CELLULAR REACTIONS TO WAXES FROM MYCO-BACTERIUM LEPRAE

By F. R. SABIN, M.D., K. C. SMITHBURN, M.D., AND R. M. THOMAS, M.D. (From the Laboratories of The Rockefeller Institute for Medical Research)

PLATE 34

(Received for publication, July 9, 1935)

The plan for chemical analysis of the Mycobacteria sponsored by the National Tuberculosis Association included an acid-fast strain isolated from a case of leprosy, as well as selected strains of human, bovine, and avian tubercle bacilli, and this study deals with the waxes extracted from *Mycobacterium leprae.*¹ The chemical analyses have not yet been completely reported (1, 2). The materials used have been in different stages of purification and have therefore provided the opportunity to compare the complex cellular reactions to mixtures of substances with the simple response to highly purified crystalline alcohols.

Materials and Methods

The strain of organisms used for the analysis was obtained from a case of leprosy (Apa case) in Honolulu, about 1909. It has been kept at the Hygienic Laboratory, Washington, D. C., as Strain 370; on Feb. 4, 1926, a subculture was given to the Mulford Biological Laboratories, Glenolden, Pennsylvania. It was there grown on Long's synthetic medium in quantities adequate for chemical analysis.

Five preparations of wax fractions from the Bacillus leprae have been studied.

1. A Crude Chloroform-Soluble Wax.—This was obtained in the same manner as the corresponding fraction from tubercle bacilli (3). This fraction has not yet been analyzed completely, but Dr. Anderson reports that it is almost free from phosphorus and nitrogen and hence is not a phosphatide (1). It is a red, wax-like substance, looking like beeswax except for the color.

771

¹ We are indebted for this material to Dr. R. J. Anderson and his associates at Yale University, who extracted the waxes from Mycobacterium leprae.

soluble in a mixture of ether and acetone than the phosphatide, and it came out on cooling the acetone solution (1). Concerning this product, Dr. Anderson has written us, "This product consists mainly of a solid glyceride which I call 'Leprosin.' It contains a number of fatty acids and the alcohols $C_{20}H_{42}O$ and $C_{18}H_{38}O$, identical with the alcohols isolated from the wax of the timothy grass bacillus." This material is a slightly pink, amorphous powder; that is, it carries some of the pigment so characteristic of the *B. leprae*.

3. Leprosin.—This was obtained from the purification of the preceding material. Though a glyceride instead of an alcohol, it corresponds in a general way in its physical properties to the unsaponifiable material from tubercle bacilli. It is a snow white powder, with a melting point of 54°C. It is optically active.

$[\alpha]_{n}^{2}$ in chloroform + 4.2°.

4. Leprosinic Acid.—This was obtained from the analysis of the leprosin and in physical properties is like it. It has a melting point of $61-62^{\circ}$ C. and is dextrorotatory:

$$[\alpha]_{\rm D}^{23}$$
 in chloroform + 4.7°.

All four of these materials from the B. leprae are acid-fast.

5. Alcohol II from the Unsaponifiable Matter of the B. leprae.—This is a pure white material consisting of feather-like crystals. It has a melting point of 84°C. and a formula of $C_{25}H_{44}O_2$. It is not acid-fast.

Besides this alcohol from the B. *leprae* we have also received from Dr. Anderson a similar material as follows:

Alcohol Isolated from the Wax of the Timothy Grass Bacillus.—This is a pure white material made up of fine, needle-like crystals. Its melting point is 63°C. It is dextroortatory:

$[\alpha]_{n}^{n}$ in ether + 6.93°.

Its formula is $C_{20}H_{42}O$. It is not acid-fast.

None of these preparations has been given in nujol, the method used in our first study of wax fractions from tubercle bacilli (4). Rather it has been found better to introduce these materials in the form of a dry powder through an incision under ether anesthesia. The crude waxes can be ground into a fine powder if they are first chilled with dry ice until brittle. The more purified substances, the leprosin, the leprosinic acid, and the alcohols, do not need more than simple grinding to separate the clumps of crystals.

The method of preparing these waxes from the *B. leprae* in colloidal suspension, which was developed by one of us with the unsaponifiable material from the tubercle bacillus (5), was tried with the lepra fractions but with less success. It consisted of dissolving the wax in chloroform and adding an equal amount of hot alcohol. To this solution the same amount of distilled water was added drop by

drop, which procedure threw down the wax in the form of a precipitate. The suspension obtained from the crude chloroform-soluble wax was not as fine as that from the unsaponifiable material from the tubercle bacillus but rather consisted of particles about 7μ in diameter which settled out into quite large aggregates. This material, therefore, had to be freshly prepared for each injection.

When the crude wax obtained from the purification of the lepra phosphatide was prepared for colloidal suspension, the precipitate was of more or less regular pentagonal particles, highly refractive and suggesting a crystalline form. There was no agglutination of these particles.

