Guest Editor: D. F. Mureşanu

Basic mechanisms of neurodegeneration: a critical update

Kurt A. Jellinger *

Institute of Clinical Neurobiology, Kenyongasse, Vienna, Austria

Received: November 6, 2009; Accepted: December 18, 2009

- Introduction
- Protein aggregation
- 'Toxic oligomer' hypothesis
- Protein (mis)folding
- Proteostasis and molecular chaperones
- Protein misfolding and endoplasmic reticulum stress
- Unfolded protein response
- Interaction of proteins
- The ubiquitin proteasome system
- Autophagy and neurodegeneration
- UPS in neurodegenerative disorders
- Aggresomes
- Oxidative injury
 - Metals and oxidative stress
 - Oxidative stress in Alzheimer disease

- Oxidative stress in Parkinson disease
- Oxidative stress in other neurodegenerative disorders
- Impaired bioenergetics and mitochondrial dysfunction
 - Mitochondrial dysfunction in Alzheimer and Parkinson disease
- Fragmentation of neuronal Golgi apparatus
- Disruption of cellular/axonal transport
- Axonal transport in tauopathies
 - Axonal transport in synucleinopathies
 - Axonal transport in other neurodegenerative diseases
- Dysfunction of neurotrophins
- 'Neuroinflammatory' processes
- · The final pathway: multifaceted neuronal death
- Cell death cascades in major NDDs
- Acknowledgements

Abstract

Neurodegenerative diseases are characterized by progressive dysfunction of specific populations of neurons, determining clinical presentation. Neuronal loss is associated with extra and intracellular accumulation of misfolded proteins, the hallmarks of many neurodegenerative *proteinopathies*. Major basic processes include abnormal protein dynamics due to deficiency of the ubiquitin–proteosome–autophagy system, oxidative stress and free radical formation, mitochondrial dysfunction, impaired bioenergetics, dysfunction of neurotrophins, 'neuroinflammatory' processes and (secondary) disruptions of neuronal Golgi apparatus and axonal transport. These interrelated mechanisms lead to programmed cell death is a long run over many years. Neurodegenerative disorders are classified according to known genetic mechanisms or to major components of protein deposits, but recent studies showed both overlap and intraindividual diversities between different phenotypes. Synergistic mechanisms between pathological proteins suggest common pathogenic mechanisms. Animal models and other studies have provided insight into the basic neurodegeneration and cell death programs, offering new ways for future prevention/treatment strategies.

Keywords: neurodegeneration • proteinopathies • pathogenic factors • oxidative stress • bioenergetic deficiency • mitochondrial defects • neuronal dysfunction/death

Introduction

Neurodegenerative disorders (NDD) constitute a set of pathological conditions originating from slow progressive and irreversible dysfunction and loss of neurons and synapses in selected areas of the nervous system which determine clinical presentation and course. The major basic mechanisms leading to neurodegeneration (ND) are considered multifactorial caused by genetic, environmental and endogenous factors related to aging, but their pathogenic role and their basic molecular mechanisms are not fully

*Correspondence to: Kurt A. JELLINGER, Institute of Clinical Neurobiology, Kenyongasse 18, A-1070 Vienna, Austria. understood [1, 2]. NDDs currently are classified according to known genetic mechanisms and/or to the major compounds of their protein deposits (Fig. 1). Based on critical conformational changes of proteins, these disorders are denoted as 'protein misfolding' diseases or proteinopathies [3, 4]. Recent progress with respect to the triggers of most NDDs is illustrated by the number of citations in PubMed (December 4, 2009): ND 37,019; NDDs 168,827; pathogenesis 89,008.

Tel.: +43-1-5266534 Fax: +43-1-5266534 E-mail: kurt.jellinger@univie.ac.at



Fig. 1 Algorithm for classification of neurodegenerative diseases with protein deposits (proteinopathies) (from [1]).

Common pathogenic mechanisms underlying many NDDs include:

- Abnormal protein dynamics with misfolding, defective degradation, proteasomal dysfunction and aggregation; often with actions and mutations of molecular chaperones;
- Oxidative stress (OS) and formation of free radicals/reactive oxygen species (ROS);
- Impaired bioenergetics, mitochondrial dysfunctions and DNA damage;
- 4. Fragmentation of neuronal Golgi apparatus;
- 5. Disruption of cellular/axonal transport (4 and 5 may be regarded as secondary effects);
- 6. Dysfunction of neurotrophins (NTFs) and
- 7. 'Neuroinflammatory'/neuroimmune processes.

These mechanisms are interrelated in complex vicious circles finally leading to cell dysfunction and death, the basic molecular cascades of which under discussion.

Protein aggregation

Abnormal interactions between proteins that result in aberrant intra and extracellular deposition of self-aggregating misfolded

proteins with formation of high-ordered insoluble fibrils are pathological hallmarks of many, albeit diverse, NDDs [see 1, 2, 5, 6]. In general, the identity of the underlying protein determines which neurons are affected and, hence, the clinical manifestation of each disease [7, 8]. However, the same neuronal populations can be affected by different disorders. The same neurodegenerative process and the same mutation in the genes encoding protein constituents may be associated with a variety of clinico-pathological phenotypes, whereas similar or identical phenotypes may be related to different genetic defects, resulting from complex gene-gene and gene-environmental interplay [9]. Abnormal protein-protein interactions and/or the lesions that result from them trigger vicious circles leading to dysfunction and death of neuronal and glial cells with ensuing disintegration of neuronal networks [10, 11]. Abnormal interaction between normal, highly soluble brain proteins alters their conformation, and misfolding gradually converts them into insoluble polymers with the aggregates adopting either highly ordered (cross-pleated B-sheet structures) or disordered (amorphous) forms [12]. Since deposits of natively unfolded proteins aggregated into defined fibrillar structures often display the properties of amyloid (i.e. ~10-nm-wide fibrils with crossed β -pleated sheet structures), these disorders are grouped together as brain amyloidoses [2, 13].



'Toxic oligomer' hypothesis

A causative link between the formation of protein aggregates and ND is suggested to be a result of the toxic action of substances produced during early phases, *i.e.* soluble oligomers and protofibrillar derivatives of misfolded proteins [14–16]. However, the precise biochemical mechanisms are poorly understood.

The 'toxic oligomer' hypothesis is supported by the finding that a single-domain antibody can recognize a common conformational epitope that is displayed by several disease-associated proteins, including β -amyloid (A β), α -synuclein (α Syn), τ protein, prions and polyglutamine (polyQ) peptides [17-20] (Fig. 2). Soluble AB oligomers are increased in Alzheimer's disease (AD) in both brain tissue and plasma [21], and their levels correlate with cognitive dysfunction [22]. The analysis of the assembly pathway of AB in vitro and biochemical characterization of AB deposits isolated from AD brains indicate that AB oligomerization occurs via distinct intermediates. Recent studies suggest the existence of different types of AB42 strains, some causing toxic effects and others aggregation into oligomers without adapting toxic conformations [23]. AB40 inhibits amyloid deposition in vivo [24], but 3D reconstruction of AB1-40 and AB1-42 amyloid fibrils revealed similar protofibril structures [25]. The cellular prion protein (PrPC) and amyloid precursor protein (APP)-like protein 1 (APLP1) were suggested as possible mediators of A_B-induced synaptic dysfunction [26]. AB oligomers isolated from the brains of AD patients and injected into rodents are capable to disrupt N-methyl D-aspertate acid receptors and impairing memory [27]. AB peptides have a crucial role in regulation of synapse vesicle release and might point to the primary pathological events that lead to compensatory synaptic loss in AD [28]. APLP1 forming heterodimers with APP

and being mainly present on the cell surface, is a strong candidate receptor for (toxic) AB oligomers [29]. They affect the presynaptic and terminal post-synaptic region [30, 31, 424-426], synaptic plasticity and transmission [32] correlating with synaptic loss [33]. and perturbated APP synaptic adhesion activity may contribute to synaptic dysfunction and AD pathogenesis [34, 35]. Native amylospheroids isolated from AD brains with a distinct toxic surface induce neuronal loss through a mechanism different from other AB assemblies, which have been reported to perturbate glutamatergic synapse transmission [36]. The amount of AB is determined by the balance between its production by proteolytic processing of APP via β - and γ -secretases and its clearance. Apolipoprotein E ε 4 has been identified as an AD risk factor, partly explained by its reduced ability to crosslink AB [37]. Increased AB production and/or aggregation likely contribute to progression of AD [38], while suppression of AB deposition leads to long-term reduction of AD pathology [39, 40]. Recent results provide a unifying mechanism in which AB oligomers induce τ hyperphosphorylation in AD, while an antibody that targets soluble AB oligomers blocks their attachment to synaptic binding sites and prevents τ hyperphosphorylation [41]. Autoantibodies against AB are common in AD brain and help control the plaque burden [42]. The fraction of the soluble, mainly toxic AB pool is intracellular and is recognized and targeted by conformation-sensitive antibodies [43]. Single-domain antibodies recognize selectively small oligomeric forms of AB, prevent AB-induced neurotoxicity, and inhibit fibril formation [18]. Lipid surfaces promote aggregation of AB proteins and subsequent permeability changes on lipid membranes [44], but there is a dynamic relationship between intra and extracellular pools of AB [45], intraneuronal AB accumulation particularly contributing to AD progression [46], to which changes at neuronal networks may strongly contribute [47]. A prion-like spread has been shown for A β aggregates in a tg mouse model [427], and extrusion of A β aggregates may seed extracellular amyloid plaque formation during AD pathogenesis [428].

Oligomeric and pre-aggregated forms of τ protein have been shown to be toxic *in vitro*, but the mechanisms underlying its structural changes leading to human tauopathies still remain elusive [48, 49]. The natively unfolded character of τ and its aggregation to Alzheimer-like paired helical filaments and the link between A β and τ in this relationship in AD and other NDDs have been discussed conversely (coexistence of both, axonal connection; 'dual pathway' model – [40, 50]. Granular τ oligomers slowly evolve to larger structures and eventually to filaments having a size smaller than those for PHFs purified from AD [51]. Recent studies with τ that has been believed to reside within the cytoplasm, suggested τ aggregates spreading through cell-to-cell transmission of misfolded protein [52–54], and aggregation intermediates are linked to ND [48, 55].

 α Syn assembling into highly soluble special oligomers may proceed to insoluble aggregates and plays a central role in ND in Parkinson disease (PD) [56]; it is potentially prone to misfold [57] and may lead to neuronal death, but other modes of toxicity have been proposed [58], suggesting interaction of α Svn and AB resulting in ND [59]. The lysosomal protease cathepsin D influences α Syn processing, aggregation and toxicity *in vivo* [60], and α Syn contributes to glycogen synthase kinase 3 β catalysed τ phosphorylation in PD models [61]. Interaction with synphilin-1 promotes inclusion formation of α Syn providing new insights in to the inclusion body formation in Lewy body disorders [429]. Elevated levels of soluble α Syn oligomers have been detected in post-mortem brains of DLB patients [62]. Seeding induced by α Svn oligomers may lead to spreading of α Svn pathology [63] and to the formation of Lewy body (LB)-like intracellular inclusions [64], with cell-to-cell transmission of α Syn aggregates via endocytosis, leading to nuclear fragmentation and caspase-3 activation [65]. NMR spectroscopy provided information about structure and aggregation of α Syn [66]. Methods to detect morphologically distinct oligometric forms of α Syn have been described [67].

In polyQ diseases, the protein context of expanded polyQ and its soluble mutant conformers are critical for the disease specificity [20]. Structural and dynamic aspects related to oligomerization of superoxide dismutase 1 (SOD1) and its mutants have provided better insight into SOD1 pathway in relation to familial amyotrophic lateral sclerosis (fALS) [68], which is linked to the two genes TARDBP (TDP-43, which encodes TAR-DNA binding protein) and FUS (RNA binding protein fusion) [69, 70], whereas in sporadic amyotrophic lateral sclerosis (sALS) there is lack of evidence of monomer/misfolded SOD1 [71]. As most NDDs, ALS features deposits of SOD1, TARDBP and FUS aggregates.

The discovery of new prions expands the spectrum of their biological functioning [72]. It further promotes the assembly of these pathological conformers into filaments that progressively accumulate in a disease- and protein-specific manner in the cytosol or nuclei of affected brain cells (neurons and/or glia) or in the extracellular space [16, 73]. The relationship between A β and PrPC and the role of PrPC in the pathogenesis of AD are under discussion [74]. In spinocerebellar ataxia (SCA), protein aggregation, alteration of the CO_2 homeostasis and others are important pathogenic factors [75].

Protein (mis)folding

Proteins are a heterogeneous population of different conformers which, due to their flexibility, are in a dynamic state between various conformational substrates maintained by synthesis and degradation [76]. Folding, part of the normal process that converts newly synthesized proteins to physiologically functional molecules, is controlled by molecular chaperones that prevent inappropriate interaction between non-native polypeptides, and promote the refolding of proteins that have become misfolded as a result of cellular stress [77]. Protein aggregation occurs in vivo as a result of improper folding or misfolding leading to a structural change of a normal, functional protein, inducing the formation of protein aggregates with various supramolecular organizations (Fig. 3). Aberrant proteins, the result of production errors, inherited or acquired amino acid substitutions or damage, often cannot fold correctly and will be trapped in misfolded conformations. The pathogenic pathways involve membrane permeabilization through a channel mechanism or hydrophobic interaction of prefibrillary oligomers with various cellular targets [78]. To get rid of misfolded proteins, the living cell contains a large number of intracellular proteases, which, together with the chaperones, comprise the cellular protein quality-control systems in the endoplasmic reticulum (ER). Quality control against misfolded proteins is the cytosole in a network for cell survival [79], and ER protein quality control in NDD is disordered [80].

Many proteins associated with NDDs are intrinsically disordered under physiological conditions [4, 81]. Progressive intracellular protein accumulation can result from various pathological processes: (i) abnormal synthesis and folding, (ii) abnormal interaction with other proteins, (iii) overproduction of protein constituents, (iv) impaired degradation and turnover, (v) altered post-translational modifications of newly synthesized proteins, (vi) protein oxidation, (vii) nucleic acid-induced structural conversions. (viii) abnormal proteolytic cleavage. (ix) improper expression or altered gene splicing, (x) insufficient molecular chaperone activity and (xi) impaired intracellular transport of proteins. Recent studies with mutant huntingtin (mhtt), τ protein and a Syn demonstrated propagation of misfolded protein aggregates through cell-to-cell transmission in cultured cells and tg mouse models [430-432]. Due to the central role of these phenomena in cell biology, protein misfolding and aberrant disorder-to-order conformational transitions in protein structure are associated with a large number of NDDs [82]. Causes are genetic deficits producing a single amino acid substantiation or expansion of a repeating amino acid tract, as occurs in familial diseases. However, for most NDDs that occur sporadically or in non-Mendelian familial fashion, other factors may induce the pathogenic cascade.



Fig. 3 Model of protein misfolding and fibrillation leading to deposition of aggregated proteins in cells and extracellular spaces via actions of the UPS, phagosomes and aggresomes, either causing cell death or cytoprotection.

Proteostasis and molecular chaperones

Protein formation is regulated by the proteostasis network, an integrated biological system that generates and protects the protein fold [83]. Many inherited disorders due to amino acid substitutions exhibit loss-of-function (LOF) pathogenesis because the aberrant protein is eliminated by one of the control systems. However, not all aberrant proteins can be eliminated and the misfolded protein may accumulate and form toxic oligomeric and/or aggregates. In this case, the loss of function may be accompanied by a gain-of-function (GOF) pathogenesis, which often determines the pathological and clinical features [84]. Many diseases of protein homeostasis, or 'proteostasis', due to misregulation of protein maintenance, include LOF and gain-of-toxic-function diseases (AD, PD, Huntington disease/HD). The conformational change may promote diseases either by gain of toxic activity or by the lack of biological function of the natively folded protein under adverse conditions, including OS, ER stress and aging by accumulation of damaged proteins in the cell [85]. In AD, there is no consensus as yet whether the disease acts through a LOF or GOF mechanism. On the other hand, larger protein aggregates may represent an inherent protective or defensive mechanism by sequestering or inactivating toxic oligomers and protofibrils [see 1, 6] and AB monomers have been shown to be neuroprotective. This 'abnormal protein-protein interaction' or 'fatal attractions' hypothesis describes plausible unifying mechanisms to account for the onset and progression of neurodegenerative proteinopathies.

Proteostasis is maintained by a network, which comprises pathways that control protein synthesis, folding, trafficking,

aggregation, disaggregation and degradation, including the unfolded protein response (UPR), the heat-shock response, the ubiquitin proteasome system (UPS) and epigenetic programs [86]. Decreased ability of this network to cope with inherited misfolding-prone proteins, aging, and/or metabolic/environmental stress appears to trigger or exacerbate proteostasis diseases [86]. This is influenced by genes that control aging, thus linking stress and protein homeostasis with the health and life span of the organism. Small molecules can enhance proteostasis by binding to and stabilizing specific proteins (chaperones) or by increasing the proteostasis network capacity (proteostasis regulators) [87]. Adaptation and survival requires the ability to coordinate the activities of protective stress response pathways and chaperone networks.

Molecular chaperones have essential roles in many cellular processes at the folding/degradation interface in mammalian cells [88]. Cellular molecular chaperones, which are ubiquitous, stress-induced proteins, have been found to be effective in preventing misfolding of different disease-causing proteins, essentially reducing the severity of several NDDs and other proteinopathies. Chaperones are engaged in suppressing the effect of protein misfolding-induced consequences [89, 90] inhibiting amyloid formation in the extracellular space [433] and to protect against OS [91]. They play an important role in the deterrence of protein damage during aging and in ND [92]. Conditions of stress are characterized by a robust increase in the synthesis of heat shock proteins (HSPs) that are crucial for recovery from stress-induced protein damage [93]. Almost all HSPs function as molecular chaperones, and the number of diseases that are known to be caused

by their mutations is increasing. Accumulation of aggregationprone proteins activates signal transduction pathways that control cell viability in various models of PD, HD and ALS [57, 77, 94] and there is an emerging role of chaperones in the pathogenesis of PD [434]. Several chaperone actions might be required to impede tissue pathogenesis *in vivo*, and molecular chaperones may increasingly become new targets for the therapy of NDDs [77].

Protein misfolding and endoplasmic reticulum stress

In neurons, the ER is important for the synthesis, folding and post-translational modification of transmembrane and secreted proteins. The ER response is characterized by changes in specific proteins, causing translational attenuation, induction of ER chaperones and degradation of misfolded proteins, while in prolonged or aggravated ER stress, cellular signals leading to cell death are activated. ER stress may be involved in some human neuronal diseases, such as AD, PD, prion diseases, etc. [95]. The exact mechanisms and causal effects of ER stress and the proteins involved, however, are poorly understood [96].

The mechanism by which α Syn, τ and A β protein make fibrils is an example of conformational plasticity, because these polypeptides can visit a coil or helical structure, but otherwise they convert into a pathogenic B-sheet structure highly suitable for polymerization and fibre formation [57]. Misfolding of a Syn, which is involved in neuronal functions, and can act as molecular chaperone, is a critical factor of synucleinopathies [97]. It can affect the function of several key PD-linked genes such as DJ-1, PINK-1, and perhaps also LRRK2, and parkin, which like α Svn, is also prone to misfolding, especially in the presence of age-related stress [98]. Both conformational folding of previously unfolded τ protein and cleavage causing aggregation lead to neuronal dysfunction by interference with axonal transport, neurotoxicity of early aggregated state, contributing to disruption of neuronal circuits due to synapse loss and cell death [99–101]. Extracellular τ , α Syn and prion aggregates can transmit a misfolded state from the outside to the inside of a cell, indicating propagation of protein misfolding [52, 64, 65].

The role that polyQ-induced aggregates, *e.g.* in HD, plays in disease is not yet fully determined. N-terminal mutant huntingtin (mhtt) proteins activate cellular pathways linked to ER stress and cause protein misfolding inside the cells and form toxic amyloid-like aggregates, preceded by a series of protein misfolding steps termed poly-Q-fibrillation as a prerequisite for the underlying mechanisms of ND [102]. They could be part of a compensatory detoxification process by the cell in which pathogenic oligomers are sequestered [103], suggesting that compounds targeting ER stress may be considered in designing novel approaches for treatment of HD and possibly other polyQ diseases [104].

In vivo, these changes develop insidiously over the lifetime of an individual, even though they usually do not manifest clinically until middle or late life. The causes of this prolonged process due to progressive damage of specific vulnerable brain regions or neuronal networks before clinical manifestation are poorly understood. A crucial unanswered question is whether these aggregates contribute to the onset and progression of ND are mere bystanders resulting from an alternative pathway, or even may be a neuroprotective response [1, 105]. Although mutations in the genes encoding the fibrillizing proteins segregate familial forms of the corresponding diseases, the same brain lesions also can be formed by the corresponding wild-type protein in a sporadic form of the disease. Since protein misfolding may also result in altering their normal function, e.g. physiological proteins, fibrils may be inactive endproducts of a common pathogenic cascade of an otherwise deleterious process, but, in general, detection of fibrillar assemblies indicates that neurons have been exposed to a wide array of noxious agents. It becomes apparent that the pathology of AD and other proteinopathies represents effect rather than cause, or a host response to injury equaling neuroprotection [106].

Unfolded protein response

Proteins are synthesized and folded in the mitochiondria as well as in cytosol and in the ER. Alterations in cellular homeostasis that affect protein folding in the ER trigger a signalling pathway known as the UPR [107]. There are three pathways to relieve ER stress and to thus promote survival of the neuron: (i) activation of PRKlike endoplasmic reticulum kinase (PERK) inhibitory cellular protein translation, preventing further protein synthesis; (ii) increased capacity of the ER to handle unfolded proteins, mediated by the activating transcription factor 6, and inoritol-requiring kinase-1 and (iii) removal of misfolded proteins from ER by enhancing proteolytic degradation [108]. The molecular mechanisms underlying these and related processes are unclear, but recent studies suggested that the pathway starts with stress detection in the mitochondrial matrix followed by activation of UPR mitochondrial targets in the nucleus. ER stress is caused by disturbances in its structure and function with accumulation of misfolded proteins and alterations in calcium homeostasis [109]. During aging and in NDDs, cellular Ca²⁺-regulating systems are compromised resulting in synaptic dysfunction, impaired plasticity and ND. OS, perturbated energy metabolism and aggregation of disease-related proteins adversely affect Ca²⁺ homeostasis by mechanisms that have been elucidated [109, 110]. The UPR is activated in pretangle neurons in AD hippocampus [111] and increased levels of molecular chaperones in cytologically normal appearing neurons suggest its early role. Although its initial participation in AD pathogenesis could be neuroprotective, its sustained activation might initiate or mediate ND [112].

