



To Be or Not To Be Vaccinated: That Is a Question in Myasthenia Gravis

Qian Zhou, Ran Zhou, Haojun Yang and Huan Yang*

Department of Neurology, Xiangya Hospital, Central South University, Changsha, China

Myasthenia gravis (MG) is an autoimmune disease characterized by muscle weakness and abnormal fatigability due to the antibodies against postsynaptic receptors. Despite the individual discrepancy, patients with MG share common muscle weakness, autoimmune dysfunction, and immunosuppressive treatment, which predispose them to infections that can trigger or exacerbate MG. Vaccination, as a mainstay of prophylaxis, is a major management strategy. However, the past years have seen growth in vaccine hesitancy, owing to safety and efficacy concerns. Ironically, vaccines, serving as an essential and effective means of defense, may induce similar immune cross-reactivity to what they are meant to prevent. Herein, we outline the progress in vaccination, review the current status, and postulate the clinical association among MG, vaccination, and immunosuppression. We also address safety and efficacy concerns of vaccination in MG, in relation to COVID-19. Since only a handful of studies have reported vaccination in individuals with MG, we further review the current clinical studies and guidelines in rheumatic diseases. Overall, our reviews offer a reference to guide future vaccine clinical decision-making and improve the management of MG patients.

OPEN ACCESS

Edited by:

Li-Tung Huang, Kaohsiung Chang Gung Memorial Hospital, Taiwan

Reviewed by:

Hans De Haard, Argenx BVBA, Belgium Li Xiaoli, Shandong University, China Acary Oliveira, Federal University of São Paulo, Brazil

*Correspondence:

Huan Yang 403850@csu.edu.cn

Specialty section:

This article was submitted to Autoimmune and Autoinflammatory Disorders, a section of the journal Frontiers in Immunology

Received: 30 June 2021 Accepted: 01 September 2021 Published: 17 September 2021

Citation:

Zhou Q, Zhou R, Yang H and Yang H (2021) To Be or Not To Be Vaccinated: That Is a Question in Myasthenia Gravis. Front. Immunol. 12:733418. doi: 10.3389/fimmu.2021.733418 Keywords: myasthenia gravis, immunosuppression, neuromuscular disease, vaccines, infection prophylaxis, autoimmune, COVID-19, SARS-CoV-2

INTRODUCTION

Myasthenia gravis, an autoimmune antibody-mediated disease that affects the postsynaptic membrane at the neuromuscular junction, is characterized by muscle weakness and abnormal fatigability (1). Its worldwide prevalence is 150-250 cases per million individuals, with an annual incidence of approximately 8-10 cases per million person-years, although these greatly vary across

1

Abbreviations: MG, Myasthenia gravis; IST, immunosuppressive treatment; EBV, Epstein Barr virus; PRRs, patternrecognition receptors; APCs, antigen-presenting cells; DCs, Dendritic cells; TLR, toll-like receptor; AChR, acetylcholine receptor; MPLA, mono-phosphoryl lipid A; IFN-I, type-I interferon; AIIDs, autoimmune inflammatory diseases; Treg, regulatory T cells; Tfh, follicular Th; Th1, T helper type 1; PC, plasma cell; GCs, germinal centers; TECs, thymic epithelial cells; HPV, human papillomavirus; MUSK, muscle specific receptor tyrosine kinase; poly(I:C), polyinosinic-polycytidylic acid; ASIA, autoimmune/inflammatory syndrome induced by adjuvants; Abs, antibodies; dsRNA, double strands RNA; TSAs, tissue-specific antigens; CMG, childhood-onset myasthenia gravis; LA-JEV, live-attenuated Japanese encephalitis vaccination; SLE, systemic lupus erythematosus; WHO, World Health Organization; ILI, influenza-like illness; CS, corticosteroids; RTX, rituximab; PCV, Pneumococcal conjugate vaccine; PPSV, polysaccharide vaccine; ABA, abatacept; DMARDs, Diseasemodifying anti-rheumatic drugs; TNF α , Tumor necrosis factor α ; TCZ, Tocilizumab; RMDs, rheumatic and musculoskeletal diseases; HCs, healthy controls; AEs, adverse events.

MG subgroups (2). Despite differences in clinical and pathological manifestations between subgroups, all MG patients share muscle weakness, an autoimmune disease mechanism, and immunosuppressive treatment (IST), which predispose them to infectious disorders (3). Infection, as it had been proved in much research, is closely related to the initial event and exacerbation of MG.

Vaccination is a mainstay of preventing infection in such highrisk populations (4). However, natural infections can result in autoimmune disorders, posing a question as to whether vaccines containing pathogen antigens can also increase the risk of flare or exacerbate autoimmunity in susceptible individuals, in a similar fashion to infection (4). Anti-vaccination lobby groups have described a series of case reports of *de novo* initialization of autoimmune disease or flares of existing autoimmune disease after vaccine administration. Their fears have also been fueled by the insufficient immune responses in protecting MG patients with IST, especially during the pandemic.

These concerns, regarding the safety and efficacy of vaccination, have raised a fervent debate, exacerbated the hesitancy of vaccination, and posed a threat to public health. According to a previous study that investigated poor compliance among MG patients, it is evident that fear of general, non-myasthenic side effects (42.6%), the fear of a myasthenic exacerbation (31.5%), and refusal from their treating physicians (14.8%) are the main reasons for the observed hesitancy (5). A recent survey showed that experienced practitioners are more conservative in vaccine recommendation, despite existing guidelines (6). Little is known regarding the safety and efficacy of vaccination in MG patients. In this review, we will discuss infection- and vaccination-induced effects on the immune system, and address the safety and efficacy of vaccination in patients with myasthenia gravis. We also discuss the status of MG vaccination in relation to COVID-19 vaccines.

EFFECT OF INFECTION ON MG

Previous studies have shown that infections are closely associated with the onset of MG, and is the largest contributor of MG progression (30%) (3, 7, 8). Initial events triggering MG have been hypothesized inside the thymus, where thymoma accounts for 10% of MG cases and the remaining 90% have unknown causes, among which genetic factors are responsible for less than 50%. Infections act as a major external causal factor since previous reports have associated MG with various viral infections, including Hepatitis B and C, herpes simplex, HIV, West Nile virus, Zika virus, dengue virus, cytomegalovirus, parvovirus B19, human foamy virus, poliovirus, Epstein Barr virus (EBV), and SARS-CoV-2 (9-19). The thymus is a common target organ for infectious diseases that not only alter thymocyte development and export, but also affect the thymic microenvironmental compartment (20). However, MG symptoms are likely to occur long after a triggering infection, which makes it difficult to correlate with a particular infection.

Muscle weakness, IST, and immune dysfunction of MG patients shape the vulnerability to infections which in turn

account for MG deterioration. Infections have been identified as the most common cause for precipitating a life-threatening event according to a population-based study from Spain, and associated with 44.3% of emergency department visits and 39.7% of hospitalizations (7, 21). MG exacerbations caused by infections have not been linked to any specific microorganisms.

In attempts to explain the onset or exacerbation of MG after infection, numerous mechanisms have been proposed. Consequently, molecular mimicry, which refers to crossreactivity between microbial antigens and self-antigens when sharing similarities, has emerged as the most widely accepted mechanism (22). Other mechanisms about autoimmunity after infection include epitope spreading, bystander effect, release of cryptic epitopes, reactivation of memory T cells, activation of superantigens, direct inflammatory damage, formation of immune complexes, expression of MHC antigens on nonimmune cells, and patient's genetic predisposition to autoimmunity (23).

Thus, preventing such infections could protect genetically predisposed groups from developing MG. For MG patients, it is clinically important to avoid serious complications resulting from infections, hence the need for vaccines.

THE IMMUNOLOGY OF VACCINES

The Mechanism of Vaccine

Vaccination, which has rewritten human history since 1796 when Edward Jenner first introduced vaccinia, has become the most cost-effective means for combating infectious diseases. Notable examples of vaccine effectiveness include the eradication of smallpox and the decline in many lethal diseases. Its function depends on the interaction between innate and adaptive responses.

Dendritic cells (DCs), the most efficient antigen-presenting cells (APCs) in the immune system, are the key players in this unique process (24). Specifically, they detect antigens through innate receptors, known as pattern-recognition receptors (PRRs), and present antigens to T cells (24). After inducing a mixture of chemokines and cytokines, DCs transport them to the lymph nodes where activation of T and B cells, as well as production of antibodies, occurs (25). Notably, the specific innate signals received by DCs impact the magnitude and quality of the ensuing T- and B-cell responses, as well as the induction of memory cells (26).

CD4⁺ T cells, activated by antigens that are presented by HLA-class II alleles on APCs, stimulate and sustain B cells' response. With extreme diversity, HLA molecules are pivotal immune regulatory components encoded by the Major Histocompatibility complex, accounting for individual variation to the immune response against antigens. Those with "favorable" HLA alleles tend to raise optimal and protective antibody titers (27).

Vaccines have continued to evolve in the epoch of molecular biology and biotechnology, and can now be subdivided into 2 broad groups based on their components (28, 29). The first group comprises live attenuated vaccines, with weakened virulent properties, which simulate the kind of protective immunity induced in people who survive live infection. The others are non-living vaccines (**Figure 1**). Unlike live-attenuated vaccines, which are characterized by high immunogenicity and substantial protection but with a risk of virulence reversal, the second group of vaccines, especially subunit vaccines, exhibit higher safety and fewer side effects, which often impairs their immunogenicity, necessitating the use of adjuvants to improve their potency (30, 31).

Adjuvants, though often not particularly immunogenic by themselves, have been widely used to increase the magnitude and alter the quality of adaptive response, enhance immune responses to vaccine components, and prolong immunological memory. However, their cellular and molecular mechanisms of action remain unclear. Antigens, in combination with adjuvants, enable easier and more effective recognition of "non-self", enhance the innate immune system and provide a second signal for T cell activation. An ideal adjuvant acts locally at the site of injection to limit system effect, without causing severe side effects (25).

Numerous adjuvants have emerged in the past 100 years, including aluminum salt, virosomes, emulsion adjuvants. Today, many other toll-like receptor (TLR) agonists are still under clinical investigation, with only TLR4 ligand, mono-phosphoryl lipid A (MPLA), licensed (32).

Vaccines and Autoimmunity

Vaccines, though have saved numerous lives by preventing lethal infection, encountered public inquiry due to evidence of vaccineand adjuvant-induced autoimmunity using both animal models and human patients (31). As for vaccine-induced autoimmunity, certain pathogenic elements in the vaccine bearing a significant similarity with specific human proteins, can lead to immune crossreactivity and harm similar human proteins, based on molecular mimicry. Molecular mimicry is supposed to facilitate immune tolerance (33-35), but further study found that this tolerance could be attenuated by genetic susceptibility to autoimmunity and environmental factors, especially adjuvants (36). The presence of stimuli including lipopolysaccharides, altered glycosylation patterns, posttranslational modification as citrullination, and chemical modification as oxidation and glycation, can also result in breaking tolerance (37). Furthermore, "Autoimmune/Autoinflammatory Syndrome Induced by Adjuvants" (ASIA syndrome) which refers to a broad spectrum of reactions was proposed by Shoenfeld in 2010, and remains highly controversial entity (38, 39).

