Combined aliskiren and L-arginine treatment has antihypertensive effects and prevents vascular endothelial dysfunction in a model of renovascular hypertension

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Abstract

Angiotensin II is a key player in the pathogenesis of renovascular hypertension, a condition associated with endothelial dysfunction. We investigated aliskiren (ALSK) and L-arginine treatment both alone and in combination on blood pressure (BP), and vascular reactivity in aortic rings. Hypertension was induced in 40 male Wistar rats by clipping the left renal artery. Animals were divided into Sham, 2-kidney, 1-clip (2K1C) hypertension, 2K1C + ALSK (ALSK), 2K1C + L-arginine (L-arg), and 2K1C + ALSK + L-arginine (ALSK + L-arg) treatment groups. For 4 weeks, BP was monitored and endothelium-dependent and independent vasoconstriction and relaxation were assessed in aortic rings. ALSK + L-arg reduced BP and the contractile response to phenylephrine and improved acetylcholine relaxation. Endothelium removal and incubation with N-nitro-L-arginine methyl ester (L-NAME) increased the response to phenylephrine in all groups, but the effect was greater in the ALSK + L-arg group. Losartan reduced the contractile response in all groups, apocynin reduced the contractile response in the 2K1C, ALSK and ALSK + L-arg groups, and incubation with superoxide dismutase reduced the phenylephrine response in the 2K1C and ALSK groups. eNOS expression increased in the 2K1C and L-arg groups, and iNOS was increased significantly only in the 2K1C group compared with other groups. AT_1 expression increased in the 2K1C compared with the Sham, ALSK and ALSK + L-arg groups, AT_2 expression increased in the ALSK + L-arg group compared with the Sham and L-arg groups, and gp91phox decreased in the ALSK + L-arg group compared with the 2K1C and ALSK groups. In conclusion, combined ALSK + L-arg was effective in reducing BP and preventing endothelial dysfunction in aortic rings of 2K1C hypertensive rats. The responsible mechanisms appear to be related to the modulation of the local renin-angiotensin system, which is associated with a reduction in endothelial oxidative stress.

Key words: 2K1C hypertension; endothelial dysfunction; aliskiren; L-arginine; RAAS and oxidative stress

Introduction

Hypertension is manifested not only by increased arterial pressure but also by complex structural and functional alterations of its target organs. Long-term hypertension often results in left ventricular hypertrophy, which is considered a risk factor for coronary heart disease (1), and also causes structural alterations of the vascular wall characterized by endothelial dysfunction, extracellular matrix deposition, medial layer thickening due to hypertrophy/hyperplasia, and migration of vascular smooth muscle cells (VSMCs) (2). Chronic kidney artery diseases, such as renal artery stenosis, generally lead to hypertension, and a kidney-related animal model of hypertension, the 2-kidney, 1-clip (2K1C) model, is produced by subjecting a renal artery to partial stenosis by clip placement. Kidney ischemia results in an increase

of plasma renin activity and the consequent increase in angiotensinogen concentration leads to a persistent rise in blood pressure (2,3). This hypertension model is associated with increased angiotensin II levels, and this peptide produces mitogenic effects, which are critically involved in the development of the structural and functional vascular changes caused by hypertension (4). In experimental 2K1C hypertension, the overproduction of reactive oxygen species (ROS), which leads to oxidative stress, plays an important role in the pathogenesis of renovascular hypertension and enhanced oxidation-sensitive signaling pathway activation (5). Previous studies have reported that angiotensin II stimulates the production of ROS such as superoxide through the activation of membrane-bound nicotinamide adenine dinucleotide

(NADH) or nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (6).

Endothelial dysfunction has an important role in the pathogenesis and progression of hypertensive heart disease (7). Increased oxidative stress impairs endothelial function and is one of the primary mediators of the development of hypertension, atherosclerosis, diabetes, cardiac hypertrophy, heart failure, ischemia-reperfusion injury, and stroke (8).

Drugs that target the renin-angiotensin-aldosterone system (RAAS), such as angiotensin-converting enzyme (ACE) inhibitors and blockers of angiotensin receptor-1 (AT_1), are effective in reducing blood pressure and morbidity and mortality. Their low rate of side effects makes them well tolerated and therefore attractive as first-line agents for the treatment of arterial hypertension (9). Aliskiren (ALSK), a recent addition to the family of RAAS-blockers, is a direct renin inhibitor indicated for the treatment of hypertension. Several studies have previously investigated the effectiveness of ALSK both as mono-therapy and in combination with other agents in lowering blood pressure (10). Some studies have evaluated ALSK administered once a day to reduce blood pressure compared with ramipril (11), losartan (12), irbesartan (13), and hydrochlorothiazide (14). In those studies, which included patients with mild-to-moderate essential hypertension, ALSK led to a decrease in blood pressure similar to the other agents or drugs. However, whether ALSK reduces persistent hypertension, such as that produced in 2K1C models, has not been demonstrated.

Our previous results demonstrated that treatment with L-arginine, a substrate for nitric oxide (NO) production, reduces blood pressure in the 2K1C hypertension model, not only because of its known effects on NO formation and vasodilation but also because of increased renal excretion of water and sodium (15). Recently, L-arginine supplementation in patients with mild arterial hypertension was shown to stimulate NO biosynthesis and reduce oxidative stress (16). Gokce (17) reported that the L-arginine-mediated mechanisms of reduction in arterial hypertension include improvement of vasomotor functions of the endothelium, increased synthesis of NO in vessels, decreased activity of endothelin-1 and angiotensin II, modulation of hemodynamic changes in kidneys, lowering of oxidative stress, and improved insulin sensitivity.

This study investigated the effects of ALSK, L-arginine and the combination of ALSK and L-arginine on blood pressure and vascular reactivity in aortic rings in a renovascular 2K1C hypertension model, with a focus on the renin-angiotensin system and the involvement of oxidative stress in renovascular hypertension-induced endothelial dysfunction.

Material and Methods

Animals and treatment

Male Wistar rats (150-170 g, n = 8 per group) were used in these experimental procedures. The care and use of laboratory animals were in accordance with the NIH guidelines. All experiments were conducted in compliance with the Guidelines for Biomedical Research as stated by the Brazilian Societies of Experimental Biology and were approved by the Institutional Ethics Committee of the Universidade Federal do Espírito Santo (CEUA-UFES 004/2010). All rats had free access to water and were fed rat chow *ad libitum*. Rats were divided into five groups: Sham (normotensive control, 0.1 mL saline vehicle by gavage); 2K1C (hypertension control, untreated); 2K1C treated with ALSK (50 mg/kg, 0.3 mL/day by gavage); 2K1C treated with L-arginine (10 mg/kg, 0.1 mL/day L-arg by gavage), and 2K1C treated with ALSK + L-arginine (50 mg/kg ALSK, 0.3 mL/day + 10 mg/kg L-arg, 0.1 mL/day, both by gavage). At the end of treatment, rats were anesthetized by intraperioneal (*ip*) injection of pentobarbital (35 mg/kg) and killed by exsanguination. The thoracic aorta was carefully dissected and connective tissue removed. For vascular reactivity experiments, the aortas were divided into cylindrical segments 4 mm in length. For analysis of protein expression, some arteries were rapidly frozen in liquid nitrogen and stored at –80°C until analyzed.

