

Hereditary kidney diseases associated with hypomagnesemia

Felix Claverie-Martin¹, Ana Perdomo-Ramirez¹, Victor Garcia-Nieto²

¹Unidad de Investigación, Hospital Universitario Nuestra Señora de Candelaria, Santa Cruz de Tenerife, Spain ²Unidad de Nefrología Pediátrica, Hospital Universitario Nuestra Señora de Candelaria, Santa Cruz de Tenerife, Spain

In the kidney, a set of proteins expressed in the epithelial cells of the thick ascending loop of Henle and the distal convoluted tubule directly or indirectly play important roles in the regulation of serum magnesium levels. Magnesium reabsorption in the thick ascending loop of Henle occurs through a passive paracellular pathway, while in the distal convoluted tubule, the final magnesium concentration is established through an active transcellular pathway. The players involved in magnesium reabsorption include proteins with diverse functions including tight junction proteins, cation and anion channels, sodium chloride cotransporter, calcium-sensing receptor, epidermal growth factor, cyclin M2, sodium potassium adenosine triphosphatase subunits, transcription factors, a serine protease, and proteins involved in mitochondrial function. Mutations in the genes that encode these proteins impair their function and cause different rare diseases associated with hypomagnesemia, which may lead to muscle cramps, fatigue, epileptic seizures, intellectual disability, cardiac arrhythmias, and chronic kidney disease. The purpose of this review is to describe the clinical and genetic characteristics of these hereditary kidney diseases and the current research findings on the pathophysiological basis of these diseases.

Keywords: Hypomagnesemia, Magnesium handling, Mutation, Rare diseases, Renal tubulopathies

Introduction

Magnesium (Mg^{2+}) , the second-most abundant intracellular cation, is an indispensable ion for many cellular functions including energy metabolism and nucleic acid and protein synthesis [1]. Mg^{2+} is also a regulator of sodium, potassium, and calcium channels. Therefore, serum Mg^{2+} levels, which are usually 0.70 to 1.1 mmol/L, need to be precisely controlled. Hypomagnesemia is defined as serum Mg^{2+} level below 0.7 mmol/L. The predominant laboratory tests used for the diag-

nosis of hypomagnesemia are the serum Mg^{2+} concentration and the 24-hour urinary Mg^{2+} tests. In cases in which the serum Mg^{2+} level is low, a 24-hour urine Mg^{2+} higher than 24 mg/day suggests renal Mg^{2+} wasting as the cause of the hypomagnesemia, while values lower than 24 mg/day indicate deficient Mg^{2+} intake and/or gastrointestinal losses. Magnesium homeostasis is determined by intestinal absorption, renal reabsorption, and storage in bone. In the kidney and intestine, these processes involve a combination of paracellular and transcellular epithelial transport routes.

Correspondence: Felix Claverie-Martin

Copyright © 2021 by The Korean Society of Nephrology

Received: May 24, 2021; Revised: June 16, 2021; Accepted: June 20, 2021

Editor: Sungjin Chung, The Catholic University of Korea, Seoul, Republic of Korea

Unidad de Investigación, Hospital Universitario Nuestra Señora de Candelaria, Carretera del Rosario 145, 38010 Santa Cruz de Tenerife, Spain. E-mail: fclamar@gobiernodecanarias.org, fclamar@gmail.com

ORCID: https://orcid.org/0000-0003-2468-3267

[©] This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial and No Derivatives License (http:// creativecommons.org/licenses/by-nc-nd/4.0/) which permits unrestricted non-commercial use, distribution of the material without any modifications, and reproduction in any medium, provided the original works properly cited.

Hypomagnesemia may cause muscle cramps, fatigue, appetite loss, and disruptions in calcium and potassium homeostasis [1]. Acute hypomagnesemia may lead to more serious consequences like epileptic seizures, intellectual disability, and cardiac arrhythmias. Causes of hypomagnesemia include type 2 diabetes, gastrointestinal diseases, alcoholism, use of diuretics or other medications, dietary deficiency, and genetic defects. Over the last two decades, clinical and genetic studies of patients with rare hereditary disorders of Mg^{2+} handling have enabled the identification of important components of epithelial Mg^{2+} transport in the kidney [2].

After passing the glomerular filter, 90% to 95% of the filtered Mg²⁺ is subsequently reabsorbed along the nephron [1]. Approximately 10% to 25% of the Mg²⁺ is reabsorbed by the proximal tubule (PT), 50% to 70% is reabsorbed by the thick ascending limb of the loop of Henle (TAL), and 5% to 10% is reabsorbed by the distal convoluted tubule (DCT) (Fig. 1). In the PT, Mg²⁺ reabsorption occurs in a passive paracellular mode. The mechanisms that control this process are unknown. In the TAL, Mg²⁺ reabsorption also takes place through a passive paracellular pathway, which is facil-





itated by tight junction proteins claudin-16 and claudin-19 (Fig. 2) [3,4]. The basolateral calcium-sensing receptor (CaSR) controls the paracellular transport of calcium (Ca²⁺) and Mg²⁺ by regulating the claudin-16/claudin-19 channel function through a signaling pathway [5]. In contrast, the main K⁺ secretory channel in the kidney, the renal outer medullary K⁺ channel (ROMK), which is located in the apical membrane of the TAL, plays a key role in the generation of the lumen-positive potential in TAL [1]. The renal chloride (Cl⁻) channel ClC-Kb, which is located in the basolateral membrane of the TAL and also the DCT, facilitates the efflux of Cl⁻ to the interstitium [6]. The DCT establishes the final Mg²⁺ concentration through active transcellular



Figure 2. Reabsorption of Mg^{2^+} in the TAL. In this segment of the nephron, Mg^{2^+} reabsorption takes place through a passive paracellular pathway via claudin-16/claudin-19 channels located in the tight junctions. This transport depends on the driving force generated by the joint actions of Na⁺, K⁺-adenosine triphosphatase (ATPase), and Cl⁻ channel ClC-Kb, both located in the basolateral membrane, and Na⁺-K⁺-2Cl-cotransporter (NKCC2) and renal outer medullary K⁺ channel (ROMK), the main K⁺ secretory channel in the kidney, located in the apical membrane. Basolateral calcium-sensing receptor (CaSR) indirectly regulates the paracellular divalent cation transport and the transcellular NaCl transport in the TAL.

ATP, adenosine triphosphate; HNF1 β , hepatocyte nuclear factor-1 beta; PCBD1, pterin-4 α -carbinolamine dehydratase; TAL, thick ascending limb of the loop of Henle.

reabsorption, which is highly regulated, via transient receptor potential melastatin type 6 (TRPM6) Mg²⁺ channels located in the apical membrane [7] (Fig. 3). TRPM6 needs to form heterotetramers with its close homolog TRPM7 to function [8]. The protein(s) that facilitates Mg²⁺ efflux from the basolateral side to the blood compartment has not been identified [9]. Cyclin M2 (CNNM2) and solute carrier family member A1 (SLC41A1) have been proposed as likely candidates for this function, but this subject is still under debate [10–12]. Other proteins involved, albeit indirectly, in Mg²⁺



Figure 3. Reabsorption of Mg²⁺ in the DCT. In this tubular segment, Mg²⁺ is reabsorbed through a transcellular pathway. The apical Mg²⁺ channel transient receptor potential melastatin type 6 (TRPM6) facilitates transport of Mg^{2+} from the lumen to the epithelial cell. The driving force needed for Mg²⁺ transport is generated by the joint actions of Na⁺, K⁺-adenosine triphosphatase (ATPase), the Cl⁻ channel ClC-Kb and the K⁺ channel Kir4.1 in the basolateral membrane and Na⁺-Cl⁻ cotransporter (NCC) and K⁺ secretory channels Kv1.1 and renal outer medullary K⁺ channel (ROMK) in the apical membrane. Epidermal growth factor (EGF) is implicated in the regulation of Mg^{2+} reabsorption through TRPM6. HNF1 β with coactivator pterin-4 α -carbinolamine dehydratase (PCBD1) increase transcription of the γ subunit of the Na⁺, K⁺-AT-Pase. Cyclin M2 (CNNM2) is a candidate for basolateral extrusion of Mg^{2+} or a regulator of Mg^{2+} transport, but this has not been experimentally confirmed.

