## Heliyon



Received: 27 February 2018 Revised: 29 July 2018 Accepted: 4 September 2018

Cite as: Hiroyuki Miwa, Nobuyuki Itoh. Unknown genes, *Cebelin* and *Cebelinlike*, predominantly expressed in mouse brain. Heliyon 4 (2018) e00773. doi: 10.1016/j.heliyon.2018. e00773



# Unknown genes, *Cebelin* and *Cebelin-like*, predominantly expressed in mouse brain

#### Hiroyuki Miwa<sup>\*,1</sup>, Nobuyuki Itoh

Department of Genetic Biochemistry, Kyoto University Graduate School of Pharmaceutical Sciences, Sakyo, Kyoto 606-8501, Japan

\* Corresponding author.

E-mail address: hiroyukimiwa1026@gmail.com (H. Miwa).

<sup>1</sup> Present address: Department of Pathophysiology, Tokyo University of Pharmacy and Life Sciences, 1432-1 Horinouchi, Hachioji, Tokyo 192-0392, Japan.

### Abstract

We identified two genes, *Cebelin* and *Cebelin-like*, encoding unknown proteins in mice. Cebelin and Cebelin-like consist of 168 and 167 amino acids with putative secreted signal sequences. However, Cebelin and Cebelin-like are cellular proteins not secreted proteins. *Cebelin* and *Cebelin-like* were predominantly expressed in the brain among major tissues examined. The expression of *Cebelin* in the brain was predominantly detected in the internal granule layer of the cerebellum.

Keywords: Developmental biology, Biochemistry, Molecular biology, Neuroscience

#### 1. Introduction

Proteins with putative secreted signal sequences are mostly secreted or membrane proteins. Secreted proteins potentially play crucial roles as extracellular signaling molecules in cell proliferation, differentiation, and function. The identification and characterization of unknown genes encoding secreted proteins potentially provide new insights into morphogenesis, metabolism, and disease (Klee et al., 2004;

### Α

mCebelin	MTAGTVVITGGILATVILLCIIAVLCYCRLQYYCCK	36
hCEBELIN	MTAGTVVITGGILATVILLCIIAVLCYCRLQYYCCK	36
KGTDGEDAE	EEEEEEHGLSIHPRVPACNACSSHVLDGRGGLAPL	81
* * KSGTEVA	**********************DEEEEREHDLPTHPRGPTCNACSSQALDGRGSLAPL	79
TSESCSQPC	GV-ASHCTTCSPYRTPFYIRTADMVPNGGGGERLSF	125
*** ***** TSEPCSQPC	** ********** ************************	124
APTHYKEGG	TPSLKLAAPQNYPVTWPSSGHEAFTNPRAISTDV	168
*** ***** APTYYKEGG	********* ****** ** ******************	167



2 https://doi.org/10.1016/j.heliyon.2018.e00773 2405-8440/© 2018 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/). Kassai et al., 2005; Wakahara et al., 2007; Koike et al., 2007; Miwa et al., 2009; Miyake et al., 2009; Ohta et al., 2015). Additionally, genes expressed by specific cells could become useful markers in developmental biology (Miwa and Era, 2015, 2016, 2018). We identified mouse cDNAs encoding unknown proteins with putative secreted signal sequences but not putative transmembrane domains from GenBank. We termed one of them *Cebelin*, which is also referred to as *Fam163a*, as the gene was predominantly expressed in the cerebellum.

### 2. Results and discussion

The full-length cDNA was cloned by polymerase chain reaction (PCR) with mouse brain cDNA as a template. Cebelin protein consists of 168 amino acids (AAs) with a putative secreted signal sequence (30 AAs) at its amino terminus but not putative transmembrane domains (GenBank accession code NM\_177838) (Fig. 1A). Cebelin is a unique protein with no known functional motifs and no primary structure similarity to known functional proteins. Human *CEBELIN* cDNA was also identified by a homology-based search from GenBank. The AA sequence of human CEBELIN (167 AAs) with a putative secreted signal sequence (30 AAs) was highly similar (~85% AA identity) to that of mouse Cebelin (Fig. 1A). The coding region of *Cebelin* is divided with a single intron (data not shown). Mouse *Cebelin* is closely linked to *Tor1aip1*, *Toriaip2*, *Tdrd5* and *Nphs2* on chromosome 1 at q25.2-25.3, supporting that human *CEBELIN* is a human ortholog of mouse *Cebelin* (Fig. 1B).