Renovascular hypertensive model

Renovascular hypertension was induced by the Goldblatt 2K1C method as described in our previous reports (15,18). To minimize stress-induced fluctuation of systolic blood pressure (SBP), rats were trained by measuring SBP daily for at least 7 days before the 2K1C procedure or the sham operation. Then, a retro-peritoneal flank incision was performed in the rats anesthetized with sodium pentobarbital (35 mg/kg, *ip*). The left renal artery was exposed via midline laparotomy. Renovascular hypertension was induced by partial occlusion of the artery by a U-shaped silver clip with an internal diameter of 0.20 mm. Sham rats (normotensive sham operated) underwent a similar surgical procedure but without clip placement. The criterion for hypertension in the present study was an SBP>160 mmHg, and only hypertensive 2K1C rats with SBP>160 mmHg were used in the experimental procedures.

Blood pressure measurements

Indirect SBP was measured by tail-cuff plethysmography (IITC Life Science, Inc., USA). Conscious rats were restrained for 5-10 min in a warm, quiet room and conditioned to numerous cuff inflation-deflation cycles by a trained operator. SBP was measured before surgery (time 0) and a week after surgery to confirm that the procedure had been successful and resulted in hypertensive animals (time 7), and at the end of the treatment, 28 days after surgery (time 28). Blood pressure was measured 3 times on all 3 days and the mean of the 3 measurements was recorded for each time.

Vascular reactivity measurements

Aortic segments 4 mm in length were mounted between two parallel wires in a 37°C organ bath containing Krebs-Henseleit solution (KHS; 124 mM NaCl, 4.6 mM KCl, 2.5 mM $CaCl_2$, 1.2 mM $MgSO_4$, 1.2 mM KH_2PO_4, 0.01 mM EDTA, 23 mM $NaHCO_3$, 11 mM glucose) and gassed with 95% O_2-5% CO_2, pH 7.4. Arterial segments were stretched to an optimal resting tension of 1.0 g. Isometric tension was recorded using a force displacement transducer (TSD125C, Biopac Systems, USA) connected to an acquisition system (MP100A, Biopac Systems).

After a 45-min equilibration period, all aortic rings were exposed twice to 75 mM KCl. The first exposure was to determine their functional integrity, and the second exposure was to assess the maximal tension that they could be exposed to. Next, the endothelial integrity was tested with acetylcholine (ACh, 10 μM) in segments previously contracted with phenylephrine (1 μM). After a 45-min washout period, concentration-response curves to phenylephrine (10^{-10} to 3×10^{-4} M) were determined. Single curves were obtained for each segment.

In all experimental groups, the influence of the endothelium on the response of aortic segments to phenylephrine was investigated after mechanical removal of the endothelium by rubbing the lumen of the segment with a needle. The absence of endothelium was confirmed by the inability of 10 μM ACh to produce relaxation.

The role of endothelial-derived vasoactive factors on the phenylephrine-elicited contractile response was investigated. The effects of the following drugs were evaluated: *1)* the nonspecific nitric oxide synthase (NOS) inhibitor N-nitro-L-arginine methyl ester (L-NAME, 100 μM), *2)* an AT_1 antagonist (losartan, 10 μM), *3)*, an NADPH oxidase inhibitor (apocynin, 0.3 mM), and *4)* superoxide dismutase (SOD) (150 U/mL). These drugs were added to the bath 30 min before generating the phenylephrine concentration-response curves.

In another set of experiments conducted after the 45-min equilibration period, the aortic rings from all of the experimental groups were precontracted with phenylephrine (1 μM) until they reached a plateau (approximately 15 min), and concentration-response curves to ACh (10^{-10} to 3×10^{-4} M) or sodium nitroprusside (SNP: 10^{-10} to 3×10^{-4} M) were determined.

Western blot analysis

Aortas were homogenized in lysis buffer containing 150 mM NaCl, 50 mM Tris-HCl, 5 mM EDTA.2Na, 1 mM $MgCl_2$ plus protease inhibitor (Sigma Fast; Sigma, USA). The protein concentration was determined by the Lowry method (19), and bovine serum albumin (BSA) was used as a standard. Equal amounts of protein (50 μg) were separated by 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). Proteins were transferred to polyvinylidene difluoride (PVDF) membranes that were incubated with mouse monoclonal antibodies against endothelial nitric oxide synthase (eNOS, 1:1500; BD, USA), inducible nitric oxide synthase (iNOS, 1:1500; BD), gp91phox (1:1000; BD) and rabbit polyclonal antibodies for AT_1 (1:500; Santa Cruz Biotechnology, USA) and AT_2 (1:1000; Millipore, USA). After washing, the membranes were incubated with alkaline phosphatase conjugated anti-mouse IgG (1:3000, Abcam Inc., USA) or anti-rabbit (1:7000; Santa Cruz Biotechnology) antibodies. The protein bands were visualized using a nitro-blue tetrazolium/5-bromo-4-chloro-3′-indolyphosphate (NBT/BCIP) staining system (Invitrogen Corporation, USA) and quantified using the Image J software (National Institutes of Health, USA). The same membranes were used to assay β-actin expression using a mouse monoclonal antibody to β-actin (1:5000; Sigma Chemical, Co., USA), and the results are reported as the ratio of the densities of specific bands to the corresponding β-actin.

Drugs and reagents

Rasilez® (Aliskiren; Novartis, Italy), l-phenylephrine hydrochloride, L-NAME, apocynin, SOD, acetylcholine chloride, sodium pentobarbital, losartan, superoxide dismutase, sodium nitroprusside and L-arginine monohydrochloride were purchased from Sigma-Aldrich (USA). The salts and reagents used were of analytical grade and purchased from Sigma-Aldrich and Merck (Germany).

Statistical analyses

Data are reported as means ± SE. Contractile responses are reported as a percentage of the maximal response induced by 75 mM KCl. Relaxation responses to ACh or SNP are reported as the percentage of relaxation of the previous contraction. For each concentration-response curve, the maximal effect (R_{max}) and the concentration of agonist that produced 50% of the maximal response (log EC_{50}) were calculated using nonlinear regression analysis. The sensitivities of the agonists are reported as pD_2 (–log EC_{50}).