ATP, adenosine triphosphate; DCT, distal convoluted tubule; EGFR, epidermal growth factor receptor; HNF1 β , hepatocyte nuclear factor-1 beta.

transport in the DCT include epidermal growth factor (EGF), a hormone that regulates the expression of TRPM6 on the apical membrane, basolateral Na⁺, K⁺-adenosine triphosphatase (ATPase), and K⁺ channel Kir4.1, and the apical K⁺ channel Kv1.1 and Na⁺ Cl⁻ cotransporter (NCC) [13–16]. The last four proteins participate in the generation of the driving force needed for Mg²⁺ transport [9]. Mutations in any of the above proteins, except SLC41A1, which are involved directly or indirectly in Mg²⁺ transport, have been shown to cause hypomagnesemia (Table 1).

In this review, we present the current knowledge of hereditary kidney diseases associated with hypomagnesemia. We discuss the clinical characteristics and genetic information for each disease and describe the pathophysiological basis that has been proposed for some of the diseases, although in general these remain incompletely understood. We classified hypomagnesemias in three groups according to the implicated genes (Table 1). Group 1 includes hypomagnesemias associated with genes that encode proteins directly involved in Mg²⁺ transport and the regulatory proteins. Group 2 contains hypomagnesemias associated with genes encoding proteins involved in transport of other ions or their regulators, which indirectly affect Mg²⁺ handling. Group 3 includes hypomagnesemias associated with genes required for mitochondrial function, which also indirectly result in Mg²⁺ loss.

Group 1 hypomagnesemias

Familial hypomagnesemia with hypercalciuria and nephrocalcinosis types 1 and 2

Familial hypomagnesemia with hypercalciuria and nephrocalcinosis (FHHNC) is a rare autosomal recessive tubular disorder characterized by excessive urinary loss of Mg²⁺ and Ca²⁺, bilateral nephrocalcinosis, and progressive chronic kidney disease (CKD) [17]. FHHNC patients typically present during early childhood or before adolescence with recurrent urinary tract infections, polyuria, polydipsia, nephrolithiasis, and failure to thrive [18–21]. FHHNC patients may show a pronounced decline in glomerular filtration rate at the time of diagnosis, and approximately one-third of cases progress to chronic renal failure during childhood or adolescence [21,22]. In contrast to patients with other hypomagnesemias, FHHNC patients have high serum levels of

Table 1. Inherited diseases associated with hypomagnesemia

Disorder	Associated gene	e Protein ^a	Inheritance	OMIM #
Group 1				
FHHNC type 1	CLDN16	Claudin-16	AR	248250
FHHNC type 2	CLDN19	Claudin-19	AR	248190
Hypomagnesemia with secondary hypocalcemia	TRPM6	Mg ²⁺ channel TRPM6	AR	602014
Isolated recessive hypomagnesemia	EGF	Epidermal growth factor	AR	611718
NISBD2	EGFR	Epidermal growth factor receptor	AR	616069
HSMR type 1	CNNM2	Cyclin M2	AD/AR	616418/613882
Group 2				
Bartter syndrome type 3	CLCNKB	Cl ⁻ channel ClC-Kb	AR	607364
Gitelman syndrome	SLC12A3	Na ⁺ Cl ⁻ cotransporter	AR	263800
Autosomal dominant hypocalcemia with hypercalciuria	CASR	Calcium-sensing receptor CaSR	AD	601198
Episodic ataxia type 1	KCNA1	Voltage-gated K^{+} channel Kv1.1	AD	160120
Isolated dominant hypomagnesemia	FXYD2	γ subunit of Na ⁺ , K ⁺ -ATPase	AD	154020
HSMR type 2	ATP1A1	α 1 subunit of Na+, K ⁺ -ATPase	AD	618314
EAST/SeSAME syndrome	KCNJ10	K⁺ channel Kir4.1	AR	612780
ADTKD-HNF1B	HNF1B	Transcription factor HNF1β	AD	137920
TNHP	PCBD1	PCBD1, dimerization cofactor of HNF1 $\!\beta$	AR	264070
Kenny-Caffey syndrome type 2	FAM111A	Nuclear serine protease FAM111A	AD	127000
Group 3				
HHH syndrome	MT-TI	Isoleucine tRNA	Mitochondrial	500005
Kearns-Sayre syndrome	Large deletions	-	Mitochondrial	530000
HUPRA syndrome	SARS2	seryl-tRNA synthetase	AR	613845

AD, autosomal dominant; ADTKD-*HNF1B*, autosomal dominant tubulointerstitial kidney disease subtype *HNF1B*; AR, autosomal recessive; EAST/SeS-AME, epilepsy, ataxia, sensorineural deafness and tubulopathy syndrome/seizures, sensorineural deafness, ataxia, mental retardation and electrolyte imbalance; FHHNC, familial hypomagnesemia with hypercalciuria and nephrocalcinosis; HHH, hypertension, hypercholesterolemia and hypomagnesemia; HSMR, hypomagnesemia, seizures, and mental retardation disorder; HUPRA, hyperuricemia, pulmonary hypertension, renal failure and alkalosis; NISBD2, neonatal inflammatory skin and bowel disease type 2; TNHP, transient neonatal hyperphenylalaninemia and primapterinuria; tRNA, transfer RNA. ^aAll the genes associated with hereditary hypomagnesemias encode proteins, except MT-TI, which encodes a tRNA.

parathyroid hormone (PTH) before the onset of chronic renal failure [1,19,23]. In some cases, patients present amelogenesis imperfecta [24,25]. Clinical signs of severe hypomagnesemia such as seizures and muscular tetany are rare.

FHHNC is caused by recessive mutations in *CLDN16* (FHHNC type 1, OMIM #248250) or *CLDN19* (FHHNC type 2, OMIM #248190) [3,4]. Patients with mutations in *CLDN19* also present ocular abnormalities such as severe myopia, nystagmus, and macular colobamata [4,21,26]. *CLDN16* and *CLDN19* encode the tight junction proteins claudin-16 and claudin-19, respectively, which are strongly expressed in the kidney [3,4]. Claudin-19 is also expressed in peripheral neurons and retina [4,27]. Claudin-16 and claudin-19 form heteromeric paracellular cation channels in the TAL that regulate Ca²⁺ and Mg²⁺ transport [28,29]. The reabsorption of Mg²⁺ is greatly dependent on the transepithelial poten-

tial as a driving force, which is created by a transepithelial NaCl concentration gradient. *CLDN16* and *CLDN19* mutations have been shown to impair the permeating function of these channels, resulting in a reduced lumen-positive potential and the simultaneous loss of the driving force for Mg^{2+} reabsorption [22,28].

Hypomagnesemia with secondary hypocalcemia

Hypomagnesemia with secondary hypocalcemia (HSH, OMIM #602014) is a rare autosomal recessive disorder characterized by severe hypomagnesemia associated with hypocalcemia. The levels of serum Mg²⁺ in patients with HSH are usually much lower than in patients with other types of hereditary hypomagnesemias [30]. The disease usually presents in early infancy, with neurological symptoms including tetany and severe seizures that are refractory to anticonvulsant therapy [30,31]. HSH patients have very low serum levels of PTH. Persistent low levels of serum Mg^{2+} likely cause secondary hypocalcemia by inhibiting PTH secretion and inducing resistance at the receptor sites [30]. The main defect in HSH is a reduction in intestinal Mg^{2+} absorption, which is in contrast to all other identified hereditary hypomagnesemias, along with reduced Mg^{2+} reabsorption in the DCT with renal Mg^{2+} wasting [31,32].

HSH is caused by recessive loss-of-function mutations in the TRPM6 gene, which encodes the TRPM6 cation channel [31,32]. TRPM6 is predominantly expressed in the apical membrane of the intestinal and renal DCT epithelial cells, where it is involved in Mg²⁺ reabsorption [7]. Its channel activity and expression are regulated by several factors including EGF and adenosine triphosphate (ATP) [14]. TRPM6 mutations identified in HSH patients disrupt Mg^{2+} conductance through the channel, both in the colon and DCT, causing hypomagnesemia [7]. However, the mechanisms leading to this disease are not entirely known. TRPM6 interacts with its close homolog TRPM7 to form functional heteromeric TRPM6/TRPM7 Mg²⁺ channels [8]. TRPM6 and TRPM7 contain a transmembrane ion channel segment covalently joined to a cytosolic serine/ threonine protein kinase domain at the carboxy terminus. The kinase domain is cleaved from the channel segment and, after its translocation to the nucleus, it regulates the transcription of many genes involved in development [33]. Therefore, it has been suggested that TRPM6 may play a much wider role in the cell than only in Mg^{2+} homeostasis [33].

