



Draft Genome Sequences of Four Aspergillus Section Fumigati Clinical Strains

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ABSTRACT Aspergillus fungi in section Fumigati include important human pathogens. Here, we sequenced the genomes of two strains of Aspergillus hiratsukae and two strains of Aspergillus felis. The average genome sizes are 29.5 Mb for A. hiratsukae and 31.8 Mb for A. felis.

A spergillus is a highly diverse genus of industrially and medically important fungi (1, 2). The genus is taxonomically divided into 27 sections (3). Section *Fumigati* contains the major human pathogen *Aspergillus fumigatus* (4) and several so-called cryptic species, such as *Aspergillus hiratsukae* and *Aspergillus felis* (5–7), which are morphologically similar but genetically distinct from *A. fumigatus*. Cryptic species account for over 10% of cases of *Aspergillus* infection (8). Here, we sequenced the genomes of two clinical strains of *A. hiratsukae*, CNM-CM5793 and CNM-CM6106, from nail and ear infections, respectively, both from Spain. We also sequenced two clinical strains of *A. felis*, strain CNM-CM7691 from an ear infection in Spain and strain CNM-CM5623 from Portugal. All four isolates were recovered from clinical samples following standard procedures and sent to the Medical Mycology Reference Laboratory (at the National Center for Microbiology, Instituto de Salud Carlos III, Madrid, Spain) for identification and susceptibility testing. Except for infection type, no clinical data were recorded. Therefore, the fungal isolates were judged to be exempt from informed consent of the patients and institutional review board approval.

Species assignment was based on a maximum-likelihood phylogenetic analysis (Fig. 1). For genome sequencing, we grew all strains in glucose-yeast extract-peptone (GYEP) liquid medium (0.3% yeast extract and 1% peptone; Difco, Soria Melguizo) with 2% glucose (Sigma-Aldrich, Spain) for 24 to 48 h at 30°C. The mycelium was mechanically disrupted by vortex mixing with glass beads and used to extract genomic DNA using the phenol-chloroform method (9). DNA was quantified using the QuantiFluor double-stranded DNA (dsDNA) system and the QuantiFluor ST fluorometer (Promega, Madison, WI, USA). DNA quality was checked with the Agilent 2100 bioanalyzer (Agilent Technologies, Inc., Santa Clara, CA, USA). DNA libraries were prepared using the Nextera DNA library prep kit (Illumina, Inc., San Diego, CA, USA) according to the manufacturer's guidelines. Paired-end sequencing (2×150 bp) was performed using the NextSeq 500 platform following the manufacturer's protocols (Illumina, Inc.).

For all software, default parameters were used except where otherwise noted. The numbers of sequencing read pairs generated for strains CNM-CM5793, CNM-CM6106, CNM-CM7691, and CNM-CM5623 were 7,733,508, 5,237,901, 9,555,248, and 6,768,577, respectively. Quality control of the sequence reads was performed with FastQC v0.11.7 (https://www.bioinformatics.babraham.ac.uk/projects/fastqc/). Raw reads were cleaned with Trimmomatic v0.38 (10) with the following parameters: NexteraPE-PE.fa:2:30:10:2:

Citation dos Santos RAC, Rivero-Menendez O, Steenwyk JL, Mead ME, Goldman GH, Alastruey-Izquierdo A, Rokas A. 2020. Draft genome sequences of four *Aspergillus* section *Fumigati* clinical strains. Microbiol Resour Announc 9:e00856-20. https://doi.org/10.1128/ MRA.00856-20.

Editor Vincent Bruno, University of Maryland School of Medicine

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Received 25 August 2020 Accepted 9 September 2020 Published 1 October 2020



FIG 1 Maximum-likelihood phylogenetic tree of the four strains sequenced in this announcement (in bold) and related species in section *Fumigati*, based on the analysis of the four markers beta-tubulin gene (*benA*), calmodulin gene (*CaM*), actin gene (*act*), and RNA polymerase II second-largest subunit gene (*RPB2*), commonly used in *Aspergillus* taxonomy (18); sequences were obtained from reference 18 except for the sequences of the four newly sequenced strains, which were obtained by searching for markers of each strain in orthogroups generated by OrthoFinder v2.3.3 (19) using *A. fumigatus* Af293 (17) as the reference. Each marker was aligned with MAFFT v7.397 (20), and a supermatrix was generated with FASconCAT v1.11 (21). Tree inference was carried out on IQ-TREE v2.0.3 (22) with partitions with the option "MFP+MERGE," which employs ModelFinder to find the best partition scheme. The final tree was edited in FigTree v1.4.4 (http://tree.bio.ed.ac.uk/software/figtree/). Support values are based on 1,000 bootstrap replicates. *A. clavatus* (section *Clavati*) was used to root the tree. Note that the species *A. parafelis*, *A. pseudofelis*, and *A. felis* (18); thus, we infer that the two sequenced strains belong to *A. felis*.

keepBothReads, SLIDINGWINDOW:4:15, LEADING:3, TRAILING:3, and MINLEN:90. The genome sequences were assembled with SPAdes v3.14.0 (11) employing multiple k-mers (31, 41, 51, 61, 71, 81, and 91) and the --careful parameter. The genomic reads were mapped to the assembly with Bowtie v2.3.4.1 (12), followed by a single iteration

