

# **Urological Oncology**

# Diagnostic Utility of Caveolin-1 and MOC-31 in Distinguishing Chromophobe Renal Cell Carcinoma from Renal Oncocytoma

# Hyoun Wook Lee, Eun Hee Lee, Chang Hun Lee<sup>1</sup>, Hee Kyung Chang<sup>2</sup>, Seo Hee Rha<sup>3</sup>

Department of Pathology, Samsung Changwon Hospital, Sungkyunkwan University School of Medicine, Changwon, <sup>1</sup>Pusan National University Hospital, School of Medicine Pusan National University, <sup>2</sup>Kosin University College of Medicine, <sup>3</sup>Dong-A University College of Medicine, Busan, Korea

**Purpose:** Renal tumors consist of heterogeneous groups that frequently show complex and overlapping morphology, thus making it difficult to make a correct diagnosis. One of the most problematic differential diagnoses is to distinguish chromophobe renal cell carcinoma (RCC) from oncocytoma. These should be distinguished by differences in their behavior and clinical outcome. Our study was performed to identify whether caveolin-1 and MOC-31 are useful immunohistochemical markers for differentiating chromophobe RCC from oncocytoma.

**Materials and Methods:** We selected 23 chromophobe RCCs, 8 oncocytomas, and 25 clear cell RCCs and performed immunohistochemical staining for caveolin-1 and MOC-31.

**Results:** Caveolin-1 was positive in 20 (87%) of 23 chromophobe RCCs, 0 of 8 oncocytomas, and 21 (84%) of 25 clear cell RCCs. MOC-31 was positive in 22 (96%) of 23 chromophobe RCCs, 2 (25%) of 8 oncocytomas, and 14 (56%) of 25 clear cell RCCs. There was a statistically significant difference in the expression of caveolin-1 and MOC-31 between chromophobe RCC and oncocytoma (p < 0.001). In addition, clear cell RCC was also significantly different from oncocytoma in the expression of caveolin-1 (p < 0.001) and was significantly different from chromophobe RCC in the expression of MOC-31 (p < 0.001).

**Conclusions:** Caveolin-1 and MOC-31 can be useful markers in the differential diagnosis of chromophobe RCC, oncocytoma, and clear cell RCC.

**Key Words:** Adenoma, oxyphilic; Caveolin 1; MOC-31 monoclonal antibody, human; Renal cell carcinoma

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

### **Article History:**

received 10 November, 2010 accepted 21 January, 2011

### Corresponding Author:

Hyoun Wook Lee
Department of Pathology, Samsung
Changwon Hospital, Sungkyunkwan
University School of Medicine, 50,
Hapsung-dong, Masan Hoewon-gu,
Changwon 630-723, Korea
TEL: +82-55-290-6497

FAX: +82-55-290-6418 E-mail: sudowo@nate.com

### INTRODUCTION

Renal neoplasms are composed of heterogeneous groups including clear cell renal cell carcinoma (RCC), papillary RCC, chromophobe RCC, collecting duct carcinoma, oncocytoma, and others. In most cases, these categories are easily differentiated from each other on the basis of histologic features alone. Occasionally, however, there are overlapping morphological characteristics between such tumors as well as histologic heterogeneity within a single tumor. In those cases, it may be difficult to make an accu-

rate subtyping of the tumors.

A diagnostic problem can arise in distinguishing chromophobe RCC from oncocytoma. These two types of tumors have a common origin, distal nephrons, and similar phenotype, so it may be difficult to distinguish these tumors in the practice of diagnostic pathology. Chromophobe RCC and oncocytoma should be distinguished from each other because both tumors have different behaviors and clinical outcomes. Oncocytoma is a benign tumor, whereas chromophobe RCC is a subtype of RCC, namely, malignant. In recent years, several studies have been carried out to find

helpful ancillary means to distinguish chromophobe RCC from oncocytoma. However, the results of these studies are inconsistent and still unsatisfactory [1-8].

Caveolin-1, a 24-kd membrane protein, is a major structural and functional protein component of caveolae, endocytic structures of the cell membrane [9,10]. Caveolin-1 plays important roles in membrane traffic, lipid traffic, and signal transduction [9,10]. Several studies have shown that the expression of caveolin-1 is elevated in various types of malignancies such as prostate, breast, colon, esophagus, and urinary bladder cancers [11-14], but the precise roles of caveolin-1 in carcinogenesis are still unclear.