To compare the effects of endothelium denudation, L-NAME, losartan, and apocynin on the contractile responses to phenylephrine, some of the results are reported as differences in the area under the concentration-response curve (dAUC) for the control (E+) and each experimental group (E–, L-NAME, losartan, SOD and apocynin). These data indicated whether the size of the effect of endothelial denudation, L-NAME, losartan, SOD, and apocynin was significantly different in sham-treated segments and segments in the 2K1C, ALSK, L-arg and ALSK + L-arg groups. The means were compared using one-way and two-way ANOVA, followed by Tukey's *post hoc* test when appropriate.

For protein expression, data are reported as the ratio of the immunoblot densities corresponding to the protein of interest and β-actin. The means were analyzed using one-way ANOVA followed by Fisher's *post hoc* test. For
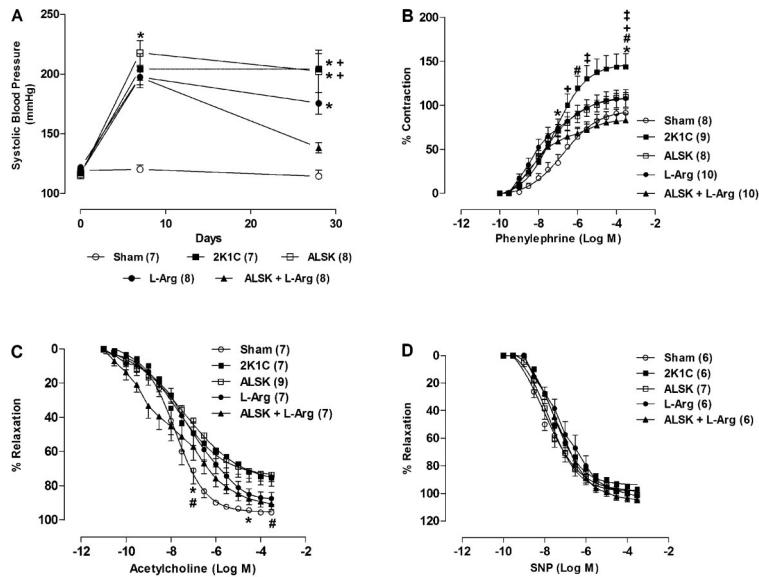
Figure 1. Effects of aliskiren (ALSK), L-arginine (L-arg) and a combination of both on systolic blood pressure throughout the experiment (*A*). Effects of ALSK and L-arg treatment in renovascular hypertension on the concentration-response curves to phenylephrine (*B*), acetylcholine (*C*) and sodium nitroprusside (SNP) (*D*) in the aortic rings. Data are reported as means ± SE. The number of animals in each group is indicated in parentheses. *P<0.05 *vs* Sham; #P<0.05 *vs* ALSK; ‡P<0.05 *vs* L-arg; +P<0.05 *vs* ALSK+L-arg (two-way ANOVA, followed by Tukey's *post hoc* test).

all analyses, the differences were considered significant at P<0.05.

Results

Effect of ALSK and L-arginine treatment on SBP

The baseline SBP (time 0) was similar in the 5 experimental groups before surgery (Sham: 112.2 ± 1.01 mmHg, n=7; 2K1C: 120.4 ± 2.11 mmHg, n=7; ALSK: 124.6 ± 1.20 mmHg, n=8; L-arg: 115.6 ± 3.3 mmHg, n=8, and ALSK+L-arg: 118.8 ± 2.70 mmHg, n=8), and no significant change in SBP was seen in the Sham group at the end of treatment (114.4 ± 5.2 mmHg, n=7). Surgical renal stenosis was associated with a significant increase in SBP compared with the sham operation, and was detectable as early as 7

Table 1. Parameters of maximal response (R_{max}) and sensitivity (pD_2) of the concentration-response curves to phenylephrine in the aortas from all experimental groups, before (E+) and after endothelial denudation (E–) and after incubation with L-NAME (100 μM), losartan (10 μM), apocynin (0.3 nM) and SOD (150 U/mL).

	Control (E+)	E–	L-NAME	Losartan	Apocynin	SOD
Sham						
R_{max}	92.4 ± 4.4[§]	130.3 ± 3.66[⊥]	133.5 ± 10.6[⊥]	93.3 ± 6.8	91.7 ± 3.6	87.4 ± 6.8
pD_2	6.77 ± 0.35	7.93 ± 0.20[⊥]	8.45 ± 0.30[⊥]	7.55 ± 0.28	7.83 ± 0.21	7.3 ± 0.21
2K1C						
R_{max}	148.1 ± 15.6	166.4 ± 7.59	163.1 ± 8.7	86.9 ± 8.6[⊥]	42.7 ± 4.32*[⊥]	62.9 ± 7.1[⊥]
pD_2	7.25 ± 0.14	8.90 ± 0.32[⊥]	7.94 ± 0.12[⊥]	7.93 ± 0.32	7.70 ± 0.17	7.8 ± 0.25[⊥]
ALSK						
R_{max}	112.3 ± 7.4	136.7 ± 12.9	218.5 ± 40.7[⊥]	78.8 ± 6.4[⊥]	60.7 ± 16.8[⊥]	72.9 ± 8.3[⊥]
pD_2	7.74 ± 0.21	8.23 ± 0.20	7.84 ± 0.28	8.47 ± 0.15	10.2 ± 1.85	7.7 ± 0.31
L-arg						
R_{max}	106.6 ± 8.8[§]	161.8 ± 10.5[⊥]	158.1 ± 9.1[⊥]	44.7 ± 9.0*[§⊥]	85.2 ± 17.1	72.2 ± 14.1
pD_2	8.20 ± 0.24*	8.60 ± 0.20	8.06 ± 0.26	10.9 ± 2.87	14.4 ± 3.05	8.3 ± 0.34
ALSK + L-arg						
R_{max}	84.39 ± 7.6[§]	162.4 ± 13.9[⊥]	187.8 ± 19.1[⊥]	52.8 ± 6.3[§⊥]	35.7 ± 5.0*[‡⊥]	67.3 ± 4.7
pD_2	8.28 ± 0.37[§]	8.13 ± 0.43	8.37 ± 0.15	7.7 ± 0.33	10.5 ± 1.60	8.5 ± 0.54