						No. (%) of complete	No. (%) of
Assembly	No. of contigs	Avg genome	GC content	N ₅₀	No. of	and single-copy	fragmented
size ^a (bp)	>1,000 bp	coverage (×)	(%)	(bp)	genes	BUSCOs	BUSCOs
29,562,918	745	63	50.38	100,935	9,685	3,487 (98.33)	31 (0.87)
29,374,270	922	39	50.37	71,695	9,663	3,466 (97.74)	38 (1.07)
31,643,783	663	47	49.93	112,776	10,161	3,494 (98.53)	26 (0.73)
31,957,614	559	70	49.93	138,232	10,243	3,503 (98.78)	18 (0.51)
	Assembly size ^a (bp) 29,562,918 29,374,270 31,643,783 31,957,614	Assembly size ^a (bp) No. of contigs >1,000 bp 29,562,918 745 29,374,270 922 31,643,783 663 31,957,614 559	Assembly size ^a (bp) No. of contigs >1,000 bp Avg genome coverage (×) 29,562,918 745 63 29,374,270 922 39 31,643,783 663 47 31,957,614 559 70	Assembly size ^a (bp) No. of contigs >1,000 bp Avg genome coverage (x) GC content (%) 29,562,918 745 63 50.38 29,374,270 922 39 50.37 31,643,783 663 47 49.93 31,957,614 559 70 49.93	Assembly size ^a (bp) No. of contigs >1,000 bp Avg genome coverage (x) GC content (%) N ₅₀ (bp) 29,562,918 745 63 50.38 100,935 29,374,270 922 39 50.37 71,695 31,643,783 663 47 49.93 112,776 31,957,614 559 70 49.93 138,232	Assembly size ^a (bp) No. of contigs >1,000 bp Avg genome coverage (×) GC content (%) No. of (bp) No. of genes 29,562,918 745 63 50.38 100,935 9,685 29,374,270 922 39 50.37 71,695 9,663 31,643,783 663 47 49.93 112,776 10,161 31,957,614 559 70 49.93 138,232 10,243	Assembly size ^a (bp) No. of contigs >1,000 bp Avg genome coverage (x) GC content (%) N ₅₀ (bp) No. of genes and single-copy BUSCOs 29,562,918 745 63 50.38 100,935 9,685 3,487 (98.33) 29,374,270 922 39 50.37 71,695 9,663 3,466 (97.74) 31,643,783 663 47 49.93 112,776 10,161 3,494 (98.53) 31,957,614 559 70 49.93 138,232 10,243 3,503 (98.78)

TABLE 1 Overall genome assembly, completeness, and annotation statistics

^a Based on contigs with more than 1,000 bp.

of Pilon v1.22 (13) used in base correction to fix misassemblies and for gap filling. Overall statistics of the final assemblies were assessed with QUAST v4.6.3 (14). Genome assembly completeness was assessed by examining for the presence of 3,546 universal single-copy orthologs in Eurotiomycetes (eurotiomycetes_odb10) with BUSCO v4.0.4 (15). We used AUGUSTUS v3.3.1 (16) for prediction of protein-coding genes using the *A. fumigatus* Af293 gene models as a reference (17).

Assembly sizes, numbers of contigs, GC content, N_{50} contig values, gene numbers, percentages of complete and single-copy BUSCOs, and percentages of fragmented BUSCOs for all four strains are reported in Table 1. We summarized the genome statistics based on contigs of greater than 1,000 bp but included all contigs of greater than 200 bp in the assemblies submitted to GenBank. Genomic information of *Aspergillus* strains that are closely related to major human pathogens is important for understanding the origin and evolution of opportunistic human pathogenicity in the genus *Aspergillus*.

Data availability. The draft genome sequences of *A. felis* strains CNM-CM5623 and CNM-CM7691 and *A. hiratsukae* strains CNM-CM5793 and CNM-CM6106 are deposited in GenBank under the accession numbers JACBAE000000000, JACBAG00000000, JACBAD00000000, and JACBAF000000000, respectively. The raw reads of *A. felis* strains CNM-CM5623 and CNM-CM7691 and *A. hiratsukae* strains CNM-CM5793 and CNM-CM6106 are deposited in the NCBI Sequence Read Archive (SRA) under accession numbers SRR11804853, SRR11804830, SRR11802685, and SRR11802449, respectively. Genome assemblies and raw data are associated with BioProject number PRJNA633131.

ACKNOWLEDGMENTS

This research was funded by Brazilian São Paulo Research Foundation (FAPESP) grant 2016/07870-9 (to G.H.G.) and scholarships 2017/21983-3 and 2019/07526-4 (to R.A.C.D.S.) and by research projects from the Fondo de Investigación Sanitaria (PI13/ 02145 and PI16CIII/00035) of the Instituto de Salud Carlos III. J.L.S. and A.R. were funded by the Howard Hughes Medical Institute through the James H. Gilliam Fellowships for Advanced Study program. A.R. also received funding from the National Institutes of Health/National Institute of Allergy and Infectious Diseases (1R56Al146096-01A1).

Computational infrastructure was provided by the Advanced Computing Center for Research and Education (ACCRE) at Vanderbilt University.

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