In attempting to prepare similar suspensions of leprosin and leprosinic acid, the addition of distilled water resulted in the separation of minute droplets of an oily nature which formed an unstable emulsion. This emulsion had a tendency to curdle and adhere to the surface of the flask. This was in marked contrast to the stability of the suspension prepared from the unsaponifiable material from H-37.

RESULTS

Reactions to the Crude Chloroform-Soluble Wax from B. leprae.—The crude chloroform-soluble wax from the B. leprae was given intraperitoneally to six rabbits, as shown in Table I.

Rabbits R 4268 and R 4269 both received the material after it had been powdered with dry ice. The material did not give a simple reaction; where the wax had lodged, the area became infiltrated with leucocytes making an abscess. Around these abscesses were monocytes, fibroblasts, and clasmatocytes, filled with leucocytes. In the omentum the reaction was not uniform; there were abscesses, bands of fibroblasts, and foci of monocytes, some showing partial fusion into giant cells. There was a marked dilatation of the vessels. The retrosternal lymph nodes had their sinuses filled with monocytes and in some places they were present also in the follicles.

The same tendency toward the formation of abscesses followed the introduction of the material in colloidal suspension.

This preparation showed a marked tendency to a clumping of the particles, with the result that the dose varied; Rabbit R 4189 received a small dose, R 4187 an average dose, and R 4188 a massive dose. The abscesses were smaller because the masses of the wax had been smaller. They were filled with clasmatocytes containing leucocytes. Around the abscesses were monocytes and giant cells, many of them vacuolated, indicating a phagocytosis of the wax. The edge of one of the abscesses with a narrow border of giant cells is shown in Fig. 1, from Rabbit R 4188. Bands of fibroblasts were extensive; in one animal of the group, R 4188, there was an extreme involvement of the retrosternal lymph nodes with

н	
TABLE	

	11
e	1
570	11
ã.	
e	11
8	11
52	Ш
Š.	н
2	11
5	Н
Wax from E	
5	11
4	11
ble Wax	
qn	11
10	IJ
25	1
۲ .	н
w.	Ш
E	Ц
5	Н
lorofori	11
<u> </u>	II.
1	11
S.	Н
te Chlorofo	11
Ŀ.	11
11	11
Ü	
e Crud	
1	li
~	11
2	Ш
:0	IJ.
8	
S.	n
ĸ	Ш
4	11
.2	Ш
2	Ш
2	
5	11
. 3	Iì.
29	11
ĩ	11
f Rabbits Which Recei	11
of Ro	
0,	11
ls	
8	11
õ	11
10	11
~	11
1	Н

				Peri	Peritoneal exudate	date	
Animal No.	no. and amount of injections	Time	Method of preparing material	P.	Percentage of	l l	Tissues
		-		PMN	Lymph.	Mono.	
		days					
R 2795		4	Dry powder through	1		1	Most of the wax in one bolus. Coccidiosis present
	20 mg.	K	cannula				
R 4268		6	Material powdered	15.0	14.0	70.80	Many abscesses, 2 to 3 cm. in diameter in omen-
	50 mg.	R	with dry ice and				tum, on cecum, in the peritoneal wall, and in
			introduced through				incision. Abscesses surrounded by monocytes
			incision under anes-				and fibroblasts, with clasmatocytes containing
			thesia				leucocytes in the border. Omentum almost too
							massive to study as film and very complex:
							zones of fibroblasts; many small foci of mono-
							cytes showing partial fusion into giant cells, the
							center of which contain leucocytes. Marked
							vascular dilatation. Retrosternal nodes have
							sinuses filled with monocytes
R 4269		9	yy yy	63.68	0	36.31	Many abscesses 2 to 3 cm. in diameter in omentum
	50 mg.	K					surrounded by monocytes, fibroblasts, and clas-
							matocytes containing leucocytes. Many small
							foreign body giant cells with vacuolated cyto-
							plasm. Diaphragm shows tubercles of giant
							cells in which there is only partial fusion of the
							monocytes. Leucocytes infiltrating the tissues.
							Bands of fibroblasts; marked dilatation of the
							vessels. Extensive reactions of the same type
							in retrosternal nodes replacing part of the
							follicles