Data are reported as means ± SE. R_{max}: maximal effect (reported as a percentage of the maximal response induced by 75 mM KCl); pD2: –log one-half R_{max}; E–: endothelium removal; L-NAME: N^G-nitro-L-arginine methyl ester; SOD: superoxide dismutase. *P<0.05 *vs* Sham; §P<0.05 *vs* 2K1C; ‡P<0.05 *vs* L-arg; and ⊥P<0.05 *vs* control E+ (two-way ANOVA, followed by Tukey's *post hoc* test).
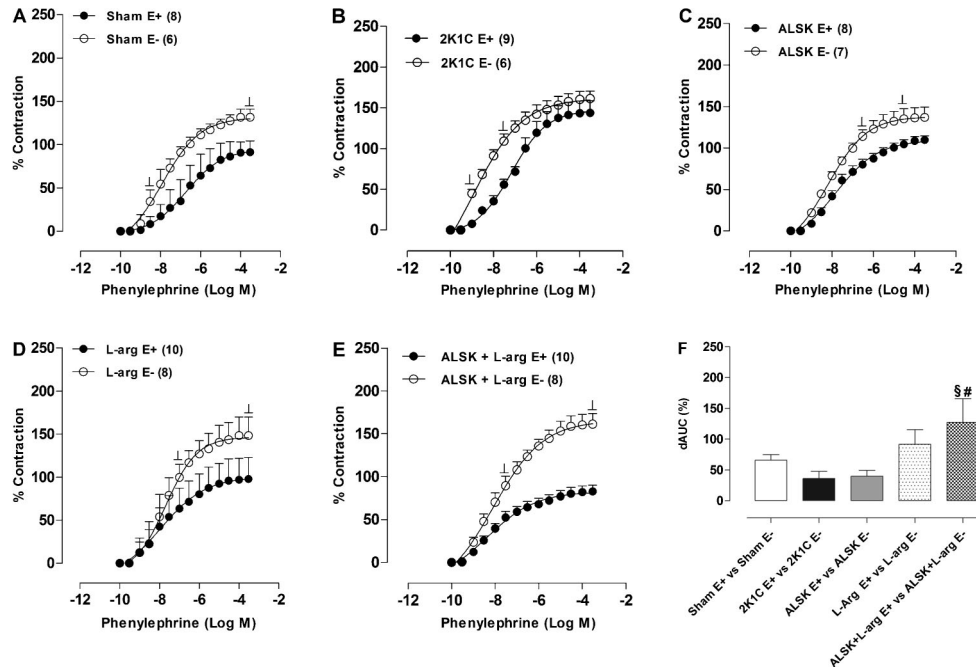
Figure 2. Effects of endothelium removal (E–) on the concentration-response curve for phenylephrine in the aortic rings from Sham (*A*), 2K1C (*B*), aliskiren (ALSK) (*C*), L-arginine (L-arg) (*D*) and ALSK + L-arg (*E*) treatment in the aortic rings with (E +) and without (E–) endothelium. The differences in the area under the concentration-response curves (dAUC) in endothelium-denuded and intact segments is shown in *F*. Data are reported as means ± SE. The number of animals in each group is indicated in parentheses. #P<0.05 *vs* ALSK; §P<0.05 *vs* 2K1C and ⊥P<0.05 *vs* E + (two-way ANOVA, followed by Tukey's *post hoc* test).

days after surgery (2K1C: 204 ± 12.7 mmHg, n = 7; ALSK: 217.8 ± 10.2 mmHg, n = 7; L-arg: 197.5 ± 8.9 mmHg, n = 8; ALSK + L-arg: 197.1 ± 6.08 mmHg, n = 8 *vs* Sham: 119.2 ± 2.51). After 21 days of treatment, only the combined administration of ALSK + L-arg (138.4 ± 4.37 mmHg, n = 8) was effective in reducing SBP (P<0.05) compared to 2K1C (204 ± 12.7 mmHg, n = 6). Additionally, the ALSK (202.4 ± 17.7 mmHg, n = 7) and L-arg (175.6 ± 9.14 mmHg, n = 7) groups maintained high SBP compared with the Sham group (114.4 ± 5.2 mmHg, n = 7; Figure 1A).

Effects of ALSK and L-arginine treatment on vascular reactivity

None of the treatments affected the response to KCl (Sham E +: 2.85 ± 0.17 g, n = 8; 2K1C E +: 2.73 ± 0.27 g, n = 9; ALSK E +: 2.78 ± 0.12 g, n = 8; L-arg E +: 2.40 ± 0.15 g, n = 10; ALSK + L-arg E +: 2.41 ± 0.13 g, n = 10; and Sham E–: 2.88 ± 0.11 g, n = 7; 2K1C E–: 2.87 ± 0.32 g, n = 8; ALSK E–: 2.38 ± 0.18 g, n = 8; L-arg E–: 2.75 ± 0.32 g, n = 8; ALSK + L-arg E–: 2.42 ± 0.21 g, n = 8; P>0.05). Renovascular hypertension (2K1C group) increased the contractile responses induced by phenylephrine in rat aortas (Figure 1B). It also increased R_{max} compared with the Sham, L-arg and ALSK + L-arg groups, but not the sensitivity to phenylephrine (Table 1).

The concentration-dependent relaxation induced by ACh showed impairment at some concentrations in the 2K1C and ALSK groups compared with the Sham group (Figure 1C), but no differences were seen in R_{max} and sensitivity to phenylephrine (Table 1). The response induced by SNP did not change in any of the groups (Figure 1D).

Effects of ALSK and L-arginine treatment on the endothelial modulation of vasoconstrictor responses

To evaluate the influence of endothelium on phenylephrine-induced contraction, we mechanically removed that layer. The reactivity increased, but the responses were smaller in the 2K1C group and in the ALSK group (Figure 2). This difference was clearly seen when dAUC was compared (2K1C: 36.3 ± 11.5; ALSK: 39.8 ± 9.5 *vs* ALSK + L-arg: 127.3 ± 38.3, P<0.05; Figure 2F). Similarly, R_{max} was increased in the Sham, L-arg and ALSK + L-arg groups compared with the control (E +), and the sensitivity to phenylephrine was altered in both the Sham and 2K1C groups (Table 1).