Isolated recessive hypomagnesemia

Isolated recessive renal hypomagnesemia is a rare disorder characterized by hypomagnesemia and normocalciuria [34]. Hypomagnesemia is due to renal Mg²⁺ wasting. Patients show seizures and neurodevelopmental delay during childhood. Only two affected girls from a consanguineous family have been reported, and no other biochemical abnormalities were identified in these patients. A homozygous missense mutation in the *EGF* gene coding for pro-EGF was identified as the underlying genetic defect [34]. Pro-EGF is a type I membrane-bound precursor protein that is proteolytically cleaved to generate the soluble EGF peptide hormone. EGF binds with high affinity to the renal epidermal growth factor receptor (EGFR) at the basolateral membrane of the DCT. EGFR activation triggers a signaling cascade leading to an increase of TRPM6 channels on the apical membrane and increased Mg^{2+} reabsorption [14]. The disease-causing *EGF* mutation results in diminished sorting of pro-EGF, preventing adequate secretion of the EGF hormone [34]. This leads to inadequate stimulation of the EGFR, and therefore insufficient activation of the TRPM6 channel, which results in reduced reabsorption of Mg^{2+} .

Neonatal inflammatory skin and bowel disease type 2

Using whole-exome sequencing, a rare homozygous missense mutation (p.Gly428Asp) in *EGFR* was identified in a child with an inflammatory syndrome affecting the skin, bowel, and lungs (OMIM #616069) [35]. The pregnancy was complicated by polyhydramnios and the child was born prematurely. Laboratory tests revealed low serum levels of magnesium. The child showed failure to thrive and died at 2.5 years of age from widespread cutaneous and pulmonary infections in addition to electrolyte imbalance. Results of a skin biopsy and immunofluorescence microscopy studies revealed that the mutation p.Gly428Asp reduces EGFR plasma membrane localization [35].

Hypomagnesemia, seizures, and mental retardation disorder type 1

Hypomagnesemia, seizures, and mental retardation disorder (HSMR) type 1 is a complex rare condition characterized by renal Mg²⁺ loss that results in hypomagnesemia, infantile or juvenile epileptic seizures, and intellectual disability [36,37]. Patients also show autistic features, aggressive behavior, variable degrees of delayed psychomotor development, speech limitations, impaired motor skills, and in some cases obesity [37,38]. HSMR type 1 is caused by loss-offunction mutations in the CNNM2 gene, which encodes the transmembrane protein CNNM2 [36-38]. Most HSMR type 1 patients carry heterozygous mutations that are generated de novo or inherited in an autosomal dominant pattern (OMIM #616418) [36-38]. However, a recessive mode of inheritance has been reported for several families [37,39]. Patients with recessive CNNM2 mutations show a severe phenotype, including brain malformations, refractory epilepsy, and acute intellectual disability (OMIM #613882).

CNNM2 is expressed in many organs and tissues including brain and kidney. In the kidney, *CNNM2* is predominantly expressed at the basolateral membrane of the DCT cells, where it is associated with Mg^{2+} reabsorption [36,40]. Whether CNNM2 is itself an Mg^{2+} transporter or a regulator of Mg^{2+} transport is unclear [11]. However, *CNNM2* pathogenic mutations reduce its expression in the plasma membrane, resulting in defective Mg^{2+} reabsorption and Mg^{2+} wasting [11,38].

Furthermore, recent studies revealed that Mg^{2+} -ATP binding to the intracellular C-terminus of CNNM2 is required for protein dimerization and Mg^{2+} efflux [41]. Disease-causing mutations that are located in the Mg^{2+} -ATP-binding site abolish ATP binding and Mg^{2+} efflux activity. The basis of the neurological defects remains unknown.

Group 2 hypomagnesemias

Bartter syndrome type 3

Bartter syndrome (BS) includes a group of several tubulopathies characterized by renal salt wasting, hypokalemia, hypochloremic metabolic alkalosis, hyperreninemia, hyperaldosteronism, and low to normal blood pressure [42,43]. Patients usually present during the first years of life with failure to thrive, polyuria, and polydipsia. The main pathogenic mechanism in these tubulopathies is defective salt reabsorption predominantly in the TAL. Five different types of BS have been identified based on the gene involved [43]. Patients with BS type 3 or classic BS (OMIM #607364) develop hypomagnesemia during childhood or later in life [44]. This disorder is characterized by a great clinical variability, and there is a correlation between the severity of mutations and younger age at diagnosis [44,45].

BS type 3 is caused by recessive loss-of-function mutations of the *CLCNKB* gene, which encodes the kidney-specific Cl⁻ channel ClC-Kb that is involved in NaCl reabsorption in the renal tubule [6]. The ClC-Kb protein is expressed in the basolateral membrane of epithelial cells in the TAL and the DCT. In these tubular segments, Cl⁻ exits the cell through ClC-Ka and ClC-Kb channels. Mutations in *CLCNKB* alter the intracellular Cl⁻ regulation, which subsequently interferes with the generation of the lumen-positive potential and results in salt wasting and possibly hypomagnesemia.

Gitelman syndrome

Gitelman syndrome (GS, OMIM #263800) is an autosomal recessive salt-losing tubulopathy characterized by hypokalemic metabolic alkalosis, low or normal blood pressure, hypocalciuria, and hypomagnesemia with renal Mg²⁺ wasting [46]. GS is the most common cause of hereditary hypomagnesemia and is usually detected during adolescence or adulthood. GS may be asymptomatic or associated with mild symptoms including chronic fatigue, muscle weakness, thirst, salt craving, nocturia, and cramps, which can considerably reduce the quality of life [47]. Severe complications such as cardiac arrhythmias have been reported in some cases [46].

GS is caused by recessive inactivating mutations of the *SLC12A3* gene encoding the thiazide-sensitive NCC, which is localized in the apical membrane of DCT cells, where it plays a fundamental role in the reabsorption of Na⁺ and Cl⁻ [48,49]. NCC mutations lead to reduced reabsorption, which results in Na⁺ and Cl⁻ wasting, hypovolemia, and subsequent hyperaldosteronism with metabolic alkalosis [50]. The pathogenesis of hypocalciuria and hypomagnesemia may be explained by the compensatory paracellular reabsorption of Na⁺ and Ca²⁺ in the PT due to volume reduction and by decreased apical expression of TRPM6 [51]. There is phenotypic variability in GS, including in patients with the same *SLC12A3* mutation.

Autosomal dominant hypocalcemia with hypercalciuria

Autosomal dominant hypocalcemia with hypercalciuria (ADHH, OMIM #601198) is a rare disorder of calcium homeostasis characterized by variable levels of hypocalcemia and low or normal serum levels of PTH [52]. Patients also present with hypomagnesemia, hypermagnesuria, hyperphosphatemia, and hypercalciuria [52,53]. Hypocalcemia is a derived effect of hypomagnesemia as a result of parathyroid failure or PTH resistance [30]. ADHH patients may develop hypocalcemic symptoms (paresthesias, carpopedal spasm, and seizures), and some have renal and basal ganglia calcifications, but others are asymptomatic [52].

ADHH is caused by heterozygous gain-of-function mutations in the *CASR* gene [52,54]. This gene encodes the extracellular CaSR, a G protein-coupled receptor that is highly expressed in parathyroid glands and kidneys [53]. In the kidney, CaSR is highly expressed in the basolateral membrane of the TAL, where it regulates Ca^{2+} reabsorption independently of PTH [53,55]. A signaling pathway including two microRNAs and the tight junction proteins claudin-14 and claudin-16 mediates the effect of CaSR on renal Ca^{2+} and Mg^{2+} excretion [5]. Activating mutations of CaSR increase the expression of claudin-14, which binds to claudin-16 and blocks the cation permeability of the claudin-16/claudin-19 channel.

Episodic ataxia type 1

Loss-of-function mutations in the *KCNA1* gene are typically associated with an autosomal dominant neurological disorder called episodic ataxia type 1 (EA1, OMIM #160120), which is characterized by recurring episodes of ataxia and myokymia from early childhood [13]. The clinical phenotype in EA1 patients can include seizures, epilepsy, and, in some cases, paroxysmal kinesigenic dyskinesia, cataplexy, myokymia, and hypomagnesemia [56]. These symptoms can appear alone or in combination with EA1. A genotype-phenotype correlation analysis in a large cohort of EA1 patients revealed high inter-and intrafamilial variability of symptoms, but the penetrance of hypomagnesemia has not been evaluated [57].

