

REVIEW ARTICLE

Open Access

GSK3 β : a plausible mechanism of cognitive and hippocampal changes induced by erythropoietin treatment in mood disorders?

Becky Inkster^{1,2,3}, Gwyneth Zai^{2,4,5,6}, Gemma Lewis⁷ and Kamilla W. Miskowiak^{6,8}

Abstract

Mood disorders are associated with significant psychosocial and occupational disability. It is estimated that major depressive disorder (MDD) will become the second leading cause of disability worldwide by 2020. Existing pharmacological and psychological treatments are limited for targeting cognitive dysfunctions in mood disorders. However, growing evidence from human and animal studies has shown that treatment with erythropoietin (EPO) can improve cognitive function. A recent study involving EPO-treated patients with mood disorders showed that the neural basis for their cognitive improvements appeared to involve an increase in hippocampal volume. Molecular mechanisms underlying hippocampal changes have been proposed, including the activation of anti-apoptotic, antioxidant, pro-survival and anti-inflammatory signalling pathways. The aim of this review is to describe the potential importance of glycogen synthase kinase 3-beta (GSK3 β) as a multi-potent molecular mechanism of EPO-induced hippocampal volume change in mood disorder patients. We first examine published associations between EPO administration, mood disorders, cognition and hippocampal volume. We then highlight evidence suggesting that GSK3 β influences hippocampal volume in MDD patients, and how this could assist with targeting more precise treatments particularly for cognitive deficits in patients with mood disorders. We conclude by suggesting how this developing area of research can be further advanced, such as using pharmacogenetic studies of EPO treatment in patients with mood disorders.

Mood disorders and cognitive deficits

Mood disorders affect ~20% of the general population¹ and for individuals suffering from a mood disorder, there is a 5–6% lifetime risk of completed suicide². Major depressive disorder (MDD) is ranked as the third most prevalent condition associated with disability³ and is estimated to be the second leading cause of disability worldwide by 2020⁴. Bipolar disorder (BD) is also on the

top ten list of most debilitating mental illnesses³ and is associated with significant psychosocial and occupational disability⁵. Both mood disorders, MDD and BD, are debilitating and chronic psychiatric disorders that cause significant suffering and burden in individuals with these illnesses and their families and friends, as well as reducing their quality of life^{6–8}.

Treatment of MDD and BD has focused on reducing mood symptoms;⁹ however, cognitive deficits are a core symptom domain of mood disorders¹⁰ that prolongs illness duration and reduces the likelihood of recovery^{11,12}. Cognitive dysfunction also contributes to socio-occupational impairment^{13,14}, which represents the largest economic cost of mood disorders for society^{15,16}.

Correspondence: Becky Inkster (becky.inkster@gmail.com) or Gemma Lewis (gemma.lewis@ucl.ac.uk)

¹Wolfson College, University of Cambridge, Cambridge, UK

²Department of Psychiatry, University of Cambridge, Cambridge, UK

Full list of author information is available at the end of the article.

These authors contributed equally: Becky Inkster and Gwyneth Zai

© The Author(s) 2018



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

Patients with MDD have consistently displayed difficulties in attention (e.g., in effortful attention, as well as automatic processing), declarative memory (e.g., verbal learning and memory, visuospatial learning and memory and episodic memory), and executive function (e.g., response inhibition, problem solving and planning, verbal fluency, decision-making and mental flexibility)¹⁷. These deficits are particularly pronounced in response to information that is emotionally or socially relevant. Similar but more severe deficits, specifically in verbal learning, spatial working memory, set-shifting and sustained attention, have been reported in patients with BD^{18,19}. While neurobiological mechanisms of cognitive impairments in mood disorders are unclear, converging preclinical, human neuroimaging and post-mortem evidence suggest that they may arise from disrupted neuroplasticity and associated structural changes in hippocampal volume^{20–22}. This highlights the potential of novel treatments with direct and lasting effects on neuroplasticity changes to induce enduring structural alterations and effectively alleviate cognitive deficits.

Pharmacological treatments for mood disorders have limited effects on cognitive dysfunction^{23,24} and are, in some cases, associated with adverse effects on cognition due to anticholinergic, sedative, extrapyramidal and/or blunting effects²⁵, which may exacerbate patients' persistent cognitive impairments during periods of remission (i.e., when patients are relatively symptom-free)²⁶. Existing cognitive enhancing drugs (i.e., medications aiming to improve cognitive functions) have shown limited pro-cognitive effects in depressed patients²⁷. Among the most promising cognition treatments are vortioxetine, which has shown replicated effects on psychomotor speed in symptomatic MDD²⁸, modafinil that improved some aspects of cognition in a study of remitted MDD²⁹, transcranial direct current stimulation that improved working memory in symptomatic MDD^{30,31}, lurasidone that improved a global measure of cognition in remitted BD³² and erythropoietin (EPO) that improved several cognitive domains in symptomatic MDD and remitted BD^{33,34}. However, despite these promising findings, there are no clinically available effective treatments for cognitive impairment in mood disorders to date^{35,36}. Indeed, many studies have examined the efficacy of existing and novel interventions to reduce cognitive dysfunction in patients with mood disorder;^{35,36} however, cognition trials in this area have faced some important methodological challenges that may negate the interpretations and significance of findings^{36,37}. Although preliminary evidence showed promising effects of psychological interventions for cognitive dysfunction, such as cognitive remediation in patients with MDD^{33,38}, we recently demonstrated a lack of beneficial effects of this intervention for BD patients in a randomized, controlled clinical trial³⁹. Notably, this trial

was limited by a small sample size ($n = 44$), short follow-up times (12 weeks) and lack of enrichment for the primary outcome (objectively-assessed verbal memory dysfunction). Indeed, emerging evidence indicates that cognitive remediation programs may be useful in BD and there are several ongoing cognitive remediation trials in BD.

Recent randomized, placebo-controlled trials demonstrated that 8 weekly doses of erythropoietin (EPO) reduced cognitive dysfunction in patients with treatment-resistant depression (TRD)³³ and in patients with BD in partial remission³⁴. Treatment-resistant depression was defined as lack of remission after ≥ 2 adequate antidepressant treatments with 2 different classes of antidepressant drugs in previous or current depressive episodes³³. The improvement of verbal memory after EPO vs. saline treatment across TRD patients and BD patients was of a moderate effect size (change in RAVLT total score, mean [SD]: EPO: 6.4 [8.8]; saline: 2.1 [8.0]; $d = 0.54$). Structural magnetic resonance imaging (MRI) assessments of patients from these two trials revealed that memory improvement was associated with normalization of volume loss in a subfield of the left hippocampus corresponding to the cornu ammonis 1–3 (CA1–3) and subiculum⁴⁰. Post hoc exploratory assessments of the mean surface displacement values revealed that the subfield hippocampal volume change was of a large effect size (hippocampal surface displacement, mean [SD]: EPO: 0.04 [0.08]; saline: -0.05 [1.0]; $d = 0.90$). However, the biological mechanisms linking EPO to increased hippocampal volume in mood disorders remain unknown.

EPO biology

EPO is a glycoprotein hormone cytokine that plays important roles in regulating red blood cell synthesis (i.e., hematopoiesis)⁴¹, trafficking of immune cells, anti-apoptotic actions, neurodevelopment⁴², neuroprotection and cognitive function^{43,44}. EPO and its receptor are expressed in multiple organ systems and have been shown to interact closely with the nervous, vascular, immune and reproductive systems^{45–47}. EPO is produced and secreted predominantly in the kidney, but it is also expressed in brain regions including the hippocampus, amygdala, temporal cortex, prefrontal cortex, internal capsule and midbrain^{45,48,49} as well as the liver and the uterus⁴⁷. Expression of EPO and its receptor have also been found in neurons, glial cells, endothelial cells and adult neural progenitor cells. Expression levels are high during human embryonic brain development, but remain present in adulthood⁴⁵. EPO functions in a hypoxia-sensitive manner meaning that stimuli such as hypoxia and stress (i.e., cellular changes such as hypoglycaemia, electrolyte imbalance, anaemia, infections and loss of endogenous anti-oxidants, etc.) can affect EPO and its receptor^{45–47},