L-NAME (100 μM) was used to investigate the putative role of NO in the effects of ALSK and L-arginine treatment on the contractile response induced by phenylephrine. The concentration-response curve for phenylephrine was left-shifted in the aortic segments from all
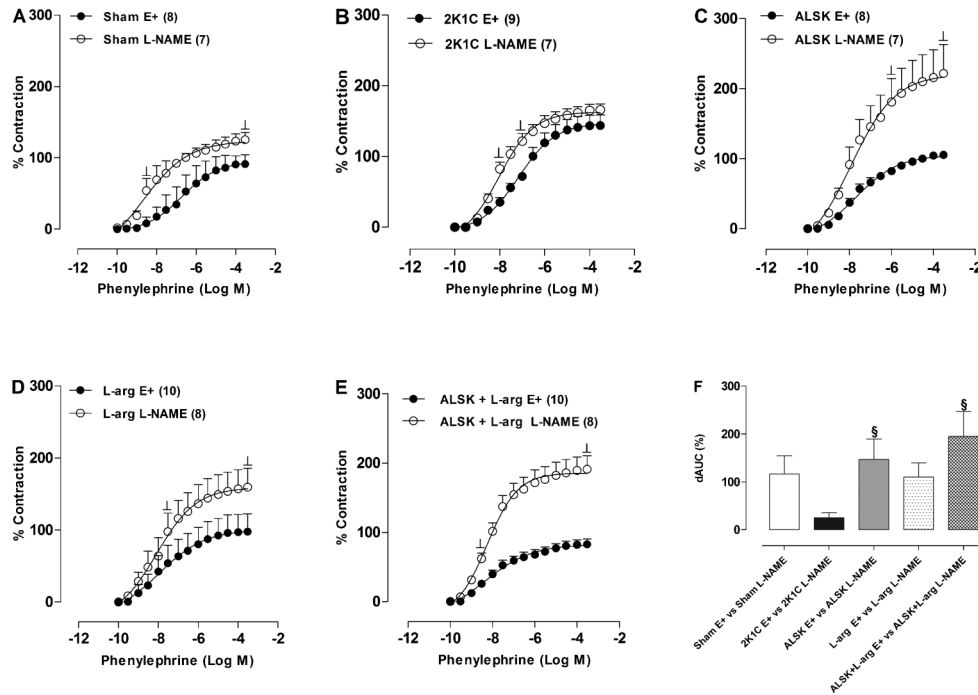
Figure 3. Effects of N^G-nitro-L-arginine methyl ester blocker (L-NAME, 100 μM) on the concentration-response curve for phenylephrine in the aortic rings from Sham (*A*), 2K1C (*B*), aliskiren (ALSK) (*C*), L-arginine (L-arg) (*D*) and ALSK + L-arg (*E*) groups in aortic rings in the presence (L-NAME) and absence (E+) of L-NAME blocker. The differences in the area under the concentration-response curves (dAUC) in the presence and absence of L-NAME is shown in *F*. Data are reported as means ± SE. The number of animals in each group is indicated in parentheses. §P<0.05 *vs* 2K1C and $^\perp$P<0.05 *vs* E+ (two-way ANOVA, followed by Tukey's *post hoc* test).

groups (Figure 3A-E). However, this effect was smaller in the ring preparations from the 2K1C group than from the ALSK and ALSK + L-arg treatment groups, as indicated by the dAUC values (2K1C: 25.2 ± 10.5 *vs* ALSK: 147.1 ± 42.2 and ALSK + L-arg: 195 ± 51.7; Figure 3F). The R$_{max}$ was increased in the Sham, ALSK, L-arg and ALSK + L-arg groups compared to the controls (E+), and the sensitivity to phenylephrine was increased in the Sham and 2K1C groups (Table 1).

These results indicated that renovascular hypertension induces endothelial dysfunction in the conductance arteries, thereby reducing endothelial NO modulation of the vasoconstrictor responses. The protein expression of

eNOS (Figure 4A) increased in the 2K1C hypertension and L-arg groups; treatment with either ALSK or ALSK + L-arg reduced eNOS protein expression in the aorta (Figure 4A). In addition, the protein expression of iNOS (Figure 4B) increased significantly in the 2K1C group compared to the Sham, ALSK, L-arg and ALSK + L-arg groups (Figure 4B).

Role of the RAAS in the effects of ALSK and L-arginine treatment on the phenylephrine response

To investigate whether the local RAAS was involved in alterations of the vascular reactivity to phenylephrine induced by 2K1C and the effects of ALSK and L-arginine
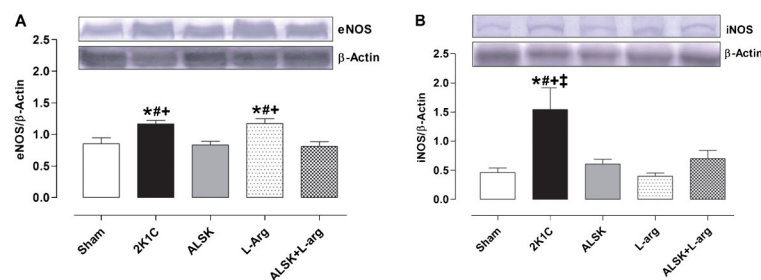

Figure 4. Effects of aliskiren (ALSK) and L-arginine (L-arg) treatment in renovascular hypertension on the densitometric analyses of Western blotting for endothelial nitric oxide synthase (eNOS) (*A*) and inducible nitric oxide synthase (iNOS) (*B*). Data are reported as means ± SE. *P<0.05 *vs* Sham; #P<0.05 *vs* ALSK; ‡P<0.05 *vs* L-arg; +P<0.05 *vs* ALSK + L-arg (one-way ANOVA, followed by Fisher's *post hoc* test).
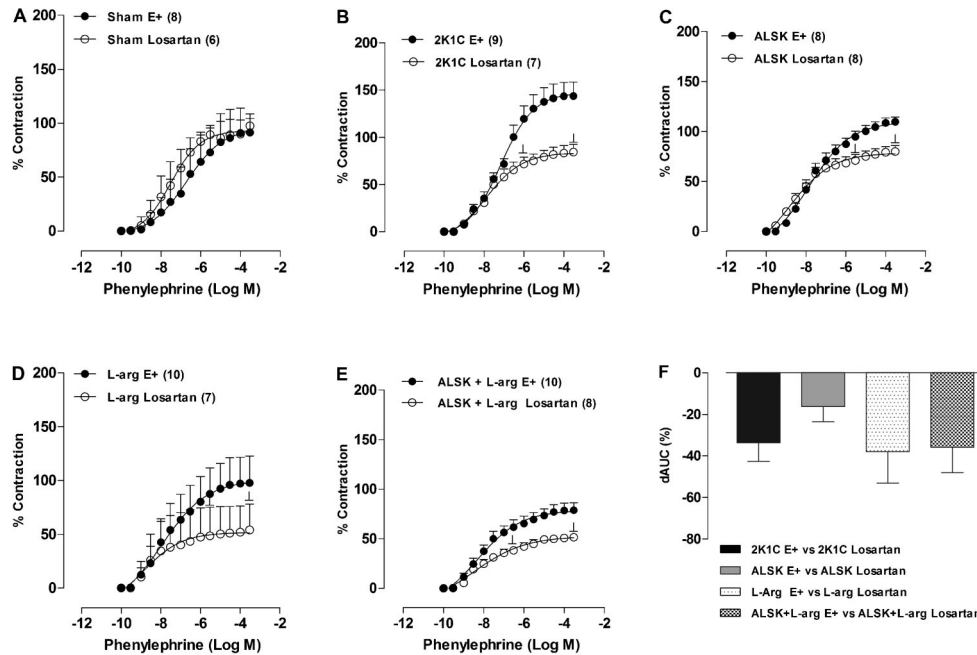
Figure 5. Effects of losartan (10 mM) on the concentration-response curves to phenylephrine in endothelium-intact aortic segments from Sham (*A*), 2K1C (*B*), aliskiren (ALSK) (*C*), L-arginine (L-arg) (*D*), and ALSK + L-arg (*E*) treatments in aortic rings in the presence (losartan) and absence (E +) of losartan blocker. The differences in the area under the concentration-response curves (dAUC) in the presence and absence of losartan are shown in *F*. The number of animals in each group is indicated in parentheses. $^{\perp}P < 0.05$ *vs* E + (two-way ANOVA, followed by Tukey's *post hoc* test).

