Open access Review

#### eGastroenterology

# Risk factors of primary liver cancer initiation associated with tumour initiating cell emergence: novel targets for promising preventive therapies

Arthur Brouillet. 1,2 Fouad Lafdil 1,2,3

**To cite:** Brouillet A, Lafdil F. Risk factors of primary liver cancer initiation associated with tumour initiating cell emergence: novel targets for promising preventive therapies. *eGastroenterology* 2023;**1**:e100010. doi:10.1136/egastro-2023-100010

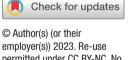
➤ Prepublication history for this paper is available online. To view these files, please visit the journal online (http://dx.doi.org/ 10.1136/egastro-2023-100010).

Received 08 February 2023 Accepted 18 March 2023

#### **ABSTRACT**

Primary liver cancers ranked as the sixth most commonly diagnosed cancers and the third-leading cause of cancerrelated death in 2020. Despite encouraging findings on diagnosis and treatments, liver cancer remains a lifethreatening disease with a still increasing incidence. Therefore, it is of interest to better characterise and understand the mechanistic process occurring at early steps of carcinogenesis. Inflammatory responses in liver diseases participate in the activation of liver progenitor cells (LPCs) facultative compartment but also to their transformation into cancer stem cells (CSCs) and give rise to primary liver cancer including hepatocellular carcinoma and cholangiocarcinoma. Higher intratumoural heterogeneity has been associated with poorer prognosis and linked to tumour escape from the immune surveillance and to resistance to chemotherapy. A better understanding of the malignant transformation of LPC as tumour initiating cells (ie, CSC) should also provide a potential new therapeutic target for anticancer therapy. In this review, we summarise the recent reports identifying underlying mechanisms by which chronic liver inflammatory responses could trigger the early steps in liver carcinogenesis, notably through the transformation of LPCs into tumour initiating cells.

#### INTRODUCTION



employer(s)) 2023. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

<sup>1</sup>UMR-S955, Université Paris-Est, Créteil, France <sup>2</sup>U955, Institut National de la Santé et de la Recherche Médicale (INSERM), Créteil, France <sup>3</sup>Institut Universitaire de France, Paris, France

#### **Correspondence to**

Professor Fouad Lafdil; fouad.lafdil@u-pec.fr

Primary liver cancers including hepatocellular carcinoma (HCC) and intrahepatic cholangiocarcinoma (CCA) ranked as the sixth most commonly diagnosed cancers and the third-leading cause of cancer-related death in 2020 (Globocan 2020). Among all primary liver cancers, HCC is the most common cancer accounting for more than 80% of cases with a 5-year survival rate of less than 10% in western countries. Despite significant progress in diagnosis and treatments, HCC that is often diagnosed at late stages (70% of cases) remains a life-threatening disease with an increasing incidence. Therefore, a better understanding of the underlying mechanisms triggering the early steps of tumourigenesis represent a great interest

to predict and propose more effective therapeutic options for liver cancer prevention.

Numerous studies have classified HCC according to gene expression profiling, immunohistological phenotypes and somatic mutation detection, revealing various patterns of HCC and next-generation sequencing analyses confirmed by high molecular heterogeneity within the same tumour nodule and various clonal evolution. 1-3 The origin of such heterogeneity is still debated. However, clinical and histological studies revealed that 28%-50% of HCC express progenitor/stem cell markers.4 In healthy livers, the progenitor compartment is composed of resident liver progenitor cells (LPCs) defined as bipotent intrahepatic quiescent cells. They are activated in chronic liver diseases in cases of massive tissue damage or prolonged chronic insult altering the proliferative capacities of remaining healthy hepatocytes, and participate in liver regeneration, fibrogenesis and tissue repair<sup>5</sup> (figure 1). LPC accumulation, known as ductular reaction, is frequently observed in diverse chronic liver diseases, such as in preneoplastic cirrhotic livers with a worse prognosis.<sup>6</sup> In addition, it is admitted that LPCs have the potential to initiate tumours because of their likelihood to transform into cancer stem cells (CSCs) that ultimately lead to the development of heterogeneous lineages of cancer cells.<sup>5</sup> 7-9 The underlying mechanisms leading LPCs to become tumour-initiating cells is not yet fully understood, but recent studies reported some promising clues that deserve to be further considered.

Due to its enriched cell composition with high density of immune cells including myeloid immune cells such as resident macrophages called Kupffer cells (KCs), neutrophils or lymphoid cells such as Natural Killer (NK), Natural Killer T (NKT), T and





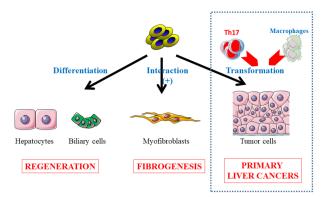


Figure 1 Liver progenitor cell compartment and properties. Liver progenitor cells are bipotent cells involved in: (1) the liver regenerative process by differentiating into hepatocytes or biliary cells; (2) fibrogenesis by tightly interacting with hepatic myofibroblasts and favouring their accumulation; (3) cancer initiation by undergoing a transforming process into cells harbouring a cancer stem cell phenotype in an inflammatory microenvironment shaped by infiltrated-Th17 cells and macrophages.

B lymphocytes, the liver is considered as an immunological organ. During chronic liver diseases whatever the aetiology, the liver is frequently subjected to a continuous regenerative process occurring in a particularly active inflammatory context.

Increasing evidence suggests that LPC expansion is carried out within a particular microenvironment requiring cellular interactions with non-parenchymal and immune cells. Chronic non-resolving inflammation and tissue damage participate not only in the activation of LPC compartment, but also regulate several mechanisms leading to their transformation into CSCs, including metabolic and epigenetic reprograming pathways.