Interaction of proteins

Despite differences in the molecular composition of these filamentous lesions, the brain regions and cell types they affect, growing evidence supports the notion that they engage common downstream pathogenic targets. Although the different localizations of hallmark protein aggregations (extracellular AB deposits in AD, cytoplasmic LBs in PD, TDP-43 intracytoplasmic and intranuclear inclusions in AD, frontotemporal lobar degeneration, ALS, intranuclear inclusions in HD) may suggest that each protein strikes a single cellular domain, recent studies indicate the possibility of common pathogenesis. In addition to showing common morphological and chemical features, the inclusion bodies co-localize with several other proteins that are not structural elements of abnormal fibres, but are binding to them, including various chaperones, proteins implicated in cellular responses, signal transduction. cytoskeletal, cell cycle and others [105, 113]. Proteomic analysis of cortical LBs revealed 296 proteins [114], while in brainstem LBs 90 proteins were identified, differing from Pick bodies and suggesting a complex formation process [115]. In vivo protein aggregation exhibits a remarkable specificity, suggesting that conserved mechanisms underlie it depending on selective interactions and result in the formation of fibrillary structures. This might reflect an irreversible sequestration and subsequent loss of functions and/or a failed attempt to refold and degrade aggregated proteins.

The cellular response to these aggregates includes (1) the recruitment of chaperones or proteins involved in the folding of nascent translational products and in the resolubilization of aggregated polypeptides, and (2) the ubiquitination of aggregates, suggesting cellular attempts to degrade deposits of these mutant proteins *via* the UPS [116].

The frequent co-occurrence of AD and PD and of inclusions characteristic of both disorders in several other NDDs suggests the involvement of a common underlying process [117-119], which may be caused by hybrid oligomer interactions [120]. A collision of two or more processes may occur in the same brain region or even within single cells, e.g. in rare familial forms of DLB [121]. A direct link between τ and synaptic pathology is supported by accumulation of abnormally phosphorylated τ and α Syn within synaptic terminals in AD brains and APP Swedish mutant to mice [122, 123]. Induction of hyperphosphorylation of τ by α Syn in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) model of parkinsonism [124], co-location of phospho- τ and α Syn in both neurofibrillary tangles (NFTs) and LBs [125], and co-occurrence of abnormal deposition of τ , α Syn and TDP-43 in AD, DLB and other NDDs, probably related to genetic factors [118, 126], highlight the interface between these two and other misfolded proteins. Oxidatively modified α Syn degraded by the proteasome promotes the recruitment of τ to protein inclusions in oligodendroglia cells in synucleinopathies [127]. Accumulation of AB and phospho- τ are thought to be coexistent, AB initiating hyperphosphorylation of τ [40]. Both can be linked by separate mechanisms driven by a common upstream driver [50]. Interactions between AB, aSyn and τ may be a molecular mechanism in the overlapping pathology of AD and PD/DLB [128, 129], probably generated by the same stimulus with the outcome possibly having an inverse relationship depending on genetic background or environmental factors. Although recent data documented co-localization of AB and phospho- τ in synaptosomes [130], and A β and τ protein show synergistic, age-associated effects in triple tg AD mice [131]. Why τ and A β pathologies are so intimately associated, remains one of the major unresolved questions of AD pathophysiology. TDP-43 proteinopathies are distinct from most other NDDs because they are due to protein misfolding without amyloidosis, while TDP-43⁺ inclusions show abnormal hyperphosphorylation, ubiquitination and C-terminal fragments [132]. It is unclear, whether there is a common underlying pathogenic mechanism inducing both ND and fibrillary protein aggregates that are typical for different disease processes (double or triple amyloidosis) or if they represent a common final pathology leading to ND.

The ubiquitin proteasome system

All cellular components are subjected to continuous surveillance by intracellular control systems. The UPS, the major non-lysosomal system for regulating protein turnover, and the autophagylysosome pathway (ALP) are the two most important mechanisms for regulating protein handling [133]. These cellular protein quality control systems in the ER are important in the pathogenesis of NDDs, but also in neuroprotection [80, 134]. The presence of ubiquitinated proteins within neuronal inclusions is one of the hallmarks of ND. The inclusions contain various components of the UPS which operates as an intracellular protein-clearing system.

The UPS is a substrate-specific non-lysosomal, ATP-dependent proteolytic system that plays a major role controlling the initial steps of gene expression, DNA repair, nuclear quality-control, cell cycle and signalling control [133, 135]. It appears to be at the intersection of whether a toxic protein is degraded or whether it is packed into an inclusion which may be one of the strategies of the cell to process damaged and/or mutated potentially toxic proteins and that, given a chance, the cell will recover from such stress. Proteasome regulators (activators and inhibitors) have been reviewed recently [136]. Proteasomal dysfunction may be caused by misexpression of one or more of its subunits. The UPS becomes activated during OS and works to process misfolded protein-mediating reactions that link abnormal proteins with multiple ubiquitine (Ub) molecules as a signal for degradation domains and may promote their degradation by Ub ligases [137. 138]. Aggregated proteins are transported to perinuclear microtubule-organizing centres (centrosomes), where they are encapsulated by intermediary filaments to form aggressomes, subsequently subjected to proteolysis and removed by autophagy [139]. Ubiguitylation and endocytosis of plasma membrane proteins are regulated by Ub ligase adaptors [140] mediated by Ub ligase Ubr1 [141]. The proteasome also contains deubiquitinating enzymes that can remove Ub before substrate degradation initiates, thus allowing some substrates to escape degradation [142]. The UPS becomes abnormally activated also during abnormal protein cleavage, and altered or inappropriate gene splicing. In these conditions, it may not be able to degrade damaged proteins producing their aggregation in the cell and ultimately causing neuronal dysfunction (Fig. 3). A reduction in its efficacy increases the storage of aggregation-prone proteins and could explain the accumulation of Ub and substrates in diseased neurons, which disrupt normal cell activity [116]. However, ubiquitination of protein substrates without proteolysis and proteasome-independent function of Ub in endocytosis and signalling, and Ub-independent proteolytic degradation of proteins have been described [133]. Thus, derangements in the Ub could potentially lead to alterations in processes that are unrelated to protein degradation.

Ubiguitination, the hallmark of protein degradation by the 26S proteasome, is a multistep process, in which Ub-activating enzymes (E1s, E2s and E3s) are key mediators and are able to regulate the chain formation as downstream signalling pathways [137, 143]. Mono-Ub and Ub dimers may regulate the enzymatic functions of UB carboxy-terminal hydrolase L1 (UCH-L1) and L3, two of the deubiquitinating enzymes expressed in the brain, in vivo [144]. The family of proteins containing Ub-like and Ub-associated domains has been implicated in proteasomal degradation, thus regulating the proper turnover of proteins [133] [145]. Control of iron homeostasis is performed by Fe-regulated ubiquitin ligase [146]. Through a process termed sumoylation, SUMO (small Ub-like modifier) monomers are conjugated to target proteins. In some cases they may act to protect proteins from Ub-dependent degradation or appear to trigger polyubiquitination, and there are relationships between Ub, SUMO and DNA repair pathways. Monoubiguitination of the p53 tumour suppressor protein causes it to be transported from the nucleus to the cytoplasm by multifaceted effects of SUMO [133, 147, 148]. p53 protein requires multiple layers of regulatory control to ensure correct functions [435], and p53 proteasomal degradation may be Ubindependent [436]. The complexity of the Ub system in protein regulation and its role in the regulation of cell death are still not fully elucidated [149, 437], but quantitative proteomics gave insight into the function of unconventional Ub chains in proteasomal degradation [150].

Inhibition of lysosomal functions by higher levels of chaperones reduces proteasomal activity [151]. Under conditions of proteolytic stress, the cell may switch to a non-proteolytic form of ubiquitination to help divert misfolded proteins away from an overloaded proteasome, and thus can preserve its proteasomal functions over prolonged periods of stress and recover thereafter [152].

Many expanded proteins are resistant to proteolysis by the UPS, and the lack of its efficacy may be responsible for accumulating expanded proteins within affected cells, leading to *neuronal inclusions*. Ubiquitinated protein aggregates provide a nuclear centre for the formation of inclusion bodies, but there are several differences that distinguish these inclusions, some being aggregated in the cytoplasm, others are prevalent in the nucleus [105]. Differential activities of the UPS in neurons *versus* glia may account for the preferential accumulation of misfolded proteins [153].

Misregulation of degradation of misfolded proteins leads to their accumulation with inhibition of axonal transport, thus facilitating the accumulation of ubiquitinated proteins in the cell body and cell dysfunction. This interaction may involve (1) a loss of function, (2) a gain of function or (3) an inflammatory stimulus.

Autophagy and neurodegeneration

Another pathway for cytosolic protein clearance and organelle degradation through lysosomes is autophagy [154-156], a cytosolic, non-specific bulk degradation system, important as a clearance route for many cytosolic, aggregate-prone proteins. Model systems in increasing numbers are available to study the role of autophagy in the central nervous system (CNS) [154, 157]. Proteins are transferred to the lysosomal membrane and, through binding to a receptor lysosome-associated membrane protein (autophagosome), they are translocated into the lysosomal lumen and degraded [158]. Ub has a role in selective autophagy [139]. The molecular mechanism involved in the three autophagic pathways: chaperone-mediated autophagy (CMA), microautophagy (without vesicles) and macroautophagy, the most common type associated with mitochondria and cell death [159, 160]. Macroautophagy is responsible for clearing of aggregated or aggregate-prone proteins, microautophagy for degradation of organelles, and CMA uses the lysosomal protein LAMP2a as a receptor that recognizes proteins with a specific pentapeptide for lysosomal degradation. Their relevance in ND has been reviewed recently [159, 161]. The autophagy machinery is regulated by different types of stress at various levels, from transcriptional activation to post-translation of protein modification [162]. Dysregulation of macroautophagy has been implicated in synaptic dysfunction, cellular stress and neuronal cell death [163] with superoxide being the major ROS regulator [164], and contributes to the pathogenesis of several human diseases, including NDDs [158], while its activation seems to be protective in certain NDDs [165]. The process, known as microautophagy, is poorly understood [166, 167]. Degradation of mutated α Syn by the CMA is impaired, which could explain the selective degeneration of PD dopaminergic neurons [168, 169]. The autophagy-related protein beclin-1 does not only play an important role in the intracellular degradation of α Syn either directly or indirectly through the autophagy pathway [170], thus ameliorating the ND in α Syn models of PD, but also shows reduced expression in early AD [171], while x-box-binding protein, a key UPR transcription factor that regulates genes involved in protein folding and quality control, protects against ALS by increasing autophagy [172]. Tissue-specific impairment of autophagy in the CNS tissue causes massive loss of neurons resulting in ND [173]. Emerging evidence supports the view that induction of autophagy is a neuroprotective process and that inadequate or defective autophagy, rather than excessive autophagy, compromises degradation of UPS substrates and, thus, promotes cell death [174, 175]. The UPS and autophagy are two signalling pathways that can provide protection against ND [138]. However, how autophagy protects cells from damage leading to ND is not yet clear [176, 177].

UPS in neurodegenerative disorders

Dysregulation in the UPS appears to be both a cause and a result of ND processes. Its dysfunction has already been implicated in the pathogenesis of PD [178, 179], and the demonstration that α Syn is degraded by both proteasome and autophagy indicates a possible linkage between the UPS and ALP [168]. Mutated aSyn inhibits ALP by binding to the receptor on the lysosomal membrane for autophagy (Greek: self-digestion) pathway supporting the assumption that the ALP may be related to the development of PD [180]. LBs, the morphological hallmark of PD and DLB, have a distinct central parkin and Ub^+ domain, with α Syn in the periphery, but it is incorporated into LBs and dystrophic neurites before ubiquitination [181]. Co-localization of α Syn and parkin within LBs suggests that parkin plays a role in ubiquitination and posttranslational modification of α Syn [182]. The Parkinson-associated protein PINK1 interacts with Beclin1 and promotes autophagy [438]; on the other hand, PARKIN is cytoprotective, partially by increasing the removal of cellular AB through a proteasomedependent pathway [439]. There is conflicting evidence concerning the pathogenesis of α Svn aggregation [66]. A functional Ub-3-ligase complex consisting of early-onset familial PD associated Parkin, PINK-1 (PARK-6) and DJ-1 (PARK-7) mutations promotes degradation of un- or misfolded proteins and may constitute a pathogenic mechanism for recessive forms of inherited PD [81, 1831. LRRK2 mutations (PARK8), a major cause of late inset parkinsonism, have clinical features comparable to sporadic PD [184, 185], display variable neuropathology, including α Syn and τ inclusions [186, 187], thus suggesting an upstream role of LRRK2 in protein aggregation [188].

The role of UPS pathology in AD has been reviewed [133, 189]. AB inhibits the proteasome and enhances amyloid and τ accumulation [190], but proteasome inhibition increases accumulation and insolubility of τ protein independent of τ phosphorylation, and JNK inhibition may be partially responsible for the relatively decreased phosphorylation of τ in tg models [191]. ATPase subunit 6b (S6b) transcript up-regulation may be related to tauopathies, while in synucleinopathies it appears not to be involved [192]. The importance of decreased UPS function has been observed also in other NDDs, e.g. in HD, where protein guality control regulates polyQ-induced toxicity [20], but its global status in HD brain is not yet fully elucidated [193]. Neuronal polyQ protein inclusions are present predominantly in the nucleus, which is not accessible to autophagy. Intracellular degradation of polyQ aggregates by the UPS is triggered by some nuclear Ub ligases [194], and inclusion bodies are suggested to have a protective cellular response to mhtt mediated by improving intracellular protein degradation [195], while UPS activity in synapses of HD is impaired [196].

Dysfunction of the UPS is also important in ALS showing skein-like inclusions in motor neurons rich in Ub proteasome and some chaperone proteins, in SCAs, with neuronal intranuclear inclusions containing ataxin-1 and several chaperones [197], in ALS-parkinsonism-dementia complex of Guam (ALS/PDS), neuronal intranuclear inclusion disease, and other NDDs (for review see [133]. However, despite the notion that inclusions might be protective, involved neurons ultimately fail to compensate for the abnormal and/or toxic protein accumulation and, finally, die. While inhibition of the UPS might be expected to worsen most NDDs, its augmentation possesses unique challenges, such as delivery of its components to the nervous system or identification of drugs that enhance the degradation of toxic proteins without compromising normal UPS function [116].

Aggresomes

When the capacity of the proteasome system to degrade misfolded proteins is overwhelmed, aggregation occurs and proteins are moved to a Ub-rich structure termed the 'aggresome' [198]. It forms part of the cellular response to aggregated proteins and appears as inclusions in a number of proteinopathies. Aggresomes have been reported for SOD, parkin, α Syn [181] and prion proteins, the formation of which has been associated with activation of caspases and apoptosis. Ubiguitination and sequestration of protein (e.g. aSyn, synphilin-1) in aggresomes and cytoplasmic inclusions may represent a mechanism of cell protection. On the other hand, α Svn aggregation is associated with decline in proteasome and lysosome, which may be involved in the pathogenesis of cell degeneration in PD [199]. However, it is not clear whether aggresome formation is causative or protective [200], although data suggested that they serve a cytoprotective function. facilitating the degradation of toxic proteins.

Oxidative injury

OS occurs when the production of free radicals or their products are in excess of antioxidant defence mechanisms. It can damage biological molecules and initiate a cascade of events, including dysfunction of mitochondrial respiration, excitotoxicity, and a fatal rise in cytosolic calcium leading to cellular dysfunction together with nitric oxide and reactive nitrogen species. Thus, it is a major factor of the cytopathology of many diseases including NDDs and their models [see 201-204]. The generation of ROS during earlystage protein aggregation is a common fundamental molecular mechanism underlying their pathogenesis. A major source of ROS in neurons may result from escaping electrons from the respiratory chain reacting with oxygen. Other contributors are metal-ironassociated Fenton reactions, lipid peroxidation and nitric oxide induced protein nitrosylation [203, 205]. Generation of excessive nitric oxide and ROS can mediate protein misfolding in the absence of genetic predisposition [206]. Among the various free radicals generated in the living organism, hydroxyl radical and peroxynitrite are the most potent and can damage cells via nonselective oxidation of proteins, lipids, fatty acids and nucleic acid [204]. They are formed via the Haber-Weiss and Fenton reaction between H_2O_2 and reduced transition metals. Hydrogen peroxide is subsequently converted to hydroxyl radicals by the addition of Fe(II) by Fenton's reaction, one of the fundamental mechanisms underlying neurodegenerative processes. The accumulation and precipitation of proteins may be aggravated by OS, and may, in turn, cause more oxidative damage by interfering with the function of the proteasome, that increases levels of OS not only to proteins but also to other biomolecules [203].

Proteins are initial targets of ROS, and protein radicals generated by ROS can oxidize reduced glutathione, which triggers dysregulation of calcium homeostasis, reactive astrogliosis and other changes observed in ND [207, 208]. Superoxide (0^{2^-}) is suggested to be the major reactive OS regulating autophagy [164]. Cells which fail to compensate for oxidative imbalance (stress) enter apoptosis with rapid cell death, while those with compensatory response to ROS (antioxidant enzymes, low molecular weight reductants, etc.) may show long-term survival. There is no clear and defined mechanism showing how OS plays a key role in the regulation of cell survival and death in NDDs, but sublethal RNA oxidation may be a pathogenic mechanism for NDDs [209].

Increased levels of oxidative damage to DNA, lipids and proteins have been detected in post-mortem tissues from patients with PD, AD, ALS, progressive supranuclear pals (PSP) and aging, some of which are environment-induced [210]. Oxidative damage to nuclear and mitochondrial DNA (mtDNA) occurs in the earliest detectable phase of AD, PD and HD, but also in normal aging [202, 204, 211–213]. Although the precise sources of increased oxidative damage are not fully clear, increased localization of redoxactive transition metals in brain regions most affected by ND is consistent with their contribution to OS. Free radical oxygen chemistry plays an important pathogenic role in all these conditions, though it is as yet undetermined what types of oxidative damage occur early in the pathogenic cascade and which ones are secondary manifestation of dying neurons [214, 215]. HSPs have been shown to protect against OS [91].

Metals and oxidative stress

Alterations in metal homeostasis inducing increased production of free radicals, suggests a direct cause-effect relationship between disruption in metal homeostasis and the increased oxidative damage. In addition to the generation of O^{2-} and H_2O_2 , the availability of redox-active Fe is a major determinant of ROS-mediated cellular damage. Elevated levels of redox-active Fe, derived from degenerating mitochondria, accumulate in in the normal aging brain and in several NDDs [216].

Iron is a powerful promoter of free radical damage, able to catalyse generation of highly reactive radicals from H_2O_2 and lipid peroxides, generating OS. Although most iron in the brain is stored in ferritin, 'catalytic' iron is readily mobilized from injured brain tissue. In AD, aberrant metal homeostasis may contribute to the formation of ROS and toxic A β oligomers, and increased lipid peroxidation precedes A β plaque formation in animal models of AD. Imbalance in iron homeostasis is a precursor of the ND

process leading to AD [440]. Conditions such as neuroferritinopathy [217] and Friedreich ataxia are associated with mutations in genes that encode proteins involved in iron metabolism [218], inducing OS and cell death [441] the brain ages, iron accumulates in regions that are affected by AD and PD [219], and dopamine (DA) complexes of iron are important in the pathogenesis of PD [220].

Oxidative stress in Alzheimer disease

The role of OS in various NDDs, including AD and PD, has recently been reviewed [see 1, 221]. Here, only some essentials will be summarized.

In both human AD and transgenic (tg) mouse models of AD, oxidative damage occurs preceding AB deposition that further contributes to OS and ND [222]. In the initial stages of AD, AB deposition and phospho- τ may represent compensatory responses to ensure that neurons may not succumb to oxidative damage. OS induces macroautophagy of AB protein and ensuing apoptosis [223]. Mutant APP and its derivates are involved in the generation of free radicals in mitochondria and cause mitochondrial oxidative damage, linking AB, generation of free radicals and oxidative damage in the pathogenesis of AD [224]. OS or neurotoxic by-products of lipid peroxidation may damage DNA and lead to programmed cell death (PCD) in AD. In AD and other NDDs, the production of advanced glycation end products (AGEs) and their receptors has been observed [225-227]. Being both markers of transitional metal-induced OS and inducers of protein crosslinking and free radical formation, they may reflect early disease-specific changes rather than late epiphenomena. AGEs co-localize with inducible nitric oxide synthase in AD, supporting an AGE-induced OS [227]; and contribution of AGE receptors (RAGE) to AB cytotoxicity [228], while infusion of soluble RAGE in tg animals decreases AB content and amyloid load [225]. Oxidative imbalance is met by a series of complex reactions (mitochondrial dysfunction, impaired synaptic transmission, disruption of membrane integrity, etc.) to establish a disease-related homeostasis balance [204]. Excess AB may induce OS and/or disturb cellular calcium homeostasis through disproportionate PrPC-receptor crosslinking, but the precise mechanism between mitochondrial oxidative damage and the role of oligomeric AB has not yet been explicated [229]. Oxidative damage to DNA and RNA repair is particularly severe in the hippocampus, the earliest and most severely involved brain area, and nitrated brain proteins are seen in both early and late stages of AD [230]. In AD RNA is extensively modified and, while clearly damaged, its rapid turnover may also serve a protective function [204]. Stress-activated protein kinase pathways triggered by OS, are excessively activated in AD, even in early stages, deregulating cyclin-dependent kinase 5, compromising the cellular antioxidant defence system and causing mitochondrial damage [231]. S-nitrosylation of dynamin-related protein 1 (Drp1) that is increased in AD brains, mediates AB-related mitochondrial fission and neuronal injury [232]. Antioxidants may protect neurons in AD and PD, reducing the risk of ND [233].

