



REVIEW

Recent Advances in Understanding Cholangiocarcinoma [version 1; referees: 2 approved]

Lindsey Kennedy^{1,2}, Laura Hargrove¹, Jennifer Demieville², Nicole Francis³,
Rowan Seils¹, Sara Villamaria¹, Heather Francis ¹⁻³

¹Department of Internal Medicine, Texas A&M Health Science Center, College of Medicine, Bryan, TX, USA

²Research, Central Texas Veterans Health Care System, Temple, TX, USA

³Baylor Scott & White Health Digestive Disease Research Center, Temple, TX, USA

v1 **First published:** 09 Oct 2017, 6(F1000 Faculty Rev):1818 (doi: 10.12688/f1000research.12118.1)
Latest published: 09 Oct 2017, 6(F1000 Faculty Rev):1818 (doi: 10.12688/f1000research.12118.1)

Abstract

Cholangiocarcinoma (CCA) is an aggressive malignancy that arises from damaged epithelial cells, cholangiocytes, and possibly de-differentiated hepatocytes. CCA has a poor overall survival rate and limited therapeutic options. Based on this data, it is imperative that new diagnostic and therapeutic interventions be developed. Recent work has attempted to understand the pathological mechanisms driving CCA progression. Specifically, recent publications have delved into the role of cancer stem cells (CSCs), mesenchymal stem cells (MSCs), and microRNAs (miRNAs) during CCA pathology. CSCs are a specific subset of cells within the tumor environment that are derived from a cell with stem-like properties and have been shown to influence recurrence and chemoresistance during CCA. MSCs are known for their anti-inflammatory activity and have been postulated to influence malignancy during CCA, but little is known about their exact functions. miRNAs exert various functions via gene regulation at both the transcriptional and the translational levels, giving miRNAs diverse roles in CCA progression. Additionally, current miRNA-based therapeutic approaches are in clinical trials for various liver diseases, giving hope for similar approaches for CCA. However, the interactions among these three factors in the context of CCA are unknown. In this review, we focus on recently published data (within the last 3 years) that discuss the role of CSCs, MSCs, and miRNAs and their possible interactions during CCA pathogenesis.

Open Peer Review

Referee Status:

	Invited Referees	
	1	2
version 1 published 09 Oct 2017		

F1000 Faculty Reviews are commissioned from members of the prestigious F1000 Faculty. In order to make these reviews as comprehensive and accessible as possible, peer review takes place before publication; the referees are listed below, but their reports are not formally published.

- 1 **Li Wang**, University of Connecticut, USA
- 2 **Jesper B Anderson**, University of Copenhagen, Denmark

Discuss this article

Comments (0)

Corresponding author: Heather Francis (HFrancis@medicine.tamhsc.edu)

Author roles: **Kennedy L:** Conceptualization, Resources, Supervision, Validation, Writing – Review & Editing; **Hargrove L:** Conceptualization, Investigation, Writing – Original Draft Preparation, Writing – Review & Editing; **Demieville J:** Writing – Review & Editing; **Francis N:** Writing – Review & Editing; **Seils R:** Writing – Review & Editing; **Villamaria S:** Writing – Review & Editing; **Francis H:** Writing – Review & Editing

Competing interests: The authors declare that they have no competing interests.

How to cite this article: Kennedy L, Hargrove L, Demieville J *et al.* **Recent Advances in Understanding Cholangiocarcinoma [version 1; referees: 2 approved]** *F1000Research* 2017, 6(F1000 Faculty Rev):1818 (doi: [10.12688/f1000research.12118.1](https://doi.org/10.12688/f1000research.12118.1))

Copyright: © 2017 Kennedy L *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution Licence](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Grant information: Portions of this work were supported by (i) a VA Merit Award (1101BX003031) from the US Department of Veterans Affairs, Biomedical Laboratory Research and Development Service and (ii) the National Institute of Health National Institute of Diabetes and Digestive and Kidney Diseases (DK108959). This material is the result of work supported with resources and the use of facilities at the Central Texas Veterans Health Care System (Temple, TX, USA). The content is the responsibility of the authors alone and does not necessarily reflect the views or policies of the Department of Veterans Affairs or the US Government.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

First published: 09 Oct 2017, 6(F1000 Faculty Rev):1818 (doi: [10.12688/f1000research.12118.1](https://doi.org/10.12688/f1000research.12118.1))

Introduction

Cholangiocarcinoma (CCA) is a hepatobiliary malignancy with a poor 5-year survival rate and limited treatment options. CCA arises from damaged cholangiocytes, the epithelial cells that line the biliary tree. CCA can be classified into intrahepatic, perihilar, or distal subtypes¹. Intrahepatic CCA, the second most common form of liver cancer, is generally located proximal to the second-order bile ducts, perihilar CCA is located between the second-order bile ducts and the intersection of the cystic duct into the common bile duct, and distal CCA is found in areas between the cystic duct and the ampulla of Vater (Figure 1)². Aside from being categorized by anatomic location, CCA is categorized on the basis of histopathological analysis and by growth-type patterns as well^{3,4}. For instance, CCA that has been derived from large cholangiocytes is predominantly categorized as perihilar CCA with well to moderately differentiated mucin-producing cells and periductal infiltrating growth pattern, whereas CCA that is derived from small cholangiocytes is predominantly categorized as intrahepatic CCA and largely contains non-mucin-producing cells and has mass-forming growth patterns^{3,4}. CCA is highly heterogeneous not only in initiation and location but also in progression, making it difficult to categorize CCA into distinct subtypes.

CCA incidence has been steadily increasing⁵. One of the first lines of treatment for CCA is resection; however, options for resection are limited since the disease is generally too advanced at the time of diagnosis⁶. Recently, it has been reported that following resection in patients with intrahepatic CCA the 5-year

and 10-year survival rates are 32.3% and 8.4%, respectively⁷. If unresectable, liver transplantation is another option for patients with intrahepatic CCA, and post-liver transplantation 5-year survival rates are 51%⁸. For patients with perihilar CCA, resection may be an option but outcomes are poor; 5-year survival rates are 10%⁹. It has also been suggested that some subsets of perihilar CCA may benefit from neoadjuvant chemoradiation therapy or biliary drainage or both, but outcomes are variable¹⁰⁻¹². For patients with distal CCA following resection, 5-year survival rates are 23%⁹. Furthermore, these patients should not be subjected to biliary drainage, since they tend to have increased complications related to this surgery¹³. It is apparent that our current approaches for CCA are lacking, as demonstrated by the poor survival rates and limited treatment options. For these reasons, it is critical that new and effective therapies be developed.