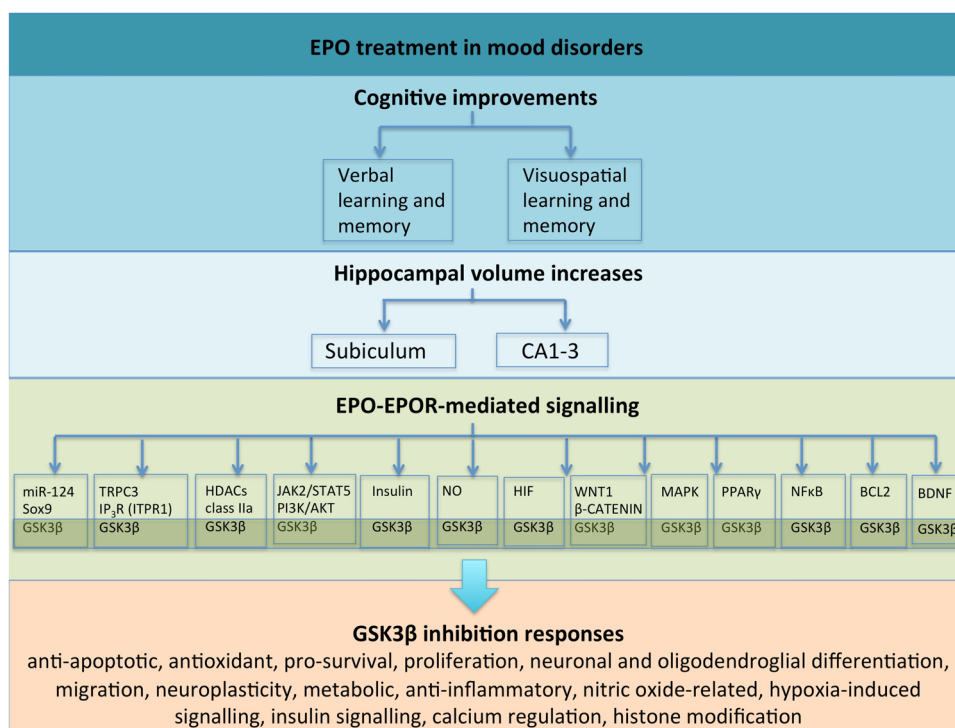


Fig. 1 This overview schematic summarizes the complex, interrelated relationships between EPO treatment in mood disorders, cognitive deficits, hippocampal changes and EPO’s potential mechanisms of action through the GSK3β inhibition . Notably, complex relationships exist across signalling pathways and molecules, which have not been illustrated

which can have pleiotropic effects in the modulation of apoptotic and immune activities⁵⁰ as well as neurotrophic and neuroprotection effects⁴⁶. Specifically, hypoxia-inducible factor (HIF) rapidly upregulates the expression of the EPO receptor, EPO-R, in cells of the Central Nervous System (CNS) and of EPO synthesis by neurons and astrocytes⁴⁵. Extracellular EPO then binds to EPO-R on the cell membrane, which triggers the intracellular JAK2 (janus kinase 2) signalling. This results in the activation of several signal transduction pathways including STAT5 (signal transducer and activator of transcription 5), PI3K (phosphatidylinositol 3-kinase)/Akt (protein kinase B), NFκB (nuclear factor-κB) and MAPK (mitogen-activated protein kinase). These pathways switch on signalling cascades that lead to long-lasting biological protective and reparative responses, which may be important for future treatment of cognitive impairments in neuropsychiatric disorders including depression⁴⁶. Specifically, relevant down-stream effects of these signalling cascades include activation of anti-apoptotic, antioxidant and anti-inflammatory signalling in neurons, glial and cerebrovascular endothelial cells, and promotion of dendritic sprouting, neurogenesis, hippocampal brain-derived neurotrophic factor (BDNF) and long-term potentiation^{51–53}. Erythropoietin was also shown to exert neuroprotective effects by inhibiting the activity of the enzyme

glycogen synthase kinase 3-beta (GSK3β)^{54,55}, as will be discussed in greater detail later in this review. This may be particularly relevant in relation to mood disorders since GSK3β is a key activator of cell death and other functions involved in mood disorders, hippocampal volume, glucocorticoid regulation and neuroplasticity^{56–58}.

It was a conceptual break-through that systemic administration of high-dose (> 500 International Units [IU]/kg) EPO was shown to cross the blood-brain barrier (BBB)⁴⁹ and facilitate neuroprotection and neuroplasticity in animal models of neurodegenerative and neuropsychiatric conditions⁵⁹ in addition to after acute neural injury^{60–62}. While it is unclear whether EPO crosses the BBB via an active transport mechanism or in an unspecific manner, it is evident that systemically administered high-dose EPO enters the brain to an extent that is sufficient for neuroprotection (ibid.). Accordingly, administration of such high doses of EPO to humans (through injections of 40,000–48,000 IU/ml)^{33,34,63–65} improved brain function and cognition after short-term (1 week) and longer-term (8–12 weeks) treatment. In contrast, short-term administration (3 days) administration of lower-dose EPO (30,000 IU to men of 74 ± 7 kg [mean ± SD]; corresponding to < 500 IU/kg) produced no cognitive benefits in healthy men⁶⁶ and 12 weeks low-dose EPO treatment (8000 IU/ml) produced no neural or cognitive benefits

with schizophrenia^{63,67}. Although no more precise pharmacokinetic or pharmacodynamic studies have been performed, this evidence indicates that high doses of EPO are required for neuroprotection and cognitive enhancement.

EPO has also been used to treat anaemia, ischaemia and reperfusion injuries (i.e., stroke, heart attack)⁶⁸, neurological disorders (i.e., seizures⁶⁹, spinal cord ischaemia, Alzheimer's disease, Parkinson's disease and demyelinating disease⁴⁷), and retinal disease⁴⁷ and neuropsychiatric disorders^{33,34,46}. Thus, knowledge of the underlying mechanisms of EPO may provide important insights for future therapeutic strategies for the treatment of neuropsychiatric, neurodegenerative, inflammatory and autoimmune-related disorders.

In this review, we highlight evidence collectively suggesting that inhibition of GSK3 β acts as a multi-potent molecular mechanism that may mediate multi-potent effects of EPO on hippocampal volume changes in depression (Fig. 1). Understanding the complex relationship between EPO and GSK3 β (and its pleiotropic regulatory role across its large genetic network) on cognitive functioning in depressed patients may help reveal new drug targets (both upstream and downstream), aid precision medicine, and ultimately reduce disability and mortality for mood disorders.

Narrative review search methodology

The following search terms were included in this review: cognition, cognitive functions or dysfunction or impairment or deficits, cognitive enhancers or enhancement, mood disorders, depression, bipolar disorder (BD), major depressive disorder (MDD), treatment-resistant depression (TRD), erythropoietin (EPO), glycogen synthase kinase-3 beta (*GSK3 β*), hippocampus, hippocampal volume or structure, molecular pathway and biology or biological. Several search engines were used, including PubMed and Medline. This review has mainly focused on unipolar and bipolar depression and therefore, only the most recent reviews on other disorders such as neurological and cardiac diseases have been included for references. Two factors led us to choosing a narrative style for the review paper: firstly, to our knowledge, this is the first review paper to bridge these complex interrelated topics in the literature and, secondly, it was not our intention to perform an extensive systematic search for each of the topics independently as this would be an enormous undertaking beyond the scope of our narrative approach.

EPO treatment and cognitive function

Studies in patients with schizophrenia, and multiple sclerosis, have shown that 8–12 weeks of high-dose

(40,000–48,000 IU) EPO treatment improves cognitive functioning that lasts for up to 6 months after treatment completion, long beyond red blood cell normalization^{63,64}. This indicates that the pro-cognitive effects of EPO are not directly related to changes in the vascular system. Indeed, the effects of EPO on neurocognitive function in humans seem to be mediated through neurobiological actions rather than indirect increases in red blood cells^{65,70}. In particular, these studies demonstrated that a single high dose of EPO (40,000 IU) versus saline improves neural and cognitive measures of memory and executive functioning in healthy volunteers without affecting red blood cells (*ibid.*). Based on this evidence, Miskowiak et al.^{33,34} conducted a randomised, placebo-controlled clinical trial examining the effects of 8 weekly infusions of EPO (40,000 IU) on mood symptoms and cognitive dysfunction in patients with TRD and patients with BD in partial remission. EPO treatment improved verbal memory in TRD patients and speed of complex cognitive processing across attention, memory and executive function in BD patients relative to placebo treatment. These cognitive changes were independent of changes in mood symptoms and were maintained several weeks after red blood cell normalisation at a 6-week follow-up at which time EPO-treated patients displayed structural increase in the left hippocampus⁴⁰ and changes in task-related neural activity within a fronto-parietal network^{71,72}. Importantly, post hoc analyses showed that the structural hippocampal increase and task-related neural activity change correlated with the observed improvements in EPO-treated patients' cognitive functions, whereas no influence was found of changes in red blood cells, mood symptoms, diagnosis, age or gender^{40,71,72}.

Effects of EPO have also been demonstrated on neural and cognitive responses to facial expressions in healthy volunteers^{70,73} and were subsequently replicated in a sample of patients with acute depression⁷⁴. Long-term EPO treatment did not improve the primary measure of depression severity in an 8-week trial (Hamilton Depression Rating Scale [HDRS] score), but this may be a result of suboptimal statistical power⁷⁵ and the use of HDRS, which might underestimate other less relevant depressive symptom domain and burden of illness that correlate poorly with depression severity;⁷⁶ however, improvement in several other depression-relevant outcomes including self-rated depression and quality of life were observed, suggesting that further investigations of the antidepressant efficacy of EPO in larger-scale trials are warranted³³. Given this evidence demonstrating the potential impact of EPO on cognitive function and mood symptoms, it is important to elucidate the biological mechanisms underlying alterations of neural processing.

GSK3 β : biological mechanism of mood disorders

GSK3 β is a highly active proline-directed serine-threonine protein kinase. It contributes to diverse cellular functions including gene expression, neurogenesis, neuroplasticity, cell survival, differentiation, migration, stress responses, cell structure, cell death, the immune system, neurotransmitter systems, metabolism and other functions^{77–80}. GSK3 β inhibitors increase proliferation, migration and differentiation of neural stem cells in the adult hippocampal dentate gyrus⁸¹. GSK3 β is ubiquitously expressed throughout the brain, most prominently in the cerebral cortex and hippocampus (Allen Brain Atlas). GSK3 β is a particularly unique protein kinase⁸² that can be inactivated through the action of various kinases, such as Akt/protein kinase B, protein kinase A and protein kinase C on the ninth position of serine (Ser9)⁸³.

