RESEARCH ARTICLE

Toward quantitative neuroimaging biomarkers for Friedreich's ataxia at 7 Tesla: Susceptibility mapping, diffusion imaging, R_2 and R_1 relaxometry



Sina Straub¹ Stephanie Mangesius^{2,3} Julian Emmerich^{1,4} Michaelicato⁵ Katja S. Degenhardt^{1,4} Katja S. Degenhardt^{1,4} Mark E. Ladd^{1,4,6} Sylvia Boesch⁵ Elke R. Gizewski^{2,3}

¹Division of Medical Physics, German Cancer Research Center (DKFZ), Heidelberg, Germany

²Department of Neuroradiology, Medical University of Innsbruck, Innsbruck, Austria

³Neuroimaging Core Facility, Medical University of Innsbruck, Innsbruck, Austria

⁴Faculty of Physics and Astronomy, Heidelberg University, Heidelberg, Germany

⁵Department of Neurology, Medical University of Innsbruck, Innsbruck, Austria

⁶Faculty of Medicine, Heidelberg University, Heidelberg, Germany

Correspondence

Stephanie Mangesius, Department of Neuroradiology, Medical University of Innsbruck, Anichstrasse 35, 6020 Innsbruck, Austria.

Email: stephanie.mangesius@i-med.ac.at

Abstract

Friedreich's ataxia (FRDA) is a rare genetic disorder leading to degenerative processes. So far, no effective treatment has been found. Therefore, it is important to assist the development of medication with imaging biomarkers reflecting disease status and progress. Ten FRDA patients (mean age 37 \pm 14 years; four female) and 10 ageand sex-matched controls were included. Acquisition of magnetic resonance imaging (MRI) data for quantitative susceptibility mapping, R₁, R₂ relaxometry and diffusion imaging was performed at 7 Tesla. Results of volume of interest (VOI)-based analyses of the quantitative data were compared with a voxel-based morphometry (VBM) evaluation. Differences between patients and controls were assessed using the analysis of covariance (ANCOVA; p < 0.01) with age and sex as covariates, effect size of group differences, and correlations with disease characteristics with Spearman correlation coefficient. For the VBM analysis, a statistical threshold of 0.001 for uncorrected and 0.05 for corrected p-values was used. Statistically significant differences between FRDA patients and controls were found in five out of twelve investigated structures, and statistically significant correlations with disease characteristics were revealed. Moreover, VBM revealed significant white matter atrophy within regions of the brainstem, and the cerebellum. These regions overlapped partially with brain regions for which significant differences between healthy controls and patients were found in the VOI-based quantitative MRI evaluation. It was shown that two

Abbreviations: ASPIRE, computationally efficient combination of multi-channel phase data from multi-echo acquisitions; B₁, transverse RF magnetic field; BET, brain extraction tool; CP, cerebral peduncle; CS, corticospinal fibers; DARTEL, diffeomorphic anatomical registration using exponentiated lie algebra; DN, dentate nuclei; DTI, diffusion tensor imaging; EPI, echo planar imaging; FA, fractional anisotropy; FAST, FMRIB's automated segmentation tool; FLIRT, FMRIB's linear image registration tool; FMRIB, Oxford Centre for functional MRI of the brain; FNIRT, FMRIB's non-linear image registration tool; FSL, FMRIB software library; GRAPPA, generalized autocalibrating partially parallel acquisitions; GRE, gradient echo; ICE, image calculation environment; ICP, inferior cerebellar peduncle; ID, identification number; JHU, Johns Hopkins University; MCP, middle cerebellar peduncle; MITK, Medical Imaging Interaction Toolkit; ML, medial lemniscus; MNI, Montreal Neurological Institute; MP2RAGE, magnetization prepared 2 rapid acquisition gradient echoes; MRI, magnetic resonance imaging; PCT, pontine crossing tract; PTR, posterior thalamic radiation (including optic radiation); QSM, quantitative susceptibility mapping; RESOLVE, readout segmentation of long variable echo-trains; RN, red nucle; SCP, superior cerebellar peduncle; SN, substantia nigra; SNR, signal-to-noise ratio; SPM, statistical parametric mapping; SS, sagittal stratum (including inferior longitudinal fasciculus and inferior fronto-occipital fasciculus); STAR-QSM, dipole inversion the streaking artifact reduction for QSM; STEAM, stimulated echo acquisition mode; TSE, turbo spin echo; VOI/VOIs, volume of interest/volumes of interest; V-SHARP, variable-kernel sophisticated harmonic artifact reduction for phase data.

Edited by Jeremy Hogeveen. Reviewed by Alexander Rauscher and Andrea Martinuzzi.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. Journal of Neuroscience Research published by Wiley Periodicals LLC

2220

independent analyses provided overlapping results. Moreover, positive results on correlations with disease characteristics were found, indicating that these quantitative MRI parameters could provide more detailed information and assist the search for effective treatments.

KEYWORDS

brainstem, cerebellum, diffusion imaging, fiber tracts, Friedreich's ataxia, magnetic resonance imaging, midbrain, MR relaxometry, nuclei, quantitative MRI, quantitative susceptibility mapping

1 | INTRODUCTION

Friedreich's ataxia (FRDA) is a rare genetic disorder affecting the FXN gene which produces the protein frataxin within the mitochondria of the cells (Koeppen, 2011). More precisely, most patients inherit trinucleotide (GAA) expansions within the first intron of the FXN gene, and those with a higher number of repeats (>500) are more likely to present at an earlier age and with different clinical symptoms (Schols et al., 1997), whereas longer GAA repeats lead to lower frataxin levels (Sacca et al., 2011). A lack of frataxin disrupts the iron metabolism (Gibson, Koonin, Musco, Pastore, & Bork, 1996; Zhang et al., 2006) leading to degenerative processes especially in tissues with a high metabolic rate such as the heart, pancreas, as well as the nervous system (Koeppen, 2013). Within the nervous system, white matter fiber tracts, spinal nerves, and the cerebellum are affected (Lamarche, Lemieux, & Lieu, 1984). FRDA is the most common inherited ataxia presenting with early onset of clinical manifestations (Koeppen, 2011). So far, no effective treatment has been found that can stop or reverse the progress of FRDA. Therefore, it is important to assist the development of efficient medication by finding reliable imaging biomarkers that reflect the disease status and progress in FRDA patients.

An extensive body of literature of cerebral abnormalities in FRDA exists (Selvadurai, Harding, Corben, & Georgiou-Karistianis, 2018) in which functional and structural cerebral and cerebellar white and gray matter changes are assessed: Volumetric, voxel-based morphometry (VBM), and tract-based spatial statistics (TBSS) magnetic resonance imaging (MRI) studies have shown a significant atrophy of the cerebral and cerebellar cortex in FRDA patients (Cocozza et al., 2020; Selvadurai et al., 2016) and the dentate nuclei (DN; Solbach et al., 2014; Stefanescu et al., 2015), a degeneration of white matter tracts (Della Nave et al., 2008; Pagani et al., 2010; Selvadurai et al., 2020), as well as a significant correlation of those findings with disease severity or certain symptoms (Akhlaghi et al., 2011) such as the impairment of the visuospatial functions (Cocozza et al., 2020). Another study including VBM and TBSS analyses found atrophy in several parts of the brainstem, and cerebellum, but did not observe any focal signal abnormalities on proton-density and on T₂-weighted images (Pagani et al., 2010). A recent study (Selvadurai et al., 2020), that used multiple diffusion imaging-based metrics as well as magnetization transfer imaging to assess microstructural and T_1 -weighted

Significance

In five out of twelve investigated structures of the brainstem and the cerebellum, statistically significant differences were found in Friedreich's ataxia patients versus controls with quantitative magnetic resonance imaging at 7 Tesla (quantitative susceptibility mapping, R_1 , R_2 relaxometry and diffusion imaging), and statistically significant correlations with disease characteristics were revealed. The regions in which voxel-based morphometry showed significant white matter atrophy partially overlapped with the brain regions for which significant differences between controls and patients were found in the volume-of-interest-based quantitative MRI evaluation. Quantitative MRI parameters could provide more detailed information about the disease and assist the search for effective treatments.

imaging to assess macrostructural white matter changes found extensive disruption of cerebello-cerebral and intracerebral white matter tracts. Quantitative methods such as quantitative susceptibility mapping (QSM; Ward et al., 2019) and T_2 mapping (da Silva et al., 2014) seemed to provide useful information on longitudinal change of disease severity that correlated with longitudinal change in DN magnetic susceptibility. Most conclusive among several studies is the relation of disease severity and degenerative changes in the superior cerebellar peduncles and in peridentate white matter as well as the DN pathology which is an established observation in FRDA.