MOC-31 is a kind of cell surface glycoprotein and is expressed in most benign and malignant epithelia. It is known that MOC-31 is useful for distinguishing adenocarcinoma from mesothelioma as well as hepatocellular carcinoma from cholangiocarcinoma and metastatic adenocarcinoma in the liver [15,16].

In this study, we intended to identify whether there were

any differences in immunohistochemical reactivity for caveolin-1 and MOC-31 between chromophobe RCC and oncocytoma and to determine whether the expression of these proteins has a benefit as useful immunohistochemical markers in distinguishing the tumors. In addition, clear cell RCC, particularly the granular cell type, may also have a phenotype similar to the previously mentioned two types of tumors and cause a diagnostic problem. Accordingly, we also investigated the immunohistochemical reactivity of clear cell RCC for caveolin-1 and MOC-31 and evaluated the differences among clear cell RCC, chromophobe RCC, and oncocytoma.

### MATERIALS AND METHODS

### 1. Case selection

Twenty-three chromophobe RCCs and 8 oncocytomas were retrieved from the surgical pathology archives between 1997 and 2006 at three institutes. Twenty-five clear cell

Table 1. Clinicopathologic data of chromophobe RCC and oncocytoma

Case No.	Age (yr)/ Sex	Tumor size (cm)	Perinephric tissue invasion	Renal vein invasion	Tumor stage	EM	Caveolin-1	MOC-31
Chromoph	obe RCC							
1	67/M	8.0	_	_	pT2a	Done	Diffuse positive	Diffuse positive
2	76/M	5.4	_	_	pT1b	Not done	Diffuse positive	Negative
3	47/F	4.5	_	_	pT1b	Done	Diffuse positive	Diffuse positive
4	64/M	3.7	_	_	pT1a	Done	Negative	Diffuse positive
5	70/M	9.5	_	_	pT2a	Not done	Diffuse positive	Diffuse positive
6	49/M	5.0	_	_	pT1b	Not done	Diffuse positive	Diffuse positive
7	41/F	7.2	_	_	pT2a	Not done	Diffuse positive	Diffuse positive
8	17/M	8.0	_	_	pT2a	Not done	Diffuse positive	Diffuse positive
9	59/M	15.0	_	_	pT2b	Not done	Diffuse positive	Diffuse positive
10	43/F	10.0	_	_	pT2a	Not done	Diffuse positive	Diffuse positive
11	65/M	3.5	_	_	pT1a	Not done	Diffuse positive	Diffuse positive
12	63/F	6.5	_	_	pT2b	Not done	Diffuse positive	Diffuse positive
13	41/F	5.0	_	_	pT1b	Not done	Diffuse positive	Diffuse positive
14	39/F	24.0	+	+	pT4	Not done	Negative	Diffuse positive
15	64/M	2.2	_	_	pT1a	Not done	Diffuse positive	Diffuse positive
16	49/F	4.4	_	_	pT1b	Not done	Diffuse positive	Diffuse positive
17	41/F	2.5	+	_	pT3a	Not done	Negative	Diffuse positive
18	43/F	3.3	_	_	pT1a	Not done	Diffuse positive	Diffuse positive
19	40/F	5.0	_	_	pT1b	Not done	Diffuse positive	Diffuse positive
20	65/M	13.0	+	+	pT3a	Not done	Diffuse positive	Diffuse positive
21	49/F	11.5	_	_	pT2b	Not done	Diffuse positive	Diffuse positive
22	42/M	5.0	_	+	pT3a	Not done	Diffuse positive	Diffuse positive
23	77/M	5.0	+	+	pT3a	Not done	Diffuse positive	Diffuse positive
Oncocytom	ıa				-		-	-
1	46/M	4.5	_	_		Done	Negative	Negative
<b>2</b>	68/M	5.8	_	_		Done	Negative	Negative
3	65/F	3.5	_	_		Not done	Negative	Negative
4	80/F	2.7	_	_		Done	Negative	Negative
5	38/M	3.0	_	_		Not done	Negative	Negative
6	45/F	2.0	_	_		Done	Negative	Negative
7	52/F	4.4	_	_		Not done	Negative	Diffuse positive
8	64/F	5.0	_	_		Not done	Negative	Diffuse postive