Adjuvants improve vaccines' efficacy through the innate immune system. Generally, pathogens can naturally generate their adjuvant effect through multiple PRRs, although most vaccines need the aid of adjuvants. Most adjuvants are considered immunostimulatory agents and ligands for PRRs, responsible for shaping the innate immune response and producing type-I interferon (IFN-I) which is critical not only in anti-viral mechanism, but also in the pathogenesis of autoimmune inflammatory diseases (AIIDs) (31). Furthermore, signaling *via* TLR3, TLR4, TLR7, TLR8, and TLR9 improves T helper cells type 1 (Th-1) immune responses, while signaling via TLR2 (along with TLR1 or TLR6) and TLR5 enhances Th2-type responses (40, 41). These responses have also been shown to influence regulatory T cells (Treg) and Th17 development, which is particularly relevant for AIIDs (42). Although Coffman et al. (31, 43) believed that this innate immune stimulation is shortlived, circumscribed, and does not trigger AIIDs due to insufficient autoreactivity, this hypothesis was refuted with the use of self-antigens or its analogs, and genetic predisposition. For example, the similarity between a peptide sequence of the influenza nucleoprotein A and an extracellular domain of Hypocretin receptor was found to result in narcolepsy after Pandemrix vaccination in the 2009 H1N1 pandemic. Interestingly, HLA-DQB *06:02 allele was found in over 90% of narcolepsy patients, while Canadian vaccinated population without such genetic component did not experience this disorder (36).

Animal Models

Investigation of viral pathogenesis, safety and efficacy of vaccines, and autoimmune mechanism require animal models which entitle researchers to apply translational research to predict human responses. Non-human primates, a traditional and fundamental model to test new vaccines in clinical trials, turn out to be unreliable, compared with murine which share the highest levels of heptapeptide with pathogens and humans (44). However, rodents, the most widely used models in both preclinical and mechanistic evaluation of vaccines, still have substantial discordance with regards to immune responses with humans. Important distinctions, including the expression and function of TLR, between rodent models and humans, have been found (40). Cross-species differences complicate the interpretation of animal-derived data on humans.

Mechanism of Vaccination and MG

Though numerous autoimmune case reports after vaccination lead to the peaking passion in the exploration of mechanism, little is known regarding the vaccines' role in triggering or exacerbating MG (Figure 2). MG, a prototypic but multifaceted autoimmune disease, involves a spectrum of antibodies (Abs) and a specific mechanism of action. Although the actual underlying mechanisms of MG are unclear, presence of ectopic germinal centers (GCs) and loss of central as well as peripheral tolerance have been discussed (1). High numbers of Th17 cells, follicular Th (Tfh) cells, and dysfunction of Tregs have been shown to promote autoantibody production from B cells and plasma cells (PCs), thereby exacerbating MG pathogenesis (45). Interestingly, the pattern of MG pathogenesis, to some extent, overlaps with the underlying mechanism of vaccine action. For example, Tfh cells are required for GCs, Abs, and long-lived PC responses, not only in MG but also in vaccine immunization (45, 46). Nonetheless, the link between the underlying mechanism of vaccination action and MG remains unclear.

Activation of the TLRs signaling pathways plays a crucial role in triggering and sustaining the inflammatory response and chronicity of MG. Accumulating evidence has indicated that

Types of vaccines	Advantages	Disadvantages	Used examples against pathogens	COVID-19 vaccines
Live-attenuated vaccines	Retain most of pathogen's immunogenicity; less revaccination; induce strong immune response and long-lasting protec- tion, even persisting for a whole life; no adjuvants required	Virulence reversal; unstable; difficult preservation; compli- cated production process; inappropriate for immunosuppres- sive patients	Measles, mumps, rubella (MMR); Rotavirus; Smallpox; Varicella; Yellow fever	COVI-VAC
Killed/ inactivated vaccines	Heat-stable; no virulence reversal; easy preserva- tion	More expensive; require multiple adminstrations; possible presence of unwanted contami- nants	Hepatitis A Influenza (injection); Polio (injection) Rabies	CoronaVac; BBIBP-CorV
Subunit vaccines	High safety and stability	High requirement for antigen; focus immune responses only against the relevant targets; complex preparation; lower immunogenicity and require adjuvants		NVX-CoV2373; EpiVacCorona ier
Viral Vector-Based vaccines viral vector gene viral vector pathogen	Safety; induce strong responses; lower cytotoxicity r gene	Potential risk for infection, chromosomal integration and oncogenesis; vary efficacy between different ethnic group	l Ebola	ChAox1-S-(A ZD1222)
DNA vaccines	Cheap, safe and relatively easy to design and manufac- ture; long-term stability; possibility of oral formulation	Poor immune responses; be administered with a gene gun; potential risk of genetic integation	No approved DNA vaccines for human; only in veterinary use: vaccine against West Nile Virus in horse, canine melanoma	GLS-5310; COVIGEN
RNA Vaccines	Safe and rapid develop- ment and production; more protein produced compared to naked pDNA; controllable due to transient nature; cannot modify the genetic information of the host	Easy degradation; require cold chain transportation	mRNA-1273; BNT162b2	mRNA-1273; BNT162b2
Virus-like particles	None infectivity; enable strong immunity due to dense epitope; induce cross immunity; Longer half-life in serum; stable quality	Require higher cost and technology	Human papillomavirus ; Hepatitis B virus; Hepatitis C virus	VBI-2902a

TLRs, mainly TLR2, TLR3, TLR4, TLR7, TLR8, and TLR9 in MG, coupled with aberrant expression or persistent triggering of these receptors, can result in self-sustained inflammation (47–52). For example, a previous study demonstrated that MG thymus with persistent poliovirus infection can synthesize similar quantities of plus and minus RNA strands and VP1 (a viral protein), and express more TLR4, indicating that chronic inflammation and dysregulation of the TLR4 pathways in MG thymus could be attributed to viral persistence (17). Moreover, research has revealed crosstalk between intrathymic EBV with TLR7 and TLR9, a phenomenon that consequently drives massive proinflammatory cytokines, chemokines, and IFN-I produced by thymic epithelial cells (TECs) and plasmacytoid

DCs in a genetic susceptibility context (53). INF-I, produced by TLR7 and other TLRs, may form an exacerbating vicious cycle by maintaining TLR7 expression (53). TLR pathways are also involved in vaccine responses, where they control the activation of adaptive immunity. Interestingly, vaccines may contain TLR ligands as adjuvants, or the primary antigens can be TLR ligands themselves, implying that vaccines can trigger and sustain this autoimmune activity. These results suggest that the TLR pathway is an intersection between MG pathogenesis and vaccine action.

MPLA, a licensed TLR4 adjuvant and a derivative of lipopolysaccharide, is far less toxic. Unlike TLR4 in controlling of both the MyD88 and TRIF signaling pathways, MPLA has a

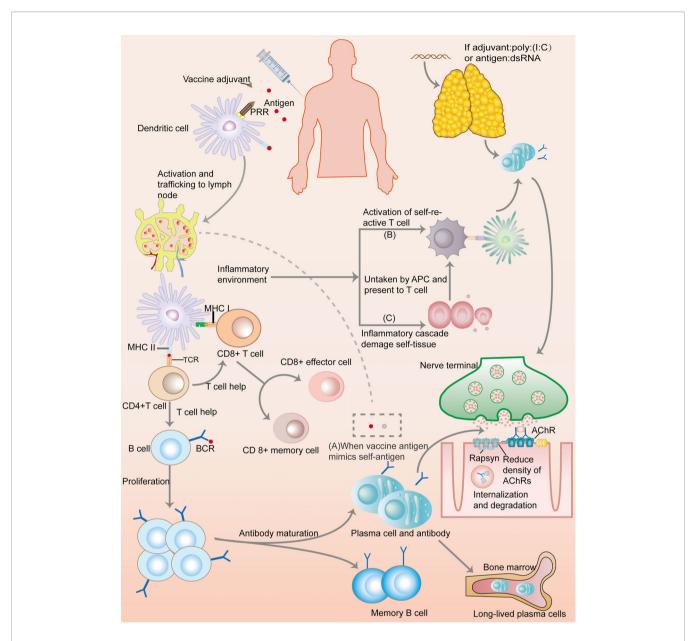


FIGURE 2 | Immunology of a conventional protein vaccine and the possible mechanisms of triggering myasthenia gravis. After a vaccine is injected, antigens would be taken up by dendritic cells (DCs). Combining with adjuvant, pattern recognition receptors (PRRs) deliver danger signal into the DCs, inducing DCs activation and transportation to lymph nodes where the presentation of vaccine antigen by MHC molecules would lead to the activation of T cells through combining to T cell receptor (TCR). With the help of CD4+T cells, B cells undergo activation and proliferation, leading to antibody maturation and the appearance of short-lived and long-lived plasma cells, as well as memory B cells. CD8+T cells also develop into CD8+effector cell and memory cells. This immunization processes create a special inflammatory environment which correlates with the mechanisms of autoimmune: (A) Molecular mimicry: If vaccine antigens share great similarity with self-antigens, autoimmune reaction may ensue because the immune response to the vaccine antigens can crossreact with self-antigens. (B) Bystander effect: immune responses to vaccine antigens can induce inflammatory signals, inadvertently resulting in the activation of self-reactive T cells, and thus the autoimmune processes. (C) Epitope spreading: inflammatory cascades cause tissue damage, and the presentation of self-antigens would trigger the activation of self-reactive T cells. Additionally, if a vaccine contains dsRNA or its analog, such as poly(I:C), it can trigger a thymic overexpression of IFN-β, leading to MG through overexpression of AChR, DCs autosensitization, abnormal recruitment of B cells, and GC formation.

bias toward TRIF signaling, and induces high IL-10 expression that could counterbalance the proinflammatory response (40). Currently, there is not report associating MPLA with MG.

However, Polyinosine-polycytidylic acid (poly(I:C)), a TLR3 agonist, can induce thymic changes and trigger MG symptoms

by mimicking double strands RNA (dsRNA), a viral nucleic acid associated with viral replication and antiviral mechanism (50). Poly(I:C) can specifically result in the overexpression of acetylcholine receptor α (α -AChR), but not other tissuespecific antigens (TSAs) in TECs. This process is mediated by

Vaccination in Myasthenia Gravis

TLR3, protein kinase R and IFN- β , etc., which are involved in antiviral responses, and observed in MG thymus. Further studies reveal IFN- β plays a central role in thymic events leading to specific overexpression of α -AChRs, TECs apoptosis, and DC autosensitization against AChR. It can cause overexpression of CXCL13 and CCL21 that are associated with GCs development, and induce the B cell-activating factor (BAFF) that favors autoreactive B cell survival (54). Moreover, poly(I:C) are proved to engage in thymic involution by disrupting T-cell development (55, 56). Thus, the injection of vaccines containing specific molecules, no matter from pathogen antigens or adjuvants, which mimics dsRNA and activate TLR3 pathway, can possibly lead to a specific autoimmune reaction against AChR.