Ultra-high field MRI has been shown to outperform conventional clinical MRI in the delineation of very small anatomical structures (Deistung, Schafer, Schweser, Biedermann, Gullmar, et al., 2013; Straub et al., 2019), and in the detection of small pathological changes such as in epilepsy (Zeineh et al., 2009) and micro-hemorrhage (Theysohn et al., 2011) due to the higher resolution as well as the higher contrast-to-noise ratios available at higher field strengths (Ladd et al., 2018). To the best of our knowledge, no study has assessed whether quantitative MRI other than diffusion tensor imaging (DTI), magnetization transfer ratio, or spectroscopy (Blair et al., 2019) could provide complementary or additional information to evaluate the current stage and the course of Friedrich's ataxia. Compared to 3 T MRI, it has been shown that 7 T MRI can provide contrast-tonoise ratios up to almost fourfold higher for certain structures in the brainstem (Straub et al., 2019). Utilizing the higher contrast-to-noise ratio available at ultra-high field, this study investigates the degeneration of fiber tracts throughout the brainstem as well as of the DN, the red nuclei (RN), and the substantia nigra (SN) in FRDA with quantitative MR parameters (susceptibility, diffusion anisotropy, and R_{2} and R_{1} relaxometry) by assessing a patient and a control group, and compares its results with a VBM analysis of the same cohorts. QSM was chosen as a contrast due to its ability to quantify tissue iron content (Langkammer, Schweser, et al., 2012) and its sensitivity to myelin (Lee et al., 2012). R_1 and R_2 relaxometry was used since its known to be influenced by myelin and iron (Harkins et al., 2016; Koenig, 1991; Langkammer, Krebs, et al., 2012; MacKay et al., 2006; Ogg & Steen, 1998). DTI has long been utilized to assess white matter fiber integrity (Chanraud, Zahr, Sullivan, & Pfefferbaum, 2010), and was therefore also included as a quantitative MRI parameter. Whether the used quantitative MRI methods provide sufficiently large effects to reveal differences between FRDA patients and healthy controls as well as correlations with disease characteristics at clinical field strengths are beyond the scope of this study and left to further investigations.

2 | METHODS AND MATERIALS

2.1 | Patient and control cohorts

This study was conducted in accordance with the Declaration of Helsinki. Institutional review board approval was obtained and all subjects provided written informed consent. Inclusion criteria were diagnosis of FRDA, physical and mental capability of providing informed consent and participating in the study. Exclusion criteria involved pregnancy and contraindications for 7 T MRI. As FRDA is a rare disease, the inclusion of 10 patients was the goal to achieve a sufficiently large patient cohort for the analyses. Ten genetically confirmed FRDA patients (mean age 37 ± 14 years; four female) and 10 age- and sex-matched healthy controls (mean age 37 ± 14 years; four female) were included in the study. Patients were recruited at the Department for Neurology, Medical University of Innsbruck, Austria, and MR images were generated at the DKFZ Heidelberg, Germany. All participants finished the study. Nine patients had homozygous GAA triplet expansions and one patient was heterozygous (one GAA triplet expansion and one point mutation). For eight patients, length of GAA triplet repeats was available. Table 1 summarizes the patient cohort.

2.2 | Data acquisition

Data were acquired at a whole-body 7 T system (Magnetom 7 Tesla) with an 8Tx/32Rx-channel head coil (Nova Medical Inc., Wakefield, MA, USA) by use of an in-house-constructed butler matrix. A monopolar three-dimensional multi-echo gradient echo (GRE), a twodimensional (2D) RESOLVE (readout segmentation of long variable echo-trains; Heidemann et al., 2010; Porter & Heidemann, 2009) with 11 shots in stimulated acquisition mode (STEAM; Merboldt, Hanicke, & Frahm, 1991; Tanner, 1970) with mixing time 50 ms and with b-values 50 and 800 s/mm² and 20 diffusion directions, an EPI (echo planar imaging) factor of 68, echo spacing of 0.32 ms, and fat saturation, and a vendor-provided MP2RAGE (magnetization prepared 2 rapid acquisition gradient echo; Marques et al., 2010) sequence were acquired using the same parameters as in a previous study (Straub et al., 2019). The RESOLVE sequence is equipped with navigator-based reacquisition which was used for the acquisition of the DTI data. For dictionary-based T_2 mapping, a 2D multiecho turbo spin echo (ME-TSE) with turbo factor 5 and a turbo flash sequence for B₁ mapping (Chung, Kim, Breton, & Axel, 2010) were acquired as well. All sequences were acquired in strictly transversal orientation with phase-encoding direction right-left. Sequence parameters are summarized in Table 2. Patients and controls received

TABLE 1 Patient characteristics including patient age, sex, age at disease onset in years, and the number of GAA repeats

Patient number	Age in years	Age of disease onset in years	GAA1 repeats	GAA2 repeats	Sex
1	22	12	>700	>700	Male
2	49	28	250	250	Female
3	62	22	120	150	Male
4	46	34	350	350	Male
5	50	27	350	350	Female
6	34	20	500	600	Female
7	31	17	400	950	Male
8	18	15	700	970	Female
9	37	17	-	n.a.	Male
10	23	15	-	-	Male

Abbreviations: GAA1/GAA2, GAA repeats; n.a., not applicable (point mutation).

22	N	jour eui	NAL O	cie	nc	e R	ese	arc	ch−		
Acquisition	time min:sec	18:09		23:55		13:21		8:08		1:32	
Partial Fourier (slice,	phase)	6/8, 7/8		I		I		6/8, 7/8		ı	
GRAPPA (factor/ref.	lines)	2/36		2/68		3/27		3/43		I	
	BW in Hz/px	140		631		130		240		490	
FA ::	'n	12		180		120		4,5		7	
	II in ms	I		I		I		900, 2750		I	
TT :	I E in ms	10.5, 21, 31.5		59, 99		15, 88, 161,235		4.01		2.36	
TR in	ms	38		2700		7400		5000		3000	
Resolution in mm ³ ,	matrix size	$0.47 \times 0.47 \times 0.47$	464 imes 348 imes 192	$1.2 \times 1.2 \times 2.4$	176 imes 142 imes 14	$0.49 \times 0.49 \times 2.0$	$448 \times 336 \times 30$	$0.69 \times 0.69 \times 0.69$	$320 \times 240 \times 192$	1.15 imes 1.15 imes 4.0	192 imes 144 imes 15
		ME-GRE		RESOLVE		ME-TSE		MP2RAGE		Turbo flash (B_1)	

anonymized subject identifications numbers (IDs), and all MR images contained theses IDs only.

2.3 Data processing

2.3.1 | Quantitative susceptibility mapping

For susceptibility map generation, single-channel data were combined using the computationally efficient combination of multichannel phase data from multi-echo acquisitions (ASPIRE; Eckstein et al., 2018) on the scanner and echo-wise unwrapped with Laplacianbased phase unwrapping (Li, Avram, Wu, Xiao, & Liu, 2014; Li, Wu, & Liu, 2011; Wu, Li, Guidon, & Liu, 2012). After background field removal with the variable-kernel sophisticated harmonic artifact reduction for phase data (V-SHARP; Li et al., 2014; Wu, Li, Guidon, et al., 2012) method (kernel size up to 12 mm), all echoes were averaged (Wu, Li, Avram, Gho, & Liu, 2012), and a dipole inversion was performed using the streaking artifact reduction for QSM (STAR-QSM) algorithm (Wei et al., 2015). Background field removal required a brain mask which was generated with FSL-BET (Brain Extraction Tool; Smith, 2002), and manually corrected in the Medical Imaging Interaction Toolkit (MITK; Maleike, Nolden, Meinzer, & Wolf, 2009; Nolden et al., 2013). Volumes of interest (VOIs) for cerebrospinal fluid (CSF) in the atria of the lateral ventricles were manually drawn in MITK and used to reference susceptibility maps (Straub et al., 2017). Except for the coil combination, all processing steps were performed in Matlab (Matlab, R2017b, MathWorks, Natick, USA).