RCC: renal cell carcinoma, EM: electron microscopic examination, M: male, F: female

98 Lee et al

RCCs, including 10 cases of clear cell type and 15 cases of granular cell type, were also selected from the surgical pathology files of one institute. Those cases were composed of only radical surgical specimens. Two pathologists, one of whom has expertise in genitourinary pathology, reviewed all cases and achieved consensus on classification according to the recommendations of the International Union Against Cancer [17] and the World Health Organization Classification of Tumors [18]. Most tumors were categorized on the basis of light microscopic histology. Electron microscopic examination was performed for 9 cases because they showed obscure and overlapping histologic features on light microscopic, histochemical, and immunohistochemical examinations. Two cases having unclear features despite the electron microscopic examination were excluded from this study. Staging of chromophobe RCC was performed according to the recently updated staging system (7th edition) of the American Joint Committee on Cancer (AJCC) (7th edition) and the Union Internationale Contre le Cancer (UICC) [19]. The clinicopathologic data of all chromophobe RCCs and oncocytomas are summarized in Table 1.

### 2. Immunohistochemistry

The immunohistochemical staining was performed with the DAKO EnVision Kit (Dako, Denmark) on sections prepared from formalin-fixed and paraffin-embedded specimens that were dewaxed and rehydrated with graded concentrations of alcohol. Endogenous peroxidase was blocked by dipping the sections in 3% aqueous hydrogen peroxide for 10 min, and antigen retrieval was performed with a 10-min microwave treatment in 10 mmol/l citrate buffer, pH 6.0. For immunohistochemical staining of MOC-31, Target Retrieval Solution (Dako, Denmark) was used for antigen retrieval. Diluted primary antibodies for caveolin-1 (1:200, Transduction Laboratories, US) and MOC-31 (1:50, Dako, Denmark) were treated at room temper-

ature for 30 min and 1 h, respectively. After the primary antibody incubation, the sections were incubated with the secondary antibody. The sections were lightly counterstained with hematoxylin.

Immunoreactivity for caveolin-1 and MOC-31 was considered as diffuse positive when 50% or more of the tumor cells were stained, as focal positive when 10% to 49% of the tumor cells were stained, and as negative when less than 10% of the tumor cells were stained. Internal positive controls were endothelial cells and distal tubular epithelial cells for caveolin-1 and MOC-31, respectively.

## 3. Statistical analysis

Data were analyzed by the Fisher's exact test with SPSS ver. 12.0 (SPSS Inc., Chicago, IL, USA) to evaluate correlation between the tumor types and expression of immunohistochemical stain for caveolin-1 and MOC-31. p-values less than 0.05 were considered statistically significant.

### **RESULTS**

### 1. Immunoreactivity for Caveolin-1

In normal kidney tissue, expression of caveolin-1 was seen

TABLE 2. Expression of caveolin-1 and MOC-31 in normal kidney tissue

	Caveolin-1	MOC-31
Glomerular capillary	_	_
Visceral epithelium	_	_
Parietal epithelium	+	_
Proximal tubule	_	_
Distal tubule	_	+
Collecting duct	_	+
Endothelial cell	+	_
Smooth muscle cell	+	_

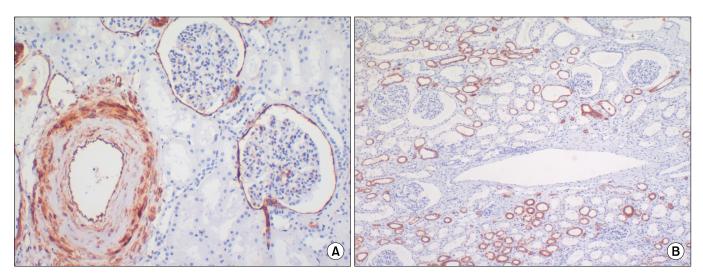


FIG. 1. Expression of caveolin-1 and MOC-31 in normal kidney tissue. (A) Caveolin-1 is expressed in parietal epithelial cells of Bowman capsules, endothelial cells, and smooth muscle cells of blood vessels. (B) MOC-31 is expressed only in epithelial cells of distal tubules.

in parietal epithelia of Bowman capsules, endothelial cells, and smooth muscle cells of blood vessels but not in endothelial cells of glomeruli or any other epithelia (Table 2, Fig. 1A).