THE SAFETY OF VACCINATION IN MG

Would Vaccination Trigger MG?

Previous case reports have shown that vaccines can cause MG occurrence, as evidenced by the relationship between MG onset with intravesical Bacillus Calmette-Guerin for bladder cancer and human papillomavirus (HPV) vaccination (57–59). Unfortunately, the mechanism is unclear and requires further investigation. Recent investigation has validated the massive commonality between HPV L1 epitopes and human proteins, i.e., the immune attacks against HPV L1 epitopes may cross-react with human proteins that share the epitopic peptide sequence (37). MG is also reportedly induced and exacerbated by Hepatitis B vaccination, although there is no molecular mimicry between HBsAg and nicotinic AChR (11).

A recent investigation, exploring the molecular and clinical relationship between childhood-onset myasthenia gravis (CMG) and live-attenuated Japanese encephalitis vaccination (LA-JEV), implicated vaccinations in the development of MG, after excluding genetic factors and viral infection (60). Exposing BALB/c mice to a group of Chinese planned immunization programs, researchers found LA-JEV can cause MG-like symptoms in the mice (60). Results later revealed that an 86% identity in a 7-amino-acid-long region was shared by both the human/murine AChR a-subunit (TWTYDGS) and RdRp (TWTYHGS) that is generated during the replication of LA-JEV in vivo (60). Based on these findings, it was evident that antibodies against RdRp can cross react with AChR. Notably, the LA-JEV vaccinates nearly 20-30 million children in China every year, but it is in the expanded immunization program in Japan and Taiwan, and not incorporated in the national immunization programs across Europe, North America, and Australia, which might explain the low incidence of CMG in these areas (60). Although this research presents strong evidence owing to the study design that combined retrospective and cohort nature, animal models, epidemiology, and molecular mechanism, the pathology caused by low antibody titer, as well as knowledge of the additional factors associated with immunological profile remains unclear, requiring further clarification.

The analysis of the potential link between vaccines and MG is puzzling. Currently, there exist no general criteria for diagnosing

vaccine-related MG, necessitating a case-by-case assessment (61). Consequently, the World Health Organization (WHO) has formulated four basic principles for assessing the adverse events (AEs) of vaccines, namely consistency, strength, specificity, and temporal relation (61, 62). Epidemiology and case reports are valuable clues for deciphering the relationship between vaccines and MG. However, there is a need to consider background incidence, genetic predisposition, environmental factors, and time relevance (clinical data in most clinical trials are collected within 6 weeks but recent research proposes one year after the last vaccination as a maximum theoretical risk interval) (63).

The rate of autoimmune reactions after vaccination is less than 0.01% of all vaccinations worldwide, possibly biased by underreporting. In most countries, reporting such vaccinerelated AEs is based on voluntary rather than obligation. Thus, clinicians and patients should be encouraged to report such correlations.

Though for most people, the benefits of immunization trump the potential risks since developing post-vaccination MG is rare, compared with the number of vaccines administered. Then who should be cautious of vaccination? Soriano et al. (64) purposed four groups at risk (1): Persons with prior-vaccination autoimmune phenomena; (2) Persons with a medical history of autoimmunity; (3) Persons with a history of allergic reactions, especially vaccine-related reactions; (4) Persons who are susceptible to autoimmunity, with a family history of autoimmune diseases, presence of autoantibodies, certain genetic profiles, etc.

Would Vaccination Exacerbate MG?

Influenza vaccines, have been assessed for their safety in MG patients (**Table 1**). In 2009, a population-based study was conducted in Canada, over a 14-year period based on data from healthcare databases. Notably, no association was found between the administration of influenza vaccines and the hospitalization of MG patients (65). However, several factors may have compromised the findings: limited generalizability due to clinicians' withholding vaccination from younger patients, exclusion of mild worsening of MG, and inability to measure the risk level of influenza vaccination across certain subgroups of MG. A similar conclusion was drawn in an Israel study in 2011, where researchers believed that seasonal influenza and H1N1 vaccines were safe for MG patients (5).

In 2017, a Korean study used a recall-based self-report questionnaire to demonstrate that the risk of MG symptom exacerbation following seasonal influenza vaccination was very low (1.5%), while the occurrence of influenza-like illness (ILI) was significantly associated with exacerbation of MG symptoms (40%) (68). Therefore, influenza infection is an apparent risk factor for MG progression, whereas vaccination seems to be an effective way to prevent the progression. However, data in this study did not show whether vaccination conferred protection due to imbalance baseline demography and the poor match between the vaccine strain and the circulating influenza viruses. Unlike the apparent bias presented in the former studies, the Korean study focused on a single influenza season when almost

Vaccine	Researchers	Study design	Subjects	Safety	Efficacy
Influenza vaccine	ZINMAN <i>et al.</i> (65)	Self- matched, case-series method	513 MG patients who were hospitalized within 42 weeks after vaccination; risk interval: first 6w; control interval: final 24w	Relative incidence of hospitalization in the risk interval was 0.84 (95% Cl: 0.65-1.09)	-
Influenza and H1N1 vaccines	AURIEL <i>et al.</i> (5)	Cohort study; using a questionnaire	74 MG patients (38 patients received influenza vaccine,24 H1N1 vaccine, and 20 both)	 In H1N1 vaccine group: the upper 95% corrected Cl for neurological side effect is 0.142, and the upper 99% corrected Cl is 0.167 In influenza vaccine group: the upper 95% corrected Cl is 0.013, and the upper 99% corrected Cl is 0.013. 	-
Diphtheria and tetanus	Csuka <i>et al.</i> (66)	Cohort study	279 SLE; 158 MG; 208 HCs	_	A-DIPHTH levels: HCs and MG (0.14[0.06-0.33] vs 0.10 [0.04-0.29], p=0.02) A-TET levels: HCs and MG (1.01[0.37-2.49] vs 1.00 [0.38-2.65], p=0.73)
Non- adjuvanted influenza vaccination (Mutagrip [®])	Tackenberg <i>et al.</i> (67)	A double- blind randomized controlled trial	62 MG patients (vaccine group; placebo group)	The difference between groups for AChR-ab-titer change 4.0% [-13.3%, 4.5%] (p=0.28) and for QMG change 0.00 [-0.17%, 0.00] (p=0.79). AE was comparable between groups.	-
Influenza vaccine (Trivalent unadjuvanted inactivated vaccines)	Seok <i>et al.</i> (68)	Case- controlled study, using a questionnaire	258 patients (112 men, 185 generalized MG, 133 received an influenza vaccination, and 121 had a common cold or ILI)	MG symptoms aggravated in 10 (40%) patients after ILI; only 2 (1.5%) aggravated after influenza vaccination.	-
Tetanus revaccination	Strijbos <i>et al.</i> (69)	A prospective, placebo- controlled study	65 MG patients (51 AChR MG, 6 MUSK MG, 9 LEMS); HC group: 20.	Titers of disease-specific autoantibodies remained unchanged after revaccination. Mean increase of QMG score was 1.08 point (P=0.01, 95%: 0.5-1.7) in AChR-MG group. Mean decrease of MG-ADL score was 0.86 point (95%CI: 1.6-0.2)	 Compared with HCs, AChR-MG have lower anti- TT IgG (pre, p=0.003, post, p=0.03) All have protective anti-TT IgG titer before TR. IM+ and IM- have lower GMT (IM-, p=0.02; IM+, p<0.01); only IM+ have lower GMT 4w after TR (p<0.01) Increase factor of anti-TT titer after TR were comparable.
Influenza vaccine (contains strains H1N1 pdm09, H3N2 and B/ Brisbane/060/ 08)	Strijbos <i>et al.</i> (70)	A prospective, double-blind, randomized, placebo- controlled study	47 AChR MG patients; 47 HCs	No change in AChR-ab titer was observed 4w after influenza vaccination. Total scores of the MGC, QMG, MG-ADL outcome measures were the same before and after vaccination between MG vaccination group and placebo group.	 (1) Seroprotective titers (HI>1:40) was achieved in (3) Seroprotective titers (HI>1:40) was achieved in (4) Seroprotective titers for H3N2, 95.7% vs 97.9% (5) rH1N1, and 46.8% vs 51% for the B-strain. (5) Seroprotective titer for these strains was reached (40.4% of MG and in 51% of HCs. (H3N2, p=0.2; (1) H1N1, p=0.7, B-strain, p=0.9) (2) No significant effect on serological response (2) between IM- and IM+ (H3N2, p=0.2; H1N1, p=0.2; and (3) B-strain, p=0.9)

TABLE 1 | Summary of epidemiologic studies on immune response in MG vaccinated patients.

MG, myasthenia gravis; AChR, acetylcholine receptor; MUSK, muscle specific receptor tyrosine kinase; w, week; HC, healthy control; Cl, confidence interval; AE, adverse events; ILI, influenza-like illness; LEMS, Lambert-Eaton myasthenic syndrome; A-DIPHTH, diphtheria-antitoxin-IgG; A-TET, tetanus-anti-toxoid-IgG; IM, immunosuppressant; TT, tetanus toxoid; TR, tetanus revaccination; GMT, Geomean titers

all influenza vaccines used in Korea were trivalent unadjuvanted inactivated vaccines, and it used stratified analysis to minimize biases.

In 2017, the first double-blind, randomized, placebocontrolled trial was reported, providing class 1b evidence that influenza vaccination (Mutagrip[®]) had a positive risk-benefit ratio in MG patients (67). Similar studies later revealed that influenza vaccines (Vaxigrip, RVG 22306) and tetanus revaccination were safe for MG patients, as evidenced by lack of clinical or immunological exacerbation in clinical outcome scores and elevated AChR antibody titers (69, 70).

AChR-ab titer, whose change in the individual is a useful indicator of disease activity, did not see a clinically relevant increase in influenza and tetanus vaccination. Interestingly, one study even observed a pronounced AChR-ab reduction in a subgroup of patients without immunosuppressive therapy, while 3 of 4 examined anti-influenza antibody immunoglobulin classes had a more potent increase, casting doubt on whether the

influenza vaccine could inhibit an AChR-ab boost and suppress autoimmune effective B and T cells (67). Additionally, clinical outcome scores, a useful tool to evaluate the disease stability and life quality, did not show an obvious change. QMG scores in one study suggested some worsening of MG with a statistically significant increase of 1 point after 4 weeks, which is far less than the 2 or 3 points accepted as minimal clinically relevant difference, and MG-ADL showed improvement, especially 12 weeks after revaccination (69). However, the use of clinical scores as primary outcome measures have their limitation owing to the large variations in individual levels between pre- and post- revaccination.

Notably, unblinded vaccination of MG patients resulted in more side effects and subjective complaints, relative to blinded subjects given a placebo injection, possibly due to the prejudice among MG patients who considered vaccination to be harmful (69).

These studies did not include patients with severe or unstable MG; hence no meaningful conclusion could be drawn from the groups. In fact, most studies were conducted in the stable and quiescent stage to avoid any misunderstanding associated with safety issues, because some AEs attributed to vaccination actually occur due to underlying disease conditions (71).