Relaxometry and diffusion tensor imaging 2.3.2

The longitudinal relaxation rate R_1 was calculated as $R_1 = 1/T_1$ from the vendor-provided T_1 maps that were acquired with the MP2RAGE sequence. The vendor-provided diffusion fractional anisotropy maps were used for the diffusion data evaluation. Maps of the transverse relaxation rate $R_2 = 1/T_2$ were calculated from ME-TSE data and B_1 maps using a dictionary-based method (Emmerich et al., 2019): A dictionary for the echo intensities was computed for the ME-TSE sequence according to the sequence parameters (Table 2) using a combination of Bloch simulations to compute the slice profiles and subsequently extended phase graph model (Weigel, 2015) simulations to compute the resulting signal intensities. The measured signal intensities and B₁ maps were then fitted to this precalculated dictionary to obtain T_2 values.

2.4 | Data analysis

time; TI, inversion time; TR, repetition time

2.4.1 | Volumes of interest generation

Volumes of interest (Figure 1) were generated for nine fiber tracts (middle cerebellar peduncle-MCP, pontine crossing tract-PCT, corticospinal tract-CS, medial lemniscus-ML, inferior cerebellar peduncle-ICP,

2223



FIGURE 1 Overview of the volumes of interest. JHU atlas labels in MNI space that were used as volumes of interest for the middle cerebellar peduncle (MCP), pontine crossing tract (PCT), corticospinal tract (CS), medial lemniscus (ML), inferior cerebellar peduncle (ICP), superior cerebellar peduncle (SCP), cerebral peduncle (CP), posterior thalamic radiation (PTR; including optic radiation), sagittal stratum (SS; including inferior longitudinal fasciculus and inferior fronto-occipital fasciculus), and manually drawn volumes of interest for substantia nigra (SN), red nuclei (RN), and dentate nuclei (DN) on susceptibility maps (QSM) are shown [Color figure can be viewed at wileyonlinelibrary.com]

superior cerebellar peduncle-SCP, cerebral peduncle-CP, posterior thalamic radiation (including the optic radiation)-PTR, sagittal stratum (including the inferior longitudinal, fasciculus, and the inferior frontooccipital fasciculus)-SS) by registering JHU-ICBM-DTI-81 white matter labels (Mori et al., 2008) to the MP2RAGE data of this study with FSL-FLIRT (Jenkinson & Smith, 2001) and FSL-FNIRT (Andersson, Jenkinson, & Smith, 2010) using the FSL MNI152_T1_0.5mm template. For DN, RN, and SN, bilateral VOIs were generated manually on the susceptibility maps using MITK including all slices on which the structures were morphologically visible for each patient/control individually. All VOIs were subsequently registered to the other imaging contrasts with FSL-FLIRT and automatically corrected for registration errors by thresholding to exclude CSF pixels in the case of R_1 , R_2 , and FAmaps to eliminate voxels from the VOIs for which $R_1 < \frac{1}{3000} \frac{1}{m}$, $R_2 < \frac{1}{170} \frac{1}{m}$, and FA < 300, respectively. In the case of susceptibility maps, a CSF mask that was generated with FSL-FAST (Zhang, Brady, & Smith, 2001) was used for automatic correction. A manual VOI assessment ensued, as well as a correction in cases in which pixels were included in the VOI that clearly did not belong to the corresponding structure, i.e. pixels that were wrongly positioned in the CSF or belonged to a different

anatomical structure. As the DTI data only covered an approximately four centimeter slab, not all fiber tracts were feasible for DTI analysis, the brain nuclei were not included in DTI analysis as well.

2.4.2 | Voxel-based morphometry

VBM (Ashburner & Friston, 2000; Mechelli, Price, Friston, & Ashburner, 2005) was performed using the MP2RAGE data and Statistical Parametric Mapping (SPM12, https://www.fil.ion.ucl. ac.uk/spm/software/spm12/) in Matlab. Following (Ashburner, 2010), first, grey matter, white matter, CSF, skull, and head were segmented using default parameters and generating the input data for the next step. Then, a template was generated from all controls and patients using the "Diffeomorphic Anatomical Registration Using Exponentiated Lie Algebra" (DARTEL) algorithm (Ashburner, 2007) with default parameters. Finally, DARTEL tools were used to normalize white matter images to the Montreal Neurological Institute (MNI) space which included smoothing with an 8-mm Gaussian kernel, spatially normalizing, and scaling by the Jacobian determinant.

2.4.3 | Statistical analysis

All statistical analyses were performed in Matlab, and with SPM12 for the general linear model analysis for VBM evaluation. Mean susceptibility values, relaxation rates, and mean diffusion fractional anisotropy (FA) were calculated. Differences between the patients and the healthy controls were assessed using the analysis of covariance (ANCOVA) with age and sex as covariates, and effect size of the group differences between healthy controls and patients was calculated using Cohen's d. Correlations with FRDA characteristics as well as with patient age (Table 1) were assessed using the Spearman correlation coefficient. p-Values of less than 0.01 were considered statistically significant. To analyze statistically significant differences in white matter volume from the VBM evaluation between the patient and the control group, a two-sided t-test was performed in SPM12 with age and sex as covariates, first uncorrected with a statistical threshold of 0.001, and secondly using the SPM12 familywise error correction to correct for multiple comparisons with a statistical threshold of 0.05. No data were removed prior to analysis. Data processing was performed blinded to disease characteristics.

3 | RESULTS

3.1 | Differences between FRDA patients and controls

In Table 3, the effect size for the differences between the patient and the control group is given by Cohen's *d*. Largest effect size (above 2) is observed for R_2 relaxometry of the RN and for diffusion fractional anisotropy of the SCPs. For mean susceptibility values of the DN,

	QSM	R ₁	R ₂	FA
MCP		0.65	0.66	0.53
PCT				0.55
CS				1.18
ML		0.76	0.95	0.57
ICP		1.06		1.77
SCP		1.46	1.22	2.28
СР		0.61	1.70	1.06
PTR		0.71	0.83	n.a.
SS		0.60		n.a.
DN	1.46			n.a.
RN	1.62	1.61	2.91	n.a.
SN			1.16	n.a.
	0.5-0.99	1-1.99	2-2.99	

TABLE 3 Effects size (Cohen's d) of group differences

STRAUB ET AL.

and RN, for mean R_1 values of the ICPs and SCPs, and RN, for mean R_2 values of the SCPs, CPs, RN, and SN, as well as for mean diffusion fractional anisotropy of the CS, the ICPs, CPs, the effect size is larger than 1. For mean R_1 values of five other fiber tracts (MCP, ML, CP, PTR, and SS), mean R_2 values of three other fiber tracts (MCP, ML, and PTR), and diffusion fractional anisotropy of three other fiber tracts (MCP, PCT, and ML), the effect size is larger than 0.5.

In five brain regions, which coincide with the regions of largest effect size (Table 3, Cohen's d 1.46 or larger), statistically significant differences (indicated by white stars) were found between healthy controls and patients. Figure 2a summarizes these differences as well as the covariates (age and sex) by showing the *p*-values of the ANCOVA. In Figure 2b, box plots for the VOIs and contrasts, for which *p*-values were low (\ll 0.05), are shown and significant differences are indicated with a black star. The corresponding region is indicated within each box plot. Susceptibility mapping revealed significant differences between healthy controls and patients for the DN and RN for which susceptibility values where higher in patients. In R₁ maps, values for RN were significantly lower in patients and significantly higher in SCP. R₂ mapping showed significant differences between controls and patients for RN, and for one fiber tract (CP), where R₂ was significantly lower/higher. Diffusion fractional was significantly lower in patients in two fiber tracts (ICP and SCP). For SN, there was a significant correlation with age.