To examine whether Cebelin is a secreted protein, Myc and His<sub>6</sub> tags-fused Cebelin was overexpressed in mammalian cells, COS-7 cells. Both the medium and lysate of the cultured cells were examined by Western blotting using anti-Myc tag antibody. We could detect no bands in the medium or lysate of the control. A band was detected in the lysate but not the medium of the *Cebelin*-overexpressed cells, indicating that Cebelin is a cellular protein but not a secreted protein (Fig. 2A). This result was discrepant from the previous study (Vasudevan et al., 2009). The observed molecular mass (~25 kDa) was larger than the calculated molecular mass of the recombinant Cebelin protein (~20.5 kDa), indicating that Cebelin protein might be subjected to post-translational modification.

Fig. 1. Molecular analysis of *Cebelin*. A: Comparison of AA sequences of mouse Cebelin and human CEBELIN. The numbers refer to AA positions of mouse Cebelin and human CEBELIN. Asterisks represent identical residues of the sequences. Underlines represent putative secreted signal sequences. Dashes represent introduced gaps to align the sequences. B: Syntenic relationship between mouse chromosome 1G3 and human chromosome 1q25.2-25.3. The mouse *Cebelin* and human *CEBELIN* genes are closely linked to the mouse *Torlaip1*, *Torlaip2*, *Tdrd5*, or *Nphs2* genes and human *TORLAIP1*, *TORLAIP2*, *TDRD5*, or *NPHS2* genes, respectively. mb, megabase.







We also examined the cellular localization of Cebelin in the cells by immunocytochemical analysis using anti-Myc tag antibody. No signals were detected in the control. In contrast, Cebelin was widely detected in the *Cebelin*-overexpressed cells. Cebelin was most intensely co-localized with Mannosidase II, a marker protein for the Golgi apparatus (Moremen and Touster, 1986), indicating that Cebelin was most intensely detected in the Golgi apparatus (Fig. 2B). Cebelin is a cellular protein with a putative secreted signal sequence. As hydrophobic segments at the amino termini were reported to potentially function as type II membrane protein signal anchors (Yokoyama-Kobayashi et al., 1999), the putative secreted signal sequence in Cebelin might function as the type II signal anchor.

The expression of *Cebelin* was examined in adult mouse tissues (postnatal day 56, P56) by reverse transcription (RT)-PCR using the specific primers for *Cebelin*. Although all the tissues examined expressed  $\beta$ -*Actin* (Tokunaga et al., 1986), the expression of *Cebelin* was predominantly detected in the brain (Fig. 3A). We also examined the expression of *Cebelin* in the brain at respective developmental stages (embryonic day 12.5, E12.5-P56). The expression of *Cebelin* was more abundantly detected in the postnatal brain than the embryonic brain (Fig. 3B).

The expression of *Cebelin* was also examined in the adult brain by in situ hybridization using the antisense *Cebelin* RNA probe. Essentially we could detect no grains



**Fig. 3.** Expression of *Cebelin* in adult mouse tissues and brain at respective developmental stages, and localization of *Cebelin* in adult mouse brain. A: The expression of *Cebelin* was examined in adult mouse tissues (P56) by RT-PCR.  $\beta$ -Actin was a control. The expected sizes of *Cebelin* and  $\beta$ -Actin cDNA are 574 and 408 base pairs, respectively. Fig. S2A and B are full images of the gels. B: The expression of *Cebelin* was examined in mouse brain at respective developmental stages (E12.5-P56) by RT-PCR. C: The localization of *Cebelin* was examined in adult mouse brain (P56) by in situ hybridization using the sense (a, c, e, g, i, k, m, o, q) or antisense (b, d, f, h, j, l, n, p, r) *Cebelin* RNA probe. Black grains show the location of *Cebelin*. The sections of the brain were counterstained with cresyl-violet (a'-r'). Scale bars = 5 mm.

on any sections with the sense probe as a control. In contrast, the expression of *Cebelin* shown by black grains was predominantly detected in the internal granule layer of the cerebellum with the antisense probe (Fig. 3C). However, the expression of *Cebelin* was not significantly detected in any other region of the brain.

