INSIGHT



## **BRAIN NETWORKS**

## **Modeling breathing rhythms**

Computational models are helping researchers to understand how certain properties of neurons contribute to respiratory rhythms.

## JAN-MARINO RAMIREZ AND NATHAN A BAERTSCH

**Related research article** Phillips RS, John TT, Koizumi H, Molkov YI, Smith JC. 2019. Biophysical mechanisms in the mammalian respiratory oscillator re-examined with a new data-driven computational model. *eLife* **8**:e41555. DOI: 10.7554/eLife.41555

rchestral music is beautiful, rich and complex, yet analyzing the contribution of any one instrument can be difficult. In many ways, neuroscientists face similar challenges when they try to understand how the networks in the brain work. These circuits are made of many types of neurons, each of which has different properties. Moreover, the properties of a neuron can change because of its interactions with other cells in the network.

Researchers have started to unravel this complexity by building computational models of both neurons and networks of neurons, and by focusing on pairs of interacting properties. This is similar to how one might study how the left and right hand of a violinist work together to produce melody and rhythm, and then use this knowledge to better understand other stringed instruments and their roles in the orchestra.

Within the 'rhythmogenic' networks of the brain, different types of neurons work together to create the body rhythms that are essential for life. For example, a complex network generates breathing rhythms, and it is often used to understand rhythmogenic circuits in general. The balance between excitatory and inhibitory connections between neurons is critical to shape network activity (*Ramirez and Baertsch, 2018*). Equally important, but less well understood, are the interactions between intrinsic properties that are built into individual neurons.

In particular, two intrinsic properties are thought to play a role in controlling breathing rhythms and network activity in general. The first is a persistent sodium current  $(I_{NaP})$ , which is slowly activated and inactivated by changes in the voltage across the neuronal membrane. The second does not depend on voltage: rather, the calcium-activated non-selective cation current  $(I_{CAN})$  is triggered when the level of calcium ions inside the cell increases. The source of these calcium ions is unknown, but there is experimental evidence that they could be provided by mechanisms at the synapses between neurons (Del Negro et al., 2010; Del Negro et al., 2011). Both I<sub>NaP</sub> and I<sub>CAN</sub> allow some neurons to generate rhythms on their own, without being stimulated by their neighbors. Yet, in the actual network, these properties also enhance the signals transmitted by excitatory synapses (Ramirez et al., 2004). Because I<sub>NaP</sub> and I<sub>CAN</sub> interact with synaptic properties, as well as with each other, it is difficult to isolate their relative contributions.

Pharmacological and genetic manipulations have shed light on how  $I_{NaP}$  and  $I_{CAN}$  work in the respiratory network (see, for example, *Koizumi et al., 2018; Picardo et al., 2019*), but there is still no consensus on how they contribute to rhythmogenesis (*Feldman and Del Negro, 2006; Ramirez et al., 2004*). Now, in eLife, Jeffrey Smith of the National Institute Neurological Disorders Stroke (NINDS) and colleagues – including Ryan Phillips as first author – report results from a computational modeling approach that re-examines how  $I_{NaP}$  and  $I_{CAN}$ control breathing (*Phillips et al., 2019*).

© Copyright Ramirez and Baertsch. This article is distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use and redistribution provided that the original author and source are credited.



**Figure 1.** How the intrinsic properties of neurons contribute to the breathing rhythm. Phillips et al. established a model of the brain network that controls breathing, and used it to deduce how two built-in properties of neurons, the  $I_{NaP}$  sodium current (blue) and the  $I_{CAN}$  current (green), control the breathing rhythm. Without  $I_{NaP}$ , the network is silent, with the neurons exhibiting non-rhythmic activity (grey). However, a small number of neurons with high levels of  $I_{NaP}$  activity can produce a weak rhythm, even in the absence of  $I_{CAN}$ . In turn, the model suggests that  $I_{CAN}$  is activated by calcium ions coming from synapses; the current would then amplify excitatory interactions between neurons. This amplification leads to additional neurons participating in the rhythm (red), producing a robust functional network.