Moreover, the differences revealed by the statistical analysis are also visually recognizable. In Figure 3, representative slices of susceptibility, R_1 and R_2 maps, as well as fractional anisotropy maps from diffusion-weighted imaging are shown for brain regions for which significant differences between healthy controls and patients were found. Images from different patients (indicated by patient number P) and age-/sex-matched controls are shown. On susceptibility maps,

Abbreviations: CP, cerebral peduncle; CS, corticospinal tract; DN, dentate nuclei; FA, fractional anisotropy; ICP, inferior cerebellar peduncle; MCP, the middle cerebellar peduncle; ML, medial lemniscus; PCT, pontine crossing tract; PTR, posterior thalamic radiation (including optic radiation); QSM, quantitative susceptibility mapping; RN, red nuclei; SCP, superior cerebellar peduncle; SN, substantia nigra; SS, sagittal stratum (including inferior longitudinal fasciculus and inferior fronto-occipital fasciculus).

2225



FIGURE 2 Significant differences between healthy controls and patients indicated with a star. (a) *p*-Values of ANCOVA for differences and covariates (age and sex) in susceptibility values (χ), R_1 and R_2 values, and diffusion fractional anisotropy (FA) of mean VOI values of the middle cerebellar peduncle (MCP), pontine crossing tract (PCT), corticospinal tract (CS), medial lemniscus (ML), inferior cerebellar peduncle (ICP), superior cerebellar peduncle (SCP), cerebral peduncle (CP), posterior thalamic radiation (PTR; including optic radiation), sagittal stratum (SS; including inferior longitudinal fasciculus and inferior fronto-occipital fasciculus), dentate nuclei (DN), red nuclei (RN), and substantia nigra (SN). For PTR, SS, DN, RN, and SN, FA values were not available from all patients and controls due to insufficient brain coverage of the field of view. (b) Box plots of mean susceptibility values, mean R_1 , mean R_2 , and mean diffusion fractional anisotropy (b) for all patients and healthy controls. Only box plots for low *p*-values (see a) are shown. The whiskers represent the 9th and the 91st percentile [Color figure can be viewed at wileyonlinelibrary.com]

conspicuous differences in DN (first row) and RN (second row) can be observed with a higher susceptibility on the patient images compared to healthy controls. On R_1 maps, higher R_1 relaxation rates are visible in the SCP (third row). Higher R_2 relaxation rates can be seen in the RN, and the CP (fourth row) on R_2 maps for the patient data. On fractional anisotropy maps, a lower fractional anisotropy can be observed for the SCP (fifth row) in the patient data.

3.2 | Voxel-based morphometry

Compared to the age- and sex-matched controls, statistically significant, white matter atrophy was found in the patient cohort in the uncorrected t-test analysis (p < 0.001). Figure 4 shows suprathreshold t-values (transparent red) representing the significant white matter atrophy in the patient cohort, and the JHU-ICBM-DTI-81 white matter labels, which were used in this study, overlaid on the 0.5 mm MNI space template. Except for the VOIs of SS and PTR, suprathreshold t-values partially overlap with all white matter VOIs used in this study. A large overlap can be observed for the cerebellar

peduncles (ICP and SCP). Moreover, suprathreshold t-values were found in the regions of the RN and SN (VOIs not shown), in peridentate white matter, as well as in parts of the thalamus (Figure 4e, arrow), and in large parts of the midbrain (Figure 4e, arrow head). For the corrected t-test analysis, no statistically significant clusters were found.

3.3 | Correlation analysis

In Figure 5, the correlations between the different imaging contrasts and disease characteristics are visualized. Figure 5a shows the correlation coefficients and *p*-values for the correlations between age at disease onset, patient age, GAA1 repeats, GAA2 repeats (Table 1) and mean susceptibility, R_1 , R_2 , fractional anisotropy values in the different VOIs. Statistically significant correlations are indicated by white stars; green stars indicate that additionally significant differences have been found for the respective contrast/VOI combination. For the significant correlations between mean susceptibility values and disease characteristics, a significant correlation with



FIGURE 3 Representative axial slices of susceptibility, R_1 , R_2 , and fractional anisotropy maps (top to bottom) from four different patients and controls. The patient number is indicated in each slice. On susceptibility maps, arrows point to dentate nuclei (DN), and red nuclei (RN), on the R_1 map to superior cerebellar peduncles (SCP), on the R_2 map to the red nuclei (RN), and the cerebral peduncle (CP), and on the fractional anisotropy map arrows point to superior cerebellar peduncles (SCP) [Color figure can be viewed at wileyonlinelibrary.com]

patient age was revealed as well, possibly indicating a bias. The significant correlations between disease characteristics and contrast/ VOI combinations for which no significant correlation with patient age was found are indicated with black circles. Figure 5b shows the scatterplots of mean R_1 values in ML, mean R_2 values in CP and RN, and mean FA values in ICP. All correlation coefficients and *p*-values are given in Table S1.

4 | DISCUSSION

In this study, statistically significant (p < 0.01, no correction for multiple comparisons) differences between a group of 10 FRDA patients and a group of age- and sex-matched healthy controls were found in five (CP, DN, ICP, SCP, and RN) out of twelve investigated structures of the brainstem, and the cerebellum using quantitative MRI (susceptibility, relaxation rates (R_2 , R_1), diffusion fractional anisotropy) at 7 Tesla. It was also shown that the effect size of group differences for susceptibility, R2, R1, and fractional anisotropy values between healthy controls and patients were medium to large (0.53-2.91) for several fiber tracts and brain nuclei, indicating that the used methods in the investigated structures could be useful biomarkers in larger studies. In the present study, only VOI/contrast combinations with an effect size of 1.46 (Cohen's d) or larger for group differences provided significant differences between patients and controls. Moreover, using VBM, statistically significant white matter atrophy was found within regions of the brainstem, midbrain, and the cerebellum that partially overlapped with brain regions for which significant differences between healthy controls and patients were found in the VOI-based quantitative MRI evaluation part of this study. The largest overlap was observed for SCP for which significant differences between patients and healthy controls were found using R_1 relaxometry and diffusion-weighted imaging indicating white matter damage in patients. The atrophy of SCP shown in FRDA previously (Akhlaghi et al., 2011; Della Nave et al., 2008; Pagani et al., 2010) is in accordance with the findings of this study. Moreover, SCP atrophy as well as the atrophy of the peridentate white matter were suggested to correlate with disease symptoms (Akhlaghi et al., 2011; Pagani et al., 2010). Using VBM, atrophy in FRDA patients was also found in the central portion of the medulla oblongata, the dorsal upper pons, the central portion of the midbrain, the medial portion of the right CP, and optic chiasm (Pagani et al., 2010) which partly coincides with the VBM findings of this study. Atrophy of the DN (Solbach et al., 2014) and the RN (Rezende et al., 2019) as well as higher iron content in the DN have been reported in FRDA (Koeppen et al., 2007; Waldvogel, van Gelderen, & Hallett, 1999). These findings are in accordance with the findings of this study, in which higher susceptibility values, higher R_2 values, and lower R_1 values were found in the RN as well as higher susceptibility values in the DN. Iron accumulation or iron redistribution and DN atrophy are well-known features in the cerebral pathology of FRDA (Selvadurai et al., 2018), and pathological changes in RN are supposed to be due to their tight link to the DN, and consequently to the loss or reduction of input through the SCP (Akhlaghi et al., 2014; Habas, Guillevin, & Abanou, 2010; Nioche, Cabanis, & Habas, 2009) which is in accordance with the findings of this study in which pathologic differences were found in DN (with QSM), RN (with QSM, R_1 and R_2 relaxometry), and SCP (with R_1 relaxometry and fractional anisotropy). Even though a dictionary-based R_2 mapping method was used that compensates for B_1 field inhomogeneities, very low B_1 in parts of the cerebellum might affect R_2 in the DN. This could explain why no significant differences between patients and controls were found in DN in contrast to the findings of this (da Silva et al., 2014) 3 T study. Gradient echo techniques such as susceptibility mapping are much less affected by B₁ field inhomogeneities that occur particularly strong at ultra-high field. However, susceptibility values calculated with standard QSM methods depend on fiber orientation (Aggarwal, Kageyama, Li, & van Zijl, 2016), which might be the reason why no significant differences were found with QSM in this study.