774

Totter.	,			100	40	20 24	W
K_410/	20 mg.	4 4	Comonan suspension	1.10	61.22		abscesses in body wall surrounded by fibro-
		K 5 days after					blasts and clasmatocytes containing leucocytes.
		third injec-					Omentum has no large abscesses but many small
		tion	-				tubercles of foreign body giant cells and mono-
							cytes, some vacuolated. In the center of the
							tubercles were leucocytes. Dilatation of the
						_	vessels and marked bands of fibroblasts. Few
							giant cells in retrosternal lymph node. One
							small hemorrhage in one lung, infiltrated with
							monocytes
R 4188	3	4	8 8	8.88	24.36	66.75	No symptoms. No rise in temperature. Ab-
	20 mg.	4	_				scesses 4 to 10 mm. in diameter on surface of
		K 5 days after					cecum, in capsule of liver and spleen, on dia-
		third injec-					phragm, in body wall, and in the omentum.
		tion					Abscesses contain clasmatocytes filled with leu-
							cocytes and are surrounded by a narrow band
							of monocytes. Omentum has many small tuber-
							cles of foreign body giant cells and monocytes,
							some of them vacuolated. Dilatation of ves-
							sels and bands of fibroblasts. Lungs had several
							small, translucent nodules which were made up
							of monocytes. Retrosternal nodes show ex-
							treme involvement with monocytes and giant
							cells both in the sinuses and in the follicles. In
				- 1			the fresh tissue, typical epithelioid cells were
							seen, confirmed in sections
R 4189	3	4	77	0.25	42.45	. 57.28	Very slight reaction; a few abscesses in the body
	20 mg.	4					wall and a few foreign body giant cells in the
		K 5 days after					omentum
	,	tinira injec-					
		non	-		_		

Ħ
BLE
TA]

Protocols of Rabbits Which Received the Wax Obtained in the Purification of the Phosphatide of B. leprae

Animal No. R 2715	No. and amount of injections 1 20 mg.	Time days K	Method of preparing material Dry powder through cannula blown in with air	Peri Peri 1.02	Peritoneal exudate Percentage of <u>PMN</u> Lymph. <u>M</u> 1.02 5.10 93	date of 93.87	Tissues Omentum showed increased size of milk spots with particles of wax on them. Increase in mono- cytes; occasional fusion into giant cells, some having as many as 50 nuclei. Few epithelioid cells. Infiltration with neutrophilic leucoytes and a few myelocytes. No increase in fbro- blasts. Retrosternal node has many monocytes
R 2800	1 20 mg.	¥ ¥	Dry powder through incision under anes- thesia	1.13		1.13 97.72	in the sinuses Omentum showed increased size of milk spots with particles of wax on them. Around the particles were young monocytes showing no signs of phagocytosis; that is, no vacuoles staining with neutral red. Few neutrophilic and eosinophilic leucocytes. No increase in fibroblasts. Eosino-
R 2798	2 20 mg.	4 D	Dry powder through cannula. Died after second injec- tion which punc- tured the liver			1	philc myelocytes prominent in the bone marrow Wax in single bolus and reaction slight. Few foreign body giant cells in omentum. Blood in retrosternal lymph nodes 5 min. after puncture of the liver

776

d 3 hrs. after the second injection the animal had a convulsion, was extremely sensitive on being	touched, became cyanotic, pulse was about 250 and respiration was slow. Temperature was	111.5° and the animal was killed. Small ab-	ocesses in mesencery, utapiragin, and omenum. Omentum showed increased density of the milk	spots with monocytes and giant cells, dilatation	of the vessels, neutrophilic leucocytes, both free	and in clasmatocytes, and a few epithelioid	cells. Marked damage of the lymphocytes in	the nodes, probably due to the temperature.	Congestion of the lungs and liver. Hemorrhage	in the bone marrow	49.10 No symptoms; no rise in temperature. Small	abscesses on peritoneal wall near places of in-	jections, on the large intestine, on the dia-	phragm, and in the omentum. Milk spots of	omentum increased in size and number. Ab-	scesses of omentum contain many clasmatocytes	filled with leucocytes and have a wide border	of giant cells. Many giant cells in tubercles	and diffusely scattered. Marked vasculariza-	tion, bands of fibroblasts, and increase in fibrous	tissue. Acid-fast stain negative. Retrosternal	nodes show extreme numbers of monocytes in	the sinuses and the follicles are almost replaced	by them. Small, translucent nodules in lungs	made up of monocytes
Cells all dead											45.29											<u> </u>			
Cell											5.59	•			~,					<u></u> .					
Dry powder through cannula blown in	with air				<u> </u>						Colloidal suspension	in 4 cc. water													
4 K 3 <u>4</u> hrs. after	second in- jection										4	4	K 6 days after	the third in-	jection										
2 20 mg.)										.	20 mg.									-				
R 2763			·							****	R 4184														