The CCA microenvironment contains a plethora of cell types, extracellular structures, and secreted factors that influence the progression of the tumor. Cancer cells are surrounded by stroma, which is made up of various stromal cells and extracellular matrix¹⁴. Additionally, cytokines, chemokines, growth factors, and proteinases secreted from cancer cells and stromal cells can promote a pro-inflammatory environment¹⁴. This type of tumor microenvironment can lead to myofibroblast activation, cancer stem cell (CSC) initiation, and the recruitment of various inflammatory cell types¹⁴. The CCA microenvironment is a complex network that is influenced by a multitude of soluble factors and cell types.

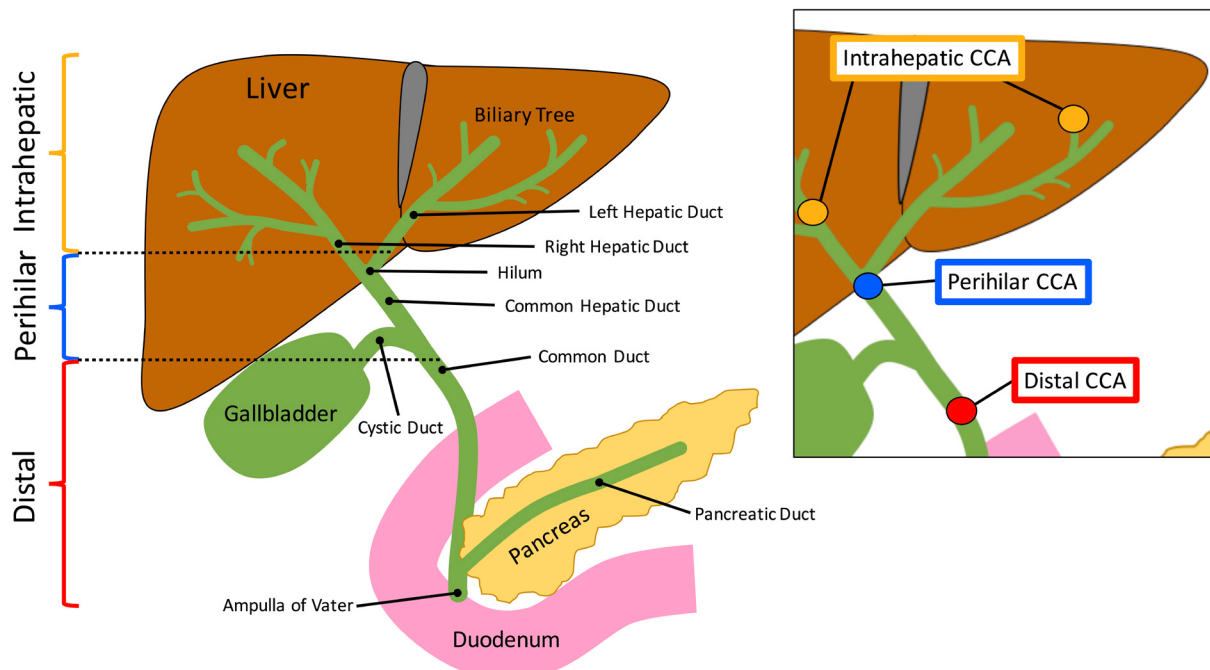


Figure 1. Schematic image of the anatomical locations of intrahepatic, perihilar, and distal cholangiocarcinoma (CCA). CCA is categorized on the basis of its location within the biliary tree. Intrahepatic CCA is found proximal to the second-order bile ducts, perihilar CCA is found between the second-order bile ducts and the convergence of the cystic duct and the common duct, and distal CCA is found between the cystic duct and ampulla of Vater.

This review will focus on recent publications (within the last 3 years) that advance our understanding of CCA and will analyze how their findings could lead to new diagnostic or therapeutic targets. Overall, these publications discuss their findings within the context of all three subtypes of CCA (intrahepatic, perihilar, and distal), and for this reason we discuss the findings of the articles in the context of general CCA. Among the recent publications, many of the articles fell into similar categories. Specifically, publications tended to focus on the functional and mechanistic roles of (i) CSCs, (ii) mesenchymal stem cells (MSCs), or (iii) microRNAs (miRNAs) in CCA progression. We will be discussing how these three categories influence the CCA microenvironment, either alone or in relation to one another.

Cancer stem cells

The tumor microenvironment comprises many different cell types with various functions. Recently, CSCs have been identified in the tumor microenvironment and add complexity to our understanding of CCA. Specifically, CSCs have been identified for their role in tumor initiation, progression, and relapse. CSCs function in a manner similar to normal stem cells, wherein they are capable of self-renewal, can differentiate into multiple different cell types, and have unlimited division¹⁵. CSCs promote an inflammatory environment through the activation of stromal cells and the recruitment of inflammatory cells¹⁶. The cell type from which CSCs are derived is still under debate, but it has been hypothesized that CSCs may be derived from a stem-like cell that acquires a cancer-promoting alteration¹⁷. Multiple studies have found that hepatocellular carcinoma (HCC) progression is driven by CSCs^{18–21}; however, little is known about the role of CSCs in CCA.

As previously stated, the CCA microenvironment is a complex network of various cell types. Among the various cell types, macrophages are associated with CCA tumor progression and are significantly correlated with a poorer prognosis and metastasis⁴. A recent article by Raggi *et al.* hypothesized that CSCs secrete factors that promote macrophage differentiation toward the tumor-associated macrophage (TAM) subtype²². First, CCA spheres were cultured and medium was collected and placed on cultured macrophages. Following treatment with CCA sphere medium, macrophages expressed cluster of differentiation 68 (CD68), CD115, human leukocyte antigen-D related, and CD206, indicating an activated phenotype. These activated macrophages had TAM-like features, including increased invasiveness. To identify whether these findings were clinically relevant, macrophages were isolated from human CCA resections, and it was noted that these isolated macrophages expressed a phenotype similar to that of the treated cultured macrophages. The authors then identified the factors secreted from CCA spheres that were driving this TAM-like phenotype and found that interleukin-13 (IL-13), IL-34, and osteoactivin were detected in CCA sphere medium and in serum of patients with CCA. Furthermore, these factors were associated with CSC presence, and the authors concluded that the secreted factors driving the TAM-like phenotype must be derived from CSCs. These findings present a novel mechanism by which CCA-associated CSCs influence the activation of TAMs that promote CCA progression²². Given that much of the

work was done with CCA sphere medium, more work regarding the exact factors secreted from CSCs is necessary to definitively demonstrate that the activation of TAMs is specifically driven by CSCs.