Oxidative stress in Parkinson disease

In PD, many biochemical changes indicate compromised antioxidant systems suggested to underlie cellular vulnerability to progressive OS which generates excessive ROS or free radicals in substantia nigra (SN) with subsequent cell damage [see 234-237]. Increase of iron in the SNp with a shift of Fe(II):Fe(III) of 2:1 as compared to 1:2 in controls and aggregation of τ , can promote DA synthesis with accompanying increased generation of reactive metabolites, leading to degeneration. Protein misfolding of sporadic PD has been associated with ROS formed as products of O₂ reduction by combination of DA and Fe [220]. In pigmented DAergic neurons of human substantia nigra pars compacta (SNc) recent proteomics have identified L-ferritin in neuromelanin granules, suggesting it as an important player in the complicated network governing Fe homeostasis in human DA neurons [238]. Both reduced glutathione, and glutathione peroxidase activity are decreased in SN of PD and in incidental LB disease (preclinical PD), probably preceding both complex I and DA loss [239]. Peroxynitrite, formed by reduced SOD, induces aggregation of aSyn in situ, and nitrated aSyn is found in the core of LBs indicating their in damaging structural proteins [236]. Cross-linking of α Syn by AGEs in PD and in incidental LB disease suggests that AGE-promoted LB formation may reflect early disease-specific changes, accelerating inclusion body formation [240]. Protein misfolding in sPD has been associated with ROS as products of O2 reduction by combination of DA and Fe [220]. Abnormal α Syn as found in LBs produces hydrogen peroxide and the neurons are capable of dissecting antioxidant enzymes to regions of OS, and that glutathione peroxidase-positive microglia are involved in neuroprotection in PD and DLB [241]. aSyn is up-regulated in a subset of neurons in response to chronic OS and is associated with neuroprotection from relatively low levels of OS. Formation of aSyn protofibrils is stimulated by translational modifications that occur under conditions of OS, while its aggregation is inhibited by antioxidants and various proteins with chaperone activity [242]. Many of the above factors demonstrated in Human PD and animal models indicate a multicomponent process in this NDD [see 243] and cell death pathways in PD are caused by many interacting factors [244, 245].

Oxidative stress in other neurodegenerative disorders

OS is also recognized as a major pathogenic factor in other NDDs [see 1]. In HD, where in both human brain and in tg mouse models, increased indices of OS markers have been reported, the crucial initiation mechanism induced by mhtt is still unclear, but early and critical involvement of defects in mitochondrial function and CNS energy metabolism may trigger the disease. Proteomic and OS analyses in human brain samples of HD indicate that OS and damage to specific macromolecules would participate in disease progression [246].

In PSP, increased lipid peroxidation in cerebral cortex [247] is proportional to the extent of τ pathology, increased activity of antioxidant systems, e.g. SOD and glutathione, is seen in multiple brain areas, and oxidative damage affects regions vulnerable to PSP and argyrophilic grain disease [248]. Increased OS is important in both sALS and fALS. The two principal hypotheses on the pathogenesis of ALS are: (i) oxidative damage stemming from aberrant SOD1 redox chemistry, and (ii) misfolding of the mutant protein. Enhanced basal oxidoradical products, lipid peroxide, perturbed calcium homeostasis and increased nitrotyrosine in lower motoneurons of both to mice and human ALS are present. At terminal stages disruption of glutathione peroxidase PrxII/GPxI-overexpression mechanism was observed, suggesting that the breakdown of this redox system accelerates neuronal degeneration and/or death. Because only a subset of ALS cases can be attributed to one particular deficit. *e.g.* mutation of SOD1. ALS aetiology is likely to be multifactorial [249]. Mutant SOD1 localized in the intermembrane space of mitochondria is sufficient to determine mitochondrial abnormalities and neuronal toxicity, thus contributing to ALS pathogenesis [250, 251]. Extracellular mutant SOD1 induces microglial-mediated motoneuron injury [442]. Two genes have recently be linked to fALS-TARDPD (TDP-43) and FUS (RNA binding protein fusion [69, 70], which comprises 3% and 4% of fALS, respectively. sALS has been associated with ECP3 gene, which encodes the catalytic subunit of the histone acetvltransferase complex elongator protein [69]. Identification of mutations in the gene encoding TDP-43, identified in both fALS and sALS stress the importance of TDP-43 in the pathogenesis of both types of ALS [252], but the precise mechanisms by which SOD1 leads to ND have not been defined. OS and cytoskeletal abnormalities causing increased protein glutathionylation leading to abnormalities in neuronal function are essential in Friedreich's ataxia [218].

In summary, free radical-mediated damage to lipids, proteins and nucleic acids is at least a part of the pathogenic events in the majority of NDDs. OS should be considered a window to view both some of the basic pathogenic cascades leading to ND as a means to design strategies to modify fundamental abnormalities, preventing or delaying disease progress. Induction of heat shock proteins has recently been suggested for protection against OS [91].

Impaired bioenergetics and mitochondrial dysfunction

Mitochondria, the 'energy powerhouse of the cell', are key cytoplasmic organelles vital for the function and survival of neurons. They provide energy from aerobic metabolism; oxidative phosphorylation *via* the oxidative phosphorylation system (OXPHOS) is the principal source of high-energy compounds [253]. These proteins link mitochondrial function and dynamics to the regulation of metabolism, cell-cycle control, development and cell death, giving evidence that provide molecular definition to mitochondria as a central platform in the execution of diverse cellular events, including cell death [254, 255].

Mitochondria are both targets and important sources of ROS. Generation of reactive oxidants, including ROS, is increased in damaged mitochondria and in cells with compromised mitochondrial function. Due to the heterogeneity of NS mitochondria and variations in mitochondrial functioning related to regionally distinct regulatory influences and dependence of their energy production to local demands, they show selective vulnerability to injury [256]. Recent studies have shown that healthy aging is associated with reduced neuronal mitochondrial metabolism and altered glial mitochondrial metabolism, which may be in part responsible for decline in brain function [443]. High levels of oxidants can induce the mitochondrial permeability transition, uncouple oxidative phosphorylation with catastrophic effects on mitochondrial energetics, and contribute to cytotoxicity via necrosis and/or apoptosis. Two major mechanisms of mitochondrial damage are the respiratory chain enzyme and mitochondrial DNA. OS and damage to mtDNA during aging impair mitochondrial energy metabolism and ion homeostasis in neurons, thereby rendering them vulnerable to degeneration [253, 257]. Disturbances of the mitochondrial proteolytic system affect neuronal maintenance and axonal function [444]. In NDD, there is a reciprocal interaction between mitochondrial fusion, fission, transport and mitophagy [258]. Impaired bioenergetics and dysfunction of mitochondrial energy metabolism lead to reduced ATP production, impaired calcium buffering and generation of ROS, representing a 'deadly triad in ND' [259]. Mitochondrial dysfunction initiates and propagates neuronal dysfunction in all age-associated NDDs [445].

In a variety of tissues, cumulative OS, disrupted mitochondrial respiration and oxidative mitochondrial damage are associated with, and may promote cell death [203, 253–255, 259]. Dysfunctions of mitochondria disturb cell function, cause mtDNA damage, sensitize cells to neurotoxic insults and may initiate cell death, all significant phenomena in the pathogenesis of NDDs [253, 257, 260]. The mtDNA may play an essential role in the pathogenesis of HD, that shows multiple mtDNA deletions in over 60% of HD patients, probably caused by CAG repeats instability and mhtt [261], that directly and indirectly impairs mitochondrial function [262].

OXPHOS disorders ranging from *Leigh disease* to PD and AD are caused by defects of mitochondrial energy metabolism, related to complex I deficiency [263]. There are several pathways by which both mitochondrial dysfunction and protein aggregation may interact (Fig. 4). Both aggregated SOD and A β in the mitochondrial matrix could contribute to cell death. The mitochondrial permeability transition pore, being crucial in both necrotic and apoptotic cell death, is controlled by anti-apoptotic members of the Bax/Bcl-2 family [264]. Mitochondrial Bax interacts with the voltage-dependent anion channel to accelerate opening of the permeability transition pore which is favoured by Ca²⁺ and oxidizing agents, contributing to cytochrome c release. Calcium regulation controls mitochondria motility and morphology in steady state,

stressed and pathological conditions [265, 266]. A consequence of mitochondrial dysfunction is increased generation of free radicals and oxidative damage [254].

A decrease in the DNA base excision repair activity in NDDs suggests that that the brain might be subjected to the double insult of increased DNA damage, as well as deficiencies of DNA repair pathways [267–269]. There is evidence of impaired DNA repair in both the aging brain and AD [270]. Although the molecular basis underlying the decline in mitochondrial function is not really understood, recent experimental evidence has shed some light on the pivotal role of mitochondrial morphology control and dysregulated mitochondrial fusion and fission events in ND [271]. Whether these changes are a secondary consequence of other factors or whether they cause eventual cell death is unknown. However, the identification of mutations in genes that encode proteins functioning in DNA repair and damage establish a mechanistic precedent that clearly links DNA damage and repair anomalies with progressive ND [268].

The importance of microRNAs, brain-enriched, small non-coding RNAs that participate in RNA translational and protein expression regulation, has been emphasized [272], suggesting their involvement in the emergence or progression of NDDs, including AD and prion-induced NDD [273–276].

Mitochondrial dysfunction in Alzheimer and Parkinson disease

In AD, there is a large body of evidence implicating impaired energy metabolism and oxidative damage [see 204, 209]. AD brains show an average 50% reduction in mtRNA content that, together with other changes, is likely to reduce oxidative phosphorylation. The 'mitochondrial cascade hypothesis', that has recently been updated, could explain many of the biochemical, genetic and pathological features of sporadic AD [277, 278]. Significant increase in oxidative damage to mtDNA in AD brain may cause abnormal mitochondrial dynamics and dysfunction that ultimately damage AD neurons [279]. AB plays a role in the formation of mitochondrial permeability transition pore (mPTP) [446]. AB-induced mitochondrial dysfunction is an early event in AD tg models [280] and is a trigger of AD pathophysiology [447]. Patterns in protein changes in early AD suggest activation of mitochondrial pathways that include proteins responsible for transport and utilization of ATP. The most prominent changes may occur in early AD [448]. Despite the evidence of mitochondrial dysfunction in AD, no causative mutations in the mtDNA have been detected so far, and results of studies on the role of mtDNA haplogroups in AD are controversial, although a primary role of the mitochondrial genome is suggested [278]. Both APP and γ -secretase, as well as an isoform of insulin-degrading enzyme, which regulates AB levels, are present in mitochondria [224], and mitochondrial OS causes hyperphosphorylation of τ [281]. Mitochondrial dysfunction in AD is caused by oligomeric AB [449], leading to impairment of ATP production and increase of OS. It may also be

Fig. 4 Schematic interaction of proteins in neurodegenerative diseases and mitochondria. Accumulation of mitochondrial DNA mutations may induce ROS production and cause oxidative damage in aging. In AD, ROS production and decreased ATP may contribute to production of AB peptides that may enter mitochondria, induce free radicals, decrease cytochrome oxidase activity and inhibit ATP generation. APP is transported to outer mitochondrial membranes, blocks import of nuclear cytchrome oxidase proteins to mitchondria, and may decrease cytochrome oxidase activity. In AD and models. Aß is found in mitochondrial matrix, produces free radicals and causes mitochondrial dysfunction. N-terminal portion of ApoE4, γ -secretase complex proteins, e.g. presenilins and nicastrin, associated



with mitochondria, may contribute to A β production and cause oxidative damage. In HD neurons, Htt binds to outer mitochondrial membrane und induces free radical production; H₂O₂ may also interrupt calcium uptake. In PD neurons, mutant proteins of α Syn, Parkin, PINK1 and DJ1 are associated with mitochondria and cause their dysfunction. In ALS, mutant SOD1, localized in inner and outer mitochondrial membranes and matrix, induces oxidative damage; associated with impairment of complexes II and IV. Frataxin, a gene product in FRA, is a mitochondrial protein responsible for heme biosynthesis and formation of iron-sulfur clusters, facilitating the accumulation of iron and inducing free radicals (modified from [257]).

caused by impaired axonal transport with proximal collection of mitochondria which could contribute to loss of distal synapses [282]. Calcium dysregulation occurring early in AD, related to PSEN mutations, A β production and tau phosphorylation [450] leads to synaptic and neuronal dysfunction, underlying dementia associated with the disease [451]. Caspase-cleaved τ expression results in mitochondrial dysfunction in cortical neurons *via* increased calcineurin activity in AD brains [283]. Significant increase in oxidative damage to mtDNA in AD brain may cause abnormal mitochondrial dynamics and dysfunction that ultimately damage AD neurons [279]. Mitochondrial ATP synthase in the entorhinal cortex is a target of OS in the first, clinically silent periods of AD pathology [284].

Mitochondrial alterations and OS are important parts of the multifactorial pathogenic process of PD [267, 285–288, 452]. In both sPD and fPD, abnormal mitochondrial paradigms include impaired function of the electron transport chain, damage to mtDNA, impaired calcium buffering, and anomalies in mitochondrial morphology and dynamics. Mitochondrial dysfunction triggers an increased free turbulin, which destabilizes the microtubule network and promotes α Syn oligomerization [453]. Since the UPS requires ATP at several steps, impaired mitochondrial

function may impair its activity. More importantly, increased production of ROS leads to damaged misfolded proteins requiring degradation by the UPS. Parkin plays a role in maintaining mitochondrial homeostasis [289]. It improves mitochondrial dysfunction, alters the intrinsic threshold for mitochondrial cytochrome c release, regulates their remodelling, promotes their autophagy, promotes DNA repair and protects against genotoxicity, promotes intracellular A β_{42} clearance [289–293] and regulates DNA repair [294], while PINK1 (PARK6), a fPD-associated gene, modulates mitochondrial morphogenesis and distribution [295]. Both are causes of autosomal recessive PD and induce the autophagic pathway (mitophagy) or defective oxidative phosphorylation [454, 455].

A specific protein–protein interaction of α Syn and COX, a key enzyme of the mitochondrial respiratory chain, suggests that α Syn aggregation may contribute to enhanced mitochondrial dysfunction [296].

Acute action of rotenone induces mitochondrial ROS production disrupting Ca²⁺ homeostasis in SNc but not in substantia nigra pars reticulata (SNr) neurons [297]. Both rotenone and chronic MPTP result in oxidative damage [298, 299] and α Syn aggregates closely resembling LBs, which were not reproduced by others [300, 301]. Mitochondrial complex I inhibition is not required in the models for DAergic cell death [302]. Subacute rotenone treatment of SH-SY5Y neuroblastoma cells reproduced Lewy neurites but not LB inclusions, reduces mitochondrial movement and slowly kills neural cells [303]. Mutations in parkin which cause recessive early-onset PD, are associated with marked mitochondrial abnormalities and less resistance to OS induced by paraquat. Reduction of cerebral mitochondrial dysfunction is a primary or secondary event, or part of a multifactorial pathogenic process remains to be elucidated [267]. mtDNA abnormalities and mitochondrial dysfunction are implicated in PD pathogenesis, and a link between mitochondrial dysfunction, OS, protein misfolding and abnormal autophagy is becoming increasingly prominent [304].

For clarification of NDDs with genetic causes of bioenergetic deficits see [1]. Energetic defects in major NDDs only in part have been proven by the identification of genetic defects that are causally linked to mitochondrial dysfunction. If this has a causative role to disease pathogenesis, then a number of therapeutic targets are implicated, including the permeability transition pore, cytochrome c release, free radical scavengers that could result in novel treatments. Mhtt causes calcium homeostasis and mitochondrial dysfunction in striatal neurons and excitotoxic neuronal death [305, 306, 456], and in models of Machado-Joseph disease, decreased synthesis of the protein Hsp27 and increased mitochondrial DNA damage are seen [307, 308]. Mitochondrial dysfunction has been linked to the ALS variants of SOD1 which is preferentially associated with mitochondria and subsequently impairs mitochondrial function [457] via the mitochondrial permeability transition pore (mPTP) [458].

Fragmentation of neuronal Golgi apparatus

Fragmentation of the neuronal Golgi apparatus-probably a secondary effect of the above mentioned basic mechanisms - was reported in ALS, corticobasal degeneration, AD, PD, Creutzfeldt-Jakob disease, and in SCA type 2 (SCA2) [309]. Mechanisms involved in Golgi fragmentation include: (i) dysregulation by mutant SOD1 of the microtubule-destabilizing protein Stathmin, (ii) disruption by mutant SOD1 of the neuronal cytoplasmic dynein, (iii) coprecipitation of mutant SOD1 with Hsp25 and Hsp27, (*iv*) reduction of detyrosinated microtubules by aggregated τ which resulted in non-apoptotic cell death and (v) disruption by mutant growth hormone of the trafficking from the rough ER to the Golgi apparatus [310]. α Syn has been shown to block ER Golgi traffic causing Golgi fragmentation and neuronal Golgi fragmentation is an early and probably irreversible lesion in ND in nigral neurons with α Syn⁺ inclusions in PD [311], caused by a variety of mechanisms including accumulation of misfolded proteins.

Disruption of cellular/axonal transport

There is growing evidence that defective neuronal and axonal transport due to early axonal dysfunction play a mechanistic role in most NDDs [282, 312, 313]. Whether misregulation of axonal transport has a direct role in the pathogenesis of these disorders or is a secondary phenomenon remains to be elucidated. Most of the transport uses the microtubule system that is stabilized by dystrophin [314] and forms a network of trafficking highways. Cargo is linked to the cytoskeleton by unidirectional proteins made of a motor domain that reversibly interacts with the cytoskeleton and converts chemical energy into motion. All axonal components are synthesized in the cell body and transported from there into the axonal processes (anterograde transport). A complementary mechanism transports cargo in the opposite direction, *i.e.* away from the axon into the cell body (retrograde transport). It is powered by members of the kinesin protein transport family, fast retrograde and slow anterograde transport, e.g. of neurofilament proteins, by dynein and is regulated by neurofilament subunit head domain phosphorylation [315-317]. Slow transport is a function of neuronal intermediate neurofilaments [318]. For intracellular transport which is fundamental for cellular functions, survival and morphogenesis, the kinesin superfamily transport proteins are important [319]. Neurofilament light chain head domain phosphorylation regulates axonal transport of neurofilaments [315].

Three mechanisms highlight the significance of disrupted cellular/axonal transport in human NDDs: (i) human motor protein mutations in these disorders. (ii) axonal transport defects in animal and in vitro cellular models harbouring human mutations and (*iii*) roles for pathogenic proteins like APP, τ , presenilin and synuclein, in the regulation of axonal transport [320]. The main pathogenic stimuli of these conditions, though often not definitely determined, result in an initial perturbation of the axon and its cytoskeleton, which then results in slow neuronal degeneration and loss of connectivity [312]. Mitochondrial transport in axons uses motors of kinesin families and cytoplasmic dynein, to translocate along microtubules, and bidirectional movement may be coordinated through interaction between dynein and kinesin-1. Protein defects that are directly or indirectly linked to axonal transport could be reversed by specific interventions stabilizing microtubules and repair or protect axons. The axonal transport of τ occurs via a mechanism utilizing fast transport motors, including the kinesin family of proteins, and that α Syn transport in neurons may involve both kinesin and dynein proteins [313].

Axonal transport in tauopathies

Axonal transport is impaired in AD and other tauopathies [321, 322], probably early in their pathogenesis. Proteolytic cleavage of APP occurs before its sorting into axonal transport vesicles [323]. It travels *via* fast axonal transport and in vesicular complexes containing presenilin and β -site APP-cleaving enzyme 1 (BACE1) and

acts as a receptor for the anterograde motor kinesin. Decreased retrograde transport of nerve growth factor (NGF) in human brain and mouse models leads to loss of neuronal markers and shrinkage of neurons in the cholinergic basal forebrain, rather than due to decreased synthesis. Misregulation of APP can transduce into misregulation of fast axonal transport, which is a pathogenic mechanism for intraneuronal AB [324]. There are multiple initiating factors converging upon pathways of amyloid induced defects in neuronal transport [325]. τ , is a microtubule-binding protein that, after its hyperphosphorylation and segregation into tangles, is unable to bind microtubuli, causing their destabilization and, thereby, disrupting axonal transport movement. The 3- and 4-repeat τ proteins show differential effects on mitochondrial axonal transport [326]. Interfering with axonal transport may activate stress kinase pathways initiating a biochemical cascade that drives normal τ proteins into a pathogenic state [327]. However, axonal transport rates in vivo are unaffected by τ deletion, or hyperexpression, in mice [328].

Axonal transport in synucleinopathies

 α Syn is a highly conserved protein, strongly expressed in neurons and enriched in presynaptic terminals. It is mainly transported in the slow, but a part also in fast transport. Significant age-related transport retardation may lead to accumulation of α Syn over time and produces pathology suggesting axonal transport abnormalities in synucleinopathies [329]. Accumulations of small α Syn aggregates in presynaptic terminals in the cortex of DLB and PD associated with loss of dendritic spines suggest a pathological impact on synaptic functions leading to ND, implicating PD as a synaptopathy [330].

Axonal transport in other neurodegenerative diseases

In SOD-1 tg models of ALS show retarded transport of neurofilament proteins and deficits in the delivery of mitochondria to the axon by fast transport, even before symptom onset implicated impaired transport as an early defect in the progress of ND, but, to date, no motor protein gene mutation in human ALS has been reported [331]. Mutant SOD1 interfering with axonal transport and protein turnover may influence the function and viability of motor neurons in ALS [332].

HD shows striatal degeneration with aggregation of huntingtin (htt) within vesicles and moves in the fast axonal transport component. In animal models, this was inhibited by infusion with pathological polyQ repeats and by disruption of the *Drosophila* htt gene. Htt plays a role in protein trafficking, vesicle transport, postsynaptic signalling, transcriptional regulation and apoptosis. A loss of function of the normal protein and a toxic gain of function of mhtt contribute to the disruption of multiple intracellular pathways [193]. An htt-binding protein HAP1 interacts with the dynactin complex, important for dynein and possibly kinesin fast movements, and accumulated vesicles and organelles in dystrophic axons from human HD patients suggest that aggregates of polyQ repeats disrupt fast axonal transport, and there is enhanced sensibility of striatal neurons to axonal transport defects induced by mhtt [333].