In chromophobe RCCs, 20 (87%) of 23 cases showed diffuse positivity for caveolin-1. All positive cases showed expression of caveolin-1 in 90% or more of the tumor cells and moderate to strong intensity except for one case that had relatively weak intensity. The pattern of immunohistochemical staining was diffusely cytoplasmic and granular. In most of the cases, the cell membrane was also stained and the staining intensity was stronger in the membrane than in the cytoplasm (Fig. 2A). In the three negative cases, none of the tumor cells expressed caveolin-1.

Oncocytomas showed negative results for caveolin-1 in all eight cases (Fig. 2B). None of the tumor cells expressed caveolin-1 except for one case in which caveolin-1 was expressed in only 5% or less of the tumor cells.

Clear cell RCCs were diffuse positive in 19, focal positive in 2, and negative in 4. The total number of positive cases was 21 (84%) of 25 clear cell RCCs. All 10 cases of the clear cell type were diffuse positive, whereas 9 (60%) of 15 cases of the granular cell type were diffuse positive. There was a slight difference in the pattern of immunohistochemical staining between the clear and granular cell types. All cases of the clear cell type showed dominantly membranous staining without cytoplasmic staining (Fig. 3A), whereas most cases of the granular cell type exhibited a similar staining pattern to that of chromophobe RCC, namely, a diffusely cytoplasmic and granular pattern with focal membranous condensation (Fig. 3B).

The incidence of caveolin-1 expression showed statisti-

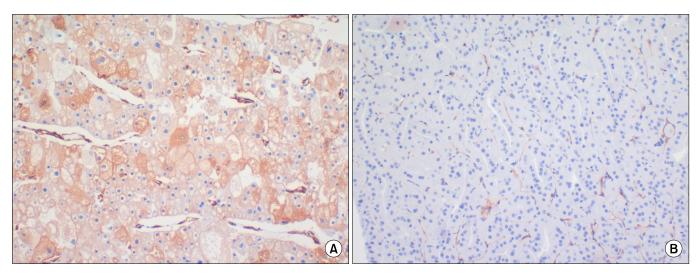


FIG. 2. Expression of caveolin-1 in chromophobe renal cell carcinoma and oncocytoma. (A) Chromophobe renal cell carcinoma is diffuse, moderate to strong positive for caveolin-1, showing granular and cytoplasmic staining with membranous condensation. (B) Oncocytoma is negative for caveolin-1.

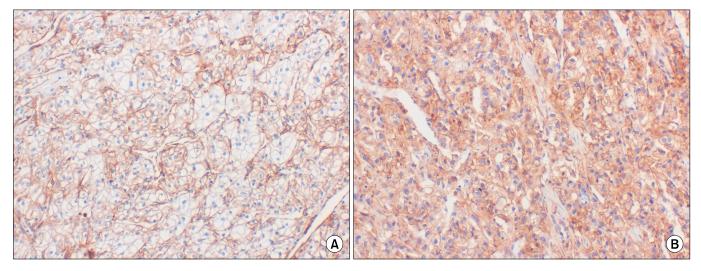


FIG. 3. Expression of caveolin-1 in clear cell renal cell carcinoma. (A) Clear cell type shows membranous staining of caveolin-1. (B) Granular cell type shows diffuse cytoplasmic staining with membranous condensation similar to chromophobe renal cell carcinoma.

100 Lee et al

cally significant differences between chromophobe RCCs and oncocytomas (p < 0.001) and between clear cell RCCs and oncocytomas (p < 0.001) but not between chromophobe and clear cell RCCs (p=0.466) (Table 3, 4).

### 2. Immunoreactivity for MOC-31

MOC-31 was positive in distal tubules and collecting ducts of normal kidney tissue but negative in other epithelial and mesenchymal tissues (Table 2, Fig. 1B).

MOC-31 was diffuse positive in 22~(96%) of 23 chromophobe RCCs, showing a strong membranous pattern of immunohistochemical staining (Fig. 4A), and negative in only 1 case.