Laminins are a family of extracellular matrix proteins that are mainly found in the basement membrane and are composed of α , β , and γ chains^{23,24}. The γ 2-chain of laminin-332 (Ln-332) is highly elevated in HCCs expressing the biliary marker cytokeratin-19 (CK-19) and is correlated with a poor prognosis for patients with HCC or intrahepatic CCA^{25–27}. The aim of the publication by Govaere *et al.* was to characterize the CSC niche and determine the possible cell of origin²⁸. This publication slightly deviates from the others discussed in this review because it looks at CSCs in mixed HCC/CCA samples. It was noted that elevated biliary and hepatic progenitor cell (HPC) markers were seen with increased expression of *LAMC2*, the gene that encodes the γ 2-chain of Ln-332. Immunopositivity for the γ 2-chain of Ln-332 was found surrounding small HPC-like tumor cells that had a low proliferation rate, identified as possible CSCs. Lastly, the γ 2-chain of Ln-332 was strongly co-expressed in ductular areas with low proliferative capacity but was expressed at low levels in the hepatocyte areas of HCC/CCA. The authors concluded that Ln-332 maintains the CSC niche and supports stem-like properties in these cells²⁸. While this article is one of a few that looks at the factors that maintain CSCs, future work needs to be performed to look at the role of Ln-332 specifically in CCA samples.

Given these findings, it is evident that the formation of CSCs strongly impacts the surrounding tumor microenvironment, which can impact disease progression. Specifically, these articles identify the pro-inflammatory role of CSCs during CCA. First, CSCs were shown to promote macrophage transition to a TAM-like phenotype. Furthermore, CSC accumulation of laminins, a key component of extracellular matrix, may promote the survival, migration, and invasion of tumor cells as well as stromal activation. It is evident that CSCs are key in maintaining and promoting a pro-inflammatory environment during CCA initiation and progression.

Mesenchymal stem cells

Recently, interest in the role of MSCs in tumor progression and metastasis has increased. MSCs are non-hematopoietic stem cells that primarily reside in the bone marrow but are recruited to injured tissues, inflammatory sites, and primary tumors^{29–31}. As stated above, the presence of CSCs promotes an inflammatory environment, which may promote MSC migration and the infiltration of CCA tumors. MSCs have low immunogenicity and, following recruitment to injured tissues, are able to maintain their multi-differentiation capacity³². Specifically, MSCs may differentiate into cancer-associated fibroblasts, myofibroblasts, or hepatic stellate cells to promote tumor progression^{33,34}. These various cell types increase the inflammatory capacity of the stromal environment, further promoting the initiation of CSCs and the recruitment of MSCs. Moreover, MSCs secrete various cytokines that promote an inflammatory microenvironment^{35,36}. For these reasons, the role of MSCs in CCA progression and inflammatory response has become a topic of interest.

While MSCs are known to influence the inflammatory environment of tumors, the exact microenvironment necessary for these effects is unknown. Also, it has been postulated that inflammatory conditions are a major activator of MSC-associated immunosuppression³⁷. Recent work from Zhong *et al.* found that tumor necrosis factor- α (TNF- α) and interferon- γ (IFN- γ) were able to stimulate the expression of TNF- α , C-C motif chemokine ligand 5 (CCL5), IL-6 and indoleamine 2,3-dioxygenase, and activated nuclear factor kappa B (NF- κ B) signaling in MSCs³⁸. Secreted factors from these stimulated MSCs were able to induce CCA cell migration and metastasis *in vitro* and *in vivo*. Furthermore, CCA cells treated with supernatants from the stimulated MSCs had increased expression of C-C motif chemokine receptor 5 (CCR5). Increased CCL5/CCR5 signaling in CCA cells was able to increase the expression of matrix metalloproteinase-2 (MMP-2) and MMP-9. Overall, the authors deduced that TNF- α and IFN- γ stimulate MSCs to secrete CCL5, which in turn activates CCR5 on CCA cells to promote inflammation and metastasis³⁸. Inhibition of TNF- α or IFN- γ signaling or both could block MSC recruitment, thereby reducing CCA inflammation and metastasis to hopefully contribute to an improved outcome.

Chemoresistance is a major obstacle in cancer treatment. Previously, it was demonstrated that MSC-secreted IL-8 induces doxorubicin resistance in triple-negative breast cancer³⁹. Considering these findings, Wang *et al.* examined the role of MSCs in the progression of CCA development⁴⁰. Using human umbilical cord-derived MSCs, the authors noted that cultured CCA cells that were co-cultured with these MSCs had increased cell proliferation, metastasis, and resistance to the anti-cancer drug compound K *in vivo*. *In vitro*, CCA cells co-cultured with MSCs had increased colony formation and invasion. The tumors from mice injected with CCA cells co-cultured with MSCs demonstrated enhanced Wnt/ β -catenin signaling. These findings were corroborated *in vitro* where MSCs and their secreted factors were shown to stimulate Wnt signaling via nuclear translocation of β -catenin, upregulation of Wnt, and increased expression of the downstream targets MMP-2, cyclin D1, and c-Myc in cultured CCA cells. Overall, this study indicates that MSCs increase CCA metastasis and chemoresistance via increased Wnt/ β -catenin signaling⁴⁰. Targeting the Wnt/ β -catenin signaling pathway may prove therapeutic for patients with CCA.

It is interesting to note that TNF- α and IFN- γ are necessary to drive MSC migration to CCA tumors. Given that CSCs promote an inflammatory environment, this may play a role in the recruitment of MSCs to the CCA tumor. Based on these articles, the presence of MSCs in CCA is indicative of increased CCA migration and invasion, and inhibition of MSC migration or induction of MSC apoptosis may be therapeutic for patients with CCA.

microRNAs

miRNAs are short, non-coding RNAs that regulate the expression of specific mRNAs⁴¹. Specifically, miRNAs will recognize and bind to complementary sequences found within the 3' untranslated region (3' UTR) of specific mRNAs to regulate their expression levels^{42,43}. The role of miRNAs varies, depending on the cellular process, in both physiological and pathological conditions⁴⁴⁻⁴⁶. The

function of miRNAs during CCA has increasingly become a topic of interest; multiple human clinical trials evaluating the efficacy of miRNA-based therapeutics during various liver diseases are under way^{47,48}. miRNA signaling has been noted in all three subtypes of CCA⁴⁹⁻⁵¹; however, the identification of subtype-specific miRNAs has not been made. It is known that miRNAs can regulate MSC and CSC function⁵²⁻⁵⁴, but this regulation has not been reported in the context of CCA. Further understanding the impact of miRNAs on CCA development and progression may bring about novel diagnostic tools or therapeutic interventions. In this section, we will be discussing the role of miRNAs during CCA progression but also analyzing their potential impacts on CSC and MSC biology.