The *KCNA1* gene encodes the α subunit of the voltage-gated potassium channel Kv1.1, which is abundantly expressed in specific neurons and plays an important role in regulating neuronal excitability in the central and peripheral nervous system [57]. In the kidney, KCNA1 is exclusively expressed at the apical membrane of the DCT cells alongside the Mg²⁺ transporter TRPM6 [58]. Previous studies suggested that potassium (K⁺) secretion via Kv1.1 provides the electrochemical gradient needed for Mg²⁺ reabsorption by the TRPM6 channel [58,59]. Interestingly, two specific KCNA1 heterozygous mutations, p.Asn255Asp and p.Leu328Val, have been associated with hypomagnesemia, leading to muscle cramps and tetanic episodes [58,59]. Electrophysiological analyses showed that both amino acid substitutions result in nonfunctional Kv1.1 channels [58–60]. The reduction in the K⁺ conductance depolarizes the apical membrane of DCT cells, reducing the electrical driving force and leading to renal Mg²⁺ loss. The frequency of hypomagnesemia in patients with KCNA1 mutations may be greater than reported, since serum Mg²⁺ levels have not been examined in all EA1 patients. Additional research is needed to understand the association of *KCNA1* mutations with hypomagnesemia.

Isolated dominant hypomagnesemia

Isolated dominant hypomagnesemia is a rare autosomal dominant disorder characterized by hypomagnesemia, hypocalciuria, and occasionally chondrocalcinosis (OMIM #154020) [61]. Some patients suffer from muscle cramps, episodes of convulsions, or chondrocalcinosis [61,62]. This disease has been identified in only three families who carry the same missense mutation, p.Gly41Arg, in the FXYD2 gene and appear to be descendants of a common ancestor [62,63]. FXYD2 encodes the kidney-specific regulatory γ-subunit of basolateral Na⁺, K⁺-ATPase, which is composed of a catalytic α subunit and an auxiliary β subunit. The Na⁺, K⁺-ATPase complex maintains the electrochemical gradients of Na⁺ and K⁺ across the basolateral plasma membrane that provide the driving force for transepithelial Mg²⁺ transport particularly in the DCT [15]. There are two splice variants of the γ -subunit: FXYD2a, which is expressed mainly in the TAL and PT, and FXYD2b, which is expressed exclusively in the basolateral membrane of the DCT and collecting duct [64]. Expression studies showed that the p.Gly41Arg mutation causes incorrect trafficking of the mutant y FXYD2b subunit, preventing its interaction with the α and β Na⁺, K⁺-ATPase subunits [62]. This leads to destabilization and reduction of Na⁺, K⁺-ATPase activity in the DCT, which eventually results in decreased Mg²⁺ uptake and renal Mg²⁺ loss.

Hypomagnesemia, seizures, and mental retardation disorder type 2

Recently, three children from non-consanguineous families were reported who presented with generalized seizures in infancy associated with severe intellectual disability, massive renal Mg²⁺ wasting, and hypomagnesemia (HSMR type 2; OMIM #618314) [65]. Other findings included significant developmental delay and limited motor skills. Mutational analysis identified heterozygous *de novo* mutations in the *ATP1A1* gene encoding the catalytic α 1 subunit of the Na⁺, K⁺-ATPase [65]. This ubiquitously expressed subunit is the major α isoform in the kidney and is present in practically all cell types of the central nervous system [15]. Dominant and *de novo* mutations in *ATP1A1* are also the cause of autosomal dominant Charcot-Marie-Tooth disease type 2 (CMT2) [66]. To the best of our knowledge, serum Mg^{2+} levels have not been studied in CMT2 patients. Functional studies of the mutant Na⁺, K⁺-ATPase α 1 subunits revealed loss of pump activity and anomalous cation permeability, leading to membrane depolarization [65]. These discoveries emphasize the essential task of the catalytic α 1 subunit of the Na⁺, K⁺-ATPase in renal tubular Mg²⁺ transport and neuronal activity. Since *ATP1A1* is expressed in the TAL and DCT, it could affect transport in both tubular segments.

Epilepsy, ataxia, sensorineural deafness, and tubulopathy/seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance syndrome

Epilepsy, ataxia, sensorineural deafness, and tubulopathy (EAST) syndrome or seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance (SeSA-ME) syndrome (OMIM #612780) is an autosomal recessive disease characterized by early-onset epilepsy, delayed psychomotor development, ataxia, sensorineural deafness, and a salt-wasting tubulopathy with or without mental retardation [67,68]. The renal phenotype develops during the course of the disease and comprises polyuria, hypokalemia, metabolic alkalosis, hypocalciuria, and hypomagnesemia [69,70]. Plasma renin and aldosterone levels are increased and blood pressure is at the low end of the normal range. Urinary findings include K^+ , Mg^{2+} , and Na^+ wasting.

This complex disorder is caused by loss-of-function mutations in the KCNJ10 gene encoding Kir4.1, one of the components of the inwardly rectifying K⁺ channel Kir4.1/ Kir5.1 [16,67,68]. KCNJ10 is mainly expressed in glial cells of the brain, the stria vascularis of the inner ear, and the kidney [69]. The renal phenotype of patients with EAST/SeS-AME syndrome closely resembles that of patients with GS, suggesting that KCNJ10 mutations mainly impair transport in the DCT. In the kidney, Kir4.1 is expressed in the basolateral membrane of the DCT and is involved in K⁺ recycling, which is needed for the activity of Na⁺, K⁺-ATPase, and generation of a negative transmembrane potential [16]. Mutations that inactivate Kir4.1 function lead to a depolarization of the basolateral membrane and to a reduction of the driving force for anion channels and Na⁺-coupled exchangers. This alteration in membrane voltage could also affect other transport processes, such as those for Cl^- and Mg^{2+} , which could explain the Mg^{2+} loss detected in patients with EAST/ SeSAME syndrome.

Autosomal dominant tubulointerstitial kidney disease caused by *HNF1B* mutations

Autosomal dominant tubulointerstitial kidney disease (ADTKD) comprises a group of rare kidney disorders characterized by tubular damage and interstitial fibrosis without glomerular lesions [71]. Affected individuals usually develop CKD and end-stage renal disease in adulthood. Heterozygous mutations in several genes cause ADTKD, and this disease is subdivided into several subtypes based on the mutated gene [71].

ADTKD subtype *HNF1B* (ADTKD-*HNF1B*, OMIM #137920) is associated with hypomagnesemia [72,73]. The phenotypes detected in ADTKD-*HNF1B* patients are very heterogeneous and may appear during pregnancy, in childhood, or in adulthood [73]. Symptoms include renal cysts, kidney malformations, abnormalities of the genital tract and liver, and maturity-onset diabetes of the young (MODY) type 5 [72,74]. Hypomagnesemia and hypermagnesuria are observed in approximately 50% of patients [72,73].

ADTKD-HNF1B is caused by heterozygous mutations of the *HNF1B* gene, which encodes the developmentally regulated transcription factor hepatocyte nuclear factor-1ß $(HNF1\beta)$ [72,74,75]. These mutations are inherited in a dominant inheritance pattern or appear *de novo*. HNF1β regulates tissue-specific gene expression in epithelial cells of several organs, including the kidneys, pancreas, liver, and urogenital tract [76]. In the adult kidney, HNF1 β is expressed in epithelial cells of all tubular segments. However, its role in renal Mg²⁺ reabsorption seems to take place in the DCT, since hypomagnesemia in ADTKD-HNF1B patients is frequently accompanied by hypocalciuria [72,73]. Several transcriptional targets of HNF1^β have been identified, including the FXYD2 gene [72], which is involved in reabsorption of Mg²⁺ in the DCT, as described above. Therefore, inactivating mutations of HNF1ß would lead to reduced expression of FXYD2 and, consequently, to renal Mg²⁺ wasting and hypomagnesemia [72]. HNF1 β also regulates the transcription of *KCNJ10*, a gene that can affect Mg²⁺ transport in the DCT, as described above [77].