In oncocytomas, MOC-31 was negative in 6 of 8 cases and diffuse positive in the other 2 cases (25%) (Fig. 4B). Three of the six negative cases expressed MOC-31 in less than 10% of the tumor cells, and the other three cases showed no staining. In the former three cases, MOC-31 was characteristically expressed in the tumor cells showing tubular structures but not solid nests or trabecular structures. In addition, two oncocytomas showing diffuse expression of MOC-31 were composed of predominantly tubular structures, and solid or trabecular structures were only a minor portion. MOC-31 was expressed in such tubular structures rather than in solid or trabecular structures (Fig. 5).

Clear cell RCCs showed relatively variable MOC31 staining results. Clear cell RCCs were diffuse positive for MOC-31 in 12, focal positive in 2, and negative in 11. The total number of positive cases was 14 (56%) of 25 cases. In particular, the granular cell type of clear cell RCC showed less positivity for MOC-31 than did the clear cell type. Only 6 (40%) of 15 cases of the granular cell type were positive for MOC-31. All positive cases showed a membranous staining pattern as for the chromophobe RCCs.

The incidence of MOC-31 expression showed statistically significant differences between chromophobe RCCs and oncocytomas (p  $\!<\!0.001)$  and between chromophobe and clear cell RCCs (p  $\!<\!0.001)$  but not between clear cell

TABLE 4. Statistical results of the Fisher's exact test

		p-value			
	Chromophobe	Chromophobe	Clear cell		
	RCC vs.	RCC vs. Clear	RCC vs.		
	Oncocytoma	cell RCC	Oncocytoma		
Caveolin-1	<0.001	0.466	< 0.001		
MOC-31	<0.001	<0.001	0.215		

RCC: renal cell carcinoma

Table 3. Results of immunohistochemical staining for caveolin-1 and MOC-31

	Caveolin-1, n (%)			MOC-31, n (%)		
	Negative	Focal positive	Diffuse positive	Negative	Focal positive	Diffuse positive
Chromophobe RCC (n=23)	3 (13)	0 (0)	20 (87)	1 (4)	0 (0)	22 (96)
Oncocytoma (n=8)	8 (100)	0 (0)	0 (0)	6 (75)	0 (0)	2(25)
Clear cell RCC (n=25)	4 (16)	2(8)	19 (76)	11 (44)	2(8)	12 (48)
Clear (n=10)	0 (0)	0 (0)	10 (100)	2(20)	0 (0)	8 (80)
Granular (n=15)	4(27)	2(13)	9 (60)	9 (60)	2 (13)	4(27)

RCC: renal cell carcinoma

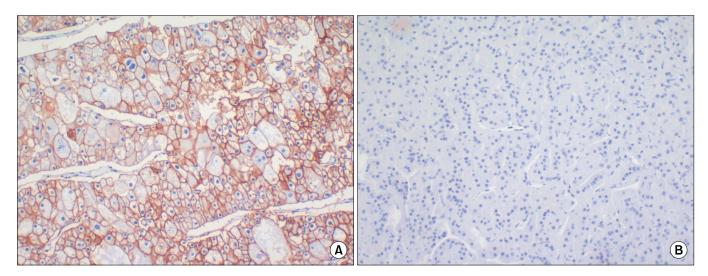
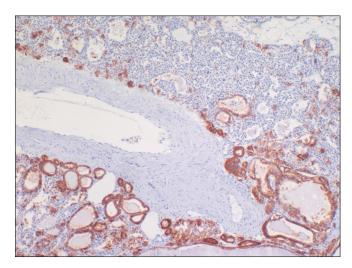


FIG. 4. Expression of MOC-31 in chromophobe renal cell carcinoma and oncocytoma. (A) Chromophobe renal cell carcinoma shows diffuse, strong, and membranous staining for MOC-31. (B) Oncocytoma shows no staining for MOC-31.



**FIG. 5.** Expression of MOC-31 in oncocytoma with tubular differentiation. In a few cases of oncocytomas, MOC-31 is expressed in the tumor cells showing tubular differentiation but not solid or trabecular nests.

RCCs and oncocytomas (p=0.215) (Table 3, 4).

### DISCUSSION

Subtypes of renal cell tumors are various and complex. Although each renal cell tumor has unique characteristics, they may occasionally have overlapping histologic features that make it difficult to make a proper diagnosis. One of the most problematic differential diagnoses is to distinguish chromophobe RCC from oncocytoma.