777

	6
	ġ
	-3
	<u> </u>
	- 6
	Ŷ
	- 1
	Ė
	님
	Ĕ
	7
	TABLE

			AADO	TWC		110			SU1	0.00	L	CFI		5			
	Tissues			like those of R 4184, except less formation of fibers in the omentum and less reaction in the	retrosternal nodes. Nodules of monocytes in	No symptoms after the first injection. After the	second, rapid respiration and temperature fell	temperature fell 2.5° and then rose 4.8°. One	of the injections had lodged in the sheath of the	external oblique muscle, where there was a	them vacualities, and monocytes, many of	cytes but many cosinophilic. Omentum showed	increased density of the milk spots due to mono-	cytes and giant cells, whose cytoplasm was filled	with vacuoles. Some wax still to be seen on	the milk spots. Some typical epithelioid cells.	Moderate increase in fibroblasts
late	l l	Mono.	54.05			81.25											
toneal exu	ercentage (Lymph.	18.31			13.54											
Perù	Å	PMN	27.62			5.20			<u> </u>								-
	Method of preparing material		Colloidal suspension	in 4 cc. water		Dry powder through	cannula										
	Time		days 4	4 K 6 days after	the third in- iection	4	4 K 5 daws after	the third in-	jection								
No. and	amount of injections		3	20 mg.		3	20 mg.										
			R 4185			R 2771											
		Retioneal exudate Method of preparing Percentage of	No. and amount of injections Time Method of preparing Percentage of PMN Lymph. Mono.	No. and amount of injections Time Method of preparing Peritoneal exudate amount of injections Pinne Percentage of adoys days PMN Lymph. 3 4 Colloidal suspension 27.62 18.31	No. and amount of linjections Time Time Method of preparing Peritoneal exudate Amount of linjections Time Method of preparing Peritoneal exudate amount of linjections PMN Lymph. Mono. adors days Colloidal suspension 27.62 18.31 54.05 X 6 days after in 4 cc. water N N N	No. and amount of injections Time Time Method of preparing Peritoneal exudate amount of injections Time Method of preparing Percentage of amount of injections amount of Percentage of Mono. abys days Colloidal suspension 27.62 18.31 54.05 20 mg. 4 in 4 cc. water in 4 cc. water in ection	No. and amount of injections Time Time Method of preparing Peritoneal exudate Mount of injections Time Method of preparing Percentage of amount of injections FMN Lymph. Mono. 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 in 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25	No. and injectionsTimeMethod of preparingPeritoneal exudateNo. and injectionsTimeMethod of preparingPercentage ofamount of injectionsdaysExample.Mono.34Colloidal suspension27.6218.3154.0534in 4 cc. water27.6218.3154.0534Dry powder through5.2013.5481.2520 mg.4Dry powder through5.2013.5481.25	No. and amount of injectionsTimeMethod of preparingPeritoneal exudateNo. and injectionsTimeMethod of preparingPercentage of IPMNIXmph.anount of injectionsaoysColloidal suspension27.6218.3154.0534In 4 cc. water27.6218.3154.0534In 4 cc. water27.6218.3154.0534Dry powder through5.2013.5481.2520 mg.4Canula5.2013.5481.2534Dry powder through5.2013.5481.2520 mg.K 5 days afterthe third in-the third in-13.5481.25	No. and injectionsTimeMethod of preparingPeritoneal exudateNo. and injectionsTimeMethod of preparingPercentage of34Colloidal suspension27.6218.3154.0534Colloidal suspension27.6218.3154.0534In 4 cc. water27.6218.3154.0534Dry powder through5.2013.5481.2534Carys aftersamula5.2013.5481.25910Fibrid in-fibrid in-fibrid in-fibrid in-34Dry powder through5.2013.5481.251011Fibrid in-fibrid in-fibrid in-10fibrid in-fibrid in-fibrid in-fibrid in-10fibrid in-fibrid in-fibrid in-fibrid in-11fibrid in-fibrid in-fibrid in-fibrid in-10fibrid in-fibrid in-fibrid in-fibrid in-10fibrid in-fibrid in-fibrid in-fibrid in-10fibrid in-fibrid in-fibrid in-fibrid in-11fibrid in-fibrid in-fibrid in-11fibrid in-fibrid in-fibrid in-11fibrid in-fibrid in-fibrid in-12fibrid in-fibrid in-fibrid in-13fibrid in-fibrid in-fibrid in-13fibrid in-fibrid in-fibrid in- <t< td=""><td>No. and injectionsTimeMethod of preparingPeritoneal exudateNo. and injectionsTimeMethod of preparingPercentage of34Colloidal suspension27.6218.3154.0534Colloidal suspension27.6218.3154.0520 mg.K 6 days after in 4 cc. water27.6218.3154.0534Dry powder through5.2013.5481.2520 mg.K 5 days after ijectionS.2013.5481.25</td><td>No. and injectionsTimeMethod of preparingPeritoneal exudateNo. and injectionsTimeMethod of preparingPercentage of34Colloidal suspension27.6218.3154.0534Colloidal suspension27.6218.3154.0520 mg.4in 4 cc. water27.6218.3154.0534Dry powder through5.2013.5481.2520 mg.K 5 days aftercannula5.2013.5481.25jectionjectionjectionjection13.5481.25</td><td>No. and injections Time amount of material Method of preparing material Peritoneal exudate Anound injections Anon Aays Anon Aays Percentage of Anon Mono. 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 in 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 3 4 cannula 5.20 13.54 81.25 5 13.54 13.54 81.25 4 cannula 5.20 13.54 81.25</br></br></td><td>No. and amount of injections Time Time Method of preparing Peritoneal exudate Amount of injections Time Method of preparing Percentage of 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 In 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 3 4 Canula cannula 5.20 13.54 81.25 3 4 Canula in- fortion 5.20 13.54 81.25 3 4 Canula fortion 5.20 13.54 81.25</td><td>No. and injections Time amount of material point Method of preparing material point Peritoneal exudate No. and injections Time for material point Method of preparing Peritoneal exudate 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 In 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 20 mg. 4 cannula 5.20 13.54 81.25 9 jection jection jection 5.20 13.54 81.25</td><td>No. and injections Time material material Method of preparing material Peritoneal exudate Anonotic injections Time material Method of preparing Peritoneal exudate 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 In 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 20 mg. 4 cannula 5.20 13.54 81.25 9 jection jection jection 5.20 13.54 81.25</td><td>No. and injections Time Method of preparing material material Peritoneal exudate 3 4 PMN Lymph. Mono. 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 In 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 3 4 cannula 5.20 13.54 81.25</td></t<>	No. and injectionsTimeMethod of preparingPeritoneal exudateNo. and injectionsTimeMethod of preparingPercentage of34Colloidal suspension27.6218.3154.0534Colloidal suspension27.6218.3154.0520 mg.K 6 days after in 4 cc. water27.6218.3154.0534Dry powder through5.2013.5481.2520 mg.K 5 days after ijectionS.2013.5481.25	No. and injectionsTimeMethod of preparingPeritoneal exudateNo. and injectionsTimeMethod of preparingPercentage of34Colloidal suspension27.6218.3154.0534Colloidal suspension27.6218.3154.0520 mg.4in 4 cc. water27.6218.3154.0534Dry powder through5.2013.5481.2520 mg.K 5 days aftercannula5.2013.5481.25jectionjectionjectionjection13.5481.25	No. and injections Time amount of material Method of preparing material Peritoneal exudate Anound 	No. and amount of injections Time Time Method of preparing Peritoneal exudate Amount of injections Time Method of preparing Percentage of 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 In 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 3 4 Canula cannula 5.20 13.54 81.25 3 4 Canula in- fortion 5.20 13.54 81.25 3 4 Canula fortion 5.20 13.54 81.25	No. and injections Time amount of material point Method of preparing material point Peritoneal exudate No. and injections Time for material point Method of preparing Peritoneal exudate 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 In 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 20 mg. 4 cannula 5.20 13.54 81.25 9 jection jection jection 5.20 13.54 81.25	No. and injections Time material material Method of preparing material Peritoneal exudate Anonotic injections Time material Method of preparing Peritoneal exudate 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 In 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 20 mg. 4 cannula 5.20 13.54 81.25 9 jection jection jection 5.20 13.54 81.25	No. and injections Time Method of preparing material material Peritoneal exudate 3 4 PMN Lymph. Mono. 3 4 Colloidal suspension 27.62 18.31 54.05 3 4 In 4 cc. water 27.62 18.31 54.05 3 4 Dry powder through 5.20 13.54 81.25 3 4 cannula 5.20 13.54 81.25