FIGURE 4 Agreement of voxel-based morphometry and volume of interest-based analysis. Suprathreshold *t*-values (*p* < 0.001, uncorrected) show regions of significant white matter atrophy overlaid on three representative axial (a–d) and one sagittal slice (e) of the 0.5 mm MNI_T1 space template. JHU labels for the middle cerebellar peduncle (MCP), the pontine crossing tract (PCT), the corticospinal tract (CS), the medial lemniscus (ML), the inferior cerebellar peduncle (ICP), the superior cerebellar peduncle (SCP), the cerebral peduncle (CP), the posterior thalamic radiation (PTR), and the sagittal stratum (SS) are overlaid as well. The arrow indicates the region of the thalamus and the arrow head indicates the midbrain [Color figure can be viewed at wileyonlinelibrary.com]

Moreover, this study evaluated if significant correlations of quantitative MRI parameters of the investigated VOIs and disease characteristics were present. For all MRI contrasts significant correlations with one or more disease characteristics were found. Mean R₁ values within the ML positively correlated with age at disease onset, as well as mean R_2 values within the CP which could indicate more white matter damage in the patients with the earlier disease onset agreeing with current literature (Selvadurai et al., 2020). Mean R₂ values in the RN positively correlated with GAA1 and GAA2 repeats which are in accordance with the relation of GAA repeat length and frataxin levels (Sacca et al., 2011). Fractional anisotropy in the ICP negatively correlated with GAA1 and GAA2 repeats which also agrees with findings in the literature (Selvadurai et al., 2020) that found a correlation of ICP volume and GAA1 repeats. For the significant susceptibility/VOI correlation pairs, a significant correlation with patient age was revealed as well. Magnetic susceptibility measured with QSM is known to be sensitive for age-dependent iron accumulation within the brain nuclei (Acosta-Cabronero, Betts, Cardenas-Blanco, Yang, & Nestor, 2016; Hallgren & Sourander, 1958; Zhang et al., 2018). In this study, this could mask any disease-related changes within the patient group as higher susceptibility values due to iron accumulation could be related to higher patient age or to increased disease severity (that is, longer GAA repeats/earlier age at onset) which cannot be differentiated with the studied cohort. As the control cohort was age-matched, significant differences could very well be observed and results consequently did not suffer from an age-bias.

This study has several limitations. The low number of included patients limits achievable statistical significance. Since FRDA is a rare disease, studies are often rather small (Akhlaghi et al., 2011; Pagani et al., 2010), however, there exist studies using 3 T MRI that include approximately 30 FRDA patients (Selvadurai et al., 2020). Moreover, the patient cohort was relatively inhomogeneous consisting of three young patients (between 18 and 23 years) with early onset (before the age of 25 years) of FRDA, three patients in their 30s as well as one patient aged 62 with early onset, and three patients older than 45 years with rather late onset (older than 25 years). Sex was also only considered as covariate, and sex-related differences were not assessed separately due to the rather low patient number; however, all patients and controls were age- and sex-matched. To further evaluate the value of quantitative MRI for assessing disease status as well as progression and correlation with clinical parameters, studies including larger homogeneous cohorts at high or ultra-high field might be helpful. Moreover, due to a stronger presence of field inhomogeneities at ultra-high field, DTI is even more prone to artifacts compared to clinical field strength, therefore, a readout-segmented EPI sequence with navigator-based reacquisition was chosen at the expense of longer acquisition times. To avoid prohibitively long acquisition times, only 20 diffusion directions were used as well as a lower coverage of the brain. JHU-ICBM-DTI-81 white matter labels were co-registered to the MP2RAGE, as these possessed the largest brain coverage, data of this study and then subsequently co-registered to all other contrasts which could be prone to registration



FIGURE 5 Correlations between quantitative MRI parameters and FRDA patients' disease characteristics as well as with patient age. (a) Correlation coefficients (p, first row) and p-values (p, second row) for correlations between mean volumes of interest (VOIs) values for the different imaging contrasts (susceptibility χ , relaxation rates R_1 and R_2 , diffusion fractional anisotropy FA) and age at onset (first column), patient age (second column), GAA1 repeats (third column), and GAA2 repeats (fourth column). White stars indicate significant correlations, green stars indicate that for this contrast/VOI combination, additionally significant differences between patients and healthy controls have been found (see Figure 2). Correlations for contrast/VOI combinations, for which no correlation with patient age was found, are marked with a black circle. (b) Scatterplots for the contrasts and VOIs for which significant correlations with disease characteristic, but not with patient age were found [Color figure can be viewed at wileyonlinelibrary.com]

errors despite the automatic and manual corrections applied to the VOIs. The fact that the JHU-ICBM-DTI-81 is in standard space, however, not a standard space template might further be a source of error. For the VBM analysis, no normalization with total intracranial volume was used since the MP2RAGE sequence did not cover the entire brain in all patients. Finally, VBM itself has received critique concerning its reliability and precision (Ashburner & Friston, 2001; Bookstein, 2001; Michael, Evans, & Moore, 2016; Scarpazza, Tognin, Frisciata, Sartori, & Mechelli, 2015).

Although the present results are promising, access to ultrahigh-field MRI is still limited as 7 T MRI scanners are currently more expensive and less available in clinical and research centers, whereas 3 T can routinely be applied. However, 7 T MRI scanners have various advantages in comparison to the routine MRI: Higher contrast-to-noise and signal to-noise ratios at ultra-high field allow for higher possible spatial resolutions, and consequently result in higher sensitivity to tissue changes and anatomical details, which thus produces clearer tissue boundaries and helps visualize small structures in vivo within reasonable scan time (Deistung, Schafer, Schweser, Biedermann, Turner, et al., 2013; Karamat, Darvish-Molla, & Santos-Diaz, 2016; Straub et al., 2019; Thomas et al., 2008). This

study aimed at investigating quantitative MRI parameters which could assist to gain further insight in the mechanisms of the pathology in FRDA. To further evaluate the added value of the 7 T study over 3 T MRI for clinical routine application, a consecutive study is planned in the same study cohort. Replication of the present study results would facilitate their application at clinical field strengths, which is, however, beyond the scope of this study and therefore left to further investigations.

22

I/S

ĥ

.5 21

20

520

480

520

⊈ 500

480

0

0

⊈ 500

00

20 30 age at onset

0

200 400 600

°

500

GAA2 repeats

GAA1 repeats

00

0

0 8

0

10

0

0

RN

600

RN

1000

МІ

0

O

CP

ICP

ICP

C

1000

CONCLUSION 5

It was shown in this study that two independent analyses, a VOIbased evaluation of quantitative MRI data and a VBM analysis, provided overlapping results. Susceptibility mapping provided significant differences between FRDA patients and controls, as well as R_1 , R_2 relaxometry and diffusion imaging for two brain nuclei and three fiber tracts. Moreover, positive results on correlations with disease characteristics were found, indicating that these quantitative MRI parameters could provide more detailed information than volumetric assessment alone, and may assist the search for effective treatments.

DECLARATION OF TRANSPARENCY

The authors, reviewers and editors affirm that in accordance to the policies set by the *Journal of Neuroscience Research*, this manuscript presents an accurate and transparent account of the study being reported and that all critical details describing the methods and results are present.

ACKNOWLEDGMENTS

The use of the ASPIRE gradient echo sequence and corresponding ICE program for coil combination of the 7 T GRE data provided by Korbinian Eckstein and Simon D. Robinson is kindly acknowledged.

CONFLICT OF INTERESTS

The authors report no conflict of interest.

AUTHOR CONTRIBUTIONS

Conceptualization, S.S., S.M., M.E.L., S.B., and E.G.; Methodology, S.S., S.M., M.E.L., S.B., and E.G.; Software, S.S., J.E., and K.S.D.; Validation, S.S., S.M., J.E., K.S.D., M.E.L., S.B., and E.G.; Formal Analysis, S.S., S.M., J.E., and K.S.D.; Investigation, S.S., S.M., E.I., W.N., and E.G.; Resources, S.S., S.M., M.E.L., S.B., and E.G.; Data Curation, S.S., S.M., E.I., W.N., and E.G.; Writing – Original Draft, S.S.; Writing – Review & Editing, S.M., J.E., E.I., W.N., K.S.D., M.E.L., S.B., and E.G.; Visualization, S.S., S.M., J.E., K.S.D., M.E.L., and S.B.; Supervision, M.E.L. and S.B.; Project Administration, S.S., S.M., E.I., W.N., and E.G.