treatment in this response, AT_1 receptors were blocked with losartan (10 mM). As shown in Figure 5, losartan reduced the vasoconstrictor response induced by phenyl-ephrine in aortas from the 2K1C (Figure 5B), ALSK (Figure 5C), L-arg (Figure 5D), and ALSK + L-arg (Figure 5E) groups, but there were no differences in the dAUC values (Figure 5F). The R_{max} was decreased in the 2K1C, ALSK, L-arg and ALSK + L-arg groups compared to the control (E +), but not sensitivity to phenylephrine (Table 1). Additionally, R_{max} of L-arg and ALSK + L-arg were reduced compared to the 2K1C and Sham groups (Table 1). These findings suggested that 2K1C hypertension stimulated the local RAAS, and that only the combination of ALSK + L-arginine was able to correct this dysfunction.

To further investigate the involvement of the local RAAS on the effects of 2K1C hypertension and ALSK and L-arginine treatment, expression of the angiotensin AT_1 and AT_2 receptors was evaluated. Western blot analyses showed increased levels of AT_1 receptor protein expression in the aortas from the 2K1C group compared with the Sham, ALSK and ALSK + L-arg groups (Figure 6A). AT_2 receptor protein expression was increased in the aortas from the ALSK + L-arg group compared with the Sham and L-arg groups (Figure 6B). Together, these results indicated that 2K1C hypertension induced endothelial dysfunction in conductance arteries through an upregula-tion of AT_1 receptor expression, and the ALSK and

L-arginine combination prevented these responses and also promoted an upregulation of AT_2 receptor expression.

Role of free radicals in the effects of ALSK and L-arginine treatment on the phenylephrine response

To determine whether the endothelial changes observed in the aortic rings after 2K1C hypertension and ALSK and L-arginine treatment were related to changes in superoxide anion production, the effects of the superoxide anion scavenger SOD and the NADPH oxidase inhibitor, apocynin, on the vasoactive responses were analyzed. SOD reduced vascular reactivity to phenylephrine in the 2K1C (Figure 7B) and ALSK (Figure 7C) groups (P<0.05). However, the magnitude of this response, as shown by the differences in the dAUC, was significantly greater in the 2K1C than in the ALSK group (2K1C: −49.9 ± 5.91% *vs* ALSK: −29.6 ± 6.93%, P<0.05, Figure 7F). Additionally, SOD reduced the R_{max} of the 2K1C and ALSK groups compared with the control E + group and increased the sensitivity (pD_2) of 2K1C compared with control E +. On the other hand, apocynin, an inhibitor of NADPH oxidase, reduced the phenylephrine responses in the aortic segments from group 2K1C (Figure 8B), ALSK (Figure 8C), and ALSK + L-arg treated rats (Figure 8E), but the decrease was smaller in the ALSK + L-arg group than in the 2K1C group; this difference was clearly seen when
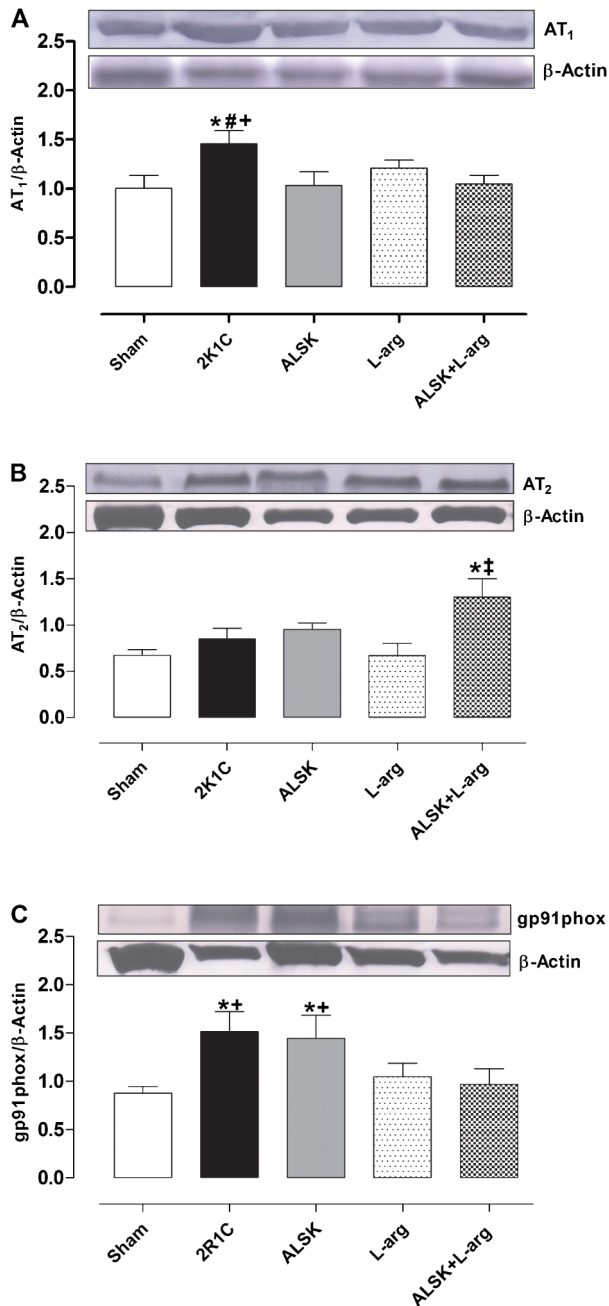
Figure 6. Densitometric analyses of angiotensin receptor-1 (AT$_1$) (*A*), AT$_2$ (*B*) and gp91phox (*C*) in aortas from Sham, 2K1C, aliskiren (ALSK), L-arginine (L-arg), and ALSK+L-arg treated rats. Data are reported as means±SE. *P<0.05 *vs* Sham; #P<0.05 *vs* ALSK; ‡P<0.05 *vs* L-arg; +P<0.05 *vs* ALSK+L-arg (one-way ANOVA, followed by Fisher's *post hoc* test).

dAUC were compared (2K1C: −64.6±6.57% *vs* ALSK+L-arg: −18.68 ±10.3%, P<0.05, Figure 8F). Incubation with apocynin reduced the R$_{max}$ of 2K1C and ALSK+L-arg groups compared with the Sham group.