In this review, we propose to summarise the latest findings that identified molecular inflammatory mechanisms that trigger the LPC growth, accumulation and their transformation into CSCs considered actively participating in the rise of primary liver cancers. These recent findings will allow to open discussion and thought for novel therapeutic strategies aiming at preventing liver cancer initiation.

## LPC COMPARTMENT ACTIVATION AS A RISK FACTOR FOR TRIGGERING EARLY STEPS OF LIVER CARCINOGENESIS

HCC and CCA have been considered as independent tumours that originate from distinct parenchymal cells (ie, hepatocytes vs biliary cells respectively). However, accumulating evidence suggests that both types of primary liver cancer could also originate from a common tumourinitiating cell population. Converging data demonstrated that CSCs give rise to primary liver cancers including HCC or CCA<sup>10</sup> and increasing evidence indicates that CSC are mainly responsible for tumour aggressiveness, relapse, metastasis and therapeutic failures to conventional anticancer treatments. As LPCs and CSCs were found in already established HCC<sup>4</sup> and CCA, LPCs were

defined as tumour initiating cells that ultimately evolve into heterogeneous lineages of cancer cells addressing tumour heterogeneity and plasticity in primary liver cancer. 913 A non-resolving inflammatory process observed in various chronic liver diseases could not only participate in the activation of LPC facultative compartment, but also to their transformation into CSC.<sup>8</sup> <sup>14</sup> This small subset of cancer cells acquiring robust stem cell properties within tumours could originate from a 'dedifferentiation' of non-tumour LPCs considered as normal intrahepatic cells in the liver. Another point of view explaining the accumulation of CSC is to consider that differentiation of LPC into mature parenchymal cells is impaired.<sup>8 9 13–15</sup> The underlying mechanisms leading to such CSC accumulation involve regulatory signalling pathways that induce or maintain the stemness properties in the CSC niche surrounded by stroma cells and immune cells. 16 17

### IMPACT OF SUSTAINED LIVER INFLAMMATORY RESPONSE ON LPC TRANSFORMATION INTO CSC

Chronic inflammation is known to actively participate in liver carcinogenesis by favouring tumour initiation and progression. <sup>16-18</sup> Unambiguously, immune regulation plays a key role in primary liver cancers in all chronic liver disease. <sup>19</sup> Several cytokines exert their potential mutagenic effect by the induction of genomic instability through the production of highly reactive molecules that can damage DNA. <sup>20</sup>

### Impact of liver macrophages on LPC activation and transformation into CSCs

During the liver inflammatory response, both innate and adaptive immune cells are critical for activation of the LPC compartment. KC depletion reduced LPC proliferation in mice fed with Choline-Deficient and Ethionine supplemented (CDE) diet<sup>21</sup> or in 2-acetylaminofluorene (AAF)/Partial Hepatectomy (PH) rat model, <sup>22</sup> which was associated with reduced recruitment of circulating monocytes and liver regeneration. Moreover, the proinflammatory cytokines produced by KC such as Interleukin (IL)-6 and Tumour Necrosis Factor (TNF)-α or TNF-like weak inducer of apoptosis were found to support LPC proliferation.<sup>23</sup> Long-term treatment of rat LPC WB-F344 cells by TNF-\alpha (but not IL-6) promotes spheroid formation in vitro and tumour after subcutaneous inoculation into immunodeficient NOD/SCID mice. Aberrant expression of LPC markers such as Alpha-fetoprotein (AFP), Cytokeratin (CK)-19 and Ovalbumin (OV) 6 was detected in tumours from TNFα-treated WB-F344 cells revealing their malignant transformation.<sup>8</sup> Interestingly, chronic TNF- $\alpha$ exposure was also able to trigger chromosomal instability due to an aberrant expression of ubiquitin D and checkpoint kinase 2. Incorporation of the KrasG12D mutation into murine LPC increased their proliferative capacity and their capability to form colonies in vitro, revealing a mechanism of LPC transformation by horizontal transfer of oncogene.<sup>24</sup>



Furthermore, in the context of an already established tumour immune environment, tumour-associated macrophages (TAMs) produce cytokines which induce CSC accumulation, <sup>25</sup> such as IL-6 and TNF- $\alpha$  through the STAT3 signalling pathway<sup>8</sup> or by inducing Epithelio-Mesenchymal Transition (EMT) and stemness features via the Wnt/ $\beta$ -catenin pathway. <sup>26</sup>

### Involvement of lymphocytes on LPC-driven transformation into CSCs

Activation of LPC compartment was drastically weakened in mice lacking T cells<sup>27</sup> and T-cell-mediated hepatitis induced by concanavalin A triggers NK cell-sensitive LPC expansion following partial hepatectomy.<sup>28</sup> Moreover, other inflammatory-related proteins including lymphotoxin  $\beta$ , interferon- $\gamma$ , IL-22, galectin-3 promote liver regeneration from LPCs. <sup>23</sup> <sup>29</sup> <sup>30</sup> In patients with various chronic liver diseases, LPC accumulation correlates with the recruitment of IL-17 producing cells and the severity of ductular reaction.<sup>31</sup> Among the key players in modulating liver inflammation, T helper-(Th)17 lymphocytes have been implicated in several types of liver diseases and the proinflammatory IL-17 cytokine involved in the crosstalk between innate and adaptive immunity.<sup>32</sup> The key role of IL-17 in LPC activation was evidenced in an experimental model of mice fed with a CDE diet, showing that IL-17 is responsible for macrophage-induced IL-27 expression that favours LPC differentiation into hepatocytes. 31 These results highlight collaborative work between IL-17 and IL-27 that is required to properly achieve liver regeneration from LPC.