Furthermore, we identified mouse cDNA encoding another unknown protein of 167 AAs (GenBank accession code NM\_175427) (Fig. 4). As the protein is significantly similar ( $\sim$ 43% AA identity) to Cebelin, we named it Cebelin-like, which is also referred to as *Fam163b*. Human *CEBELIN-LIKE* cDNA was also identified. The AA sequence of human CEBELIN-LIKE (166 AAs) was highly similar ( $\sim$ 90% identity) to that of mouse Cebelin-like (Fig. 4).

Cebelin-like was overexpressed in CHO-S cells in the same way as Cebelin was. Both the medium and lysate of the cultured cells were examined by Western blotting. The result indicates that Cebelin-like is also a cellular protein, whereas Brorin-like is a secreted protein as described previously (Miwa et al., 2009) (Fig. 5A).

To examine the cellular localization of Cebelin-like in the cells, a green fluorescent protein (GFP)-fused Cebelin-like was overexpressed in COS-7 cells. In the result,

mCbn	MTAGTVVI	TGGILATV	ILLCIIAV	′LCYCRLQ	YYCCKKGTD	40
	******	******	******	*****	*****	
mCbl	MTAGTVVI	TGGILATV	ILLCIIAV	LCYCRLQ	YYCCKK	37
	******	******	******	******	*****	
hCBL	MTAGTVVI	TGGILATV	ILLCIIAV	'LCYCRLQ	YYCCKK	37
GEDAI	EEEEEEEF	IGLSIHPRV	PACNACSS	HVLDGRG	GLAPLTSES	85
4	* ** ***	*	×	**	* * *	
DH	ESEEDEEEF	DFAVHSHL	PPLHSNRN	ILVLTNGP	ALYPAATTS	79
*	*******	******	******	******	**** * **	
DH	ESEEDEEEF	PDFAVHSHL	PPLHSNRN	ILVLTNGP	ALYPTASTS	79
CSQP	CGVA-SHCI	TCSPYRTP	-FYIRTA-	DMVPN	GGGGERLSF	125
**	* *	** * *	¥	*	** *	
FSQKS	SPQARALCF	RSCSHYEPP	TFFLQEPE	DEDFEGV	RNGGGRVAY	124
****	*******	**** ***	******	* * *	*** ** *	
FSQKS	SPQARALCF	RSCSHCEPP	TFFLQEPF	PEEE-EDV	LNGGERVLY	123
APTHY	YKEGGTPSI	.KLAAPQNY	PVTWPSSO	HEAFTNP	RAISTDV	168
	**	*	*	***	* ****	
KSIS	QEDVELPSA	SFGGLQAL	NPNRLSAM	IREAFSRS	RSVSTDV	167
** **	*****	*****	******	**** **	** ****	
KSVS	QEDVELPPG	GFGGLQAL	NPNRLSAM	IREAFARS	RSISTDV	166

Fig. 4. Molecular analysis of *Cebelin-like*. Comparison of AA sequences of mouse Cebelin-like (mCbl), human CEBELIN-LIKE (hCBL), and mouse Cebelin (mCbn). The numbers refer to AA positions of mouse Cebelin-like, human CEBELIN-LIKE, and mouse Cebelin. Asterisks represent identical residues of the sequences. Dashes represent introduced gaps to align the sequences.