ALSK+L-arg treatment also reduced R$_{max}$ compared with L-arg treatment (Table 1). To further investigate the involvement of the local oxidative stress on the effects of 2K1C hypertension and ALSK and L-arginine treatment, the expression of the gp91phox, the heme binding subunit of the superoxide-generating NADPH oxidase, was analyzed. Western blot analysis revealed increased levels of gp91phox-containing NADPH oxidase protein expression in the aortas from the 2K1C and ALSK groups compared with the Sham group. ALSK+L-arg treatment reduced the expression of this enzyme compared with expression in the 2K1C and ALSK groups (Figure 6C).

Discussion

The present study demonstrated the effects of a 21-day treatment with ALSK and L-arginine, alone or in combination, on blood pressure and vascular reactivity to phenylephrine in rats with renovascular hypertension. The major findings of this study were as follows: *i*) the high levels of blood pressure promoted by the 2K1C model were partially restored by L-arg treatment, and were fully restored with the combination of L-arg and ALSK; *ii*) all treatments reduced the vasoconstrictor response to phenylephrine and prevented endothelial dysfunction; *iii*) the mechanisms related to the reduction in blood pressure and prevention of endothelial dysfunction in the ALSK+L-arg group were most likely associated with improvements in the vascular RAAS and the reduction in oxidative stress. This is the first study to evaluate the effects of these treatments on vascular reactivity in this model of hypertension.

Renovascular hypertension is caused by an increased generation of angiotensin II owing to increased renal renin release. Therefore, excess angiotensin II production via several different effector pathways is at least partially responsible for the establishment and development of hypertension, left ventricular hypertrophy, and endothelial dysfunction (6,7), which may result from the interplay of several mechanisms (20). We demonstrated that only the combination of ALSK and L-arg normalized blood pressure in rats with 2K1C hypertension, suggesting possible additive effects associated with combined therapy. ALSK induced negligible antihypertensive effects, but those effects were associated with a functional improvement in aorta reactivity to phenylephrine, suggesting that renin is a mediator in the pathogenesis of 2K1C hypertensive-induced vascular alterations. Additional studies are needed to establish the mechanisms responsible for these responses. 2K1C hypertension increases vasoconstriction to phenylephrine in the aorta (2), which could be caused by a reduction in NO availability (5), or increased vascular superoxide anion production by activating vascular NADPH oxidase (21,22).

To investigate endothelial modulation, the endothelium was removed. Following removal, we observed that
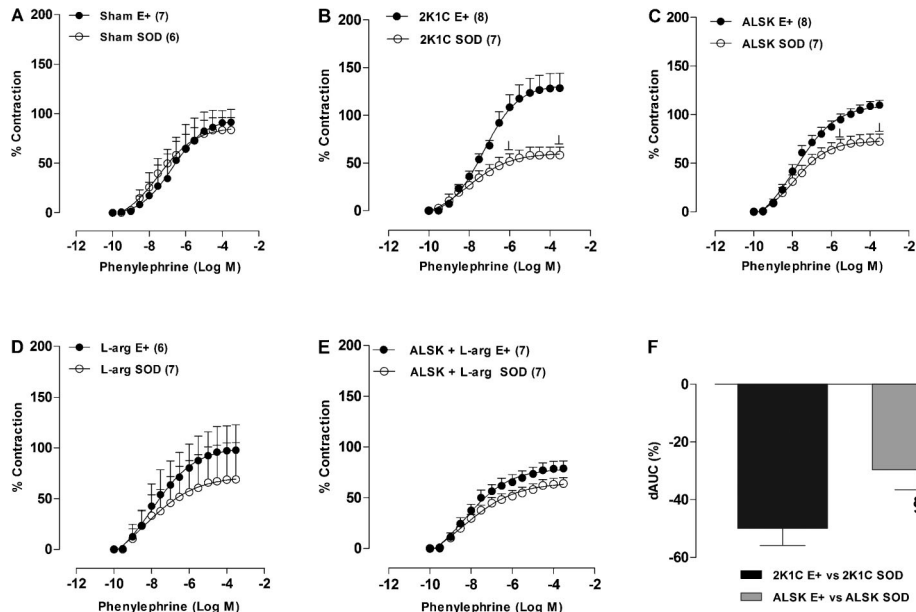
Figure 7. Effects of superoxide dismutase (SOD, 150 U/mL) on the concentration-response curves to phenylephrine in endothelium-intact aortic segments from Sham (*A*), 2K1C (*B*), aliskiren (ALSK) (*C*), L-arginine (L-arg) (*D*), and ALSK + L-arg (*E*) treatments in aortic rings in the presence (SOD) and absence (E+) of SOD incubation. The differences in the area under the concentration-response curves (dAUC) in the presence and absence of SOD are shown in *F*. Data are reported as means ± SE. The number of animals in each group is indicated in parentheses. [§]$P < 0.05$ *vs* 2K1C and [⊥]$P < 0.05$ *vs* E+ (two-way ANOVA, followed by Tukey's *post hoc* test).
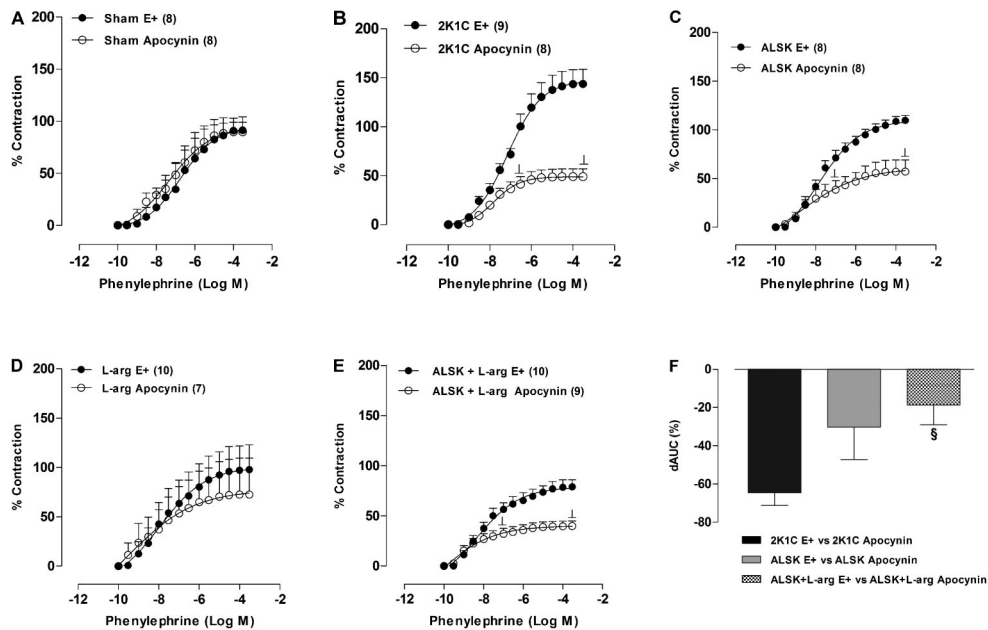