Transient neonatal hyperphenylalaninemia and primapterinuria

Transient neonatal hyperphenylalaninemia and primapterinuria (TNHP, OMIM #264070) is an autosomal recessive disorder characterized by mild transient hyperphenylalaninemia and elevated urinary levels of 7-biopterin [78]. Affected individuals are asymptomatic and exhibit normal psychomotor development. A follow-up study of TNHP patients showed that patients may also develop hypomagnesemia with renal Mg²⁺ wasting and MODY type diabetes [79]. TNHP is caused by mutations in the *PCBD1* gene, which encodes the bifunctional protein pterin-4 α-carbinolamine dehydratase (PCBD1). PCBD1 functions as a cytosolic enzyme that is implicated in the regeneration of the essential cofactor tetrahydrobiopterin as well as a coactivator of HN-F1 β -mediated transcription within the nucleus [80]. Gene expression studies combined with immunohistochemical analysis showed that in the kidney, PCBD1 is expressed predominantly in the DCT [79]. PCBD1 interacts with the dimerization domain of HNF1ß and regulates the formation of a transcriptionally active tetrameric complex [81]. Binding of PCBD1 to HNF1 β stimulates the *FXYD2* promoter in the DCT, and PCBD1 mutations identified in TNHP patients cause defective dimerization and degradation of the PCBD1 protein, leading to decreased *FXYD2* promoter activity [79]. The reduced expression of FXYD2 would cause hypomagnesemia in these patients.

Kenny-Caffey syndrome type 2

Kenny-Caffey syndrome type 2 (KCS type 2, #OMIM 127000) is characterized by severe short stature, impaired skeletal development, eve abnormalities, hypomagnesemia, and hypoparathyroidism [82,83]. KCS type 2 patients may also have frequent episodes of low Ca²⁺ levels in serum triggered by hypoparathyroidism. This multisystem disease is caused by heterozygous missense mutations in the FAM111A gene, which encodes the nuclear trypsin-like serine protease FAM111A [82,83]. FAM111A is involved in the regulation of PTH production, calcium homeostasis, bone development and growth, but the specific mechanisms have not been determined [82,83]. FAM111A mutations identified in patients usually appear de novo, but some cases with autosomal dominant inheritance have been described [84]. These mu-

tations affect the peptidase domain of FAM111A and may impair its catalytic activity [83,85].

FAM111A, first identified as an antiviral restriction factor, is ubiquitously expressed, and its nuclear localization suggests that it might be involved in transcriptional regulation [82,83,86]. FAM111A mutations result in hyperactivation of the FAM111A intrinsic protease activity, which could cause abnormal degradation of DNA-binding proteins or FAM111A depletion through hyper-autoproteolytic cleavage [87-89]. Hyperactive FAM111A is cytotoxic, disrupting nuclear structure and pore distribution in a protease-dependent manner [89]. Targets of FAM111A protease activity have been recently identified and include nucleoporins and the associated germinal center-associated nuclear protein transcription/replication factor [89]. Hypomagnesemia in KCS type 2 patients could be attributed to degradation of transcription factors involved in magnesium homeostasis. Tan et al. [90] suggested a potential role of FAM111A in regulation of CaSR, since KCS type 2 shares phenotypic characteristics with ADHH-like hypocalcemia and hypoparathyroidism.

Group 3 hypomagnesemias

Mitochondria are abundant in kidneys, as the kidney requires high amounts of energy to enable the reabsorption of ions in different tubular segments [91]. Mitochondrial dysfunction leads to reduced ATP synthesis and loss of renal function. Because the function and biogenesis of mitochondria are under the genetic control of both mitochondrial DNA (mtDNA) and nuclear DNA, mutations in either genome can be the cause of disease [92]. mtDNA mutations are usually inherited from the patient's mother but they can also appear *de novo*. The clinical features of these patients can be very variable since they depend not only on the type of mutation but also on the number of mitochondria affected [92,93]. Mitochondrial diseases manifest in infancy and are multisystemic, and there are no effective treatment options for the patients. Some mitochondrial diseases are associated with hypomagnesemia but the mechanism underlying how they cause hypomagnesemia remains unsolved. These diseases may affect the TAL, the DCT, or both.

Hypertension, hypercholesterolemia, and hypomagnesemia

This disorder was identified in a large pedigree and includes hypertension, hypercholesterolemia, and hypomagnesemia (OMIM #500005) [94]. Affected individuals also have hypocalciuria and hypokalemia. Each of these traits was transmitted via the maternal lineage with a pattern indicating mitochondrial inheritance. Other clinical characteristics include migraine headache, sensorineural hearing loss, and hypertrophic cardiomyopathy, all usually associated with mitochondrial dysfunction. Sequence analvsis of the mitochondrial genome in the maternal lineage of the pedigree identified a C to T transition in the MT-TI gene, which encodes isoleucine transfer RNA (tRNA^{1le}) [94]. Further analysis of affected members revealed that all copies of the mtDNA contain this mutation. The affected C is located immediately 5' to the tRNA^{Ile} anticodon, and biochemical studies showed that the substitution of T significantly weakens ribosome binding [95]. These studies suggest that this mutation results in the loss of mitochondrial function. Hypomagnesemia associated with hypocalciuria points to a primary defect in the DCT [96].

Kearns-Sayre syndrome

Kearns-Sayre syndrome (KSS, OMIM #530000) is a rare progressive multisystem disease characterized by ophthalmoparesis/ptosis and pigmentary retinopathy. The onset is typically before 20 years of age, and patients typically present one of the following: cerebellar ataxia, cardiac conduction defects, deafness, short stature, cognitive involvement, tremor, and cardiomyopathy [97]. Some cases develop hypoparathyroidism and renal tubular dysfunction resulting in severe hypomagnesemia, hypocalcemia with hypermagnesuria, and hypokalemia [98,99]. KSS is caused by large deletions in mtDNA, which can reduce considerably ATP production [93]. In most cases, the deletions arise de novo but some are transmitted through maternal inheritance. Patients' cells usually contain a mixture of wild-type and mutant mtDNA molecules in variable quantities, which is critical in determining the level of cellular dysfunction [92]. A defect in Mg²⁺ reabsorption in the TAL or DCT, due to ATP depletion, could be the cause of hypomagnesemia in these patients.

Hyperuricemia, pulmonary hypertension, renal failure, and alkalosis syndrome

Hyperuricemia, pulmonary hypertension, renal failure and alkalosis (HUPRA) syndrome (OMIM #613845) is an autosomal recessive disease characterized by early-onset progressive renal failure, hyperuricemia, metabolic alkalosis, pulmonary hypertension, developmental delay, and, in some cases, hypomagnesemia [100]. This rare disorder has been diagnosed in only six children from three families and it is caused by homozygous missense mutation in SARS2 on chromosome 19, which encodes the mitochondrial seryl-tRNA synthetase (SARS2) [100,101]. SARS2 catalyzes the serine aminoacylation of two mitochondrial tRNAs. Only two mutations have been identified in patients with HUPRA syndrome, p.Asp390Gly and p.Arg402His. The p.Asp390Gly mutation significantly reduces the aminoacylation of one mitochondrial tRNA leading to its degradation, which would cause alterations in the synthesis of mitochondrial proteins and consequently in energy supply [100]. Decreased energy production may account for diminished Na⁺, K⁺-ATPase activity in the TAL and DCT, which could explain the salt wasting and hypomagnesemia observed in HUPRA patients.

Concluding remarks

 Mg^{2+} is an essential ion that plays a key role in the regulation of many cellular functions. In the kidney, proteins expressed in the apical and basolateral membranes of the TAL and DCT maintain serum Mg^{2+} levels within a narrow physiological range. Mutations in the genes that encode these proteins cause different types of hypomagnesemia. Consequently, serum Mg^{2+} levels should be determined in patients presenting with seizures, muscle cramps, and arrhythmias.

In this review, we have described the current research findings on hereditary hypomagnesemias. Some of these diseases share common characteristics and thus precise diagnosis requires the identification of the causative mutation. Therefore, we suggest a diagnostic approach for clinicians based on previously reported data [2,102] (Fig. 4). Many questions are still unanswered. For instance, the underlying mechanisms for hypomagnesemia in most hereditary diseases are still unclear. In addition, the physiological functions of *CNNM2*



Figure 4. Flowchart for the diagnosis of inherited hypomagnesemias. Group 3 hypomagnesemias were not included because the clinical features of these patients are extremely variable.

