# SOX2 for Stem Cell Therapy and Medical Use: Pros or Cons?

Cell Transplantation Volume 29: 1–7 © The Author(s) 2020 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/0963689720907565 journals.sagepub.com/home/cll SAGE

Hong-Meng Chuang<sup>1,2</sup>, Mao-Hsuan Huang<sup>1,3</sup>, Yu-Shuan Chen<sup>1,2</sup>, and Horng-Jyh Harn<sup>1,4</sup>

#### Abstract

Stem cell transplantation is a fast-developing technique, which includes stem cell isolation, purification, and storage, and it is in high demand in the industry. In addition, advanced applications of stem cell transplantation, including differentiation, gene delivery, and reprogramming, are presently being studied in clinical trials. In contrast to somatic cells, stem cells are self-renewing and have the ability to differentiate; however, the molecular mechanisms remain unclear. SOX2 (sex-determining region Y [SRY]-box 2) is one of the well-known reprogramming factors, and it has been recognized as an oncogene associated with cancer induction. The exclusion of SOX2 in reprogramming methodologies has been used as an alternative cancer treatment approach. However, the manner by which SOX2 induces oncogenic effects remains unclear, with most studies demonstrating its regulation of the cell cycle and no insight into the maintenance of cellular stemness. For controlling certain critical pathways, including Shh and Wnt pathways, SOX2 is considered irreplaceable and is required for the normal functioning of stem cells, particularly neural stem cells. In this report, we discussed the functions of SOX2 in both stem and cancer cells, as well as how this powerful regulator can be used to control cell fate.

#### **Keywords**

SOX2, cell transplantation, stem cells, cancer, stemness

## Introduction

Stem cell transplantation is a well-established technique that has been part of the clinical treatment strategies for both malignant (e.g., acute myeloid leukemia and Hodgkin's lymphoma) and nonmalignant (e.g., thalassemia and sickle cell anemia) diseases and disorders since 1959<sup>1</sup>. In response to the increasing medical need, stem cells are now being isolated and transplanted from a variety of sources, including umbilical cord blood, placenta, amniotic fluid, dental pulp, and adipose tissue $^{2-4}$ . In addition, the discovery of induced pluripotent stem cells (iPSCs) has dramatically broadened the field<sup>5</sup>. These improvements are a reflection of the unmet medical need of regenerative medicine and the limitations of drugs and medical devices<sup>6</sup>. Despite organ transplantation having been successfully performed for the heart, kidney, liver, lung, pancreas, intestine, and thymus, organ sources are in very limited supply<sup>7,8</sup>. Even if a patient receives an organ and the transplantation is considered a success, the patient is required to take antirejection drugs, which have the risk of severe side effects. As such, the development of artificial organs may represent a promising solution; however, effective and efficient techniques for tissue engineering pose numerous challenges<sup>9</sup>.

Stem cells and/or progenitor cells may be a good choice for compensation of the functions of target tissues and for the secretion of appropriate cytokines and growth factors, including those that are considered immunomodulatory<sup>10–12</sup>. However, the majority of studies have revealed that delivered cells rarely transdifferentiate into their target type and the survival time remains insufficient<sup>13,14</sup>. Nonetheless, while the mechanisms remain unclear, most experimental and

<sup>4</sup> Department of Pathology, Hualien Tzu Chi Hospital & Tzu Chi University, Hualien, Republic of China

Submitted: November 6, 2019. Revised: January 14, 2020. Accepted: January 27, 2020.

#### **Corresponding Author:**

Horng-Jyh Harn, Department of Pathology, Hualien Tzu Chi Hospital & Tzu Chi University, Hualien 970, Republic of China. Email: arthewduke@gmail.com



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

<sup>&</sup>lt;sup>1</sup> Buddhist Tzu Chi Bioinnovation Center, Tzu Chi Foundation, Hualien, Republic of China

<sup>&</sup>lt;sup>2</sup> Department of Medical Research, Hualien Tzu Chi Hospital, Hualien, Republic of China

<sup>&</sup>lt;sup>3</sup> Department of Stem Cell Applied Technology, Gwo Xi Stem Cell Applied Technology, Hsinchu, Republic of China

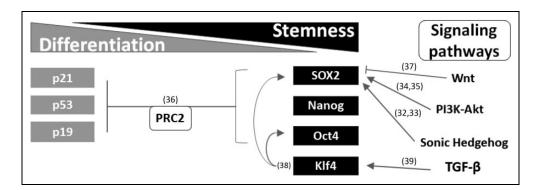


Fig. 1. Molecular interactions illustrate various SOX2 signaling partners.<sup>32–39</sup>

clinical results have shown positive results<sup>15–17</sup>. As such, the technologies underlying stem cell therapy continue to improve regarding cellular function and survival duration. The most important breakthrough has been the reprogramming of somatic cells into pluripotent stem cells with the exogenous expression of certain transcription factors<sup>18</sup>. The potential applications of stem cells, including cellular transplantation and organ development, have been tremendously enhanced after the discovery of iPSCs<sup>19,20</sup>.