Menin is a known tumor-suppressor gene that is expressed in all tissues⁵⁵, but its role in CCA is poorly defined. miR-24 has previously been recognized as an oncogene in multiple gastrointestinal cancers and has been shown to target menin, but this interaction in the context of CCA is not understood⁵⁶⁻⁵⁹. A recent article by Ehrlich *et al.* evaluated the role of menin during CCA proliferation and angiogenesis and its regulation by miR-24⁵¹. The authors found that human advanced CCA tumor sections, as well as *in vitro* human CCA cell lines, had increased miR-24 expression alongside decreased menin expression. *In vitro*, human CCA cell lines treated with an miR-24 inhibitor showed increased menin levels with a subsequent reduction in cell proliferation, angiogenesis, migration, and invasion. It was then demonstrated that miR-24 negatively regulates menin expression. *In vivo*, the authors noted that tumor size and the expression of proliferative and angiogenic markers were significantly reduced in the miR-24-inhibited tumor group compared with controls. Interestingly, fibrogenesis was enhanced in the miR-24-inhibited tumor group when compared with controls. From this study, it is evident that miR-24 acts as an oncogene to suppress menin expression, thereby increasing tumor burden, proliferation, angiogenesis, migration, and invasion⁵¹. However, further work is necessary to delineate the exact downstream mechanisms regulating these events as well as understand the primary cellular source of miR-24 during CCA, which may help us understand why fibrogenesis increased even as tumor burden and angiogenesis decreased. While this study focuses on the role of miR-24 in cultured CCA cells, miR-24 has previously been shown to promote stemness in embryonic stem cells⁶⁰. In this context, miR-24 may promote the formation of CSCs in CCA, leading to increased CCA migration and invasion.

Circadian rhythms are endogenous oscillations that are regulated by clock genes and are present in the central nervous system, peripheral tissues, and single cells⁶¹. While circadian oscillations are a normal physiological process, dysregulation of these oscillations promotes tumor development^{62,63}; however, little is known about the role of the circadian rhythm during CCA. A recent publication identified that the expression of Per1, which negatively regulates circadian oscillations, was decreased in human CCA samples and cultured human CCA cell lines⁶⁴. In addition, overexpression of Per1 decreased cell proliferation and increased apoptosis in cultured human CCA cells. These findings were mimicked *in vivo*, wherein immunocompromised mice injected with cultured human CCA cells overexpressing Per1 had decreased tumor growth, proliferation, angiogenesis, and metastasis. Per1

was found to be a target of miR-34a, and following treatment with a miR-34a inhibitor, human cultured CCA cells had decreased proliferation, migration, and invasion. These findings are the first to identify that the disruption of clock genes regulated by miR-34a contributes to CCA malignancy⁶⁴. As stated, circadian clock genes are present in all cells and tissues, and previous work has shown that circadian clock genes regulate MSC differentiation, migration, and cell cycle⁶⁵. It is possible that the regulation of core clock genes via miR-34a plays a role in MSC migration and activation during CCA. Also, the disruption of core clock genes in CCA cells may contribute to the tumor niche, thereby supporting CSC formation or MSC recruitment or both.

Tumor growth and metastatic potential are regulated by a myriad of factors, including epigenetic changes, such as DNA methylation^{66,67}. Specifically, it has been shown that DNA methylation plays a prominent role in the progression of CCA⁶⁸. On the basis of this information, Zhou *et al.* set out to investigate the functional and mechanistic role of miR-191 in CCA⁶⁹. It was first noted that miR-191 expression was increased in CCA tumors when compared with adjacent normal bile duct tissue. miR-191 expression was found to be an independent risk factor for a worse prognosis for human CCA patients. *In vivo* and *in vitro* analysis revealed that overexpression of miR-191 was associated with enhanced proliferation, invasion, and migration and reduced ten-eleven translocation 1 (TET1) expression, which induces DNA demethylation and was shown to be a direct target of miR-191. The authors then found that TET1 expression allows for the methylation of CpG-rich regions in the gene transcription start site of p53, a tumor suppressor, leading to a reduction of p53 expression and subsequent increase in tumor burden. These findings suggest that the overexpression of miR-191 is associated with CCA progression via miR-191/TET1/p53 signaling⁶⁹. Aside from its role in p53 expression⁷⁰, miR-191 has been shown to promote CSC-like properties in bronchial epithelial cells, alluding to the fact that miR-191 may influence the CSC niche in CCA as well. This publication identifies a novel therapeutic target for the treatment of CCA and suggests that miR-191 levels may be used as a diagnostic or prognostic factor or both.

Aside from p53, N-myc downstream-regulated gene 2 (NDRG2) is another tumor suppressor that plays a role in the progression of various cancers^{71,72}. NDRG2 has been shown to be downregulated during tumor progression, and it has been postulated that NDRG2 inhibits the metastasis of HCC⁷¹⁻⁷³. miR-181 is upregulated in several cancers⁷⁴⁻⁷⁶; however, miR-181 has also been shown to inhibit tumor formation⁷⁷⁻⁷⁹. Leukemia inhibitory factor (LIF) is a member of the IL-6 cytokine family whose dysregulation has been observed in different cancers⁷⁹. The potential function of NDRG2, LIF, and miR-181c in the development and progression of CCA is not fully understood, and a recent publication evaluated the potential roles and mechanisms of these factors⁸⁰. The authors found that the expression of NDRG2 was decreased and miR-181c and LIF increased in human CCA compared with non-tumor tissues. Furthermore, downregulation of NDRG2 alongside overexpression of miR-181c or LIF indicated a poorer overall survival in patients with CCA. *In vivo* and *in vitro*, it was shown that overexpression of NDRG2 was able to inhibit CCA

cell proliferation, chemoresistance, and metastasis. LIF was able to activate miR-181c, while NDRG2 inhibited LIF transcription. These findings identify that NDRG2 and LIF/miR-181c counteract each other, and dysregulation of one of these pathways may contribute to carcinogenesis and metastasis⁸⁰. Given that LIF is an IL-6 cytokine, miR-181c regulation of LIF may influence the inflammatory properties of the microenvironment so that it favors CSC formation or MSC recruitment. Though complex, this novel pathway may serve as a potential therapeutic target for the treatment of CCA.