The researchers, who are based at NINDS, the University of New Hampshire and Georgia State University, established a simplified model of the breathing network, which only takes into account excitatory synaptic interactions. Then, they tuned the contributions of three variables,  $I_{NaP}$ ,  $I_{CAN}$ , and the source of intracellular calcium ions, and explored how this affected the freguency and amplitude of breathing. First, they examined how changes in the source of calcium ions influenced the contribution of  $I_{\mbox{\scriptsize CAN}}$  to the network. If calcium came from within neurons, I<sub>CAN</sub> controlled the frequency, but not the strength, of breathing. On the other hand, if calcium depended on synaptic activity, I<sub>CAN</sub> acted as a synaptic amplifier to control the strength of the rhythm, but it had little effect on its frequency. This scenario best matched experimental data and prior conclusions (Del Negro et al., 2011; Koizumi et al., 2018), prompting Phillips

et al. to conclude that  $I_{CAN}$  is activated by calcium ions that are primarily of synaptic origin.

The group then went on to demonstrate that  $I_{CAN}$  does not establish the rhythm in their computational model, but that it increases the strength of breathing by recruiting more neurons to participate in the network. Indeed, when  $I_{CAN}$  was eliminated from the model network, a weak breathing rhythm remained, which was generated by a small number of neurons with high levels of  $I_{NaP}$  activity (*Figure 1*). Eliminating this 'rhythmogenic kernel' from the model stopped the rhythm altogether, in agreement with some (*Peña et al., 2004*), but not all (*Pace et al., 2007*) experimental data.

Unlike the model built by Phillips et al., the actual network that controls breathing is not exclusively excitatory, but is subjected to important inhibitory and neuromodulatory control. Further, alternative mechanisms of rhythmogenesis that do not depend on I<sub>NaP</sub> have also been

proposed (**Del Negro et al., 2010; Rubin et al., 2009**). Could different rhythmogenic mechanisms therefore contribute to breathing, depending on the demands of the network? In particular, could the contributions of  $I_{NaP}$  and  $I_{CAN}$  change based on the neuromodulatory state of the network? Addressing such questions will require further back-and-forth between experiments and increasingly complex models.

Overall, the results of Phillips et al. allow us to understand how intrinsic neuronal properties independently control the strength and frequency of the breathing rhythm. Their model is also a useful framework in which to explore how changes in the way  $I_{NaP}$  and  $I_{CAN}$  interact can dynamically impact rhythmogenic properties. For example, it could shed light on the way how the network reconfigures when the body lacks oxygen. Ultimately, describing the duet between  $I_{NaP}$  and  $I_{CAN}$  in the respiratory network may help dissect how rhythmic activity is controlled in other regions of the brain (*Penn et al., 2016*; *Riquelme et al., 2018*).

Jan-Marino Ramirez is in the Center for Integrative Brain Research, Seattle Children's Research Institute, and the departments of Pediatrics and Neurological Surgery, University of Washington, Seattle, United States

jan.ramirez@seattlechildrens.org b http://orcid.org/0000-0002-5626-3999

Nathan A Baertsch is in the Center for Integrative Brain Research, Seattle Children's Research Institute, Seattle, United States

b http://orcid.org/0000-0003-1589-5575

Competing interests: The authors declare that no competing interests exist. Published 25 March 2019

## References

Del Negro CA, Hayes JA, Pace RW, Brush BR, Teruyama R, Feldman JL. 2010. Synaptically activated burst-generating conductances may underlie a grouppacemaker mechanism for respiratory rhythm generation in mammals. *Progress in Brain Research* **187**:111–136. DOI: https://doi.org/10.1016/B978-0-444-53613-6.00008-3, PMID: 21111204 Del Negro CA, Hayes JA, Rekling JC. 2011. Dendritic calcium activity precedes inspiratory bursts in preBötzinger complex neurons. *Journal of Neuroscience* **31**:1017–1022. DOI: https://doi.org/10. 1523/JNEUROSCI.4731-10.2011, PMID: 21248126 Brain Networks | Modeling breathing rhythms