As a member of the SOX gene family and SOXB group, which includes SOX1, SOX2, and SOX3, SOX2 (sexdetermining region Y [SRY]-box 2) encodes a 34.3 kD protein<sup>21</sup>. As a key regulator of self-renewal, SOX2 protein binds to octamer-binding transcription factor 4 (Oct4) and enhances the expression of Nanog<sup>22,23</sup>. However, Tanaka et al. indicated that SOX2 is unnecessary as an enhancer, suggesting that it modulates the expression of  $Oct4^{24-26}$ . The coupling of SOX2 to paired box protein 6 (PAX6) and BRN2 (encoded by POU3F2 in humans) has been shown to regulate eye and neural primordial cell functions<sup>27</sup>. Interestingly, SOX2 and/or the partner protein are not considered sufficient for transcriptional activation, but this complex is<sup>28</sup>. Once the complex is formed, downstream genes such as undifferentiated embryonic cell transcription factor 1 and fibroblast growth factor 4 activate and enhance embrionic stem cell development and survival<sup>29</sup>. Accordingly, the knockdown of Sox2 expression in mouse embryonic stem cells (ESCs) results in the failure of this self-renewal property and leads to differentiation<sup>22</sup>. In contrast to tumorigenesis, the expression level of SOX2 correlates with lower survival and treatment resistance<sup>30</sup>. Therefore, we evaluated the relationship between SOX2 and its functions in both stem and cancer cells and discovered a potential approach for improving stem cells and deteriorating cancer cells.

## SOX2 Is Associated With an Enormous Expression Network

The characteristics of stemness are associated with the target genes of SOX2. In addition, stem cells possess regulatory mechanisms to maintain the appropriate expression of SOX2. For mouse ESCs, the exogenous elevated expression of Sox2 leads to differentiation of ESCs into a wide range of cell types, including neuroectoderm, mesoderm, and trophectoderm (TE)<sup>31</sup>. Moreover, feedback regulation involved in the Akt pathway reactivates endogenous Sox2 expression and serves to retain cellular stemness (Fig. 1)<sup>40</sup>. However, in comparison with iPSCs, the expression of SOX2 is artificial and lacks interactive control. Nevertheless, to reprogram cells into iPSCs, four genes, namely, Oct4, Klf-4, SOX2, and c-Myc (abbreviated to OKSM), are exogenously activated and these genes need a specific ratio to work adequately. Since the OKSM is necessary for pluripotency, other accessory factors such as Nanog and Sal-like protein 4 can only increase the efficiency of reprogramming and cannot replace SOX2 or OCT4<sup>41,42</sup>. For example, a ratio increase of Klf4 is recommended in one of the commercial cellular reprogramming kits. Moreover, the expression of SOX2 is activated by the VP16 transactivator and further improves reprogramming efficiency<sup>43</sup>. These findings indicate that the OKSM acts as a driving force in the fertilization stage and should be tightly restricted or the cells may get out of control. Thus, the upstream and downstream regions of the SOX2 coding sequence contain large untranslated regions (UTRs) or the so-called gene deserts to prevent mutations and false binding<sup>44</sup>. Certain enhancers, including miRNA, long-noncoding RNA, and posttranslational modification have been shown to reversely regulate both transcriptional and posttranslational activity. At least 19 miRNAs, including miR-200b and miR-145, can influence the expression of SOX2<sup>45</sup>. Indeed, miR-200 facilitates the reprogramming of fibroblasts into iPSCs in the presence of OKSM<sup>46</sup>. Moreover, miR-145 targets the 3'-UTR of SOX2 and inhibits self-renewal in human ESCs and iPSCs<sup>47</sup>. In other words, simply elevating the expression of SOX2 serves to attract negative regulation molecules such as the cyclin-dependent kinase inhibitor 1A  $(p21^{Cip1})^{48}$ . Conversely, the zygote has a massive demethylation pattern compared with reprogrammed cells<sup>49</sup>. Although existing techniques, including single-cell RNA-Seq, methylation array, and bioinformatics, have confirmed that the methylation state changes in zygotes and somatic cells, the secrets of dedifferentiation remain a mystery<sup>50</sup>.

## SOX2 Utilization in Stem Cell Therapy

Except for the stemness regulation of SOX2, the existence for cell differentiation and development is necessary. Notably, in the development of ESCs, when the blastocyst is formed and then divides into the TE and inner cell mass, the expression of SOX2 is decreased. However, the TE will not form if SOX2 is impaired or knocked down by siRNA<sup>51</sup>. This change is due to the complex formed with Oct4 and Nanog. For example, Oct4 and Nanog bind to SOX2 and regulate its functions of self-renewal and differentiation inhibition<sup>52</sup>. In adult humans, the olfactory nerve proliferates and is replaced every 3 to 4 weeks. The SOX2/PAX6-expressed epithelium plays an important role in maintaining the multipotency of the olfactory nerve<sup>53</sup>. These findings suggest further applications in the transplantation from iPSC-differentiated neural stem cells (NSCs). In particular, the in vitrotranscribed mRNA of SOX2 has been shown to induce NSC morphology in human dermal fibroblasts<sup>54</sup>. In addition, another study revealed that exogenous Sox2 expression in rat bone marrow-derived stem cells (BMSCs) benefits the cell transplantation treatment in a rat traumatic brain injury (TBI) model<sup>55</sup>. Especially, BMSCs retain their self-renewal property via the expression of Sirtuin1 (SIRT1)<sup>56</sup>. SIRT1 is a lysine deacetylase that contributes in maintaining SOX2 content by avoiding the acetylation and ubiquitination of SOX2<sup>57</sup>. Moreover, proliferation and differentiation potential is conferred by the forced SOX2 expression of BMSC<sup>58</sup>. Using MRI tracking, Jiang et al. found that NSCs migrate into the injury site of rats with TBI<sup>59</sup>. Therefore, the existence of SOX2 is essential for the maintenance of selfrenewal and multipotency. These studies suggested that Sox2-positive cells may play a role in neuron regeneration, enhancing neural functions after brain injury<sup>60</sup>.