Several neurogenetics studies have investigated associations between *GSK3 β* and mood disorders. The GSK3 β gene (*GSK3 β* ; OMIM 605004) was mapped to chromosome 3q13.3⁸⁴. Functional single nucleotide polymorphisms (SNPs) have been identified in *GSK3 β* ; for example, a promoter T to C polymorphism at position –50 (rs334558) with the T allele having a higher in vitro transcriptional activity and an intron 5T to C polymorphism at position –157 (rs6438552) with the T allele lacking exons 9 and 11 and has been associated with an increased level of GSK3 β ⁸⁵. Several studies have investigated genetic variants in *GSK3 β* as risk factors for MDD^{86,87} and BD⁸⁸. Other studies have focused on anxiety symptoms in MDD and P300 waveform⁸⁹, psychotic symptoms in MDD and BD⁹⁰, age of onset in MDD⁹¹ and BD⁹², suicidal behaviour in MDD⁹³ and combined cases of MDD and schizophrenia patients⁹⁴. Furthermore, *GSK3 β* polymorphisms have been examined as a predictor of antidepressant response⁹⁵ and lithium response^{96,97}.

Neuroimaging genetic studies of mood disorders have reported associations between *GSK3 β* variation and hippocampal volume. A genetic association study of numerous *GSK3 β* SNPs and brain-wide grey matter volume using MRI-based voxel-based morphometry was conducted in a sample of 134 patients with recurrent MDD and 144 healthy controls⁵⁶. Disease modulated associations were reported between grey matter volume in the right hippocampus and bilateral temporal cortex and a functional intronic *GSK3 β* polymorphism, rs6438552. The same direction of association was observed in a larger, independent sample of healthy volunteers between the same *GSK3 β* polymorphism and hippocampal volume using different neuroimaging methods⁹⁸. This polymorphism has also been associated with altered resting state networks in MDD patients⁹⁹. Based on in vitro work, this polymorphism alters the splice acceptor site leading to exclusion of exons 9 and 11, which alters the protein's

function to then hyperphosphorylate the substrate, microtubule-associated protein tau⁸⁵. Further in vivo and in vitro work is required to understand how this modified GSK3 β protein regulates other substrates. Additional associations between hippocampal volume and genetic variation involving GSK3 β -related pathways and other directly interacting proteins have also been reported^{57,58}.

Identifying putative connections between GSK3 β , erythropoietin, hippocampus, cognition and mood disorders

The hippocampus is an important brain region implicated in mood disorders. Specifically, changes in the neural circuitry of the hippocampus have been implicated in cognitive deficits in patients with mood disorders^{100–102}, which may arise in part from the disruption of neuroplasticity⁶⁷. Disturbance in hippocampal neuroplasticity has been hypothesised to play an aetiological role in mood disorders and may result from chronic inflammatory processes and over-activation of stress responses^{103–105}. This is consistent with evidence showing that stress-induced glucocorticoid production is associated with reduced hippocampal neurogenesis, hippocampal memory deficits and depression-like behaviour in animals^{106–109}. Moreover, a recent meta-analysis¹¹⁰ supported an overall significant hippocampal volume reduction in patients with MDD relative to controls and several additional studies reported hippocampal subiculum shape abnormalities in patients with depression^{111–113}.

The involvement of GSK3 β in EPO-mediated neuroprotection via PI3K/AKT is well documented in the literature (e.g. see refs ^{45,114–116}). In the context of primary hippocampal neurons, EPO treatment triggers pro-survival mechanisms by activation of PI3K/AKT⁴⁵, which suppresses downstream target GSK3 β (i.e., by increasing phosphorylation of Ser9 in GSK3 β)¹¹⁷. In contrast, PI3K/AKT pathway inactivation results in GSK3 β pro-apoptotic functions. In a recent study, Ma and colleagues¹¹⁶ administered exogenous EPO to rats for 4 weeks using an animal model of vascular dementia. Their results indicated improvements in memory impairment, promotion of hippocampal dendritic spine growth as well as deactivation of GSK3 β via an EPO-R/JAK2/STAT5/PI3K/Akt/GSK3 β pathway¹¹⁶ (Fig. 1).

Another mechanism of action of EPO treatment that could be linked with GSK3 β function is through the central role that GSK3 β plays in neuronal and oligodendroglial differentiation. A recent study by Hassouna and colleagues¹¹⁸ examined the effects of EPO in young, healthy mice administered EPO for 3 weeks. The authors reported an approximately 20% increase in hippocampal CA1/CA3 neurons and oligodendrocytes, and they detected a significant enhancement of neuronal and oligodendroglial differentiation rather than proliferation¹¹⁸.

Using neural stem cells and hippocampal cultures, the authors found that EPO administration decreased the transcription factor Sry-box 9 (Sox9) and increased micro RNA 124 (miR-124). miR-124 is known to regulate Sox9 function and drive neuronal differentiation¹¹⁸. We highlight evidence showing an interconnected relationship between GSK3 β , Sox9 and miR-124. Sox9 interacts with GSK3 β via its targets in the Wnt signalling pathway^{119,120}. For example, Sox9 inhibits the GSK3 β -dependent Wnt/beta-catenin signaling pathway in chondrocyte differentiation by promoting beta-catenin phosphorylation in the nucleus. This finding is in keeping Hassouna and colleagues¹¹⁸ in that EPO inhibits Sox9 although it should be noted that different tissues and models were used and so further investigations are warranted for mood disorders. Other Sox-related genes should also be explored given that, for example, Sox17 regulates the Wnt/ β -catenin signaling pathway via GSK3 β in oligodendrocyte progenitor cells¹²¹. Furthermore, miR-124 co-regulates neuronal differentiation and dendritic architecture via the AKT/GSK3 β -dependent pathway¹²². Its regulation of GSK3 β hippocampal expression may have implications for chronic stress and mood disorder pathophysiology^{123,124}. Further evidence has shown that miR-124 regulates HDAC4 and GSK3 β expression in the hippocampus, which may have important implications for chronic stress and depression¹²⁴ and another study identified associations between *HDAC4* genetic variation and reduced hippocampal volume in two independent MDD cohorts⁵⁸. Notably, another class IIa histone deacetylases (HDAC5) has been implicated in the therapeutic action of EPO whereby researchers found that EPO regulates phosphorylation at two different sites stimulating nuclear export of HDAC5 in rat hippocampal neurons¹²⁵. Collectively and indirectly, these diverse studies provide a plausible link between EPO treatment and its downstream effects on GSK3 β function (Fig. 1), which require much greater examination in order to delineate specific and selective effects.

Research has shown that EPO stimulates calcium influx. In terms of biological mechanisms, one study demonstrated that interactions between inositol 1,4,5-trisphosphate (ITPR1; alias, IP₃R) and transient receptor potential cation channel subfamily C member 3 (TRPC3) is required for epo-modulated Ca²⁺ influx, which was reduced under conditions of mutated or deleted IP₃R binding sites on TRPC3¹²⁶. *ITPR1* (alias, IP₃R) genetic variation was recently associated with reduced hippocampal volume in two independent MDD cohorts, which lead the authors to speculate that mood disorders, and specifically cognitive changes, may involve mechanisms related to ITPR, endoplasmic reticulum (ER) stress, the unfolded protein response (UPR) system and GSK3 β signalling⁵⁶.

Another possible way in which EPO treatment could be linked with GSK3 β function is through anti-apoptotic mechanisms. Several biological models have implicated GSK3 β as a key activator of cell death⁷⁹ and so inactivation of GSK3 β may therefore promote cell viability. For example, evidence has demonstrated a molecular relationship between EPO, GSK3 β and the mitochondrial cell death pathway; EPO suppresses 6-hydroxydopamine (6-OHDA)-induced apoptosis by increasing phosphorylation of Ser9 in GSK3 β (i.e., increasing GSK3 β inhibition)⁵⁴. Neuroprotective effects against apoptosis were observed for both EPO and the GSK3B inhibitor 4-benzyl-2-methyl-1, 2,4-thiadiazolidine-3, 5-dione (TDZD8). In contrast, 6-OHDA decreased phosphorylation of Ser9 in GSK3 β (i.e., increased GSK3 β activity). In this study, decreases in mitochondrial expression of the anti-apoptotic gene B-cell lymphoma 2 (*Bcl-2*) were also observed (Fig. 1). Other related work has also described a relationship between EPO treatment, increased phosphorylation of Ser9 in GSK3 β , and oxidant stress-induced apoptosis^{127,128}. Further investigation is crucial to understand how EPO treatment interacts with GSK3 β function in different brain tissue types, cellular environments and diseases.

An additional relationship between EPO and GSK3 β involves the downstream increase in hippocampal brain-derived neurotrophic factor (BDNF) expression, neurite growth and spine density^{53,129} (Fig. 1). BDNF is highly involved in neuroplasticity, cell survival, differentiation and cell death^{130,131} as well as learning and memory^{132–134}. Evidence has shown that GSK3 β interacts with BDNF at the protein level; GSK3 β overexpression inhibits BDNF-induced cAMP response element-binding (CREB) phosphorylation^{135,136}. *GSK3 β* and *BDNF* genotype combinations have been associated with MDD⁸⁶.