PEER REVIEW

The peer review history for this article is available at https://publo ns.com/publon/10.1002/jnr.24701.

DATA AVAILABILITY STATEMENT

The authors take full responsibility for the data, the analyses and interpretation, and the conduct of the research and have full access to all of the data, of which we have the right to publish any and all data in the absence of a sponsor. Anonymized data, not published in the article, will be shared on reasonable request from a qualified investigator.

ORCID

Sina Straub D https://orcid.org/0000-0002-9167-8778 Stephanie Mangesius https://orcid.org/0000-0001-5790-2724 Julian Emmerich https://orcid.org/0000-0001-5870-8472 Elisabetta Indelicato https://orcid.org/0000-0003-0217-8630 Elke R. Gizewski https://orcid.org/0000-0001-6859-8377

REFERENCES

- Acosta-Cabronero, J., Betts, M. J., Cardenas-Blanco, A., Yang, S., & Nestor, P. J. (2016). In vivo MRI mapping of brain iron deposition across the adult lifespan. *Journal of Neuroscience*, 36(2), 364–374. https://doi.org/10.1523/JNEUROSCI.1907-15.2016
- Aggarwal, M., Kageyama, Y., Li, X., & van Zijl, P. C. (2016). B-0-orientation dependent magnetic susceptibility-induced white matter contrast in the human brainstem at 11.7T. *Magnetic Resonance in Medicine*, 75(6), 2455–2463. https://doi.org/10.1002/mrm.26208

- Akhlaghi, H., Corben, L., Georgiou-Karistianis, N., Bradshaw, J., Storey, E., Delatycki, M. B., & Egan, G. F. (2011). Superior cerebellar peduncle atrophy in Friedreich's ataxia correlates with disease symptoms. *Cerebellum*, 10(1), 81–87. https://doi.org/10.1007/s1231 1-010-0232-3
- Akhlaghi, H., Yu, J., Corben, L., Georgiou-Karistianis, N., Bradshaw, J. L., Storey, E., ... Egan, G. F. (2014). Cognitive deficits in Friedreich ataxia correlate with micro-structural changes in dentatorubral tract. *Cerebellum*, 13(2), 187–198. https://doi.org/10.1007/s1231 1-013-0525-4
- Ashburner, J. (2007). A fast diffeomorphic image registration algorithm. *NeuroImage*, 38(1), 95–113. https://doi.org/10.1016/j.neuro image.2007.07.007
- Ashburner, J. (2010). VBM tutorial. https://www.fil.ion.ucl.ac.uk/~john/ misc/VBMclass10.pdf
- Ashburner, J., & Friston, K. J. (2000). Voxel-based morphometry–The methods. NeuroImage, 11(6), 805–821. https://doi.org/10.1006/ nimg.2000.0582
- Ashburner, J., & Friston, K. J. (2001). Why voxel-based morphometry should be used. *NeuroImage*, 14(6), 1238–1243. https://doi. org/10.1006/nimg.2001.0961
- Blair, I. A., Farmer, J., Hersch, S., Larkindale, J., Lynch, D. R., Napierala, J., ... Subramony, S. H. (2019). The current state of biomarker research for Friedreich's ataxia: A report from the 2018 FARA biomarker meeting. *Future Science OA*, 5(6), FSO398. https://doi.org/10.2144/ fsoa-2019-0026
- Bookstein, F. L. (2001). "Voxel-based morphometry" should not be used with imperfectly registered images. *NeuroImage*, 14(6), 1454–1462. https://doi.org/10.1006/nimg.2001.0770
- Chanraud, S., Zahr, N., Sullivan, E. V., & Pfefferbaum, A. (2010). MR diffusion tensor imaging: A window into white matter integrity of the working brain. *Neuropsychology Review*, 20(2), 209–225. https://doi. org/10.1007/s11065-010-9129-7
- Chung, S., Kim, D., Breton, E., & Axel, L. (2010). Rapid B-1(+) mapping using a preconditioning RF pulse with TurboFLASH readout. *Magnetic Resonance in Medicine*, 64(2), 439–446. https://doi.org/10.1002/ mrm.22423
- Cocozza, S., Costabile, T., Pontillo, G., Lieto, M., Russo, C., Radice, L., ... Sacca, F. (2020). Cerebellum and cognition in Friedreich ataxia: A voxel-based morphometry and volumetric MRI study. *Journal of Neurology*, 267(2), 350–358. https://doi.org/10.1007/s00415-019-09582-9
- da Silva, C. B., Bergo, F. P. G., D'Abreu, A., Cendes, F., Lopes-Cendes, I., & Franca, M. C. (2014). Dentate nuclei T2 relaxometry is a reliable neuroimaging marker in Friedreich's ataxia. *European Journal of Neurology*, 21(8), 1131–1136. https://doi.org/10.1111/ene.12448
- Deistung, A., Schafer, A., Schweser, F., Biedermann, U., Gullmar, D., Trampel, R., ... Reichenbach, J. R. (2013). High-resolution MR imaging of the human brainstem in vivo at 7 Tesla. *Frontiers in Human Neuroscience*, 7, 710. https://doi.org/10.3389/fnhum.2013.00710
- Deistung, A., Schafer, A., Schweser, F., Biedermann, U., Turner, R., & Reichenbach, J. R. (2013). Toward in vivo histology: a comparison of quantitative susceptibility mapping (QSM) with magnitude-, phase-, and R2*-imaging at ultra-high magnetic field strength. *NeuroImage*, 65, 299–314. https://doi.org/10.1016/j.neuroimage.2012.09.055
- Della Nave, R., Ginestroni, A., Tessa, C., Salvatore, E., Bartolomei, F., Salvi, F., ... Mascalchia, M. (2008). Brain white matter tracts degeneration in Friedreich ataxia. An in vivo MRI study using tract-based spatial statistics and voxel-based morphometry. *NeuroImage*, 40(1), 19–25. https://doi.org/10.1016/j.neuroimage.2007.11.050
- Eckstein, K., Dymerska, B., Bachrata, B., Bogner, W., Poljanc, K., Trattnig, S., & Robinson, S. D. (2018). Computationally efficient combination of multi-channel phase data from multi-echo acquisitions (ASPIRE). *Magnetic Resonance in Medicine*, 79(6), 2996–3006. https://doi. org/10.1002/mrm.26963