Although MG could be theoretically exacerbated by vaccine immunogen contents and adjuvants, results from previous clinical trials indicate that influenza and tetanus vaccines are safe for patients with stable diseases. Since randomized, placebocontrolled trials are the best approaches to assess the potential causal relationship between an AE and vaccination, we suggest that more trials of this kind should be performed to enable accurate evaluation of the safety of other vaccines in MG patients.

THE EFFICACY OF VACCINATION IN MG

Although vaccination represents a feasible option for managing a substantial number of infections, some previous studies have indicated that the seroprotection rate is significantly lower in immunocompromised than in immunocompetent patients, casting doubt on the efficacy of vaccination (72, 73). MG, a paradigm of autoimmune disease, requires symptomatic treatment and IST. Studies and guidelines in rheumatic diseases indicate that most live vaccines are contraindicated in patients with IST, while inactivated vaccines generally exhibit a similar safety pattern in immunosuppressed and immunocompetent patients, although the immune response to vaccination can be impaired or even absent with regards to magnitude, breadth, and persistence (74, 75). However, data on the efficacy of vaccines in MG remains scarce.

Efficacy of vaccines refers to suppression of disease incidence among vaccinated subjects, relative to unvaccinated ones, while immunogenicity is the ability of a vaccine to elicit an immune response in a vaccinated individual (76). Research on vaccine efficacy has always been hampered and often used immunogenicity as a proxy, owing to the requirement of complex study designs, large cohorts, and long follow-up periods (77). Besides, the level of protection afforded by vaccination is affected by many factors, including immunosuppressants, age, HLA alleles, disease status, vaccine types, doses, and schedule.

A previous prospective study investigating the humoral immune response to tetanus revaccination in patients with MG or Lambert-Eaton myasthenic syndrome showed that IST slightly reduced pre- and post-tetanus antibody titers, but a significant humoral response could still be evoked with a median 6-fold increase factor of the tetanus antibody titer (69). These findings corroborated those from a prior study which found no significant differences in antibody responses and immune protection against diphtheria and tetanus in patients with systemic lupus erythematosus (SLE), and MG, relative to healthy subjects (66). Another study evaluating the efficacy and safety of influenza vaccines in MG patients in 2019 reported that a post-vaccination seroprotective titer for three strains of seasonal influenza vaccine was reached in 40.4% and 51% of AChR MG subjects and healthy controls, respectively (70). In these studies, IST comprised prednisolone, azathioprine, mycophenolic acid, and cyclosporine, as well as a combination of IST. Although IS therapy or thymectomy status did not significantly influence post-vaccination geomean titers, these studies did not measure the specific effects of a single drug, due to the small size of treatment subgroups and frequent combination of IST.

Because of the limited data on the efficacy of vaccines in MG, we analyzed results from studies in other AIIDs and concluded whether IST diminishes the vaccine response depends on the immunosuppressive mechanism and the immune profile of vaccines. AIIDs patients with IST tend to exhibit both similar and lower responses to vaccines compared with healthy individuals. It is ironic that those who most need protection due to their increased infection risk, least benefit from vaccination (78). Our findings indicate corticosteroids has a contradictory effect on the development of protective antibodies after vaccination (79, 80), while rituximab (RTX), a B-lymphocyte-depleting monoclonal antibody, severely inhibits immune response, which is partially restored 6-10 months after RTX administration (72, 73).

Pneumococcal polysaccharide vaccine (PPSV) consists almost exclusively of capsular polysaccharides, which provokes a minimal T cell-mediated antibody production in B cells, while Pneumococcal conjugate vaccine (PCV), conjugating to the diphtheria toxoid CRM197, elicits a robust T cell-dependent immune response (77). Thus, immune responses should be more profound after PCV than after PPSV. However, the effect is exactly contrariwise in patients with IST (77). It is hypothesized that IST mainly compromises cellular immunity, reducing the response rate more severely in PCV than PPSV (77). However, abatacept (ABA), a selective T-cell costimulation modulator, can impair PPSV serologic responses (81). This can be explained by the inhibition of B-cell differentiation which requires T-cell help, and polysaccharide antigens cannot be considered completely T-cell independent because they are enhanced by T-cell help.

A latest meta-analysis proved AIID patients with IST presented impaired serologic response, compared to healthy

individuals and patients without IST. Patients using anti-tumor necrosis factor α (TNF α) maintained a more favorable immune response to pneumococcal vaccination, compared with those with other IST (77). Likewise, Hua and Kapetanovic reported PPV-23 responses were normal in anti-TNFa, but reduced in Methotrexate (MTX) (82, 83). Disease-modifying anti-rheumatic drugs (DMARDs) exert their effect by blocking of clonal expansion of effector T- and B-cells, while anti-TNFa specifically interferes with the immune system by reducing migration of dendritic cells, inhibiting T cell activation, and reducing memory cell survival (77). This may account for the less negative effect on the immune response with anti-TNF α than with DMARDs. IL-6 also plays a vital role in B cells differentiating into plasma cells. Tocilizumab (TCZ), an IL-6 inhibitor that should hamper antibodies, did not impact response compared with healthy controls (HCs) (84). Thus, some researchers deduce the immune response to vaccination is not influenced in the same way by biologic agents. Treatment target on B cells (RTX) or T cells (ABA) may exert deleterious impact, whereas anti-cytokine therapies (TCZ and anti-TNF α) could maintain humoral responses to vaccines (85).

However, these results, mainly from rheumatoid arthritis, contradict in Crohn's disease, in which anti-TNF α also severely lowers the antibody titer after pneumococcal vaccination, posing a question of whether results attained from other diseases can apply in MG (86).

Patients exposed to different ISTs are expected to exhibit varied responses to different vaccines, due to the different underlying mechanisms of immunosuppressive drugs and variable immunological patterns of vaccines. Efficacy of vaccination in MG patients with different ISTs requires further clarification, since results obtained from other AIIDs do not directly apply to it. High-quality clinical trials are required to affirm vaccine efficacy in MG patients treated with different IST.

Age of vaccination is another factor that affects vaccine efficacy (87). For example, previous studies have shown that it is somewhat difficult to induce strong immune responses in the first year of life or in older periods, particularly among individuals older than 75 years (66, 68). Previous evidence has also indicated that growing older compromises protection after vaccination (66, 68). Moreover, vaccine doses and administrative schedules also impact protection, since immunocompromised patients often require larger doses or frequent administration (77). Thus, better vaccine-mediated protection can be obtained before the initiation of immunosuppressive treatment.

RISK-BENEFIT ASSESSMENT OF VACCINATION IN MG

Though previously mentioned research on vaccination in MG patients is scarce and mainly about influenza vaccines, nearly all the evidence supports vaccine-related worsening of MG is rare (5, 65, 67, 68, 70). The antibody response in MG is not different from that in healthy subjects, even in those with IST except RTX (70). Study evaluating the cause of death in Swedish MG patients

reveals influenza/pneumonia is a striking contributor (88). A 10year longitudinal study found a significant 48% reduction in mortality and a 27% reduction in hospital admissions after influenza vaccination in AIID patients (89). Thus, most MG specialists believe the benefits of vaccination outweigh any small risks in possible transient MG symptoms exacerbation (90). Guidelines also recommend influenza and pneumococcal vaccine for AIID patients, which preferably be administered during quiescent disease (91). Patients before Eculizumab treatment should consider be immunized with meningococcal vaccines (92). Discussion with doctors when considering a vaccine is necessary to assess the risk-benefit of vaccination (90, 91).

COVID-19 VACCINES IN MG PATIENTS

SARS-CoV-2 Infection and MG

COVID-19 was declared as a pandemic by WHO on March 11th, 2020. To date, accumulating evidence has confirmed that SARS-CoV-2 is linked to MG. 10 case reports of new-onset MG following COVID-19 have been analyzed with the following features: mean age 51 years, male gender (6), time interval between COVID-19 and MG (5-56 days), generalized (7), bulbar and/or ocular symptoms (5), anti-AChR antibodies (9) and anti-muscle specific kinase antibodies (anti-MUSK) (2) (93, 94).

Many plausible mechanisms have arisen to explain such causal relations. First, COVID-19 may be a disease of the nicotinic cholinergic system (95). Sequences on the SARS-CoV-2 proteins, similar to neurotoxin, can bind to AChRs and block the function of acetylcholine. Second, SARS-CoV-2 peptide CFLGYFCTCYFGLF aligns neuronal acetylcholine receptor subunit alpha-2 with 7 residue matches, affirming that antibodies against SARS-CoV-2 can crossreact with the human tissue, thereby causing autoimmunity (96). However, these studies did not analyze conformational or non-linear epitope (97, 98). Third, researchers have also speculated that SARS-CoV-2 can unmask a previously non-symptomatic MG or COVID-19 patients incidentally suffer an overlapping occurrence of MG (99). Furthermore, SARS-CoV-2 produces a proinflammatory milieu and cytokine storm, resulting in immune dysregulation and disrupted self-tolerance. IL-6, which is associated with a higher mortality rate in COVID-19 patients, also correlates with MG progression (100). Lastly, drugs like hydroxychloroquine sulfate and azithromycin during treatment can also trigger MG.

Moreover, mounting case reports show COVID-19 links to MG progression. Viral infection is perceived to trigger autoimmunity through augmentation of T cell signaling, thereby contributing to the proinflammatory environment, triggering hyper-reactive antiviral immune responses, epitope spreading, and effects of fever on the neuromuscular junction (3, 101).

The point is of interest since often, the autoimmunity after infection somewhat corresponds to the autoimmunity after vaccination. The aforementioned occurrence and deterioration of MG after COVID-19 infection implies that the protective antiviral antibody immune response can become a pathogenic attack against the human organism, while the adjuvanted anti-SARS-CoV-2 vaccines with higher immunogenicity can also elicit autoimmune response compared to SARS-CoV-2 infection.

SARS-CoV-2 Vaccines Hesitation

Vaccines, serving as a cornerstone in mastering the COVID-19 pandemic and achieving herd immunity, is rapidly evolving. Unlike traditional vaccines whose development pipeline span several years, sometimes even 15 before approval and distribution, SARS-CoV-2 vaccines development has been revolutionized, with 18 vaccines in Phase III clinical trial and 5 ending Phase III with positive results within 2 years (102). Consequently, such rapid speed to attain an insufficiently vetted vaccine increases the risk of trading freedom from COVID-19 to an autoimmune assault (97). Unfortunately, the risk of crossreactivity increases, since the current preclinical tests cannot measure it (44, 103). Although interim Phase III safety data from some vaccine trials have been largely positive, these studies failed to measure rare adverse effects due to a limited number of participants coupled with a lack of long-term efficacy because of trial results obtained from the last few months. It could be hard to identify some AEs of such vaccines produced utilizing new technologies. Moreover, Phase III trials currently underway mainly targetted on a healthy population.

An online survey on people's acceptance of COVID-19 vaccines showed that 54.9% of patients with rheumatic and musculoskeletal diseases (RMDs) were willing to get vaccinated, compared to the rate of HCs, 82.3%. Their refusal was mostly disease-linked, and reasons included fear of AEs and disease worsening (104).