In the context of inflammation-induced tumourigenesis, IL-17 is a crucial cytokine produced by immune cells, especially activated T-helper 17 cells, which contribute to the initiation and progression of several cancers through activation of stem/progenitor compartment in gastrointestinal, <sup>33</sup> skin, <sup>34</sup> ovarian, <sup>35</sup> pancreatic, <sup>36</sup> breast <sup>37</sup> and prostate <sup>38</sup> cancers. In the liver, the proinflammatory IL-17 cytokine drives progression of steatohepatitis, 39 hepatic fibrosis and LPC expansion<sup>31</sup> which are a known risk factor of HCC development. 40 41 The tumour-promoting inflammatory impact of IL-17 was recently confirmed on a cohort of 404 patients with cirrhosis, where plasma IL-17 and AFP combination effectively predicts imminent HCC occurrence within a year. 42 Recently, IL-17-producing cells were localised within ductular reaction close to LPC and CSC in human preneoplastic cirrhotic livers from diverse aetiologies. 43 The direct effect of IL-17 on LPC transformation was evidenced in vitro with long-term IL-17 stimulation of murine and human LPCs, which promotes expression of CSC (such as Cd133, EpCAM) and tumour (alpha-fetoprotein and glypican-3) markers and acquired self-renewal capacity highlighted by spheroid formation. These data demonstrated that chronic exposure to IL-17 induces the conversion of LPCs into cells acquiring CSC phenotype. The potential tumourigenic effect of IL-17 was confirmed in

vivo by increased expansion of the tumour mass after a subcutaneous engraftment of IL17-pretreated LPCs into immunodeficient NOD/SCID mice, while no significant cell proliferation was observed in control mice engrafted with non-pretreated LPC. Interestingly when compared with untreated LPC, tumours from IL-17-pretreated LPC exhibited an aggressive phenotype with histopathological features of mixed HCC and CCA phenotype, in accordance with malignant transformation of bipotential LPC.

Gamma delta ( $\gamma\delta$ ) T-cells represent a major T-cell population IL-17 participate in IL-17 production<sup>23</sup> have been reported to promote tissue repair by interacting with tissue-resident stem cells and participate in HCC.<sup>44</sup> Moreover, lymphatic endothelial cells could secrete IL-17 to potentiate the self-renewal and tumourigenesis properties of CSCs by upregulating programmed cell deathligand 1 (PD-L1) expression. Then, IL-17 may participate in hepatocarcinogenesis by driving the transformation of LPCs into CSCs and favouring their ability to escape from antitumour immune response.

### THE ROLE OF NON-CODING RNAS IN CONTROLLING LPC TRANSFORMATION INTO CSCs

non-coding RNAs including micro-RNAs (miRNAs) are actively involved in both physiological and pathophysiological processes in the liver. In chronic liver diseases, the alteration of miRNAs expression is associated with metabolic syndrome, liver injury and fibrogenesis, and to tumourigenesis, making theses miRNAs attractive targets for the diagnosis and treatment of liver diseases. 45 A dysregulated expression of numerous miRNAs has been identified as key component in the carcinogenic process. Indeed, they harbour a high capacity of regulating many genes involved in tumour cell expansion, and regulating immune response.46 Recent studies also reported miRNA as potential biomarkers. 47 Among the downregulated miRNAs in HCC, miR-122 which is the most abundant miRNA in the normal liver is downregulated in almost 50% of diagnosed HCC patients.  $^{48\,49}$ 

### Impact of the inflammatory response on epigenetic cell reprograming

The role of IL-17 in driving the tumour-initiating abilities of CSC includes epigenetic alteration that was identified in carcinogenesis. Interestingly in IL-17-treated LPC, expression of miR-122 which accounts for 70% of the liver's total miRNAs that was considered as a tumour suppressor miRNA, was sharply decreased in experiments performed in vitro and in vivo. Recently, long-term exposure to IL-17 cytokine has been shown to strongly downregulate miR-122 expression in LPCs allowing their dedifferentiation and transformation into CSCs with increased risk of primary liver cancer. In addition, transfection of miR-122 mimic into LPC was sufficient to abolish their acquired self-renewal



capacities by long-term-IL-17 pretreatment. The mechanism by which IL-17 reduces miR-122 levels could be the consequence of the STAT3-mediated inhibition of Hepatocyte Nuclear Factor (HNF) 4- $\alpha$  expression <sup>31</sup> that is involved in the transcriptional regulation of mir-122. <sup>52</sup>

### Non-coding RNA transfer through exosomes as a significant mean of cell communication controlling HCC development

Exosomes are micro or nano extracellular vesicles that originate from cell membranes and able to influence the proliferative and migration rates of cancer cells. Depending on their cargo, exosomes can either suppress or promote tumour cell progression. They can act as means of communication between cells by transferring their contain in miRNAs, long non-coding RNAs (lncRNAs) or circular RNAs (circRNAs) and modulate other molecular signalling pathways such as PTEN and PI3K/Akt in cancer.