Dysfunction of neurotrophins

There is growing evidence that reduced neurotrophic support is a significant factor in the pathogenesis of NDDs [334]. NTFs affect neuronal survival: influence synaptic function and plasticity. NTFs bind to different receptors, to a common receptor, and each of them also binds to one of the family of Trk receptors. Since NTFs in neurons are subject to retrograde and to anterograde transport from and to targeting neurons, their effects may be related to synthesis in local or remote sites or to changes in axonal transport. In CNS disorders, such as AD, PD and HD, OS appears linked to the loss of neurotrophic support [201]; it can cause down-regulation of NTFs which, in turn, up-regulates antioxidant enzymes and promotes the expression of antioxidant proteins. Brain-derived neurotrophic factor (BDNF) levels in to mouse models of AD are negatively correlated with A β levels, suggesting that the effect of A β on decreased BDNF expression is specific to the aggregation state of AB and is dependent on large oligomers [335]. AD brain shows increase of the precursor form of NTF and decrease in BDNF in surviving neurons of hippocampus and neocortex, and decrease of TrkA in cortex and nucleus basalis [321]. AB may induce the TrkA pathway activation and promote NGF secretion [336, 337]. AB may act as a neurotrophic factor that mimics the activity of NGF. However, at higher concentrations, the amyloid behaves as an antagonist of NGF, contributing to the advent of AD [338]. AB42 may induce neuronal death through the p75 NTF receptor, and p75-Aβ-PrPC complexes could provide reactive OS and elevated intracellular calcium required for p75 signalling [339]. Postmortem studies point to a lack of NGF action in early AD stages, whereas NGF is found in enhanced concentrations in brains with severe AD partly due to a pathologically altered axonal transport of NGF in the neurons [340]. Advanced glycation and lipoxidation end-products modification of pro-NGF offers a novel pathway in the etiopathogenesis of AD [459]. Inhibition of soluble NTF signalling in a mouse model of AD prevents/slows pre-plaque amyloid-associated neuropathology, and potentially the progressive neuron 'loss' in AD [341, 342].

New steps and targets in the pathogenesis of AD may be: NGF and BDNF signalling through neurotrophic tyrosine kinases prevent the cleavage of APP by BACE1, which, together with γ -secretase, converts APP into toxic forming A β fragments, and a soluble N-terminal fragment (N-APP). In AD, this extracellular fragment of APP, and death receptor G may engage tumour necrosis factor (TNF) receptor family member 21 (TNFRSF21) which activates a widespread caspase-dependent self-destructing process – *via*

activating caspase-6 apoptosis-related cystein peptidase (casp6, MCH2)-related enzymes—as a major culprit behind AD [343]. This pro-apoptotic pathway proposed by these data may be relevant for other NDDs.

Studies in rodent and primate models of AD showed that BDNF prevents lesion-induced death of entorhinal neurons, reverses neuronal atrophy and ameliorates age-related cognitive impairment. A β -induced NGF dysmetabolism in AD may explain the paradoxical up-regulation of the precursor form of NGF (pro-NGF) in AD accompanied by atrophy of forebrain cholinergic neurons [344]. Protectional effects of BDNF on cortical neuronal circuitry involving AD, acting through amyloid-independent mechanisms [345], indicate a prominent role of NGF in both the aetiology and treatment of AD [346].

In PD, decreased neuronal content and their receptors in SN indicate a reduction of neurotrophic support and alterations in axon guidance in early stages of cellular stress, leading to dopaminergic cell death [334]. In HD, the mhtt induces a downregulation of BDNF in the basal ganglia, leading to neuronal loss, opening up the possibility of BDNF therapy. Transcription from BDNF promoter II and IV is down-regulated in human HD cortex from an early symptomatic stage. In addition to the reduction in BDNF mRNA, there may be unbalanced neurotrophic receptor signalling in HD [347]. In ALS, NGF concentrations and BDNF are up-regulated in early stages of the disease, whereas the levels of other NGFs gradually increase during the course of the disorder. In ALS spinal cords, TrkA was up-regulated, but the results of trials in both animal models and human patients were controversial. Therefore, comparative in situ data for transcription levels and protein contents in NTFs and their receptors in both sites of neuronal origin and termination in human brain are needed to understand their potential role in new treatment strategies [201, 345].

'Neuroinflammatory' processes

Chronic inflammatory reactions and signs of immune activation in the CNS, with major histocompatibility complex (MHC) class II expression, glial reaction, T-cell infiltration, and blood-brain barrier dysfunction are prominent pathological features in the pathogenesis and progression of NDDs [348]. Iron has various pro- and anti-inflammatory activities, and inappropriate iron chelation may be a major pathogenic contributor to inflammatory and NDDs [349]. AD 'inflammation' has been considered toxic or, on the contrary, useful [350]. However, a better knowledge about the interplay between the nervous system and the local and systemic immune system will be important for the understanding of pathogenic mechanisms of ND [351], the role of innate 'protective autoimmunity' and of reparative functions of autoimmunity in NDDs [352]. It has been suggested that immune alterations may occur prior to amyloid deposition and neuronal degeneration. Complement activation exacerbates the pathology of AD, and it has been discussed whether it could be the outcome of the innate immunity defence in the brain [353]. A central question is whether immune and inflammatory pathways become hyperactivated with age and promote ND or whether insufficient immune responses, which fail to cope with age-related stress, may contribute to NDDs [354]. Age-related neuroinflammatory changes negatively impact neuronal function [460]. Breakdown of the normal blood-brain barrier with influx of blood-born molecules (plasma proteins) have been suggested to cause local damage as starting mechanisms of some NDDs [348]. A leaky BBB, fibrinogen infiltration and microglial activity may cause neuronal damage in 'inflamed' AD brain [461].

Activated microglia may be both neurotoxic and neuroprotective, depending on several factors including aging, but the causes for transformation between the two actions remains largely unknown [355]. Microglia form the brain's innate immune cell type, but also are associated with constant phagocytotic clearance of cell debris, representing an essential link between degeneration and regeneration. Insufficient clearance by microglia, present in several NDDs and declining with aging, is associated with an inadequate regenerative response [356], while astrocytes produce anti-inflammatory and neuroprotective agents [357].

Components related to AD neuroinflammation include microglia and astrocytes, the classic and alternate pathways of the complement system, the pentraxins, acute-phase proteins, neuronal-type nicotinic acetylcholine receptors, peroxisomal proliferation-activated receptors, as well as 'pro-inflammatory' cytokines and chemokines. In animal models and human brains, both the microglia and astrocytes may generate AB, which itself may act as a pro-inflammatory agent inducing the activation of glia and many inflammatory components [358]. The patterns of cytochemokine involvement strengthen the inflammatory theory of AD and raise a pathophysiological role for selective alteration of this network [359]. Footprints of radicals and peroxynitrite attack have been detected in post-mortem AD brain, which, at least in part, are produced by activated microglia, and, together with anomalous release of cytokines and chemokines, but also systemic inflammation with increase in serum TNF α , may be a progression factor in AD [350, 360, 361]. In both aged and AD brain and in animal models, deposition of AB is likely to trigger inflammatory cascades with increased production of pro-inflammatory factors, part of which are localized in the AB plaques and in the surrounding activated glia [355]. Inflammation potentially increases brain levels of AB by three mechanisms: increased influx, decreased efflux, and increased neuronal production [362]. JNK-AP1 signalling pathway may be responsible for ABinduced neuroinflammation in cultured human brain endothelial cells and AD brain and this signalling pathway may serve as a therapeutic target for relieving A_B-induced inflammation [363], and TNF α gene polymorphism may affect the risk of developing AD [364]. AB oligomers and fibrils stimulate differential activation of microglia [358]. Inactivation of pro-inflammatory cytokines such as TNF, despite known adverse effects, makes it an attractive target for therapeutic development to treat NDDs [365]. Inhibiting p53 pathways in microglia alternates microglial

evolved neurotoxicity following exposure to Alzheimer peptides [462]. The problems of AD and tg animal models treated by immunotherapy causing plaque removal and increase in severity of cerebral amyloid angiopathy, but not preventing progressive ND, are not discussed here [see 366–368].

On the other hand, in human AD brain deposits of A\beta devoid of τ^+ structures were found to be co-localized with non-activated, ramified dystrophic microglia suggesting that Aβ does not trigger microglial activation [369]. These findings and recent experimental evidence [370] argue against the hypothesis that neuroinflammatory changes contribute to AD. They support the idea that progressive, age-related microglial degeneration and loss of microglial neuroprotection rather than induction of microglial activation contribute to the onset of sporadic AD [369].

In PD, SN cell degeneration is accompanied by astroglial reaction and proliferation of MHC class II positive microglia releasing pro-inflammatory cytokines, nitric oxide, complements, and OS that may be both inducing factors or sequelae of neuronal death [371, 463]. The pattern of humoural immune reactivity is consistent with activation of microglia leading to the targeting of DA nigral neurons for the destruction in both idiopathic and genetic cases of PD [372]. Parkin deficiency increases vulnerability to inflammation-related nigral degeneration, while human neuromelanin induces neuroinflammation and degeneration in the rat SN [373], and accregated α Syn activates microglia. DA cell death is probably influenced by the innate immune system, and in an MPTP mouse model T cell-mediated DAergic toxicity is almost exclusively arbitrated by CD4⁺ T cells and requires the Fas/FasL but not the IFNc cytotoxic pathway [374]. Intrastriatal lipopolysaccharide injection served as a model for inflammatory-induced mitochondrial dysfunction causing NDD in the nigrostriatal system [375]. Microglial activation and corresponding DAergic terminal loss in early PD support the notion that neuroinflammatory responses by intrinsic microglia contribute to the progressive degeneration in PD and related diseases [376]. Part of the specific vulnerability of the SN could be a consequence of $h-TNF\alpha$ hypomethylation [377], TNF α overexpression inducing apoptosis of neuronal cells. On the other hand, microglia may be affected by the disease process and may therefore not be capable of exerting sufficient neuroprotective function, such as glutathione peroxidase expression [241].

Microglia activation in HD has been shown to correlate with severity of the disease [378], with a close spatial and temporal relationship to neuronal dysfunction, and has already been detected in presymptomatic HD gene carriers [379]. BDNF and its receptor levels have been assessed in human cortices affected by HD [347]. Alterations of the adaptative immune system in sALS suggest early involvement of a 'neuroinflammative' process in the pathobiology of ALS [350, 380]. The role of inflammation in tg models of NDDs, in particular in mouse models of AD, have been reviewed recently [381]. In conclusion, whereas recent studies provide evidence for microglial abnormalities in several NDDs including ALS, HD and Creutzfeldt-Jakob disease [382, 383], the involvement of microglia in the pathogenesis of AD and PD remains controversial [369, 384].

The final pathway: multifaceted neuronal death

Neurons undergo diverse forms of cell death depending on the nature and severity of the stress. The nature, time course and molecular causes of cell death in NDDs and their relations to basic processes are still a matter of discussion [see 1]. Based on distinct morphologic criteria and biochemical features. PCD is classified into three major types: apoptosis (PCD type I), autophagy (PCD type II) and oncotic necrosis, a passive killing of the cell (PCD type III) [385-387]. Morphologically, apoptotis, dependent on caspase activation, is characterized by chromatin condensation (pyknosis), nuclear fragmentation, cell shrinkage and plasma membrane blabbing. The cell breaks into small membrane-surrounded fragments (apoptotic bodies), which are cleared by phagocytosis in vivo without inciting an inflammatory response. phagocytotic activity being balanced by positive and negative signals. Apoptosis can occur locally, without damaging healthy adjacent cells. This is in contrast to necrosis, an accidental and uncontrolled mode of cell death (PCD type III, not dependent on caspase activation), but triggered by OS [388], which exhibits rapid cell swelling and subsequent rupture of the plasma membrane that, due to an inflammatory response, usually induces substantial secondary cell damage in the surrounding tissue [389]. Autophagy (PCD type II), known as the process by which molecules and organelles undergoe lysosomal clearance/degradation in order to help maintain cellular homeostasis [160], also depends on caspase activation and a number of autophagy-related proteins and genes such as LC3 (light chain 3) and beclin-1 [171, 172]. Hundreds of caspase substartes have been described but only for a few of them the function of their cleavage by caspasin is well understood in the pathogenesis of NDDs [390]. There are many reasons and ways for a neuron to die, among which apoptosis is a specific form that is processed in two major signalling pathways. the extrinsic (death receptor) pathway and the intrinsic (mitochondria based) pathways, with several avenues of cross-talk between them (Fig. 5).

Many of the morphological differences between apoptotic and necrotic processes are thought to be a consequence of the action of cysteine proteases, while caspase and calpain functions in cell death are bridging the gap between apoptosis and necrosis. However, caspase-independent cell death also exists [388]. Tissue-specific impairment of autophagy in CNS tissue causes massive loss of neurons resulting in ND. Autophagic cell death. more correctly cell death with autophagy rather than by autophagy [391], probably represents a failure of neuroprotective mechanisms [392]. The prominence of autophagy in neurons contributing to the build-up of dysfunctional mitochondria and protein aggregates may lead to neuronal cell death, either due to an insufficient autophagic process or to up-regulation of autophagy [391]; both indicating a cross-talk between autophagy and the apoptotic pathways. Although markers for autophagy have been identified in different NDDs [393], it is not vet possible to be certain that these are indicative of the occurrence of autophagic cell death.



Fig. 5 Overview of apoptotic signalling through the receptor-mediated ('extrinsic') and mitochondria bases ('intrinsic') pathways, showing all key molecular players of apoptosis, the importance of the caspase cascade *via* interaction with different death domaines, and the role of effective caspase driving the execution of the cell death program.

Neuronal cell death may exhibit morphologic features of autophagy or necrosis, which differs from that of canonical apoptosis, or autophagic vacuolation can precede apoptotic cell death; this argues against the clear distinction between apoptotic and autophagic cell death [394]. Recent data point to the existence of multiple non-apoptotic, regulated cell death mechanisms, *e.g.* in tauopathies [101], some of which overlap or are mutually exclusive with apoptosis. Increasing evidence suggests that the regulation of neuronal cell death is complex, utilizing multiple pathways that are dependent on the damaging insult [395], each demonstrating specificity of function, regulation and pathway involvement, influenced by subtle differences among cell phenotypes (see Fig. 6).

Whilst post-mortem human brain tissue or those from animal models of injury or disease provide clues as to the diversity of neuronal death, the clearest information about molecular mecha-



Fig. 6 Diverse pathways leading to cell death, illustrated by the concept of the apoptosis-necrosis continuum that integrates the various death pathways and subsequent intracellular signalling pathways (ER stress, UPS, ATP loss, etc.) to help explain the complex patterns of neuronal death (mix of PCD-types I, II and /or III) (modified from [245]).

nisms leading to death comes from laboratory studies at the cellular level using cell models of injury or disease. Detailed studies of the molecular cell biology of appropriate cultured primary neurones, or immortalized cells with neuronal properties, have enabled the dissection of cellular responses and death outcome. There is an initial primary response to the trigger that may be external (*e.g.* toxic molecules impinging on the cell) or internal (*e.g.* inappropriate intracellular protein aggregation). The primary distortion of cellular biochemistry and physiology, if not rectified by the cell promptly and effectively, activates signalling pathways, both upstream and downstream of mitochondria, causing a crosstalk of PDC processes [245, 257]. Indeed, dying or dead cells may simultaneously manifest characteristics of more than one type of death pathway.

Cell death cascades in major NDDs

Despite demonstration of DNA fragmentation and up-regulation of pro-apoptotic and cell death regulator proteins, it is still unclear, whether apoptotic or necrotic modes are responsible for cell death in NDDs. It is preceded by the activation of caspases and altered expression of pro-apoptotic members of the Bcl-2 family and other ARPs. Multiple caspases and elevated caspase mRNAs have been detected in post-mortem tissue from AD brains [396], while others observed no apoptotic morphology in AD [397].

Frequent DNA fragmentation and the 'pro-apoptotic' environment in AD brain indicate increased vulnerability of neurons to metabolic and other noxious factors. Recent data suggest that the interaction between APP and/or AB and the cell death mediates p75 (NTR) (the common NTF receptor) and its interaction with pro-apoptotic ligands cause a selective vulnerability of neurons in the cholinergic forebrain in APP [341]. Caspase-3 has been found to be enriched in post-synaptic densities and increased in AD [398] and activation of caspases by PSEN1 gene and its inhibition by secretase inhibition were reported [399]. In surviving neurons, it can be suggested that viability is, in part, maintained by the lack of distal transmission of the caspase-mediated apoptotic signals. This phenomenon of apoptotic avoidance termed abortive apoptosis or abortosis, may represent an exit from the caspase-induced death program [400]. That, given the robust survivals of neurons with NFTs [401] and recent evidence that NFTs in experimental models do not directly correlate with neuron loss [402] suggests that affected neurons may be able to withhold NFT formation for a long time before they degenerate. Despite a lack of clear evidence linking τ aggregates to neuronal loss, the presence of a tangle in a neuron is likely to be harmful, leading to changes in axonal transport and loss of synapses [403]. Associations between tangle bearing neurons with caspase activation suggests that tangles are at least markers of neuronal disease [101]. Although considered neuroprotective, like other significant intracellular and neuritic protein aggregations they finally contribute to dysfunction and death of involved neurons.

Recent data indicating a link between the development of Aβ and NFT/ τ pathologies may be due to caspase activation and cleavage of APP which facilitate production of Aβ and by cleavage of τ may initiate or accelerate the development of tangle pathology [404, 405]. In several tg τ models, an increased number of TUNEL⁺ neurons and ultrastructural features of both apoptosis and necrosis, but no activated caspases were observed [406], while in aging PS/APP mice a crosstalk between apoptosis and autophagy was identified, which influences neuronal survival in AD-related ND [407]. However, many questions remain open about how changes in τ lead to ND and the relationship of NFT pathology to neuronal death, which recently is proposed to be a non-apoptotic caspase-associated form of death [101].

Cell death is a significant part of the pathology of PD. Although the process is mysterious, the prime suspect for a toxic protein is α Syn and its aberrant forms, including increased expression of the normal gene [408]. Apoptosis may be involved in PD, although the molecular mechanisms that initiate dopaminergic neuron loss are not known. These include, among others, JNK signalling, p53 activation, cell-cycle reactivation and signalling through Bcl-2 proteins [244]. Whether PCD actually occurs within the human PD brain remains controversial, and this possibility has been neither confirmed by numerous studies nor definitely excluded [see 1]. The question whether LBs and other α Syn aggregates are harmful or cytoprotective still remains unresolved. Although their formation may reflect one of several response patterns by the CNS to upstream dysregulation of α Syn metabolism, the α Syn pathway appears to be an essential factor for the selective multisystemic loss of neurons and glia in many synucleinopaties, *e.g.* PD, PDD, DLB, multiple system atrophy (MSA), and related disorders, showing widespread occurrence of LBs and dystrophic neurites with neuronal loss in many regions of the central, peripheral and autonomic nervous system. In sporadic ALS persitent cleavage and nuclear translocation of AIP in motor neurons in the spinal cord have been described recently [409].

A specific issue is whether a particular NDD is cellautonomous, where neurons form the primary defective entities in isolation from surrounding cells. Additionally, there are also functional changes in glial cells, such as astrocytes, which normally sustain neurones. There is a glutamatergic interplay between neurons and astrocytes, disturbances of which may be involved in neuronal derangement and contribute to disease development [410-412]. Such glial perturbations (non-cell-autonomous) are thought to apply in several NDDs [413]. The heterogeneous astroglial responses in various synucleinopathies, tauopathies and other NDDs indicate distinct underlying pathogenic mechanisms in each disorder [414], while in other diseases, e.g. MSA, there is a definite relation between oligodendrogliopathy and neuronal degeneration [415, 416]. While astrocytic dysfunction could play its part in the progression and severity of AD [417], the role of astrocytes in PD, ALS and related disorders is poorly understood [414, 418, 419]. Astrocytes display very different biological profiles when they become 'reactive' and an emergent literature indicates the normal homeostatic or sustenance relationships, are disrupted turning them instead into 'collaborators in neurotoxicity' [see 420, 421].

Post-mortem analysis can bridge some but not all of our knowledge gaps; the results are still controversial, and we need a better understanding of the molecular basis and pathways that drive the yin-yang between neuronal survival and death. The combined use of *in vitro* models and new analytical models should help to map environmental and toxic pathways in the aetiology of NDDs [422]. Understanding neuronal death pathways and their cross-talk not only informs the detailed pathobiology but also suggests novel therapeutic modalities, some of which have been reviewed recently [2, 91, 133, 154, 198, 201, 254, 366, 405, 423]. An extensive discussion of effective prophylactic and treatment strategies appears to be outside the goal of the present review.

Acknowledgements

The author thanks Mr. E. Mitter-Ferstl, Ph.D., for secretarial and computer work. The study was supported by the Society for Supporting the Research in Experimental Neurology, Vienna, Austria.