ADHH, autosomal dominant hypocalcemia with hypercalciuria; ADTKD-*HNF1B*, autosomal dominant tubulointerstitial kidney disease subtype *HNF1B*; BS3, Bartter syndrome type 3; EA1, episodic ataxia type 1; EAST, epilepsy, ataxia, sensorineural deafness, and tubulopathy; FHHNC1, familial hypomagnesemia with hypercalciuria and nephrocalcinosis type 1; FHHNC2, familial hypomagnesemia with hypercalciuria and nephrocalcinosis type 2; GS, Gitelman syndrome; HNF1B, hepatocyte nuclear factor-1 beta; HSH, hypomagnesemia with secondary hypocalcemia; HSMR1, hypomagnesemia, seizure, and mental retardation disorder type 1; HSMR2, hypomagnesemia, seizures, and mental retardation disorder type 2; IDH, isolated dominant hypomagnesemia; IRH, isolated recessive hypomagnesemia; KCS2, Kenny-Caffey syndrome type 2; MODY, maturity-onset diabetes of the young; NISBD2, neonatal inflammatory skin and bowel disease type 2; SeSAME, seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance; TNHP, transient neonatal hyperphenylalaninemia and primapterinuria.

^aThese clinical signs do not appear in all patients.

and *SLC41A1* remain to be clarified. The identification of the protein(s) that facilitate the basolateral extrusion of Mg²⁺ will greatly improve our understanding of Mg²⁺ transport in the DCT. A better knowledge of the pathophysiology of these diseases is needed to develop specific therapeutic strategies. So far, the only therapeutic option to treat hypomagnesemia in patients with GS, BS type 3, EAST/SeSAME, HSH, or isolated dominant hypomagnesemia is essentially

supportive and involves oral or intravenous magnesium supplementation. However, this approach has been found to be unsuccessful in patients with other hypomagnesemias like FHHNC, HSMR, and ADTKD-*HNF1B*.

Conflicts of interest

All authors have no conflicts of interest to declare.

Funding

This work was financially supported by the "Instituto de Salud Carlos III-Subdirección General de Evaluación y Fomento de la Investigación" and the European Regional Development Fund "Another way to build Europe" (grant number PI20/00652, Project RenalTube).

Disclosure

Figures were created using the web application BioRender. com.

Authors' contributions

Conceptualization: All authors Writing original draft: FCM, APR Review and editing: All authors All authors read and approved the final manuscript.

ORCID

Felix Claverie-Martin, https://orcid.org/0000-0003-2468-3267 Ana Perdomo-Ramirez, https://orcid.org/0000-0002-2562-2119 Victor Garcia-Nieto, https://orcid.org/0000-0003-1890-7076

References

- 1. de Baaij JH, Hoenderop JG, Bindels RJ. Magnesium in man: implications for health and disease. *Physiol Rev* 2015;95:1–46.
- 2. Viering DH, de Baaij JH, Walsh SB, Kleta R, Bockenhauer D. Genetic causes of hypomagnesemia, a clinical overview. *Pediatr Nephrol* 2017;32:1123–1135.
- **3.** Simon DB, Lu Y, Choate KA, et al. Paracellin-1, a renal tight junction protein required for paracellular Mg2+resorption. *Science* 1999;285:103–106.
- 4. Konrad M, Schaller A, Seelow D, et al. Mutations in the tight-junction gene claudin 19 (CLDN19) are associated with renal magnesium wasting, renal failure, and severe ocular involvement. *Am J Hum Genet* 2006;79:949–957.
- 5. Gong Y, Renigunta V, Himmerkus N, et al. Claudin-14 regulates renal Ca⁺⁺ transport in response to CaSR signalling via a novel microRNA pathway. *EMBO J* 2012;31:1999–2012.
- 6. Simon DB, Bindra RS, Mansfield TA, et al. Mutations in the chloride channel gene, CLCNKB, cause Bartter's syndrome type III.

Nat Genet 1997;17:171–178.

- 7. Voets T, Nilius B, Hoefs S, et al. TRPM6 forms the Mg2+influx channel involved in intestinal and renal Mg2+ absorption. *J Biol Chem* 2004;279:19–25.
- 8. Chubanov V, Waldegger S, Mederos y Schnitzler M, et al. Disruption of TRPM6/TRPM7 complex formation by a mutation in the TRPM6 gene causes hypomagnesemia with secondary hypocalcemia. *Proc Natl Acad Sci U S A* 2004;101:2894–2899.
- **9.** Franken GA, Adella A, Bindels RJ, de Baaij JH. Mechanisms coupling sodium and magnesium reabsorption in the distal convoluted tubule of the kidney. *Acta Physiol (Oxf)* 2021;231:e13528.
- 10. Kolisek M, Nestler A, Vormann J, Schweigel-Röntgen M. Human gene SLC41A1 encodes for the Na+/Mg2+ exchanger. Am J Physiol Cell Physiol 2012;302:C318–C326.
- Arjona FJ, de Baaij JH. CrossTalk opposing view: CNNM proteins are not Na+ /Mg2+ exchangers but Mg2+ transport regulators playing a central role in transepithelial Mg2+ (re)absorption. J Physiol 2018;596:747–750.
- Arjona FJ, Latta F, Mohammed SG, et al. SLC41A1 is essential for magnesium homeostasis in vivo. *Pflugers Arch* 2019;471:845– 860.
- **13.** Browne DL, Gancher ST, Nutt JG, et al. Episodic ataxia/myokymia syndrome is associated with point mutations in the human potassium channel gene, KCNA1. *Nat Genet* 1994;8:136–140.
- 14. Thebault S, Alexander RT, Tiel Groenestege WM, Hoenderop JG, Bindels RJ. EGF increases TRPM6 activity and surface expression. J Am Soc Nephrol 2009;20:78–85.
- **15.** Mayan H, Farfel Z, Karlish SJ. Renal Mg handling, FXYD2 and the central role of the Na,K-ATPase. *Physiol Rep* 2018;6:e13843.
- 16. Su XT, Ellison DH, Wang WH. Kir4.1/Kir5.1 in the DCT plays a role in the regulation of renal K + excretion. *Am J Physiol Renal Physiol* 2019;316:F582-F586.
- 17. Claverie-Martin F. Familial hypomagnesaemia with hypercalciuria and nephrocalcinosis: clinical and molecular characteristics. *Clin Kidney J* 2015;8:656–664.
- Praga M, Vara J, González-Parra E, et al. Familial hypomagnesemia with hypercalciuria and nephrocalcinosis. *Kidney Int* 1995;47:1419–1425.
- Weber S, Schneider L, Peters M, et al. Novel paracellin-1 mutations in 25 families with familial hypomagnesemia with hypercalciuria and nephrocalcinosis. *J Am Soc Nephrol* 2001;12:1872– 1881.
- **20.** Blanchard A, Jeunemaitre X, Coudol P, et al. Paracellin-1 is critical for magnesium and calcium reabsorption in the human thick ascending limb of Henle. *Kidney Int* 2001;59:2206–2215.

- **21.** Claverie-Martín F, García-Nieto V, Loris C, et al. Claudin-19 mutations and clinical phenotype in Spanish patients with familial hypomagnesemia with hypercalciuria and nephrocalcinosis. *PLoS One* 2013;8:e53151.
- 22. Konrad M, Hou J, Weber S, et al. CLDN16 genotype predicts renal decline in familial hypomagnesemia with hypercalciuria and nephrocalcinosis. *J Am Soc Nephrol* 2008;19:171–181.
- **23.** Vall-Palomar M, Burballa C, Claverie-Martín F, Meseguer A, Ariceta G. Heterogeneity is a common ground in familial hypomagnesemia with hypercalciuria and nephrocalcinosis caused by CLDN19 gene mutations. *J Nephrol* 2021;34:2053-2062.
- 24. Yamaguti PM, Neves FA, Hotton D, et al. Amelogenesis imperfecta in familial hypomagnesaemia and hypercalciuria with nephrocalcinosis caused by CLDN19 gene mutations. *J Med Genet* 2017;54:26–37.
- **25.** Bardet C, Courson F, Wu Y, et al. Claudin-16 deficiency impairs tight junction function in ameloblasts, leading to abnormal enamel formation. *J Bone Miner Res* 2016;31:498–513.
- 26. Godron A, Harambat J, Boccio V, et al. Familial hypomagnesemia with hypercalciuria and nephrocalcinosis: phenotype-genotype correlation and outcome in 32 patients with CLDN16 or CLDN19 mutations. *Clin J Am Soc Nephrol* 2012;7:801–809.
- 27. Miyamoto T, Morita K, Takemoto D, et al. Tight junctions in Schwann cells of peripheral myelinated axons: a lesson from claudin-19-deficient mice. J Cell Biol 2005;169:527–538.
- 28. Hou J, Renigunta A, Konrad M, et al. Claudin-16 and claudin-19 interact and form a cation-selective tight junction complex. *J Clin Invest* 2008;118:619–628.
- 29. Hou J, Renigunta A, Gomes AS, et al. Claudin-16 and claudin-19 interaction is required for their assembly into tight junctions and for renal reabsorption of magnesium. *Proc Natl Acad Sci U S A* 2009;106:15350–15355.
- **30.** Schlingmann KP, Sassen MC, Weber S, et al. Novel TRPM6 mutations in 21 families with primary hypomagnesemia and secondary hypocalcemia. *J Am Soc Nephrol* 2005;16:3061–3069.
- **31.** Schlingmann KP, Weber S, Peters M, et al. Hypomagnesemia with secondary hypocalcemia is caused by mutations in TRPM6, a new member of the TRPM gene family. *Nat Genet* 2002;31:166–170.
- **32.** Walder RY, Landau D, Meyer P, et al. Mutation of TRPM6 causes familial hypomagnesemia with secondary hypocalcemia. *Nat Genet* 2002;31:171–174.
- **33.** Krapivinsky G, Krapivinsky L, Renthal NE, Santa-Cruz A, Manasian Y, Clapham DE. Histone phosphorylation by TRPM6's cleaved kinase attenuates adjacent arginine methylation to reg-