monocytes and giant cells, as shown in Fig. 3 which is a section through the sinuses. The follicles of this node were also extensively replaced by similar cells. Besides these reactions, typical epithelioid cells were found in the omenta and the retrosternal lymph nodes of these animals, both with the supravital technique and in sections.

Reactions to Wax from the Purification of the Phosphatide of B. leprae.—The wax obtained from the purification of the lepra phosphatide was given to seven rabbits, as shown in Table II. This was the only material given which was followed by any symptoms and they proved to be inconstant.

The early reaction was shown by two rabbits, R 2715 and R 2800, one of which received the material through a cannula and the other through an incision. In neither animal were there symptoms after the injection. The wax had lodged on the milk spots of the omentum, and around the particles of the wax there had been a multiplication of monocytes; in places these monocytes had started to fuse into giant cells. There were occasional epithelioid cells but there was no increase in fibroblasts.

Rabbit R 2763 showed extreme disturbance after the second injection, 3 hours later it had a convulsion, the temperature rose to 111.5° , and the animal was therefore killed. A second rabbit, R 2771, had a rise in temperature of 3.3° after a third injection but survived. We did not determine the cause of the fever in Rabbit R 2763. The material injected showed no bacteria. The brain and cord were normal; there were small abscesses in the omentum, in the mesentery, and on the diaphragm, but not as many as in the rabbits of the preceding series. There was the same reaction of monocytes and giant cells around the wax which had lodged on the milk spots of the omentum.

Rabbit R 2771 showed similar cellular reactions, complex in type; there were foreign body giant cells around the particles of wax, a few neutrophilic leucocytes, and many eosinophilic leucocytes. There were epithelioid cells and an increase in fibroblasts.