Feldman JL, Del Negro CA. 2006. Looking for inspiration: new perspectives on respiratory rhythm. *Nature Reviews Neuroscience* **7**:232–241. DOI: https:// doi.org/10.1038/nrn1871, PMID: 16495944 **Koizumi H**, John TT, Chia JX, Tariq MF, Phillips RS, Mosher B, Chen Y, Thompson R, Zhang R, Koshiya N, Smith JC. 2018. Transient receptor potential channels TRPM4 and TRPC3 critically contribute to respiratory motor pattern formation but not rhythmogenesis in rodent brainstem circuits. *Eneuro* **5**:ENEURO.0332-17. 2018. DOI: https://doi.org/10.1523/ENEURO.0332-17. 2018, PMID: 29435486

Pace RW, Mackay DD, Feldman JL, Del Negro CA.
2007. Role of persistent sodium current in mouse preBötzinger complex neurons and respiratory rhythm generation. *The Journal of Physiology* 580:485–496.
DOI: https://doi.org/10.1113/jphysiol.2006.124602
Peña F, Parkis MA, Tryba AK, Ramirez JM. 2004.
Differential contribution of pacemaker properties to the generation of respiratory rhythms during normoxia and hypoxia. *Neuron* 43:105–117. DOI: https://doi. org/10.1016/j.neuron.2004.06.023, PMID: 15233921
Penn Y, Segal M, Moses E. 2016. Network

synchronization in hippocampal neurons. *PNAS* **113**: 3341–3346. DOI: https://doi.org/10.1073/pnas. 1515105113, PMID: 26961000

**Phillips RS**, John TT, Koizumi H, Molkov YI, Smith JC. 2019. Biophysical mechanisms in the mammalian respiratory oscillator re-examined with a new datadriven computational model. *eLife* **8**:e41555. DOI: https://doi.org/10.7554/eLife.41555

Picardo MCD, Sugimura YK, Dorst KE, Kallurkar PS, Akins VT, Ma X, Teruyama R, Guinamard R, Kam K, Saha MS, Del Negro CA. 2019. Trpm4 ion channels in pre-Bötzinger complex interneurons are essential for breathing motor pattern but not rhythm. *PLOS Biology* **17**:e2006094. DOI: https://doi.org/10.1371/journal. pbio.2006094, PMID: 30789900

Ramirez JM, Tryba AK, Peña F. 2004. Pacemaker neurons and neuronal networks: an integrative view. *Current Opinion in Neurobiology* **14**:665–674. DOI: https://doi.org/10.1016/j.conb.2004.10.011, PMID: 15582367

Ramirez JM, Baertsch N. 2018. Defining the rhythmogenic elements of mammalian breathing. *Physiology* **33**:302–316. DOI: https://doi.org/10.1152/ physiol.00025.2018, PMID: 30109823

**Riquelme D**, Silva I, Philp AM, Huidobro-Toro JP, Cerda O, Trimmer JS, Leiva-Salcedo E. 2018. Subcellular localization and activity of TRPM4 in medial prefrontal cortex layer 2/3. *Frontiers in Cellular Neuroscience* **12**:12. DOI: https://doi.org/10.3389/ fncel.2018.00012, PMID: 29440991

Rubin JE, Hayes JA, Mendenhall JL, Del Negro CA. 2009. Calcium-activated nonspecific cation current and synaptic depression promote network-dependent burst oscillations. *PNAS* **106**:2939–2944. DOI: https:// doi.org/10.1073/pnas.0808776106, PMID: 19196976