMT in different malignant tumors, changes in the expression of EMTrelated markers in the MDA231/si-ZEB2-AS1 cells and si-NC/ MDA231 cells were investigated. Meanwhile, mesenchymal markers and epithelial markers were decreased and increased, respectively, when LncRNA-ZEB2-AS1 knocked down, suggesting that knocking down IncRNA-ZEB2-AS1 results in the inhibition of the EMT of breast cancer (Figure 4C,D). These results were supported by the EMT-related markers detected by immunofluorescence (Figure 4E). Subsequently, we attempted to determine the important IncRNA-ZEB2-AS1 mechanisms that inhibit the EMT of the MDA231 cells. We discovered that the PI3K/Akt/GSK3<sup>β</sup>/Zeb2 signaling pathway has a pivotal role in the regulation of the EMT of the MDA231 cells. The phosphorylation levels of protein that played a key function in the PI3K/Akt/GSK3<sub>β</sub>/Zeb2 signaling pathway was tested by Western blot. Our data showed that the phosphorylation levels of GSK3<sup>β</sup> and Akt in MDA231/si-ZEB2-AS1 cells were considerably depressed when IncRNA-ZEB2-AS1 expression was knocked down (Figure 4F). All the results validated that knockdown of IncRNA-ZEB2-AS1 leads to the down-regulation of ZEB2 expression and subsequently to the suppression of EMT through the PI3K/Akt/GSK3β/Zeb2 signaling pathway.

#### 3.5 | Reduction of IncRNA-ZEB2-AS1 suppressed EGF-induced F-actin polymerization in MDA231 cells

F-actin polymerization is an important mechanism for the invasion and metastasis of cancer cells. To confirm the putative that the depression of IncRNA-ZEB2-AS1 could suppress EGF-induced F-actin polymerization of MDA231 cells, we performed the F-actin polymerization assay to detect the effect of IncRNA-ZEB2-AS1 reduction on the invasion and metastasis capacity of MDA231 cells with EGF



FIGURE 3 Knockdown of IncRNA-ZEB2-AS1 suppressed the tumorigenesis and metastasis of MDA231 cells in SCID mice. A, Tumor volume growth curves in SCID mice. B, Tumor weight at the end point. C, Number of tumor nodules at the end point. D, Pulmonary metastasis and HE staining pulmonary metastasis of SCID mice (scale bars =  $200 \mu m$ ) (\*, P < 0.05).

stimulation. The data showed that EGF led to short actin polymerization in si-NC/MDA231 cells and was notably decreased in the MDA231/si-ZEB2-AS1 cells, indicating that IncRNA-ZEB2-AS1 played a vital function in modulating cytoskeleton rearrangement by EGF stimulation (Figure 5A,B). Meanwhile, LIMK and cofilin are two vital proteins related to the regulation of F-actin polymerization.<sup>20</sup> We detected the phosphorylation status of LIMK and cofilin, to confirm whether IncRNA-ZEB2-AS1 inhibits EGF-induced rearrangement of cytoskeletons in the MDA231 cells (Figure 5C). As shown in Figure 5C, the reduction of IncRNA-ZEB2-AS1 expression depressed the phosphorylation levels of LIMK and cofilin, and the cytoskeleton was abolished. These results indicated that reduction of IncRNA-ZEB2-AS1 suppressed EGF-induced F-actin polymerization in MDA231 cells.

#### 3.6 | Clinical relevance of IncRNA-ZEB2-AS1 between ZEB2 and EMT related markers in human breast cancer

In order to further indicate the clinically relevant by which IncRNA-ZEB2-AS1 modulated ZEB2 and EMT related markers. We detected the IncRNA-ZEB2-AS1 expression and its association with the expression levels of ZEB2 in the clinical tissues of breast cancer paired with breast cancer specimens and ANT specimens. The relationship between the expression levels of EMT related markers and IncRNA-ZEB2-AS1 was investigated. In Figure 6, a positive correlation among IncRNA-ZEB2-AS1, ZEB2, and vimentin in 30 tested clinical specimens was observed, whereas IncRNA-ZEB2-AS1 was negatively correlated with E-cadherin. All the results suggested that IncRNA-ZEB2-AS1 in the ZEB2 and EMT-related markers is clinically relevant in breast cancer.