## **Direct Evidence of SOX2 Initiating Tumorigenesis**

SOX2 is generally considered an oncogene; however, its role in tumorigenesis remains controversial<sup>61,62</sup>. As part of the same lineage of breast cancer cells, the SOX2-positive population shows a greater colony-forming ability and would be abolished by SOX2 knockdown<sup>63,64</sup>. SOX2 is amplified in patients with cancer, and it contributes to the same stemness property observed in stem cells of patients with lung, brain, breast, and colon tumors<sup>65</sup>.

The clinical implications of SOX2 and cancers vary depending on the type of cancer, influencing patient survival and prognosis<sup>66</sup>. These molecules and pathways include VEGF, MAPK, Notch-Shh, BMP, Jak-STAT, and others, depending on the types of tumors<sup>67-69</sup>. In brief, SOX2 regulates downstream genes and microRNAs by direct DNA binding, resulting in the alteration of thousands of genes and 3

hundreds of microRNAs<sup>35</sup>. Moreover, a glioma cell subset with high levels of SOX2 has been shown to be resistant to platelet-derived growth factor (PDGF)- and insulin-like growth factor 1 (IGF-1)-receptor inhibitors<sup>70</sup>. Conversely, SOX2 may play a role in the maintenance of PDGF and IGF-1 pathways in cancer cells as well as in stem cells and may produce dysplasia or tumor cell initiation. However, in patients with gastric cancer, the overall survival rate is lower with SOX2 methylation than with unmethylated SOX2. Moreover, the exogenous expression of SOX2 results in cell cycle arrest through cyclin-dependent kinase inhibitor 1B (p27<sup>Kip1</sup>) and Rb phosphorylation<sup>71</sup>. Indeed, Sox2 expression in the mouse respiratory epithelium does not cause pulmonary tumors but induces the cellular proliferation of respiratory epithelial cells<sup>72</sup>. Direct induced-NSCs can also be obtained by SOX2 expression in human and mouse fibroblasts without tumorigenesis<sup>73</sup>.

## **Future Prospects of SOX2 Utilization**

Due to the close relationship between SOX2 and cancer, studies that have investigated SOX2-dependent gene manipulation are limited. However, cell differentiation and proliferation has been achieved, including SOX2-expressing dental pulp stem cells, which lead to the differentiation of odontoblasts<sup>74</sup>. SOX2 also cooperates with various cofactors, including Oct4 for stemness, BRN2 for neural differentiation, and PAX3 for melanocyte maturation. These studies suggested that SOX2 (and possibly other members in the family) is one of the masters regulating cell fate and is associated with different kinds of cofactors<sup>75</sup>.

An urgent question we would ask is how to take advantage of SOX2 without eliciting detrimental effects? For the nerve system, SOX2 may prove to be a useful regulator for the maintenance of progenitor characteristics, allowing the cells to retain their ability to self-renew and differentiate into neurons, astrocytes, and oligodendrocytes<sup>76,77</sup>. Moreover, Hagey and Muhr found that decreased expression of Sox2 resulted in the differentiation of radial glia cells into their more developed progeny, intermediate progenitor cells<sup>78</sup>. However, mouse NSCs and progenitors have reduced differentiation ability and SOX2 expression with the loss of the E2f3a transcription factor, indicating that the expression of SOX2 is essential for neurogenesis<sup>79</sup>. These studies indicated that SOX2 is required for stemness but is unfavorable for differentiation. In other words, the expression of SOX2 should be controlled or there is a risk of the cells going corrupt.

Conversely, the SOX2-induced stemness ability is devastating and problematic when it appears in tumor cells. Since the existence of SOX2 is associated with the cell membrane, specific antibodies are not effective for disrupting its function. To control the stemness and metastatic properties, Tuhin et al. found that the downregulation effect of actinomycin D induced the cell death of breast cancer stem cells<sup>80</sup>. Moreover, SOX2 knockdown or small molecules reduced **Table I.** A Summary Table of SOX2 Potential in Medical Use and Their Main Concerns for Cancer Therapy and Stem Cell Transplantation.