The role of miR-16 in tumorigenesis has been studied in various cancers⁸¹⁻⁸³, but its role in CCA is unknown. Yes-associated protein 1 (YAP1) is inhibited by the hippo signaling pathway, but in the absence of this inhibition, YAP1 acts as an oncogene in numerous cancers⁸⁴⁻⁸⁸; however, mechanisms regulating YAP1 expression in human CCA are largely unknown. Recent work found that in human CCA tissues miR-16 expression was significantly downregulated and correlated with tumor size, metastasis, and staging⁸⁹. Furthermore, downregulation of miR-16 was strongly associated with increased tumor progression and worsened overall survival in human CCA patients. *In vivo* and *in vitro* experimentation demonstrated that overexpression of miR-16 inhibited CCA cell proliferation, invasion, and metastasis. The authors then found that YAP1 was a direct target of miR-16. In support of these findings, it was noted that YAP1 levels are greatly enhanced in human CCA tissues, which was inversely correlated with the above findings that miR-16 is largely downregulated in human CCA. This publication concludes that miR-16 acts as a tumor suppressor in CCA through downregulation of YAP1⁸⁹. The miR-16/YAP1 interaction may not only be a therapeutic target but also act as a reliable prognostic marker for CCA. Additionally, YAP1 has been shown to control the self-renewal and differentiation of MSCs; therefore, miR-16 regulation of YAP1 may influence MSC function during CCA⁹⁰.

It has long been known that miRNAs play a role in the initiation and progression of CCA, but miRNA regulation of MSCs and CSCs during CCA is unknown. While these publications discuss the role of miRNAs in CCA cells and tissues, we can draw comparisons to the impact that they may have on MSC and CSC function. Furthermore, we can note that miRNA signaling may manipulate the microenvironment to become pro-inflammatory, which will favor the initiation of CSC formation as well as the recruitment of MSCs to the injured tissue. However, the discussion of miRNA regulation of MSCs and CSCs during CCA is largely speculative, and further research is necessary to fully understand this signaling process.

Conclusions

Currently, therapeutic options for patients with CCA are extremely limited, a worrying predicament given the increase in incidence. In addition, options for patients with CCA are further limited since the disease tends to be at an advanced stage at the time of diagnosis. These facts indicate the imperative need to develop better diagnostic tools to help identify CCA at an earlier time point as well as more sophisticated therapeutic tools to improve survival rates. The roles of MSCs and CSCs during CCA are a new area of study with

little information known about these cells. For this reason, more studies are needed to fully elucidate the impact that MSCs and CSCs have on CCA development and progression. It is possible that therapies targeting these cells will reduce chemoresistance and recurrence, but only future studies will be able to fully define this. There are a large number of publications regarding miRNA regulation of CCA progression; however, miRNA regulation of MSCs and CSCs during CCA is unknown. Given that miRNA therapies for various liver diseases are currently in clinical trials, there is hope that new miRNA-based therapies are developed for CCA, specifically those that may impact MSC and CSC function. While we have made strides in terms of understanding the CCA tumor environment and the pathways regulating progression, few translational findings have been developed. In the future, translational studies are necessary to help find diagnostic, prognostic, and therapeutic tools for the treatment of CCA.

Abbreviations

CCA, cholangiocarcinoma; CCL5, C-C motif chemokine ligand 5; CCR5, C-C motif chemokine receptor 5; CD, cluster of differentiation; CSC, cancer stem cell; HCC, hepatocellular carcinoma; HPC, hepatic progenitor cell; IFN- γ , interferon-gamma; IL, interleukin; LIF, leukemia inhibitory factor; Ln-332, laminin-332; miRNA, microRNA; MMP, matrix metalloproteinase; MSC,

mesenchymal stem cell; NDRG2, N-myc downstream-regulated gene 2; TAM, tumor-associated macrophage; TET1, ten-eleven translocation 1; TNF- α , tumor necrosis factor-alpha; YAP1, yes-associated protein 1.

Competing interests

The authors declare that they have no competing interests.

Grant information

Portions of this work were supported by (i) a VA Merit Award (1101BX003031) from the US Department of Veterans Affairs, Biomedical Laboratory Research and Development Service and (ii) the National Institute of Health National Institute of Diabetes and Digestive and Kidney Diseases (DK108959). This material is the result of work supported with resources and the use of facilities at the Central Texas Veterans Health Care System (Temple, TX, USA). The content is the responsibility of the authors alone and does not necessarily reflect the views or policies of the Department of Veterans Affairs or the US Government.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

References



- Oliveira IS, Kilcoyne A, Everett JM, *et al.*: **Cholangiocarcinoma: classification, diagnosis, staging, imaging features, and management.** *Abdom Radiol (NY)*. 2017; **42**(6): 1637–49.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Razumilava N, Gores GJ: **Combination of gemcitabine and cisplatin for biliary tract cancer: a platform to build on.** *J Hepatol*. 2011; **54**(3): 577–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Cardinale V, Bragazzi MC, Carpino G, *et al.*: **Cholangiocarcinoma: increasing burden of classifications.** *Hepatobiliary Surg Nutr*. 2013; **2**(5): 272–80.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Banales JM, Cardinale V, Carpino G, *et al.*: **Expert consensus document: Cholangiocarcinoma: current knowledge and future perspectives consensus statement from the European Network for the Study of Cholangiocarcinoma (ENS-CCA).** *Nat Rev Gastroenterol Hepatol*. 2016; **13**(5): 261–80.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Bridgewater J, Galle PR, Khan SA, *et al.*: **Guidelines for the diagnosis and management of intrahepatic cholangiocarcinoma.** *J Hepatol*. 2014; **60**(6): 1268–89.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Guglielmi A, Ruzzenente A, Campagnaro T, *et al.*: **Intrahepatic cholangiocarcinoma: prognostic factors after surgical resection.** *World J Surg*. 2009; **33**(6): 1247–54.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Si A, Li J, Xiang H, *et al.*: **Actual over 10-year survival after liver resection for patients with intrahepatic cholangiocarcinoma.** *Oncotarget*. 2017; **8**(27): 44521–32.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Eishamy M, Presser N, Hammad AY, *et al.*: **Liver transplantation in patients with incidental hepatocellular carcinoma/cholangiocarcinoma and intrahepatic cholangiocarcinoma: a single-center experience.** *Hepatobiliary Pancreat Dis Int*. 2017; **16**(3): 264–70.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- DeOliveira ML, Cunningham SC, Cameron JL, *et al.*: **Cholangiocarcinoma: thirty-one-year experience with 564 patients at a single institution.** *Ann Surg*. 2007; **245**(5): 755–62.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Darwish Murad S, Kim WR, Harnois DM, *et al.*: **Efficacy of neoadjuvant chemoradiation, followed by liver transplantation, for perihilar cholangiocarcinoma at 12 US centers.** *Gastroenterology*. 2012; **143**(1): 88–98.e3; quiz e14.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Devieire J, Baize M, de Toeuf J, *et al.*: **Long-term follow-up of patients with hilar malignant stricture treated by endoscopic internal biliary drainage.** *Gastrointest Endosc*. 1988; **34**(2): 95–101.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Bhat M, Hathcock M, Kremers WK, *et al.*: **Portal vein encasement predicts neoadjuvant therapy response in liver transplantation for perihilar cholangiocarcinoma protocol.** *Transpl Int*. 2015; **28**(12): 1383–91.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- van der Gaag NA, Rauws EA, van Eijck CH, *et al.*: **Preoperative biliary drainage for cancer of the head of the pancreas.** *N Engl J Med*. 2010; **362**(2): 129–37.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Brivio S, Cadamuro M, Strazzabosco M, *et al.*: **Tumor reactive stroma in cholangiocarcinoma: The fuel behind cancer aggressiveness.** *World J Hepatol*. 2017; **9**(9): 455–68.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Pattabiraman DR, Weinberg RA: **Tackling the cancer stem cells - what challenges do they pose?** *Nat Rev Drug Discov*. 2014; **13**(7): 497–512.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Romano M, De Francesco F, Gringeri E, *et al.*: **Tumor Microenvironment Versus Cancer Stem Cells in Cholangiocarcinoma: Synergistic Effects?** *J Cell Physiol*. 2016; **231**(4): 768–76.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Raggi C, Invernizzi P, Andersen JB: **Impact of microenvironment and stem-like plasticity in cholangiocarcinoma: molecular networks and biological concepts.** *J Hepatol*. 2015; **62**(1): 198–207.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Marquardt JU, Raggi C, Andersen JB, *et al.*: **Human hepatic cancer stem cells are characterized by common stemness traits and diverse oncogenic pathways.** *Hepatology*. 2011; **54**(3): 1031–42.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Ma S, Chan KW, Hu L, *et al.*: **Identification and characterization of tumorigenic liver cancer stem/progenitor cells.** *Gastroenterology*. 2007; **132**(7):