References

- Jellinger KA. Recent advances in our understanding of neurodegeneration. J Neural Transm. 2009; 116: 1111–62.
- Skovronsky DM, Lee VM-Y, Trojanowski JQ. Neurodegenerative diseases: new concepts of pathogenesis and their therapeutic implications. *Annu Rev Pathol Mech Dis.* 2006; 1: 151–70.
- Golde TE, Miller VM. Proteinopathyinduced neuronal senescence: a hypothesis for brain failure in Alzheimer's and other neurodegenerative diseases. *Alzheimers Res Ther.* 2009; 1: 5.
- Uversky VN. Intrinsic disorder in proteins associated with neurodegenerative diseases. *Front Biosci.* 2009; 14: 5188–238.
- Herczenik E, Gebbink MF. Molecular and cellular aspects of protein misfolding and disease. *FASEB J.* 2008; 22: 2115–33.
- Ovádi J, Orosz F. Protein folding and misfolding: neurodegenerative diseases. Netherlands: Springer; 2009. p. 274.
- Dickson DW. Neuropathology of non-Alzheimer degenerative disorders. Int J Clin Exp Pathol. 2010; 3: 1–22.
- Kovacs GG, Budka H. Protein-based neuropathology and molecular classification of human neurodegenerative diseases. In: Ovádi J, Orosz F, editors. Protein folding and misfolding: neurodegenerative diseases. Netherlands: Springer; 2009. pp. 251–72.
- Migliore L, Coppede F. Genetics, environmental factors and the emerging role of epigenetics in neurodegenerative diseases. *Mutat Res.* 2009; 667: 82–97.
- Palop JJ, Chin J, Mucke L. A network dysfunction perspective on neurodegenerative diseases. *Nature*. 2006; 443: 768–73.
- Seeley WW, Crawford RK, Zhou J, et al. Neurodegenerative diseases target largescale human brain networks. *Neuron*. 2009; 62: 42–52.
- 12. Ecroyd H, Carver JA. Unraveling the mysteries of protein folding and misfolding. *IUBMB Life.* 2008; 60: 769–74.
- Morris AM, Watzky MA, Finke RG. Protein aggregation kinetics, mechanism, and curve-fitting: a review of the literature. *Biochim Biophys Acta*. 2009; 1794: 375–97.
- Israeli E, Sharon R. Beta-synuclein occurs in vivo in lipid-associated oligomers and forms hetero-oligomers with alpha-synuclein. J Neurochem. 2009; 108: 465–74.
- 15. Selkoe DJ. Soluble oligomers of the amyloid beta-protein impair synaptic plasticity

and behavior. *Behav Brain Res.* 2008; 192: 106–13.

- Soto C, Estrada LD. Protein misfolding and neurodegeneration. *Arch Neurol.* 2008; 65: 184–9.
- Aguzzi A, Sigurdson C, Heikenwaelder M. Molecular mechanisms of prion pathogenesis. Annu Rev Pathol. 2008; 3: 11–40.
- Lafaye P, Achour I, England P, et al. Single-domain antibodies recognize selectively small oligomeric forms of amyloid beta, prevent Abeta-induced neurotoxicity and inhibit fibril formation. *Mol Immunol.* 2009; 46: 695–704.
- Moore RA, Taubner LM, Priola SA. Prion protein misfolding and disease. *Curr Opin Struct Biol.* 2009; 19: 14–22.
- Williams AJ, Paulson HL. Polyglutamine neurodegeneration: protein misfolding revisited. *Trends Neurosci.* 2008; 31: 521–8.
- Xia W, Yang T, Shankar G, et al. A specific enzyme-linked immunosorbent assay for measuring beta-amyloid protein oligomers in human plasma and brain tissue of patients with Alzheimer disease. *Arch Neurol.* 2009; 66: 190–9.
- Tomic JL, Pensalfini A, Head E, et al. Soluble fibrillar oligomer levels are elevated in Alzheimer's disease brain and correlate with cognitive dysfunction. *Neurobiol Dis.* 2009; 35: 352–8.
- Harmeier A, Wozny C, Rost BR, et al. Role of amyloid-beta glycine 33 in oligomerization, toxicity, and neuronal plasticity. J Neurosci. 2009; 29: 7582–90.
- Kim J, Onstead L, Randle S, et al. Abeta40 inhibits amyloid deposition in vivo. J Neurosci. 2007; 27: 627–33.
- Schmidt M, Sachse C, Richter W, et al. Comparison of Alzheimer Abeta(1–40) and Abeta(1–42) amyloid fibrils reveals similar protofilament structures. Proc Natl Acad Sci USA. 2009; 106: 19813–8.
- Lauren J, Gimbel DA, Nygaard HB, et al. Cellular prion protein mediates impairment of synaptic plasticity by amyloid-beta oligomers. *Nature*. 2009; 457: 1128–32.
- Shankar GM, Li S, Mehta TH, et al. Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat Med.* 2008; 14: 837–42.
- Abramov E, Dolev I, Fogel H, et al. Amyloid-beta as a positive endogenous regulator of release probability at hip-

pocampal synapses. *Nat Neurosci.* 2009; 12: 1567–76.

- Nygaard HB, Strittmatter SM. Cellular prion protein mediates the toxicity of betaamyloid oligomers: implications for Alzheimer disease. Arch Neurol. 2009; 66: 1325–8.
- Nimmrich V, Ebert U. Is Alzheimer's disease a result of presynaptic failure? Synaptic dysfunctions induced by oligomeric beta-amyloid. *Rev Neurosci.* 2009; 20: 1–12.
- Dinamarca MC, Colombres M, Cerpa W, et al. Beta-amyloid oligomers affect the structure and function of the postsynaptic region: role of the Wnt signaling pathway. Neurodegener Dis. 2008; 5: 149-52.
- Moreno H, Yu E, Pigino G, et al. Synaptic transmission block by presynaptic injection of oligomeric amyloid beta. Proc Natl Acad Sci USA. 2009; 106: 5901–6.
- Deshpande A, Kawai H, Metherate R, et al. A role for synaptic zinc in activitydependent Abeta oligomer formation and accumulation at excitatory synapses. J Neurosci. 2009; 29: 4004–15.
- Wang Z, Wang B, Yang L, et al. Presynaptic and postsynaptic interaction of the amyloid precursor protein promotes peripheral and central synaptogenesis. J Neurosci. 2009; 29: 10788–801.
- Hoe HS, Fu Z, Makarova A, et al. The effects of amyloid precursor protein on postsynaptic composition and activity. *J Biol Chem.* 2009; 284: 8495–506.
- Noguchi A, Matsumura S, Dezawa M, et al. Isolation and characterization of patient-derived, toxic, high mass amyloid beta-protein (Abeta) assembly from Alzheimer disease brains. J Biol Chem. 2009; 284: 32895–905.
- Gunzburg MJ, Perugini MA, Howlett GJ. Structural basis for the recognition and cross-linking of amyloid fibrils by human apolipoprotein E. *J Biol Chem.* 2007; 282: 35831–41.
- Vintem AP, Henriques AG, da Cruz ESOA, et al. PP1 inhibition by Abeta peptide as a potential pathological mechanism in Alzheimer's disease. *Neurotoxicol Teratol.* 2009; 31: 85–8.
- Karlnoski RA, Rosenthal A, Kobayashi D, et al. Suppression of amyloid deposition leads to long-term reductions in Alzheimer's pathologies in Tg2576 mice. J Neurosci. 2009; 29: 4964–71.

- Huang HC, Jiang ZF. Accumulated amyloid-beta peptide and hyperphosphorylated tau protein: relationship and links in Alzheimer's disease. *J Alzheimers Dis.* 2009; 16: 15–27.
- De Felice FG, Wu D, Lambert MP, et al. Alzheimer's disease-type neuronal tau hyperphosphorylation induced by A beta oligomers. *Neurobiol Aging.* 2008; 29: 1334–47.
- Kellner A, Matschke J, Bernreuther C, et al. Autoantibodies against beta-amyloid are common in Alzheimer's disease and help control plaque burden. Ann Neurol. 2009; 65: 24–31.
- Meli G, Visintin M, Cannistraci I, et al. Direct in vivo intracellular selection of conformation-sensitive antibody domains targeting Alzheimer's amyloid-beta oligomers. J Mol Biol. 2009; 387: 584–606.
- Relini A, Cavalleri O, Rolandi R, et al. The two-fold aspect of the interplay of amyloidogenic proteins with lipid membranes. Chem Phys Lipids. 2009; 158: 1–9.
- Oddo S, Caccamo A, Smith IF, et al. A dynamic relationship between intracellular and extracellular pools of Abeta. Am J Pathol. 2006; 168: 184–94.
- LaFerla FM, Green KN, Oddo S. Intracellular amyloid-beta in Alzheimer's disease. *Nat Rev Neurosci.* 2007; 8: 499–509.
- Savioz A, Leuba G, Vallet PG, et al. Contribution of neural networks to Alzheimer disease's progression. Brain Res Bull. 2009; 80: 309–14.
- Sahara N, Maeda S, Takashima A. Tau oligomerization: a role for tau aggregation intermediates linked to neurodegeneration. *Curr Alzheimer Res.* 2008: 5: 591–8.
- Brunden KR, Trojanowski JQ, Lee VM. Evidence that non-fibrillar tau causes pathology linked to neurodegeneration and behavioral impairments. *J Alzheimers Dis.* 2008; 14: 393–9.
- Small SA, Duff K. Linking Abeta and tau in late-onset Alzheimer's disease: a dual pathway hypothesis. *Neuron.* 2008; 60: 534–42.
- Barrantes A, Sotres J, Hernando-Perez M, et al. Tau aggregation followed by atomic force microscopy and surface plasmon resonance, and single molecule tau-tau interaction probed by atomic force spectroscopy. J Alzheimers Dis. 2009; 18: 141–51.
- 52. Frost B, Jacks RL, Diamond MI. Propagation of tau misfolding from the

outside to the inside of a cell. *J Biol Chem.* 2009; 284: 12845–52.

- Ren PH, Lauckner JE, Kachirskaia I, et al. Cytoplasmic penetration and persistent infection of mammalian cells by polyglutamine aggregates. Nat Cell Biol. 2009; 11: 219–25.
- Clavaguera F, Bolmont T, Crowther RA, et al. Transmission and spreading of tauopathy in transgenic mouse brain. Nat Cell Biol. 2009; 11: 909–13.
- Jeganathan S, von Bergen M, Mandelkow EM, et al. The natively unfolded character of tau and its aggregation to Alzheimer-like paired helical filaments. *Biochemistry*. 2008; 47: 10526–39.
- Kazantsev AG, Kolchinsky AM. Central role of alpha-synuclein oligomers in neurodegeneration in Parkinson disease. *Arch Neurol.* 2008; 65: 1577–81.
- Uversky VN. Neuropathology, biochemistry, and biophysics of alpha-synuclein aggregation. J Neurochem. 2007; 103: 17–37.
- Waxman EA, Giasson BI. Molecular mechanisms of alpha-synuclein neurodegeneration. *Biochim Biophys Acta*. 2009; 1792: 616–24.
- Crews L, Tsigelny I, Hashimoto M, et al. Role of synucleins in Alzheimer's disease. *Neurotox Res.* 2009; 16: 306–17.
- Cullen V, Lindfors M, Ng J, et al. Cathepsin D expression level affects alphasynuclein processing, aggregation, and toxicity in vivo. *Mol Brain.* 2009; DOI: 10.1186/1756-6606-2-5.
- Duka T, Duka V, Joyce JN, et al. Alpha-Synuclein contributes to GSK-3beta-catalyzed Tau phosphorylation in Parkinson's disease models. FASEB J. 2009; 23: 2820–30.
- Paleologou KE, Kragh CL, Mann DM, et al. Detection of elevated levels of soluble alpha-synuclein oligomers in post-mortem brain extracts from patients with dementia with Lewy bodies. Brain. 2009; 132: 1093–101.
- Danzer KM, Krebs SK, Wolff M, et al. Seeding induced by alpha-synuclein oligomers provides evidence for spreading of alpha-synuclein pathology. J Neurochem. 2009; 111: 192–203.
- Luk KC, Song C, O'Brien P, et al. Exogenous alpha-synuclein fibrils seed the formation of Lewy body-like intracellular inclusions in cultured cells. Proc Natl Acad Sci USA. 2009; 106: 20051–6.
- Desplats P, Lee HJ, Bae EJ, et al. Inclusion formation and neuronal cell death through neuron-to-neuron transmis-

sion of alpha-synuclein. *Proc Natl Acad Sci USA*. 2009; 106: 13010–5.

- Li C, Lutz EA, Slade KM, et al. 19F NMR studies of alpha-synuclein conformation and fibrillation. *Biochemistry*. 2009; 48: 8578–84.
- Emadi S, Kasturirangan S, Wang MS, et al. Detecting morphologically distinct oligomeric forms of alpha-synuclein. *J Biol Chem.* 2009; 284: 11048–58.
- Banci L, Bertini I, Boca M, et al. Structural and dynamic aspects related to oligomerization of apo SOD1 and its mutants. Proc Natl Acad Sci USA. 2009; 106: 6980–5.
- Kwiatkowski TJ Jr, Bosco DA, Leclerc AL, et al. Mutations in the FUS/TLS gene on chromosome 16 cause familial amyotrophic lateral sclerosis. *Science*. 2009; 323: 1205–8.
- Neumann M, Sampathu DM, Kwong LK, et al. Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science*. 2006; 314: 130–3.
- Liu HN, Sanelli T, Horne P, et al. Lack of evidence of monomer/misfolded superoxide dismutase-1 in sporadic amyotrophic lateral sclerosis. Ann Neurol. 2009; 66: 75–80.
- Alberti S, Halfmann R, King O, et al. A systematic survey identifies prions and illuminates sequence features of prionogenic proteins. *Cell.* 2009; 137: 146–58.
- Fontaine SN, Brown DR. Mechanisms of prion protein aggregation. Protein Pept Lett. 2009; 16: 14–26.
- Kellett KA, Hooper NM. Prion protein and Alzheimer disease. *Prion*. 2009: 3: 190–4.
- Matilla-Duenas A, Sanchez I, Corral-Juan M, et al. Cellular and molecular pathways triggering neurodegeneration in the spinocerebellar ataxias. *Cerebellum.* 2009; DOI: 10.1007/s12311-009-0144-2.
- Schwartz AL, Ciechanover A. Targeting proteins for destruction by the ubiquitin system: implications for human pathobiology. Ann Rev Pharmacol Toxicol. 2009; 49: 73–96.
- Muchowski PJ, Wacker JL. Modulation of neurodegeneration by molecular chaperones. Nat Rev Neurosci. 2005; 6: 11–22.
- Hartl FU, Hayer-Hartl M. Converging concepts of protein folding in vitro and in vivo. *Nat Struct Mol Biol.* 2009; 16: 574–81.
- Kubota H. Quality control against misfolded proteins in the cytosol: a network for cell survival. *J Biochem.* 2009; 146: 609–16.
- Scheper W, Hoozemans JJ. Endoplasmic reticulum protein quality control in neurodegenerative disease: the good, the bad and the therapy. *Curr Med Chem.* 2009; 16: 615–26.

- Eliezer D. Biophysical characterization of intrinsically disordered proteins. *Curr Opin Struct Biol.* 2009; 19: 23–30.
- Mendoza-Espinosa P, Garcia-Gonzalez V, Moreno A, et al. Disorder-to-order conformational transitions in protein structure and its relationship to disease. *Mol Cell Biochem.* 2009; 330: 105–20.
- Balch WE, Morimoto RI, Dillin A, et al. Adapting proteostasis for disease intervention. Science. 2008; 319: 916–9.
- Winklhofer KF, Tatzelt J, Haass C. The two faces of protein misfolding: gain- and loss-of-function in neurodegenerative diseases. *EMBO J.* 2008; 27: 336–49.
- Mirzaei H, Regnier F. Protein:protein aggregation induced by protein oxidation. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2008; 873: 8–14.
- Hutt DM, Powers ET, Balch WE. The proteostasis boundary in misfolding diseases of membrane traffic. *FEBS Lett.* 2009; 583: 2639–46.
- Powers ET, Morimoto RI, Dillin A, et al. Biological and chemical approaches to diseases of proteostasis deficiency. Annu Rev Biochem. 2009; 78: 959–91.
- Arndt V, Rogon C, Hohfeld J. To be, or not to be-molecular chaperones in protein degradation. *Cell Mol Life Sci.* 2007; 64: 2525–41.
- Broadley SA, Hartl FU. The role of molecular chaperones in human misfolding diseases. *FEBS Lett.* 2009; 583: 2647–53.
- Roelofs J, Park S, Haas W, et al. Chaperone-mediated pathway of proteasome regulatory particle assembly. *Nature*. 2009; 459: 861–5.
- Kalmar B, Greensmith L. Induction of heat shock proteins for protection against oxidative stress. *Adv Drug Deliv Rev.* 2009; 61: 310–8.
- Calderwood SK, Murshid A, Prince T. The shock of aging: molecular chaperones and the heat shock response in longevity and aging-a mini-review. *Gerontology*. 2009; 55: 550-8.
- Goldbaum O, Riedel M, Stahnke T, et al. The small heat shock protein HSP25 protects astrocytes against stress induced by proteasomal inhibition. *Glia.* 2009; 57: 1566–77.
- Roodveldt C, Bertoncini CW, Andersson A, et al. Chaperone proteostasis in Parkinson's disease: stabilization of the Hsp70/alpha-synuclein complex by Hip. EMBO J. 2009; 28: 3758–70.
- 95. Malhotra JD, Kaufman RJ. The endoplasmic reticulum and the unfolded protein

response. *Semin Cell Dev Biol*. 2007; 18: 716–31.

- 96. Lindholm D, Wootz H, Korhonen L. ER stress and neurodegenerative diseases. *Cell Death Differ.* 2006; 13: 385–92.
- Uversky VN. Alpha-synuclein misfolding and neurodegenerative diseases. *Curr Protein Pept Sci.* 2008; 9: 507–40.
- Tan JM, Wong ES, Lim KL. Protein misfolding and aggregation in Parkinson disease. *Antioxid Redox Signal.* 2009; 11: 2119–34.
- Mondragon-Rodriguez S, Basurto-Islas G, Binder LI, et al. Conformational changes and cleavage; are these responsible for the tau aggregation in Alzheimers disease? Future Neurol. 2009; 4: 39–53.
- 100. **Takashima A.** Hyperphosphorylated tau is a cause of neuronal dysfunction in tauopathy. *J Alzheimers Dis.* 2008; 14: 371–5.
- 101. Spires-Jones TL, Stoothoff WH, de Calignon A, et al. Tau pathophysiology in neurodegeneration: a tangled issue. *Trends Neurosci.* 2009; 32: 150–9.
- Hatters DM. Protein misfolding inside cells: the case of huntingtin and Huntington's disease. *IUBMB Life*. 2008; 60: 724–8.
- Okamoto S, Pouladi MA, Talantova M, et al. Balance between synaptic versus extrasynaptic NMDA receptor activity influences inclusions and neurotoxicity of mutant huntingtin. *Nat Med.* 2009; 15: 1407–13.
- 104. Reijonen S, Putkonen N, Norremolle A, et al. Inhibition of endoplasmic reticulum stress counteracts neuronal cell death and protein aggregation caused by N-terminal mutant huntingtin proteins. Exp Cell Res. 2008; 314: 950–60.
- 105. Woulfe J. Nuclear bodies in neurodegenerative disease. *Biochim Biophys Acta*. 2008; 1783: 2195–206.
- 106. Castellani RJ, Zhu X, Lee H-G, et al. Molecular pathogenesis of Alzheimer disease: reductionist versus expansionist approaches. Int J Mol Sci. 2009; 10: 1386–406.
- 107. **Austin RC.** The unfolded protein response in health and disease. *Antioxid Redox Signal.* 2009; 11: 2279–87.
- 108. Schroder M. Endoplasmic reticulum stress responses. *FASEB J.* 2008; 65: 862–94.
- Bezprozvanny I. Calcium signaling and neurodegenerative diseases. *Trends Mol Med.* 2009; 15: 89–100.
- Marambaud P, Dreses-Werringloer U, Vingtdeux V. Calcium signaling in neurodegeneration. *Mol Neurodegener*. 2009; 4: 20.

- 111. Hoozemans JJ, van Haastert ES, Nijholt DA, et al. The unfolded protein response is activated in pretangle neurons in Alzheimer's disease hippocampus. Am J Pathol. 2009; 174: 1241–51.
- 112. Hoozemans JJM, Scheper W. Endoplasmic reticulum stress in neurodegeneration. In: Ovádi J, Orosz F, editors. Protein folding and misfolding: neurodegenerative diseases. Netherlands: Springer; 2009. pp. 111–32.
- 113. Inoue H, Kondo T, Lin L, *et al.* Protein misfolding and axonal protection in neurodegenerative diseases. In: Ovádi J, Orosz F, editors. Protein folding and misfolding: neurodegenerative diseases. Netherlands: Springer; 2009. pp. 97–110.
- Leverenz JB, Umar I, Wang Q, et al. Proteomic identification of novel proteins in cortical lewy bodies. *Brain Pathol.* 2007; 17: 139–45.
- 115. Burns SA, Moran CR, Pingerelli PL, et al. Redefining the Lewy body: evidence of distinct protein composition and nonrandom assembly process (abstract). *Mov Disord*. 2008; 23: S22.
- Tai HC, Schuman EM. Ubiquitin, the proteasome and protein degradation in neuronal function and dysfunction. *Nat Rev Neurosci.* 2008; 9: 826–38.
- 117. **Strobel G.** The spectrum series: grappling with the overlap between Alzheimer's and Parkinson's diseases. *J Alzheimers Dis.* 2009; 18: 625–40.
- 118. Arai T, Mackenzie IR, Hasegawa M, et al. Phosphorylated TDP-43 in Alzheimer's disease and dementia with Lewy bodies. Acta Neuropathol. 2009; 117: 125–36.
- 119. Fujishiro H, Tsuboi Y, Lin WL, et al. Colocalization of tau and alpha-synuclein in the olfactory bulb in Alzheimer's disease with amygdala Lewy bodies. Acta Neuropathol. 2008; 116: 17–24.
- 120. Tsigelny IF, Crews L, Desplats P, et al. Mechanisms of hybrid oligomer formation in the pathogenesis of combined Alzheimer's and Parkinson's diseases. *PLoS One.* 2008; 3: e3135.
- 121. Clarimon J, Molina-Porcel L, Gomez-Isla T, et al. Early-onset familial lewy body dementia with extensive tauopathy: a clinical, genetic, and neuropathological study. J Neuropathol Exp Neurol. 2009; 68: 73–82.
- 122. Takahashi RH, Capetillo-Zarate E, Lin MT, et al. Co-occurrence of Alzheimer's disease beta-amyloid and tau pathologies at synapses. *Neurobiol Aging*. 2008; in press: doi:10.1016/j.neurobiolaging. 2008.07.021.