Exosomal miRNA has been described reflecting the miRNA expressed by tumour cells and for this reason, it has been considered as plasma biomarkers for cancers. In the liver, various elevated levels of exosomal miRNA were positively or negatively associated with HCC progression, including miR-122.<sup>53'54</sup> Numerous studies revealed that exosomal miRNA participate in hepatocarcinogenesis by regulating cell proliferation, immune response and EMT.<sup>55</sup> For instance, HCC cell-derived exosomes containing miR-21 lead to tumour cell expansion through the inhibition of PTENp1 and PTEN expression.<sup>56</sup> MiR-142 and miR-223 expressed in macrophages were found to be transferred by exosomes in HCC cells and affected posttranscriptional regulation of proteins leading to the inhibition of the tumour cell growth.<sup>57</sup> In a different way of communication, tumour-derived exosomes were shown to possibly activate Dendritic Cells (DCs), or to induce the proliferation of immature T cells, and their differentiation into an antigen-specific cytotoxic T lymphocytes (CTLs) phenotype, that ultimately allow increasing the antitumour immune response. 58 59 In ectopic and orthotopic mouse models of HCC growth, it has been reported that treatment with dendritic cells pulsed with tumour cell-derived exosomes was able to trigger a strong immune response characterised by an increased number of T cells and in interferon gamma (IFN-γ) production, and in parallel, by a decrease in IL-10 and Transforming Growth Factor (TGF)-β, resulting in a significant decrease in tumour growth.<sup>60</sup>

Extracellular vesicles coming from neutrophils were shown to transfer their contain to hepatocytes. For instance, miR-223 were reported to prevent Non-Alcoholic Fatty Liver Diseases (NAFLD) and Non-Alcoholic Steatohepatitis (NASH) progression via hepatocyte uptake of neutrophil-derived extracellular vesicles that consequently exerted its anti-inflammatory and antifibrotic properties, <sup>61</sup> making it a potential therapeutic target for NASH and liver cancer treatments. Recently, it was demonstrated in two models

of inflammation-associated HCC that miR-223 attenuates hypoxia-induced tumour immunosuppression and angiogenesis in HCC via the inhibition of hypoxia-inducible factor  $1\alpha$ . Then, gene delivery of miR-223 via adenovirus in hepatocytes may improve the efficacy of the current therapy for HCC with PD-1/PD-L1 inhibitors combined with antiangiogenic agents.  $^{62}$ 

In a cohort of 112 HCC patients compared with healthy individuals, 31 miRNAs, 4 lncRNAs and 2 circRNAs were identified in exosomes and are considered as biomarkers to predict HCC prognosis. <sup>63</sup> In a TCGA analysis from 371 HCC samples, 5 exosome-related lncRNA were associated with poor outcome and also related to overexpressed immune checkpoints. <sup>64</sup> This lncRNA-exosomal signature may help to propose an adapted and personalised immunotherapy for each HCC patient. Interestingly, one of these lncRNA, muskelin 1 antisense RNA (MKLN1-AS), was shown to promote tumour growth <sup>65</sup> or drive EMT of liver cancer cells. <sup>66</sup>

Then, altered expression of non-coding RNAs are being investigated as biomarkers for early diagnosis and prognosis of primary liver cancers. Their implication on intercellular communication via extracellular vesicles between parenchymal and LPC with immune cells and liver CSC niche reveals their usefulness as therapeutic targets in liver carcinogenesis process.<sup>45</sup> Accumulating data have demonstrated that various miRNAs/lncRNAs/circRNAs regulate activation of LPC compartment<sup>67-69</sup> and are essential for maintaining CSC properties in liver cancer. 70 71 Although the involvement of non-coding RNAs is well described in liver injury and liver fibrosis, the exact role of miRNAs/ lncRNAs/circRNAs in the initiation and promotion of hepatocarcinogenesis has rarely been explored, notably tumours resulting in malignant transformation of LPCs, including the capacity of progenitor cell dedifferentiation to form CSCs (figure 2).

### PROMISING THERAPEUTIC STRATEGIES FOR LIVER CANCER PREVENTION

In terms of therapy research aiming at preventing liver cancer occurrence, maintaining LPC compartment integrity by limiting their activation/transformation could be considered as attractive therapeutic strategy. In addition to the inflammatory microenvironment related to the chronic liver diseases, the exosome-mediated bidirectional communication between tumour cells and their microenvironment brings novel insight to decipher the underlying molecular mechanisms involved in early steps of liver cancer development.

#### **Targeting immunomodulatory components**

In that context, recent studies evidenced the tumourigenic potential of proinflammatory cytokines including IL-6, TNF- $\alpha$  and IL-17 as presented above (figure 2). Therefore, targeting and inhibiting the release or action

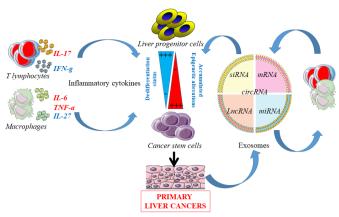


Figure 2 Transforming process of liver progenitor cells (LPCs) into cancer stem cells (CSCs). In a chronic inflammatory context, epigenetic changes occur in LPCs: (1) through a sustained stimulation by cytokines that either promotes dedifferentiation and transformation of LPCs to CSCs (IL-17, TNF-α and IL-6), or in contrast avoiding their transformation (IFN-γ, IL-27); (2) by hosting small non-coding RNAs transferred by exosomes coming from already established tumour cells or immune cells composing the microenvironment with a possible bidirectional communication. circRNA, circular RNA; IFN, interferon; IL, interleukin; LncRNA, long non-coding RNA; miRNA, microRNA; mRNA, messenger RNA; TNF, tumour necrosis factor; siRNA, small interference RNA.

of such cytokines could be considered to prevent LPC acquirement of tumour-initiating phenotype.

For instance, in a murine HCC xenograft model, the use of anti-IL-6 tocilizumab antibody showed reduced HCC growth and prevented the occurrence of CSCs. <sup>25</sup>

In same lines of evidence, clinically TNF- $\alpha$  expression was correlated to LPCs activation and HCC recurrence. In an experimental model of TNF- $\alpha$ -inhibition or deletion, LPC activation and proliferation were inhibited. At last, preclinical benefits of anti-TNF- $\alpha$  treatments have also been reported for multiple types of cancers, including HCC.