- 2542–56.
[PubMed Abstract](#) | [Publisher Full Text](#)
20. Raggi C, Factor VM, Seo D, *et al.*: **Epigenetic reprogramming modulates malignant properties of human liver cancer.** *Hepatology*. 2014; 59(6): 2251–62.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
21. Yamashita T, Forgues M, Wang W, *et al.*: **EpCAM and alpha-fetoprotein expression defines novel prognostic subtypes of hepatocellular carcinoma.** *Cancer Res*. 2008; 68(5): 1451–61.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
22. Raggi C, Correnti M, Sica A, *et al.*: **Cholangiocarcinoma stem-like subset shapes tumor-initiating niche by educating associated macrophages.** *J Hepatol*. 2017; 66(1): 102–15.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
23. Kallias YN, Robson AJ, Fallowfield JA, *et al.*: **Remodelling of extracellular matrix is a requirement for the hepatic progenitor cell response.** *Gut*. 2011; 60(4): 525–33.
[PubMed Abstract](#) | [Publisher Full Text](#)
24. Lorenzini S, Bird TG, Boulter L, *et al.*: **Characterisation of a stereotypical cellular and extracellular adult liver progenitor cell niche in rodents and diseased human liver.** *Gut*. 2010; 59(5): 645–54.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
25. Govaere O, Komuta M, Berkers J, *et al.*: **Keratin 19: a key role player in the invasion of human hepatocellular carcinomas.** *Gut*. 2014; 63(4): 674–85.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
26. Sulpice L, Rayar M, Desille M, *et al.*: **Molecular profiling of stroma identifies osteopontin as an independent predictor of poor prognosis in intrahepatic cholangiocarcinoma.** *Hepatology*. 2013; 58(6): 1992–2000.
[PubMed Abstract](#) | [Publisher Full Text](#)
27. Giannelli G, Fransvea E, Bergamini C, *et al.*: **Laminin-5 chains are expressed differentially in metastatic and nonmetastatic hepatocellular carcinoma.** *Clin Cancer Res*. 2003; 9(10 Pt 1): 3684–91.
[PubMed Abstract](#)
28. Govaere O, Wouters J, Petz M, *et al.*: **Laminin-332 sustains chemoresistance and quiescence as part of the human hepatic cancer stem cell niche.** *J Hepatol*. 2016; 64(3): 609–17.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
29. Karp JM, Leng Teo GS: **Mesenchymal stem cell homing: the devil is in the details.** *Cell Stem Cell*. 2009; 4(3): 206–16.
[PubMed Abstract](#) | [Publisher Full Text](#)
30. Chamberlain G, Fox J, Ashton B, *et al.*: **Concise review: mesenchymal stem cells: their phenotype, differentiation capacity, immunological features, and potential for homing.** *Stem Cells*. 2007; 25(11): 2739–49.
[PubMed Abstract](#) | [Publisher Full Text](#)
31. Spaeth E, Klopp A, Dembinski J, *et al.*: **Inflammation and tumor microenvironments: defining the migratory itinerary of mesenchymal stem cells.** *Gene Ther*. 2008; 15(10): 730–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
32. Yagi H, Soto-Gutierrez A, Parekkadan B, *et al.*: **Mesenchymal stem cells: Mechanisms of immunomodulation and homing.** *Cell Transplant*. 2010; 19(6): 667–79.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
33. Spaeth EL, Dembinski JL, Sasser AK, *et al.*: **Mesenchymal stem cell transition to tumor-associated fibroblasts contributes to fibrovascular network expansion and tumor progression.** *PLoS One*. 2009; 4(4): e4992.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
34. Sawitza I, Kordes C, Götz S, *et al.*: **Bile acids induce hepatic differentiation of mesenchymal stem cells.** *Sci Rep*. 2015; 5: 13320.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
35. Keating A: **Mesenchymal stromal cells: new directions.** *Cell Stem Cell*. 2012; 10(6): 709–16.
[PubMed Abstract](#) | [Publisher Full Text](#)
36. Nauta AJ, Fibbe WE: **Immunomodulatory properties of mesenchymal stromal cells.** *Blood*. 2007; 110(10): 3499–506.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
37. Krampera M, Cosmi L, Angelini R, *et al.*: **Role for interferon-gamma in the immunomodulatory activity of human bone marrow mesenchymal stem cells.** *Stem Cells*. 2006; 24(2): 386–98.
[PubMed Abstract](#) | [Publisher Full Text](#)
38. Zhong W, Tong Y, Li Y, *et al.*: **Mesenchymal stem cells in inflammatory microenvironment potentially promote metastatic growth of cholangiocarcinoma via activating Akt/NF- κ B signaling by paracrine CCL5.** *Oncotarget*. 2017; 8: 73693–73704.
[Publisher Full Text](#) | [F1000 Recommendation](#)
39. Chen DR, Lu DY, Lin HY, *et al.*: **Mesenchymal stem cell-induced doxorubicin resistance in triple negative breast cancer.** *Biomed Res Int*. 2014; 2014: 532161.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
40. Wang W, Zhong W, Yuan J, *et al.*: **Involvement of Wnt/ β -catenin signaling in the mesenchymal stem cells promote metastatic growth and chemoresistance of cholangiocarcinoma.** *Oncotarget*. 2015; 6(39): 42276–89.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
41. Friedman RC, Farh KK, Burge CB, *et al.*: **Most mammalian mRNAs are conserved targets of microRNAs.** *Genome Res*. 2009; 19(1): 92–105.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