Safety of SARS-CoV-2 Vaccines in MG

To date, several types of COVID-19 vaccines have undergone clinical trials and some have been approved for emergency use. Among them, RNA-based vaccines, including Pfizer/BioNTech (BNT162b2) and Moderna (mRNA-1273) mRNA, which were the earliest to be approved, serve as both immunogen and adjuvant, and have generated contrasting effects on innate immune responses (105, 106). Specifically, they are considered safe, without infectivity and integrating vector. With possible modification, they can be effective by increasing mRNA translation, and maintain the production of IFN-I and proinflammatory cytokines, which are desirable in countering the virus (106). On the other hand, the adjuvanticity of these vaccines is based on TLR 3,7,8,9 agonists, distinct from previous vaccines, and is a common pathogenic mechanism in autoimmunity (107). Researchers believe that some modifications, including the incorporation of modified nucleosides, can increase vaccine efficacy but reduce innate immune activation (106).

Following the strict guidelines from WHO to assess the causality of adverse events following immunization after COVID-19, 2 newonset MG cases were identified after the second dose of BNT162b2 vaccine, one being severe (107). A case of MG crisis after the second dose of COVID-19 vaccine has also been reported (108). The newonset and flare of MG all occurred within 1 week after the second dose. Noteworthily, before vaccination, this MG patient with a 5-year course still maintained on prednisone 7.5-milligram tablet daily and pyridostigmine 60-milligram tablet six times daily, which meant he didn't have MG remission. Though steroids can reduce cytokine expression, lymphocyte differentiation, and proliferation, it is possible that this patient was not medically optimized and still developed a cytokine storm (108).

There is few research hitherto about MG course after COVID-19 vaccination. Several studies investigating the safety of SARS-CoV-2 mRNA vaccines in patients with stable RMDs revealed minor side effects and no distinct impact on RMD activity (109, 110). A recent multicenter study reported no evidence of significant disease flares, but the occurrence of herpes zoster after BNT162b2 mRNA vaccination in subjects with low and even no IST is worth investigating (80).

Efficacy of SARS-CoV-2 Vaccines in MG

Research investigating the efficacy of SAR-CoV-2 vaccines in MG patients is scarce. Extrapolating from studies in RMD, rituximab, a B-cell depleting agent, is significantly associated with a higher seronegative rate (80, 111, 112). Thus, most guidelines suggest patients on RTX should be vaccinated either one month before initiation of the therapeutic scheme or 6-8 months after the RTX infusion (113). However, previous reports showed some patients with RMD on RTX therapy still developed adequate titers of antibodies against SARS-CoV-2 despite having undetectable B cells (114, 115). B cells are suspected to play a comparatively lesser role in the clearance of virions. Preclinical and early results from human trials showed that human after mRNA vaccination had Th-1 skewed T cell immune responses with RBD-specific CD8+ and CD4+ T cells. Those on B cell-depleted treatment with a deficient humoral response may still be protected by the cellular response (116). Moreover, those who had adequate antibodies used lower doses of RTX or just initiated this therapy (115).

Patients on the scheme of mycophenolate mofetil or abatacept exhibited significantly lower SARS-CoV-2 antibody titers (80, 111, 112). The blunted response was also associated with glucocorticoids, even if its doses were low (6.2mg/day), which precluded the consideration of dose-dependent effect (80). MTX, as monotherapy or in combination with other drugs except abatacept or rituximab, elicited a slightly reduced response (80). Older age is another factor contributing to the reduced immunogenicity (80).

From the standpoint of particular autoimmunity, rheumatoid arthritis, antineutrophil cytoplasmic antibody-associated vasculitis, and idiopathic inflammatory myositis are at risk of blunted response to the vaccine compared with other diseases (80). Further studies are needed to determine the factors, including drugs, doses, duration, timing of IST that influence the antibody responses to COVID-19 in MG patients.

Risk-Benefit Assessment and Should MG Patients Be Vaccinated?

Today, MG patients with COVID-19 infections experience a highly variable course and outcome. The percentage of patients who need hospitalization and/or experienced MG exacerbation varies across studies (117). MG patients might face a higher risk of severe outcome, especially those with respiratory muscle

weakness, older age, other and other medical or neurologic diseases (118). Data from a global, physician-reported registry of 91 MG with COVID-19 infection presented 63(69%) demand for hospitalization, 36 (40%) requiring rescue therapy (e.g., IVIG, PE, or steroids) due to disease worsening or crisis of MG, 39 (43%) complete recovery or discharging home, and 22 (24%) death (119). These figures are valuable because of analyzing a number of patients, but they did not consider international variations in the infection course. Unexpectedly, studies in the French and Polish cohorts showed COVID-19 had limited effect on most MG patients, while high myasthenia gravis foundation of America (MGFA) class (≥IV) before COVID-19 was associated with severe COVID-19 (117, 120). IST did not have a significant impact on infection risk (120, 121). Another important aspect influencing the infection course in MG patients is international differences in the number of vaccinated populations as those who get vaccinated will form a "cocoon effect" to protect those who cannot be vaccinated (117).

Owing to a lack of clinical vaccine trials in patients in MG and the inconsistency between RMD and MG, there is no direct evidence about COVID-19 vaccine safety and efficacy in MG patients. Recently, MGFA supports the potential benefits of SARS-CoV-2 mRNA vaccines outweigh the risks, and recommend patients with IST to discuss getting an additional mRNA vaccine dose with their treating providers (122). For healthy subjects without the conditions mentioned by Soriano et al. (64), it is unlikely to develop new-onset MG after vaccination. For MG patients who must be vaccinated, they had better reach minimal manifestation stage or at least disease quiescent stage.

To ensure the uttermost safety and best efficacy, patients should consult their clinicians to discuss the following: their attitudes, intent, safety and efficacy concerns about vaccination, local incidence of COVID-19, individual circumstances (e.g., disease activity, medications, comorbidities) (123). Risk and benefit assessment should be carried out on a case-by-case basis. The usual post-vaccination observation time that lasts 15-30 minutes does not fully capture the AEs that may appear 1-2 weeks after vaccination (108). Both patients and clinicians should carefully monitor AEs since those who had previously been COVID-19 infected might experience AEs after first doses, and who had not been infected could get AEs after the second dose (124).

Which Vaccine Is Preferable in MG Patients?

All vaccines target at least on the epitopes of S protein or its RBD sequence that either vectored or presented in different ways

REFERENCES

- Gilhus NE, Tzartos S, Evoli A, Palace J, Burns TM, Verschuuren J. Myasthenia Gravis. Nat Rev Dis Primers (2019) 5(1):30. doi: 10.1038/ s41572-019-0079-y
- Carr AS, Cardwell CR, McCarron PO, McConville J. A Systematic Review of Population Based Epidemiological Studies in Myasthenia Gravis. *BMC Neurol* (2010) 10:46. doi: 10.1186/1471-2377-10-46

(125). Right now, with a lack of evidence, there is no preference for one COVID-19 vaccine over another. Firstly, it will take a long time before industrial policies and national political issues allow such comparative assessment of these vaccines and set international standards. Secondly, geographic contexts, economic conditions, supply, storage, and schedule of administration are also necessary elements to determine the choice. The evaluation will continue. Future cohorts to monitor vaccine effectiveness and safety among individuals with rare conditions will guide preferred orientation according to age, immune and medical status, etc.

DISCUSSION AND PERSPECTIVES

The relationship between humans and vaccines is no doubt intricate and complex, with new layers still unraveling. Although numerous vaccines have been developed, totally transforming human life, their underlying mechanism of action, as well as the association with myasthenia gravis remain unknown, necessitating further studies. Theoretically, vaccines could trigger and enhance the progression of MG, in a similar fashion to infection, and immunosuppressive treatment may impair immune response, but recent experimental results have demonstrated the safety and efficacy of influenza and tetanus vaccines. Molecular mimicry is a commonplace topic, a validated reality rather than a fantasy. Thus, to select peptide epitopes for safe vaccine design, analyzing already proven immunoreactive epitopes and filtering out the peptide with cross-reactive potential becomes necessary.

Since the direct evidence about COVID-19 vaccines in MG patients is limited, more data is required to validate their safety and efficacy. Future studies should not only focus on the molecular aspects, but also aim at providing more clinical evidence to enable MG patients and their physicians to make decisions.

AUTHOR CONTRIBUTIONS

HY developed the idea, participated in the study design and critical revision of the manuscript. QZ did the literature research, framed the outline, and wrote the manuscript. RZ gave helpful advice, and critically revised the draft. HJY gave helpful advice. All authors contributed to the article and approved the submitted version.

- Gilhus NE, Romi F, Hong Y, Skeie GO. Myasthenia Gravis and Infectious Disease. J Neurol (2018) 265(6):1251–8. doi: 10.1007/s00415-018-8751-9
- De Martino M, Chiappini E, Galli L. Vaccines and Autoimmunity. Int J Immunopathol Pharmacol (2013) 26(2):283-90. doi: 10.1177/ 039463201302600201
- Auriel E, Regev K, Dori A, Karni A. Safety of Influenza and H1N1 Vaccinations in Patients with Myasthenia Gravis, and Patient Compliance. *Muscle Nerve* (2011) 43(6):893–4. doi: 10.1002/mus.22077