In addition, in a murine model of HCC developed on a fibrotic context, recapitulating the clinical and histological features reported in human, IL-17-deficiency or anti-IL-17 therapy was shown to reduce liver tumour growth by repressing LPC transformation into CSCs.  $^{43}$  Interestingly, several ongoing clinical trials using anti-IL17 or anti-TNF- $\alpha$  therapies to treat autoimmune diseases including psoriasis,  $^{74}$  showed a well-tolerated treatment suggesting their potential use for other diseases to prevent or cure HCC.

#### **Targeting exosomal transfer**

Exosomes not only are considered as biomarkers for HCC but also could constitute a new target for HCC treatment. Numerous studies revealed that HCC-derived exosomes elicit HCC initiation and progression. Then, inhibiting the release of exosomal oncomiR or increasing the effect of tumour suppressor miRNA may constitute a novel strategy to treat HCC initiation and also progression. For

example, blockade of miR-23a-3p in HCC cells prevent their exosomal release and upregulation of PD-L1 expression in macrophages, which plays an important role in tumour cell escape from antitumour immunity.<sup>75</sup> Similarly, targeting exosomal miRNA released from HCC cells which stimulate angiogenesis<sup>76</sup> or favouring tumour immune escape<sup>77</sup> may provide new therapeutic tools in HCC treatment.

Origin of exosomes from other cell types could carry and effectively deliver therapeutic miRNAs. For example, mesenchymal stem cells (MSCs) are known to produce large amounts of exosomes and their transfection with miR-122 can effectively package this liver-specific miRNA in released exosomes. Consequently, the transfer of miR-122 from adipose tissue-derived MSCs into recipient HCC cells was successfully observed, rendering cancer cells sensitive to chemotherapeutic agents. Additionally, intratumour injection of miR122-exosomes significantly increased the antitumour efficacy of sorafenib on HCC in vivo. 78 Similar result was obtained with lentivirus-mediated transfection of miR-199a by increasing HCC sensitivity to doxorubicin in vitro and in vivo through inhibiting the mTOR pathway.<sup>79</sup> Among stromal cells, cancer-associated fibroblasts (CAF) can also transfer miRNA to HCC and it was demonstrated that  $miR-320a^{80}$  and  $miR-150-3p^{81}$ could function as an antitumour miRNA. Then, transfer of these CAF-derived tumour suppressor miRNA might be a potential treatment option to inhibit HCC progression. Similarly, delivery of miR-214 which function as a tumour suppressor in HCC by human endothelial cellderived exosomes, in combination with anticancer drugs (sorafenib and oxaliplatin) reduces the viability and invasion of HCC cells compared with monotherapy.<sup>82</sup> On the contrary, transfer of oncomiR 1228-3p from CAFderived exosomes into HCC cells increases their resistance to sorafenib,83 requiring to propose a targeting of this exosomal miRNA. Thus, exosomes considered as biomarkers for HCC could be used as an efficient tool to target and control the tumour cell development. Taken together, these recent studies in early steps of tumourigenesis, cell-derived exosomes may become an important tool for not only early cancer diagnosis but also useful for therapeutic drug delivery.

#### **CONCLUSION**

Primary liver cancers develop mainly in an inflammatory context evidenced in virtually all chronic liver diseases. Despite significant advances in liver cancer diagnosis and therapies, the current anticancer treatments remain poorly effective in advanced stages of the disease over the past decade. Chronic non-resolving inflammation drives malignant initiation, tumour growth by increasing cancer stemness (self-renewal, EMT, chromosomal instability, immune escape), cancer metastasis and recurrence. Implication of LPCs in carcinogenic processes may enlighten the plasticity of these cells with high capacity to self-renew and their putative malignant features responsible for



genetic heterogeneity observed in tumour development. Dissecting and evaluating the contribution of stromal and immune cells allows understanding the intricated crosstalk between these cells localised in CSC niche, which control the LPC transformation into CSCs and giving rise to HCC and CCA. In this inflammatory context, Th17 cells secreting IL-17 are crucial components among infiltrating immune cells that drives transformation of LPC into CSC. Therefore, treatment aiming at neutralising IL-17 production in combination with other therapeutic strategies (targeting angiogenesis, immune escape, metastasis) may constitute a novel strategy for CSC eradication and could prevent liver cancer initiation from LPC origin.

**Contributors** AB and FL wrote the manuscript. FL designed the review. Both authors approved the final manuscript.

**Funding** The authors have not declared a specific grant for this research from any funding agency in the public, commercial or not-for-profit sectors.

Competing interests None declared.

Patient and public involvement Patients and/or the public were not involved in the design, or conduct, or reporting, or dissemination plans of this research.

Patient consent for publication Not applicable.

Provenance and peer review Not commissioned; externally peer reviewed.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.