42. Pillai RS, Artus CG, Filipowicz W: **Tethering of human Ago proteins to mRNA mimics the miRNA-mediated repression of protein synthesis.** *RNA*. 2004; 10(10): 1518–25.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
43. Bartel DP: **MicroRNAs: target recognition and regulatory functions.** *Cell*. 2009; 136(2): 215–33.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
44. Katsumi T, Ninomiya M, Nishina T, *et al.*: **MIR-139-5p is associated with inflammatory regulation through c-FOS suppression, and contributes to the progression of primary biliary cholangitis.** *Lab Invest*. 2016; 96(11): 1165–77.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
45. Francis H, McDaniel K, Han Y, *et al.*: **Regulation of the extrinsic apoptotic pathway by microRNA-21 in alcoholic liver injury.** *J Biol Chem*. 2014; 289(40): 27526–39.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
46. Kennedy LL, Meng F, Venter JK, *et al.*: **Knockout of microRNA-21 reduces biliary hyperplasia and liver fibrosis in cholestatic bile duct ligated mice.** *Lab Invest*. 2016; 96(12): 1256–67.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
47. Shibata C, Otsuka M, Kishikawa T, *et al.*: **Current status of miRNA-targeting therapeutics and preclinical studies against gastroenterological carcinoma.** *Mol Cell Ther*. 2013; 1: 5.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
48. Baek J, Kang S, Min H: **MicroRNA-targeting therapeutics for hepatitis C.** *Arch Pharm Res*. 2014; 37(3): 299–305.
[PubMed Abstract](#) | [Publisher Full Text](#)
49. Palumbo T, Poultsides GA, Kouraklis G, *et al.*: **A functional microRNA library screen reveals miR-410 as a novel anti-apoptotic regulator of cholangiocarcinoma.** *BMC Cancer*. 2016; 16(1): 353.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
50. Haga H, Yan I, Takahashi K, *et al.*: **Emerging insights into the role of microRNAs in the pathogenesis of cholangiocarcinoma.** *Gene Expr*. 2014; 16(2): 93–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
51. Ehrlich L, Hall C, Venter J, *et al.*: **miR-24 Inhibition Increases Menin Expression and Decreases Cholangiocarcinoma Proliferation.** *Am J Pathol*. 2017; 187(3): 570–80.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
52. Du K, Li Z, Fang X, *et al.*: **Ferulic acid promotes osteogenesis of bone marrow-derived mesenchymal stem cells by inhibiting microRNA-340 to induce β -catenin expression through hypoxia.** *Eur J Cell Biol*. 2017; 96(6): 496–503.
[PubMed Abstract](#) | [Publisher Full Text](#)
53. Pakravan K, Babashah S, Sadeghizadeh M, *et al.*: **MicroRNA-100 shuttled by mesenchymal stem cell-derived exosomes suppresses *in vitro* angiogenesis through modulating the mTOR/HIF-1 α /VEGF signaling axis in breast cancer cells.** *Cell Oncol (Dordr)*. 2017; 40(5): 457–470.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
54. Yu M, Xue Y, Zheng J, *et al.*: **Linc00152 promotes malignant progression of glioma stem cells by regulating miR-103a-3p/FEZF1/CDC25A pathway.** *Mol Cancer*. 2017; 16(1): 110.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
55. Chandrasekharappa SC, Guru SC, Manickam P, *et al.*: **Positional cloning of the gene for multiple endocrine neoplasia-type 1.** *Science*. 1997; 276(5311): 404–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
56. Vijayaraghavan J, Maggi EC, Crabtree JS: **miR-24 regulates menin in the endocrine pancreas.** *Am J Physiol Endocrinol Metab*. 2014; 307(1): E84–92.
[PubMed Abstract](#) | [Publisher Full Text](#)
57. Meng FL, Wang W, Jia WD: **Diagnostic and prognostic significance of serum miR-24-3p in HBV-related hepatocellular carcinoma.** *Med Oncol*. 2014; 31(9): 177.
[PubMed Abstract](#) | [Publisher Full Text](#)
58. Liu YX, Long XD, Xi ZF, *et al.*: **MicroRNA-24 modulates aflatoxin B1-related hepatocellular carcinoma prognosis and tumorigenesis.** *Biomed Res Int*. 2014; 2014: 482926.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
59. Dong W, Li B, Wang Z, *et al.*: **Clinical significance of microRNA-24 expression in esophageal squamous cell carcinoma.** *Neoplasma*. 2015; 62(2): 250–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
60. Lee SH, Chen TY, Dhar SS, *et al.*: **A feedback loop comprising PRMT7 and miR-24-2 interplays with Oct4, Nanog, Klf4 and c-Myc to regulate stemness.** *Nucleic Acids Res*. 2016; 44(22): 10603–18.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
61. Balsalobre A, Damiola F, Schibler U: **A serum shock induces circadian gene expression in mammalian tissue culture cells.** *Cell*. 1998; 93(6): 929–37.
[PubMed Abstract](#) | [Publisher Full Text](#)
62. Filipski E, King VM, Li X, *et al.*: **Host circadian clock as a control point in tumor progression.** *J Natl Cancer Inst*. 2002; 94(9): 690–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
63. Viswanathan AN, Scherhammer ES: **Circulating melatonin and the risk of**