ulate gene expression. *Proc Natl Acad Sci U S A* 2017;114:E7092–E7100.

- **34.** Groenestege WM, Thébault S, van der Wijst J, et al. Impaired basolateral sorting of pro-EGF causes isolated recessive renal hypomagnesemia. *J Clin Invest* 2007;117:2260–2267.
- **35.** Campbell P, Morton PE, Takeichi T, et al. Epithelial inflammation resulting from an inherited loss-of-function mutation in EGFR. *J Invest Dermatol* 2014;134:2570–2578.
- **36.** Stuiver M, Lainez S, Will C, et al. CNNM2, encoding a basolateral protein required for renal Mg2+ handling, is mutated in dominant hypomagnesemia. *Am J Hum Genet* 2011;88:333–343.
- **37.** Arjona FJ, de Baaij JH, Schlingmann KP, et al. CNNM2 mutations cause impaired brain development and seizures in patients with hypomagnesemia. *PLoS Genet* 2014;10:e1004267.
- **38.** Franken GA, Müller D, Mignot C, et al. The phenotypic and genetic spectrum of patients with heterozygous mutations in cyclin M2 (CNNM2). *Hum Mutat* 2021;42:473–486.
- **39.** Accogli A, Scala M, Calcagno A, et al. CNNM2 homozygous mutations cause severe refractory hypomagnesemia, epileptic encephalopathy and brain malformations. *Eur J Med Genet* 2019;62:198–203.
- 40. de Baaij JH, Stuiver M, Meij IC, et al. Membrane topology and intracellular processing of cyclin M2 (CNNM2). *J Biol Chem* 2012;287:13644–13655.
- **41.** Chen YS, Kozlov G, Fakih R, et al. Mg 2+-ATP sensing in CNNM, a putative magnesium transporter. *Structure* 2020;28:324–335.
- 42. Rodríguez-Soriano J. Bartter and related syndromes: the puzzle is almost solved. *Pediatr Nephrol* 1998;12:315–327.
- 43. Konrad M, Nijenhuis T, Ariceta G, et al. Diagnosis and management of Bartter syndrome: executive summary of the consensus and recommendations from the European Rare Kidney Disease Reference Network Working Group for Tubular Disorders. *Kidney Int* 2021;99:324–335.
- 44. Seys E, Andrini O, Keck M, et al. Clinical and genetic spectrum of Bartter syndrome type 3. *J Am Soc Nephrol* 2017;28:2540–2552.
- 45. Konrad M, Vollmer M, Lemmink HH, et al. Mutations in the chloride channel gene CLCNKB as a cause of classic Bartter syndrome. *J Am Soc Nephrol* 2000;11:1449–1459.
- 46. Blanchard A, Bockenhauer D, Bolignano D, et al. Gitelman syndrome: consensus and guidance from a Kidney Disease: Improving Global Outcomes (KDIGO) Controversies Conference. *Kidney Int* 2017;91:24–33.
- **47.** Cruz DN, Shaer AJ, Bia MJ, Lifton RP, Simon DB; Yale Gitelman's and Bartter's Syndrome Collaborative Study Group. Gitelman's syndrome revisited: an evaluation of symptoms and health-re-

lated quality of life. *Kidney Int* 2001;59:710–717.

- 48. Simon DB, Nelson-Williams C, Bia MJ, et al. Gitelman's variant of Bartter's syndrome, inherited hypokalaemic alkalosis, is caused by mutations in the thiazide-sensitive Na-Cl cotransporter. *Nat Genet* 1996;12:24–30.
- **49.** Vargas-Poussou R, Dahan K, Kahila D, et al. Spectrum of mutations in Gitelman syndrome. *J Am Soc Nephrol* 2011;22:693–703.
- McCormick JA, Ellison DH. Distal convoluted tubule. *Compr Physiol* 2015;5:45–98.
- 51. Nijenhuis T, Vallon V, van der Kemp AW, Loffing J, Hoenderop JG, Bindels RJ. Enhanced passive Ca2+ reabsorption and reduced Mg2+ channel abundance explains thiazide-induced hypocalciuria and hypomagnesemia. *J Clin Invest* 2005;115:1651–1658.
- **52.** Pearce SH, Williamson C, Kifor O, et al. A familial syndrome of hypocalcemia with hypercalciuria due to mutations in the calcium-sensing receptor. *N Engl J Med* 1996;335:1115–1122.
- **53.** Hannan FM, Kallay E, Chang W, Brandi ML, Thakker RV. The calcium-sensing receptor in physiology and in calcitropic and noncalcitropic diseases. *Nat Rev Endocrinol* 2018;15:33–51.
- 54. Pollak MR, Brown EM, Estep HL, et al. Autosomal dominant hypocalcaemia caused by a Ca(2+)-sensing receptor gene mutation. *Nat Genet* 1994;8:303–307.
- 55. Loupy A, Ramakrishnan SK, Wootla B, et al. PTH-independent regulation of blood calcium concentration by the calcium-sensing receptor. *J Clin Invest* 2012;122:3355–3367.
- 56. Paulhus K, Ammerman L, Glasscock E. Clinical spectrum of KCNA1 mutations: new insights into episodic ataxia and epilepsy comorbidity. *Int J Mol Sci* 2020;21:2802.
- **57.** Graves TD, Cha YH, Hahn AF, et al. Episodic ataxia type 1: clinical characterization, quality of life and genotype-phenotype correlation. *Brain* 2014;137(Pt 4):1009–1018.
- 58. Glaudemans B, van der Wijst J, Scola RH, et al. A missense mutation in the Kv1.1 voltage-gated potassium channel-encoding gene KCNA1 is linked to human autosomal dominant hypomagnesemia. *J Clin Invest* 2009;119:936–942.
- 59. van der Wijst J, Konrad M, Verkaart SA, et al. A de novo KCNA1 mutation in a patient with tetany and hypomagnesemia. *Nephron* 2018;139:359–366.
- **60.** van der Wijst J, Glaudemans B, Venselaar H, et al. Functional analysis of the Kv1.1 N255D mutation associated with autosomal dominant hypomagnesemia. *J Biol Chem* 2010;285:171–178.
- **61.** Geven WB, Monnens LA, Willems JL, Buijs W, Hamel CJ. Isolated autosomal recessive renal magnesium loss in two sisters. *Clin Genet* 1987;32:398–402.
- 62. Meij IC, Koenderink JB, van Bokhoven H, et al. Dominant

isolated renal magnesium loss is caused by misrouting of the Na(+),K(+)-ATPase gamma-subunit. *Nat Genet* 2000;26:265–266.