Heliyon







**Fig. 5.** Detection of recombinant Cebelin-like. A: CHO-S cells were transfected with the recombinant Cebelin-expression vector (Cbn) (a, b), the recombinant Cebelin-like-expression vector (Cbl) (c, d) or the recombinant Brorin-like-expression vector (Brl) (e, f), which was a control. The lysate (a, c, e) and medium (b, d, f) of the transfected CHO-S cells were examined by Western blotting using anti-Myc tag antibody. Fig. S3A and B are full images of the blots. B: COS-7 cells transfected with the GFP-fused Cebelin-expression vector (*Cbn*) (a–d, i–l) or the GFP-fused Cebelin-like-expression vector (*Cbn*) (a–d, i–l) or the GFP-fused Cebelin-like-expression vector (*Cbl*) (e–h, m–p) were examined by immunocytochemical using anti-EEA1 antibody for the endosome or anti-GRP78 antibody for the endoplasmic reticulum. The signals obtained by GFP (a, e, i, m), Hoechst (b, f, j, n), and immunocytochemical using anti-EEA1 antibody (c, g) or anti-GRP78 antibody (k, o) were merged (d, h, l, p). Scale bar = 50  $\mu$ m.



**Fig. 6.** Expression of *Cebelin-like* in adult mouse tissues and brain at respective developmental stages. The expression of *Cebelin-like* was examined in adult mouse tissues (P56) and brain at respective developmental stages (E12.5-P56) by RT-PCR.  $\beta$ -Actin was a control. The expected sizes of *Cebelin-like* and  $\beta$ -Actin cDNA are 543 and 408 base pairs, respectively. Fig. S4A and B are full images of the gels.

the localization of Cebelin-like was similar to that of Cebelin and only partly overlapped EEA1, a marker protein for the endosome (Mu et al., 1995), or GRP78, the endoplasmic reticulum (Kozutsumi et al., 1988) (Fig. 5B).

The expression of *Cebelin-like* was examined in the embryonic brains and adult tissues by RT-PCR. The expression profiles of *Cebelin-like* are also similar to those of *Cebelin* (Fig. 6).

In conclusion, we identified two genes, *Cebelin* and *Cebelin-like*, encoding unknown proteins in mice and human. Both Cebelin and Cebelin-like are cellular proteins not secreted proteins and predominantly expressed in the brain. The present findings indicate that *Cebelin* and *Cebelin-like* are unknown genes encoding cellular proteins that potentially play roles in the cerebellum.

### 3. Experimental

### 3.1. Mice

The Animal Research Committee of Kyoto University Graduate School of Pharmaceutical Sciences approved all study protocols. All mice were purchased from Shimizu Laboratory Supplies.

### **3.2.** Identification of Cebelin and Cebelin-like in mice and humans

AA sequences predicted from mouse cDNAs of unknown function in nucleotide sequence databases were randomly analyzed using PSORT. The cDNAs encoding putative secreted proteins were identified and cloned in pGEM-T Easy vector (Promega). We named two of the cDNAs mouse *Cebelin* and *Cebelin-like*. Human

*CEBELIN* or *CEBELIN-LIKE* cDNA was also identified in a homology-based search of human cDNA sequences in nucleotide sequence databases with the AA sequence of mouse Cebelin or Cebelin-like.

### **3.3.** Forced expression of Cebelin or Cebelin-like cDNA in COS-7 cells and CHO-S cells

The *Cebelin* or *Cebelin-like* cDNA with a DNA fragment encoding a Myc tag and a  $His_6$  tag or a GFP at the 3' terminus of the coding region was constructed in pcDNA3.1(+) vector (Thermo Fisher Scientific).

COS-7 cells and CHO-S cells were transfected with the respective vectors using Lipofectamine 2000 (Thermo Fisher Scientific) and cultured at 37  $^{\circ}$ C in a humidified atmosphere of 5% CO<sub>2</sub> in air.