A complex relationship between EPO, GSK3 β , the hippocampus and depression may exist, in part, through nitric oxide (NO)-related pathways. In brief, increased GSK3 β mRNA expression was found in post-mortem hippocampal samples from MDD patients, which is consistent with previous animal studies of depression. GSK3 β mRNA expression was also significantly correlated with nitric oxide synthase 1 (NOS1) in these same patients¹³⁷, which is in keeping with previous evidence suggesting that nitric oxide activates GSK-3 β . EPO can influence oxygen delivery through stimulation of NO production⁴⁵, which may contribute to its neuroprotective role; however, this relationship is complex and very much dependant on the cell and tissue type, and different dose-time exposure conditions (i.e., short-term versus long-term exposure, hypoxia versus normoxia conditions etc.). Possible relationships between EPO, GSK-3 β , NOS and hypoxia may exist, although specific mechanisms remain unclear. Hypoxia modulates NOS mRNA and protein levels under

specific conditions¹³⁸, GSK-3 β overexpression is associated with reduced hypoxia-inducible transcription factor 1 α (HIF-1 α)¹³⁹, while EPO-R expression is rapidly upregulated by HIF⁴⁵. More detailed work in this area is needed to understand the isoform-specific interactions however (e.g., the role of HIF-1 α versus HIF-2 α etc.).

GSK3 β acts centrally in the canonical Wnt signalling pathway, which is essential for regulating neurodevelopment as well as synaptic maintenance and plasticity in the adult brain¹⁴⁰. Independent evidence has implicated the Wnt pathway in mood disorders^{57,141}. Biological interactions between EPO and the canonical Wnt signalling pathway have been observed in elevated D-glucose models of diabetes¹⁴². These authors¹⁴² found that EPO triggered anti-apoptotic responses via the modulation of Wnt1 protein expression that subsequently promoted β -catenin translocation. The authors¹⁴² also reported that Wnt1 gene silencing and Wnt1 antagonist administration prevented the protective EPO treatment. Notably, biological interactions between EPO and Wnt signalling have also been a proposed mechanism of action for neurodegenerative diseases⁴⁷.

Peroxisome proliferator-activated receptor-gamma co-factor 1A (*PPARGC1A*) is involved in the PPAR- γ system, which interacts with numerous pathways including the Wnt signalling pathway⁵⁷. Evidence has shown that *PPARGC1A* genetic variation is associated with altered brain volume in MDD patients⁵⁷. Activation of the PPAR- γ system has been shown to improve depressive-like behaviours¹⁰⁵. It has been proposed that PPAR- γ plays a protective role against ER stress¹⁰⁵ and that PPAR- γ pro-survival activity is inhibited by HDAC4 activation¹⁴³. Furthermore, the PPAR- γ system has been linked to EPO function¹⁴⁴. For example, a study examining the therapeutic implications of EPO in type 2 diabetes and insulin resistance found that EPO regulates the PI3K/AKT signalling pathway via PPAR γ -dependent activation¹⁴⁴ (Fig. 1).

Insulin signalling pathways also share complex relationships with both EPO and GSK3 β . Evidence has shown that insulin-like growth factor leads to increased EPO and EPOR expression in neuronal cells⁴⁶ and that GSK3 β is inhibited by insulin-mediated mechanisms¹⁴⁵. It has been proposed that impaired insulin receptor-mediated regulation of GSK3 β activity is involved with the cognition and depression¹⁴⁶.

EPO, GSK3 β and pharmacological treatments

Evidence from animal studies suggests that inhibition of GSK3 β is a potential mechanism contributing to the antidepressant-like effects of lithium, ketamine¹⁴⁷ and valproate¹⁴⁸. Lithium is considered to be the gold standard pharmacological treatment for BD and has pleiotropic effects on multiple cellular systems and

pathways¹⁴⁹. Additionally, lithium treatment results in significant inhibition of GSK3 activity^{150,151}, which has been shown to mediate neuroprotective, anti-oxidative and neurotransmission mechanisms. The effect of lithium-induced GSK3 inhibition has also previously been shown to reduce tauopathy and neurodegeneration¹⁵², and another study demonstrated that lithium (Li⁺) inhibits GSK3 by competition for magnesium (Mg²⁺)¹⁵³. With regard to ketamine, while the literature is inconclusive, there is an indication that ketamine may be effective at treating depression¹⁵⁴, in particular severe depression, TRD and acute suicidality. With its fast-acting properties¹⁵⁵, ketamine has been shown to interact with EPO^{156,157}. The combination of EPO and ketamine may offer new areas of investigation for mood disorder treatments. The antidepressant actions of ketamine involve GSK3 β inhibition¹⁴⁷. Lithium and other selective GSK3 β inhibitors enhance the effects of low doses of ketamine¹⁵⁸ and the authors suggested that GSK3 β activation is an underlying mechanism related to ketamine-induced apoptosis. Low-dose interactions may be of particular interest for reducing the risk of side effects and possible misuse given prior evidence implicating ketamine with misuse and addiction¹⁵⁹. Ketamine has been shown to modulate inflammatory responses¹⁶⁰. Acute or chronic use of ketamine has been found to induce cognitive impairments with hyperphosphorylation of tau and apoptosis¹⁶¹, and transient behavioural changes similar to schizophrenia (i.e., motor and social behavioural disturbances)^{162,163}. However, studies have previously shown that ketamine has anti-inflammatory effects under inflammatory conditions and has been used in surgical procedures in patients with sepsis^{164,165}, chronic stress-induced depression¹⁶⁶, mood disorders in general¹⁶⁷ and severe TRD¹⁶⁸. The effect of ketamine in reducing depressive symptoms has been shown to be fast-onset but short-lived and requires continual or maintenance treatment; however, the safety of long-term ketamine use has not yet been examined¹⁶⁹. Evidence has suggested the involvement of the serotonergic and dopaminergic systems in addition to the glutamate N-methyl-D-aspartate (NMDA) receptor and BDNF¹⁶⁹. Studies have postulated that excessive or ill-timed NMDA antagonism by ketamine may induce glutamate excitotoxicity, which further complicates the role of ketamine in neuroprotection or neurotoxicity, and its clinical utility¹⁶⁹. Future clinical trials that examine the EPO-ketamine combination treatment would be of interest, especially in patients who have molecular measures of GSK3 β given its interactions with EPO and ketamine. GSK3 β cellular signalling is extremely fast acting and responsive to cellular changes. Animal studies have also shown that the monoamine reuptake inhibitor antidepressants, fluoxetine and imipramine, increase the inhibitory control of

phosphorylation of Ser9 in GSK3^{170,171}. Furthermore, valproate directly inhibits GSK3 β and was shown to protect cells from ER stress and apoptosis¹⁴⁸. Inhibition of GSK3 β is therefore a possible mechanism of action shared by several classes of antidepressant medication and other emerging medications (i.e., ketamine) for the treatment of depression. Whether the pharmacological effects of these antidepressants on GSK3 β contribute to the reduction of depressive symptoms is yet to be established.

Future directions and challenges

Here we have reviewed literature that examines the relationship between EPO, mood disorders, cognition and the hippocampus. We then speculated that EPO inhibits GSK3 β activity and subsequently might alter complex signalling cascades to improve cognition via hippocampal brain changes. A key limitation of this review is that the selective molecular effects of the treatment with EPO remain unclear. Also, other antidepressant therapies have strong overlap with these cellular pathways presented in this review indicating that much more work is required to unravel directly relevant versus secondary molecular events in the context of cognitive and hippocampal in mood disorders. One key area that needs prioritising is to explore cellular differentiation molecular mechanisms. Future studies investigating the effects of EPO on different cellular networks mediated by GSK3 β are highly warranted to identify its common and specific roles in the treatment of mood disorders and other neuropsychiatric illnesses. Given the similarities and known differences between MDD and BD, further exploration of the underlying mechanism that differentiates unipolar and bipolar depression is necessary for novel treatment of these debilitating and chronic mood disorders. Nevertheless, the beneficial effects of EPO on cognition and hippocampal volume have been observed across several neuropsychiatric diseases including MDD, BD⁴⁰ and schizophrenia⁶⁷, suggesting that EPO modulates common signalling pathways involved in neuroplasticity and cognition across these disorders. Preliminary evidence suggests that GSK3 β inhibition may play a role in improving a range of cognitive deficits^{172,173}. We therefore recommend that further studies directly test for associations between hippocampal-related cognitive measures in mood disorders and GSK3 β -related genetic networks (e.g., ITPR1) as well as considering co-treatment designs with EPO, such as ER stress inhibitors⁵⁸. This may lead to future potential treatment options more targeted for illness-related cognitive impairments¹⁷³.

Additional research is required to elucidate the role of the *EPO* gene (OMIM: 133170) and its related genetic variation; surprisingly, this has not been studied in mood disorders (or psychiatric disorders more generally). To our knowledge, only one study to date has investigated

genetic variants across *EPO* and *EPOR* in schizophrenia, which showed initial promising results in cognitive modulation⁴³. Given the caveats for genetic association studies and recruitment challenges for EPO patient studies, interactions between GSK3 β and *EPO/EPOR* also require further examination using large scale, well powered healthy participant populations. In vivo work examining the functional effects of *EPO/EPOR* and GSK3 β will also be an important avenue for further investigation.