Chromophobe RCC was first described by Thoenes et al in 1985 [20]. Chromophobe RCC consists of tumor cells showing abundant pale cytoplasm with a perinuclear halo and peripheral condensation and hyperchromatic, irregular, wrinkled nuclei. By contrast, oncocytomas have tumor cells showing abundant eosinophilic granular cytoplasm and regular and round nuclei. Both tumors can be distinguished on the basis of these histologic features. However, occasionally, when the tumors show obscure microscopic findings, it can be difficult to make a differential diagnosis. Therefore, many investigators have searched for helpful ancillary means to discriminate these two types of renal tumors.

Electron microscopic examination is an important and useful tool in the differential diagnosis of chromophobe RCC and oncocytoma. However, electron microscopic examination is technically difficult, expensive, and not available in many laboratories. Several studies have suggested that Hale's colloidal iron stain is helpful for the differential diagnosis of chromophobe RCC from oncocytoma [1]. However, Hale's colloidal iron stain is also technically demanding and difficult to interpret, and therefore reproducibility is relatively low [2].

Several studies with various immunohistochemical markers, including kidney-specific cadherin, CK7, EMA, CD10, parvalbumin, RCC, c-KIT, and RON proto-onco-

gene, have been performed to find useful markers for distinguishing chromophobe RCC from oncocytoma. However, the results of these studies are inconsistent or turned out to be not satisfactory. Accordingly, the use of these markers in differential diagnosis is limited [2-8,21,22].

Caveolin-1 is a major protein component of caveolae, which are endocytic structures of the cell membrane, and is localized to the cytosolic surface of caveolae. Caveolin-1 interacts with both lipids and lipid anchors on proteins; therefore, it has multiple functions in signal transduction, membrane traffic, and intracellular and extracellular lipid transport [9,10]. Expression of this molecule has been studied in various types of tumors, including prostate, breast, colon, esophagus, and urinary bladder cancers [11-14]. However, only a small number of studies have recently reported a relationship between the expression of caveolin-1 and renal neoplasms, including chromophobe RCC and oncocytoma [23-25].

In the present study, caveolin-1 was expressed in 20 (87%) of 23 chromophobe RCCs but in 0 of 8 oncocytomas. All positive cases of chromophobe RCC showed diffuse, moderate to strong staining in 90% or more of tumor cells and cytoplasmic and granular staining with membranous condensation. This clear discrimination enables the use of caveolin-1 for differential diagnosis between chromophobe RCC and oncocytoma (p < 0.001).

Carrion et al and Mete et al reported that most oncocytomas express caveolin-1 with a diffuse cytoplasmic staining pattern and that chromophobe RCCs show a relatively decreased expression of caveolin-1 [23,24]. This difference probably originated from the artifact that they did not use a blocking step for endogenous biotin despite using an avidin-biotin peroxidase kit. Endogenous biotin in oncocytomas may cause a false-positive result. Nevertheless, there was a difference in staining pattern between chromophobe RCCs and oncocytomas in their studies. According to them, chromophobe RCCs showed peripheral cytoplasmic staining, whereas oncocytomas showed diffuse cytoplasmic staining. There was an obvious difference in the incidence and staining pattern of caveolin-1 expression between both tumors.

In contrast with these findings, Garcia and Li reported that caveolin-1 was positive in all 21 chromophobe RCCs but was focal positive, with staining in less than 20% of the tumor cells, in only 3 of 26 oncocytomas [25]. This result is consistent with and supports our result.

Occasionally, it may also be difficult to distinguish clear cell RCC, particularly the granular cell type, from chromophobe RCC or oncocytoma in practice. Therefore, our present study also included clear cell RCCs. Twenty-one (84%) of 25 clear cell RCCs were positive for caveolin-1, including 2 focal positive cases. Granular cell type tumors of clear cell RCCs were very similar to chromophobe RCC in staining pattern as well as extent, whereas clear cell type tumors differed from chromophobe RCC, that is, showing membranous staining. On the basis of these results, the granular cell type of clear cell RCC could be differentiated from

102 Lee et al

oncocytoma (p < 0.001) but not chromophobe RCC (p= 0.466).