- 123. Muntane G, Dalfo E, Martinez A, et al. Phosphorylation of tau and alpha-synuclein in synaptic-enriched fractions of the frontal cortex in Alzheimer's disease, and in Parkinson's disease and related alphasynucleinopathies. *Neuroscience*. 2008; 152: 913–23.
- 124. Duka T, Rusnak M, Drolet RE, et al. Alpha-synuclein induces hyperphosphorylation of Tau in the MPTP model of parkinsonism. FASEB J. 2006; 20: 2302–12.
- 125. Shao CY, Crary JF, Rao C, et al. Atypical protein kinase C in neurodegenerative disease II: PKCiota/lambda in tauopathies and alpha-synucleinopathies. J Neuropathol Exp Neurol. 2006: 65: 327–35.
- 126. Higashi S, Iseki E, Yamamoto R, et al. Concurrence of TDP-43, tau and alphasynuclein pathology in brains of Alzheimer's disease and dementia with Lewy bodies. Brain Res. 2007; 1184: 284–94.
- 127. Riedel M, Goldbaum O, Richter-Landsberg C. alpha-Synuclein promotes the recruitment of tau to protein inclusions in oligodendroglial cells: effects of oxidative and proteolytic stress. J Mol Neurosci. 2009: 39: 226–34.
- 128. Obi K, Akiyama H, Kondo H, et al. Relationship of phosphorylated alphasynuclein and tau accumulation to Abeta deposition in the cerebral cortex of dementia with Lewy bodies. *Exp Neurol.* 2008; 210: 409–20.
- 129. Mandal PK, Pettegrew JW, Masliah E, et al. Interaction between Abeta peptide and alpha synuclein: molecular mechanisms in overlapping pathology of Alzheimer's and Parkinson's in dementia with Lewy body disease. Neurochem Res. 2006: 31: 1153–62.
- Fein JA, Sokolow S, Miller CA, et al. Colocalization of amyloid beta and tau pathology in Alzheimer's disease synaptosomes. *Am J Pathol.* 2008; 172: 1683–92.
- 131. Rhein V, Song X, Wiesner A, et al. Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. Proc Natl Acad Sci USA. 2009; 106: 20057–62.
- 132. Kwong LK, Uryu K, Trojanowski JQ, et al. TDP-43 proteinopathies: neurodegenerative protein misfolding diseases without amyloidosis. *Neurosignals*. 2008; 16: 41–51.
- Lehman NL. The ubiquitin proteasome system in neuropathology. *Acta Neuropathol.* 2009; 118: 329–47.
- 134. Hirsch C, Gauss R, Horn SC, et al. The ubiquitylation machinery of the

endoplasmic reticulum. *Nature*. 2009; 458: 453–60.

- Ciechanover A. Tracing the history of the ubiquitin proteolytic system: the pioneering article. *Biochem Biophys Res Commun.* 2009; 387: 1–10.
- Huang L, Chen CH. Proteasome regulators: activators and inhibitors. *Curr Med Chem.* 2009; 16: 931–9.
- 137. Schulman BA, Harper JW. Ubiquitin-like protein activation by E1 enzymes: the apex for downstream signalling pathways. *Nat Rev Mol Cell Biol.* 2009; 10: 319–31.
- Nedelsky NB, Todd PK, Taylor JP. Autophagy and the ubiquitin-proteasome system: collaborators in neuroprotection. *Biochim Biophys Acta*. 2008; 1782: 691–9.
- Kirkin V, McEwan DG, Novak I, et al. A role for ubiquitin in selective autophagy. *Mol Cell.* 2009; 34: 259–69.
- Leon S, Haguenauer-Tsapis R. Ubiquitin ligase adaptors: regulators of ubiquitylation and endocytosis of plasma membrane proteins. *Exp Cell Res.* 2009; 315: 1574–83.
- 141. **Eisele F, Wolf DH.** Degradation of misfolded protein in the cytoplasm is mediated by the ubiquitin ligase Ubr1. *FEBS Lett.* 2008; 582: 4143–6.
- Finley D. Recognition and processing of ubiquitin-protein conjugates by the proteasome. *Annu Rev Biochem.* 2009; 78: 477–513.
- 143. Ye Y, Rape M. Building ubiquitin chains: E2 enzymes at work. *Nat Rev Mol Cell Biol.* 2009; 10: 755–64.
- 144. Setsuie R, Sakurai M, Sakaguchi Y, et al. Ubiquitin dimers control the hydrolase activity of UCH-L3. *Neurochem Int.* 2009; 54: 314–21.
- 145. Su V, Lau AF. Ubiquitin-like and ubiquitinassociated domain proteins: significance in proteasomal degradation. *Cell Mol Life Sci.* 2009; 66: 2819–33.
- 146. Vashisht AA, Zumbrennen KB, Huang X, et al. Control of iron homeostasis by an iron-regulated ubiquitin ligase. Science. 2009; 326: 718–21.
- Bergink S, Jentsch S. Principles of ubiquitin and SUMO modifications in DNA repair. *Nature*. 2009: 458: 461–7.
- 148. **Stehmeier P, Muller S.** Regulation of p53 family members by the ubiquitin-like SUMO system. *DNA Repair.* 2009; 8: 491–8.
- Komander D. The emerging complexity of protein ubiquitination. *Biochem Soc Trans*. 2009; 37: 937–53.
- 150. Xu P, Duong DM, Seyfried NT, et al. Quantitative proteomics reveals the function of unconventional ubiquitin chains in

proteasomal degradation. *Cell.* 2009; 137: 133–45.

- 151. Qiao L, Zhang J. Inhibition of lysosomal functions reduces proteasomal activity. *Neurosci Lett.* 2009; 456: 15–9.
- 152. Wong ESP, Tan JMM, Lim K-L. Dynamic role of ubiquitination in the management of misfolded proteins associated with neurdegenerative diseases. In: Ovádi J, Orosz F, editors. Protein folding and misfolding: neurodegenerative diseases. Netherlands: Springer; 2009. pp. 77–95.
- 153. Tydlacka S, Wang CE, Wang X, et al. Differential activities of the ubiquitin-proteasome system in neurons versus glia may account for the preferential accumulation of misfolded proteins in neurons. J Neurosci. 2008; 28: 13285–95.
- 154. Martinet W, Agostinis P, Vanhoecke B, et al. Autophagy in disease: a doubleedged sword with therapeutic potential. *Clin Sci.* 2009; 116: 697–712.
- Galluzzi L, Vicencio JM, Kepp O, et al. To die or not to die: that is the autophagic question. Curr Mol Med. 2008; 8: 78–91.
- Lee JA. Autophagy in neurodegeneration: two sides of the same coin. *BMB Rep.* 2009; 42: 324–30.
- 157. Jaeger PA, Wyss-Coray T. All-youcan-eat: autophagy in neurodegeneration and neuroprotection. *Mol Neurodegener*. 2009; 4: 16.
- Eskelinen EL, Saftig P. Autophagy: a lysosomal degradation pathway with a central role in health and disease. *Biochim Biophys Acta*. 2009; 1793: 664–73.
- Levine B, Kroemer G. Autophagy in the pathogenesis of disease. *Cell.* 2008; 132: 27–42.
- 160. Todde V, Veenhuis M, van der Klei IJ. Autophagy: principles and significance in health and disease. *Biochim Biophys Acta*. 2009; 1792: 3–13.
- 161. Yue Z, Friedman L, Komatsu M, et al. The cellular pathways of neuronal autophagy and their implication in neurodegenerative diseases. *Biochim Biophys* Acta. 2009; 1793: 1496–507.
- He C, Klionsky DJ. Regulation mechanisms and signaling pathways of autophagy. Annu Rev Genet. 2009.
- 163. Chu CT, Plowey ED, Dagda RK, et al. Autophagy in neurite injury and neurodegeneration: in vitro and in vivo models. *Methods Enzymol.* 2009; 453: 217–49.
- 164. Chen Y, Azad MB, Gibson SB. Superoxide is the major reactive oxygen species regulating autophagy. *Cell Death Differ*. 2009; 16: 1040–52.

- 165. Puyal J, Ginet V, Vaslin A, et al. [The two faces of autophagy in the nervous system]. Med Sci. 2009; 25: 383–90.
- 166. **Tolkovsky AM.** Mitophagy. *Biochim Biophys Acta*. 2009; 1793: 1508–15.
- Rubinsztein DC. The roles of intracellular protein-degradation pathways in neurodegeneration. *Nature*. 2006; 443: 780–6.
- Cuervo AM, Stefanis L, Fredenburg R, et al. Impaired degradation of mutant alpha-synuclein by chaperone-mediated autophagy. Science. 2004; 305: 1292–5.
- Martinez-Vicente M, Talloczy Z, Kaushik S, et al. Dopamine-modified alpha-synuclein blocks chaperone-mediated autophagy. J Clin Invest. 2008; 118: 777–88.
- 170. **Spencer B, Potkar R, Trejo M, et al.** Beclin 1 gene transfer activates autophagy and ameliorates the neurodegenerative pathology in alpha-synuclein models of Parkinson's and Lewy body diseases. *J Neurosci.* 2009; 29: 13578–88.
- 171. **Pickford F, Masliah E, Britschgi M, et al.** The autophagy-related protein beclin 1 shows reduced expression in early Alzheimer disease and regulates amyloid beta accumulation in mice. *J Clin Invest.* 2008; 118: 2190–9.
- 172. Hetz C, Thielen P, Matus S, et al. XBP-1 deficiency in the nervous system protects against amyotrophic lateral sclerosis by increasing autophagy. *Genes Dev.* 2009; 23: 2294–306.
- 173. Uchiyama Y, Shibata M, Koike M, *et al.* Autophagy-physiology and pathophysiology. *Histochem Cell Biol.* 2008; 129: 407–20.
- 174. Korolchuk VI, Mansilla A, Menzies FM, et al. Autophagy inhibition compromises degradation of ubiquitin-proteasome pathway substrates. *Mol Cell.* 2009; 33: 517–27.
- 175. Cherra SJ, Chu CT. Autophagy in neuroprotection and neurodegeneration: a question of balance. *Future Neurol.* 2008; 3: 309–23.
- 176. Tooze SA, Schiavo G. Liaisons dangereuses: autophagy, neuronal survival and neurodegeneration. *Curr Opin Neurobiol.* 2008; 18: 504–15.
- Rami A. Review: autophagy in neurodegeneration: firefighter and/or incendiarist? *Neuropathol Appl Neurobiol.* 2009; 35: 449–61.
- 178. Li H, Guo M. Protein degradation in Parkinson disease revisited: it's complex. *J Clin Invest.* 2009; 119: 442–5.
- 179. Lim KL. Ubiquitin-proteasome system dysfunction in Parkinson's disease: current evidence and controversies. *Expert Rev Proteomics*. 2007; 4: 769–81.

- Pan T, Kondo S, Le W, *et al.* The role of autophagy-lysosome pathway in neurodegeneration associated with Parkinson's disease. *Brain.* 2008; 131: 1969–78.
- 181. Eliezer D. Synuclein structure and function in Parkinson's disease. In: Ovádi J, Orosz F, editors. Protein folding and misfolding: neurodegenerative diseases. Netherlands: Springer; 2009. pp. 159–74.
- 182. Schlossmacher MG, Frosch MP, Gai WP, et al. Parkin localizes to the Lewy bodies of Parkinson disease and dementia with Lewy bodies. Am J Pathol. 2002; 160: 1655–67.
- Xiong H, Wang D, Chen L, et al. Parkin, PINK1, and DJ-1 form a ubiquitin E3 ligase complex promoting unfolded protein degradation. J Clin Invest. 2009; 119: 650–60.
- 184. Schiesling C, Kieper N, Seidel K, et al. Review: familial Parkinson's disease-genetics, clinical phenotype and neuropathology in relation to the common sporadic form of the disease. Neuropathol Appl Neurobiol. 2008; 34: 255–71.
- Haugarvoll K, Rademakers R, Kachergus JM, et al. LRRK2 R1441C parkinsonism is clinically similar to sporadic Parkinson disease. *Neurology*. 2008; 70: 1456–60.
- Zimprich A, Biskup S, Leitner P, et al. Mutations in LRRK2 cause autosomaldominant parkinsonism with pleomorphic pathology. *Neuron.* 2004; 44: 601–7.
- Whaley NR, Uitti RJ, Dickson DW, et al. Clinical and pathologic features of families with LRRK2-associated Parkinson's disease. J Neural Transm Suppl. 2006: 221–9.
- Santpere G, Ferrer I. LRRK2 and neurodegeneration. Acta Neuropathol. 2009; 117: 227–46.
- 189. Oddo S. The ubiquitin-proteasome system in Alzheimer's disease. J Cell Mol Med. 2008; 12: 363–73.
- 190. **Tseng BP, Green KN, Chan JL, et al.** Abeta inhibits the proteasome and enhances amyloid and tau accumulation. *Neurobiol Aging.* 2008; 29: 1607–18.
- Liu YH, Wei W, Yin J, et al. Proteasome inhibition increases tau accumulation independent of phosphorylation. *Neurobiol Aging.* 2009; 30: 1949–61.
- 192. Zouambia M, Fischer DF, Hobo B, et al. Proteasome subunit proteins and neuropathology in tauopathies and synucleinopathies: consequences for proteomic analyses. Proteomics. 2008; 8: 1221–36.
- 193. Gil JM, Rego AC. Mechanisms of neurodegeneration in Huntington's disease. *Eur J Neurosci.* 2008; 27: 2803–20.

- 194. Iwata A, Nagashima Y, Matsumoto L, et al. Intranuclear degradation of polyglutamine aggregates by the ubiquitin-proteasome system. J Biol Chem. 2009; 284: 9796–803.
- 195. Mitra S, Tsvetkov AS, Finkbeiner S. Single neuron ubiquitin-proteasome dynamics accompanying inclusion body formation in Huntington disease. J Biol Chem. 2009; 284: 4398–403.
- 196. Wang J, Wang CE, Orr A, et al. Impaired ubiquitin-proteasome system activity in the synapses of Huntington's disease mice. J Cell Biol. 2008; 180: 1177–89.
- 197. Davies JE, Sarkar S, Rubinsztein DC. The ubiquitin proteasome system in Huntington's disease and the spinocerebellar ataxias. *BMC Biochem.* 2007; 8: S2.
- Olzmann JA, Li L, Chin LS. Aggresome formation and neurodegenerative diseases: therapeutic implications. *Curr Med Chem.* 2008; 15: 47–60.
- 199. Chu Y, Dodiya H, Aebischer P, et al. Alterations in lysosomal and proteasomal markers in Parkinson's disease: relationship to alpha-synuclein inclusions. *Neurobiol Dis.* 2009; 35: 385–98.
- 200. Mishra A, Godavarthi SK, Maheshwari M, et al. The ubiquitin ligase E6-AP is induced and recruited to aggresomes in response to proteasome inhibition and may be involved in the ubiquitination of Hsp70-bound misfolded proteins. J Biol Chem. 2009; 284: 10537–45.
- Gardiner J, Barton D, Overall R, Marc J. Neurotrophic support and oxidative stress: converging effects in the normal and diseased nervous system. *Neuroscientist*. 2009; 15: 47–61.
- Sultana R, Butterfield DA. Oxidatively modified, mitochondria-relevant brain proteins in subjects with Alzheimer disease and mild cognitive impairment. J Bioenerg Biomembr. 2009; 41: 441–6.
- Halliwell B. Oxidative stress and neurodegeneration: where are we now? J Neurochem. 2006; 97: 1634–58.
- 204. Nunomura A, Hofer T, Moreira PI, et al. RNA oxidation in Alzheimer disease and related neurodegenerative disorders. Acta Neuropathol. 2009; 118: 151–66.
- Martinez A, Portero-Otin M, Pamplona R, et al. Protein targets of oxidative damage in human neurodegenerative diseases with abnormal protein aggregates. Brain Pathol. 2009. DOI: 10.1111/j.1750-3639.2009. 00326.x
- Nakamura T, Lipton SA. Cell death: protein misfolding and neurodegenerative diseases. *Apoptosis*. 2009; 14: 455–68.

- 207. Bond CE, Greenfield SA. Multiple cascade effects of oxidative stress on astroglia. *Glia.* 2007; 55: 1348–61.
- Ballatori N, Krance SM, Notenboom S, et al. Glutathione dysregulation and the etiology and progression of human diseases. *Biol Chem.* 2009; 390: 191–214.
- Castellani RJ, Nunomura A, Rolston RK, et al. Sublethal RNA oxidation as a mechanism for neurodegenerative disease. Int J Mol Sci. 2008; 9: 789–806.
- Migliore L, Coppede F. Environmentalinduced oxidative stress in neurodegenerative disorders and aging. *Mutat Res.* 2009; 674: 73–84.
- Browne SE, Beal MF. Oxidative damage in Huntington's disease pathogenesis. Antioxid Redox Signal. 2006; 8: 2061–73.
- Wang J, Markesbery WR, Lovell MA. Increased oxidative damage in nuclear and mitochondrial DNA in mild cognitive impairment. J Neurochem. 2006; 96: 825–32.
- Pratico D, Sung S. Lipid peroxidation and oxidative imbalance: early functional events in Alzheimer's disease. J Alzheimers Dis. 2004; 6: 171–5.
- Mancuso M, Coppede F, Migliore L, et al. Mitochondrial dysfunction, oxidative stress and neurodegeneration. J Alzheimers Dis. 2006; 10: 59–73.
- Sayre LM, Perry G, Smith MA. Oxidative stress and neurotoxicity. *Chem Res Toxicol.* 2008; 21: 172–88.
- Ward RJ, Crichton RR. Iron and its role in neurodegenerative diseases. In: Sigel A, Sigel H, Sigel RKO, editors. Metal ions in life sciences. Vol. 1. New York: Wiley & Sons: 2006. pp. 227–79.
- Mancuso M, Davidzon G, Kurlan RM, et al. Hereditary ferritinopathy: a novel mutation, its cellular pathology, and pathogenetic insights. J Neuropathol Exp Neurol. 2005; 64: 280–94.
- Sparaco M, Gaeta LM, Santorelli FM, et al. Friedreich's ataxia: oxidative stress and cytoskeletal abnormalities. J Neurol Sci. 2009; 287: 111–8.
- Altamura S, Muckenthaler MU. Iron toxicity in diseases of aging: Alzheimer's disease, Parkinson's disease and atherosclerosis. J Alzheimers Dis. 2009; 16: 879–95.
- 220. Arreguin S, Nelson P, Padway S, et al. Dopamine complexes of iron in the etiology and pathogenesis of Parkinson's disease. J Inorg Biochem. 2009; 103: 87–93.
- 221. Sultana R, Perluigi M, Butterfield DA. Oxidatively modified proteins in Alzheimer's disease (AD), mild cognitive impairment and animal models of AD: role of Abeta in pathogenesis. Acta

Neuropathol. 2009; in press: doi 10.1007/s00401-009-0517-0.

- 222. Sonnen JA, Breitner JC, Lovell MA, et al. Free radical-mediated damage to brain in Alzheimer's disease and its transgenic mouse models. *Free Radic Biol Med.* 2008; 45: 219–30.
- Zheng L, Kagedal K, Dehvari N, et al. Oxidative stress induces macroautophagy of amyloid beta-protein and ensuing apoptosis. *Free Radic Biol Med.* 2009; 46: 422–9.
- 224. Reddy PH, Beal MF. Amyloid beta, mitochondrial dysfunction and synaptic damage: implications for cognitive decline in aging and Alzheimer's disease. *Trends Mol Med.* 2008; 14: 45–53.
- 225. Yan SD, Bierhaus A, Nawroth PP, et al. RAGE and Alzheimer's disease: a progression factor for amyloid-beta-induced cellular perturbation? J Alzheimers Dis. 2009; 16: 833–43.
- 226. Origlia N, Righi M, Capsoni S, et al. Receptor for advanced glycation end product-dependent activation of p38 mitogenactivated protein kinase contributes to amyloid-beta-mediated cortical synaptic dysfunction. J Neurosci. 2008; 28: 3521–30.
- 227. Wang MY, Ross-Cisneros FN, Aggarwal D, et al. Receptor for advanced glycation end products is upregulated in optic neuropathy of Alzheimer's disease. Acta Neuropathol. 2009; 118: 381–9.
- Takuma K, Fang F, Zhang W, et al. RAGEmediated signaling contributes to intraneuronal transport of amyloid-beta and neuronal dysfunction. Proc Natl Acad Sci USA. 2009; 106: 20021–6.
- Pedersen PL. Mitochondrial matters of the brain: amyloid formation and Alzheimer's disease introduction. J Bioenerg Biomembr. 2009; 41: 403–5.
- Reed TT, Pierce WM Jr, Turner DM, et al. Proteomic identification of nitrated brain proteins in early Alzheimer's disease inferior parietal lobule. J Cell Mol Med. 2008; 13: 2019–29.
- Sun KH, de Pablo Y, Vincent F, et al. Deregulated Cdk5 promotes oxidative stress and mitochondrial dysfunction. J Neurochem. 2008; 107: 265–78.
- Cho DH, Nakamura T, Fang J, et al. S-nitrosylation of Drp1 mediates betaamyloid-related mitochondrial fission and neuronal injury. *Science*. 2009; 324: 102–5.
- Zhao B. Natural antioxidants protect neurons in Alzheimer's disease and Parkinson's disease. *Neurochem Res.* 2009; 34: 630–8.