Potential in Medical Use	Main Concerns	Ways to Breakthrough
Manipulate cell fate	The complicated upstream and downstream effectors of SOX2	Use the necessary domain of SOX2 <sup>66</sup>
Prolong stem cell lifespan during transplantation	Risk of neoplastic or teratoma growth	Use an additional molecular beacon that indicates abnormal SOX2 expression <sup>89</sup>
Retains stem cell potential Target as a tumor marker	May be involved in tumorigenesis Constant expression in normal cells	The development of tumor-specific siRNA or antibodies <sup>90</sup>

SOX2 expression and inhibited the stemness and metastatic properties<sup>61,81</sup>. A significant factor in patients with cancer is the selection of  $SOX2^+$  cells after anticancer therapies, including radiotherapy and chemotherapy, and SOX2induced drug-resistant genes have been characterized in numerous studies<sup>82,83</sup>. These *SOX2*<sup>+</sup> cells then form a new tumor bulk, with the most well-studied type being the quiescent Sonic Hedgehog subgroup medulloblastoma, which is activated into medulloblastoma-propagating cells after antimitotic drug treatment<sup>84</sup>. Moreover, the inhibition of the SOX2-driven transcriptional network arrests GBM growth by treatment with mithramycin, which is an antineoplastic antibiotic<sup>85</sup>. Indeed, SOX2 antigen and antibody were found in small-cell lung cancer (SCLC) cell lines and sera in SCLC patients. However, neither the antigen nor the antibody of SOX2 or other SOX group B genes exist in normal sera, suggesting that SOX2 might be a potential tumor target or marker<sup>86</sup>. However, these effects should be precisely targeted toward tumor cells to take advantage of the attributes of SOX2.

## Summary

With the use of iPSC-derived retinal pigment epithelial cells transplanted in clinical trials, a large research and development effort has been undertaken to not only evaluate the effectiveness and safety but also improve the associated techniques<sup>87,88</sup>. In this report, we discussed the stemness-prone properties and some possible risks of *SOX2* in Table 1. In addition, the safety issue in iPSC-derived cell therapy is the most important topic; therefore, Larsson et al. established a molecular beacon to compliment SOX2 mRNA, displaying the fluorescent signal when SOX2 is expressed<sup>89</sup>. Although the interaction networks remain incomplete, reactions with different expression levels of *SOX2* are more apparent than ever. In summary, the potential applications of *SOX2* are

extremely promising but precise targeting and expression in the right place and at the right dosage are crucial.

#### Acknowledgments

We are grateful to Enago (www.enago.tw) for English proofreading.

## **Author Contributions**

Writing—original draft preparation, HM Chuang; writing—review, MS Huang and YS Chen; conceptualization, HJ Harn.

#### **Declaration of Conflicting Interests**

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

### Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study was funded by Buddhist Tzu Chi Bioinnovation Center, Tzu Chi Foundation, Hualien, Taiwan (project title: Development of a small molecule regulates transcription activity of SOX2 in type I collagen synthesis in fibroblasts for treating pulmonary fibrosis, MF00A130SS01) and Ministry of Science and Technology, Taiwan (MOST 106-2320-B-303-001-MY3 and MOST 106-2320-B-303-002-MY3).

### ORCID iDs

Hong-Meng Chuang https://orcid.org/0000-0003-1978-3618 Horng-Jyh Harn https://orcid.org/0000-0001-6777-3284

#### References

- Copelan EA. Hematopoietic stem-cell transplantation. N Engl J Med. 2006;354(17):1813–1826.
- Lynch W, Rezai S, Henderson CE. Human amniotic fluid: a source of stem cells for possible therapeutic use. Am J Obstet Gynecol. 2016;215(3):401.
- 3. Kfoury Y, Scadden DT. Mesenchymal cell contributions to the stem cell niche. Cell Stem Cell. 2015;16(3):239–253.
- Mead B, Logan A, Berry M, Leadbeater W, Scheven BA. Concise review: dental pulp stem cells: A novel cell therapy for retinal and central nervous system repair. Stem Cells. 2017; 35(1):61–67.
- Hayashi R, Ishikawa Y, Sasamoto Y, Katori R, Nomura N, Ichikawa T, Araki S, Soma T, Kawasaki S, Sekiguchi K, Quantock AJ, et al. Co-ordinated ocular development from human iPS cells and recovery of corneal function. Nature. 2016;531(7594):376–380.
- Chang CY, Ting HC, Liu CA, Su HL, Chiou TW, Harn HJ, Lin SZ. Induced pluripotent stem cells: a powerful neurodegenerative disease modeling tool for mechanism study and drug discovery [published online ahead of print January 1, 2018]. Cell Transplant. 2018;27(11):1588–1602.
- Martin-Gandul C, Mueller NJ, Pascual M, Manuel O. The impact of infection on chronic allograft dysfunction and allograft survival after solid organ transplantation. Am J Transplant. 2015;15(12):3024–3040.