#### **REFERENCES**

- 1 Calderaro J, Couchy G, Imbeaud S, et al. Histological subtypes of hepatocellular carcinoma are related to gene mutations and molecular tumour classification. J Hepatol 2017;67:727–38.
- 2 Khaliq M, Ko S, Liu Y, et al. Stat3 regulates liver progenitor celldriven liver regeneration in Zebrafish. Gene Expr 2018;18:157–70.
- 3 Ringelhan M, Pfister D, O'Connor T, et al. The immunology of hepatocellular carcinoma. *Nat Immunol* 2018;19:222–32.
- 4 Durnez A, Verslype C, Nevens F, et al. The clinicopathological and prognostic relevance of cytokeratin 7 and 19 expression in hepatocellular carcinoma. A possible progenitor cell origin. *Histopathology* 2006;49:138–51.
- 5 Lukacs-Kornek V, Lammert F. The progenitor cell dilemma: cellular and functional heterogeneity in assistance or escalation of liver injury. J Hepatol 2017;66:619–30.
- 6 Hou X-J, Ye F, Li X-Y, et al. Immune response involved in liver damage and the activation of hepatic progenitor cells during liver tumorigenesis. Cell Immunol 2018;326:52–9.
- 7 Akin A, Akin U. The mediating role of social safeness on the relationship between Facebook(®) use and life satisfaction. *Psychol Rep* 2015;117:341–53.
- 8 Li X-F, Chen C, Xiang D-M, et al. Chronic inflammation-elicited liver progenitor cell conversion to liver cancer stem cell with clinical significance. *Hepatology* 2017;66:1934–51.
- 9 Sia D, Villanueva A, Friedman SL, et al. Liver cancer cell of origin, molecular class, and effects on patient prognosis. Gastroenterology 2017;152:745–61.
- 10 Ko S, Russell JO, Molina LM, et al. Liver progenitors and adult cell plasticity in hepatic injury and repair: knowns and unknowns. Annu Rev Pathol 2020;15:23–50.
- 11 Correnti M, Binatti E, Gammella E, et al. The emerging role of tumor microenvironmental stimuli in regulating metabolic rewiring of liver cancer stem cells. Cancers (Basel) 2022;15:5.
- 12 Komuta M, Spee B, Vander Borght S, et al. Clinicopathological study on cholangiolocellular carcinoma suggesting hepatic progenitor cell origin. Hepatology 2008;47:1544–56.
- 13 Batlle E, Clevers H. Cancer stem cells revisited. Nat Med 2017;23:1124–34.

- 14 Fang X, Yan Q, Liu S, et al. Cancer stem cells in hepatocellular carcinoma: intrinsic and extrinsic molecular mechanisms in stemness regulation. Int J Mol Sci 2022;23:12327.
- 15 Thorgeirsson SS. Stemness and reprogramming in liver cancer. Hepatology 2016;63:1068–70.
- Müller L, Tunger A, Plesca I, et al. Bidirectional crosstalk between cancer stem cells and immune cell subsets. Front Immunol 2020:11:140
- 17 Oshimori N, Guo Y, Taniguchi S. An emerging role for cellular crosstalk in the cancer stem cell niche. J Pathol 2021;254:384–94.
- 18 Yu L-X, Ling Y, Wang H-Y. Role of nonresolving inflammation in hepatocellular carcinoma development and progression. NPJ Precis Oncol 2018;2:6.
- 19 Leone V, Ali A, Weber A, et al. Liver inflammation and hepatobiliary cancers. Trends Cancer 2021;7:606–23.
- 20 Neganova M, Liu J, Aleksandrova Y, et al. Therapeutic influence on important targets associated with chronic inflammation and oxidative stress in cancer treatment. Cancers (Basel) 2021;13:6062.
- 21 Elsegood CL, Chan CW, Degli-Esposti MA, et al. Kupffer cell-monocyte communication is essential for initiating murine liver progenitor cell-mediated liver regeneration. Hepatology 2015:62:1272–84
- 22 Xiang S, Dong H-H, Liang H-F, et al. Oval cell response is attenuated by depletion of liver resident macrophages in the 2-AAF/partial hepatectomy rat. PLoS One 2012;7:e35180.
- 23 So J, Kim A, Lee S-H, et al. Liver progenitor cell-driven liver regeneration. Exp Mol Med 2020;52:1230–8.
- 24 Olivera-Salazar R, García-Arranz M, Sánchez A, et al. Oncological transformation in vitro of hepatic progenitor cell lines isolated from adult mice. Sci Rep 2022;12:3149.
- 25 Wan S, Zhao E, Kryczek I, et al. Tumor-associated macrophages produce interleukin 6 and signal via STAT3 to promote expansion of human hepatocellular carcinoma stem cells. Gastroenterology 2014;147:1393–404.
- 26 Chen Y, Wen H, Zhou C, et al. TNF-A derived from M2 tumor-associated macrophages promotes epithelial-mesenchymal transition and cancer stemness through the WNT/B-catenin pathway in SMMC-7721 hepatocellular carcinoma cells. Exp Cell Res 2019:378:41–50
- 27 Strick-Marchand H, Masse GX, Weiss MC, et al. Lymphocytes support oval cell-dependent liver regeneration. J Immunol 2008;181:2764–71.
- 28 Hines IN, Kremer M, Isayama F, et al. Impaired liver regeneration and increased oval cell numbers following T cell-mediated hepatitis. Hepatology 2007;46:229–41.
- 29 Feng D, Kong X, Weng H, et al. Interleukin-22 promotes proliferation of liver stem/progenitor cells in mice and patients with chronic hepatitis B virus infection. Gastroenterology 2012;143:188–98.
- 30 Hsieh W-C, Mackinnon AC, Lu W-Y, et al. Galectin-3 regulates hepatic progenitor cell expansion during liver injury. Gut 2015;64:312–21.
- 31 Guillot A, Gasmi I, Brouillet A, et al. Interleukins-17 and 27 promote liver regeneration by sequentially inducing progenitor cell expansion and differentiation. *Hepatol Commun* 2018;2:329–43.
- 32 Lafdil F, Miller AM, Ki SH, et al. TH17 cells and their associated cytokines in liver diseases. Cell Mol Immunol 2010;7:250–4.
- 33 Jiang Y-X, Yang S-W, Li P-A, et al. The promotion of the transformation of quiescent gastric cancer stem cells by IL-17 and the underlying mechanisms. Oncogene 2017;36:1256–64.
- 34 Wu L, Chen X, Zhao J, et al. A novel IL-17 signaling pathway controlling keratinocyte proliferation and tumorigenesis via the Traf4-Erk5 axis. J Exp Med 2015;212:1571–87.
- 35 Xiang T, Long H, He L, et al. Interleukin-17 produced by tumor microenvironment promotes self-renewal of CD133+ cancer stemlike cells in ovarian cancer. Oncogene 2015;34:165–76.
- 36 Zhang Y, Zoltan M, Riquelme E, et al. Immune cell production of interleukin 17 induces stem cell features of pancreatic intraepithelial neoplasia cells. Gastroenterology 2018;155:210–23.
- 37 Mombelli S, Cochaud S, Merrouche Y, et al. IL-17A and its Homologs IL-25/IL-17E recruit the C-RAF/S6 kinase pathway and the generation of pro-oncogenic LMW-E in breast cancer cells. Sci Rep 2015;5:11874.
- 38 Zhang Q, Liu S, Parajuli KR, et al. Interleukin-17 promotes prostate cancer via MMP7-induced epithelial-to-mesenchymal transition. Oncogene 2017;36:687–99.
- 39 Ma H-Y, Yamamoto G, Xu J, et al. IL-17 signaling in steatotic hepatocytes and macrophages promotes hepatocellular carcinoma in alcohol-related liver disease. J Hepatol 2020;72:946–59.
- 40 Drescher HK, Bartsch LM, Weiskirchen S, et al. Intrahepatic TH17/ TReg cells in homeostasis and disease-it's all about the balance. Front Pharmacol 2020;11:588436.