The evidence to date suggests that EPO has potential clinical utility to reduce cognitive deficits in patients with depression. There is no known pharmacokinetic drug–drug interaction and no adverse events were observed in the recent EPO clinical trials^{33,34}. Nevertheless, significant adverse events for EPO treatment have been reported including tumour progression and thromboembolic events. Given these potential risks of EPO treatment, extensive screening is necessary prior to starting EPO therapy and EPO-treated patients must also be closely monitored (for details, see ref. ⁷⁴). Furthermore, the long-term benefits and use of EPO in patients need to be demonstrated in clinical studies with longer-term follow-up times regarding its potential benefits and risks. Specifically, studies using six months follow-up assessments of cognition and functioning are highly warranted given the short (6 weeks) follow-up times in the recent trials in BD and MDD. Knowing the biological and molecular genetic mechanisms, and pharmacogenetics underlying the effects of EPO, may guide clinicians and patients in understanding who will tolerate and respond to EPO treatment. This will allow clinicians to choose the best medications for each individual patient for precision medical care.

Given the highly pleiotropic effects of GSK3 β in triggering multiple pathways and processes, including cancer development and tumour growth¹⁷⁴, it is important to extensively investigate molecular targets that act with less potency and greater GSK3 β downstream specificity (i.e., its numerous substrates). However, in vivo and in vitro evidence is currently limited and many unknown putative GSK3 β substrates may exist. The identification of more GSK3 β substrates is therefore of great importance for understanding the larger impact GSK3 β plays in hippocampal volume of individuals with mood disorders. Algorithms that estimate the likelihood of proteins binding to GSK3 β should facilitate this work¹⁷⁵. Furthermore, exploration of protein kinases required for priming phosphorylation prior to GSK3 β protein docking should also be explored, as is being explored in cancer research¹⁷⁴. Also crucial will be harnessing the power of statistical methods, such as machine learning, to better understand how genotypic combinations combinations interact across these substrates and upstream regulating proteins as part of

15. Olesen, J., Gustavsson, A., Svensson, M., Wittchen, H. U. & Jönsson, B. CDBE2010 study group, European Brain Council. The economic cost of brain disorders in Europe. *Eur. J. Neurol.* **19**, 155–162 (2012).
16. Wyatt, R. J. & Henter, I. (1995): An economic evaluation of manic-depressive illness–1991. *Soc. Psychiatry Psychiatr. Epidemiol.* **30**, 213–219 (1995).
17. Murrough, J. W., Iacoviello, B., Neumeister, A., Charney, D. S. & Iosifescu, D. V. Cognitive dysfunction in depression: neurocircuitry and new therapeutic strategies. *Neurobiol. Learn. Mem.* **96**, 553–563 (2011).
18. Raust, A. et al. Neurocognitive performance as an endophenotype for bipolar disorder. *Front Biosci. (Elite Ed.)* **6**, 89–103 (2014).
19. Gualtieri, C. T. & Morgan, D. W. The frequency of cognitive impairment in patients with anxiety, depression, and bipolar disorder: an unaccounted source of variance in clinical trials. *J. Clin. Psychiatry* **69**, 1122–1130 (2008).
20. Canales-Rodríguez, E. J. et al. Structural abnormalities in bipolar euthymia: a multicontrast molecular diffusion imaging study. *Biol. Psychiatry* **76**, 239–248 (2014).
21. McKinnon, M. C., Yucel, K., Nazarov, A. & MacQueen, G. M. A meta-analysis examining clinical predictors of hippocampal volume in patients with major depressive disorder. *J. Psychiatry Neurosci.* **34**, 41–54 (2009).
22. Marsden, W. N. Synaptic plasticity in depression: molecular, cellular and functional correlates. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **43**, 168–184 (2013).
23. Bora, E., Harrison, B. J., Yucel, M. & Pantelis, C. Cognitive impairment in euthymic major depressive disorder: a meta-analysis. *Psychol. Med.* **43**, 2017–2026 (2013).
24. Bourne, C. et al. Neuropsychological testing of cognitive impairment in euthymic bipolar disorder: an individual patient data meta-analysis. *Acta Psychiatr. Scand.* **128**, 149–162 (2013).
25. Dias, V. V. et al. Pharmacological approaches in bipolar disorders and the impact on cognition: a critical overview. *Acta Psychiatr. Scand.* **2012**, 315–331 (2012).
26. Mora, E., Portella, M. J., Forcada, I., Vieta, E. & Mur, M. Persistence of cognitive impairment and its negative impact on psychosocial functioning in lithium-treated, euthymic bipolar patients: a 6-year follow-up study. *Psychol. Med.* **43**, 1187–1196 (2013).
27. Goeldner, C. et al. Cognitive impairment in major depression and the mGlu2 receptor as a therapeutic target. *Neuropharmacology* **64**, 337–346 (2013).
28. McIntyre, R. S., Harrison, J., Loft, H., Jacobson, W. & Olsen, C. K. The effects of vortioxetine on cognitive function in patients with major depressive disorder: a meta-analysis of three randomized controlled trials. *Int. J. Neuropsychopharmacol.* **24**, pyw055 (2016).
29. Kaser, M. et al. Modafinil improves episodic memory and working memory cognition in patients with remitted depression: a double-blind, randomized, placebo-controlled study. *Biol. Psychiatry Cogn. Neurosci. Neuroimaging.* **2**, 115–122 (2017).
30. Fregni, F., Boggio, P. S., Nitsche, M. A., Rigonatti, S. P. & Pascual-Leone, A. Cognitive effects of repeated sessions of transcranial direct current stimulation in patients with depression. *Depress Anxiety* **23**, 482–484 (2006).
31. Wolkenstein, L. & Plevnia, C. Amelioration of cognitive control in depression by transcranial direct current stimulation. *Biol. Psychiatry* **73**, 646–651 (2013).
32. Yatham, L. N. et al. Lurasidone versus treatment as usual for cognitive impairment in euthymic patients with bipolar I disorder: a randomised, open-label, pilot study. *Lancet Psychiatry* **4**, 208–217 (2017).
33. Miskowiak, K. W. et al. Recombinant human erythropoietin for treating treatment-resistant depression: a double-blind, randomized, placebo-controlled phase 2 trial. *Neuropsychopharmacology* **39**, 1399–1408 (2014).
34. Miskowiak, K. W., Ehrenreich, H., Christensen, E. M., Kessing, L. V. & Vinberg, M. Recombinant human erythropoietin to target cognitive dysfunction in bipolar disorder: a double-blind, randomized, placebo-controlled phase 2 trial. *J. Clin. Psychiatry* **75**, 1347–1355 (2014).
35. Miskowiak, K. W., Ott, C. V., Petersen, J. Z. & Kessing, L. V. Systematic review of randomized controlled trials of candidate treatments for cognitive impairment in depression and methodological challenges in the field. *Eur. Neuropsychopharmacol.* **26**, 1845–1867 (2016).
36. Miskowiak, K. W., Carvalho, A. F., Vieta, E. & Kessing, L. V. Cognitive enhancement treatments for bipolar disorder: A systematic review and methodological recommendations. *Eur. Neuropsychopharmacol.* **26**, 1541–1561 (2016).
37. Carvalho, A. F. et al. Cognitive dysfunction in depression - pathophysiology and novel targets. *Cns. Neurol. Disord. Drug. Targets* **13**, 1819–1835 (2014).
38. Porter, R. J., Bowie, C. R., Jordan, J. & Malhi, G. S. Cognitive remediation as a treatment for major depression: a rationale, review of evidence and recommendations for future research. *Aust. N. Z. J. Psychiatry* **47**, 1165–1175 (2013).
39. Demant, K. M., Vinberg, M., Kessing, L. V. & Miskowiak, K. W. Effects of short-term cognitive remediation on cognitive dysfunction in partially or fully remitted individuals with bipolar disorder: results of a randomised controlled trial. *PLoS ONE* **2015**, e0127955 (2015).
40. Miskowiak, K. W. et al. Effects of erythropoietin on hippocampal volume and memory in mood disorders. *Biol. Psychiatry* **78**, 270–277 (2015).
41. Jacobson, L. O., Goldwasser, E., Fried, W. & Plzak, L. F. Studies on erythropoiesis. VII. The role of the kidney in the production of erythropoietin. *Trans. Assoc. Am. Physicians* **70**, 305–317 (1957).
42. Siren, A. L., Fasshauer, T., Bartels, C. & Ehrenreich, H. Therapeutic potential of erythropoietin and its structural or functional variants in the nervous system. *Neurotherapeutics* **2009**, 6, 108–127 (2009).
43. Kastner, A. et al. Common variants of the genes encoding erythropoietin and its receptor modulate cognitive performance in schizophrenia. *Mol. Med.* **18**, 1029–1040 (2012).
44. Sargin, D., Friedrichs, H., El-Kordi, A. & Ehrenreich, H. Erythropoietin as neuroprotective and neuroregenerative treatment strategy: comprehensive overview of 12 years of preclinical and clinical research. *Best. Pract. Res. Clin. Anaesthesiol.* **24**, 573–594 (2010).
45. Noguchi, C. T., Asavaritkrai, P., Teng, R. & Jia, Y. Role of erythropoietin in the brain. *Crit. Rev. Oncol. Hematol.* **2007**, 64, 159–171 (2007).
46. Ma, C. et al. Erythropoietin pathway: a potential target for the treatment of depression. *Int. J. Mol. Sci.* **17**, E677 (2016).
47. Maiese, K., Chong, Z. Z., Shang, Y. C. & Wang, S. Erythropoietin: new directions for the nervous system. *Int. J. Mol. Sci.* **13**, 11102–11129 (2012).
48. Marti, H. H. et al. Erythropoietin gene expression in human, monkey and murine brain. *Eur. J. Neurosci.* **8**, 666–676 (1996).
49. Brines, M. & Cerami, A. Emerging biological roles for erythropoietin in the nervous system. *Nat. Rev. Neurosci.* **2005**, 6, 484–494 (2005).
50. Nairz, M., Sonnweber, T., Schroll, A., Theurl, I. & Weiss, G. The pleiotropic effects of erythropoietin in infection and inflammation. *Microbes Infect.* **14**, 238–246 (2012).
51. Byts, N. & Sirén, A. L. Erythropoietin: a multimodal neuroprotective agent. *Exp. Transl. Stroke Med.* **1**, 4 (2009).
52. Girgenti, M. J. et al. Erythropoietin induction by electroconvulsive seizure, gene regulation, and antidepressant-like behavioral effects. *Biol. Psychiatry* **66**, 267–274 (2009).
53. Leconte, C. et al. Comparison of the effects of erythropoietin and its carbamylated derivative on behaviour and hippocampal neurogenesis in mice. *Neuropharmacology* **60**, 354–364 (2011).
54. Ge, X. H., Zhu, G. J., Geng, D. Q., Zhang, Z. J. & Liu, C. F. Erythropoietin attenuates 6-hydroxydopamine-induced apoptosis via glycogen synthase kinase 3beta-mediated mitochondrial translocation of Bax in PC12 cells. *Neuro. Sci.* **33**, 1249–1256 (2012).
55. Li, Y. P. et al. Erythropoietin attenuates Alzheimer-like memory impairments and pathological changes induced by amyloid β 42 in mice. *Brain Res.* **1618**, 159–167 (2015).
56. Inkster, B. et al. Association of GSK3beta polymorphisms with brain structural changes in major depressive disorder. *Arch. Gen. Psychiatry* **66**, 721–728 (2009).
57. Inkster, B. et al. Pathway-based approaches to imaging genetics association studies: Wnt signaling, GSK3beta substrates and major depression. *Neuroimage* **2010**, 53, 908–917 (2010).
58. Inkster, B. et al. Unravelling the GSK3 β -related genotypic interaction network influencing hippocampal volume in recurrent major depressive disorder. PG-D-17-00038 accepted, *Psychiatric Genetics* (2018).
59. Brines, M. L. et al. Erythropoietin crosses the blood-brain barrier to protect against experimental brain injury. *Proc. Natl Acad. Sci. USA* **97**, 10526–10531 (2000).
60. Catania, M. A. et al. Erythropoietin prevents cognition impairment induced by transient brain ischemia in gerbils. *Eur. J. Pharmacol.* **437**, 147–150 (2002).
61. Mogensen, J. et al. Erythropoietin improves place learning in fimbria-fornix-transected rats and modifies the search pattern of normal rats. *Pharmacol. Biochem. Behav.* **77**, 381–390 (2004).
62. Siren, A. L. et al. Global brain atrophy after unilateral parietal lesion and its prevention by erythropoietin. *Brain* **129**, 480–489 (2006).