- Manara AR, Murphy PG, O'Callaghan G. Donation after circulatory death. Br J Anaesth. 2012;108(Suppl 1):i108–i121.
- 9. Marx V. Tissue engineering: organs from the lab. Nature. 2015;522(7556):373–377.
- Gao F, Chiu SM, Motan DA, Zhang Z, Chen L, Ji HL, Tse HF, Fu QL, Lian Q. Mesenchymal stem cells and immunomodulation: current status and future prospects. Cell Death Dis. 2016; 7:e2062.
- Liao S, Zhang Y, Ting S, Zhen Z, Luo F, Zhu Z, Jiang Y, Sun S, Lai WH, Lian Q, Tse HF. Potent immunomodulation and angiogenic effects of mesenchymal stem cells versus cardiomyocytes derived from pluripotent stem cells for treatment of heart failure. Stem Cell Res Ther. 2019;10(1):78.
- Chuang HM, Shih TE, Lu KY, Tsai SF, Harn HJ, Ho LI. Mesenchymal stem cell therapy of pulmonary fibrosis: improvement with target combination [published online ahead of print January 1, 2018]. Cell Transplant. 2018;27(11): 1581–1587.
- Hou D, Youssef EA, Brinton TJ, Zhang P, Rogers P, Price ET, Yeung AC, Johnstone BH, Yock PG, March KL. Radiolabeled cell distribution after intramyocardial, intracoronary, and interstitial retrograde coronary venous delivery: implications for current clinical trials. Circulation. 2005;112(9 Suppl):1150–1156.
- Kurtz A. Mesenchymal stem cell delivery routes and fate. Int J Stem Cells 2008;1(1):1–7.
- Squillaro T, Peluso G, Galderisi U. Clinical trials with mesenchymal stem cells: an update. Cell Transplant. 2016; 25(5):829–848.
- Trounson A, McDonald C. Stem cell therapies in clinical trials: progress and challenges. Cell Stem Cell. 2015;17(1):11–22.
- Sykova E, Rychmach P, Drahoradova I, Konradova S, Ruzickova K, Vorisek I, Forostyak S, Homola A, Bojar M. Transplantation of mesenchymal stromal cells in patients with amyotrophic lateral sclerosis: results of phase I/IIa clinical Trial. Cell Transplant. 2017;26(4):647–658.
- Maza I, Caspi I, Zviran A, Chomsky E, Rais Y, Viukov S, Geula S, Buenrostro JD, Weinberger L, Krupalnik V, Hanna S, et al. Transient acquisition of pluripotency during somatic cell transdifferentiation with iPSC reprogramming factors. Nat Biotechnol. 2015;33(7):769–774.
- Suchy F, Yamaguchi T, Nakauchi H. iPSC-Derived organs in vivo: challenges and promise. Cell Stem Cell. 2018;22(1): 21–24.
- Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell 2006;126(4):663–676.
- Iwafuchi-Doi M, Yoshida Y, Onichtchouk D, Leichsenring M, Driever W, Takemoto T, Uchikawa M, Kamachi Y, Kondoh H. The Pou5f1/Pou3f-dependent but SoxB-independent regulation of conserved enhancer N2 initiates Sox2 expression during epiblast to neural plate stages in vertebrates. Dev Biol. 2011; 352(2):354–366.
- 22. Chew JL, Loh YH, Zhang W, Chen X, Tam WL, Yeap LS, Li P, Ang YS, Lim B, Robson P, Ng HH. Reciprocal transcriptional regulation of Pou5f1 and Sox2 via the Oct4/Sox2

complex in embryonic stem cells. Mol Cell Biol. 2005; 25(14):6031-6046.

- Mato Prado M, Frampton AE, Stebbing J, Krell J.Gene of the month: NANOG. J Clin Pathol. 2015;68(10):763–765.
- 24. Masui S, Nakatake Y, Toyooka Y, Shimosato D, Yagi R, Takahashi K, Okochi H, Okuda A, Matoba R, Sharov AA, Ko MS, et al. Pluripotency governed by Sox2 via regulation of Oct3/4 expression in mouse embryonic stem cells. Nat Cell Biol. 2007;9(6):625–635.
- 25. Chen SM, Lee MS, Chang CY, Lin SZ, Cheng EH, Liu YH, Pan HC, Lee HC, Su HL. Prerequisite OCT4 maintenance potentiates the neural induction of differentiating human embryonic stem cells and induced pluripotent stem cells. Cell Transplant. 2015;24(5):829–844.
- Tanaka S, Kamachi Y, Tanouchi A, Hamada H, Jing N, Kondoh H. Interplay of SOX and POU factors in regulation of the Nestin gene in neural primordial cells. Mol Cell Biol. 2004; 24(20):8834–8846.
- Matsushima D, Heavner W, Pevny LH. Combinatorial regulation of optic cup progenitor cell fate by SOX2 and PAX6. Development. 2011;138(3):443–454
- Kamachi Y, Uchikawa M, Tanouchi A, Sekido R, Kondoh H. Pax6 and SOX2 form a co-DNA-binding partner complex that regulates initiation of lens development. Genes Dev. 2001; 15(10):1272–1286.
- Yuan H, Corbi N, Basilico C, Dailey L. Developmentalspecific activity of the FGF-4 enhancer requires the synergistic action of Sox2 and Oct-3. Genes Dev. 1995;9(21):2635–2645.
- Chung JH, Jung HR, Jung AR, Lee YC, Kong M, Lee JS, Eun YG. SOX2 activation predicts prognosis in patients with head and neck squamous cell carcinoma. Sci Rep. 2018;8(1):1677.
- Kopp JL, Ormsbee BD, Desler M, Rizzino A. Small increases in the level of Sox2 trigger the differentiation of mouse embryonic stem cells. Stem Cells. 2008;26(4):903–911.
- 32. Rimkus TK, Carpenter RL, Qasem S, Chan M, Lo HW. Targeting the sonic hedgehog signaling pathway: review of smoothened and GLI inhibitors. Cancers (Basel). 2016;8(2):E22.
- 33. Kar S, Sengupta D, Deb M, Pradhan N, Patra SK. SOX2 function and Hedgehog signaling pathway are co-conspirators in promoting androgen independent prostate cancer. Biochim Biophys Acta Mol Basis Dis. 2017;1863(1):253–265.
- Sarlak G, Vincent B. The roles of the stem cell-controlling sox2 transcription Factor: from neuroectoderm development to Alzheimer's disease? Mol Neurobiol. 2016;53(3): 1679–1698.
- Schaefer T, Lengerke C. SOX2 protein biochemistry in stemness, reprogramming, and cancer: the PI3K/AKT/SOX2 axis and beyond. Oncogene. 2020;39(2):278–292.
- 36. Boyer LA, Plath K, Zeitlinger J, Brambrink T, Medeiros LA, Lee TI, Levine SS, Wernig M, Tajonar A, Ray MK, Bell GW, et al. Polycomb complexes repress developmental regulators in murine embryonic stem cells. Nature. 2006;441(7091): 349–353.
- 37. Basu-Roy U, Seo E, Ramanathapuram L, Rapp TB, Perry JA, Orkin SH, Mansukhani A, Basilico C. Sox2 maintains self