- 41 Zhao J, Chen X, Herjan T, et al. The role of Interleukin-17 in tumor development and progression. J Exp Med 2020;217:e20190297.
- 42 Liang K-H, Lai M-W, Lin Y-H, et al. Plasma interleukin-17 and alphafetoprotein combination effectively predicts imminent hepatocellular carcinoma occurrence in liver cirrhotic patients. BMC Gastroenterol 2021;21:177.
- 43 Gasmi I, Machou C, Rodrigues A, et al. Interleukin-17 programs liver progenitor cell transformation into cancer stem cells through miR-122 downregulation with increased risk of primary liver cancer initiation. Int J Biol Sci 2022;18:1944–60.
- 44 Ko S, Shin D. Chemical screening using a Zebrafish model for liver progenitor cell-driven liver regeneration. *Methods Mol Biol* 2019;1905:83–90.
- 45 Wang X, He Y, Mackowiak B, et al. Micrornas as regulators, biomarkers and therapeutic targets in liver diseases. Gut 2021:70:784–95.
- 46 Berindan-Neagoe I, Monroig P del C, Pasculli B, et al. Micrornaome genome: a treasure for cancer diagnosis and therapy. CA Cancer J Clin 2014;64:311–36.
- 47 Yang J, Ma D, Fesler A, et al. Expression analysis of microRNA as prognostic biomarkers in colorectal cancer. Oncotarget 2017;8:52403–12.
- 48 Hou J, Lin L, Zhou W, et al. Identification of miRNomes in human liver and hepatocellular carcinoma reveals miR-199A/B-3P as therapeutic target for hepatocellular carcinoma. Cancer Cell 2011;19:232–43.
- 49 Chun KH. Molecular targets and signaling pathways of microRNA-122 in hepatocellular carcinoma. *Pharmaceutics* 2022;14:1380.
- 50 Gan X, Dai Z, Ge C, et al. FTO promotes liver inflammation by suppressing M6A mRNA methylation of IL-17Ra. Front Oncol 2022;12:989353.
- 51 Zeisel MB, Pfeffer S, Baumert TF. miR-122 acts as a tumor suppressor in hepatocarcinogenesis in vivo. J Hepatol 2013;58:821–3.
- 52 Aydin Y, Kurt R, Song K, et al. Hepatic stress response in HCV infection promotes STAT3-mediated inhibition of HNF4A-miR-122 feedback loop in liver fibrosis and cancer progression. Cancers (Basel) 2019;11:1407.
- 53 Wang Y, Zhang C, Zhang P, et al. Serum exosomal microRNAs combined with alpha-fetoprotein as diagnostic markers of hepatocellular carcinoma. Cancer Med 2018;7:1670–9.
- 54 Xue X, Zhao Y, Wang X, et al. Development and validation of serum exosomal microRNAs as diagnostic and prognostic biomarkers for hepatocellular carcinoma. J Cell Biochem 2019;120:135–42.
- 55 Zelli V, Compagnoni C, Capelli R, et al. Role of exosomal microRNAs in cancer therapy and drug resistance mechanisms: focus on hepatocellular carcinoma. Front Oncol 2022;12:940056.
- 56 Li X, Li X, Zhang B, et al. The role of cancer stem cell-derived exosomes in cancer progression. Stem Cells Int 2022;2022:9133658.
- 57 Aucher A, Rudnicka D, Davis DM. Micrornas transfer from human macrophages to hepato-carcinoma cells and inhibit proliferation. J Immunol 2013;191:6250–60.
- 58 Li J, Huang S, Zhou Z, et al. Exosomes derived from rAAV/AFP-transfected dendritic cells elicit specific T cell-mediated immune responses against hepatocellular carcinoma. Cancer Manag Res 2018;10:4945–57.
- 59 Chen R, Xu X, Tao Y, et al. Exosomes in hepatocellular carcinoma: a new horizon. Cell Commun Signal 2019;17:1.
- Rao Q, Zuo B, Lu Z, et al. Tumor-derived exosomes elicit tumor suppression in murine hepatocellular carcinoma models and humans in vitro. Hepatology 2016;64:456–72.
  He Y, Rodrigues RM, Wang X, et al. Neutrophil-to-hepatocyte
- 61 He Y, Rodrigues RM, Wang X, et al. Neutrophil-to-nepatocyte communication via LDLR-dependent miR-223-enriched extracellular vesicle transfer ameliorates nonalcoholic steatohepatitis. J Clin Invest 2021;131:e141513.
- 62 Fu Y, Mackowiak B, Feng D, et al. Microrna-223 attenuates Hepatocarcinogenesis by blocking hypoxia-driven angiogenesis and immunosuppression. Gut 2023. 10.1136/gutjnl-2022-327924 [Epub ahead of print 2 Jan 2023].