63. Ehrenreich, H. et al. Improvement of cognitive functions in chronic schizophrenic patients by recombinant human erythropoietin. *Mol. Psychiatry* **12**, 206–220 (2007).
64. Ehrenreich, H. et al. Exploring recombinant human erythropoietin in chronic progressive multiple sclerosis. *Brain* **130**, 2577–2588 (2007).
65. Miskowiak, K., O'Sullivan, U. & Harmer, C. J. Erythropoietin enhances hippocampal response during memory retrieval in humans. *J. Neurosci.* **27**, 2788–2792 (2007).
66. Rasmussen, P. 1 et al. Effects of erythropoietin administration on cerebral metabolism and exercise capacity in men. *Appl. Physiol.* **109**, 476–483 (2010).
67. Wüstenberg, T. 1 et al. Recombinant human erythropoietin delays loss of gray matter in chronic schizophrenia. *Mol. Psychiatry* **16**, 26–36 (2011).
68. Nekoui, A. & Blaise, G. Erythropoietin and Nonhematopoietic Effects. *Am. J. Med. Sci.* **2017**, 353, 76–81 (2017).
69. Mikati, M. A., El Hokayem, J. A. & El Sabban, M. E. Effects of a single dose of erythropoietin on subsequent seizure susceptibility in rats exposed to acute hypoxia at P10. *Epilepsia* **48**, 175–181 (2007).
70. Miskowiak, K., O'Sullivan, U. & Harmer, C. J. Erythropoietin reduces neural and cognitive processing of fear in human models of antidepressant drug action. *Biol. Psychiatry* **62**, 1244–1250 (2007).
71. Miskowiak, K. W. et al. Effects of erythropoietin on memory-relevant neurocircuitry activity and recall in mood disorders. *Acta Psychiatr. Scand.* **134**, 249–259 (2016).
72. Miskowiak, K. W. et al. Neural correlates of improved executive function following erythropoietin treatment in mood disorders. *Psychol. Med.* **46**, 1679–1691 (2016).
73. Miskowiak, K. et al. Erythropoietin improves mood and modulates the cognitive and neural processing of emotion 3 days post administration. *Neuropsychopharmacology* **33**, 611–618 (2008).
74. Miskowiak, K. W. et al. Effects of erythropoietin on depressive symptoms and neurocognitive deficits in depression and bipolar disorder. *Trials* **11**, 97 (2010).
75. Ostergaard, S. D., Bech, P., Miskowiak, K. W. Fewer study participants needed to demonstrate superior antidepressant efficacy when using the Hamilton melancholia subscale (HAM-D) as outcome measure. *J. Affect. Disord.* pii: S0165-0327 00678-8 (2014).
76. Hieronymus, F., Emilsson, J. F., Nilsson, S. & Eriksson, E. Consistent superiority of selective serotonin reuptake inhibitors over placebo in reducing depressed mood in patients with major depression. *Mol. Psychiatry* **21**, 523–530 (2016).
77. Hur, E. M. & Zhou, F. Q. GSK3 signalling in neural development. *Nat. Rev. Neurosci.* **11**, 539–551 (2010).
78. Doble, B. W. & Woodgett, J. R. GSK-3: tricks of the trade for a multi-tasking kinase. *J. Cell. Sci.* **116**, 1175–1186 (2003).
79. Frame, S. & Cohen, P. GSK3 takes centre stage more than 20 years after its discovery. *Biochem. J.* **359**, 1–16 (2001).
80. Jope, R. S. & Johnson, G. V. The glamour and gloom of glycogen synthase kinase-3. *Trends Biochem. Sci.* **29**, 95–102 (2004).
81. Morales-García, J. A. et al. Glycogen synthase kinase 3 inhibition promotes adult hippocampal neurogenesis in vitro and in vivo. *ACS Chem. Neurosci.* **3**, 963–971 (2012).
82. Rayasam, G. V., Tulasi, V. K., Sodhi, R., Davis, J. A. & Ray, A. Glycogen synthase kinase 3: more than a namesake. *Br. J. Pharmacol.* **156**, 885–898 (2009).
83. Molz, S. et al. Neuroprotective effect of guanosine against glutamate-induced cell death in rat hippocampal slices is mediated by the phosphatidylinositol-3 kinase/Akt/ glycogen synthase kinase 3beta pathway activation and inducible nitric oxide synthase inhibition. *J. Neurosci. Res.* **89**, 1400–1408 (2011).
84. Shaw, P. C. et al. Isolation and chromosomal mapping of human glycogen synthase kinase-3 alpha and -3 beta encoding genes. *Genome* **41**, 720–727 (1998).
85. Kwok, J. B. et al. GSK3B polymorphisms alter transcription and splicing in Parkinson's disease. *Ann. Neurol.* **58**, 829–839 (2005).
86. Zhang, K. et al. Genetic association of the interaction between the BDNF and GSK3B genes and major depressive disorder in a Chinese population. *J. Neural Transm.* **117**, 393–401 (2010).
87. Yang, C. et al. The combined effects of the BDNF and GSK3B genes modulate the relationship between negative life events and major depressive disorder. *Brain Res.* **1355**, 1–6 (2010).
88. Ronai, Z. et al. Glycogen synthase kinase 3 beta gene structural variants as possible risk factors of bipolar depression. *Am. J. Med. Genet. B. Neuropsychiatr. Genet.* **165B**, 217–222 (2014).
89. Liu, S. et al. Possible association of the GSK3beta gene with the anxiety symptoms of major depressive disorder and P300 waveform. *Genet. Test. Mol. Biomark.* **16**, 1382–1389 (2012).
90. Serretti, A. et al. Association between GSK-3beta -50T/C polymorphism and personality and psychotic symptoms in mood disorders. *Psychiatry Res.* **158**, 132–140 (2008).
91. Saus, E. et al. A haplotype of glycogen synthase kinase 3beta is associated with early onset of unipolar major depression. *Genes. Brain. Behav.* **9**, 799–807 (2010).
92. Benedetti, F. et al. A single nucleotide polymorphism in glycogen synthase kinase 3-beta promoter gene influences onset of illness in patients affected by bipolar disorder. *Neurosci. Lett.* **355**, 37–40 (2004).
93. Yoon, H. K. & Kim, Y. K. Association between glycogen synthase kinase-3beta gene polymorphisms and major depression and suicidal behavior in a Korean population. *Prog Neuropsychopharmacol. Biol. Psychiatry* **34**, 331–334 (2010).
94. Chen, J. et al. The GSK3B gene confers risk for both major depressive disorder and schizophrenia in the Han Chinese population. *J. Affect. Disord.* **185**, 149–155 (2015).
95. Tsai, S. J., Liou, Y. J., Hong, C. J., Yu, Y. W. & Chen, T. J. Glycogen synthase kinase-3beta gene is associated with antidepressant treatment response in Chinese major depressive disorder. *Pharm. J.* **8**, 384–390 (2008).
96. Benedetti, F. et al. Long-term response to lithium salts in bipolar illness is influenced by the glycogen synthase kinase 3-beta -50 T/C SNP. *Neurosci. Lett.* **376**, 51–55 (2005).
97. Adli, M. et al. Response to lithium augmentation in depression is associated with the glycogen synthase kinase 3-beta -50T/C single nucleotide polymorphism. *Biol. Psychiatry* **62**, 1295–1302 (2007).
98. Verchinski, B. A. et al. Effects of a common variant in GSK3β on hippocampal volume in healthy human volunteers. *Human Brain Mapping* conference 2010; abstract.
99. Liu, Z. et al. A combined study of GSK3β polymorphisms and brain network topological metrics in major depressive disorder. *Psychiatry Res.* **223**, 210–217 (2014).
100. Dietsche, B. et al. Altered neural function during episodic memory encoding and retrieval in major depression. *Hum. Brain. Mapp.* **35**, 4293–4302 (2014).
101. Hall, J. et al. Hippocampal function in schizophrenia and bipolar disorder. *Psychol. Med.* **40**, 761–770 (2010).
102. Carlson, P. J., Singh, J. B., Zarate, C. A. Jr, Drevets, W. C. & Manji, H. K. Neural circuitry and neuroplasticity in mood disorders: insights for novel therapeutic targets. *NeuroRx* **3**, 22–41 (2006).
103. Duman, R. S. & Monteggia, L. M. A neurotrophic model for stress-related mood disorders. *Biol. Psychiatry* **59**, 1116–1127 (2006).
104. Miller, A. H., Maletic, V. & Raison, C. L. Inflammation and its discontents: the role of cytokines in the pathophysiology of major depression. *Biol. Psychiatry* **65**, 732–741 (2009).
105. Gold, P. W., Licinio, J. & Pavlatou, M. G. Pathological parainflammation and endoplasmic reticulum stress in depression: potential translational targets through the CNS insulin, IGF1 and PPAR-gamma systems. *Mol. Psychiatry* **18**, 154–165 (2013).
106. Borcel, E. et al. Chronic stress in adulthood followed by intermittent stress impairs spatial memory and the survival of newborn hippocampal cells in aging animals: prevention by FGL, a peptide mimetic of neural cell adhesion molecule. *Behav. Pharmacol.* **19**, 41–49 (2008).
107. Kitraki, E., Kremmyda, O., Youlatos, D., Alexis, M. & Kittas, C. Spatial performance and corticosteroid receptor status in the 21-day restraint stress paradigm. *Ann. N. Y. Acad. Sci.* **1018**, 323–327 (2004).
108. Yun, J. et al. Chronic restraint stress impairs neurogenesis and hippocampus-dependent fear memory in mice: possible involvement of a brain-specific transcription factor Npas4. *J. Neurochem.* **114**, 1840–1851 (2010).
109. Duman, R. S., Malberg, J. & Thome, J. Neural plasticity to stress and antidepressant treatment. *Biol. Psychiatry* **46**, 1181–1191 (1999).
110. Schmaal, L. et al. Subcortical brain alterations in major depressive disorder: findings from the ENIGMA Major Depressive Disorder working group. *Mol. Psychiatry* **21**, 806–812 (2016).
111. Elvsashagen, T. et al. Evidence for reduced dentate gyrus and fimbria volume in bipolar II disorder. *Bipolar Disord.* **15**, 167–176 (2013).