renewal of tumor-initiating cells in osteosarcomas. Oncogene. 2012;31(18):2270–2282.

- Tang Y, Tian XC. JAK-STAT3 and somatic cell reprogramming. JAKSTAT 2013;2(4):e24935.
- Gordeeva O. TGFbeta Family Signaling pathways in pluripotent and teratocarcinoma stem cells' fate decisions: balancing between self-renewal, differentiation, and cancer. Cells 2019; 8(12):E1500.
- Ormsbee Golden BD, Wuebben EL, Rizzino A. Sox2 expression is regulated by a negative feedback loop in embryonic stem cells that involves AKT signaling and FoxO1. PLoS One. 2013;8(10):e76345.
- 41. Gao Z, Cox JL, Gilmore JM, Ormsbee BD, Mallanna SK, Washburn MP, Rizzino A. Determination of protein interactome of transcription factor Sox2 in embryonic stem cells engineered for inducible expression of four reprogramming factors. J Biol Chem. 2012;287(14):11384–11397.
- Hanna J, Saha K, Pando B, van Zon J, Lengner CJ, Creyghton MP, van Oudenaarden A, Jaenisch R. Direct cell reprogramming is a stochastic process amenable to acceleration. Nature. 2009;462(7273):595–601.
- Narayan S, Bryant G, Shah S, Berrozpe G, Ptashne M. OCT4 and SOX2 work as transcriptional activators in reprogramming human fibroblasts. Cell Rep. 2017;20(7):1585–1596.
- Zhou HY, Katsman Y, Dhaliwal NK, Davidson S, Macpherson NN, Sakthidevi M, Collura F, Mitchell JA. A Sox2 distal enhancer cluster regulates embryonic stem cell differentiation potential. Genes Dev. 2014;28(24):2699–2711.
- 45. Vencken SF, Sethupathy P, Blackshields G, Spillane C, Elbaruni S, Sheils O, Gallagher MF, O'Leary JJ. An integrated analysis of the SOX2 microRNA response program in human pluripotent and nullipotent stem cell lines. BMC Genomics. 2014;15:711.
- 46. Wang G, Guo X, Hong W, Liu Q, Wei T, Lu C, Gao L, Ye D, Zhou Y, Chen J, Wang J, et al. Critical regulation of miR-200/ ZEB2 pathway in Oct4/Sox2-induced mesenchymal-toepithelial transition and induced pluripotent stem cell generation. Proc Natl Acad Sci U S A. 2013;110(8):2858–2863.
- Xu N, Papagiannakopoulos T, Pan G, Thomson JA, Kosik KS. MicroRNA-145 regulates OCT4, SOX2, and KLF4 and represses pluripotency in human embryonic stem cells. Cell. 2009;137(4):647–658.
- 48. Marques-Torrejon MA, Porlan E, Banito A, Gomez-Ibarlucea E, Lopez-Contreras AJ, Fernandez-Capetillo O, Vidal A, Gil J, Torres J, Farinas I. Cyclin-dependent kinase inhibitor p21 controls adult neural stem cell expansion by regulating Sox2 gene expression. Cell Stem Cell. 2013;12(1):88–100.
- Seisenberger S, Peat JR, Hore TA, Santos F, Dean W, Reik W. Reprogramming DNA methylation in the mammalian life cycle: building and breaking epigenetic barriers. Philos Trans R Soc Lond B Biol Sci. 2013;368(1609):20110330.
- Huang Y, Liu H, Du H, Zhang W, Kang X, Luo Y, Zhou X, Li L. Developmental features of DNA methylation in CpG islands of human gametes and preimplantation embryos. Exp Ther Med. 2019;17(6):4447–4456.