#### DISCUSSION 4

An increasing number of studies have reported that LncRNA considerably affects cellular processes, such invasion, metastasis, metabolism, and apoptosis.<sup>21,22</sup> LncRNAs monitor carcinogenesis principally via modulating the expression levels of oncogenes or tumor suppressors.<sup>19</sup> Many studies revealed that LncRNA-ZEB2-AS1 decreases tumor growth and metastasis in HCC.<sup>23</sup> LncRNA-ZEB2-AS1 is a newly discovered lncRNA, whose clinical meaning, biological roles, and explicit mechanism are poorly understood in breast cancer. We first confirmed that IncRNA-ZEB2-AS1 is abnormally expressed in breast cancer specimens, and this abnormality is related



**FIGURE 4** Knockdown of IncRNA-ZEB2-AS1 down-regulated ZEB2 expression through PI3K/Akt/GSK3 $\beta$ /Zeb2 signaling pathway and inhibited EMT. A, Left: ZEB2 expression was tested using western blot. Right: quantification of ZEB2 protein. B, The expression of ZEB2 mRNA in MDA231 cells. C, The expression of EMT related markers was examined in MDA231 cells. D, Quantification of EMT related markers. E, Immunofluorescence stains (scale bars = 50 µm). F, Phosphorylated Akt<sup>Ser473</sup> and GSK3 $\beta$ <sup>Ser9</sup> in EGF-induced MDA231 cells were examined (\*, P < 0.05).

to the overall survival and clinicopathologic features of patients with breast cancer. Mechanistically, IncRNA-ZEB2-AS1 affects the proliferation, metastasis, and EMT of MDA231 cells.

ZEB1 and ZEB2 cardinal proteins of the ZEB family. These proteins promote EMT in cancer. Transcription factor attributing to especial Zinc finger protein families, usually located in the downstream of some signaling pathways executing EMT process in normal and pathological conditions.<sup>24</sup> Previous discuss have indicated that there is ectopic of ZEB in tumor and is associated with poor prognosis in patients.<sup>25</sup> ZEB1 is a novel protein and pivotal transcription factor in EMT progress. ZEB2 protein might have a tumor promoter role because it controls cell cycle progression and cell differentiation.<sup>26</sup> Wang et al found that E2F1 promoted EMT by modulating the promoter region of ZEB2 in NSCLC.<sup>27</sup> ZEB2 is a transcription factor that participates in adjusting different biological activities, including the inhibition of apoptosis of vascular endothelial cells via the MAPK pathway activation.<sup>28</sup> Meanwhile, the mechanisms by which IncRNA-ZEB2-AS1 regulates EMT in breast cancer remain unknown. In this study, we found that knocking down IncRNA-ZEB2-AS1 results in the down-regulation of ZEB2 expression through the PI3K/ Akt/GSK3 $\beta$ /Zeb2 signaling pathway, inactivation of AKT and GSK3 $\beta$ phosphorylation, inhibition of EMT in breast cancer.

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Distant metastasis in early phases of development is a typical biological characteristic of cancer cells, and F-actin polymerization



**FIGURE 5** Reduction of IncRNA-ZEB2-AS1 suppressed EGF-induced F-actin polymerization in MDA231 cells. A, Cytoskeleton rearrangement in the MDA231 cells was shown by fluorescence assay (scale bars =  $50 \mu m$ ). B, F-actin polymerization in the MDA231 cells under different periods of stimulation with EGF. C, P-LIMK<sup>Thr508</sup> and p-cofilin<sup>Ser3</sup> in MDA231 cells under EGF stimulation (\*, P < 0.05).