- Roy B, Litchman T, Torabi T, Nowak RJ. The Influenza Vaccination in Autoimmune Neuromuscular Diseases: A Survey of Current Practices and Perceptions. *Muscle Nerve* (2021) 63(6):918–23. doi: 10.1002/mus.27224
- Gummi RR, Kukulka NA, Deroche CB, Govindarajan R. Factors Associated With Acute Exacerbations of Myasthenia Gravis. *Muscle Nerve* (2019) 60 (6):693–9. doi: 10.1002/mus.26689
- Kassardjian CD, Widdifield J, Paterson JM, Kopp A, Nagamuthu C, Barnett C, et al. Serious Infections in Patients With Myasthenia Gravis: Population-Based Cohort Study. *Eur J Neurol* (2020) 27(4):702–8. doi: 10.1111/ ene.14153
- Eddy S, Wim R, Peter VE, Tanja R, Jan T, Werner VS. Myasthenia Gravis: Another Autoimmune Disease Associated With Hepatitis C Virus Infection. *Dig Dis Sci* (1999) 44(1):186–9. doi: 10.1023/a:1026683007277
- Hung WL, Lin YH, Wang PY, Chang MH. HIV-Associated Myasthenia Gravis and Impacts of HAART: One Case Report and a Brief Review. *Clin Neurol Neurosurg* (2011) 113(8):672–4. doi: 10.1016/j.clineuro.2011.03.019
- Stübgen JP. Neuromuscular Disorders Associated With Hepatitis B Vaccination. J Neurol Sci (2010) 292(1-2):1–4. doi: 10.1016/j.jns.2010.02.016
- 12. Meyer A, Levy Y. Geoepidemiology of Myasthenia Gravis [Corrected]. Autoimmun Rev (2010) 9(5):A383–6. doi: 10.1016/j.autrev.2009.11.011
- Molko N, Simon O, Guyon D, Biron A, Dupont-Rouzeyrol M, Gourinat AC. Zika Virus Infection and Myasthenia Gravis: Report of 2 Cases. *Neurology* (2017) 88(11):1097–8. doi: 10.1212/wnl.000000000003697
- Liu JJ, Chiu CY, Chen YC, Wu HC. Molecular Mimicry of Human Endothelial Cell Antigen by Autoantibodies to Nonstructural Protein 1 of Dengue Virus. *J Biol Chem* (2011) 286(11):9726–36. doi: 10.1074/jbc.M110.170993
- Restivo DA, Centonze D, Alesina A, Marchese-Ragona R. Myasthenia Gravis Associated With SARS-CoV-2 Infection. Ann Intern Med (2020) 173 (12):1027–8. doi: 10.7326/l20-0845
- Cavalcante P, Cufi P, Mantegazza R, Berrih-Aknin S, Bernasconi P, Le Panse R. Etiology of Myasthenia Gravis: Innate Immunity Signature in Pathological Thymus. *Autoimmun Rev* (2013) 12(9):863–74. doi: 10.1016/ j.autrev.2013.03.010
- Cavalcante P, Barberis M, Cannone M, Baggi F, Antozzi C, Maggi L, et al. Detection of Poliovirus-Infected Macrophages in Thymus of Patients With Myasthenia Gravis. *Neurology* (2010) 74(14):1118–26. doi: 10.1212/ WNL.0b013e3181d7d884
- Saïb A, Canivet M, Giron ML, Bolgert F, Valla J, Lagaye S, et al. Human Foamy Virus Infection in Myasthenia Gravis. *Lancet* (1994) 343(8898):666. doi: 10.1016/s0140-6736(94)92657-3
- Gong L, Li Y, Li X, Tu Q, Mou X, Wang S, et al. Detection of Human Parvovirus B19 Infection in the Thymus of Patients With Thymic Hyperplasia-Associated Myasthenia Gravis. *Clin Microbiol Infect* (2019) 25 (1):109.e7-.e12. doi: 10.1016/j.cmi.2018.03.036
- 20. Savino W. The Thymus Is a Common Target Organ in Infectious Diseases. *PloS Pathog* (2006) 2(6):e62. doi: 10.1371/journal.ppat.0020062
- Ramos-Fransi A, Rojas-García R, Segovia S, Márquez-Infante C, Pardo J, Coll-Cantí J, et al. Myasthenia Gravis: Descriptive Analysis of Life-Threatening Events in a Recent Nationwide Registry. *Eur J Neurol* (2015) 22(7):1056–61. doi: 10.1111/ene.12703
- Hussein HM, Rahal EA. The Role of Viral Infections in the Development of Autoimmune Diseases. Crit Rev Microbiol (2019) 45(4):394–412. doi: 10.1080/1040841x.2019.1614904
- Kivity S, Agmon-Levin N, Blank M, Shoenfeld Y. Infections and Autoimmunity–Friends or Foes? *Trends Immunol* (2009) 30(8):409–14. doi: 10.1016/j.it.2009.05.005
- 24. Akira S, Uematsu S, Takeuchi O. Pathogen Recognition and Innate Immunity. Cell (2006) 124(4):783-801. doi: 10.1016/j.cell.2006.02.015
- van der Laan JW, Gould S, Tanir JY. Safety of Vaccine Adjuvants: Focus on Autoimmunity. *Vaccine* (2015) 33(13):1507–14. doi: 10.1016/j.vaccine. 2015.01.073
- Nguyen XH, Saoudi A, Liblau RS. Vaccine-Associated Inflammatory Diseases of the Central Nervous System: From Signals to Causation. *Curr* Opin Neurol (2016) 29(3):362–71. doi: 10.1097/wco.000000000000318
- Ragone C, Meola S, Fiorillo PC, Penta R, Auriemma L, Tornesello ML, et al. HLA Does Not Impact on Short-Medium-Term Antibody Response to Preventive Anti-SARS-Cov-2 Vaccine. *Front Immunol* (2021) 12:734689. doi: 10.3389/fimmu.2021.734689

- Del Giudice G, Pizza M, Rappuoli R. Molecular Basis of Vaccination. *Mol Aspects Med* (1998) 19(1):1–70. doi: 10.1016/s0098-2997(98)00002-8
- Pulendran B, Ahmed R. Immunological Mechanisms of Vaccination. Nat Immunol (2011) 12(6):509–17. doi: 10.1038/ni.2039
- Pellegrino P, Clementi E, Radice S. On Vaccine's Adjuvants and Autoimmunity: Current Evidence and Future Perspectives. Autoimmun Rev (2015) 14(10):880-8. doi: 10.1016/j.autrev.2015.05.014
- Guimarães LE, Baker B, Perricone C, Shoenfeld Y. Vaccines, Adjuvants and Autoimmunity. *Pharmacol Res* (2015) 100:190–209. doi: 10.1016/ j.phrs.2015.08.003
- Reed SG, Tomai M, Gale MJJr. New Horizons in Adjuvants for Vaccine Development. Curr Opin Immunol (2020) 65:97–101. doi: 10.1016/ j.coi.2020.08.008
- Kanduc D. Quantifying the Possible Cross-Reactivity Risk of an HPV16 Vaccine. J Exp Ther Oncol (2009) 8(1):65–76.
- Trost B, Lucchese G, Stufano A, Bickis M, Kusalik A, Kanduc D. No Human Protein Is Exempt From Bacterial Motifs, Not Even One. *Self Nonself* (2010) 1(4):328–34. doi: 10.4161/self.1.4.13315
- Kanduc D. Peptide Cross-Reactivity: The Original Sin of Vaccines. Front Biosci (Schol Ed) (2012) 4:1393–401. doi: 10.2741/s341
- Segal Y, Shoenfeld Y. Vaccine-Induced Autoimmunity: The Role of Molecular Mimicry and Immune Crossreaction. *Cell Mol Immunol* (2018) 15(6):586–94. doi: 10.1038/cmi.2017.151
- Kanduc D, Shoenfeld Y. Human Papillomavirus Epitope Mimicry and Autoimmunity: The Molecular Truth of Peptide Sharing. *Pathobiology* (2019) 86(5-6):285–95. doi: 10.1159/000502889
- Shoenfeld Y, Agmon-Levin N. 'ASIA' Autoimmune/Inflammatory Syndrome Induced by Adjuvants. J Autoimmun (2011) 36(1):4–8. doi: 10.1016/j.jaut.2010.07.003
- Hawkes D, Benhamu J, Sidwell T, Miles R, Dunlop RA. Revisiting Adverse Reactions to Vaccines: A Critical Appraisal of Autoimmune Syndrome Induced by Adjuvants (ASIA). J Autoimmun (2015) 59:77–84. doi: 10.1016/j.jaut.2015.02.005
- Duthie MS, Windish HP, Fox CB, Reed SG. Use of Defined TLR Ligands as Adjuvants Within Human Vaccines. *Immunol Rev* (2011) 239(1):178–96. doi: 10.1111/j.1600-065X.2010.00978.x
- Gnjatic S, Sawhney NB, Bhardwaj N. Toll-Like Receptor Agonists: Are They Good Adjuvants? *Cancer J* (2010) 16(4):382–91. doi: 10.1097/ PPO.0b013e3181eaca65
- Jin B, Sun T, Yu XH, Yang YX, Yeo AE. The Effects of TLR Activation on T-Cell Development and Differentiation. *Clin Dev Immunol* (2012) 2012:836485. doi: 10.1155/2012/836485
- Coffman RL, Sher A, Seder RA. Vaccine Adjuvants: Putting Innate Immunity to Work. *Immunity* (2010) 33(4):492–503. doi: 10.1016/ j.immuni.2010.10.002
- Kanduc D, Shoenfeld Y. Medical, Genomic, and Evolutionary Aspects of the Peptide Sharing Between Pathogens, Primates, and Humans. *Glob Med Genet* (2020) 7(2):64–7. doi: 10.1055/s-0040-1716334
- 45. Uzawa A, Kuwabara S, Suzuki S, Imai T, Murai H, Ozawa Y, et al. Roles of Cytokines and T Cells in the Pathogenesis of Myasthenia Gravis. *Clin Exp Immunol* (2020) 203(1):366–74. doi: 10.1111/cei.13546
- Crotty S. T Follicular Helper Cell Biology: A Decade of Discovery and Diseases. *Immunity* (2019) 50(5):1132–48. doi: 10.1016/j.immuni.2019.04.011
- 47. Wang YZ, Yan M, Tian FF, Zhang JM, Liu Q, Yang H, et al. Possible Involvement of Toll-Like Receptors in the Pathogenesis of Myasthenia Gravis. *Inflammation* (2013) 36(1):121–30. doi: 10.1007/s10753-012-9526-6
- Bernasconi P, Barberis M, Baggi F, Passerini L, Cannone M, Arnoldi E, et al. Increased Toll-Like Receptor 4 Expression in Thymus of Myasthenic Patients With Thymitis and Thymic Involution. *Am J Pathol* (2005) 167 (1):129–39. doi: 10.1016/s0002-9440(10)62960-4
- Robinet M, Maillard S, Cron MA, Berrih-Aknin S, Le Panse R. Review on Toll-Like Receptor Activation in Myasthenia Gravis: Application to the Development of New Experimental Models. *Clin Rev Allergy Immunol* (2017) 52(1):133–47. doi: 10.1007/s12016-016-8549-4
- Cufi P, Dragin N, Weiss JM, Martinez-Martinez P, De Baets MH, Roussin R, et al. Implication of Double-Stranded RNA Signaling in the Etiology of Autoimmune Myasthenia Gravis. *Ann Neurol* (2013) 73(2):281–93. doi: 10.1002/ana.23791