- Keramari M, Razavi J, Ingman KA, Patsch C, Edenhofer F, Ward CM, Kimber SJ. Sox2 is essential for formation of trophectoderm in the preimplantation embryo. PLoS One. 2010; 5(11):e13952.
- Boyer LA, Lee TI, Cole MF, Johnstone SE, Levine SS, Zucker JP, Guenther MG, Kumar RM, Murray HL, Jenner RG, Gifford DK, et al. Core transcriptional regulatory circuitry in human embryonic stem cells. Cell 2005;122(6):947–956.
- Guo Z, Packard A, Krolewski RC, Harris MT, Manglapus GL, Schwob JE. Expression of pax6 and sox2 in adult olfactory epithelium. J Comp Neurol. 2010;518(21):4395–4418.
- 54. Kim BE, Choi SW, Shin JH, Kim JJ, Kang I, Lee BC, Lee JY, Kook MG, Kang KS. Single-factor SOX2 mediates direct neural reprogramming of human mesenchymal stem cells via transfection of in vitro transcribed mRNA. Cell Transplant. 2018;27(7):1154–1167.
- 55. Hao Q, Zheng J, Hu Y, Wang H. Bone marrow mesenchymal stem cells combined with Sox2 increase the functional recovery in rat with traumatic brain injury. Chinese Neurosurgical Journal. 2019;5(1):11.
- Yuan HF, Zhai C, Yan XL, Zhao DD, Wang JX, Zeng Q, Chen L, Nan X, He LJ, Li ST. SIRT1 is required for long-term growth of human mesenchymal stem cells. J Mol Med (Berl) 2012;90(4):389–400.
- 57. Yoon DS, Choi Y, Jang Y, Lee M, Choi WJ, Kim SH, Lee JW. SIRT1 directly regulates SOX2 to maintain self-renewal and multipotency in bone marrow-derived mesenchymal stem cells. Stem Cells. 2014;32(12):3219–3231.
- Go MJ, Takenaka C, Ohgushi H. Forced expression of Sox2 or Nanog in human bone marrow derived mesenchymal stem cells maintains their expansion and differentiation capabilities. Exp Cell Res. 2008;314(5):1147–1154.
- Jiang L, Li R, Tang H, Zhong J, Sun H, Tang W, Wang H, Zhu J. MRI tracking of iPS cells-induced neural stem cells in traumatic brain injury rats. Cell Transplant 2019;28(6):747–755.
- Niu W, Zang T, Smith DK, Vue TY, Zou Y, Bachoo R, Johnson JE, Zhang CL. SOX2 reprograms resident astrocytes into neural progenitors in the adult brain. Stem Cell Reports. 2015; 4(5):780–794.
- Yen SY, Chuang HM, Huang MH, Lin SZ, Chiou TW, Harn HJ. n-Butylidenephthalide regulated tumor stem cell genes EZH2/AXL and reduced Its migration and invasion in glioblastoma. Int J Mol Sci. 2017;18(2):E372.
- 62. Ferone G, Song JY, Sutherland KD, Bhaskaran R, Monkhorst K, Lambooij JP, Proost N, Gargiulo G, Berns A. SOX2 Is the determining oncogenic switch in promoting lung squamous cell carcinoma from different cells of origin. Cancer Cell. 2016;30(4):519–532.
- 63. Wu F, Zhang J, Wang P, Ye X, Jung K, Bone KM, Pearson JD, Ingham RJ, McMullen TP. Identification of two novel phenotypically distinct breast cancer cell subsets based on Sox2 transcription activity. Cell Signal. 2012;24(11):1989–1998.
- 64. Gangemi RM, Griffero F, Marubbi D, Perera M, Capra MC, Malatesta P, Ravetti GL, Zona GL, Daga A, Corte G. SOX2 silencing in glioblastoma tumor-initiating cells causes stop of

proliferation and loss of tumorigenicity. Stem Cells. 2009; 27(1):40-48.

- 65. Liu K, Lin B, Zhao M, Yang X, Chen M, Gao A, Liu F, Que J, Lan X. The multiple roles for Sox2 in stem cell maintenance and tumorigenesis. Cell Signal. 2013;25(5): 1264–1271.
- Weina K, Utikal J. SOX2 and cancer: current research and its implications in the clinic. Clin Transl Med. 2014;3:19.
- 67. Chen S, Xu Y, Chen Y, Li X, Mou W, Wang L, Liu Y, Reisfeld RA, Xiang R, Lv D. SOX2 gene regulates the transcriptional network of oncogenes and affects tumorigenesis of human lung cancer cells. PLoS One. 2012;7(5):e36326.
- Fang X, Yu W, Li L, Shao J, Zhao N, Chen Q, Ye Z, Lin SC, Zheng S, Lin B. ChIP-seq and functional analysis of the SOX2 gene in colorectal cancers. OMICS. 2010;14(4):369–384.
- 69. Engelen E, Akinci U, Bryne JC, Hou J, Gontan C, Moen M, Szumska D, Kockx C, van Ijcken W, Dekkers DH. Sox2 cooperates with Chd7 to regulate genes that are mutated in human syndromes. Nat Genet. 2011;43(6):607–611.
- Hagerstrand D, He X, Bradic Lindh M, Hoefs S, Hesselager G, Ostman A, Nister M. Identification of a SOX2-dependent subset of tumor- and sphere-forming glioblastoma cells with a distinct tyrosine kinase inhibitor sensitivity profile. Neuro Oncol. 2011;13(11):1178–1191.
- Otsubo T, Akiyama Y, Yanagihara K, Yuasa Y. SOX2 is frequently downregulated in gastric cancers and inhibits cell growth through cell-cycle arrest and apoptosis. Br J Cancer. 2008;98(4):824–831.
- Tompkins DH, Besnard V, Lange AW, Keiser AR, Wert SE, Bruno MD, Whitsett JA. Sox2 activates cell proliferation and differentiation in the respiratory epithelium. Am J Respir Cell Mol Biol. 2011;45(1):101–110.
- 73. Ring KL, Tong LM, Balestra ME, Javier R, Andrews-Zwilling Y, Li G, Walker D, Zhang WR, Kreitzer AC, Huang Y. Direct reprogramming of mouse and human fibroblasts into multipotent neural stem cells with a single factor. Cell Stem Cell. 2012; 11(1):100–109.
- 74. Yang Y, Zhao Y, Liu X, Chen Y, Liu P, Zhao L. Effect of SOX2 on odontoblast differentiation of dental pulp stem cells. Mol Med Rep. 2017;16(6):9659–9663.
- Julian LM, McDonald AC, Stanford WL. Direct reprogramming with SOX factors: masters of cell fate. Curr Opin Genet Dev. 2017;46:24–36.
- Graham V, Khudyakov J, Ellis P, Pevny L. SOX2 functions to maintain neural progenitor identity. Neuron. 2003;39(5): 749–765.
- 77. Zappone MV, Galli R, Catena R, Meani N, De Biasi S, Mattei E, Tiveron C, Vescovi AL, Lovell-Badge R, Ottolenghi S, Nicolis SK. Sox2 regulatory sequences direct expression of a (beta)-geo transgene to telencephalic neural stem cells and precursors of the mouse embryo, revealing regionalization of gene expression in CNS stem cells. Development. 2000; 127(11):2367–2382.