**FIGURE 6** Clinical relevance of lncRNA-ZEB2-AS1 between ZEB2 and EMT-related marker in human breast cancer. A, Correlation between lncRNA-ZEB2-AS1 and ZEB2. B, Correlation between lncRNA-ZEB2-AS1 and E-cadherin, C, correlation between lncRNA-ZEB2-AS1 and vimentin were all confirmed through clinical correlation coefficient analysis

is considered to be an early, pivotal step for invasion and metastasis. The number arrays of actin binding proteins are the modulators of F-actin polymerization and lamellipodium formation.<sup>29</sup> LIMK and cofilin are essential controllers.<sup>30,31</sup> B. Kalyanaraman et al recent research also found that cancer cell invasion needs essential regulations of cell cytoskeleton, cell-to-matrix and cellular adhesions.<sup>32</sup> Wang et al found that migration and invasion inhibitory proteins inhibit endometrial carcinoma migration via the cytoskeleton reorganization when the number of lamellipodia are markedly reduced.<sup>33</sup> It is still an unknown molecular mechanism that lncRNA-ZEB2-AS1 modulates F-actin polymerization of MDA231 cells. In the current research, we detected the rearrangement of cytoskeletal protein when lncRNA-ZEB2-AS1 was abrogated in breast cancer cells. Meanwhile, the phosphorylation levels of LIMK and cofilin were decreased after lncRNA-ZEB2-AS1

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was knocked down, verifying that IncRNA-ZEB2-AS1 was related to F-actin polymerization in MDA231 cells.

Collectively, our results showed that IncRNA-ZEB2-AS1/PI3K/ Akt/GSK3 $\beta$ /Zeb2 axis facilitates tumor progression and is a potential prevention target in breast cancer.

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#### CONFLICT OF INTEREST

The authors confirm that there are no conflicts of interest.

Guoxin Zhang, Hongli Li, Zhiyi Yang, and Yuanyuan Liu performed experiments and wrote the manuscript. Zhaoyan Wang and Yuling Yang analyzed data. Ruimei Sun, Peirui Li and Chonggao Yin assisted with the design of experiments.

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#### REFERENCES

- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer J Clin. 2011;61:69-90.
- Linos E, Spanos D, Rosner BA, et al. Effects of reproductive and demographic changes on breast cancer incidence in China: a modeling analysis. J Natl Cancer Inst. 2008;100:1352-1360.
- Zhang B, Li H, Yin C, et al. Dock1 promotes the mesenchymal transition of glioma and is modulated by MiR-31. *Neuropathol Appl Neurobiol*. 2017;43:419-432.
- Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. J Clin Invest. 2009;119:1420-1428.
- Chen DL, Chen LZ, Lu YX, et al. Long noncoding RNA XIST expedites metastasis and modulates epithelial-mesenchymal transition in colorectal cancer. *Cell Death Dis.* 2017;8:e3011.
- Iwatsuki M, Mimori K, Yokobori T, et al. Epithelial-mesenchymal transition in cancer development and its clinical significance. *Cancer Sci.* 2010;101:293-299.
- Guo F, Parker Kerrigan BC, Yang D, et al. Post-transcriptional regulatory network of epithelial-to-mesenchymal and mesenchymal-toepithelial transitions. J Hematol Oncol. 2014;7:19.
- Geisler S, Coller J. RNA in unexpected places: long non-coding RNA functions in diverse cellular contexts. *Nat Rev Mol Cell Biol.* 2013;14:699-712.
- 9. Huarte M. The emerging role of IncRNAs in cancer. Nat Med. 2015;21:1253-1261.
- 10. **Takahashi** Y, **Sawada** G, **Kurashige** J, et al. Amplification of PVT-1 is involved in poor prognosis via apoptosis inhibition in colorectal cancers. *Br J Cancer*. 2014;110:164-171.
- Hong Q, Li O, Zheng W, et al. LncRNA HOTAIR regulates HIF-1alpha/ AXL signaling through inhibition of miR-217 in renal cell carcinoma. *Cell Death Dis.* 2017;8:e2772.
- Yao XM, Tang JH, Zhu H, Jing Y. High expression of LncRNA CASC15 is a risk factor for gastric cancer prognosis and promote the proliferation of gastric cancer. *Eur Rev Med Pharmacol Sci.* 2017;21:5661-5667.
- Takahashi K, Yan I, Haga H, Patel T. Long noncoding RNA in liver diseases. *Hepatology*. 2014;60:744-753.
- Lv QL, Hu L, Chen SH, et al. A long noncoding RNA ZEB1-AS1 promotes tumorigenesis and predicts poor prognosis in Glioma. Int J Mol Sci. 2016;17.
- Tay Y, Rinn J, Pandolfi PP The multilayered complexity of ceRNA crosstalk and competition. *Nature*. 2014; 505: 344–352.
- Zhong W, Chen S, Qin Y, et al. Doxycycline inhibits breast cancer EMT and metastasis through PAR-1/NF-kappaB/miR-17/E-cadherin pathway. Oncotarget. 2017;8:104855-104866.
- 17. Jia L, Liu W, Cao B, Li H, Yin C. MiR-507 inhibits the migration and invasion of human breastcancer cells through Flt-1 suppression. *Oncotarget*. 2016;7:36743-36754.
- 18. Yin C, Li H, Zhang B, et al. RAGE-binding S100A8/A9 promotes the migration and invasion of human breast cancer cells through