- Cordiglieri C, Marolda R, Franzi S, Cappelletti C, Giardina C, Motta T, et al. Innate Immunity in Myasthenia Gravis Thymus: Pathogenic Effects of Toll-Like Receptor 4 Signaling on Autoimmunity. *J Autoimmun* (2014) 52:74–89. doi: 10.1016/j.jaut.2013.12.013
- Cavalcante P, Galbardi B, Franzi S, Marcuzzo S, Barzago C, Bonanno S, et al. Increased Expression of Toll-Like Receptors 7 and 9 in Myasthenia Gravis Thymus Characterized by Active Epstein-Barr Virus Infection. *Immunobiology* (2016) 221(4):516–27. doi: 10.1016/j.imbio.2015.12.007
- Cavalcante P, Barzago C, Baggi F, Antozzi C, Maggi L, Mantegazza R, et al. Toll-Like Receptors 7 and 9 in Myasthenia Gravis Thymus: Amplifiers of Autoimmunity? Ann N Y Acad Sci (2018) 1413(1):11–24. doi: 10.1111/nyas.13534
- Cufi P, Dragin N, Ruhlmann N, Weiss JM, Fadel E, Serraf A, et al. Central Role of Interferon-Beta in Thymic Events Leading to Myasthenia Gravis. *J Autoimmun* (2014) 52:44–52. doi: 10.1016/j.jaut.2013.12.016
- Démoulins T, Abdallah A, Kettaf N, Baron ML, Gerarduzzi C, Gauchat D, et al. Reversible Blockade of Thymic Output: An Inherent Part of TLR Ligand-Mediated Immune Response. J Immunol (2008) 181(10):6757–69. doi: 10.4049/jimmunol.181.10.6757
- Anz D, Thaler R, Stephan N, Waibler Z, Trauscheid MJ, Scholz C, et al. Activation of Melanoma Differentiation-Associated Gene 5 Causes Rapid Involution of the Thymus. *J Immunol* (2009) 182(10):6044–50. doi: 10.4049/ jimmunol.0803809
- Chung JY, Lee SJ, Shin B-S, Kang HG. Myasthenia Gravis Following Human Papillomavirus Vaccination: A Case Report. *BMC Neurol* (2018) 18(1):222. doi: 10.1186/s12883-018-1233-y
- Takizawa T, Kojima M, Suzuki S, Osada T, Kitagawa S, Nakahara J, et al. New Onset of Myasthenia Gravis After Intravesical Bacillus Calmette-Guerin: A Case Report and Literature Review. *Med (Baltimore)* (2017) 96 (46):e8757. doi: 10.1097/md.000000000008757
- Davalos L, Kushlaf H. New Onset of Seropositive Generalized Myasthenia Gravis Following Intravesical Bacille Calmette-Guerin Treatment for Bladder Cancer: A Case Study. *Muscle Nerve* (2019) 59(1):E1-e2. doi: 10.1002/mus.26328
- 60. He D, Zhang H, Xiao J, Zhang X, Xie M, Pan D, et al. Molecular and Clinical Relationship Between Live-Attenuated Japanese Encephalitis Vaccination and Childhood Onset Myasthenia Gravis. *Ann Neurol* (2018) 84(3):386–400. doi: 10.1002/ana.25267
- Wraith DC, Goldman M, Lambert PH. Vaccination and Autoimmune Disease: What Is the Evidence? *Lancet* (2003) 362(9396):1659–66. doi: 10.1016/s0140-6736(03)14802-7
- Bellavite P. Causality Assessment of Adverse Events Following Immunization: The Problem of Multifactorial Pathology. *F1000Res* (2020) 9:170. doi: 10.12688/f1000research.22600.2
- 63. Tavares Da Silva F, De Keyser F, Lambert PH, Robinson WH, Westhovens R, Sindic C. Optimal Approaches to Data Collection and Analysis of Potential Immune Mediated Disorders in Clinical Trials of New Vaccines. *Vaccine* (2013) 31(14):1870–6. doi: 10.1016/j.vaccine.2013.01.042
- Soriano A, Nesher G, Shoenfeld Y. Predicting Post-Vaccination Autoimmunity: Who Might be at Risk? *Pharmacol Res* (2015) 92:18–22. doi: 10.1016/j.phrs.2014.08.002
- 65. Zinman L, Thoma J, Kwong JC, Kopp A, Stukel TA, Juurlink DN. Safety of Influenza Vaccination in Patients With Myasthenia Gravis: A Population-Based Study. *Muscle Nerve* (2009) 40(6):947-51. doi: 10.1002/mus.21440
- 66. Csuka D, Czirják L, Hóbor R, Illes Z, Bánáti M, Rajczy K, et al. Effective Humoral Immunity Against Diphtheria and Tetanus in Patients With Systemic Lupus Erythematosus or Myasthenia Gravis. *Mol Immunol* (2013) 54(3-4):453–6. doi: 10.1016/j.molimm.2013.01.012
- 67. Tackenberg B, Schneider M, Blaes F, Eienbröker C, Schade-Brittinger C, Wellek A, et al. Acetylcholine Receptor Antibody Titers and Clinical Course After Influenza Vaccination in Patients With Myasthenia Gravis: A Double-Blind Randomized Controlled Trial (ProPATIent-Trial). *EBioMedicine* (2018) 28:143–50. doi: 10.1016/j.ebiom.2018.01.007
- Seok HY, Shin HY, Kim JK, Kim BJ, Oh J, Suh BC, et al. The Impacts of Influenza Infection and Vaccination on Exacerbation of Myasthenia Gravis. *J Clin Neurol* (2017) 13(4):325–30. doi: 10.3988/jcn.2017.13.4.325
- Strijbos E, Huijbers MG, van Es IE, Alleman I, van Ostaijen-ten Dam MM, Bakker J, et al. A Prospective, Placebo Controlled Study on the Humoral

Immune Response to and Safety of Tetanus Revaccination in Myasthenia Gravis. *Vaccine* (2017) 35(46):6290–6. doi: 10.1016/j.vaccine.2017.09.078

- 70. Strijbos E, Tannemaat MR, Alleman I, de Meel RHP, Bakker JA, van Beek R, et al. A Prospective, Double-Blind, Randomized, Placebo-Controlled Study on the Efficacy and Safety of Influenza Vaccination in Myasthenia Gravis. *Vaccine* (2019) 37(7):919–25. doi: 10.1016/j.vaccine.2019.01.007
- Dell' Era L, Esposito S, Corona F, Principi N. Vaccination of Children and Adolescents With Rheumatic Diseases. *Rheumatol (Oxford)* (2011) 50 (8):1358–65. doi: 10.1093/rheumatology/ker102
- 72. van Assen S, Holvast A, Benne CA, Posthumus MD, van Leeuwen MA, Voskuyl AE, et al. Humoral Responses After Influenza Vaccination Are Severely Reduced in Patients With Rheumatoid Arthritis Treated With Rituximab. Arthritis Rheum (2010) 62(1):75–81. doi: 10.1002/art.25033
- Bingham CO3rd, Looney RJ, Deodhar A, Halsey N, Greenwald M, Codding C, et al. Immunization Responses in Rheumatoid Arthritis Patients Treated With Rituximab: Results From a Controlled Clinical Trial. *Arthritis Rheum* (2010) 62(1):64–74. doi: 10.1002/art.25034
- 74. Bijl M, Agmon-Levin N, Dayer JM, Israeli E, Gatto M, Shoenfeld Y. Vaccination of Patients With Auto-Immune Inflammatory Rheumatic Diseases Requires Careful Benefit-Risk Assessment. *Autoimmun Rev* (2012) 11(8):572–6. doi: 10.1016/j.autrev.2011.10.015
- Rubin LG, Levin MJ, Ljungman P, Davies EG, Avery R, Tomblyn M, et al. 2013 IDSA Clinical Practice Guideline for Vaccination of the Immunocompromised Host. *Clin Infect Dis* (2014) 58(3):e44–100. doi: 10.1093/cid/cit684
- Rákóczi É, Szekanecz Z. Pneumococcal Vaccination in Autoimmune Rheumatic Diseases. *RMD Open* (2017) 3(2):e000484. doi: 10.1136/ rmdopen-2017-000484
- van Aalst M, Langedijk AC, Spijker R, de Bree GJ, Grobusch MP, Goorhuis A. The Effect of Immunosuppressive Agents on Immunogenicity of Pneumococcal Vaccination: A Systematic Review and Meta-Analysis. *Vaccine* (2018) 36(39):5832–45. doi: 10.1016/j.vaccine.2018.07.039
- Shapiro ED, Berg AT, Austrian R, Schroeder D, Parcells V, Margolis A, et al. The Protective Efficacy of Polyvalent Pneumococcal Polysaccharide Vaccine. *N Engl J Med* (1991) 325(21):1453–60. doi: 10.1056/nejm199111213252101
- Westra J, Rondaan C, van Assen S, Bijl M. Vaccination of Patients With Autoimmune Inflammatory Rheumatic Diseases. *Nat Rev Rheumatol* (2015) 11(3):135–45. doi: 10.1038/nrrheum.2014.206
- 80. Furer V, Eviatar T, Zisman D, Peleg H, Paran D, Levartovsky D, et al. Immunogenicity and Safety of the BNT162b2 mRNA COVID-19 Vaccine in Adult Patients With Autoimmune Inflammatory Rheumatic Diseases and in the General Population: A Multicentre Study. *Ann Rheum Dis* (2021). doi: 10.1136/annrheumdis-2021-220647
- Tay L, Leon F, Vratsanos G, Raymond R, Corbo M. Vaccination Response to Tetanus Toxoid and 23-Valent Pneumococcal Vaccines Following Administration of a Single Dose of Abatacept: A Randomized, Open-Label, Parallel Group Study in Healthy Subjects. *Arthritis Res Ther* (2007) 9(2):R38. doi: 10.1186/ar2174
- Hua C, Barnetche T, Combe B, Morel J. Effect of Methotrexate, Anti-Tumor Necrosis Factor α, and Rituximab on the Immune Response to Influenza and Pneumococcal Vaccines in Patients With Rheumatoid Arthritis: A Systematic Review and Meta-Analysis. *Arthritis Care Res (Hoboken)* (2014) 66(7):1016–26. doi: 10.1002/acr.22246
- 83. Kapetanovic MC, Saxne T, Sjöholm A, Truedsson L, Jönsson G, Geborek P. Influence of Methotrexate, TNF Blockers and Prednisolone on Antibody Responses to Pneumococcal Polysaccharide Vaccine in Patients With Rheumatoid Arthritis. *Rheumatol (Oxford)* (2006) 45(1):106–11. doi: 10.1093/rheumatology/kei193
- 84. Crnkic Kapetanovic M, Saxne T, Jönsson G, Truedsson L, Geborek P. Rituximab and Abatacept But Not Tocilizumab Impair Antibody Response to Pneumococcal Conjugate Vaccine in Patients With Rheumatoid Arthritis. *Arthritis Res Ther* (2013) 15(5):R171. doi: 10.1186/ar4358
- McKinnon JE, Maksimowicz-McKinnon K. Autoimmune Disease and Vaccination: Impact on Infectious Disease Prevention and a Look at Future Applications. *Trans Res* (2016) 167(1):46–60. doi: 10.1016/ j.trsl.2015.08.008
- Kantsø B, Halkjær SI, Thomsen O, Belard E, Gottschalck IB, Jørgensen CS, et al. Immunosuppressive Drugs Impairs Antibody Response of the Polysaccharide

and Conjugated Pneumococcal Vaccines in Patients With Crohn's Disease. Vaccine (2015) 33(41):5464-9. doi: 10.1016/j.vaccine.2015.08.011