MOC-31, a kind of cell surface glycoprotein, is expressed in various benign and malignant epithelia, but its function is still unclear. MOC-31 has been studied for differential diagnosis between hepatocellular carcinoma and cholangiocarcinoma or metastatic adenocarcinoma in the liver or between mesothelioma and adenocarcinoma [15,16]. Studies associated with renal neoplasms have rarely been performed.

In our study, MOC-31 showed diffuse, strong, and membranous staining in 22 (96%) of 23 chromophobe RCCs and diffuse cytoplasmic staining in only 2 (25%) of 8 oncocytomas. The other six cases of oncocytomas were considered to be negative, but three of them expressed MOC31 with cytoplasmic staining in less than 10% of the tumor cells. In these three cases and the diffuse positive two cases of oncocytomas, MOC-31 was predominantly expressed in the tumor cells showing tubular differentiation rather than solid or trabecular structures. This seems to be related to the fact that MOC-31 is expressed in distal tubular cells of normal kidney. Although most oncocytomas do not express MOC-31, some oncocytomas may express MOC-31 if they are differentiated toward tubules and have similar structural features to normal distal tubules. Further molecular analysis is needed to uncover the precise function of MOC-31 in tubular differentiation and tumorigenesis.

According to our result for MOC31 expression, diffuse membranous expression of MOC-31 is a feature of chromophobe RCC rather than of oncocytoma, particularly not showing tubular differentiation (p<0.001). Pan et al showed that 23 (82%) of 28 chromophobe RCCs were positive for MOC-31, whereas all 7 oncocytomas were completely negative [7]. This result is consistent with ours.

Clear cell RCCs expressed MOC-31 in 14 (56%) of 25 cases, including 2 focal positive cases. Considering only the granular cell type, 6 (40%) of 15 cases were positive for MOC-31. As previously mentioned, clear cell RCC had a statistically significant difference in the expression of MOC-31 from chromophobe RCC (p < 0.001) but not oncocytoma (p=0.215). For example, if a tumor is negative for MOC-31, it is more likely to be clear cell RCC than chromophobe RCC.

### **CONCLUSIONS**

The results of the present study showed an obvious discrimination between chromophobe RCC and oncocytoma in the expression of caveolin-1 and MOC-31. In other words, these immunohistochemical markers are thought to be considerably useful for distinguishing these tumors. In addition, clear cell RCC also showed a different expression of caveolin-1 and MOC-31 from oncocytoma and chromophobe RCC, respectively. If used together with other known markers such as c-KIT or RCC marker, both caveolin-1 and MOC-31 may be helpful in the differentiation of clear cell RCC from chromophobe RCC or oncocytoma.

#### **Conflicts of Interest**

The authors have nothing to disclose.