- Emmerich, J., Flassbeck, S., Schmidt, S., Bachert, P., Ladd, M. E., & Straub, S. (2019). Rapid and accurate dictionary-based T-2 mapping from multi-echo turbo spin echo data at 7 Tesla. *Journal of Magnetic Resonance Imaging*, 49(5), 1253–1262. https://doi.org/10.1002/ jmri.26516
- Gibson, T. J., Koonin, E. V., Musco, G., Pastore, A., & Bork, P. (1996). Friedreich's ataxia protein: Phylogenetic evidence for mitochondrial dysfunction. *Trends in Neurosciences*, 19(11), 465–468. https://doi. org/10.1016/S0166-2236(96)20054-2
- Habas, C., Guillevin, R., & Abanou, A. (2010). In vivo structural and functional imaging of the human rubral and inferior olivary nuclei: A mini-review. *Cerebellum*, 9(2), 167–173. https://doi.org/10.1007/ s12311-009-0145-1
- Hallgren, B., & Sourander, P. (1958). The effect of age on the non-haemin iron in the human brain. *Journal of Neurochemistry*, 3(1), 41–51. https://doi.org/10.1111/j.1471-4159.1958.tb12607.x
- Harkins, K. D., Xu, J. Z., Dula, A. N., Li, K., Valentine, W. M., Gochberg, D. F., ... Does, M. D. (2016). The microstructural correlates of T-1 in white matter. *Magnetic Resonance in Medicine*, 75(3), 1341–1345. https://doi.org/10.1002/mrm.25709
- Heidemann, R. M., Porter, D. A., Anwander, A., Feiweier, T., Heberlein, K., Knosche, T. R., & Turner, R. (2010). Diffusion imaging in humans at 7T using readout-segmented EPI and GRAPPA. *Magnetic Resonance in Medicine*, 64(1), 9–14. https://doi.org/10.1002/mrm.22480
- Andersson, JLR, Jenkinson, M, & Smith, S. (2010). Non-linear registration, aka spatial normalisation. FMRIB Technical Report, TR07JA2.
- Jenkinson, M., & Smith, S. (2001). A global optimisation method for robust affine registration of brain images. *Medical Image Analysis*, 5(2), 143–156. https://doi.org/10.1016/S1361-8415(01)00036-6
- Karamat, M. I., Darvish-Molla, S., & Santos-Diaz, A. (2016). Opportunities and challenges of 7 tesla magnetic resonance imaging: A review. *Critical Reviews in Biomedical Engineering*, 44(1–2), 73–89. https://doi. org/10.1615/CritRevBiomedEng.2016016365
- Koenig, S. H. (1991). Cholesterol of myelin is the determinant of graywhite contrast in MRI of brain. *Magnetic Resonance in Medicine*, 20(2), 285–291. https://doi.org/10.1002/mrm.1910200210
- Koeppen, A. H. (2011). Friedreich's ataxia: Pathology, pathogenesis, and molecular genetics. *Journal of the Neurological Sciences*, 303(1–2), 1–12. https://doi.org/10.1016/j.jns.2011.01.010
- Koeppen, A. H. (2013). Nikolaus Friedreich and degenerative atrophy of the dorsal columns of the spinal cord. *Journal of Neurochemistry*, 126, 4–10. https://doi.org/10.1111/jnc.12218
- Koeppen, A. H., Michael, S. C., Knutson, M. D., Haile, D. J., Qian, J., Levi, S., ... Lamarche, J. B. (2007). The dentate nucleus in Friedreich's ataxia: the role of iron-responsive proteins. *Acta Neuropathologica*, 114(2), 163–173. https://doi.org/10.1007/s00401-007-0220-y
- Ladd, M. E., Bachert, P., Meyerspeer, M., Moser, E., Nagel, A. M., Norris, D. G., ... Zaiss, M. (2018). Pros and cons of ultra-high-field MRI/MRS for human application. *Progress in Nuclear Magnetic Resonance Spectroscopy*, 109, 1–50. https://doi.org/10.1016/j.pnmrs. 2018.06.001
- Lamarche, J. B., Lemieux, B., & Lieu, H. B. (1984). The neuropathology of typical friedreichs ataxia in quebec. *Canadian Journal of Neurological Sciences*, 11(4), 592–600. https://doi.org/10.1017/S031716710 0035113
- Langkammer, C., Krebs, N., Goessler, W., Scheurer, E., Yen, K., Fazekas, F., & Ropele, S. (2012). Susceptibility induced gray-white matter MRI contrast in the human brain. *NeuroImage*, 59(2), 1413–1419. https:// doi.org/10.1016/j.neuroimage.2011.08.045
- Langkammer, C., Schweser, F., Krebs, N., Deistung, A., Goessler, W., Scheurer, E., ... Reichenbach, J. R. (2012). Quantitative susceptibility mapping (QSM) as a means to measure brain iron? A post mortem validation study. *NeuroImage*, 62(3), 1593–1599. https://doi. org/10.1016/j.neuroimage.2012.05.049

- Lee, J., Shmueli, K., Kang, B. T., Yao, B., Fukunaga, M., van Gelderen, P., ... Duyn, J. H. (2012). The contribution of myelin to magnetic susceptibility-weighted contrasts in high-field MRI of the brain. *NeuroImage*, 59(4), 3967–3975. https://doi.org/10.1016/j.neuro image.2011.10.076
- Li, W., Avram, A. V., Wu, B., Xiao, X., & Liu, C. L. (2014). Integrated Laplacian-based phase unwrapping and background phase removal for quantitative susceptibility mapping. NMR in Biomedicine, 27(2), 219–227. https://doi.org/10.1002/nbm.3056
- Li, W., Wu, B., & Liu, C. L. (2011). Quantitative susceptibility mapping of human brain reflects spatial variation in tissue composition. *NeuroImage*, 55(4), 1645–1656. https://doi.org/10.1016/j.neuro image.2010.11.088
- MacKay, A., Laule, C., Vavasour, I., Bjarnason, T., Kolind, S., & Madler, B. (2006). Insights into brain microstructure from the T-2 distribution. *Magnetic Resonance Imaging*, 24(4), 515–525. https://doi. org/10.1016/j.mri.2005.12.037
- Maleike, D., Nolden, M., Meinzer, H. P., & Wolf, I. (2009). Interactive segmentation framework of the Medical Imaging Interaction Toolkit. *Computer Methods and Programs in Biomedicine*, 96(1), 72–83. https:// doi.org/10.1016/j.cmpb.2009.04.004
- Marques, J. P., Kober, T., Krueger, G., van der Zwaag, W., Van de Moortele, P. F., & Gruetter, R. (2010). MP2RAGE, a self bias-field corrected sequence for improved segmentation and T-1-mapping at high field. *NeuroImage*, 49(2), 1271–1281. https://doi.org/10.1016/j.neuro image.2009.10.002
- Mechelli, A., Price, C. J., Friston, K. J., & Ashburner, J. (2005). Voxelbased morphometry of the human brain: Methods and applications. Current Medical Imaging Reviews, 1(2), 105–113. https://doi. org/10.2174/1573405054038726
- Merboldt, K. D., Hanicke, W., & Frahm, J. (1991). Diffusion imaging using stimulated echoes. *Magnetic Resonance in Medicine*, 19(2), 233–239. https://doi.org/10.1002/mrm.1910190208
- Michael, A. M., Evans, E., & Moore, G. J. (2016). Influence of group on individual subject maps in SPM voxel based morphometry. *Frontiers* in Neuroscience, 10, 522. https://doi.org/10.3389/fnins.2016.00522
- Mori, S., Oishi, K., Jiang, H. Y., Jiang, L., Li, X., Akhter, K., ... Mazziotta, J. (2008). Stereotaxic white matter atlas based on diffusion tensor imaging in an ICBM template. *NeuroImage*, 40(2), 570–582. https:// doi.org/10.1016/j.neuroimage.2007.12.035
- Nioche, C., Cabanis, E. A., & Habas, C. (2009). Functional connectivity of the human red nucleus in the brain resting state at 3T. American Journal of Neuroradiology, 30(2), 396–403. https://doi.org/10.3174/ ajnr.A1375
- Nolden, M., Zelzer, S., Seitel, A., Wald, D., Muller, M., Franz, A. M., ... Wolf, I. (2013). The Medical Imaging Interaction Toolkit: Challenges and advances. *International Journal of Computer Assisted Radiology and Surgery*, 8(4), 607–620. https://doi.org/10.1007/s11548-013-0840-8
- Ogg, R. J., & Steen, R. G. (1998). Age-related changes in brain T1 are correlated with iron concentration. *Magnetic Resonance in Medicine*, 40(5), 749–753. https://doi.org/10.1002/mrm.1910400516
- Pagani, E., Ginestroni, A., Della Nave, R., Agosta, F., Salvi, F., De Michele, G., ... Mascalchi, M. (2010). Assessment of brain white matter fiber bundle atrophy in patients with Friedreich ataxia. *Radiology*, 255(3), 882–889. https://doi.org/10.1148/radiol.10091742
- Porter, D. A., & Heidemann, R. M. (2009). High resolution diffusion-weighted imaging using readout-segmented echo-planar imaging, parallel imaging and a two-dimensional navigator-based reacquisition. *Magnetic Resonance in Medicine*, 62(2), 468–475. https://doi. org/10.1002/mrm.22024
- Rezende, T. J. R., Martinez, A. R. M., Faber, I., Girotto Takazaki, K. A., Martins, M. P., de Lima, F. D., ... Franca, M. C. (2019). Developmental and neurodegenerative damage in Friedreich's ataxia. *European Journal* of Neurology, 26(3), 483–489. https://doi.org/10.1111/ene.13843