112. Huang, Y. et al. Structural changes in hippocampal subfields in major depressive disorder: a high-field magnetic resonance imaging study. *Biol. Psychiatry* **74**, 62–68 (2013).
113. Tae, W. S. et al. Hippocampal shape deformation in female patients with unremitting major depressive disorder. *AJNR Am. J. Neuroradiol.* **32**, 671–676 (2011).
114. Ma, R. et al. Erythropoietin protects PC12 cells from beta-amyloid(25-35)-induced apoptosis via PI3K/Akt signaling pathway. *Neuropharmacology* **56**, 1027–1034 (2009).
115. Somerville, T. C., Linch, D. C. & Khwaja, A. Growth factor withdrawal from primary human erythroid progenitors induces apoptosis through a pathway involving glycogen synthase kinase-3 and Bax. *Blood* **98**, 1374–1381 (2001).
116. Ma, S. et al. Erythropoietin rescues memory impairment in a rat model of chronic cerebral hypoperfusion via the EPO-R/JAK2/STAT5/PI3K/Akt/GSK-3 β pathway. *Mol. Neurobiol.* <https://doi.org/10.1007/s12035-017-0568-5> (2017).
117. Maurer, U., Preiss, F., Brauns-Schubert, P., Schlicher, L. & Charvet, C. GSK-3 – at the crossroads of cell death and survival. *J. Cell. Sci.* **127**, 1369–1378 (2014).
118. Hassouna, I. et al. Revisiting adult neurogenesis and the role of erythropoietin for neuronal and oligodendroglial differentiation in the hippocampus. *Mol. Psychiatry* **21**, 1752–1767 (2016).
119. Zhu, Z., Dai, J., Liao, Y. & Wang, T. Sox9 Protects against Human Lung Fibroblast Cell Apoptosis Induced by LPS through Activation of the AKT/GSK3 β Pathway. *Biochem. (Mosc.)* **82**, 606–612 (2017).
120. Topol, L., Chen, W., Song, H., Day, T. F. & Yang, Y. Sox9 inhibits Wnt signaling by promoting beta-catenin phosphorylation in the nucleus. *J. Biol. Chem.* **284**, 3323–3333 (2009).
121. Chew, L. J. et al. SRY-box containing gene 17 regulates the Wnt/ β -catenin signaling pathway in oligodendrocyte progenitor cells. *J. Neurosci.* **31**, 13921–13935 (2011).
122. Xue, Q. et al. miR-9 and miR-124 synergistically affect regulation of dendritic branching via the AKT/GSK3 β pathway by targeting Rap2a. *Sci. Rep.* **6**, 26781 (2016).
123. Roy, B., Dunbar, M., Shelton, R. C. & Dwivedi, Y. Identification of microRNA-124-3p as a putative epigenetic signature of major depressive disorder. *Neuropsychopharmacology* **42**, 864–875 (2017).
124. Higuchi, F. et al. Hippocampal MicroRNA-124 Enhances Chronic Stress Resilience in Mice. *J. Neurosci.* **36**, 7253–7267 (2016).
125. Jo, H. R., Kim, Y. S. & Son, H. Erythropoietin and carbamylated erythropoietin promote histone deacetylase 5 phosphorylation and nuclear export in rat hippocampal neurons. *Biochem. Biophys. Res. Commun.* **470**, 220–225 (2016).
126. Tong, Q. et al. TRPC3 is the erythropoietin-regulated calcium channel in human erythroid cells. *J. Biol. Chem.* **283**, 10385–10395 (2008).
127. Otori, K. et al. Ser9 phosphorylation of mitochondrial GSK-3 β is a primary mechanism of cardiomyocyte protection by erythropoietin against oxidant-induced apoptosis. *Am. J. Physiol. Heart Circ. Physiol.* **295**, H2079–H2086 (2008).
128. Nishihara, M. et al. Erythropoietin affords additional cardioprotection to preconditioned hearts by enhanced phosphorylation of glycogen synthase kinase-3 β . *Am. J. Physiol. Heart Circ. Physiol.* **291**, H748–H755 (2006).
129. Ozalp, S. S., Eren, C. Y., Bostancioglu, R. B. & Kopal, A. T. Induction of apoptosis and inhibition of cell proliferation by the cyclooxygenase enzyme blocker nimesulide in the Ishikawa endometrial cancer cell line. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **164**, 79–84 (2012).
130. Jones, K. R. & Reichardt, L. F. Molecular cloning of a human gene that is a member of the nerve growth factor family. *Proc. Natl Acad. Sci. USA* **87**, 8060–8064 (1990).
131. Numakawa, T. et al. BDNF function and intracellular signaling in neurons. *Histol. Histopathol.* **25**, 237–258 (2010).
132. Cowansage, K. K., LeDoux, J. E. & Monfils, M. H. Brain-derived neurotrophic factor: a dynamic gatekeeper of neural plasticity. *Curr. Mol. Pharmacol.* **3**, 12–29 (2010).
133. Duman, R. S. Synaptic plasticity and mood disorders. *Mol. Psychiatry* **7**, S29–S34 (2002).
134. Tyler, W. J., Perrett, S. P. & Pozzo-Miller, L. D. The role of neurotrophins in neurotransmitter release. *Neuroscientist* **8**, 524–531 (2002).
135. Mai, L., Jope, R. S. & Li, X. BDNF-mediated signal transduction is modulated by GSK3 β and mood stabilizing agents. *J. Neurochem.* **82**, 75–83 (2002).
136. Foulstone, E. J., Tavare, J. M. & Gunn-Moore, F. J. Sustained phosphorylation and activation of protein kinase B correlates with brain-derived neurotrophic factor and insulin stimulated survival of cerebellar granule cells. *Neurosci. Lett.* **264**, 125–128 (1999).
137. Oh, D. H., Park, Y. C. & Kim, S. H. Increased glycogen synthase kinase-3 β mRNA level in the hippocampus of patients with major depression: a study using the stanley neuropathology consortium integrative database. *Psychiatry Investig.* **7**, 202–207 (2010).
138. Ho, J. J., Man, H. S. & Marsden, P. A. Nitric oxide signaling in hypoxia. *J. Mol. Med. (Berl.)* **90**, 217–231 (2012).
139. Flügel, D., Görlach, A., Michiels, C. & Kietzmann, T. Glycogen synthase kinase 3 phosphorylates hypoxia-inducible factor 1 α and mediates its destabilization in a VHL-independent manner. *Mol. Cell. Biol.* **27**, 3253–3265 (2007).
140. Lie, D. C. et al. Wnt signalling regulates adult hippocampal neurogenesis. *Nature* **437**, 1370–1375 (2011).
141. Sani, G. et al. The wnt pathway in mood disorders. *Curr. Neuropharmacol.* **10**, 239–253 (2012).
142. Chong, Z. Z. 1, Hou, J., Shang, Y. C., Wang, S. & Maiese, K. EPO relies upon novel signaling of Wnt1 that requires Akt1, FoxO3a, GSK-3 β , and β -catenin to foster vascular integrity during experimental diabetes. *Curr. Neurovasc. Res.* **8**, 103–120 (2011).
143. Yang, Y. et al. Peroxisome proliferator-activated receptor γ is inhibited by histone deacetylase 4 in cortical neurons under oxidative stress. *J. Neurochem.* **118**, 429–439 (2011).
144. Ge, Z. et al. Erythropoietin alleviates hepatic insulin resistance via PPAR γ -dependent AKT activation. *Sci. Rep.* **5**, 17878 (2015).
145. Cross, D. A., Alessi, D. R., Cohen, P., Andjelkovich, M. & Hemmings, B. A. Inhibition of glycogen synthase kinase-3 by insulin mediated by protein kinase B. *Nature* **378**, 785–789 (1995).
146. Yanagita, T. et al. Neuronal insulin receptor signaling: a potential target for the treatment of cognitive and mood disorders. *Mood Disorders*. InTech. Chapter 11. <https://doi.org/10.5772/54389> (2013).
147. Beurel, E., Song, L. & Jope, R. S. Inhibition of glycogen synthase kinase-3 is necessary for the rapid antidepressant effect of ketamine in mice. *Mol. Psychiatry* **16**, 1068–1070 (2011).
148. Kim, A. J., Shi, Y., Austin, R. C. & Werstuck, G. H. Valproate protects cells from ER stress-induced lipid accumulation and apoptosis by inhibiting glycogen synthase kinase-3. *J. Cell. Sci.* **118**, 89–99 (2005).
149. Vosahlikova, M. & Svoboda, P. Lithium - therapeutic tool endowed with multiple beneficiary effects caused by multiple mechanisms. *Acta Neurobiol. Exp. (Wars.)* **76**, 1–19 (2016).
150. Frelund, L. & Beaulieu, J. M. Inhibition of GSK3 by lithium, from single molecules to signalling networks. *Front. Mol. Neurosci.* **5**, 14 (2012).
151. Malhi, G. S. & Outhred, T. Therapeutic mechanisms of lithium in bipolar disorder: recent advances and current understanding. *Cns. Drugs* **30**, 931–949 (2016).
152. Noble, W. et al. Inhibition of glycogen synthase kinase-3 by lithium correlates with reduced tauopathy and degeneration in vivo. *Proc. Natl. Acad. Sci. USA* **102**, 6990–6995 (2005).
153. Ryves, W. J. & Harwood, A. J. Lithium inhibits glycogen synthase kinase-3 by competition for magnesium. *Biochem. Biophys. Res. Commun.* **280**, 720–725 (2001).
154. Iadarola, N. D. et al. Ketamine and other N-methyl-D-aspartate receptor antagonists in the treatment of depression: a perspective review. *Ther. Adv. Chronic Dis.* **6**, 97–114 (2015).
155. Costemale-Lacoste, J. F., Guilloux, J. P. & Gaillard, R. The role of GSK-3 in treatment-resistant depression and links with the pharmacological effects of lithium and ketamine: a review of the literature. *Encephale* **42**, 156–164 (2016).
156. Yi, Z., Za, X. & Zhi, Z. Erythropoietin protects neuron against ketamine induced injuries. *88*, 876–879 (2008).
157. Hayley, S. & Litteljohn, D. Neuroplasticity and the next wave of antidepressant strategies. *Front. Cell. Neurosci.* **7**, 218 (2013).
158. Liu, J. R., Baek, C., Han, X. H., Shoureshi, P. & Soriano, S. G. Role of glycogen synthase kinase-3 β in ketamine-induced developmental neuroapoptosis in rats. *Br. J. Anaesth.* **110**, i3–i9 (2013).
159. Delimbeuf, N., Petit, A., Karila, L. & Lejoyeux, M. Ketamine: psychiatric indications and misuses. *Rev. Med. Liege* **69**, 434–440 (2014).
160. Li, Y. et al. Effects of ketamine on levels of inflammatory cytokines IL-6, IL-1 β , and TNF- α in the hippocampus of mice following acute or chronic administration. *Front. Pharmacol.* **8**, 139 (2017).
161. Yeung, L. Y. et al. Hyperphosphorylated tau in the brains of mice and monkeys with long-term administration of ketamine. *Toxicol. Lett.* **193**, 189–193 (2010).