The material was then given to two rabbits, R 4184 and R 4185, in three doses in colloidal suspension. None of these injections caused symptoms and there was no rise in temperature. In making the suspension, the material had been heated. There were small abscesses wherever the wax had lodged. The abscesses had wide borders of monocytes and giant cells, as is shown in Fig. 2. The width of the border of giant cells can be seen to be greater than in Fig. 1, which was taken from an animal which had received the crude chloroform-soluble wax. Indeed, almost the entire section shown in Fig. 2 is of monocytes except the abscess on the lower border. Besides this, the material from Rabbit R 4184 showed many large masses of giant cells, some in tubercles and some diffusely scattered, so that when the total reaction was compared with that in the animals

e prosin		Tissues	10.		Ż	shows marked increase in size of milk spots with	foreign body giant cells enclosing the wax.	Marked bands of fibroblasts growing as in tissue culture. No epithelioid cells. Eosinophilic leu-	cocytes in the retrosternal lymph nodes	75 In omentum the predominant reaction is an ex-	tensive formation of tubercles of foreign body	giant cells. Some have hollow centers and	dense cytoplasm; others have a vacuolated cyto-	plasm. A few abscesses in the omentum, with	walls dense with giant cells. Except for these	abscesses very slight infiltration with neutro-	philic leucocytes. Increase in fibrous tissue.	Abscess in body wall near site of injection with	monocytes and giant cells in its border. Retro-	sternal nodes show an extensive infiltration of	sinuses and follicles with foreign body giant	cells and many eosinophilic leucocytes	Ř	showed wax with foreign body giant cells. One	toneal exudate
ing Le	udate	of	Mono.		88.0				-	80.75													96.66		
s Receiv	Peritoneal exudate	Percentage of	Lymph.		5.0					13.50													3.20		
Rabbil.	Per	н,	PMN		6.5					5.0													0		_
Protocols of Rabbits Receiving Leprosin		Method of preparing material			Dry powder through	incision under anes-	thesia			Colloidal suspension													77 71		
		Time		days	4	K 4 days after	second in-	jection		7	K									-			7	K	
		amount of injections			7	20 mg.					50 mg.												1	50 mg.	
		Animal No.			R 2801					R 4264													R 4265		

TABLE III

Protocols of Rabbits Receiving Lepro

.

780

No symptoms. Omentum shows foreign body giant cells singly and in tubercles. The giant	cells have enclosed the wax and a few leucocytes. Very little vacuolization of the cytoplasm of the	giant cells. Increase in inproplasts and in bands of fibers. A few small abscesses, but almost no neutrophilic leucocytes elsewhere. No eosino-	9.64 90.35 Every milk spot of omentum has foreign body	giant cells, some extremely large. Their cyto- plasm is markedly vacuolated. Tubercles of	giant ceus seen in section. Many pands of fibroblasts and many eosinophilic leucocytes in	the omenum. Many cosmophiles in the retro- sternal nodes
(· <u></u>	<u></u>	<u>.</u>	. <u>.</u>		
			0			
Dry powder through incision under anes-			3			
Dry pow incisio	thesia		3			
days	after the sec- ond injec-		1	24 days after the sec-	tion injec-	
2 4 20 mg K 14 days	after ond	II OT	4 7	4	tion	
2 20 mg.			2	20 mg.		
R 2812			R 2807			

that received the crude chloroform-soluble wax, it was clear that this preparation had a much greater proportion of the factor, that is the wax, that produces the foreign body giant cells. Besides the giant cells there was a marked dilatation of the blood vessels, seen also in Fig. 2, as well as signs of the new formation of fibroblasts and of fibrous tissue.

Reactions to Leprosin.—Five rabbits have received the leprosin and their protocols are given in Table III. The formation of abscesses, so marked a feature of the reaction to the two preceding materials, was much reduced, appearing only in Rabbit R 4264.

There were two constant reactions to the material, an extensive formation of foreign body giant cells and a marked development of bands of fibroblasts. These bands are shown in Fig. 4, from the omentum of Rabbit R 2801. The photograph was taken from a fresh film, stained with neutral red, and was made while the cells were living. It will be noted that the bands of fibroblasts are growing in a manner to simulate a tissue culture. The giant cells occurred singly and in tubercles and had eosinophilic leucocytes around them. No epithelioid cells were seen after the injection of the leprosin.

Reactions to Leprosinic Acid.—The cellular reactions to the intraperitoneal injections of leprosinic acid are shown in five rabbits, the protocols of which are given in Table IV. In every instance the reaction to this material has been of a single cell type; namely, there has been a multiplication of monocytes around the particles of the wax and their fusion into giant cells.

In the omentum it was clear that this material also lodged only on the milk spots which became tubercles of giant cells. The differential counts of the cells of the peritoneal exudates of these animals show how small a part of the reaction is made by neutrophilic leucocytes, for only in the first animal of the series were there any of these cells and in that instance, only 2.5 per cent. Instead the tissues were infiltrated with eosinophilic leucocytes.

Reactions to Alcohols from the Waxes of the B. leprae and B. phlei.— These two highly purified, crystalline alcohols, $C_{25}H_{44}O_2$, from the unsaponifiable wax material of the B. leprae, and $C_{20}H_{42}O$, from the B. phlei, or timothy grass bacillus, were each given intraperitoneally to two rabbits in doses of 15 mg. They were given in each instance as the dry crystals through an incision under ether anesthesia.