- breast and endometrial cancer in women. *Cancer Lett.* 2009; **281**(1): 1–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
64. Han Y, Meng F, Venter J, *et al.*: miR-34a-dependent overexpression of *Per1* decreases cholangiocarcinoma growth. *J Hepatol.* 2016; **64**(6): 1295–304.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
65. **F** Boucher H, Vanneaux V, Domet T, *et al.*: Circadian Clock Genes Modulate Human Bone Marrow Mesenchymal Stem Cell Differentiation, Migration and Cell Cycle. *PLoS One.* 2016; **11**(1): e0146674.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
66. Dawson MA, Kouzarides T: Cancer epigenetics: from mechanism to therapy. *Cell.* 2012; **150**(1): 12–27.
[PubMed Abstract](#) | [Publisher Full Text](#)
67. You JS, Jones PA: Cancer genetics and epigenetics: two sides of the same coin? *Cancer Cell.* 2012; **22**(1): 9–20.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
68. Sandhu DS, Shire AM, Roberts LR: Epigenetic DNA hypermethylation in cholangiocarcinoma: potential roles in pathogenesis, diagnosis and identification of treatment targets. *Liver Int.* 2008; **28**(1): 12–27.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
69. **F** Li H, Zhou ZQ, Yang ZR, *et al.*: MicroRNA-191 acts as a tumor promoter by modulating the TET1-p53 pathway in intrahepatic cholangiocarcinoma. *Hepatology.* 2017; **66**(1): 136–51.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
70. **F** Xu W, Ji J, Xu Y, *et al.*: MicroRNA-191, by promoting the EMT and increasing CSC-like properties, is involved in neoplastic and metastatic properties of transformed human bronchial epithelial cells. *Mol Carcinog.* 2015; **54** Suppl 1: E148–61.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
71. Lee DC, Kang YK, Kim WH, *et al.*: Functional and clinical evidence for *NDRG2* as a candidate suppressor of liver cancer metastasis. *Cancer Res.* 2008; **68**(11): 4210–20.
[PubMed Abstract](#) | [Publisher Full Text](#)
72. Tepel M, Roerig P, Wolter M, *et al.*: Frequent promoter hypermethylation and transcriptional downregulation of the *NDRG2* gene at 14q11.2 in primary glioblastoma. *Int J Cancer.* 2008; **123**(9): 2080–6.
[PubMed Abstract](#) | [Publisher Full Text](#)
73. Wang J, Yin D, Xie C, *et al.*: The iron chelator Dp44mT inhibits hepatocellular carcinoma metastasis via N-Myc downstream-regulated gene 2 (*NDRG2*)/gp130/STAT3 pathway. *Oncotarget.* 2014; **5**(18): 8478–91.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
74. Tong SJ, Liu J, Wang X, *et al.*: microRNA-181 promotes prostate cancer cell proliferation by regulating *DAX-1* expression. *Exp Ther Med.* 2014; **8**(4): 1296–300.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
75. Swanson JM, Wigal S, Greenhill LL, *et al.*: Analog classroom assessment of Adderall in children with ADHD. *J Am Acad Child Adolesc Psychiatry.* 1998; **37**(5): 519–26.
[PubMed Abstract](#) | [Publisher Full Text](#)
76. Mansueto G, Forzati F, Ferraro A, *et al.*: Identification of a New Pathway for Tumor Progression: MicroRNA-181b Up-Regulation and CBX7 Down-Regulation by HMGA1 Protein. *Genes Cancer.* 2010; **1**(3): 210–24.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
77. **F** Huang P, Ye B, Yang Y, *et al.*: MicroRNA-181 functions as a tumor suppressor in non-small cell lung cancer (NSCLC) by targeting *Bcl-2*. *Tumour Biol.* 2015; **36**(5): 3381–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
78. Conti A, Aguenouz M, La Torre D, *et al.*: miR-21 and 221 upregulation and miR-181b downregulation in human grade II-IV astrocytic tumors. *J Neurooncol.* 2009; **93**(3): 325–32.
[PubMed Abstract](#) | [Publisher Full Text](#)
79. Pichler M, Winter E, Ress AL, *et al.*: miR-181a is associated with poor clinical outcome in patients with colorectal cancer treated with EGFR inhibitor. *J Clin Pathol.* 2014; **67**(3): 198–203.
[PubMed Abstract](#) | [Publisher Full Text](#)
80. **F** Wang J, Xie C, Pan S, *et al.*: N-myc downstream-regulated gene 2 inhibits human cholangiocarcinoma progression and is regulated by leukemia inhibitory factor/MicroRNA-181c negative feedback pathway. *Hepatology.* 2016; **64**(5): 1606–22.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
81. **F** Chatterjee A, Chattopadhyay D, Chakrabarti G: MiR-16 targets *Bcl-2* in paclitaxel-resistant lung cancer cells and overexpression of miR-16 along with miR-17 causes unprecedented sensitivity by simultaneously modulating autophagy and apoptosis. *Cell Signal.* 2015; **27**(2): 189–203.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
82. **F** Tang X, Jin L, Cao P, *et al.*: MicroRNA-16 sensitizes breast cancer cells to paclitaxel through suppression of *IKBKB* expression. *Oncotarget.* 2016; **7**(17): 23668–83.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
83. Chen L, Wang Q, Wang GD, *et al.*: miR-16 inhibits cell proliferation by targeting *IGF1R* and the *Raf1-MEK1/2-ERK1/2* pathway in osteosarcoma. *FEBS Lett.* 2013; **587**(9): 1366–72.
[PubMed Abstract](#) | [Publisher Full Text](#)
84. Perra A, Kowalik MA, Ghiso E, *et al.*: YAP activation is an early event and a potential therapeutic target in liver cancer development. *J Hepatol.* 2014; **61**(5): 1088–96.
[PubMed Abstract](#) | [Publisher Full Text](#)
85. **F** Lee K, Lee SS, Kim SB, *et al.*: Significant association of oncogene YAP1 with poor prognosis and cetuximab resistance in colorectal cancer patients. *Clin Cancer Res.* 2015; **21**(2): 357–64.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
86. Xia Y, Chang T, Wang Y, *et al.*: YAP promotes ovarian cancer cell tumorigenesis and is indicative of a poor prognosis for ovarian cancer patients. *PLoS One.* 2014; **9**(3): e91770.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
87. **F** Sun D, Li X, He Y, *et al.*: YAP1 enhances cell proliferation, migration, and invasion of gastric cancer *in vitro* and *in vivo*. *Oncotarget.* 2016; **7**(49): 81062–76.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
88. Wu C, Xu B, Yuan P, *et al.*: Genome-wide interrogation identifies YAP1 variants associated with survival of small-cell lung cancer patients. *Cancer Res.* 2010; **70**(23): 9721–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
89. Han S, Wang D, Tang G, *et al.*: Suppression of miR-16 promotes tumor growth and metastasis through reversely regulating YAP1 in human cholangiocarcinoma. *Oncotarget.* 2017; **8**(34): 56635–56650.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
90. **F** Tang Y, Weiss SJ: Snail/Slug-YAP/TAZ complexes cooperatively regulate mesenchymal stem cell function and bone formation. *Cell Cycle.* 2017; **16**(5): 399–405.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)

Open Peer Review

Current Referee Status:  

Editorial Note on the Review Process

F1000 Faculty Reviews are commissioned from members of the prestigious F1000 Faculty and are edited as a service to readers. In order to make these reviews as comprehensive and accessible as possible, the referees provide input before publication and only the final, revised version is published. The referees who approved the final version are listed with their names and affiliations but without their reports on earlier versions (any comments will already have been addressed in the published version).

The referees who approved this article are:

Version 1

- 1 **Jesper B Anderson** University of Copenhagen, Copenhagen, Denmark
Competing Interests: No competing interests were disclosed.
- 1 **Li Wang** University of Connecticut, Storrs, Connecticut, USA
Competing Interests: No competing interests were disclosed.