- Sacca, F., Puorro, G., Antenora, A., Marsili, A., Denaro, A., Piro, R., ... Filla, A. (2011). A combined nucleic acid and protein analysis in Friedreich ataxia: Implications for diagnosis, pathogenesis and clinical trial design. *PLoS ONE*, 6(3), e17627. https://doi.org/10.1371/ journal.pone.0017627
- Scarpazza, C., Tognin, S., Frisciata, S., Sartori, G., & Mechelli, A. (2015). False positive rates in Voxel-based Morphometry studies of the human brain: Should we be worried? *Neuroscience and Biobehavioral Reviews*, 52, 49–55. https://doi.org/10.1016/j.neubi orev.2015.02.008
- Schols, L., Amoiridis, G., Przuntek, H., Frank, G., Epplen, J. T., & Epplen, C. (1997). Friedreich's ataxia–Revision of the phenotype according to molecular genetics. *Brain*, 120, 2131–2140. https://doi.org/10.1093/ brain/120.12.2131
- Selvadurai, L. P., Corben, L. A., Delatycki, M. B., Storey, E., Egan, G. F., Georgiou-Karistianis, N., & Harding, I. H. (2020). Multiple mechanisms underpin cerebral and cerebellar white matter deficits in Friedreich ataxia: The IMAGE-FRDA study. *Human Brain Mapping*, 41(7), 1920–1933. https://doi.org/10.1002/hbm.24921
- Selvadurai, L. P., Harding, I. H., Corben, L. A., & Georgiou-Karistianis, N. (2018). Cerebral abnormalities in Friedreich ataxia: A review. *Neuroscience and Biobehavioral Reviews*, 84, 394–406. https://doi. org/10.1016/j.neubiorev.2017.08.006
- Selvadurai, L. P., Harding, I. H., Corben, L. A., Stagnitti, M. R., Storey, E., Egan, G. F., ... Georgiou-Karistianis, N. (2016). Cerebral and cerebellar grey matter atrophy in Friedreich ataxia: The IMAGE-FRDA study. *Journal of Neurology*, 263(11), 2215–2223. https://doi.org/10.1007/ s00415-016-8252-7
- Smith, S. M. (2002). Fast robust automated brain extraction. *Human Brain* Mapping, 17(3), 143–155. https://doi.org/10.1002/hbm.10062
- Solbach, K., Kraff, O., Minnerop, M., Beck, A., Schols, L., Gizewski, E. R., ... Timmann, D. (2014). Cerebellar pathology in Friedreich's ataxia: Atrophied dentate nuclei with normal iron content. *NeuroImage Clinical*, 6, 93–99. https://doi.org/10.1016/j.nicl.2014.08.018
- Stefanescu, M. R., Dohnalek, M., Maderwald, S., Thurling, M., Minnerop, M., Beck, A., ... Timmann, D. (2015). Structural and functional MRI abnormalities of cerebellar cortex and nuclei in SCA3, SCA6 and Friedreich's ataxia. *Brain*, 138, 1182–1197. https://doi.org/10.1093/ brain/awv064
- Straub, S., Knowles, B. R., Flassbeck, S., Steiger, R., Ladd, M. E., & Gizewski, E. R. (2019). Mapping the human brainstem: Brain nuclei and fiber tracts at 3 T and 7 T. NMR in Biomedicine, 32(9), e4118. https://doi.org/10.1002/nbm.4118
- Straub, S., Schneider, T. M., Emmerich, J., Freitag, M. T., Ziener, C. H., Schlemmer, H. P., ... Laun, F. B. (2017). Suitable reference tissues for quantitative susceptibility mapping of the brain. *Magnetic Resonance in Medicine*, 78(1), 204–214. https://doi.org/10.1002/ mrm.26369
- Tanner, J. E. (1970). Use of stimulated echo in NMR-diffusion studies. Journal of Chemical Physics, 52(5), 2523–2526. https://doi. org/10.1063/1.1673336
- Theysohn, J. M., Kraff, O., Maderwald, S., Barth, M., Ladd, S. C., Forsting, M., ... Gizewski, E. R. (2011). 7 Tesla MRI of microbleeds and white matter lesions as seen in vascular dementia. *Journal of Magnetic Resonance Imaging*, 33(4), 782–791. https://doi.org/10.1002/jmri.22513
- Thomas, B. P., Welch, E. B., Niederhauser, B. D., Whetsell, W. O., Jr., Anderson, A. W., Gore, J. C., ... Creasy, J. L. (2008). High-resolution 7T MRI of the human hippocampus in vivo. *Journal of Magnetic Resonance Imaging*, 28(5), 1266–1272. https://doi.org/10.1002/ jmri.21576
- Waldvogel, D., van Gelderen, P., & Hallett, M. (1999). Increased iron in the dentate nucleus of patients with Friedrich's ataxia. Annals of Neurology, 46(1), 123–125.
- Ward, P. G. D., Harding, I. H., Close, T. G., Corben, L. A., Delatycki, M. B., Storey, E., ... Egan, G. F. (2019). Longitudinal evaluation of iron

concentration and atrophy in the dentate nuclei in Friedreich ataxia. *Movement Disorders*, 34(3), 335–343. https://doi.org/10.1002/mds.27606

- Wei, H. J., Dibb, R., Zhou, Y., Sun, Y. W., Xu, J. R., Wang, N., & Liu, C. L. (2015). Streaking artifact reduction for quantitative susceptibility mapping of sources with large dynamic range. *NMR in Biomedicine*, 28(10), 1294–1303. https://doi.org/10.1002/nbm.3383
- Weigel, M. (2015). Extended phase graphs: Dephasing, RF pulses, and echoes–Pure and simple. *Journal of Magnetic Resonance Imaging*, 41(2), 266–295. https://doi.org/10.1002/jmri.24619
- Wu, B., Li, W., Avram, A. V., Gho, S. M., & Liu, C. (2012). Fast and tissue-optimized mapping of magnetic susceptibility and T2* with multi-echo and multi-shot spirals. *NeuroImage*, 59(1), 297–305. https://doi.org/10.1016/j.neuroimage.2011.07.019
- Wu, B., Li, W., Guidon, A., & Liu, C. L. (2012). Whole brain susceptibility mapping using compressed sensing. *Magnetic Resonance in Medicine*, 67(1), 137-147. https://doi.org/10.1002/mrm.23000
- Zeineh, M. M., Parvizi, J., Balchandani, P., Liu, C., Gary, G., Sawyer, A., ... Atlas, S. W. (2009). Ultra-high resolution7.0T MRI of medial temporal lobe epilepsy. Proceedings of the International Society of Magnetic Resonance in Medicine, 17, 3499.
- Zhang, Y., Lyver, E. R., Knight, S. A. B., Pain, D., Lesuisse, E., & Dancis, A. (2006). Mrs3p, Mrs4p, and frataxin provide iron for Fe-S cluster synthesis in mitochondria. *Journal of Biological Chemistry*, 281(32), 22493–22502. https://doi.org/10.1074/jbc.M604246200
- Zhang, Y., Wei, H., Cronin, M. J., He, N., Yan, F., & Liu, C. (2018). Longitudinal data for magnetic susceptibility of normative human brain development and aging over the lifespan. *Data in Brief*, 20, 623-631. https://doi.org/10.1016/j.dib.2018.06.005
- Zhang, Y. Y., Brady, M., & Smith, S. (2001). Segmentation of brain MR images through a hidden Markov random field model and the expectation-maximization algorithm. *IEEE Transactions on Medical Imaging*, 20(1), 45–57. https://doi.org/10.1109/42.906424

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

Table S1 All Spearman correlation coefficients ρ and p-values for Figure 5 are provided when applicable for all investigated brain structures, middle cerebellar peduncle (MCP), pontine crossing tract (PCT), corticospinal tract (CS), medial lemniscus (ML), inferior cerebellar peduncle (ICP), superior cerebellar peduncle (SCP), cerebral peduncle (CP), posterior thalamic radiation (PTR; including optic radiation), sagittal stratum (SS; including inferior longitudinal fasciculus and inferior fronto-occipital fasciculus), and manually drawn volumes of interest for substantia nigra (SN), red nuclei (RN), and dentate nuclei (DN) and all contrasts, susceptibility mapping (QSM), R_1 and R_2 relaxometry, and fractional anisotropy (FA). Statistically significant findings are marked green

Transparent Peer Review Report

Transparent Science Questionnaire for Authors

How to cite this article: Straub S, Mangesius S, Emmerich J, et al. Toward quantitative neuroimaging biomarkers for Friedreich's ataxia at 7 Tesla: Susceptibility mapping, diffusion imaging, R_2 and R_1 relaxometry. *J Neurosci Res.* 2020;98:2219–2231. https://doi.org/10.1002/jnr.24701