### REFERENCES

- Tickoo SK, Amin MB, Zarbo RJ. Colloidal iron staining in renal epithelial neoplasms, including chromophobe renal cell carcinoma: emphasis on technique and patterns of staining. Am J Surg Pathol 1998;22:419-24.
- Mazal PR, Exner M, Haitel A, Krieger S, Thomson RB, Aronson PS, et al. Expression of kidney-specific cadherin distinguishes chromophobe renal cell carcinoma from renal oncocytoma. Hum Pathol 2005;36:22-8.
- Shen SS, Krishna B, Chirala R, Amato RJ, Truong LD. Kidneyspecific cadherin, a specific marker for the distal portion of the nephron and related renal neoplasms. Mod Pathol 2005;18:933-40.
- Adley BP, Gupta A, Lin F, Luan C, The BT, Yang XJ. Expression of kidney-specific cadherin in chromophobe renal cell carcinoma and renal oncocytoma. Am J Clin Pathol 2006;126:79-85.
- Martignoni G, Pea M, Chilosi M, Brunelli M, Scarpa A, Colato C, et al. Parvalbumin is constantly expressed in chromophobe renal carcinoma. Mod Pathol 2001;14:760-7.
- Langner C, Wegscheider BJ, Ratschek M, Schips L, Zigeuner R. Keratin immunohistochemistry in renal cell carcinoma subtypes and renal oncocytomas: a systematic analysis of 233 tumors. Virchows Arch 2004;444:127-34.
- Pan CC, Chen PC, Ho DM. The diagnostic utility of MOC31, BerEP4, RCC marker and CD10 in the classification of renal cell carcinoma and renal oncocytoma: an immunohistochemical analysis of 328 cases. Histopathology 2004;45:452-9.
- 8. Skinnider BF, Folpe AL, Hennigar RA, Lim SD, Cohen C, Tamboli P, et al. Distribution of cytokeratins and vimentin in adult renal neoplasms and normal renal tissue: potential utility of a cytokeratin antibody panel in the differential diagnosis of renal tumors. Am J Surg Pathol 2005;29:747-54.
- Rothberg KG, Heuser JE, Donzell WC, Ying YS, Glenney JR, Anderson RG. Caveolin, a protein component of caveolae membrane coats. Cell 1992;68:673-82.
- Liu P, Rudick M, Anderson RG. Multiple functions of caveolin-1.
   J Biol Chem 2002;277:41295-8.
- 11. Yang G, Truong LD, Timme TL, Ren C, Wheeler TM, Park SH, et al. Elevated expression of caveolin is associated with prostate and breast cancer. Clin Cancer Res 1998;4:1873-80.
- 12. Fine SW, Lisanti MP, Galbiati F, Li M. Elevated expression of caveolin-1 in adenocarcinoma of the colon. Am J Clin Pathol 2001;115:719-24.
- 13. Kato K, Hida Y, Miyamoto M, Hashida H, Shinohara T, Itoh T, et al. Overexpression of caveolin-1 in esophageal squamous cell carcinoma correlates with lymph node metastasis and pathologic stage. Cancer 2002;94:929-33.
- 14. Fong A, Garcia E, Gwynn L, Lisanti MP, Fazzari MJ, Li M. Expression of caveolin-1 and caveolin-2 in urothelial carcinoma of the urinary bladder correlates with tumor grade and squamous differentiation. Am J Clin Pathol 2003;120:93-100.
- Ordóñez NG. Value of the MOC-31 monoclonal antibody in differentiating epithelial pleural mesothelioma from lung adenocarcinoma. Hum Pathol 1998;29:166-9.
- Porcell AI, De Young BR, Proca DM, Frankel WL. Immunohistochemical analysis of hepatocellular and adenocarcinoma in the liver: MOC31 compares favorably with other putative markers. Mod Pathol 2000;13:773-8.

- Störkel S, Eble JN, Adlakha K, Amin M, Blute ML, Bostwick DG, et al. Classification of renal cell carcinoma: Workgroup No. 1. Union Internationale Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC). Cancer 1997;80:987-9.
- Ebele JN, Sauter G, Epstein JI, Sesterhenn IA. Pathology and genetics of tumours of the urinary system and male genital organs: World Health Organization classification of tumours. Lyon: IARC Press; 2004;10-43.
- Edge SB, Byrd DR, Compton CC, Fritz AG, Greene FL, Trotti A, et al. AJCC cancer staging manual. New York: Springer; 2010; 479-89.
- Thoenes W, Störkel S, Rumpelt HJ. Human chromophobe cell renal carcinoma. Virchows Arch B Cell Pathol Incl Mol Pathol 1985;48:207-17.
- 21. Krüger S, Sotlar K, Kausch I, Horny HP. Expression of KIT (CD117) in renal cell carcinoma and renal oncocytoma. Oncology

- 2005;68:269-75.
- Patton KT, Tretiakova MS, Yao JL, Papavero V, Huo L, Adley BP, et al. Expression of RON proto-oncogene in renal oncocytoma and chromophobe renal cell carcinoma. Am J Surg Pathol 2004;28: 1045-50.
- Carrion R, Morgan BE, Tannenbaum M, Salup R, Morgan MB. Caveolin expression in adult renal tumors. Urol Oncol 2003;21: 191-6.
- 24. Mete O, Kilicaslan I, Gulluoglu MG, Uysal V. Can renal oncocytoma be differentiated from its renal mimics? The utility of anti-mitochondrial, caveolin 1, CD63 and cytokeratin 14 antibodies in the differential diagnosis. Virchows Arch 2005;447:938-46.
- Garcia E, Li M. Caveolin-1 immunohistochemical analysis in differentiating chromophobe renal cell carcinoma from renal oncocytoma. Am J Clin Pathol 2006;125:392-8.