162. Becker, A. et al. Ketamine-induced changes in rat behaviour: a possible animal model of schizophrenia. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **27**, 687–700 (2003).
163. Razoux, F., Garcia, R. & Léna, I. Ketamine, at a dose that disrupts motor behavior and latent inhibition, enhances prefrontal cortex synaptic efficacy and glutamate release in the nucleus accumbens. *Neuropsychopharmacology* **32**, 719–727 (2007).
164. Takahashi, T. et al. The effect of ketamine anesthesia on the immune function of mice with postoperative septicemia. *Anesth. Analg.* **111**, 1051–1058 (2010).
165. Ward, J. L., Harting, M. T., Jr, Cox, C. S. & Mercer, D. W. Effects of ketamine on endotoxin and traumatic brain injury induced cytokine production in the rat. *J. Trauma* **70**, 1471–1479 (2011).
166. Wang, N. et al. The rapid antidepressant effect of ketamine in rats is associated with down-regulation of pro-inflammatory cytokines in the hippocampus. *Ups. J. Med. Sci.* **120**, 241–248 (2015).
167. Sanacora, G. et al. A consensus statement on the use of ketamine in the treatment of mood disorders. *JAMA Psychiatry* **74**, 399–405 (2017).
168. Singh, I. et al. Ketamine treatment for depression: opportunities for clinical innovation and ethical foresight. *Lancet Psychiatry* **4**, 419–426 (2017).
169. Newport, D. J. et al. APA Council of Research Task Force on Novel Biomarkers and Treatments. Ketamine and other NMDA antagonists: early clinical trials and possible mechanisms in depression. *Am. J. Psychiatry* **172**, 950–966 (2015).
170. Beaulieu, J. M. et al. Role of GSK3 beta in behavioral abnormalities induced by serotonin deficiency. *Proc. Natl. Acad. Sci. USA* **105**, 1333–1338 (2008).
171. Li, X. et al. In vivo regulation of glycogen synthase kinase-3beta (GSK3beta) by serotonergic activity in mouse brain. *Neuropsychopharmacology* **29**, 1426–1431 (2004).
172. Lipoma, T. V. et al. Inhibition of glycogen synthase kinase 3 prevents synaptic long term depression and facilitates cognition in C57bl/6J mice. *Opera Med Physiol.* **2**, 87–102 (2016).
173. O'Leary, O. & Nolan, Y. Glycogen synthase kinase-3 as a therapeutic target for cognitive dysfunction in neuropsychiatric disorders. *Cns. Drugs* **29**, 1–15 (2015).
174. McCubrey, J. A. et al. GSK-3 as potential target for therapeutic intervention in cancer. *Oncotarget* **5**, 2881–2911 (2014).
175. Linding, R. et al. NetworkIN: a resource for exploring cellular phosphorylation networks. *Nucleic Acids Res.* **36**, D695–D699 (2008).
176. Baare, W. F. et al. Hippocampal volume changes in healthy subjects at risk of unipolar depression. *J. Psychiatr. Res.* **44**, 655–662 (2010).
177. Beurel, E., Mines, M., Song, L. & Jope, R. S. Glycogen synthase kinase-3 levels and phosphorylation undergo large fluctuations in mouse brain during development. *Bipolar Disord.* **14**, 822–830 (2012).
178. Orellana, A. M. et al. Age-related neuroinflammation and changes in AKT-GSK-3 β and WNT/ β -CATENIN signaling in rat hippocampus. *Aging* **7**, 1094–1108 (2015).
179. Bikkavilli, R. K. & Malbon, C. C. Mitogen-activated protein kinases and Wnt/ β -catenin signaling molecular conversations among signaling pathways. *Commun. Integr. Biol.* **2**, 46–49 (2009).
180. Santarelli, L. et al. Requirement of hippocampal neurogenesis for the behavioral effects of antidepressants. *Science* **301**, 805–809 (2003).