- **63.** de Baaij JH, Dorresteijn EM, Hennekam EA, et al. Recurrent FXYD2 p.Gly41Arg mutation in patients with isolated dominant hypomagnesaemia. *Nephrol Dial Transplant* 2015;30:952–957.
- 64. Arystarkhova E, Sweadner KJ. Splice variants of the gamma subunit (FXYD2) and their significance in regulation of the Na, K-ATPase in kidney. *J Bioenerg Biomembr* 2005;37:381–386.
- 65. Schlingmann KP, Bandulik S, Mammen C, et al. Germline de novo mutations in ATP1A1 cause renal hypomagnesemia, re-fractory seizures, and intellectual disability. *Am J Hum Genet* 2018;103:808–816.
- 66. Lassuthova P, Rebelo AP, Ravenscroft G, et al. Mutations in AT-P1A1 cause dominant charcot-marie-tooth type 2. *Am J Hum Genet* 2018;102:505–514.
- 67. Bockenhauer D, Feather S, Stanescu HC, et al. Epilepsy, ataxia, sensorineural deafness, tubulopathy, and KCNJ10 mutations. *N Engl J Med* 2009;360:1960–1970.
- 68. Scholl UI, Choi M, Liu T, et al. Seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance (SeSAME syndrome) caused by mutations in KCNJ10. *Proc Natl Acad Sci U S A* 2009;106:5842–5847.
- **69.** Bandulik S, Schmidt K, Bockenhauer D, et al. The salt-wasting phenotype of EAST syndrome, a disease with multifaceted symptoms linked to the KCNJ10 K+ channel. *Pflugers Arch* 2011;461:423–435.
- **70.** Scholl UI, Dave HB, Lu M, et al. SeSAME/EAST syndrome--phenotypic variability and delayed activity of the distal convoluted tubule. *Pediatr Nephrol* 2012;27:2081–2090.
- **71.** Devuyst O, Olinger E, Weber S, et al. Autosomal dominant tubulointerstitial kidney disease. *Nat Rev Dis Primers* 2019;5:60.
- 72. Adalat S, Woolf AS, Johnstone KA, et al. HNF1B mutations associate with hypomagnesemia and renal magnesium wasting. J Am Soc Nephrol 2009;20:1123–1131.
- 73. Faguer S, Decramer S, Chassaing N, et al. Diagnosis, management, and prognosis of HNF1B nephropathy in adulthood. *Kidney Int* 2011 Oct;80:768–776.
- 74. Horikawa Y, Iwasaki N, Hara M, et al. Mutation in hepatocyte nuclear factor-1 beta gene (TCF2) associated with MODY. *Nat Genet* 1997;17:384–385.
- **75.** Bingham C, Bulman MP, Ellard S, et al. Mutations in the hepatocyte nuclear factor-1beta gene are associated with familial hypoplastic glomerulocystic kidney disease. *Am J Hum Genet* 2001;68:219–224.
- 76. Ferrè S, Igarashi P. New insights into the role of HNF-1 β in kidney

(patho)physiology. Pediatr Nephrol 2019;34:1325-1335.

- 77. Kompatscher A, de Baaij JHF, Aboudehen K, et al. Loss of transcriptional activation of the potassium channel Kir5.1 by HNF1 β drives autosomal dominant tubulointerstitial kidney disease. *Kidney Int* 2017;92:1145–1156.
- **78.** Thöny B, Neuheiser F, Kierat L, et al. Hyperphenylalaninemia with high levels of 7-biopterin is associated with mutations in the PCBD gene encoding the bifunctional protein pterin-4a-carbinolamine dehydratase and transcriptional coactivator (DCoH). *Am J Hum Genet* 1998;62:1302–1311.
- **79.** Ferrè S, de Baaij JH, Ferreira P, et al. Mutations in PCBD1 cause hypomagnesemia and renal magnesium wasting. *J Am Soc Nephrol* 2014;25:574–586.
- **80.** Johnen G, Kaufman S. Studies on the enzymatic and transcriptional activity of the dimerization cofactor for hepatocyte nuclear factor 1. *Proc Natl Acad Sci U S A* 1997;94:13469–13474.
- **81.** Mendel DB, Khavari PA, Conley PB, et al. Characterization of a cofactor that regulates dimerization of a mammalian homeodomain protein. *Science* 1991;254:1762–1767.
- 82. Unger S, Górna MW, Le Béchec A, et al. FAM111A mutations result in hypoparathyroidism and impaired skeletal development. *Am J Hum Genet* 2013;92:990–995.
- **83.** Isojima T, Doi K, Mitsui J, et al. A recurrent de novo FAM111A mutation causes Kenny-Caffey syndrome type 2. *J Bone Miner Res* 2014;29:992–998.
- 84. Nikkel SM, Ahmed A, Smith A, Marcadier J, Bulman DE, Boycott KM. Mother-to-daughter transmission of Kenny-Caffey syndrome associated with the recurrent, dominant FAM111A mutation p.Arg569His. *Clin Genet* 2014;86:394–395.
- **85.** Cheng SS, Chan PK, Luk HM, Mok MT, Lo IF. Adult Chinese twins with Kenny-Caffey syndrome type 2: a potential age-dependent phenotype and review of literature. *Am J Med Genet A* 2021;185:636–646.
- 86. Fine DA, Rozenblatt-Rosen O, Padi M, et al. Identification of FA-M111A as an SV40 host range restriction and adenovirus helper factor. *PLoS Pathog* 2012;8:e1002949.
- **87.** Kojima Y, Machida Y, Palani S, et al. FAM111A protects replication forks from protein obstacles via its trypsin-like domain. *Nat Commun* 2020;11:1318.
- **88.** Hoffmann S, Pentakota S, Mund A, et al. FAM111 protease activity undermines cellular fitness and is amplified by gain-of-func-

tion mutations in human disease. EMBO Rep 2020;21:e50662.

- **89.** Nie M, Oravcová M, Jami-Alahmadi Y, et al. FAM111A induces nuclear dysfunction in disease and viral restriction. *EMBO Rep* 2021;22:e50803.
- **90.** Tan RS, Lee CHL, Dimke H, Todd Alexander R. The role of calcium-sensing receptor signaling in regulating transpithelial calcium transport. Exp Biol Med (Maywood) 2021;246:2407-2419.
- **91.** Bhargava P, Schnellmann RG. Mitochondrial energetics in the kidney. *Nat Rev Nephrol* 2017;13:629–646.
- 92. Gorman GS, Chinnery PF, DiMauro S, et al. Mitochondrial diseases. *Nat Rev Dis Primers* 2016;2:16080.
- **93.** Govers LP, Toka HR, Hariri A, Walsh SB, Bockenhauer D. Mitochondrial DNA mutations in renal disease: an overview. *Pediatr Nephrol* 2021;36:9–17.
- 94. Wilson FH, Hariri A, Farhi A, et al. A cluster of metabolic defects caused by mutation in a mitochondrial tRNA. *Science* 2004;306: 1190–1194.
- **95.** Ashraf SS, Ansari G, Guenther R, Sochacka E, Malkiewicz A, Agris PF. The uridine in "U-turn": contributions to tRNA-ribosomal binding. *RNA* 1999;5:503–511.
- 96. Konrad M, Schlingmann KP, Gudermann T. Insights into the molecular nature of magnesium homeostasis. Am J Physiol Renal Physiol 2004;286:F599–F605.
- **97.** Mancuso M, Orsucci D, Angelini C, et al. Redefining phenotypes associated with mitochondrial DNA single deletion. *J Neurol* 2015;262:1301–1309.
- 98. Katsanos KH, Elisaf M, Bairaktari E, Tsianos EV. Severe hypomagnesemia and hypoparathyroidism in Kearns-Sayre syndrome. *Am J Nephrol* 2001;21:150–153.
- 99. Emma F, Pizzini C, Tessa A, et al. "Bartter-like" phenotype in Kearns-Sayre syndrome. *Pediatr Nephrol* 2006;21:355–360.
- 100. Belostotsky R, Ben-Shalom E, Rinat C, et al. Mutations in the mitochondrial seryl-tRNA synthetase cause hyperuricemia, pulmonary hypertension, renal failure in infancy and alkalosis, HUPRA syndrome. *Am J Hum Genet* 2011;88:193–200.
- **101.** Rivera H, Martín-Hernández E, Delmiro A, et al. A new mutation in the gene encoding mitochondrial seryl-tRNA synthetase as a cause of HUPRA syndrome. *BMC Nephrol* 2013;14:195.
- 102. Horinouchi T, Nozu K, Kamiyoshi N, et al. Diagnostic strategy for inherited hypomagnesemia. *Clin Exp Nephrol* 2017;21:1003– 1010.