Rabbits R 4272 and R 4241 each received the alcohol from the *B. leprae* and were killed in 5 days. Rabbit R 4236 and R 4238 received the alcohol from the

EIV	Protocols of Rabbits Receiving Leprosinic Acid		Tissues			Cuncutum suoweu accentuateu muk spots with wax enclosed in giant cells. Tubercles of foreign body giant cells on diaphragm infiltrated with easing-	philic leucocytes. Many eosinophiles in the retrosternal lymph nodes Omentum showed flecks of wax on the milk spots surrounded by foreign body	giant cells. These giant cells were not as large as in R 4270 but there were	as many as yo to 40 on some muk spots. Foci of monocytes in septa of lungs and some giant cells in the retrosternal lymph nodes	Omentum showed flecks of wax on the milk spots surrounded by foreign body	giant cells. Some of them had only three or four nuclei; others were so large that they completely filled a low power field. Many monocytes: no large	cytes seen. No foci of monocytes found in the lungs	Omentum showed many giant cells on the milk spots, some with 30 to 40	and there was one giant cell about 100μ in diameter. A focus of giant cells	and monocytes near the point of injection was infiltrated with eosinophilic	neucocyces. A tew toot of monocytes in the fungs and extensive involve- ment of both sinuses and follicles of retrosternal lymph nodes with monocytes	Less reaction than in R 4266 but of same type. Monocytes and foreign body giant cells containing wax on the milk spots of the omentum
TABLE IV	abbits Re	late		Mono.			85.18			86.59	<u>.</u>		83.50				85.20
	ols of R	Peritoneal exudate	Percentage of	Lymph.	7.07	5	14.81			13.40		1	15.97				14.79
	Protoc	Peri	Ă	NWA	1 53	4	0			0		1	16.0				0
			Method of preparing material		Dry nourder through	cannula	Dry powder through	incision under anes-		y y		T-F5-11	umperrect colloidal suspension	4		:	:
			Time		days 5	R	22	R	<u>. </u>		4	24	к Я			ç	8 M
		No. and	amount of injections		-	20 mg.		30 mg.	,	20 1	ou nug.		40 mg.)		•	40 mg.
			No.		R 2769		R 4271	792		K 4270		7766 G				7307 C	K 420/

timothy grass bacillus and were killed in 4 and 6 days respectively. The reaction was of giant cells around the crystals of alcohol; these crystals lodged on the milk spots of the omentum. The giant cells were proportional to the size of the crystals, whether they occurred singly or in a clump. There were more neutrophilic leucocytes in the peritoneal fluid than when the time interval had been longer, ranging from 5 to 11 per cent, except in the case of Rabbit R 4238, in which there was an adhesion of a part of the omentum to the peritoneal wall and to the liver, and the leucocytes were 39.08 per cent.

These experiments have not been completed; they involve a study of the reactions to these materials in the tuberculous rabbits, as well as in the normal ones.

DISCUSSION

The materials from the lepra bacillus have given an interesting opportunity to follow the chemical separation of complex mixtures by cells instead of in the test tube. It is clear that all of these materials contain some substance which, like the solid alcohol $C_{25}H_{44}O_2$, gives the formation of the simple foreign body giant cell. The crude chloroform-soluble wax and the wax obtained from the purification of the lepra phosphatide contain, however, many more substances that give cellular reactions. With both of them the most striking phenomenon is the formation of large abscesses wherever the masses of the wax lodge. Thus there is some substance chemotaxic to leucocytes in or on the wax which acts like the tuberculo-polysaccharide. That is, this material calls leucocytes from the blood stream and so damages them that they are readily engulfed by clasmatocytes. The leucocytes are found in clasmatocytes both in the local lesions and in the spleen. These abscesses are not at all like caseation for there is no basis of dead cells, but rather it is the wax itself which becomes infiltrated with leucocytes.

Besides this reaction, there is the formation of the foreign body giant cells with the fused monocytes enclosing both the wax and the leucocytes. In the case of the wax obtained from the analysis of the lepra phosphatide, this reaction is much greater in amount than with the crude chloroform-soluble wax, indicating that the wax-like material is in much greater proportion in this fraction. The tissues also become infiltrated with eosinophilic leucocytes and lymphocytes. These materials also are irritants, causing a marked dilatation of the blood vessels and probably a new growth of vessels. They also contain substances that are remarkable stimulants for the formation of fibroblasts and of new fibers. Besides all of these properties, they contain some material that gives rise to typical epithelioid cells, exactly like the reaction to the tuberculo-phosphatide. Thus these two materials give reactions as complex as those aroused by the acetone-soluble material from the tubercle bacillus (6). The cellular reactions to them are similar to those aroused by the crude wax which can be separated from tubercle bacilli. These complex cellular reactions reflect the fact that some of all the different types of lipoids come out with the first use of lipoidal solvents on mixtures.