actin polymerization and epithelial-mesenchymal transition. Breast Cancer Res Treat. 2013;142:297-309.

- Su W, Xu M, Chen X, et al. Long noncoding RNA ZEB1-AS1 epigenetically regulates the expressions of ZEB1 and downstream molecules in prostate cancer. *Mol Cancer*. 2017;16:142.
- Lu S, Niu N, Guo H, et al. ARK5 promotes glioma cell invasion, and its elevated expression is correlated with poor clinical outcome. *Eur J Cancer.* 2013;49:752-763.
- 21. Mathieu EL, Belhocine M, Dao LT, Puthier D, Spicuglia S. Functions of IncRNA in development and diseases. *Med Sci (Paris)*. 2014;30:790-796.
- Spizzo R, Almeida MI, Colombatti A, Calin GA. Long non-coding RNAs and cancer: a new frontier of translational research? Oncogene. 2012;31:4577-4587.
- Lan T, Chang L, Wu L, Yuan Y. Downregulation of ZEB2-AS1 decreased tumor growth and metastasis in hepatocellular carcinoma. *Mol Med Rep.* 2016;14:4606-4612.
- 24. Wan Makhtar WR, Browne G, Karountzos A, et al. Short stretches of rare codons regulate translation of the transcription factor ZEB2 in cancer cells. *Oncogene*. 2017;36:6640-6648.
- 25. Sanchez-Tillo E, de Barrios O, Siles L, et al. ZEB1 Promotes invasiveness of colorectal carcinoma cells through the opposing regulation of uPA and PAI-1. *Clin Cancer Res.* 2013;19:1071-1082.
- Denecker G, Vandamme N, Akay O, et al. Identification of a ZEB2-MITF-ZEB1 transcriptional network that controls melanogenesis and melanoma progression. *Cell Death Differ*. 2014;21:1250-1261.
- Wang T, Chen X, Qiao W, Kong L, Sun D, Li Z. Transcription factor E2F1 promotes EMT by regulating ZEB2 in small cell lung cancer. BMC Cancer. 2017;17:719.
- Wang LJ, Wu ZH, Zheng XT, Long JY, Dong YM, Fang X. Zinc finger E-Box binding protein 2 (ZEB2) suppress apoptosis of vascular endothelial cells induced by high glucose through mitogen-activated protein kinases (MAPK) pathway activation. *Med Sci Monit*. 2017;23:2590-2598.
- Klopocka W, Korczynski J, Pomorski P. Cytoskeleton and nucleotide signaling in glioma C6 cells. Adv Exp Med Biol. 2013;986:103-119.
- Shishkin S, Eremina L, Pashintseva N, Kovalev L, Kovaleva M. Cofilin-1 and other ADF/Cofilin superfamily members in human malignant cells. *Int J Mol Sci.* 2016;18.
- Martin SK, Kamelgarn M, Kyprianou N. Cytoskeleton targeting value in prostate cancer treatment. Am J Clin Exp Urol. 2014;2:15-26.
- 32. **Kalyanaraman** B. Teaching the basics of cancer metabolism: Developing antitumor strategies by exploiting the differences between normal and cancer cell metabolism. *Redox Biol.* 2017;12:833-842.
- Wang Y, Hu L, Ji P, et al. Erratum to: MIIP remodels Rac1-mediated cytoskeleton structure in suppression of endometrial cancer metastasis. J Hematol Oncol. 2017;10:163.

#### SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

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