### 3.4. Detection of recombinant Cebelin or Cebelin-like protein

For Western blotting, the samples were separated by sodium dodecyl sulfatepolyacrylamide gel electrophoresis (SDS-PAGE) under reducing conditions and transferred onto Hybond-ECL (GE Healthcare). The recombinant proteins were detected using mouse monoclonal anti-Myc tag antibody (Cell Signaling Technology) (1:500) as primary antibody and HRP-conjugated rabbit anti-mouse IgG antibody (Thermo Fisher Scientific) (1:1,000) as secondary antibody. Immunoreactive bands were visualized using an enhanced chemiluminescence detection system (PerkinElmer) as described (Yamashita et al., 2002).

To detect Cebelin by immunocytochemical analysis, mouse monoclonal anti-Myc tag antibody and FITC-conjugated goat anti-mouse IgG (Sigma-Aldrich) were used as primary and secondary antibodies, respectively. To detect Mannosidase II, EEA1, and GRP78, rabbit anti-Mannosidase II antibody, anti-EEA1 antibody, and anti-GRP78 antibody (Abcam) and TRITC-conjugated goat anti-rabbit antibody (Sigma-Aldrich) were used as primary and secondary antibodies, respectively.

### 3.5. **RT-PCR**

Total RNA was purified with RNeasy Mini kit (Qiagen) and transcribed to DNA using M-MLV Reverse Transcriptase (Thermo Fisher Scientific). The cDNAs were amplified with Gene *Taq* NT (Nippon Gene) and the specific primers, which were listed in Table 1. DNA fragments were detected by agarose gel electrophoresis.

### 3.6. In situ hybridization

Mouse brain at P56 was frozen in O.C.T. compound (Sakura Finetek), and sections were cut at 10  $\mu$ m. A <sup>35</sup>S-labeled sense or antisense RNA probe was transcribed from

Table 1. Primers for RT-PCR

Gene	Sequence (forward)	Sequence (reverse)
Cebelin	5'-ATACATCTTTGCAGAGTTTGATGG-3'	5'-TGTTGGGTGTGTGCAGATTGG-3'
Cebelin-like	5'-AGGCTGTTGATGGAGAAGTGG-3'	5'-AGGATAGAGGCCTGTCACACG-3'
$\beta$ -Actin	5'-CAGAGCAAGAGAGGTATCCT-3'	5'-CGGTCAGGATCTTCATGAGG-3'

*Cebelin* cDNA clone. The signals were visualized by autoradiography using BioMax MR (Carestream) as described (Yazaki et al., 1994). The sections of mouse brain were counterstained with cresyl-violet (Nissl staining).

### Declarations

### Author Contribution statement

Hiroyuki Miwa: Conceived and designed the experiments; Performed the experiments; Analyzed and interpreted the data; Contributed reagents, materials, analysis tools or data; Wrote the paper.

Nobuyuki Itoh: Conceived and designed the experiments; Analyzed and interpreted the data; Contributed reagents, materials, analysis tools or data; Wrote the paper.

### **Funding statement**

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

### **Competing interest statement**

The authors declare no conflict of interest.

### **Additional information**

Supplementary content related to this article has been published online at https://doi. org/10.1016/j.heliyon.2018.e00773.

### References

Kassai, Y., Munne, P., Hotta, Y., Penttilä, E., Kavanagh, K., Ohbayashi, N., Takada, S., Thesleff, I., Jernvall, J., Itoh, N., 2005. Regulation of mammalian tooth cusp patterning by ectodin. Science 309, 2067–2070.

Klee, E.W., Carlson, D.F., Fahrenkrug, S.C., Ekker, S.C., Ellis, L.B., 2004. Identifying secretomes in people, pufferfish and pigs. Nucleic Acids Res. 32, 1414–1421. Koike, N., Kassai, Y., Kouta, Y., Miwa, H., Konishi, M., Itoh, N., 2007. Brorin, a novel secreted bone morphogenetic protein antagonist, promotes neurogenesis in mouse neural precursor cells. J. Biol. Chem. 282, 15843–15850.

Kozutsumi, Y., Segal, M., Normington, K., Gething, M.J., Sambrook, J., 1988. The presence of malfolded proteins in the endoplasmic reticulum signals the induction of glucose-regulated proteins. Nature 332, 462–464.