- Pollard AJ, Bijker EM. A Guide to Vaccinology: From Basic Principles to New Developments. *Nat Rev Immunol* (2020) 21(2):83–100. doi: 10.1038/ s41577-020-00479-7
- Westerberg E, Punga AR. Mortality Rates and Causes of Death in Swedish Myasthenia Gravis Patients. *Neuromuscul Disord* (2020) 30(10):815–24. doi: 10.1016/j.nmd.2020.08.355
- Glück T, Müller-Ladner U. Vaccination in Patients With Chronic Rheumatic or Autoimmune Diseases. *Clin Infect Dis* (2008) 46(9):1459– 65. doi: 10.1086/587063
- MGFA. VACCINATIONS: Myasthenia Gravis Foundation of America (2021). Available at: https://myasthenia.org/MG-Community/Managing-Your-MG/Vaccinations.
- Furer V, Rondaan C, Heijstek MW, Agmon-Levin N, van Assen S, Bijl M, et al. 2019 Update of EULAR Recommendations for Vaccination in Adult Patients With Autoimmune Inflammatory Rheumatic Diseases. Ann Rheumatic Dis (2020) 79(1):39–52. doi: 10.1136/annrheumdis-2019-215882
- Cartwright SL, Cartwright MS. Health Maintenance for Adults With Neuromuscular Diseases on Immunosuppression. *Muscle Nerve* (2019) 59 (4):397–403. doi: 10.1002/mus.26382
- Muralidhar Reddy Y, SK B, Osman S, Murthy JMK. Temporal Association Between SARS-CoV-2 and New-Onset Myasthenia Gravis: Is it Causal or Coincidental? *BMJ Case Rep* (2021) 14(7):e244146. doi: 10.1136/bcr-2021-244146
- 94. Essajee F, Lishman J, Solomons R, Abraham DR, Goussard P, Van Toorn R. Transient Acetylcholine Receptor-Related Myasthenia Gravis, Post Multisystem Inflammatory Syndrome in Children (MIS-C) Temporally Associated With COVID-19 Infection. *BMJ Case Rep* (2021) 14(8). doi: 10.1136/bcr-2021-244102
- Farsalinos K, Niaura R, Le Houezec J, Barbouni A, Tsatsakis A, Kouretas D, et al. Editorial: Nicotine and SARS-CoV-2: COVID-19 may be a Disease of the Nicotinic Cholinergic System. *Toxicol Rep* (2020) 7:658–63. doi: 10.1016/ j.toxrep.2020.04.012
- Lyons-Weiler J. Pathogenic Priming Likely Contributes to Serious and Critical Illness and Mortality in COVID-19 via Autoimmunity. J Transl Autoimmun (2020) 3:100051. doi: 10.1016/j.jtauto.2020.100051
- Vojdani A, Vojdani E, Kharrazian D. Reaction of Human Monoclonal Antibodies to SARS-CoV-2 Proteins With Tissue Antigens: Implications for Autoimmune Diseases. *Front Immunol* (2020) 11:617089. doi: 10.3389/ fimmu.2020.617089
- Adiguzel Y. Molecular Mimicry Between SARS-CoV-2 and Human Proteins. Autoimmun Rev (2021) 20(4):102791. doi: 10.1016/j.autrev.2021.102791
- Finsterer J, Scorza FA. MuSK-Positive Myasthenia May Not Only Be Triggered by SARS-CoV-2. Eur J Neurol (2021). doi: 10.1111/ene.14813
- 100. Herold T, Jurinovic V, Arnreich C, Lipworth BJ, Hellmuth JC, von Bergwelt-Baildon M, et al. Elevated Levels of IL-6 and CRP Predict the Need for Mechanical Ventilation in COVID-19. J Allergy Clin Immunol (2020) 146 (1):128–36.e4. doi: 10.1016/j.jaci.2020.05.008
- 101. Li X, Geng M, Peng Y, Meng L, Lu S. Molecular Immune Pathogenesis and Diagnosis of COVID-19. J Pharm Anal (2020) 10(2):102–8. doi: 10.1016/ j.jpha.2020.03.001
- 102. Team WHO. COVID-19 Vaccine Tracker and Landscape. World Health Organization (2021). Available at: https://www.who.int/publications/m/ item/draft-landscape-of-covid-19-candidate-vaccines.
- 103. Kanduc D, Shoenfeld Y. Molecular Mimicry Between SARS-CoV-2 Spike Glycoprotein and Mammalian Proteomes: Implications for the Vaccine. *Immunol Res* (2020) 68(5):310–3. doi: 10.1007/s12026-020-09152-6
- 104. Priori R, Pellegrino G, Colafrancesco S, Alessandri C, Ceccarelli F, Di Franco M, et al. SARS-CoV-2 Vaccine Hesitancy Among Patients With Rheumatic and Musculoskeletal Diseases: A Message for Rheumatologists. *Ann Rheum Dis* 80(7):953–4. doi: 10.1136/annrheumdis-2021-220059
- 105. Karikó K, Muramatsu H, Welsh FA, Ludwig J, Kato H, Akira S, et al. Incorporation of Pseudouridine Into mRNA Yields Superior Nonimmunogenic Vector With Increased Translational Capacity and Biological Stability. *Mol Ther* (2008) 16(11):1833–40. doi: 10.1038/mt.2008.200
- 106. Karikó K, Muramatsu H, Ludwig J, Weissman D. Generating the Optimal mRNA for Therapy: HPLC Purification Eliminates Immune Activation and

Improves Translation of Nucleoside-Modified, Protein-Encoding mRNA. Nucleic Acids Res (2011) 39(21):e142. doi: 10.1093/nar/gkr695

- 107. Watad A, De Marco G, Mahajna H, Druyan A, Eltity M, Hijazi N, et al. Immune-Mediated Disease Flares or New-Onset Disease in 27 Subjects Following mRNA/DNA SARS-CoV-2 Vaccination. Vaccines (Basel) (2021) 9(5):435. doi: 10.3390/vaccines9050435
- 108. Tagliaferri AR, Narvaneni S, Azzam MH, Grist W. A Case of COVID-19 Vaccine Causing a Myasthenia Gravis Crisis. *Cureus* (2021) 13(6):e15581. doi: 10.7759/cureus.15581
- 109. Connolly CM, Ruddy JA, Boyarsky BJ, Avery RK, Werbel WA, Segev DL, et al. Safety of the First Dose of mRNA SARS-CoV-2 Vaccines in Patients With Rheumatic and Musculoskeletal Diseases. Ann Rheum Dis (2021). doi: 10.1136/annrheumdis-2021-220231
- 110. Braun-Moscovici Y, Kaplan M, Braun M, Markovits D, Giryes S, Toledano K, et al. Disease Activity and Humoral Response in Patients With Inflammatory Rheumatic Diseases After Two Doses of the Pfizer mRNA Vaccine Against SARS-CoV-2. Ann Rheum Dis (2021). doi: 10.1136/annrheumdis-2021-220503
- 111. Ruddy JA, Connolly CM, Boyarsky BJ, Werbel WA, Christopher-Stine L, Garonzik-Wang J, et al. High Antibody Response to Two-Dose SARS-CoV-2 Messenger RNA Vaccination in Patients With Rheumatic and Musculoskeletal Diseases. Ann Rheum Dis (2021). doi: 10.1136/annrheumdis-2021-220656
- 112. Boyarsky BJ, Ruddy JA, Connolly CM, Ou MT, Werbel WA, Garonzik-Wang JM, et al. Antibody Response to a Single Dose of SARS-CoV-2 mRNA Vaccine in Patients With Rheumatic and Musculoskeletal Diseases. Ann Rheum Dis (2021). doi: 10.1136/annrheumdis-2021-220289
- Moutsopoulos HM. A Recommended Paradigm for Vaccination of Rheumatic Disease Patients With the SARS-CoV-2 Vaccine. J Autoimmun (2021) 121:102649. doi: 10.1016/j.jaut.2021.102649
- 114. D'Silva KM, Serling-Boyd N, Hsu TY, Sparks JA, Wallace ZS. SARS-CoV-2 Antibody Response After COVID-19 in Patients With Rheumatic Disease. Ann Rheum Dis (2021). doi: 10.1136/annrheumdis-2020-219808
- 115. SK C, Ahmed S, Shenoy V, Menon AR, Saijan S, Babu SA, et al. Correspondence on 'SARS-CoV-2 Antibody Response After COVID-19 in Patients With Rheumatic Disease'. Ann Rheum Dis (2021). doi: 10.1136/ annrheumdis-2021-220148
- 116. Sahin U, Muik A, Derhovanessian E, Vogler I, Kranz LM, Vormehr M, et al. COVID-19 Vaccine BNT162b1 Elicits Human Antibody and T(H)1 T Cell Responses. *Nature* (2020) 586(7830):594–9. doi: 10.1038/s41586-020-2814-7
- 117. Rzepiński Ł, Zawadka-Kunikowska M. COVID-19 Pandemic Year in a Sample of Polish Myasthenia Gravis Patients: An Observational Study. *Neurol Neurochir Pol* (2021). doi: 10.5603/PJNNS.a2021.0054
- 118. Costamagna G, Abati E, Bresolin N, Comi GP, Corti S. Management of Patients With Neuromuscular Disorders at the Time of the SARS-CoV-2 Pandemic. *J Neurol* (2021) 268(5):1580–91. doi: 10.1007/s00415-020-10149-2
- Muppidi S, Guptill JT, Jacob S, Li Y, Farrugia ME, Guidon AC, et al. COVID-19-Associated Risks and Effects in Myasthenia Gravis (CARE-Mg). *Lancet Neurol* (2020) 19(12):970–1. doi: 10.1016/s1474-4422(20)30413-0
- 120. Solé G, Mathis S, Friedman D, Salort-Campana E, Tard C, Bouhour F, et al. Impact of Coronavirus Disease 2019 in a French Cohort of Myasthenia Gravis. *Neurology* (2021) 96(16):e2109–e20. doi: 10.1212/wnl.000000000011669
- 121. Kovvuru S, Nalleballe K, Onteddu SR, Sharma R, Jasti M, Kapoor N, et al. Immunosuppression in Chronic Autoimmune Neurological Disorders During the COVID-19 Pandemic. J Neurol Sci (2021) 420:117230. doi: 10.1016/j.jns.2020.117230
- 122. MGFA. CDC Announces Authorization of Additional COVID-19 Vaccine Dose for People With Compromised Immune Systems, Including Patients With Myasthenia Gravis (MG) Taking Immunosuppressive Treatment(s) 2021. Available at: https:// myasthenia.org/MG-Community/COVID-19-Resource-Center.
- 123. Curtis JR, Johnson SR, Anthony DD, Arasaratnam RJ, Baden LR, Bass AR, et al. American College of Rheumatology Guidance for COVID-19 Vaccination in Patients With Rheumatic and Musculoskeletal Diseases: Version 2. Arthritis Rheumatol (2021) 73(8):e30–45. doi: 10.1002/art.41877
- 124. Ossato A, Tessari R, Trabucchi C, Zuppini T, Realdon N, Marchesini F. Comparison of Medium-Term Adverse Reactions Induced by the First and Second Dose of mRNA BNT162b2 (Comirnaty, Pfizer-BioNTech) Vaccine: A Post-Marketing Italian Study Conducted Between 1 January and 28 February 2021. Eur J Hosp Pharm (2021). doi: 10.1136/ejhpharm-2021-002933

125. Blumental S, Debré P. Challenges and Issues of Anti-SARS-CoV-2 Vaccines. Front Med (Lausanne) (2021) 8:664179. doi: 10.3389/ fmed.2021.664179

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Zhou, Zhou, Yang and Yang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.