Figure 8. Effects of apocynin (0.3 nM) on the concentration-response curves to phenylephrine in endothelium-intact aortic segments from Sham (*A*), 2K1C (*B*), aliskiren (ALSK) (*C*), L-arginine (L-arg) (*D*), and ALSK + L-arg (*E*) treatments in aortic rings in the presence (apocynin) and absence (E+) of apocynin blocker. The differences in the area under the concentration-response curves (dAUC) in the presence and absence of apocynin are shown in *F*. Data are reported as means ± SE. The number of animals in each group is indicated in parentheses. [§]$P < 0.05$ *vs* 2K1C and [⊥]$P < 0.05$ *vs* E+ (two-way ANOVA, followed by Tukey's *post hoc* test).

the contractile response was enhanced in all groups; however, the magnitude of this response, as assessed by the dAUC, was higher in the rats treated with ALSK+L-arg than in those given ALSK or 2K1C treatment alone. These data suggest that treatment with ALSK+L-arg was more effective in releasing an endothelium-derived relaxation factor. Other investigations have also indicated the involvement of the vascular endothelium in modulating renovascular hypertension (5,23,24). Thus, the combination of drugs appeared to restore the endothelial dysfunction induced by the 2K1C model.

To investigate the role of NO in the 2K1C model and the treatment methods, NOS was inhibited by L-NAME. We observed that the contractile response was enhanced in all groups; however, the size of this response was higher in the groups treated with ALSK+L-arg and ALSK alone than in the 2K1C group. These data suggested that 2K1C hypertension induced endothelial dysfunction in conductance arteries, thereby reducing the endothelial-induced NO modulation of the vasoconstrictor response. Moreover, treatment with ALSK was crucial for endothelial modulation in the contractile response to phenylephrine. We also observed that 2K1C hypertension increased the expression of this eNOS isoform, corroborating the results of Hiyoshi et al. (25), who have also reported that 2K1C hypertension increases aortic levels of total eNOS. Other studies have demonstrated that mechanical forces on the vascular wall, such as blood pressure and shear stress, can increase the expression of eNOS in endothelial cells (26). Therefore, the increase in eNOS may be a compensatory mechanism of the reduced endothelial NO modulation observed in this hypertension model. However, despite the improvements in the vascular responses mediated by NO, eNOS protein expression in the groups treated with ALSK was not altered, in contrast to other reports that have shown an increased expression of this enzyme in double transgenic mice expressing human renin and angiotensinogen genes (27). The mechanism of NO-mediated vascular improvement with ALSK treatment might be related to an increase in eNOS activity, as reported in the SHR model (28), as well as to the AT_1 receptor restoration in our study, which reduced the activation of NADPH oxidase and ROS release and consequently augmented NO bioavailability.

2K1C hypertension increased the expression of iNOS in the aortic rings of 2K1C rats. However, we also demonstrated that the iNOS was reduced by all treatments, suggesting that both drugs were effective in preventing the upregulation of iNOS observed in 2K1C rats. This finding is important because angiotensin II may induce an increased expression of iNOS in endothelial cells, and this effect is associated with increased oxidative stress and the generation of ROS (29,30). Moreover, previous studies have shown that the iNOS isoform is able to generate superoxide anions independent of NO production (26,31).

Previous reports have shown that an increase in the concentration of angiotensin II increases the level of ROS in the aortas of normotensive and 2K1C hypertensive rats (22,32) and that the superoxide anions, one of the most important radicals for vascular biology, can directly promote changes in vascular function and are also essential for the formation of other reactive species (33,34).

Therefore, we investigated the involvement of the local renin-angiotensin system and the role of ROS on vascular reactivity to phenylephrine and the modulation of these systems by ALSK and L-arginine treatment. The losartan-blocking effects suggest that 2K1C hypertension increased AT_1 receptor expression, which is in agreement with the upregulation of AT_1 receptor expression in the 2K1C group. These data suggest the involvement of the local renin-angiotensin system in this experimental model, which induces vasoconstriction and contributes to the increase in vascular reactivity. When the AT_1 receptor was inhibited with losartan (Table 1), the L-arginine and ALSK+L-arginine treatments reduced R_{max} compared with the 2K1C and Sham groups, demonstrating the efficacy of these treatments in modulating the AT_1 receptor, as confirmed by the reduced AT_1 receptor expression in the ALSK+L-arg group. However, expression of the AT_2 receptor was not different in the combined treatment group compared with the 2K1C group, suggesting that the enhanced vascular reactivity in the ALSK+L-arg group was most likely not mediated by this receptor.

To better understand the role of oxidative stress in contractile vascular reactivity responses in 2K1C rats, an NADPH oxidase inhibitor (apocynin) and superoxide scavenger (SOD) were used. When the aortic rings were exposed to apocynin, the contractile response to phenylephrine was reduced in the 2K1C, ALSK, and ALSK+L-arg groups; however, the magnitude of this response was lower in the ALSK+L-arg group compared with the 2K1C group, suggesting that ALSK+L-arg is accompanied by reduced ROS production. Furthermore, treatment with L-arginine alone did not alter vascular reactivity to phenylephrine, suggesting that L-arginine could be the main factor involved in reducing ROS release. We also incubated aortic rings with SOD and obtained similar results to those with apocynin, demonstrating the efficacy of the treatments in reducing vascular oxidative stress. We also demonstrated that 2K1C hypertension increases gp91phox expression, suggesting that the increased vascular reactivity to phenylephrine induced by 2K1C hypertension might be caused by an increased release of ROS, most likely resulting in a reduction of NO bioavailability. Previous studies have shown that angiotensin II leads to the activation of NADPH oxidase in all vascular layers, a process that results in the scavenging of endothelium-derived NO and subsequent attenuation of endothelium-dependent relaxation (22). However, we have demonstrated that combined ALSK and L-arg

treatment reduced the magnitude of contractile responses to phenylephrine and reduced gp91phox expression, suggesting that this combination treatment minimized the release of ROS. Jung et al. (22) demonstrated that the endothelial dysfunction observed during renovascular hypertension in mice results from the activation of endothelial gp91phox-containing NADPH oxidase, suggesting that combined ALSK and L-arg treatment could recover endothelial function.

The present study showed that combined ALSK + L-arg treatment was more effective in reducing blood pressure and preventing the endothelial dysfunction in aortic rings of 2K1C hypertensive rats than the other experimental treatments. Moreover, the mechanisms responsible for these improvements appear to be related to the modulation of RAAS receptor expression, which is associated with the reduction in endothelial oxidative stress mediated by the NADPH oxidase system.

Acknowledgments

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