- 63 Tao Q, Zhu K, Zhan Y, et al. Construction of a novel exosomesrelated gene signature in hepatocellular carcinoma. Front Cell Dev Biol 2022;10:997734.
- 64 Su D, Zhang Z, Xu Z, et al. A Prognostic exosome-related LncRNA risk model correlates with the immune microenvironment in liver cancer. Front Genet 2022;13:965329.
- 65 Pan G, Zhang J, You F, et al. ETS proto-oncogene 1-activated muskelin 1 antisense RNA drives the malignant progression of hepatocellular carcinoma by targeting miR-22-3p to Upregulate ETS proto-oncogene 1. Bioengineered 2022;13:1346–58.
- 66 Guo C, Zhou S, Yi W, et al. SOX9/MKLN1-AS axis induces hepatocellular carcinoma proliferation and epithelial-mesenchymal transition. *Biochem Genet* 2022;60:1914–33.
- 67 Ruan Z, Lai M, Shang L, et al. Regulation of long non-coding RNA-Dreh involved in proliferation and migration of hepatic progenitor cells during liver regeneration in rats. Int J Mol Sci 2019;20:2549.
- 68 Jung KH, McCarthy RL, Zhou C, et al. Microrna regulates hepatocytic differentiation of progenitor cells by targeting YAP1. Stem Cells 2016;34:1284–96.
- 69 Yan Y, Wang R, Hu X, et al. MiR-126 regulates properties of SOX9<sup>+</sup> liver progenitor cells during liver repair by targeting Hoxb6. Stem Cell Reports 2020;15:706–20.
- 70 Li L, Xun C, Yu CH. Role of microRNA-regulated cancer stem cells in recurrent hepatocellular carcinoma. World J Hepatol 2022:14:1985–96.
- 71 Lv H, Lv G, Han Q, et al. Noncoding RNAs in liver cancer stem cells: the big impact of little things. Cancer Lett 2018;418:51–63.
- 72 Jing Y, Sun K, Liu W, et al. Tumor necrosis factor-A promotes hepatocellular carcinogenesis through the activation of hepatic progenitor cells. *Cancer Lett* 2018;434:22–32.
- 73 Li W, Jian YB. Antitumor necrosis factor-alpha antibodies as a noveltherapy for hepatocellular carcinoma. Exp Ther Med 2018:16:529–36.
- 74 Dong Q, Li D, Xie BB, et al. IL-17A and TNF-alpha inhibitors induce multiple molecular changes in psoriasis. Front Immunol 2022;13:1015182
- 75 Liu J, Fan L, Yu H, et al. Endoplasmic reticulum stress causes liver cancer cells to release exosomal miR-23A-3P and up-regulate programmed death ligand 1 expression in macrophages. *Hepatology* 2019:70:241–58
- 76 Moh-Moh-Aung A, Fujisawa M, Ito S, et al. Decreased miR-200B-3P in cancer cells leads to angiogenesis in HCC by enhancing endothelial ERG expression. Sci Rep 2020:10:10418.
- 77 Guan M-C, Wang M-D, Wang W-Y, et al. Exosomes as mediators of tumor immune escape and immunotherapy in hepatocellular carcinoma. Liver Research 2022;6:132–8.
- 78 Lou G, Song X, Yang F, et al. Exosomes derived from miR-122-modified adipose tissue-derived Mscs increase chemosensitivity of hepatocellular carcinoma. J Hematol Oncol 2015;8:122.
- 79 Lou G, Chen L, Xia C, et al. Mir-199A-modified exosomes from adipose tissue-derived mesenchymal stem cells improve hepatocellular carcinoma chemosensitivity through mTOR pathway. J Exp Clin Cancer Res 2020;39:4.
- 80 Zhang Z, Li X, Sun W, et al. Loss of exosomal miR-320A from cancer-associated fibroblasts contributes to HCC proliferation and metastasis. Cancer Lett 2017;397:33–42.
- 81 Yugawa K, Yoshizumi T, Mano Y, et al. Cancer-associated fibroblasts promote hepatocellular carcinoma progression through downregulation of exosomal miR-150-3p. Eur J Surg Oncol 2021;47:384–93.
- 82 Semaan L, Zeng Q, Lu Y, et al. Microrna-214 enriched exosomes from human cerebral endothelial cells (hCEC) sensitize hepatocellular carcinoma to anti-cancer drugs. *Oncotarget* 2021;12:185–98.
- 83 Zhang Y, Pan Q, Shao Z. Extracellular vesicles derived from cancerassociated fibroblasts carry tumor-promotive microRNA-1228-3p to enhance the resistance of hepatocellular carcinoma cells to sorafenib. *Hum Cell* 2023;36:296–311.