- Hagey DW, Muhr J. Sox2 acts in a dose-dependent fashion to regulate proliferation of cortical progenitors. Cell Rep. 2014; 9(5):1908–1920.
- Julian LM, Vandenbosch R, Pakenham CA, Andrusiak MG, Nguyen AP, McClellan KA, Svoboda DS, Lagace DC, Park DS, Leone G. Opposing regulation of Sox2 by cell-cycle effectors E2f3a and E2f3b in neural stem cells. Cell Stem Cell 2013; 12(4):440–452.
- Das T, Nair RR, Green R, Padhee S, Howell M, Banerjee J, Mohapatra SS, Mohapatra S. Actinomycin D Down-regulates SOX2 Expression and Induces Death in Breast Cancer Stem Cells. Anticancer Res. 2017;37(4):1655–1663.
- Li ZR, Jiang Y, Hu JZ, Chen Y, Liu QZ. SOX2 knockdown inhibits the migration and invasion of basal cell carcinoma cells by targeting the SRPK1-mediated PI3K/AKT signaling pathway. Oncol Lett. 2019;17(2):1617–1625.
- Mu P, Zhang Z, Benelli M, Karthaus WR, Hoover E, Chen CC, Wongvipat J, Ku SY, Gao D, Cao Z. SOX2 promotes lineage plasticity and antiandrogen resistance in TP53- and RB1deficient prostate cancer. Science. 2017;355(6320):84–88.
- Li D, Zhao LN, Zheng XL, Lin P, Lin F, Li Y, Zou HF, Cui RJ, Chen H, Yu XG. Sox2 is involved in paclitaxel resistance of the prostate cancer cell line PC-3 via the PI3K/Akt pathway. Mol Med Rep. 2014;10(6):3169–3176.
- 84. Vanner RJ, Remke M, Gallo M, Selvadurai HJ, Coutinho F, Lee L, Kushida M, Head R, Morrissy S, Zhu X. Quiescent sox2(+) cells drive hierarchical growth and relapse in sonic hedgehog subgroup medulloblastoma. Cancer Cell 2014;26(1): 33–47.
- Singh DK, Kollipara RK, Vemireddy V, Yang XL, Sun Y, Regmi N, Klingler S, Hatanpaa KJ, Raisanen J. Oncogenes activate an autonomous transcriptional regulatory circuit that drives glioblastoma. Cell Rep. 2017;18(4):961–976.
- Dhodapkar KM, Gettinger SN, Das R, Zebroski H, Dhodapkar MV. SOX2-specific adaptive immunity and response to immunotherapy in non-small cell lung cancer. Oncoimmunology. 2013;2(7) e25205.
- Mandai M, Kurimoto Y, Takahashi M. Autologous Induced Stem-Cell-Derived Retinal Cells for Macular Degeneration. N Engl J Med. 2017;377(8):792–793.
- Sharma R, Khristov V, Rising A, Jha BS, Dejene R, Hotaling N, Li Y, Stoddard J, Stankewicz C, Wan Q. Clinical-grade stem cell-derived retinal pigment epithelium patch rescues retinal degeneration in rodents and pigs. Sci Transl Med. 2019; 11(475).
- Larsson HM, Lee ST, Roccio M, Velluto D, Lutolf MP, Frey P, Hubbell JA. Sorting live stem cells based on Sox2 mRNA expression. PLoS One. 2012;7(11): e49874.
- 90. Malinee M, Kumar A, Hidaka T, Horie M, Hasegawa K, Pandian GN, Sugiyama H. Targeted suppression of metastasis regulatory transcription factor SOX2 in various cancer cell lines using a sequence-specific designer pyrrole-imidazole polyamide. Bioorg Med Chem. 2020;28(3):115248.