The leprosin is an entirely different type of material; it is a pure white powder in crystalline form which does not give the cellular reactions characteristic of complex mixtures of lipoids, such as have just been described. Rather its reactions are reduced to two simple properties: It causes a remarkable new growth of fibroblasts, as is shown in Fig. 4, making them grow as they do in tissue cultures. Then it produces the formation of foreign body giant cells around the particles of the wax, and subsequently gives an infiltration of the tissue with the eosinophilic leucocytes. With the leprosinic acid the material which causes the new growth of the fibroblasts has been split off from the molecule of the leprosin, and the resulting fraction causes only the formation of the foreign body giant cells. This property is seen in its purest state with the crystalline solid alcohol, in which each giant cell is proportional in size and shape to a single crystal or a clump of crystals which may happen to lodge on a milk spot.

CONCLUSIONS

1. The waxes from the *B. leprae*, like those from tubercle bacilli, are remarkable stimulants of cells.

2. The crude wax separated from the *B. leprae* is a mixture of lipoids and other materials, and gives reactions that include the types of cells characteristic of the response to the tuberculo-polysaccharide, phosphatide, and wax.

3. The wax obtained from the purification of the lepra phosphatide shows similar cellular reactions but with a greater proportion of foreign body giant cells.

786 WAXES FROM MYCOBACTERIUM LEPRAE

4. Leprosin, though a glyceride, corresponds in its physical properties to the unsaponifiable material from the tubercle bacillus. It stimulates two strains of cells, fibroblasts and monocytes. The monocytes fuse into foreign body giant cells to engulf the wax.

5. The cellular reaction to the leprosinic acid and to the crystalline alcohols is of one type only, represented by the foreign body giant cell.

BIBLIOGRAPHY

- 1. Uyei, N., and Anderson, R. J., J. Biol. Chem., 1931-32, 94, 653.
- 2. Anderson, R. J., and Uyei, N., J. Biol. Chem., 1932, 97, 617.
- 3. Anderson, R. J., J. Biol. Chem., 1929, 83, 505; 85, 327; 85, 339.
- Sabin, F. R., Doan, C. A., and Forkner, C. E., J. Exp. Med., 1930, 52, suppl. 3, 1.
- Sabin, F. R., Smithburn, K. C., and Thomas, R. M., J. Exp. Med., 1935, 62, 751.
- 6. Smithburn, K. C., and Sabin, F. R., J. Exp. Med., 1935, 61, 771.

EXPLANATION OF PLATE 34

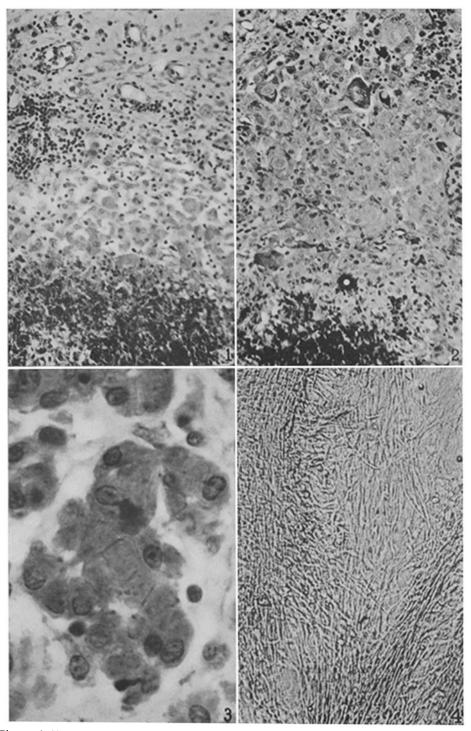
FIG. 1. Wall of an abscess in the omentum of Rabbit R 4188, which had received three intraperitoneal injections of 20 mg. each of the crude chloroformsoluble wax from the *B. leprae*. The wax was in the form of a colloidal suspension. The animal was killed 5 days after the third injection. There is a narrow band of monocytes and giant cells along the edge of the abscess. Masson stain to show the deep basophilia of the cytoplasm of the monocytes. $\times 210$.

FIG. 2. Wall of an abscess in the omentum of Rabbit R 4184, which had received three intraperitoneal injections of 20 mg. each of the wax obtained from the purification of the lepra phosphatide. The animal was killed 6 days after the third injection. There is a wide band of monocytes and giant cells along the edge of the abscess. Masson stain. $\times 250$.

FIG. 3. Sinuses of one of the retrosternal lymph nodes of the same animal as Fig. 1. It shows a pure and extensive reaction of monocytes and giant cells. Masson stain. \times 1,000.

FIG. 4. Film of the omentum of Rabbit R 2801, which had received two intraperitoneal injections of 20 mg. of leprosin in the form of a dry powder. The animal was killed 4 days after the second injection. The preparation was stained with neutral red and the photograph was taken while the cells were still living, and shows bands of fibroblasts. \times 150. THE JOURNAL OF EXPERIMENTAL MEDICINE VOL. 62

PLATE 34



Photographed by Louis Schmidt

(Sabin et al.: Waxes from Mycobacteriuml eprae)