- 234. Miller RL, James-Kracke M, Sun GY, et al. Oxidative and inflammatory pathways in Parkinson's disease. *Neurochem Res.* 2009; 34: 55–65.
- Jenner P. Oxidative stress and Parkinson's disease. *Handb Clin Neurol.* 2007; 83: 507–20.
- 236. **Tsang AH, Chung KK.** Oxidative and nitrosative stress in Parkinson's disease. *Biochim Biophys Acta.* 2008; 1792: 643–50.
- 237. Olivares D, Huang X, Branden L, et al. Physiological and pathological role of alpha-synuclein in Parkinson's disease through iron mediated oxidative stress; the role of a putative iron-responsive element. Int J Mol Sci. 2009; 10: 1226–60.
- 238. Tribl F, Asan E, Arzberger T, et al. Identification of L-ferritin in neuromelanin granules of the human substantia nigra: a targeted proteomics approach. *Mol Cell Proteomics.* 2009; 8: 1832–8.
- 239. Martin HL, Teismann P. Glutathione–a review on its role and significance in Parkinson's disease. *FASEB J.* 2009; 23: 3263–72.
- Shaikh S, Nicholson LF. Advanced glycation end products induce in vitro crosslinking of alpha-synuclein and accelerate the process of intracellular inclusion body formation. *J Neurosci Res.* 2008; 86: 2071–82.
- 241. **Power JH, Blumbergs PC.** Cellular glutathione peroxidase in human brain: cellular distribution, and its potential role in the degradation of Lewy bodies in Parkinson's disease and dementia with Lewy bodies. *Acta Neuropathol.* 2009; 117: 63–73.
- 242. Rochet J-C, Liu F. Inhibition of alphasynuclein aggregation by antioxidants and chaperones in Parkinson's disease. In: Ovádi J, Orosz F, editors. Protein folding and misfolding: neurodegenerative diseases. Netherlands: Springer; 2009. pp. 175–206.
- Przedborski S. Moving Along (Newsletter of the *Movement Disorder Soc.*) 2009; 13: 1.
- 244. Levy OA, Malagelada C, Greene LA. Cell death pathways in Parkinson's disease: proximal triggers, distal effectors, and final steps. *Apoptosis.* 2009; 14: 478–500.
- Nagley P, Higgins GC, Atkin JD, et al. Multifaceted deaths orchestrated by mitochondria in neurones. BBA – Molec Basis Dis. 2010; 1802: 167–85.
- 246. Sorolla MA, Reverter-Branchat G, Tamarit J, et al. Proteomic and oxidative stress analysis in human brain samples of Huntington disease. Free Radic Biol Med. 2008; 45: 667–78.

- 247. Martinez A, Dalfo E, Muntane G, et al. Glycolitic enzymes are targets of oxidation in aged human frontal cortex and oxidative damage of these proteins is increased in progressive supranuclear palsy. J Neural Transm. 2008; 115: 59–66.
- 248. Santpere G, Ferrer I. Delineation of early changes in cases with progressive supranuclear palsy-like pathology. Astrocytes in striatum are primary targets of tau phosphorylation and GFAP oxidation. *Brain Pathol.* 2009; 19: 177–87.
- 249. Pasinelli P, Belford ME, Lennon N, et al. Amyotrophic lateral sclerosis-associated SOD1 mutant proteins bind and aggregate with Bcl-2 in spinal cord mitochondria. *Neuron*. 2004; 43: 19–30.
- Magrane J, Hervias I, Henning MS, et al. Mutant SOD1 in neuronal mitochondria causes toxicity and mitochondrial dynamics abnormalities. *Hum Mol Genet.* 2009; 18: 4552–64.
- Barber SC, Mead RJ, Shaw PJ. Oxidative stress in ALS: a mechanism of neurodegeneration and a therapeutic target. *Biochim Biophys Acta.* 2006; 1762: 1051–67.
- 252. **Baumer D, Parkinson N, Talbot K.** TARDBP in amyotrophic lateral sclerosis: identification of a novel variant but absence of copy number variation. *Journal* of Neurology Neurosurgery and Psychiatry. 2009; 80: 1283–5.
- Yang JL, Weissman L, Bohr VA, et al. Mitochondrial DNA damage and repair in neurodegenerative disorders. DNA Repair. 2008; 7: 1110–20.
- 254. Gupta S, Kass GE, Szegezdi E, et al. The mitochondrial death pathway: a promising therapeutic target in diseases. J Cell Mol Med. 2009; 13: 1004–33.
- Perkins G, Bossy-Wetzel E, Ellisman MH. New insights into mitochondrial structure during cell death. *Exp Neurol.* 2009; 218: 183–92.
- Dubinsky JM. Heterogeneity of nervous system mitochondria: location, location, location! *Exp Neurol.* 2009; 218: 293–307.
- Reddy PH. Mitochondrial medicine for aging and neurodegenerative diseases. *Neuromolecular Med.* 2008; 10: 291–315.
- Chen H, Chan DC. Mitochondrial dynamics-fusion, fission, movement, and mitophagy-in neurodegenerative diseases. *Hum Mol Genet.* 2009; 18: R169-76.
- 259. Celsi F, Pizzo P, Brini M, et al. Mitochondria, calcium and cell death: a deadly triad in neurodegeneration. *Biochim Biophys Acta*. 2009; 1787: 335–44.

- 260. **Schapira AH.** Mitochondria in the aetiology and pathogenesis of Parkinson's disease. *Lancet Neurol.* 2008; 7: 97–109.
- 261. Banoei MM, Houshmand M, Panahi MS, et al. Huntington's disease and mitochondrial DNA deletions: event or regular mechanism for mutant huntingtin protein and CAG repeats expansion?! Cell Mol Neurobiol. 2007; 27: 867–75.
- Quintanilla RA, Johnson GV. Role of mitochondrial dysfunction in the pathogenesis of Huntington's disease. *Brain Res Bull.* 2009; 80: 242–7.
- Distelmaier F, Koopman WJ, van den Heuvel LP, et al. Mitochondrial complex I deficiency: from organelle dysfunction to clinical disease. Brain. 2009; 132: 833–42.
- 264. **Armstrong JS.** Mitochondrial membrane permeabilization: the sine qua non for cell death. *Bioessays.* 2006; 28: 253–60.
- 265. Jeyaraju DV, Cisbani G, Pellegrini L. Calcium regulation of mitochondria motility and morphology. *Biochim Biophys Acta*. 2009; 1787: 1363–73.
- Nicholls DG. Mitochondrial calcium function and dysfunction in the central nervous system. *Biochim Biophys Acta*. 2009; 1787: 1416–24.
- Bueler H. Impaired mitochondrial dynamics and function in the pathogenesis of Parkinson's disease. *Exp Neurol.* 2009; 218: 235–46.
- Martin LJ. DNA damage and repair: relevance to mechanisms of neurodegeneration. J Neuropathol Exp Neurol. 2008; 67: 377–87.
- Katyal S, McKinnon PJ. DNA repair deficiency and neurodegeneration. *Cell Cycle*. 2007; 6: 2360–5.
- Coppede F, Migliore L. DNA damage and repair in Alzheimer's disease. Curr Alzheimer Res. 2009; 6: 36–47.
- 271. Lu B. Mitochondrial dynamics and neurodegeneration. *Curr Neurol Neurosci Rep.* 2009; 9: 212–9.
- 272. **Cogswell JP, Ward J, Taylor IA**, *et al.* Identification of miRNA changes in Alzheimer's disease brain and CSF yields putative biomarkers and insights into disease pathways. *J Alzheimers Dis.* 2008; 14: 27–41.
- Saba R, Goodman CD, Huzarewich RL, et al. A miRNA signature of prion induced neurodegeneration. PLoS One. 2008; 3: e3652.
- Hebert SS, De Strooper B. Alterations of the microRNA network cause neurodegenerative disease. *Trends Neurosci.* 2009; 32: 199–206.

- Nelson PT, Wang WX, Rajeev BW. MicroRNAs (miRNAs) in neurodegenerative diseases. *Brain Pathol.* 2008; 18: 130–8.
- Bushati N, Cohen SM. MicroRNAs in neurodegeneration. *Curr Opin Neurobiol.* 2008; 18: 292–6.
- Swerdlow RH, Khan SM. The Alzheimer's disease mitochondrial cascade hypothesis: an update. *Exp Neurol.* 2009; 218: 308–15.
- Mancuso M, Calsolaro V, Orsucci D, et al. Is there a primary role of the mitochondrial genome in Alzheimer's disease? J Bioenera Biomembr. 2009: 41: 411–6.
- Wang X, Su B, Zheng L, *et al.* The role of abnormal mitochondrial dynamics in the pathogenesis of Alzheimer's disease. *J Neurochem.* 2009; 109: 153–9.
- Hauptmann S, Scherping I, Drose S, et al. Mitochondrial dysfunction: an early event in Alzheimer pathology accumulates with age in AD transgenic mice. *Neurobiol Aging.* 2009; 30: 1574–86.
- Melov S, Adlard PA, Morten K, et al. Mitochondrial oxidative stress causes hyperphosphorylation of tau. PLoS One. 2007; 2: e536.
- De Vos KJ, Grierson AJ, Ackerley S, et al. Role of axonal transport in neurodegenerative diseases. Annu Rev Neurosci. 2008; 31: 151–73.
- 283. Quintanilla RA, Matthews-Roberson TA, Dolan PJ, et al. Caspase-cleaved tau expression results in mitochondrial dysfunction in cortical neurons. Implications for the pathogenesis of Alzheimer disease. J Biol Chem. 2009; 284: 18754–66.
- 284. Terni B, Boada J, Portero-Otin M, et al. Mitochondrial ATP-Synthase in the entorhinal cortex is a target of oxidative stress at stages I/II of Alzheimer's disease pathology. Brain Pathol. 2009; DOI: 10.1111/j.1750-3639.2009.00266.x.
- 285. **Shapira IL.** The Nun study. *J Am Dent Assoc.* 2008; 139: 130–1.
- Onyango IG. Mitochondrial dysfunction and oxidative stress in Parkinson's disease. *Neurochem Res.* 2008; 33: 589–97.
- Vila M, Ramonet D, Perier C. Mitochondrial alterations in Parkinson's disease: new clues. J Neurochem. 2008; 107: 317–28.
- Yao Z, Wood NW. Cell death pathways in Parkinson's disease: role of mitochondria. *Antioxid Redox Signal.* 2009; 11: 2135–49.
- 289. Davison EJ, Pennington K, Hung CC, et al. Proteomic analysis of increased Parkin expression and its interactants provides evidence for a role in modulation of

mitochondrial function. *Proteomics*. 2009; 9: 4284–97.

- Van Laar VS, Berman SB. Mitochondrial dynamics in Parkinson's disease. *Exp Neurol.* 2009; 218: 247–56.
- Berger AK, Cortese GP, Amodeo KD, et al. Parkin selectively alters the intrinsic threshold for mitochondrial cytochrome c release. Hum Mol Genet. 2009; 18: 4317–28.
- 292. Burns MP, Zhang L, Rebeck GW, et al. Parkin promotes intracellular Abeta1-42 clearance. Hum Mol Genet. 2009; 18: 3206–16.
- 293. Park J, Lee G, Chung J. The PINK1-Parkin pathway is involved in the regulation of mitochondrial remodeling process. *Biochem Biophys Res Commun.* 2009; 378: 518–23.
- 294. Kao SY. Regulation of DNA repair by parkin. *Biochem Biophys Res Commun.* 2009: 382: 321–5.
- 295. Yang Y, Lu B. Mitochondrial morphogenesis, distribution, and Parkinson disease: insights from PINK1. J Neuropathol Exp Neurol. 2009; 68: 953–63.
- 296. Gupta A, Dawson VL, Dawson TM. What causes cell death in Parkinson's disease? Ann Neurol. 2008; 64: S3–15.
- 297. Freestone PS, Chung KK, Guatteo E, et al. Acute action of rotenone on nigral dopaminergic neurons – involvement of reactive oxygen species and disruption of Ca homeostasis. Eur J Neurosci. 2009; 30: 1849–59.
- Sherer TB, Betarbet R, Testa CM, et al. Mechanism of toxicity in rotenone models of Parkinson's disease. J Neurosci. 2003; 23: 10756–64.
- 299. Drechsel DA, Patel M. Role of reactive oxygen species in the neurotoxicity of environmental agents implicated in Parkinson's disease. *Free Radic Biol Med.* 2008; 44: 1873–86.
- Alvarez-Fischer D, Guerreiro S, Hunot S, et al. Modelling Parkinson-like neurodegeneration via osmotic minipump delivery of MPTP and probenecid. J Neurochem. 2008; 107: 701–11.
- Halliday G, Herrero MT, Murphy K, et al. No Lewy pathology in monkeys with over 10 years of severe MPTP Parkinsonism. *Mov Disord*. 2009; 24: 1519–23.
- 302. Choi WS, Kruse SE, Palmiter RD, et al. Mitochondrial complex I inhibition is not required for dopaminergic neuron death induced by rotenone, MPP+, or paraquat. *Proc Natl Acad Sci USA*. 2008; 105: 15136–41.

- 303. Borland MK, Trimmer PA, Rubinstein JD, et al. Chronic, low-dose rotenone reproduces Lewy neurites found in early stages of Parkinson's disease, reduces mitochondrial movement and slowly kills differentiated SH-SY5Y neural cells. Mol Neurodegener. 2008; 3: 21.
- Lin MT, Beal MF. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature*. 2006; 443: 787–95.
- Lim D, Fedrizzi L, Tartari M, et al. Calcium homeostasis and mitochondrial dysfunction in striatal neurons of Huntington disease. J Biol Chem. 2008; 283: 5780–9.
- Estrada Sanchez AM, Mejia-Toiber J, Massieu L. Excitotoxic neuronal death and the pathogenesis of Huntington's disease. *Arch Med Res.* 2008; 39: 265–76.
- 307. Chang WH, Tien CL, Chen TJ, et al. Decreased protein synthesis of Hsp27 associated with cellular toxicity in a cell model of Machado-Joseph disease. *Neuroscience Letters*. 2009; 454: 152–6.
- 308. Yu YC, Kuo CL, Cheng WL, et al. Decreased antioxidant enzyme activity and increased mitochondrial DNA damage in cellular models of Machado-Joseph disease. J Neurosci Res. 2009; 87: 1884–91.
- Fan J, Hu Z, Zeng L, et al. Golgi apparatus and neurodegenerative diseases. Int J Dev Neurosci. 2008; 26: 523–34.
- Gonatas NK, Stieber A, Gonatas JO. Fragmentation of the Golgi apparatus in neurodegenerative diseases and cell death. *J Neurol Sci.* 2006: 246: 21–30.
- 311. Fujita Y, Ohama E, Takatama M, et al. Fragmentation of Golgi apparatus of nigral neurons with alpha-synuclein-positive inclusions in patients with Parkinson's disease. Acta Neuropathol. 2006; 112: 261–5.
- Vickers JC, King AE, Woodhouse A, et al. Axonopathy and cytoskeletal disruption in degenerative diseases of the central nervous system. Brain Res Bull. 2009; 80: 217–23.
- Morfini GA, Burns M, Binder LI, et al. Axonal transport defects in neurodegenerative diseases. J Neurosci. 2009; 29: 12776–86.
- 314. Prins KW, Humston JL, Mehta A, et al. Dystrophin is a microtubuleassociated protein. J Cell Biol. 2009; 186: 363–9.
- 315. Yates DM, Manser C, De Vos KJ, et al. Neurofilament subunit (NFL) head domain phosphorylation regulates axonal transport of neurofilaments. Eur J Cell Biol. 2009; 88: 193–202.

- Goldstein AY, Wang X, Schwarz TL. Axonal transport and the delivery of presynaptic components. *Curr Opin Neurobiol.* 2008; 18: 495–503.
- Barry DM, Millecamps S, Julien JP, et al. New movements in neurofilament transport, turnover and disease. *Exp Cell Res.* 2007; 313: 2110–20.
- 318. Yuan A, Rao MV, Kumar A, et al. Neurofilament transport in vivo minimally requires hetero-oligomer formation. *J Neurosci.* 2003; 23: 9452–8.
- Hirokawa N, Noda Y, Tanaka Y, et al. Kinesin superfamily motor proteins and intracellular transport. Nat Rev Mol Cell Biol. 2009; 10: 682–96.
- 320. Roy S, Zhang B, Lee VM, et al. Axonal transport defects: a common theme in neurodegenerative diseases. Acta Neuropathol. 2005; 109: 5–13.
- 321. Schindowski K, Belarbi K, Buee L. Neurotrophic factors in Alzheimer's disease: role of axonal transport. *Genes Brain Behav.* 2008; 7: 43–56.
- 322. Stokin GB, Goldstein LS. Axonal transport and Alzheimer's disease. Annu Rev Biochem. 2006; 75: 607–27.
- 323. Muresan V, Varvel NH, Lamb BT, et al. The cleavage products of amyloid-beta precursor protein are sorted to distinct carrier vesicles that are independently transported within neurites. J Neurosci. 2009; 29: 3565–78.
- 324. Pigino G, Morfini G, Atagi Y, et al. Disruption of fast axonal transport is a pathogenic mechanism for intraneuronal amyloid beta. Proc Natl Acad Sci USA. 2009; 106: 5907–12.
- 325. Shah SB, Nolan R, Davis E, et al. Examination of potential mechanisms of amyloid-induced defects in neuronal transport. Neurobiol Dis. 2009; 36: 11–25.
- 326. Stoothoff W, Jones PB, Spires-Jones TL, et al. Differential effect of three-repeat and four-repeat tau on mitochondrial axonal transport. J Neurochem. 2009; 111: 417–27.
- 327. Falzone TL, Stokin GB, Lillo C, et al. Axonal stress kinase activation and tau misbehavior induced by kinesin-1 transport defects. J Neurosci, 2009: 29: 5758–67.
- 328. Yuan A, Kumar A, Peterhoff C, et al. Axonal transport rates in vivo are unaffected by tau deletion or overexpression in mice. J Neurosci. 2008; 28: 1682–7.
- 329. Li W, Hoffman PN, Stirling W, et al. Axonal transport of human alpha-synuclein slows with aging but is not affected by familial Parkinson's disease-linked mutations. J Neurochem. 2004; 88: 401–10.

- 330. Kramer ML, Schulz-Schaeffer WJ. Presynaptic alpha-synuclein aggregates, not Lewy bodies, cause neurodegeneration in dementia with Lewy bodies. *J Neurosci.* 2007; 27: 1405–10.
- Rothstein JD. Current hypotheses for the underlying biology of amyotrophic lateral sclerosis. *Ann Neurol.* 2009; 65: S3–9.
- Strom AL, Gal J, Shi P, et al. Retrograde axonal transport and motor neuron disease. J Neurochem. 2008; 106: 495–505.
- 333. Her LS, Goldstein LS. Enhanced sensitivity of striatal neurons to axonal transport defects induced by mutant huntingtin. *J Neurosci.* 2008; 28: 13662–72.
- 334. Bossers K, Meerhoff G, Balesar R, et al. Analysis of gene expression in Parkinson's disease: possible involvement of neurotrophic support and axon guidance in dopaminergic cell death. Brain Pathol. 2009: 19: 91–107.
- 335. Peng S, Garzon DJ, Marchese M, et al. Decreased brain-derived neurotrophic factor depends on amyloid aggregation state in transgenic mouse models of Alzheimer's disease. J Neurosci. 2009; 29: 9321–9.
- 336. Tapia-Arancibia L, Aliaga E, Silhol M, et al. New insights into brain BDNF function in normal aging and Alzheimer disease. Brain Res Rev. 2008; 59: 201–20.
- Bulbarelli A, Lonati E, Cazzaniga E, et al. TrkA pathway activation induced by amyloid-beta (Abeta). *Mol Cell Neurosci.* 2009; 40: 365–73.
- 338. Arevalo MA, Roldan PM, Chacon PJ, et al. Amyloid beta serves as an NGF-like neurotrophic factor or acts as a NGF antagonist depending on its concentration. Journal of Neurochemistry. 2009; 111: 1425–33.
- 339. Sotthibundhu A, Sykes AM, Fox B, et al. Beta-amyloid(1–42) induces neuronal death through the p75 neurotrophin receptor. J Neurosci. 2008; 28: 3941–6.
- 340. Schulte-Herbruggen O, Jockers-Scherubl MC, Hellweg R. Neurotrophins: from pathophysiology to treatment in Alzheimer's disease. *Curr Alzheimer Res.* 2008; 5: 38–44.
- 341. Fombonne J, Rabizadeh S, Banwait S, et al. Selective vulnerability in Alzheimer's disease: amyloid precursor protein and p75(NTR) interaction. Ann Neurol. 2009; 65: 294–303.
- 342. McAlpine FE, Lee JK, Harms AS, et al. Inhibition of soluble TNF signaling in a mouse model of Alzheimer's disease prevents pre-plaque amyloid-associated neuropathology. *Neurobiol Dis.* 2009; 34: 163–77.

- 343. Nikolaev A, McLaughlin T, O'Leary DD, et al. APP binds DR6 to trigger axon pruning and neuron death via distinct caspases. *Nature*. 2009; 457: 981–9.
- 344. Bruno MA, Leon WC, Fragoso G, et al. Amyloid beta-induced nerve growth factor dysmetabolism in Alzheimer disease. J Neuropathol Exp Neurol. 2009; 68: 857–69.
- 345. Nagahara AH, Merrill DA, Coppola G, et al. Neuroprotective effects of brainderived neurotrophic factor in rodent and primate models of Alzheimer's disease. Nat Med. 2009; 15: 331–7.
- Zuccato C, Cattaneo E. Brain-derived neurotrophic factor in neurodegenerative diseases. *Nat Rev Neurol.* 2009; 5: 311–22.
- 347. Zuccato C, Marullo M, Conforti P, et al. Systematic assessment of BDNF and its receptor levels in human cortices affected by Huntington's disease. Brain Pathol. 2008; 18: 225–38.
- 348. Stolp HB, Dziegielewska KM. Review: role of developmental inflammation and blood-brain barrier dysfunction in neurodevelopmental and neurodegenerative diseases. *Neuropathol Appl Neurobiol.* 2009; 35: 132–46.
- 349. **Kell DB.** Iron behaving badly: inappropriate iron chelation as a major contributor to the aetiology of vascular and other progressive inflammatory and degenerative diseases. *BMC Med Genomics.* 2009; 2: 2.
- Galimberti D, Fenoglio C, Scarpini E. Inflammation in neurodegenerative disorders: friend or foe? *Curr Aging Sci.* 2008; 1: 30–41.
- Piehl F, Olsson T. Inflammation and susceptibility to neurodegeneration: the use of unbiased genetics to decipher critical regulatory pathways. *Neuroscience*. 2009; 158: 1143–50.
- Graber JJ, Dhib-Jalbut S. Protective autoimmunity in the nervous system. *Pharmacol Ther.* 2009; 121: 147–59.
- 353. Salminen A, Ojala J, Kauppinen A, et al. Inflammation in Alzheimer's disease: amyloid-B oligomers trigger innate immunity defence via pattern recognition receptors. *Progr Neurobiol.* 2009; 87: 181–94.
- Lucin KM, Wyss-Coray T. Immune activation in brain aging and neurodegeneration: too much or too little? *Neuron.* 2009; 64: 110–22.
- 355. Sawada M, Sawada H, Nagatsu T. Effects of aging on neuroprotective and neurotoxic properties of microglia in neurodegenerative diseases. *Neurodegener Dis.* 2008; 5: 254–6.