Miwa, H., Era, T., 2015. Generation and characterization of PDGFRα-GFPCreERT2 knock-in mouse line. Genesis 53, 329–336.

Miwa, H., Era, T., 2016. Mesoderm differentiation from hiPS cells. Methods Mol. Biol. 1357, 403–413.

Miwa, H., Era, T., 2018. Tracing the destiny of mesenchymal stem cells from embryo to adult bone marrow and white adipose tissue via Pdgfra expression. Development 145, dev155879.

Miwa, H., Miyake, A., Kouta, Y., Shimada, A., Yamashita, Y., Nakayama, Y., Yamauchi, H., Konishi, M., Itoh, N., 2009. A novel neural-specific BMP antagonist, Brorin-like, of the Chordin family. FEBS Lett. 583, 3643–3648.

Miyake, A., Takahashi, Y., Miwa, H., Shimada, A., Konishi, M., Itoh, N., 2009. Neucrin is a novel neural-specific secreted antagonist to canonical Wnt signaling. Biochem. Biophys. Res. Commun. 390, 1051–1055.

Moremen, K.W., Touster, O., 1986. Topology of mannosidase II in rat liver Golgi membranes and release of the catalytic domain by selective proteolysis. J. Biol. Chem. 261, 10945–10951.

Mu, F.T., Callaghan, J.M., Steele-Mortimer, O., Stenmark, H., Parton, R.G., Campbell, P.L., McCluskey, J., Yeo, J.P., Tock, E.P., Toh, B.H., 1995. EEA1, an early endosome-associated protein. EEA1 is a conserved alpha-helical peripheral membrane protein flanked by cysteine "fingers" and contains a calmodulin-binding IQ motif. J. Biol. Chem. 270, 13503–13511.

Ohta, H., Konishi, M., Kobayashi, Y., Kashio, A., Mochiyama, T., Matsumura, S., Inoue, K., Fushiki, T., Nakao, K., Kimura, I., Itoh, N., 2015. Deletion of the neurotrophic factor neudesin prevents diet-induced obesity by increased sympathetic activity. Sci. Rep. 5, 10049.

Tokunaga, K., Taniguchi, H., Yoda, K., Shimizu, M., Sakiyama, S., 1986. Nucleotide sequence of a full-length cDNA for mouse cytoskeletal beta-actin mRNA. Nucleic Acids Res. 14, 2829.

Vasudevan, S.A., Shang, X., Chang, S., Ge, N., Diaz-Miron, J.L., Russell, H.V., Hicks, M.J., Ludwig, A.D., Wesson, C.L., Burlingame, S.M., Kim, E.S.,

Khan, J., Yang, J., Nuchtern, J.G., 2009. Neuroblastoma-derived secretory protein is a novel secreted factor overexpressed in neuroblastoma. Mol. Cancer Ther. 8, 2478–2489.

Wakahara, T., Kusu, N., Yamauchi, H., Kimura, I., Konishi, M., Miyake, A., Itoh, N., 2007. Fibin, a novel secreted lateral plate mesoderm signal, is essential for pectoral fin bud initiation in zebrafish. Dev. Biol. 303, 527–535.

Yamashita, T., Konishi, M., Miyake, A., Inui, K., Itoh, N., 2002. Fibroblast growth factor (FGF)-23 inhibits renal phosphate reabsorption by activation of the mitogenactivated protein kinase pathway. J. Biol. Chem. 277, 28265–28270.

Yazaki, N., Hosoi, Y., Kawabata, K., Miyake, A., Minami, M., Satoh, M., Ohta, M., Kawasaki, T., Itoh, N., 1994. Differential expression patterns of mRNAs for members of the fibroblast growth factor receptor family, FGFR-1 FGFR-4, in rat brain. J. Neurosci. Res. 37, 445–452.

Yokoyama-Kobayashi, M., Yamaguchi, T., Sekine, S., Kato, S., 1999. Selection of cDNAs encoding putative type II membrane proteins on the cell surface from a human full-length cDNA bank. Gene 228, 161–167.