- 356. Neumann H, Kotter MR, Franklin RJ. Debris clearance by microglia: an essential link between degeneration and regeneration. *Brain*. 2009; 132: 288–95.
- Lee M, Schwab C, Yu S, et al. Astrocytes produce the antiinflammatory and neuroprotective agent hydrogen sulfide. *Neurobiol Aging.* 2009; 30: 1523–34.
- Sondag CM, Dhawan G, Combs CK. Beta amyloid oligomers and fibrils stimulate differential activation of primary microglia. J Neuroinflammation. 2009; 6: 1–13.
- 359. Reale M, Iarlori C, Feliciani C, et al. Peripheral chemokine receptors, their ligands, cytokines and Alzheimer's disease. J Alzheimers Dis. 2008; 14: 147–59.
- Maccioni RB, Rojo LE, Fernandez JA, et al. The role of neuroimmunomodulation in Alzheimer's disease. Ann N Y Acad Sci. 2009; 1153: 240–6.
- Holmes C, Cunningham C, Zotova E, et al. Systemic inflammation and disease progression in Alzheimer disease. *Neurology*. 2009; 73: 768–74.
- 362. Jaeger LB, Dohgu S, Sultana R, et al. Lipopolysaccharide alters the blood-brain barrier transport of amyloid beta protein: a mechanism for inflammation in the progression of Alzheimer's disease. Brain Behavior and Immunity. 2009; 23: 507–17.
- 363. Vukic V, Callaghan D, Walker D, et al. Expression of inflammatory genes induced by beta-amyloid peptides in human brain endothelial cells and in Alzheimer's brain is mediated by the JNK-AP1 signaling pathway. *Neurobiol Dis.* 2009; 34: 95–106.
- 364. Di Bona D, Candore G, Franceschi C, et al. Systematic review by meta-analyses on the possible role of TNF-alpha polymorphisms in association with Alzheimer's disease. Brain Res Rev. 2009; 61: 60–8.
- 365. McCoy MK, Tansey MG. TNF signaling inhibition in the CNS: implications for normal brain function and neurodegenerative disease. J Neuroinflammation. 2008; 5: 45.
- 366. Foster JK, Verdile G, Bates KA, et al. Immunization in Alzheimer's disease: naive hope or realistic clinical potential? *Mol Psychiatry*. 2009; 14: 239–51.
- 367. Holmes C, Boche D, Wilkinson D, et al. Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. Lancet. 2008; 372: 216–23.
- 368. Deane R, Bell RD, Sagare A, et al. Clearance of amyloid-beta peptide across the blood-brain barrier: implication for therapies in Alzheimer's disease. CNS Neurol Disord Drug Targets. 2009; 8: 16–30.

- 369. Streit WJ, Braak H, Xue QS, et al. Dystrophic (senescent) rather than activated microglial cells are associated with tau pathology and likely precede neurodegeneration in Alzheimer's disease. Acta Neuropathol. 2009; 118: 475–85.
- 370. Chakrabarty P, Jansen-West K, Beccard A, et al. Massive gliosis induced by interleukin-6 suppresses A{beta} deposition in vivo: evidence against inflammation as a driving force for amyloid deposition. FASEB J. 2009; 10.1096/fj.09-141754.
- Hirsch EC, Hunot S. Neuroinflammation in Parkinson's disease: a target for neuroprotection? *Lancet Neurol.* 2009; 8: 382–97.
- 372. Gao HM, Kotzbauer PT, Uryu K, et al. Neuroinflammation and oxidation/nitration of alpha-synuclein linked to dopaminergic neurodegeneration. J Neurosci. 2008; 28: 7687–98.
- 373. Zecca L, Wilms H, Geick S, et al. Human neuromelanin induces neuroinflammation and neurodegeneration in the rat substantia nigra: implications for Parkinson's disease. Acta Neuropathol. 2008; 116: 47–55.
- 374. Brochard V, Combadiere B, Prigent A, et al. Infiltration of CD4+ lymphocytes into the brain contributes to neurodegeneration in a mouse model of Parkinson disease. J Clin Invest. 2009: 119: 182–92.
- 375. Hunter RL, Dragicevic N, Seifert K, et al. Inflammation induces mitochondrial dysfunction and dopaminergic neurodegeneration in the nigrostriatal system. J Neurochem. 2007; 100: 1375–86.
- 376. Kim YS, Joh TH. Microglia, major player in the brain inflammation: their roles in the pathogenesis of Parkinson's disease. *Exp Mol Med.* 2006; 38: 333–47.
- 377. Pieper HC, Evert BO, Kaut O, et al. Different methylation of the TNF-alpha promoter in cortex and substantia nigra: implications for selective neuronal vulnerability. *Neurobiol Dis.* 2008; 32: 521–7.
- Pavese N, Gerhard A, Tai YF, et al. Microglial activation correlates with severity in Huntington disease: a clinical and PET study. *Neurology*. 2006; 66: 1638–43.
- 379. Tai YF, Pavese N, Gerhard A, et al. Microglial activation in presymptomatic Huntington's disease gene carriers. Brain. 2007; 130: 1759–66.
- Mantovani S, Garbelli S, Pasini A, et al. Immune system alterations in sporadic amyotrophic lateral sclerosis patients suggest an ongoing neuroinflammatory process. J Neuroimmunol. 2009; 210: 73–9.
- 381. Schwab C, Klegeris A, McGeer PL. Inflammation in transgenic mouse models

of neurodegenerative disorders. *Biochim Biophys Acta*. 2009; DOI: 10.1016/j.bbadis. 2009.10.013.

- 382. Fendrick SE, Xue QS, Streit WJ. Formation of multinucleated giant cells and microglial degeneration in rats expressing a mutant Cu/Zn superoxide dismutase gene. J Neuroinflammation. 2007; 4: 9.
- 383. Simmons DA, Casale M, Alcon B, et al. Ferritin accumulation in dystrophic microglia is an early event in the development of Huntington's disease. *Glia.* 2007; 55: 1074–84.
- 384. Rogers J, Mastroeni D, Leonard B, et al. Neuroinflammation in Alzheimer's disease and Parkinson's disease: are microglia pathogenic in either disorder? Int Rev Neurobiol. 2007; 82: 235–46.
- 385. Kroemer G, Galluzzi L, Vandenabeele P, et al. Classification of cell death: recommendations of the nomenclature committee on cell death 2009. *Cell Death Differ*. 2009; 16: 3–11.
- Bredesen DE. Programmed cell death mechanisms in neurological disease. Curr Mol Med. 2008; 8: 173–86.
- Gorman AM. Neuronal cell death in neurodegenerative diseases: recurring themes around protein handling. *J Cell Mol Med.* 2008; 12: 2263–80.
- 388. Higgins GC, Beart PM, Nagley P. Oxidative stress triggers neuronal caspase-independent death: endonuclease G involvement in programmed cell deathtype III. Cell Mol Life Sci. 2009; 66: 2773–87.
- Henriquez M, Armisen R, Stutzin A, et al. Cell death by necrosis, a regulated way to go. *Curr Mol Med.* 2008; 8: 187–206.
- 390. Bulat N, Widmann C. Caspase substrates and neurodegenerative diseases. Brain Res Bull. 2009; 80: 251–67.
- 391. Kroemer G, Levine B. Autophagic cell death: the story of a misnomer. Nat Rev Mol Cell Biol. 2008; 9: 1004–10.
- Winslow AR, Rubinsztein DC. Autophagy in neurodegeneration and development. *Biochim Biophys Acta*. 2008; 1782: 723–9.
- 393. Klionsky DJ, Abeliovich H, Agostinis P, et al. Guidelines for the use and interpretation of assays for monitoring autophagy in higher eukaryotes. *Autophagy*. 2008; 4: 151–75.
- 394. Uchiyama Y, Koike M, Shibata M, et al. Autophagic neuron death. Methods Enzymol. 2009; 453: 33–51.
- 395. Franco R, Sanchez-Olea R, Reyes-Reyes EM, et al. Environmental toxicity, oxidative stress and apoptosis: menage a Trois. *Mutat Res.* 2009; 674: 3–22.

- 396. Ribe EM, Serrano-Saiz E, Akpan N, et al. Mechanisms of neuronal death in disease: defining the models and the players. *Biochem J.* 2008; 415: 165–82.
- 397. Woodhouse A, Dickson TC, West AK, et al. No difference in expression of apoptosis-related proteins and apoptotic morphology in control, pathologically aged and Alzheimer's disease cases. Neurobiol Dis. 2006; 22: 323–33.
- 398. Louneva N, Cohen JW, Han LY, et al. Caspase-3 is enriched in postsynaptic densities and increased in Alzheimer's disease. Am J Pathol. 2008; 173: 1488–95.
- 399. Miyoshi K, Ohyagi Y, Sakae N, et al. Enhancement of activation of caspases by presenilin 1 gene mutations and its inhibition by secretase inhibitors. J Alzheimers Dis. 2009; 16: 551–64.
- 400. Raina AK, Sayre LM, Atwood CS, et al. Apoptotic and oxidative indicators in Alzheimer disease. In: LeBlanc AC, editor. Neuromethods, Vol. 37: apoptosis: techniques and protocols, 2nd ed. Totowa, NJ: Humana Press; 2002. pp. 225–46.
- 401. Schwab C, Arai T, Hasegawa M, et al. TDP-43 pathology in familial British dementia. Acta Neuropathol. 2009; 118: 303–11.
- 402. SantaCruz K, Lewis J, Spires T, et al. Tau suppression in a neurodegenerative mouse model improves memory function. *Science.* 2005; 309: 476–81.
- 403. Katsuse O, Lin WL, Lewis J, et al. Neurofibrillary tangle-related synaptic alterations of spinal motor neurons of P301L tau transgenic mice. *Neurosci Lett.* 2006; 409: 95–9.
- 404. Cotman CW, Poon WW, Rissman RA, et al. The role of caspase cleavage of tau in Alzheimer disease neuropathology. J Neuropathol Exp Neurol. 2005; 64: 104–12.
- 405. Rohn TT, Head E. Caspases as therapeutic targets in Alzheimer's disease: is it time to "cut" to the chase? Int J Clin Exp Pathol. 2009; 2: 108–18.
- 406. Schindowski K, Belarbi K, Bretteville A, et al. Neurogenesis and cell cycle-reactivated neuronal death during pathogenic tau aggregation. *Genes Brain Behav.* 2008; 7: 92–100.
- 407. Yang DS, Kumar A, Stavrides P, et al. Neuronal apoptosis and autophagy cross talk in aging PS/APP mice, a model of Alzheimer's disease. Am J Pathol. 2008; 173: 665–81.
- Cookson MR. alpha-Synuclein and neuronal cell death. *Mol Neurodegener*. 2009; 4: 9.

- 409. Shibata N, Kakita A, Takahashi H, et al. Persistent cleavage and nuclear translocation of apoptosis-inducing factor in motor neurons in the spinal cord of sporadic amyotrophic lateral sclerosis patients. Acta Neuropathol. 2009; 118: 755–62.
- 410. **Riederer IM, Schiffrin M, Kovari E**, *et al.* Ubiquitination and cysteine nitrosylation during aging and Alzheimer's disease. *Brain Res Bull.* 2009; 80: 233–41.
- Rossi D, Volterra A. Astrocytic dysfunction: insights on the role in neurodegeneration. *Brain Res Bull.* 2009; 80: 224–32.
- 412. Chadi G, Maximino JR, de Oliveira GP. The importance of molecular histology to study glial influence on neurodegenerative disorders. Focus on recent developed single cell laser microdissection. *J Mol Histol.* 2009; 40: 241–50.
- 413. Lobsiger CS, Cleveland DW. Glial cells as intrinsic components of non-cellautonomous neurodegenerative disease. *Nat Neurosci.* 2007; 10: 1355–60.
- 414. Song YJ, Halliday GM, Holton JL, et al. Degeneration in different parkinsonian syndromes relates to astrocyte type and astrocyte protein expression. J Neuropathol Exp Neurol. 2009; 68: 1073–83.
- 415. Wenning GK, Stefanova N, Jellinger KA, et al. Multiple system atrophy: a primary oligodendrogliopathy. Ann Neurol. 2008; 64: 239–46.
- 416. Uyama N, Uchihara T, Mochizuki Y, et al. Selective nuclear shrinkage of oligodendrocytes lacking glial cytoplasmic inclusions in multiple system atrophy: a 3-dimensional volumetric study. J Neuropathol Exp Neurol. 2009; 68: 1084–91.
- 417. Simpson CL, Lemmens R, Miskiewicz K, et al. Variants of the elongator protein 3 (ELP3) gene are associated with motor neuron degeneration. *Hum Mol Genet.* 2009; 18: 472–81.
- McGeer PL, McGeer EG. Glial reactions in Parkinson's disease. *Mov Disord*. 2008; 23: 474–83.
- 419. Nagai M, Re DB, Nagata T, et al. Astrocytes expressing ALS-linked mutated SOD1 release factors selectively toxic to motor neurons. Nat Neurosci. 2007; 10: 615–22.
- 420. Maragakis NJ, Rothstein JD. Mechanisms of disease: astrocytes in neurodegenerative disease. *Nat Clin Pract Neurol.* 2006; 2: 679–89.
- 421. **Parpura V, Haydon PG.** Astrocytes in pathophysiology of the nervous system. New York: Springer; 2009.
- 422. Zurich MG, Monnet-Tschudi F. Contribution of in vitro neurotoxicology studies to the elucidation of neurodegener-

ative processes. *Brain Res Bull.* 2009; 80: 211–6.

- 423. Choonara YE, Pillay V, du Toit LC, et al. Trends in the molecular pathogenesis and clinical therapeutics of common neurodegenerative disorders. Int J Mol Sci. 2009; 10: 2510–57.
- 424. Kelly BL, Vassar R, Ferreira A. Betaamyloid-induced dynamin 1 depletion in hippocampal neurons. A potential mechanism for early cognitive decline in Alzheimer disease. J Biol Chem. 2005; 280: 31746–53.
- 425. Koffie RM, Meyer-Luehmann M, Hashimoto T, *et al.* Oligomeric amyloid beta associates with postsynaptic densities and correlates with excitatory synapse loss near senile plaques. *Proc Natl Acad Sci U S A.* 2009; 106: 4012–7.
- 426. Lacor PN, Buniel MC, Chang L, *et al.* Synaptic targeting by Alzheimer's-related amyloid beta oligomers. *J Neurosci.* 2004; 24: 10191–200.
- 427. Meyer-Luehmann M, Coomaraswamy J, Bolmont T, *et al.* Exogenous induction of cerebral beta-amyloidogenesis is governed by agent and host. *Science.* 2006; 313: 1781–4.
- 428. Hu X, Crick SL, Bu G, et al. Amyloid seeds formed by cellular uptake, concentration, and aggregation of the amyloid-beta peptide. Proc Natl Acad Sci U S A. 2009; 106: 20324–9.
- 429. Xie YY, Zhou CJ, Zhou ZR, *et al.* Interaction with synphilin-1 promotes inclusion formation of alpha-synuclein: mechanistic insights and pathological implication. *FASEB J.* 2010; 24: 196–205.
- 430. **Desplats P, Lee HJ, Bae EJ, et al.** Inclusion formation and neuronal cell death through neuron-to-neuron transmission of alpha-synuclein. *Proc Natl Acad Sci U S A*. 2009; 106: 13010–5.
- 431. Frost B, Jacks RL, Diamond MI. Propagation of tau misfolding from the outside to the inside of a cell. *J Biol Chem.* 2009; 284: 12845–52.
- Ren PH, Lauckner JE, Kachirskaia I, et al. Cytoplasmic penetration and persistent infection of mammalian cells by polyglutamine aggregates. Nat Cell Biol. 2009; 11: 219–25.
- 433. Naiki H, Nagai Y. Molecular pathogenesis of protein misfolding diseases: pathological molecular environments versus quality control systems against misfolded proteins. J Biochem. 2009; 146: 751–6.
- 434. **Bandopadhyay R, de Belleroche J.** Pathogenesis of Parkinson's disease: emerging role of molecular chaperones. *Trends Mol Med.* 2010; 16: 27–36.

- 435. Wertz IE, Dixit VM. Regulation of death receptor signaling by the ubiquitin system. *Cell Death Differ.* 2010; 17: 14–24.
- Tsvetkov P, Reuven N, Shaul Y. Ubiquitinindependent p53 proteasomal degradation. *Cell Death Differ*. 2010; 17: 103–8.
- 437. Bader M, Steller H. Regulation of cell death by the ubiquitin-proteasome system. *Curr Opin Cell Biol.* 2009; 21: 878–84.
- 438. Michiorri S, Gelmetti V, Giarda E, et al. The Parkinson-associated protein PINK1 interacts with Beclin1 and promotes autophagy. Cell Death Differ. 2010; DOI: 10.1038/cdd.2009.200.
- 439. Rosen KM, Moussa CE, Lee HK, et al. Parkin reverses intracellular beta-amyloid accumulation and its negative effects on proteasome function. J Neurosci Res. 2010; 88: 167–78.
- 440. Smith MA, Zhu X, Tabaton M, et al. Increased iron and free radical generation in preclinical Alzheimer disease and mild cognitive impairment. J Alzheimers Dis. 2010; 19: 363–72.
- 441. Cozzi A, Rovelli E, Frizzale G, et al. Oxidative stress and cell death in cells expressing L-ferritin variants causing neuroferritinopathy. *Neurobiol Dis.* 2010; 37: 77–85.
- 442. Zhao W, Beers DR, Henkel JS, et al. Extracellular mutant SOD1 induces microglial-mediated motoneuron injury. *Glia.* 2010; 58: 231–43.
- 443. Boumezbeur F, Mason GF, de Graaf RA, et al. Altered brain mitochondrial metabolism in healthy aging as assessed by in vivo magnetic resonance spectroscopy. J Cereb Blood Flow Metab. 2010; 30: 211–21.
- Martinelli P, Rugarli El. Emerging roles of mitochondrial proteases in neurodegeneration. *Biochim Biophys Acta*. 2010; 1797: 1–10.
- 445. **Gibson GE, Starkov A, Blass JP**, *et al.* Cause and consequence: mitochondrial dysfunction initiates and propagates neuronal dysfunction, neuronal death and behavioral abnormalities in age-associated neurodegenerative diseases. *Biochim Biophys Acta.* 2010; 1802: 122–34.
- 446. Du H, Yan SS. Mitochondrial permeability transition pore in Alzheimer's disease: cyclophilin D and amyloid beta. *Biochim Biophys Acta*. 2010; 1802: 198–204.
- 447. Moreira PI, Carvalho C, Zhu X, et al. Mitochondrial dysfunction is a trigger of Alzheimer's disease pathophysiology. Biochim Biophys Acta. 2010; 1802: 2–10.
- 448. Lynn BC, Wang J, Markesbery WR, Lovell MA. Quantitative changes in the mitochondrial proteome from subjects

with mild cognitive impairment, early stage, and late stage Alzheimer's disease. *J Alzheimers Dis.* 2010; 19: 325–39.

- 449. Wang X, Su B, Perry G, et al. Insights into amyloid-beta-induced mitochondrial dysfunction in Alzheimer disease. Free Radic Biol Med. 2007; 43: 1569–73.
- 450. Yu JT, Chang RC, Tan L. Calcium dysregulation in Alzheimer's disease: from mechanisms to therapeutic opportunities. *Prog Neurobiol.* 2009; 89: 240–55.
- 451. Green KN. Calcium in the initiation, progression and as an effector of Alzheimer's disease pathology. *J Cell Mol Med.* 2009; 13: 2787–99.
- 452. Winklhofer KF, Haass C. Mitochondrial dysfunction in Parkinson's disease. *Biochim Biophys Acta*. 2010; 1802: 29–44.
- 453. Estevez AR, Arduino DM, Swerdlow RH, et al. Microtubule depolymerization potentiates alpha-synuclein oligomerization. Front Ag Neurosci. 2010; 1: doi:10.3389/ neuro.24.005.2009.

- 454. Gandhi S, Wood-Kaczmar A, Yao Z, et al. PINK1-associated Parkinson's disease is caused by neuronal vulnerability to calcium-induced cell death. *Mol Cell.* 2009; 33: 627–38.
- 455. Gegg ME, Cooper JM, Schapira AH, et al. Silencing of PINK1 expression affects mitochondrial DNA and oxidative phosphorylation in dopaminergic cells. *PLoS One.* 2009; 4: e4756.
- Damiano M, Galvan L, Deglon N, et al. Mitochondria in Huntington's disease. Biochim Biophys Acta. 2010; 1802: 52–61.
- 457. Shi P, Gal J, Kwinter DM, et al. Mitochondrial dysfunction in amyotrophic lateral sclerosis. *Biochim Biophys Acta*. 2010; 1802: 45–51.
- 458. **Martin LJ.** The mitochondrial permeability transition pore: a molecular target for amyotrophic lateral sclerosis therapy. *Biochim Biophys Acta.* 2010; 1802: 186–97.
- 459. Kichev A, Ilieva EV, Pinol-Ripoll G, *et al.* Cell death and learning impairment in mice

caused by *in vitro* modified pro-NGF can be related to its increased oxidative modifications in Alzheimer disease. *Am J Pathol*, 2009: 175: 2574–85.

- 460. Lynch MA. Age-related neuroinflammatory changes negatively impact on neuronal function. *Front Ag Neurosci.* 2010; 1: Doi:10.3389/neuro.24.006.2009.
- 461. Ryu JK, McLarnon JG. A leaky blood-brain barrier, fibrinogen infiltration and microglial reactivity in inflamed Alzheimer's disease brain. J Cell Mol Med. 2009; 13: 2911–25.
- 462. Davenport CM, Sevastou IG, Hooper C, et al. Inhibiting p53 pathways in microglia attenuates microglial-evoked neurotoxicity following exposure to Alzheimer peptides. J Neurochem. 2009; 112: 552–63.
- 463. Long-Smith CM, Sullivan AM, Nolan YM. The influence of microglia on the pathogenesis of Parkinson's disease. *Prog Neurobiol.* 2009; 89: 277–87.